

Grafting of Thiazole Derivative on Chitosan Magnetite Nanoparticles for Cadmium Removal—Application for Groundwater Treatment

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Table S1. a. Reminder on equations used for modeling uptake kinetics [1,2] [3].

Model	Equation	Parameters	Ref.
PFORE	$q(t) = q_{eq,1}(1 - e^{-k_1 t})$	$q_{eq,1}$ (mmol g ⁻¹): sorption capacity at equilibrium k_1 (min ⁻¹): apparent rate constant of PFORE	[3]
PSORE	$q(t) = \frac{q_{eq,2}^2 k_2 t}{1 + k_2 q_{eq,2} t}$	$q_{eq,2}$ (mmol g ⁻¹): sorption capacity at equilibrium k_2 (g mmol ⁻¹ min ⁻¹): apparent rate constant of PSORE	[3]
RIDE	$\frac{q(t)}{q_{eq}} = 1 - \sum_{n=1}^{\infty} \frac{6\alpha(\alpha+1)\exp\left(\frac{-D_e q_n^2}{r^2} t\right)}{9 + 9\alpha + q_n^2 \alpha^2}$ With q_n being the non-zero roots of $\tan q_n = \frac{3 q_n}{3 + \alpha q_n^2}$ and $\frac{m q}{V C_0} = \frac{1}{1 + \alpha}$	D_e (m ² min ⁻¹): Effective diffusivity coefficient	[1]

(m (g): mass of sorbent; V (L): volume of solution; C₀ (mmol L⁻¹): initial concentration of the solution).

Table S1. b. Reminder on equations used for modeling sorption isotherms [2, 4, 5].

Model	Equation	Parameters	Ref.
Langmuir	$q_{eq} = \frac{q_{m,L} C_{eq}}{1 + b_L C_{eq}}$	$q_{m,L}$ (mmol g ⁻¹): Sorption capacity at saturation of monolayer b_L (L mmol ⁻¹): Affinity coefficient	[2]
Freundlich	$q_{eq} = k_F C_{eq}^{1/n_F}$	k_F and n_F : empirical parameters of Freundlich equation	[2]
Sips	$q_{eq} = \frac{q_{m,S} b_S C_{eq}^{1/n_S}}{1 + b_S C_{eq}^{1/n_S}}$	$q_{m,L}$, b_S and n_S : empirical parameters of Sips equation (based on Langmuir and Freundlich equations)	[2]
Temkin	$q_{eq} = \frac{R T}{b_T} \ln(A_T C_{eq})$	b_T : J kg mol ⁻² , Temkin isotherm constant A_T : L mol ⁻¹ , Temkin isotherm equilibrium constant	[5]

Akaike Information Criterion, AIC [6]:

$$AIC = N \ln \left(\frac{\sum_{i=0}^N (y_{i,exp.} - y_{i,model})^2}{N} \right) + 2N_p + \frac{2N_p(N_p + 1)}{N - N_p - 1}$$

Where N is the number of experimental points, N_p the number of model parameters, y_{i,exp.} and y_{i,model} the experimental and calculated values of the tested variable.

Table S2. Assignments of the metal concentration comparing with the MCL values for drinking water and livestock drinking water.

