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Short-Term Effects of Spent Mushroom Substrate Mulching Thickness on the Soil Environment, Weed Suppression, Leaf Nutrients, and Nut Characteristics in a Hazelnut Orchard

Zhong Ma 1,†, Yun-Qi Zhang 2,† [,](https://orcid.org/0000-0003-0260-0491) Lu-Jun Wang ³ , Guang-Long Hu [2](https://orcid.org/0000-0001-6060-1826) , Xiao-Qiang Gong ⁴ , Qian Bai ¹ , Shu-Chai Su 1,[*](https://orcid.org/0000-0002-9994-6896) and Jian-Xun Qi 2,*

- ¹ The Key Laboratory for Silviculture and Conservation of the Ministry of Education, Beijing Forestry University, Beijing 100083, China; mazhong@bjfu.edu.cn (Z.M.); baiqian0219@bjfu.edu.cn (Q.B.)
- ² Beijing Academy of Forestry and Pomology Sciences, Beijing 100093, China; zhyq1985@bjfu.edu.cn (Y.-Q.Z.); hglcau@gmail.com (G.-L.H.)
- ³ Anhui Academy of Forestry, Hefei 230031, China; wanglujun1984@126.com
- ⁴ State Key Joint Laboratory of Environmental Simulation and Pollution Control, School of Environment, Tsinghua University, Beijing 100084, China; gongxiaoqiang@126.com
- ***** Correspondence: sushuchai@sohu.com (S.-C.S.); qijx@263.net (J.-X.Q.)
- These authors contributed equally to this work.

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Abstract: Worldwide, a huge amount of solid fermented waste is produced every year during mushroom production. The rational utilization of spent mushroom substrate (SMS) is conducive to environmental protection and the sustainable development of agriculture. The aims of this work were to analyze the effects of the SMS mulching thickness on the soil environment, weed suppression, leaf nutrients, and nut traits in a hazelnut plantation and provide a theoretical basis for the scientific and reasonable utilization of SMS. An SMS mulching experiment with four mulching thickness treatments (MT1: 5 cm, MT2: 10 cm, MT3: 15 cm, and CK: 0 cm) was conducted in a semi-arid area of North China in 2019. The soil properties, weed control efficiency, leaf nutrient content, and nut yield and quality characteristics of these treatments were all measured and analyzed. The results showed that (1) the average soil temperatures of MT2 and MT3 at different soil depths were 0.73–1.78 \degree C and 1.18–2.50 \degree C lower than when no mulch was used in warm weather and 0.67– 1.05 ◦C and 0.99–1.56 ◦C higher than when no mulch was used in cold weather. The average soil moisture content of MT1, MT2, and MT3 at different soil depths was 6.27–8.13%, 10.66–17.23%, and 7.26–12.81% higher than that of CK, respectively. There were no significant differences in the soil bulk density or porosity among the four treatments. The average soil nutrient contents (soil organic matter and available N, P, and K) of MT1, MT2, and MT3 were 7.05–15.23%, 14.90–38.93%, and 17.42–40.11% higher than those of CK, respectively. Soil enzymatic activities of these treatments followed the order MT3 \approx MT2 > MT1 > CK. (2) MT2 and MT3 both had high levels of weed control efficiency: 55.66–92.93% and 70.43–97.90%, respectively. (3) The leaf N, P, K, and SPAD increased significantly under MT2 and MT3 by about 10% compared with the CK. (4) MT2 effectively increased the nut size, nut and kernel mass, and crude fat content of the nut (CFC). In general, the short-term field experiment clearly showed that SMS is a superior mulching material for improving the soil environment and plant growth, and 10 cm thickness is suitable for mulching practice.

Keywords: spent mushroom substrate; mulching measures; soil properties; weed control; nut traits

1. Introduction

The genus *Corylus* L. (hazelnut) includes about 20 species that are widely distributed among temperate regions of Asia, Europe, and North America. It is regarded as one of the most prevalent nut crops in the world. According to the FAO statistics from 2017, the total production of in-shell hazelnuts worldwide exceeded 1 million tons, and the main world producers were Turkey (675,000 t), Italy (131,281 t), Azerbaijan (43,000 t), USA

 $(29,030 \t{t})$, and China $(27,044 \t{t})$ [\[1\]](#page-17-0). Hazelnut is a nutrient-dense food owing to its rich concentrations of protein and oil. In addition, hazelnut provides appreciable amounts of sterols, tocopherols, polyphenols, essential minerals, and B-complex vitamins [\[2\]](#page-17-1). Owing to its pleasant nutty taste and flavor, hazelnut is widely used for confectionery and bakery products [\[3\]](#page-17-2). In recent years, hazelnut planting has been vigorously developed in China, mainly in North China and Northeast China. Although the planting area and yield have increased rapidly, outstanding problems (improper agricultural management, seasonal drought stress, soil degradation, and so on) still restrict the development of the planting industry.

Many studies have shown that mulching is an effective strategy for soil management as it improves the tillage quality by increasing the soil's biological activity, regulating the soil temperature, controlling weed growth, reducing soil evaporation, and preventing nutrient leaching [\[4\]](#page-17-3). It also provides benefits for growth, annual and perennial yield, and the health of plants, and its low cost makes it a widespread technique. Different mulching methods and materials have different effects on soil properties and plant growth. A meta-analysis was conducted based on 266 peer-reviewed publications from China, and the results showed that black film mulching is more favorable to crop growth and an increase in the crop yield and water use efficiency (WUE) in arid areas [\[5\]](#page-17-4). However, in the long term, the application of plastic film may cause serious soil pollution, and hence have negative effects on crop production [\[6\]](#page-17-5). Compared with inorganic mulches, organic mulching could have more benefits for soil health and sustainable development through supplementing with soil organic matter, increasing the water holding capacity, releasing nutrients and enhancing soil aggregation, and promoting soil biological activity [\[7\]](#page-18-0). The selection of an appropriate organic mulching material is strongly affected by the local climate, cost-effectiveness, and feasibility for the crop [\[8\]](#page-18-1).

Mushrooms have drawn a lot of interest owing to their nutritional and medicinal value, and they have been widely cultivated. Nowadays, a total of 34 billion kg of mushrooms are produced annually throughout the world [\[9\]](#page-18-2). As the largest edible mushroom producer and exporter in the world, China's main production areas are Henan, Fujian, Shandong, and Hebei province. Pingquan County, which belongs to Hebei Province, is the main *pholiota nameko* production base in northern China. After the mushroom fruiting bodies are harvested, a huge amount of solid fermented waste is produced. Because a high level of organic matter remains present in the spent mushroom substrate (SMS), it should be properly disposed of to avoid environmental pollution problems. Previously, the major applications of SMS were animal fodder, composting material, and bioenergy feedstock [\[10\]](#page-18-3). However, most SMS is still stacked on farmland or incinerated at will. SMS has become the biggest challenge for mushroom farms in terms of disposal management, although studies have confirmed that SMS is a good biofertilizer or soil improver for crop production thanks to its high amounts of macro- and micronutrients, high cation exchange capacity, near-neutral pH, high porosity, and high water-holding capacity [\[11\]](#page-18-4). SMS, with its biodegradability, availability, and sustainability, could be a promising alternative to traditional plastic mulch in the context of global plastic-reduction behavior.

At present, there is little information about the use of SMS as a mulching material and the assessment of the effects of mulching thickness on the soil–plant system. The efficacy of SMS mulch needs to be verified and evaluated. Keeping the above facts in mind, this study focused on four aspects regarding the short-term effects of mulching thickness: (1) the physico-chemical and biological properties of the soil, (2) weed suppression, (3) leaf nutrients, and (4) nut characteristics. The purpose of this study is to find a suitable SMS mulching measure to improve the soil health and nut quality. The results of this study could be used as references for SMS utilization and environmentally friendly mulching practices.

2. Materials and Methods

2.1. Description of Experimental Sites and Plant Materials

The field experiment was conducted from October 2018 to December 2019 in the North experiment station of Beijing Forestry University (118°40'46" E, 41°13'18" N), Sudayingzi village, Pingquan County, Hebei Province, China. It is located in a temperate monsoon climate zone. The annual average temperature in the past ten years has been about $7.5 \text{ }^{\circ}\text{C}$. The average annual rainfall in the past ten years has been about 540 mm, and this has been mainly concentrated in summer. The average number of annual sunshine hours in the past ten years has been about 2500 h with a frost-free period of 120–130 d. The local agrometeorological data for 2019 are depicted in Table [1.](#page-2-0) The soil texture of the experiment plot is sandy loam soil. The soil total nitrogen, available phosphorus, available potassium, alkali hydrolyzed nitrogen, and organic matter contents are 0.83 ± 0.06 , 15.67 ± 2.41 , 105.56 ± 10.63 , 32.13 ± 2.45 , and 13.43 ± 1.17 , respectively; the soil pH value is 6.5. The hazelnut plantation is mainly planted with the hybrid hazelnut *Corylus heterophylla* × Corylus avellana '82-11'. This cultivar was planted in 2013 with plant and row spacings of 2 and 3 m, respectively.

2.2. Experimental Design and Treatments

The mulching experiment was conducted in October 2018 with a total of four treatments. The treatments were non-mulching control (CK), a mulch thickness of 5 cm (MT1), a mulch thickness of 10 cm (MT2), and a mulch thickness of 15 cm (MT3). This experiment had a randomized complete block design, where each mulching treatment was replicated three times and the plot area of each treatment was 40 m^2 (8 m \times 5 m). A diagram of the experimental design is shown in Figure [1.](#page-3-0) The mulching material used was residue from the local production of *Pholiota nameko*. The main component of the residue was sawdust from the branches of *Robinia pseudoacacia* that had not been completely decomposed. The total nitrogen content, available potassium content, organic matter content, and pH value of the SMS were 30.74 ± 2.36 , 232.78 ± 20.64 , 421.69 ± 30.82 , and 5.4, respectively.

Figure 1. Schematic diagram of the randomized block experiment design. CK, control; MT, mulch thickness.

2.3. Sampling

The soil located 0–20 cm under the mushroom mulch was sampled with a stainlesssteel cylindrical drill at 15-day intervals from May to December in 2019. Five soil sampling points were selected in each experimental plot by the S-shape sampling method. In each plot, one soil sample was composed of five individual samples, and about 1 kg of soil was reserved by the coning and quartering method. The samples from different treatments were dried in a ventilation place and then sieved by stainless-steel sieves with pore sizes of 1.00 and 0.25 mm. The pH value, available nutrient content, and enzyme activity of the soil samples (<1.00 mm) were used. The soil samples (<0.25 mm) were used to determine the total nutrient content and organic matter content of soil.

About 30 mature leaves in the upper or middle parts of the branches of the tested trees were also collected from each experimental plot from May to October in 2019. The fresh leaves were dried at a low temperature (35 °C) to ensure their weights were constant. Then, the dried leaves were crushed with a pulverizer. Finally, the classified powder was obtained through a 100-mesh steel sieve and prepared for nitrogen, phosphorus, and potassium analyses.

2.4. Measurements

2.4.1. Soil Moisture and Temperature

A remote real-time monitoring instrument (Patent No. CN202648697U) was installed in each plot to measure soil moisture content (accuracy: $\pm 3\%$) and soil temperature (accuracy: ± 0.5 °C). Each instrument was connected with three moisture sensor probes and three temperature sensor probes. The moisture sensor probes were set at 15, 30, and 45 cm underground, and the temperature sensor probes were set at the surface level of the ground, 10 cm underground, and 20 cm underground. This instrument automatically records data every 10 min. The recorded data can be viewed online through the data transmission module.

2.4.2. Physico-Chemical and Biological Properties of Soil

Undisturbed topsoil cores were collected with a ring knife (diameter, 50.46 mm; length, 50 mm) and dried to a constant weight in an oven at 105 °C to measure the bulk density and total porosity [\[12\]](#page-18-5). Soil organic matter (SOM) was determined with the potassium dichromate and sulfuric acid method [\[13\]](#page-18-6). Alkaline hydrolysable-nitrogen (AH-N) was measured by the alkali solution diffusion method [\[14\]](#page-18-7). Available phosphorus (AP) was extracted with 0.5 mol/L NaHCO₃ (pH 8.5) and determined by molybdenum-antimony anti-colorimetry with an ultraviolet spectrophotometer (Lambda750, PerkinElmer Inc., Waltham, MA, USA). Available potassium (AK) was determined in ammonium acetate

extracts by flame photometry with an atomic absorption spectrophotometer (AA-240, Varian Inc., Palo Alto, CA, USA) [\[15\]](#page-18-8).

Urease activity was measured using the phenol sodium hypochlorite colorimetric method and expressed in mg of NH_4^+ -N in 1 g of dry soil after 24 h. Invertase activity was determined by the 3,5-dinitrosalicylic acid colorimetric method and shown as mg glucose in 1 g dry soil after 24 h. Catalase activity was determined using the $KMnO₄$ titration method and shown as the consumption volume (mL) of 0.1 mol·L⁻¹ KMnO₄ in 1 g of dry soil after 20 min [\[16\]](#page-18-9).

2.4.3. Weed Investigation

In order to determine the effect of mulching measures on weed growth, three small plots of 0.6 m \times 0.6 m were randomly selected from each plot to investigate weed suppression in June, August, and October of 2019. Using statistics on the number of weed species and the fresh weight of the weeds, the controlling effect of mulching measures on weeds was reflected. The weed control efficiency (WCE) can be calculated using the following formula [\[17](#page-18-10)[,18\]](#page-18-11):

$$
WCE = (DMC - DMT)/DMC \times 100
$$

where DMC is the dry matter of weeds subjected to CK treatment, and DMT is the dry matter of weeds in a particular treatment.

2.4.4. Leaf Nutrients and Chlorophyll Content

The leaf N concentration was determined using the micro-Kjeldahl method with a nitrogen determinator (Kjeltec™ 8000, FOSS Group, Hiller, Denmark) [\[19\]](#page-18-12). The total leaf P concentrations were measured with a molybdate/stannous chloride method with an ultraviolet spectrophotometer (Lambda750, PerkinElmer Inc., Waltham, MA, USA) [\[20\]](#page-18-13). The leaf K concentrations were determined by atomic absorption spectrometry with an atomic absorption spectrophotometer (AA-240, Varian Inc., Palo Alto, CA, USA) [\[21\]](#page-18-14). The chlorophyll content in leaves was estimated using a hand-held chlorophyll meter (SPAD-502, Konica Minolta Optics, Osaka, Japan) [\[22\]](#page-18-15).

2.4.5. Nut Traits

Hazelnuts exposed to different mulch treatments were harvested in October 2019, and 100 nuts were randomly selected form each treatment to determine the nut traits. The experimental procedure included the following steps: (1) three linear dimensions of hazelnuts—length (*L*), width (*W*), and thickness (*T*)—were measured in mm with an electronic digital caliper [\[23\]](#page-18-16). (2) The nuts were dried in an oven at 40 °C for 5 days to reach a constant weight, and then the dry nuts' mass (*NM*) was weighed with a laboratory balance that was accurate to 0.01 g (Practum5101-1CN, Sartorius Group, Goettingen, Germany). (3) The dry kernels were taken out of the nut shells using a hammer, and the mass of the dry kernels was weighed (*KM*). (4) The kernels were ground via ball milling (Retsch MM400, Verder Group, Haan, Germany) with liquid nitrogen to produce powder samples. (5) The powder samples were used to determine the crude protein, soluble sugar, and crude fat contents (*CPC*, *SSC*, and *CFC*).

The soluble sugar concentration was determined by colorimetry with anthrone reagent in an ultraviolet spectrophotometer at 620 nm. A standard glucose curve was plotted, and the soluble sugar and glucose concentrations were interpolated from it. The crude protein content of the kernel samples was determined with the Kjeldahl method. The protein content was calculated using a factor of 6.25 for the N content [\[24\]](#page-18-17). The total fat content was obtained using the Soxhlet extraction method [\[24\]](#page-18-17). The crude fat content was calculated by dividing the dried extracted oil mass by the dried powder sample mass.

To describe the main morphological characteristics of nuts, some physical parameters were calculated according to Equations [\[23\]](#page-18-16):

$$
V = \frac{\pi LWT}{6}, \ D = (LWT)^{\frac{1}{3}}, \ \varnothing = \frac{D}{L}, \ PK = \frac{KM}{NM} \times 100, \ NSI = \frac{W + T}{2L}
$$

where *V* is the volume of nuts (cm³), π is the circumference ratio (\approx 3.14), *L* is the length of the nut (mm), *W* is the width of the nut (mm), *T* is the thickness of the nut (mm), *D* is the geometric mean diameter of the nut (mm), ∅ is the sphericity of the nut, *PK* is the kernel percentage, *KM* is the kernel mass (g), *NM* is the nut mass (g), and *NSI* is the nut shape index.