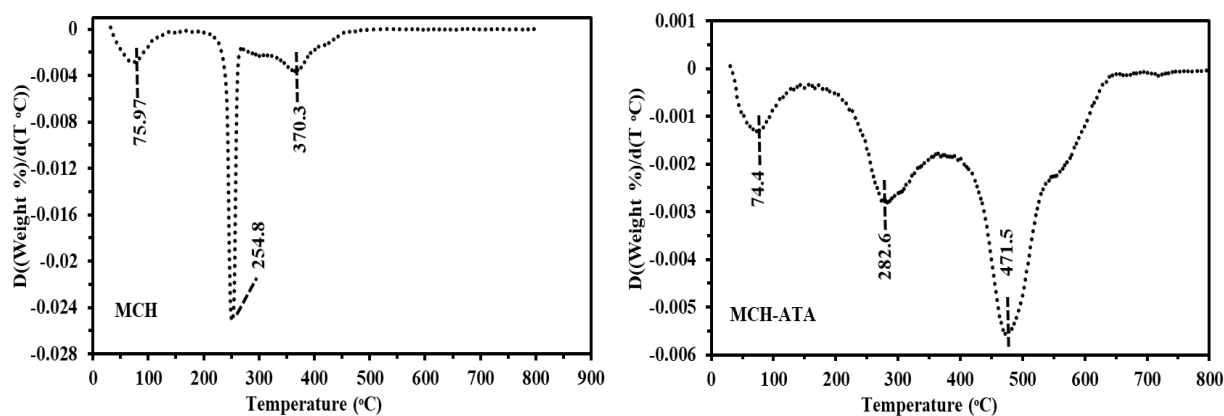
	Initial conc.	MCL (D.W)	MCL (L.W)	D.W increased time	L.W increased time
Cd (II)	1.97	0.003	0.05	656.67	39.4
Zn (II)	7.64	5	24	1.528	0.318
Pb (II)	0.997	0.01	0.1	99.7	9.97
Hg(II)	0.759	0.002	0.01	379.5	75.9
Cu(II)	2.19	1.3	0.5	1.685	4.38
Al(III)	6.943	0.2	5	34.715	1.389

Table S3. Textural properties of MCH and MCH-ATA.

Parameter	MCH	MCH-ATA
S_{BET} ($\text{m}^2 \text{g}^{-1}$)	21.17	22.5
V_{p} ($\text{cm}^3 \text{STP g}^{-1}$)	7.14	7.7

Table S4. Elemental analysis of MCH and MCH-ATA sorbents.

	MCH					MCH-ATA					
	C	N	H	O	Fe	C	N	H	O	Fe	S
[%]	26.97	4.28	4.96	31.61	32.18	18.5	6.09	4.18	35.94	30.96	4.33
Mmol	22.456	3.056	49.206	19.758	5.762	15.404	4.348	41.468	22.464	5.544	1.35

**Figure S1.** Tora Hellwan area where real contaminated samples were collected.**Figure S2.** DrTG data for MCH and after functionalization (MCH-ATA).

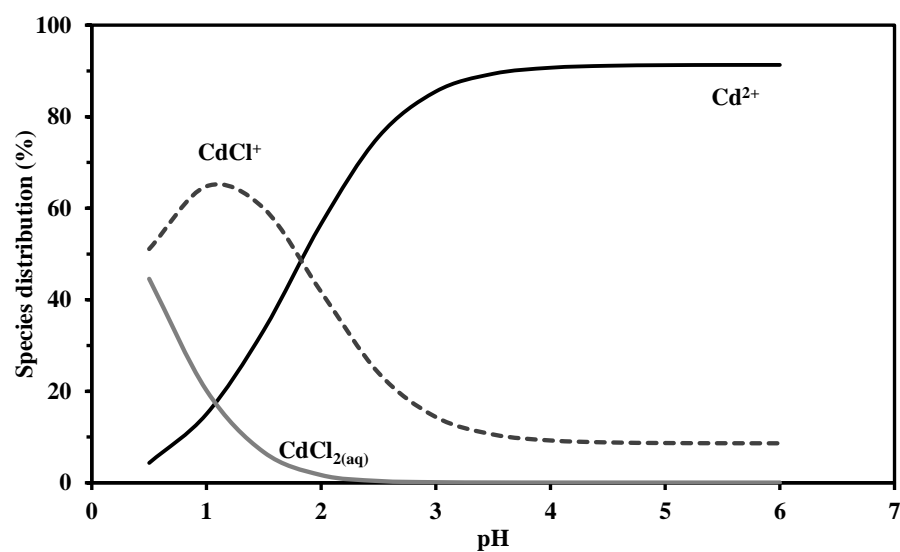


Figure S3. Speciation diagrams for Cd(II) under the experimental conditions that selected for the study of pH effect (calculation using Visual Minteq, [7]).

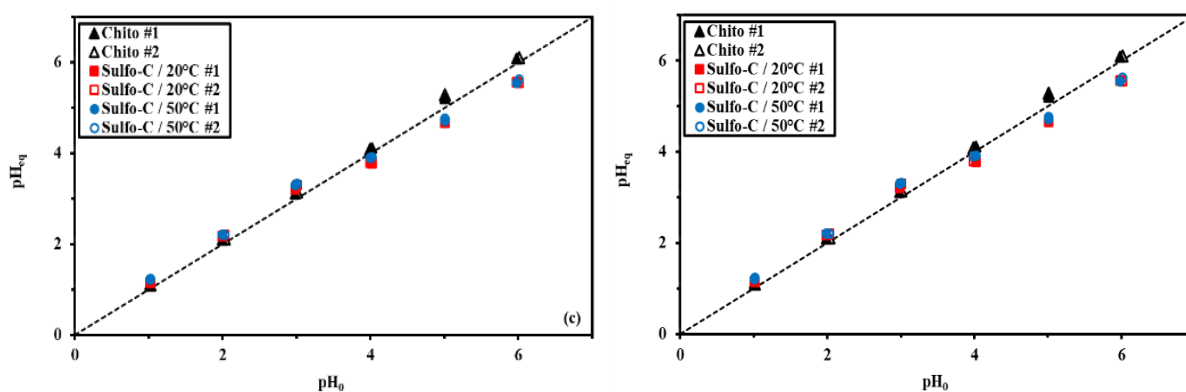


Figure S4. Variation in the pH (from 1-5) for MCH and MCH-ATA.

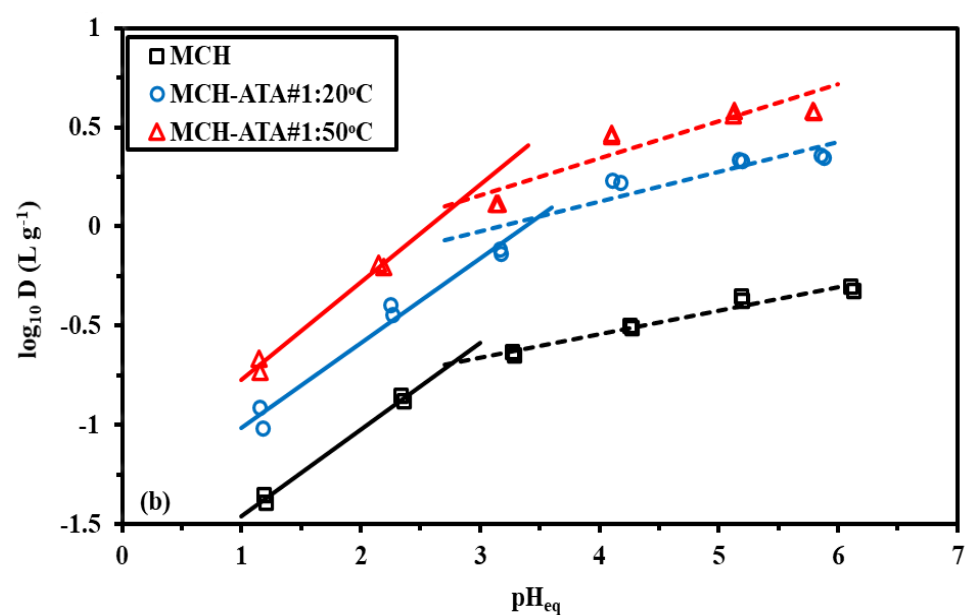


Figure S5. Plotting of pH_{eq} vs $\text{Log}_{10}D$ at 20 and 50°C.