2.5. Statistical Analysis

Each determination was performed in triplicate, and the data are reported as means \pm standard deviations. The data variables from this study were subjected to an analysis of variance using SPSS statistical software package version 18.0 (SPSS Inc., Chicago, IL, USA), which was also used to carry out Duncan's multiple comparisons test to analyze the differences in various parameters in different experimental treatments. For all statistical analyses, $p \le 0.05$ was considered significant and $p \le 0.01$ was deemed to be very significant. All figures in this manuscript were drawn with Excel 2010.

3. Results

3.1. Effect of the Mulching Thickness on Soil Physico-Chemical and Biological Properties

3.1.1. Soil Temperature

There were significant differences in the monthly variation of the soil temperature at different soil depths for each treatment (Table [2\)](#page-6-0). During the field experiment period, for each treatment, the soil temperature at different soil depths showed a trend of first rising and then falling, reaching a peak in August and a valley in December. The average topsoil temperature of CK was 0.77, 1.32, and 1.81 ◦C higher than that of MT1, MT2, and MT3 in the growth period (from May to October), respectively. However, in the dormancy period (from November to December), the average topsoil temperature of CK was 0.32, 0.75, and 1.17 ◦C lower than that of MT1, MT2, and MT3, respectively. There were significant differences in the topsoil temperature between the MT2, MT3, and CK treatments throughout the study period.

The average soil temperature during the mulching treatments was significantly lower than that of CK in the growth period at a depth of 10 cm with magnitudes of 16.64 \pm 0.14, 16.28 ± 0.16 , and 15.86 ± 0.13 °C for MT1, MT2, and MT3, respectively. During the dormancy period, the average soil temperature of CK was 0.42, 1.02, and 1.56 ◦C lower than that of MT1, MT2, and MT3, respectively. There were significant differences in the soil temperature of at a depth of 10 cm between the mulching and non-mulching treatments.

Compared with CK, at a depth of 20 cm, MT1, MT2, and MT3 decreased the soil temperature by an average of 0.30, 0.51, and 0.83 ℃ in the growth period and increased the soil temperature by 0.26, 0.71, and 1.05 \degree C in the dormancy period. There were significant differences in the soil temperature between the MT3 and CK treatments during the study period at a depth of 20 cm.

3.1.2. Soil Water

In this study, the soil water content was investigated at depths of 15, 30, and 45 cm because the fine-roots of *C. heterophylla* × *C. avellana* are mainly distributed in the depth range of 0–50 cm [\[25\]](#page-18-18). The soil water content was significantly affected by the mulching practices, and it varied spatially and temporally (Table [3\)](#page-6-1).

Table 2. Monthly average soil temperature (◦C) at different soil depths during the field experiment period for the mulch treatments.

Note: Lowercase letters represent significant differences $(p < 0.05)$ among different months for the same treatment; capital letters represent significant difference $(p < 0.05)$ among different treatments in same month.

Soil Water	Mulch Treatments	May	June	July	August	September	October	November	December
15 cm depth	MT ₁	19.64 ± 1.06 aA	22.47 ± 1.23 bcA	$23.35 \pm 1.43 \text{ bA}$	$20.11 + 1.02$ ad A	21.11 ± 1.05 acd A	21.36 ± 1.02 cdA	19.78 ± 0.97 adA	9.25 ± 0.46 eA
	MT ₂	$20.01 + 1.09$ a A	$25.54 + 1.53$ bB	$26.31 + 0.89$ bB	$22.32 + 0.96$ cdB	$22.69 + 1.10 cA$	$22.27 \pm 1.00 \text{ c}dA$	20.84 ± 0.66 adAB	$10.34 + 0.38$ eB
	MT ₃	$20.90 + 3.37$ a A	21.73 ± 1.16 aA	22.38 ± 1.10 aAC	$20.89 + 1.27$ a A B	21.05 ± 1.31 aA	$21.63 + 1.17$ a A	21.46 ± 0.47 aB	13.86 ± 0.47 bC
	CK	$15.87 + 1.25$ aB	20.32 ± 1.77 bA	$20.51 + 1.37$ bC	$19.64 + 1.32$ bcA	$20.84 + 1.11$ bA	$21.01 + 0.74$ bA	$18.33 + 0.99$ cC.	$8.78 + 0.46$ dA
30 cm depth	MT1	20.90 ± 0.50 aAB	24.54 ± 1.42 bAB	24.71 ± 1.01 bAB	21.87 ± 1.24 acAB	22.35 ± 1.07 acAB	23.27 ± 0.76 bcAB	21.24 ± 0.86 aA	12.26 ± 0.33 dA
	MT ₂	$22.66 + 1.28$ abB	$26.05 + 1.40 cA$	25.58 ± 1.06 cA	$23.34 + 0.91$ aA	$23.56 + 0.90$ a A	$23.86 + 1.04$ a A	$21.59 + 0.67$ bA	$14.52 + 0.35$ dB
	MT ₃	$22.13 + 1.88$ abB	22.15 ± 1.18 abC	$23.51 + 0.87$ aBC.	$21.54 + 0.64$ bB	$21.75 \pm 1.04 \text{ bB}$	$22.38 + 0.97$ abB	$21.77 + 0.30$ b A	18.44 ± 0.38 cC
	CK	$18.99 + 1.09$ a A	22.76 ± 0.83 bBC	22.82 ± 0.90 bC	21.14 ± 1.19 cdB	$22.18 + 0.79$ bcAB	$22.04 + 0.82$ bcdB	20.83 ± 0.63 dA	10.28 ± 0.27 eD
45 cm depth	MT ₁	24.23 ± 1.95 abA	25.45 ± 1.64 aAB	25.81 ± 0.93 aA	23.66 ± 0.52 bA	23.67 ± 0.49 bA	23.52 ± 0.61 bAB	21.76 ± 0.45 cAB	19.46 ± 0.41 dA
	MT ₂	$24.77 + 0.68$ a A	$26.86 + 0.72$ bA	27.35 ± 0.50 bB	23.81 ± 0.72 cA	23.72 ± 0.66 cA	23.79 ± 0.84 cA	22.06 ± 0.39 dA	$20.87 + 0.40$ eB
	MT3	$24.90 + 1.81$ a A	24.14 ± 1.35 abB	24.51 ± 1.01 abAC	23.08 ± 0.55 bcAB	23.78 ± 0.35 abcA	23.25 ± 0.62 bcAB	22.36 ± 0.35 cdA	21.19 ± 0.33 dB
	CK	21.62 ± 1.76 aB	23.69 ± 1.22 bB	23.57 ± 1.06 bC	22.24 ± 0.67 abB	22.57 ± 0.93 abB	22.36 ± 0.85 abB	21.25 ± 0.47 aB	17.31 ± 0.50 cC

Table 3. Monthly average soil water content (%) at different soil depths during the field experiment period for the mulch treatments.

Note: Lowercase letters represent significant differences ($p < 0.05$) among the different months for a given treatment; capital letters represent significant differences ($p < 0.05$) among different treatments in the same month.

The average soil water content of CK showed a monthly fluctuation and was strongly affected by rainfall and evaporation. The average soil water content of the mulch treatments showed a dynamic rising–stable–declining trend during the field experiment period. The soil water content of MT1, MT2, and MT3 was 8.13%, 17.23%, and 12.81% higher than that of CK, respectively, at a depth of 15 cm. At a depth of 30 cm, the soil water content of MT1, MT2 and MT3 was 6.27%, 12.48%, and 7.84% higher than that of CK, respectively. The soil water content of CK was 7.41%, 10.66%, and 7.20% lower than that of MT1, MT2, and MT3, respectively, at a soil depth of 45 cm. In this study, the average soil water content increased as the soil depth increased. The soil water content first increased and then decreased as the mulching thickness increased (from 0 to 15 cm), and the 10 cm mulching thickness treatment had a better water holding capacity. The SMS mulching not only had a positive effect by improving the soil water content, but it also effectively alleviated the soil moisture variation.

3.1.3. Soil Bulk Density and Porosity

The monthly dynamics of the soil bulk density and porosity under the different mulching thickness treatments are shown in Figure [2.](#page-7-0)

Figure 2. Variation in the soil physical properties among different treatment groups during the field experiment period: (**a**) soil bulk density and (**b**) soil porosity.