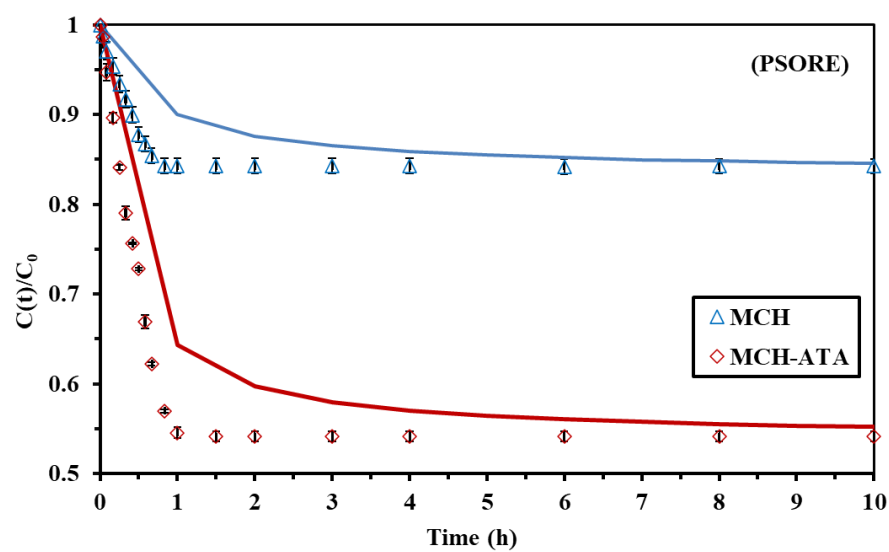


Figure S6. Average values of kinetic modeling with error bars of the triplicated experiments for PSORE of MCH (a) and MCH-ATA(b).

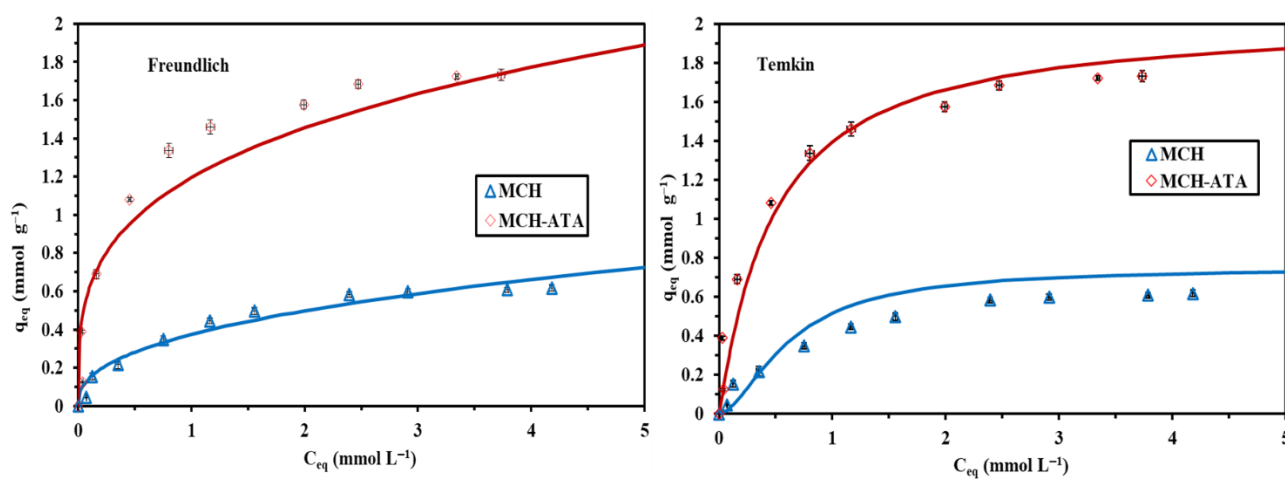


Figure S7. Average values of Freundlich and Temkin equations with error bars of the triplicated experiments for Cd(II) sorption on MCH and MCH-ATA sorbents.