Figure [2a](#page-7-0) depicts the dynamic variation in the soil bulk density during these mulch treatments throughout the field experiment period. The lowest and highest bulk densities of these treatments appeared in June and December, respectively. All treatments had similar dynamic trends, that is, the density first decreased (from May to June), then stabilized (from July to October), and finally increased (in November and December). In a field experiment period of one year, the bulk density tended to increase as the mulching thickness increased. The MT3 treatment had the highest average bulk density (1.533 g cm⁻³), followed by MT2 $(1.529 \text{ g cm}^{-3})$, MT1 $(1.524 \text{ g cm}^{-3})$, and CK $(1.517 \text{ g cm}^{-3})$.

In this study, different mulch treatments reduced the soil porosity. The general dynamic variation in the soil porosity was in contrast to that of the bulk density, showing a trend of first rising and then decreasing (Figure [2b](#page-7-0)). There were no significant differences in soil porosity among these treatments throughout the field experiment period. The highest and lowest total porosities recorded for these treatment groups all appeared in June and December, respectively. The MT3 treatment was associated with the lowest average total porosity (43.9 %), followed by MT2 (44.1 %), MT1 (44.3 %), and CK (44.7 %).

3.1.4. Soil Nutrients

Figure [3a](#page-8-0) presents the monthly dynamics of the SOM content under different mulching thicknesses. The change trend for the SOM content in MT1, MT2, and MT3 was an initial increase and then a decrease. In the CK treatment, the trend was a continuous decrease at first, followed by a stable period. There were no significant differences in the SOM content among MT1, MT2, and MT3. However, the SOM concentration of the mulched plots was significantly higher than that of the plots, with no mulch from July to October. With an increase in the mulching thickness, the SOM concentration tended to increase. During the field experiment period, MT3 had the highest average SOM concentration (16.10 ± 1.29 g kg⁻¹), slightly higher than that of MT2 (15.89 ± 1.33 g kg⁻¹) and MT1 $(15.35 \pm 1.66 \text{ g kg}^{-1})$, while the lowest average SOM content, which was only 13.65 ± 1.85 g kg⁻¹, occurred in CK.

Figure 3. Variation in soil nutrient contents among different treatments during the field experiment period: (**a**) organic matter; (**b**) alkaline hydrolyzable nitrogen; (**c**) available potassium; (**d**) available phosphorus.

Figure [3b](#page-8-0) depicts the monthly dynamics of the AH-N content with different mulch treatments. The dynamic changes in the AH-N content in these treatments were similar; that is, there was an initial rapid decrease, followed by a slight increase and, finally, an obvious decrease. The highest AH-N content of each treatment appeared in May, and the lowest value appeared in December. There were significant differences in the AH-N content not only in different months, but also for different treatments. The average AH-N content in soil exposed to the MT3 treatment was 20.39%, 12.46%, and 4.77% higher than that for CK, MT1, and MT2, respectively.

The monthly dynamics of the AK content in soil exposed to the mulch treatments are shown in Figure [3c](#page-8-0). The change trend for the AK content in soil exposed to these treatments was similar among groups, with all of them showing a decline–rise–decline fluctuation during the study period. The AK contents of MT2 and MT3 were significantly higher than that of CK. MT3 had the highest average AK concentration (114.89 \pm 4.97 mg kg⁻¹), followed by MT2 $(113.04 \pm 4.77 \text{ mg kg}^{-1})$, MT1 $(105.37 \pm 4.58 \text{ mg kg}^{-1})$, and CK $(97.85 \pm 6.26 \text{ mg kg}^{-1})$.

Figure [3d](#page-8-0) illustrates the monthly dynamics of the AP content in soil exposed to different mulch treatments. The dynamic change trend in the AP content was similar to that of the AH-N and AK contents. There were significant differences in the AP content among these treatments. The AP contents of MT2 and MT3 were much higher than those of CK and MT1 from September to December. MT3 had the highest average AP concentration (18.32 \pm 0.62 mg kg⁻¹), which was 40.11%, 21.59%, and 0.85% higher than that of CK, MT1, and MT2, respectively.

3.1.5. Soil Enzymatic Activities

Soil enzymes are some of the most active components in various biochemical processes and nutrient cycles. They play an important role in the degradation of SOM and nutrient mineralization [\[26](#page-18-19)[,27\]](#page-18-20). The dynamics of three soil enzymatic activities under different mulch treatments are shown in Figure [4.](#page-9-0)

Figure 4. Variation in soil enzymatic activities among different treatment groups during the field experiment period: (**a**) urease activity; (**b**) invertase activity; (**c**) catalase activity.

The urease activity varied significantly across the sampling period and with different mulching thickness gradients (Figure [4a](#page-9-0)). In MT1, MT2, and MT3, the urease activity went through a rising period and was maintained at a high level before sharply decreasing over time. However, the urease activity decreased in CK after the rising period. The urease activity was higher in summer and lower in winter. The average urease activity of MT1, MT2, and MT3 increased by 12.56%, 18.04%, and 17.74%, respectively, compared with that of CK.

The invertase activity dynamics of the four treatments all showed a similar trend: a slight increase at first and then a rapid decrease (Figure [4b](#page-9-0)). The invertase activity of these four treatments was higher in summer, while lower invertase activity occurred in winter. MT3 had the highest average invertase activity (8.53 \pm 0.46 mg g^{-1.}24 h⁻¹), followed by MT2 (8.41 ± 0.37 mg g^{-1.}24 h⁻¹), MT1 (8.16 ± 0.36 mg g^{-1.}24 h⁻¹), and CK $(7.95 \pm 0.47 \text{ mg g}^{-1} \cdot 24 \text{ h}^{-1}).$

Figure [4c](#page-9-0) shows the monthly dynamics of catalase activity in soil exposed to these mulch treatments. The catalase activity showed a similar trend in these treatment groups with a decline–rise–decline fluctuation during the sampling period. The catalase activity differed significantly among sample sets from different months and with different mulching thickness gradients. The catalase activity of each treatment group was higher in autumn and lower in late summer. The average catalase activity of MT1, MT2, and MT3 was 8.35%, 13.24%, and 15.76% higher, respectively, than that of CK.

3.2. Effect of Mulching Thickness on Weed Suppression

The investigation of weeds was carried out in June, August, and October. In total, 18 weed species belonging to nine families were identified from these treatments (Table [4\)](#page-11-0). Weed biomass and control efficiency are presented in Table [4.](#page-11-0)

In June, there were 8, 5, 3, and 14 weed species in MT1, MT2, MT3, and CK, respectively. The dominant weed species in CK were *Cephalanoplos segetum* (Willd.) MB., *Conyza canadensis* (L.) Cronq., *Equisetum hyemale* L., *Ixeris sonchifolia* (Bunge) Hance, and *Taraxacum mongolicum* Hand.-Mazz. *Chenopodium album* L., *Eleusine indica* (L.) Gaertn., and *Ixeris sonchifolia* (Bunge) Hance were found in all treatment groups. In August, the number of weed species decreased in MT1 (7 species) and CK (10 species). The major weeds found in CK changed to *Amaranthus retroflexus* L., *Chenopodium album* L., *Eleusine indica* (L.) Gaertn., and *Equisetum hyemale* L.. *Amaranthus retroflexus* L., *Chenopodium album* L. and *Ixeris sonchifolia* (Bunge) Hance appeared in all treatments. In October, the number of weed species in MT1 (4 species), MT2 (4 species), MT3 (1 species), and CK (7 species) reduced obviously. *Ixeris sonchifolia* (Bunge) Hance, *Equisetum hyemale* L., and *Xanthium sibiricum* Patrin ex Widder became the dominant weed species. *Ixeris sonchifolia* (Bunge) Hance was found in all plots.

The highest weed dry biomass content was recorded in June for CK (1563.86 \pm 116.33 g m⁻²), whereas the lowest weed biomass was recorded in October (318.46 \pm 20.13 g m⁻²). In early summer (June), the weed biomass of mulched plots was much less than that of plots with no mulch, and the weed control efficiency ranged from 79.01% to 97.90%. In August, the weed biomass of MT1, MT2, and MT3 increased by 37.84%, 250.46%, and 687.52%, respectively, compared with that in June, while the weed control efficiencies decreased to 48.25%, 55.66%, and 70.43%, respectively. The weed biomass of the mulched plots decreased significantly in October, and the weed control efficiency improved greatly, ranging from 80.42% to 96.63%. The highest level of weed suppression, 70.43–97.90%, was obtained with MT3, followed by MT2 (55.66–92.93%), and the lowest level was found in the MT1 plots (48.25–80.42%).