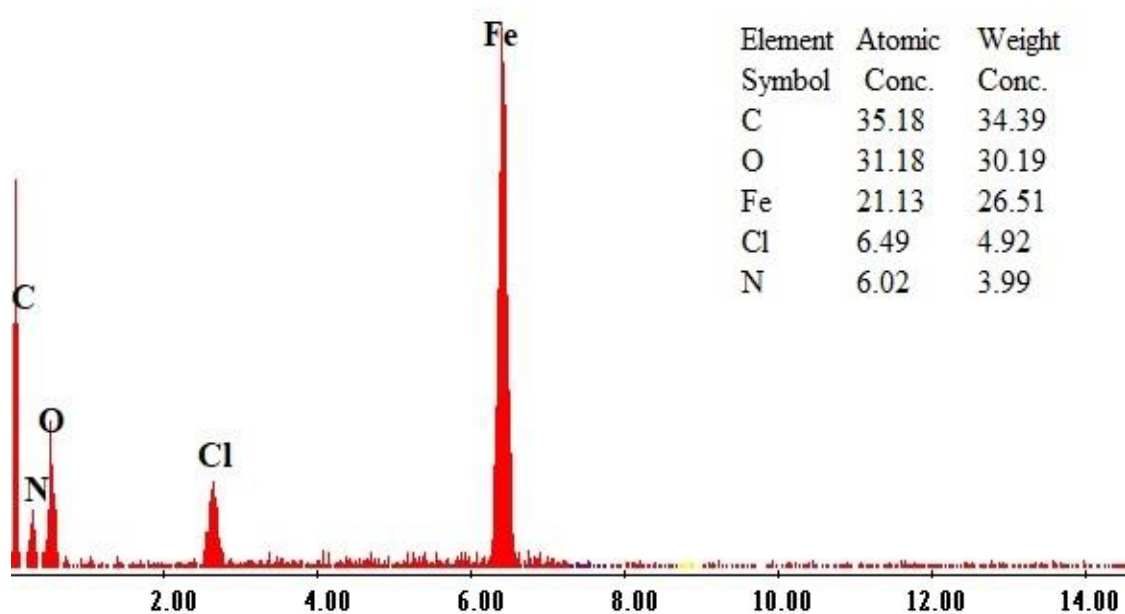


Figure S8. EDX analysis of the MCH-ATA sorbent after 5 cycles of sorption-desorption.