With an increase of mulching thickness, the number of weed species decreased obviously and the weed control efficiency increased. SMS mulching was better able to control the growth of *Conyza canadensis* (L.) Cronq., *Cephalanoplos segetum* (Willd.) MB., *Humulus scandens* (Lour.) Merr., *Portulaca oleracea* L., and *Artemisia lavandulaefolia* DC.

Months	Mulch Treatments	Weed Species	The Number of Weed Species	Weed Biomass $(g m^{-2})$	Weed Control Efficiency (%)
June	MT1	Taraxacum mongolicum Hand.-Mazz., Amaranthus retroflexus L., Chenopodium album L., Ixeris sonchifolia (Bunge) Hance, Equisetum hyemale L., Conyza canadensis (L.) Cronq., Tagetes erecta L., Eleusine indica (L.) Gaertn.	8	328.32 ± 24.46	79.01
	MT ₂	Taraxacum mongolicum Hand.-Mazz., Chenopodium album L., Ixeris sonchifolia (Bunge) Hance, Eleusine indica (L.) Gaertn., Amaranthus retroflexus L.	5	110.63 ± 10.32	92.93
	MT3	Taraxacum mongolicum Hand.-Mazz., Ixeris sonchifolia (Bunge) Hance, Eleusine indica (L.) Gaertn.	3	32.84 ± 3.26	97.90
	CK	Taraxacum mongolicum Hand.-Mazz., Amaranthus retroflexus L., Chenopodium album L., Ixeris sonchifolia (Bunge) Hance, Conyza canadensis (L.) Cronq., Cirsium setosum (WillId.) MB., Tagetes erecta L., Eleusine indica (L.) Gaertn., Descurainia sophia (L.)Webb. ex Prantl, Equisetum hyemale L., Artemisia lavandulaefolia DC., Xanthium strumarium L., Mentha haplocalyx Linnaeus, Hemisteptia lyrata (Bunge) Bunge	14	1563.86 ± 116.33	
August	MT1	Amaranthus retroflexus L., Ixeris sonchifolia (Bunge) Hance, Eleusine indica (L.) Gaertn., Equisetum hyemale L., Tagetes erecta L., Mentha haplocalyx Linnaeus, Chenopodium album L.	7	452.57 ± 28.63	48.25
	MT ₂	Amaranthus retroflexus L., Eleusine indica (L.) Gaertn., Chenopodium album L., Mentha haplocalyx Linnaeus, Ixeris sonchifolia (Bunge) Hance	5	387.71 ± 20.54	55.66
	MT3	Amaranthus retroflexus L., Eleusine indica (L.) Gaertn., Chenopodium album L., Ixeris sonchifolia (Bunge) Hance	$\overline{4}$	258.62 ± 16.71	70.43
	${\rm C}{\rm K}$	Amaranthus retroflexus L., Ixeris sonchifolia (Bunge) Hance, Artemisia lavandulaefolia DC., Eleusine indica (L.) Gaertn., Mentha haplocalyx Linnaeus, Equisetum hyemale L., Chenopodium album L., Tagetes erecta L., Cirsium setosum (Willld.) MB., Humulus scandens L.	- 10	874.48 ± 60.56	
October	MT1	Ixeris sonchifolia (Bunge) Hance, Geranium carolinianum L., Equisetum hyemale L., Chenopodium album L.	$\overline{4}$	62.36 ± 8.27	80.42
	MT ₂	Geranium carolinianum L., Xanthium strumarium L., Chenopodium album L., Ixeris sonchifolia (Bunge) Hance		38.55 ± 4.62	87.89
	MT3	Ixeris sonchifolia (Bunge) Hance		10.73 ± 3.39	96.63
	$\mathrm{C}\mathrm{K}$	Equisetum hyemale L., Chenopodium album L., Ixeris sonchifolia (Bunge) Hance, Artemisia lavandulaefolia DC., Portulaca oleracea L., Cirsium setosum (Willld.) MB., Xanthium strumarium L.	-7	318.46 ± 20.13	

Table 4. Comparison of weed species exposed to different mulch treatments.

3.3. Effect of Mulching Thickness on Leaf Nutrients and Chlorophyll Content

The monthly dynamics of the leaf N content in soil exposed to different mulch treatments are shown in Figure [5a](#page-12-0). In CK, the leaf N content first decreased in June, then increased from July to August, and finally decreased sharply in September and October. The dynamic trend for the leaf N content in the other treatment groups was an initial increase followed by a decrease. The monthly variation in the leaf N content in all treatments was significant. There were also significant differences in the leaf N content among different treatments from June to October. The average leaf N content of MT3, MT2, and MT1 was 14.03%, 12.28%, and 8.39% higher, respectively, than that of CK.

Figure 5. Variation in leaf nutrient contents among different treatment groups during the growth period: (**a**) leaf N; (**b**) leaf P; (**c**) leaf K; (**d**) SPAD.

Figure [5b](#page-12-0) shows the monthly dynamics of the leaf P content with different mulch treatments. The dynamic trend for the leaf P content in MT1, MT2, and MT3 was similar; that is, an initial slight increase from May to July and then a decrease from August to October. In CK, the trend was a decrease from May to June, then a stable period from July to August, and finally a rapid decrease in September and October. The highest leaf P content appeared in July for MT1, MT2, and MT3, while for CK, it appeared in May, and the lowest leaf P content was in October for all groups. The average leaf P content of MT3, MT2, and MT1 was 16.99%, 15.78%, and 8.55% higher, respectively, than that of CK.

Figure [5c](#page-12-0) shows the monthly dynamics of the leaf K content in soil exposed to different mulch treatments. There were significant differences in the leaf K content not only in different months, but also with different treatments. The highest leaf K content appeared in July for MT1, MT2, MT3, and CK: 11.33 ± 1.14 , 11.87 ± 1.18 , 12.02 ± 1.25 , and 10.53 ± 1.14 1.20 mg g⁻¹, respectively. The minimum leaf K content occurred in October for MT1, MT2, MT3, and CK: 4.78 ± 0.49 , 5.03 ± 0.62 , 5.24 ± 0.54 , and 4.37 ± 0.59 mg $\rm g^{-1}$, respectively. The average leaf K content of the MT3, MT2, and MT1 treatments was 12.79%, 10.61%, and 4.45% higher, respectively, than that of CK.

Figure [5d](#page-12-0) presents the monthly dynamics of SPAD in soil exposed to these mulch treatments. There were significant differences in SPAD both in different months and with different treatments. The four treatments showed similar dynamic trends; that is, an initial increase from May to August and then a decrease from September to October. The average value of SPAD in MT3, MT2, and MT1 was 9.87%, 8.96%, and 3.24% higher, respectively, than that in CK.

3.4. Effect of Mulching Thickness on the Nut Yield and Quality Traits

The nut yield and quality traits varied in soil exposed to different mulch treatments. The values of these parameters are shown in Table [5.](#page-13-0)

Nut Traits	MT1	MT ₂	MT3	CK
Individual-plant	1027.06 ± 151.71	1045.26 ± 146.41	1019.50 ± 144.15	1001.40 ± 144.54
yield (g/tree)	A	A	A	A
L (mm)	19.92 ± 2.87 AB	20.47 ± 2.23 A	20.71 ± 3.07 A	19.22 ± 2.28 B
W (mm)	$17.96 + 2.34$ A	$18.18 + 2.11$ A	$17.78 + 1.93$ AB	$16.97 + 1.90 B$
T (mm)	16.77 ± 2.29 AB	16.97 ± 1.95 A	16.53 ± 2.08 AB	$16.05 \pm 1.70 B$
D (mm)	18.13 ± 2.15 A	18.45 ± 1.76 A	18.23 ± 2.13 A	$17.34 \pm 1.70 B$
V (cm ³)	3.25 ± 1.11 A	3.38 ± 0.93 A	3.29 ± 1.01 A	$2.81 \pm 0.82 B$
Ø	0.92 ± 0.08 A	0.91 ± 0.07 AB	0.89 ± 0.06 B	0.91 ± 0.06 AB
NSI	0.88 ± 0.12 A	0.86 ± 0.11 AB	$0.84 \pm 0.09 B$	0.87 ± 0.09 AB
NM(g)	2.40 ± 0.78 A	2.56 ± 0.73 A	2.45 ± 0.69 A	$2.08 \pm 0.63 B$
KM(g)	0.95 ± 0.29 A	1.00 ± 0.30 A	0.95 ± 0.29 A	$0.82 \pm 0.21 B$
PK(%)	40.28 ± 6.43 A	40.02 ± 9.93 A	38.96 ± 7.69 A	39.69 ± 5.43 A
CPC (%)	21.16 ± 1.15 A	22.18 ± 1.15 A	21.70 ± 1.00 A	20.89 ± 1.45 A
SSC $(\%)$	4.11 ± 0.18 A	4.23 ± 0.15 A	4.18 ± 0.11 A	4.05 ± 0.24 A
CFC (%)	60.29 ± 1.68 AB	61.87 ± 1.41 A	60.63 ± 1.25 AB	59.82 \pm 1.89 B

Table 5. Comparison of weed species exposed to different mulch treatments.

Note: Capital letters represent significant differences (*p <* 0.05) among treatments.

The average values of *D*, *V*, *NM*, and *KM* were significantly lower in CK in soil exposed to mulch treatments. The highest average *L* value appeared in MT3, and this was 7.75% higher than that of CK. The highest average *D*, average *V*, average *NM*, and average *KM* values all appeared in MT2. These were 6.40%, 20.28%, 23.08%, and 21.95% higher than the values in CK, respectively. Although there were significant differences in *NM* and *KM* between soil exposed or not to mulch treatments, the difference in *PK* was not significant. MT1 treatment had the highest *PK*, followed by MT2, CK, and MT3. Sphericity and shape index are the main indicators of nut shape, and these variables are determined by nut dimensions. According to Milošević and Milošević [\[28\]](#page-18-21), an *NSI* value of 0.84 indicates that the hazelnut shape is conical, and an *NSI* value above 0.86 indicates that the hazelnut shape is globular. In this study, the average nut sphericity and nut shape index values were highest in MT1, which means that the nuts in MT1 were rounder than in other treatments. There were no significant differences in the plant yield, *CPC*, or *SSC* among these treatment groups. The individual plant yield, *CPC*, and *SSC* were the highest in MT2, 4.38%, 6.18%, and 4.44% higher than the values in CK, respectively. The *CFC* of MT2 was significantly higher than that of CK.

4. Discussion

4.1. Effect of Mulching on Soil Physico-Chemical and Biological Properties

Soil temperature is a key environmental factor that has an impact on crop growth and development by regulating water absorption and nutrient uptake in root zones [\[29\]](#page-18-22). Mulches alter the soil temperature, which affects the thermal regime of a soil [\[30\]](#page-18-23). However, the effect of mulches on the soil temperature varies depending on the capacity of the mulching materials to reflect and transmit solar energy [\[31\]](#page-18-24). Organic mulches can reduce the maximum soil temperature, but increase the minimum soil temperature, while they

significantly reduce the soil temperature [\[8\]](#page-18-1). The results of a mulching study conducted on the Loess Plateau showed that daily mean soil temperatures in soil treated with mulch at a depth of 10 cm were lower in warmer periods by $0-4$ °C and higher in colder periods by 0–2 ◦C compared with non-mulched soil [\[32\]](#page-18-25). Similarly, in our study, SMS mulching measures can efficiently regulate the soil temperature by reducing it in warm weather and increasing it in cold weather. However, the regulation ability of SMS varies with the mulching thickness and soil depth. SMS forms a temperature isolation layer on the soil surface owing to its high reflectivity and low thermal diffusivity, which can effectively reduce fluctuations in the soil temperature.

Soil moisture is also considered to be an important factor affecting plant growth and development, and even a small change in soil water storage could affect crop productivity [\[33\]](#page-18-26). A large number of previous studies have confirmed that mulching can effectively conserve soil water by reducing soil evaporation [\[29\]](#page-18-22). Mulching has a positive impact on the soil water content that depends on the type of mulching materials and thickness of mulching used [\[34\]](#page-18-27). Some studies have shown that the mulch thickness is more important than the texture of the mulch for evaporation suppression, and thicker mulch prevents soil water evaporation more effectively. Moreover, studies have shown that mineral or organic mulches must have a critical thickness to effectively reduce soil evaporation, and the crop yield and water use efficiency can be reduced if the mulching thickness is excessive [\[35\]](#page-19-0). McMillen [\[36\]](#page-19-1) compared the soil water retention capacity of three organic mulching materials with different mulching thickness. The results showed that treatment with 10 cm thick mulch increased the soil moisture by over 10% compared with bare soil, while 15 cm thick mulching treatment did not significantly reduce soil evaporation further. The results of this study are consistent with the previous research conclusion, that is, 10 cm thick mulch had a strong water holding capacity throughout the field experiment period. The soil moisture content of MT2 was significantly higher than that of CK in all three soil depths during the rainy season. Evaporation and drainage were the main factors that reduced the soil moisture content in the soil profile of 0–30 cm in the rainy season, and mulching can conserve the soil moisture for longer periods than bare soil [\[37\]](#page-19-2). With an increase in the soil depth, the difference in soil moisture among different treatments decreased.

The effect of mulching on the soil bulk density varies depending on the soil properties, mulch material used, climatic conditions, and land use. Mulching increases the soil bulk density when conventional tillage is used, but can also reduce it by adding organic matter to the soil. Whether there is an increase or decrease in the soil bulk density depends on the specific situation [\[8\]](#page-18-1). Massaccesi et al. [\[38\]](#page-19-3) supports the idea that the soil bulk density tends to increase with the application of organic mulch combined with no-tillage practices. In 2015, Kader et al. showed that the soil bulk density following straw mulching was lower than at a depth of 0–30 cm compared with that of bare soil; however, the reverse trend was found in 2016 [\[37\]](#page-19-2). Ni et al. [\[39\]](#page-19-4) reported non-significant results for the effects of mulching on the soil bulk density at a depth of 0–10 cm. The results of this study demonstrate that the soil bulk density tended to increase as the mulching thickness increased, but there was no significant difference between mulching and non-mulch treatments. Soil porosity is influenced by the agronomic practices implemented, and it decreases with the soil depth [\[40](#page-19-5)[,41\]](#page-19-6). Nzeyimana et al. [\[42\]](#page-19-7) conducted a two-year mulching experiment to quantify the effects of different organic mulches on soil properties, and the results showed that the application of mulch had a positive impact on the soil porosity. However, the effects of the interactions between the type of mulch and type of soil were site-specific. In our study, there was no significant difference in soil porosity among the different mulch treatments. SMS usually required a long time to completely mineralize [\[43\]](#page-19-8), thus it may take a longer time to improve soil structure when SMS is used as a mulching material.

A significant loss in soil nutrients occurs within 1–2 years of plastic mulching, probably owing to temperature-induced accelerated biodegradation [\[44\]](#page-19-9), which is closely linked and entangled with an increasing C/N metabolism [\[45\]](#page-19-10). However, when the soil carbon pool is maintained by organic matter input, the SOM content remains stable [\[8\]](#page-18-1). A study by Pal and Mahajan [\[46\]](#page-19-11) revealed that higher soil nutrient contents can be achieved through the application of an optimal level of organic mulch (pine-needle mulch). Similarly, our study showed that the use of SMS mulch can significantly increase the soil nutrient content compared with soil not treated with mulch. Although the highest average values of SOM, AH-N, AK, and AP were observed in MT3, there were no significant difference in soil nutrient contents between MT2 and MT3. This may be owing to the fact that nutrient release by mineralization requires an appropriate quantity of SMS mulch. The monthly dynamics of soil nutrients are strongly affected by both the mineralization rate and plant uptake and utilization. In this experiment, higher contents of AK and AP were found in soil exposed to mulch treatment. Similar results were also reported in straw mulch by Xia [\[47\]](#page-19-12) and Zhao [\[48\]](#page-19-13). The high AK content found in MT2 and MT3 may be due to the fact that K exists in SMS (plant tissue) in the free ionic form and is easily leached into the soil. However, the high content of AP in MT2 and MT3 may be related to the accumulation of P in the soil. The demand for P decreases in the nut maturity stage, while soil microorganisms accelerate the mineralization process and release a large amount of AP into the soil with mulch treatments. Microbial communities are closely associated with soil nutrients and strongly influence the soil quality.