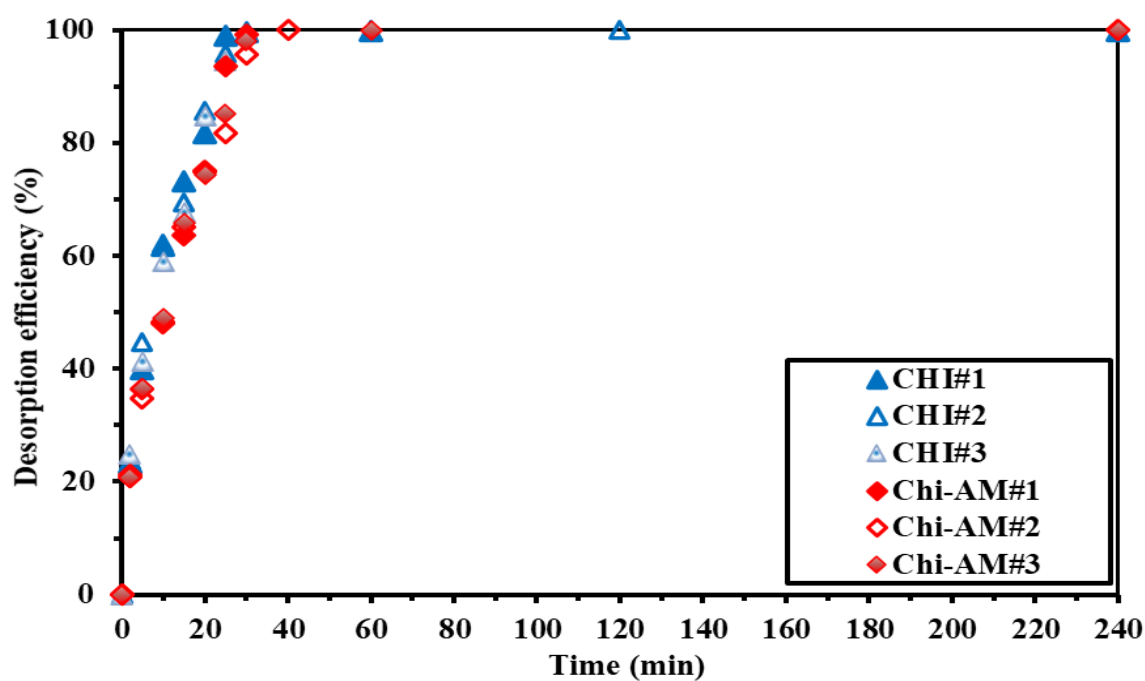


Figure S9. Desorption kinetics of MCH and MCH-ATA using 0.2M HCl solution.

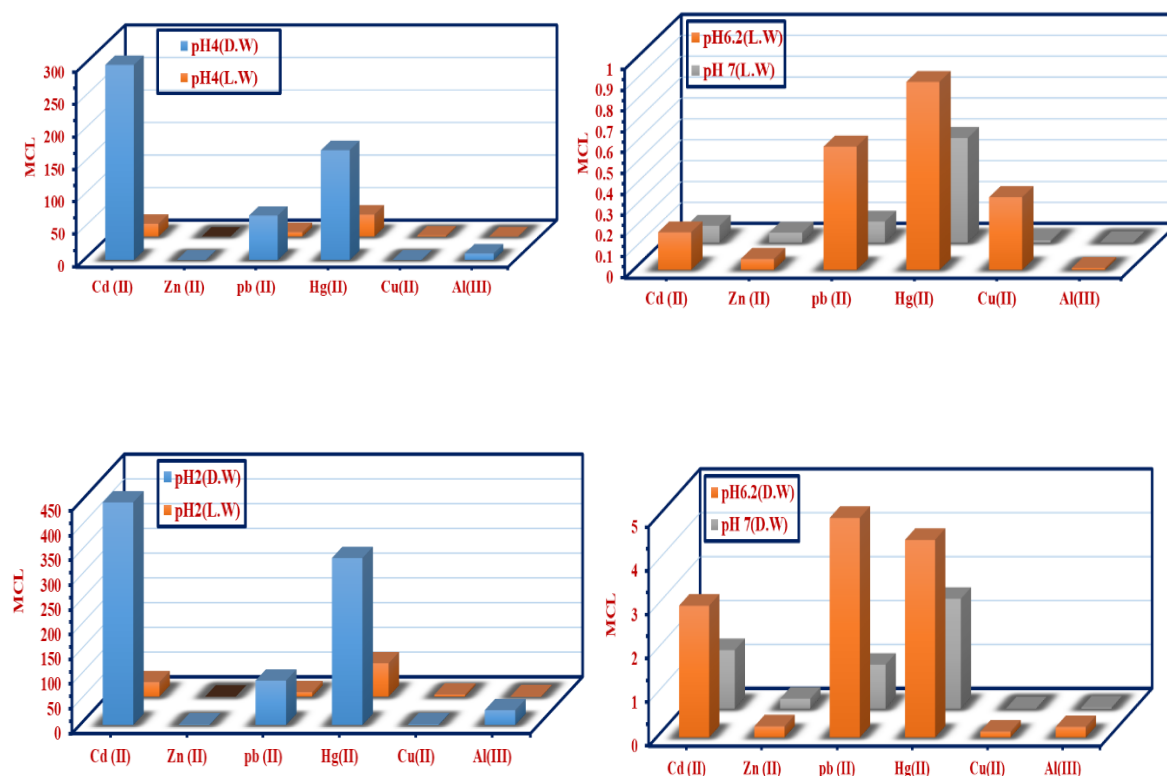


Figure S10. Comparison of the MCL levels for drinking water and Livestock drinking water for MCH-ATA at different pH values.

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