Soil enzymes mainly come from the process of microbial metabolism and are closely related to the composition, abundance, and activity of soil microbial communities. Soil enzymes are also regulated by the demand for substrates as well as by the substrate availability [\[49\]](#page-19-14). Qu and Feng [\[50\]](#page-19-15) found that straw mulching increased the activity levels of soil urease, invertase, and acid phosphatase. Similarly, this study showed that SMS mulch can effectively increase soil enzyme activities compared with soil not treated with mulch. The increase in soil enzyme activities following SMS mulching treatment might owing to the increased carbon sources and the improved soil quality. Soil enzymatic activity levels followed the order MT3 \approx MT2 > MT1 > CK. Sufficient, but not excessive application of SMS was beneficial for the carbon source utilization of fast-growing microorganisms [\[51\]](#page-19-16). In this study, the use of a moderate mulching thickness may be a better way to improve soil enzymatic activity from a cost perspective.

4.2. Effect of Mulching on Weed Suppression

Weed control is one of the biggest challenges in the farming system [\[52\]](#page-19-17). As the growth of weeds depends on the availability of light and because light can only penetrate soil to a depth of up to a few centimeters, a certain thickness (>5 cm) of organic mulch is considered to effectively suppress weed germination through the exclusion of light or by acting as a physical barrier [\[53\]](#page-19-18). In our study, SMS mulching decreased light penetration and inhibited the germination of weed seeds in the early stage. At this stage, the weed control efficiency was as high as 79.01–97.90% for mulch treatment. With the arrival of the rainy season, alien weed seeds emerged easily from thin or damaged parts of the mulch layers, resulting in a weakened inhibition effect of mulching on weeds. The weed control efficiency decreased to 48.25–70.43%. This can largely be explained by the positive effect of soil moisture on weed emergence. In addition, SMS decomposition resulted in a decrease in soil cover during the growing season and may also have positively affected weed emergence [\[54\]](#page-19-19). With a continuous decrease in temperature, some annual weeds completed their life cycle, resulting in a significant decrease in weed biomass in October. Additionally, the weed control efficiency increased to 80.42–96.63% in the nut maturity stage.

Previous studies indicated that a certain amount of organic mulch is needed for weed suppression [\[17,](#page-18-10)[18\]](#page-18-11). Thankamani et al. [\[18\]](#page-18-11) suggested that the application of 6 t·ha⁻¹ of paddy straw along with 7.5 t·ha⁻¹ of green leaf mulch at 45 and 90 days after planting could led to a higher weed control efficiency and a greater yield, as well as economic return. Taak et al. [\[55\]](#page-19-20) evaluated the effects of mulching and herbicide treatments on weed control and found that straw mulching treatment (15 t \cdot ha⁻¹) not only effectively controls weeds, but is also associated with the highest crop yield. Weed emergence and biomass decreased with increasing amounts of organic mulch [\[54\]](#page-19-19). The results of our study also support the

conclusion that SMS with a thickness of more than 10 cm is associated with a better weed control efficiency.

4.3. Effect of Mulching on the Leaf Nutrient and Chlorophyll Contents

Leaf tissue is most commonly used for nutrient analysis because it is the one of the most active sites of plant metabolism. The leaf nutrient analysis provides an indication of plant nutritional status that can reflect plant health. It is necessary to determine whether or not a plant is well nourished, as having an adequate nutritional state is a determining factor in the productivity/yield of any plant [\[21\]](#page-18-14). In our study, all SMS mulch treatments effectively increased the leaf nutrient (N, P, and K) and chlorophyll contents, while the leaf N, P, K, and chlorophyll contents of MT2 and MT3 were significantly improved. In the short-term experiment of one year, the leaf nutrient content decreased twice. The first decrease occurred in June when the nutrients of newborn leaves were easily leached out by rainfall with the arrival of the rainy season. The second decline occurred from September to October when the leaf N, P, and K contents began to be transferred to other vigorous organs (such as fruit), and the leaves entered the senescence stage. The chlorophyll content of leaves showed a trend involving an initial increase, following by a decrease. With leaf senescence, the chlorophyll content also reduced. Although studies about the effect of organic mulching on leaf nutrients are still insufficient, some existing literature supports the idea that organic mulching can improve the leaf nutrient level. Pal and Mahajan [\[46\]](#page-19-11) found a significantly higher leaf K content in *Stevia rebaudiana* Bertoni following the application of pine-needle mulching at a level of 15 t·ha⁻¹ compared with other treatments. Namaghi et al. [\[56\]](#page-19-21) also found higher levels of P and Mg in barley-straw-mulched pistachio leaves. Kader et al. [\[37\]](#page-19-2) reported that straw mulching significantly improved the leaf chlorophyll content of soybean compared with bare soil. The higher leaf nutrient and chlorophyll contents may have been due to the suitable soil microclimate created by organic mulching, which promoted root growth, and thus increased the absorption of soil nutrients [\[37\]](#page-19-2). In addition, the nutrients of organic matter were gradually released into the soil after organic mulching and significantly increased the available soil nutrients [\[56\]](#page-19-21).

4.4. Effect of Mulching on Fruit Yield and Quality

Although previous studies have reported the effect of organic mulching on the fruit yield and quality, there has been no consistent conclusion in this regard owing to differences in mulching materials, mulching practices, climatic conditions, research objects, and so on. Some studies have shown that organic mulching can markedly increase the fruit yield with no significant effect on fruit quality [\[57\]](#page-19-22). Some studies have observed that organic mulching has a significant positive effect on both fruit yield and quality. Ye et al. [\[58\]](#page-19-23) showed that mulching significantly increased the yield and quality of *Camellia oleifera*, which was consistent with previous results for apple trees [\[59\]](#page-19-24), peach trees [\[57\]](#page-19-22), jujube trees [\[60\]](#page-19-25), and pistachio trees [\[56\]](#page-19-21). The present study suggests that all mulch treatments were associated with an increasing trend in nut yield and quality compared with the nonmulching treatment. All mulch treatments were associated with the production of nuts with significantly higher geometric mean diameters and volumes than those produced with CK. A higher variability in growth of individual fruits is mainly due to the fruit-tofruit competition for nutrients and resources [\[61\]](#page-19-26). SMS mulch increased the availability of soil nutrients; therefore, the nuts produced in plots treated with mulch were larger. The larger nut size correspondingly increased the nut mass and kernel mass. These two important parameters of nuts were significantly improved under mulch treatments. The nut sphericity and nut shape index were significantly lower in MT3 than in MT1, which may be due to the rapid growth in the longitudinal diameter when the nutrient supply was relatively sufficient. Variations in fruit size and shape under different mulch treatments were also reported by Ye et al. [\[58\]](#page-19-23). Although our research revealed that short-term SMS mulching had no obvious advantage in terms of increasing the individual plant yield, *CPC*, or *SSC*, MT2 did effectively increase the *CFC* of nuts. The non-significant increase in nut

yield may have been due to the following factors: (1) organic mulch could improve soil hydro-thermal regimes [\[62\]](#page-20-0); (2) organic mulch could increase the soil nutrient content by organic matter decomposition; (3) organic mulch could prevent excessive weed growth; and (4) organic mulch could improve the nutrient status and photosynthetic capacity of leaves [\[56\]](#page-19-21). Variations in the *CPC*, *SSC*, and *CFC* of nuts among mulch treatments indicated that the formation of fruit quality is a complex physiological and biochemical process. This process is closely related to the transport, allocation, conversion, and metabolism of carbohydrates, which is strongly influenced not only by biological factors, but also by environmental factors [\[63\]](#page-20-1). The soil moisture status is often considered to be a critical factor affecting oil synthesis. When a water deficit occurs, it will lead to a decrease in the crude fat content of fruit [\[64\]](#page-20-2).

5. Conclusions

SMS mulching was found to influence soil fertility, weed control, leaf quality, and nut traits. The effects varied depending on the mulching thickness used. Although SMS mulching had no significant effect on soil structure in the short-term experiment, it significantly improved the topsoil nutrient availability. SMS mulching also decreased the number of weed species and weed biomass obviously. Short-term SMS mulching had no obvious advantages in terms of increasing the individual plant yield, *CPC*, or *SSC*, but it increased the nut size, nut and kernel mass, as well as *CFC* to a certain extent. The results showed that SMS with a 10 cm thickness could be used for mulching practices.

While SMS mulching is known to improve soil fertility and nut quality in the short term, its long-term effects require further research. In particular, more research is needed to understand the effects of SMS mulching on photosynthetic efficiency and water consumption, which are fundamental factors in the shaping of the characteristics of nuts over an extended period of time.

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References

- 1. Food and Agriculture Organization of the United Nation, Agricultural Data. Available online: [http://www.fao.org/faostat/en/](http://www.fao.org/faostat/en/#data/QC) [#data/QC](http://www.fao.org/faostat/en/#data/QC) (accessed on 17 January 2019).
- 2. Alasalvar, C.; Shahidi, F.; Liyanapathirana, C.M.; Ohshima, T. Turkish Tombul Hazelnut (*Corylus avellana* L.). 1. Compositional Characteristics. *J. Agric. Food Chem.* **2003**, *51*, 3790–3796. [\[CrossRef\]](http://doi.org/10.1021/jf0212385) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/12797745)
- 3. Felbinger, C.; Kutzsche, F.; Mnkediek, S.; Fischer, M.J.F.C. Genetic profiling: Differentiation and identification of hazelnut cultivars (*Corylus avellana* L.) using RAPD-PCR. *Food Control.* **2019**, *107*, 106791. [\[CrossRef\]](http://doi.org/10.1016/j.foodcont.2019.106791)
- 4. Jabran, K. Mulches for Enhancing Biological Activities in Soil. In *Role of Mulching in Pest Management and Agricultural Sustainability*; Ömer Halisdemir University Press: Niğde, Turkey, 2019; pp. 41-46. ISBN 978-3-030-22300-7.
- 5. Gao, H.; Yan, C.; Liu, Q.; Ding, W.; Chen, B.; Li, Z. Effects of plastic mulching and plastic residue on agricultural production: A meta-analysis. *Sci. Total Environ.* **2019**, *651*, 484–492. [\[CrossRef\]](http://doi.org/10.1016/j.scitotenv.2018.09.105) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30243168)
- 6. Qi, Y.; Yang, X.; Pelaez, A.M.; Huerta Lwanga, E.; Beriot, N.; Gertsen, H.; Garbeva, P.; Geissen, V. Macro- and micro-plastics in soil-plant system: Effects of plastic mulch flm residues on wheat (*Triticum aestivum*) growth. *Sci. Total Environ.* **2018**, *645*, 1048–1056. [\[CrossRef\]](http://doi.org/10.1016/j.scitotenv.2018.07.229)
- 7. López, R.; Burgos, P.; Hermoso, J.M.; Hormaza, J.I.; González-Fernández, J. Long term changes in soil properties and enzyme activities after almond shell mulching in avocado organic production. *Soil Tillage Res.* **2014**, *143*, 155–163. [\[CrossRef\]](http://doi.org/10.1016/j.still.2014.06.004)
- 8. Kader, M.A.; Senge, M.; Mojid, M.A.; Ito, K. Recent advances in mulching materials and methods for modifying soil environment. *Soil Tillage Res.* **2017**, *168*, 155–166. [\[CrossRef\]](http://doi.org/10.1016/j.still.2017.01.001)
- 9. Royse, D.J.; Baars, J.; Tan, Q. *Current Overview of Mushroom Production in the World: Technology and Applications*; Wiley-Blackwell Press: Hoboken, NJ, USA, 2017; pp. 5–13. ISBN 978-1-119-14941-5.
- 10. Hanafi, F.H.M.; Rezania, S.; Taib, S.M.; Din, M.F.M.; Yamauchi, M.; Sakamoto, M.; Hara, H.; Park, J.; Ebrahimi, S.S. Environmentally sustainable applications of agro-based spent mushroom substrate (SMS): An overview. *J. Mater. Cycles Waste Manag.* **2018**, *20*, 1383–1396. [\[CrossRef\]](http://doi.org/10.1007/s10163-018-0739-0)
- 11. Nakatsuka, H.; Oda, M.; Hayashi, Y.; Tamura, K. Effects of fresh spent mushroom substrate of Pleurotus ostreatus on soil micromorphology in Brazil. *Geoderma* **2016**, *269*, 54–60. [\[CrossRef\]](http://doi.org/10.1016/j.geoderma.2016.01.023)
- 12. Li, Q.; Wang, M.; Fu, Q.; Li, T.; Liu, D.; Hou, R.; Li, H.; Cui, S.; Ji, Y. Short-term influence of biochar on soil temperature, liquid moisture content and soybean growth in a seasonal frozen soil area. *J. Environ. Manag.* **2020**, *266*, 110609. [\[CrossRef\]](http://doi.org/10.1016/j.jenvman.2020.110609)
- 13. Shan, L.; Qi-Quan, L.I.; Chang-Quan, W.; Bing, L.I.; Xue-Song, G.; Yi-Ding, L.I.; De-yong, W. Spatial variability of soil bulk density and its controlling factors in an agricultural intensive area of Chengdu Plain, Southwest China. *J. Integr. Agric.* **2019**, *18*, 290–300. [\[CrossRef\]](http://doi.org/10.1016/S2095-3119(18)61930-6)
- 14. Khan, S.; Mulvaney, R.; Hoeft, R. A Simple Soil Test for Detecting Sites that are Nonresponsive to Nitrogen Fertilization. *Soil Sci. Soc. Am. J.* **2001**, *65*, 1751–1760. [\[CrossRef\]](http://doi.org/10.2136/sssaj2001.1751)
- 15. Carson, P.L. Recommended potassium test. In *Recommended Chemical Soil Test Procedures for the North Central Region*; Dahnke, W.C., Ed.; North Dakota Agricultural Experiment Station: Fargo, ND, USA, 1980; pp. 17–18.
- 16. Ge, Y.; Wang, Q.; Wang, L.; Liu, W.; Liu, X.; Huang, Y.; Christie, P. Response of soil enzymes and microbial communities to root extracts of the alien *Alternanthera philoxeroides*. *Arch. Agron. Soil Sci.* **2018**, *64*, 708–717. [\[CrossRef\]](http://doi.org/10.1080/03650340.2017.1373186)
- 17. Singh, R.; Brar, S.; Walia, U.S. Comparative efficiency of herbicides for weed control in Chickpea (*Cicer arietinum* L.). *Crop Res.* **2000**, *19*, 1–5.
- 18. Thankamani, C.K.; Kandiannan, K.; Hamza, S.; Saji, K.V. Effect of mulches on weed suppression and yield of ginger (*Zingiber officinale* Roscoe). *Sci. Hortic.* **2016**, *207*, 125–130. [\[CrossRef\]](http://doi.org/10.1016/j.scienta.2016.05.010)
- 19. Dong, Q.; Dang, T.; Guo, S.; Hao, M. Effect of different mulching measures on nitrate nitrogen leaching in spring maize planting system in south of Loess Plateau. *Agric. Water Manag.* **2019**, *213*, 654–658. [\[CrossRef\]](http://doi.org/10.1016/j.agwat.2018.09.044)
- 20. Debnath, A.; Barrow, N.J.; Ghosh, D.; Malakar, H. Diagnosing p status and p requirement of tea (*camellia sinensis* L.) by leaf and soil analysis. *Plant Soil* **2011**, *341*, 309–319. [\[CrossRef\]](http://doi.org/10.1007/s11104-010-0645-2)
- 21. Guimarães, Z.T.; Dos Santos, V.A.; Nogueira, W.L.; de Almeida Martins, N.O.; Ferreira, M.J. Leaf traits explaining the growth of tree species planted in a Central Amazonian disturbed area. *For. Ecol. Manag.* **2018**, *430*, 618–628. [\[CrossRef\]](http://doi.org/10.1016/j.foreco.2018.08.048)
- 22. Martinez, F.; Oliveira, J.A.; Calvete, E.O.; Palencia, P. Influence of growth medium on yield, quality indexes and SPAD values in strawberry plants. *Sci. Hortic.* **2017**, *217*, 17–27. [\[CrossRef\]](http://doi.org/10.1016/j.scienta.2017.01.024)
- 23. Hosseinpour, A.; Seifi, E.; Javadi, D.; Ramezanpour, S.S.; Molnar, T.J. Nut and kernel characteristics of twelve hazelnut cultivars grown in Iran. *Sci. Hortic.* **2013**, *150*, 410–413. [\[CrossRef\]](http://doi.org/10.1016/j.scienta.2012.11.028)
- 24. Wang, W.; Jung, J.; McGorrin, R.J.; Traber, M.G.; Leonard, S.W.; Cherian, G.; Zhao, Y. Investigation of drying conditions on bioactive compounds, lipid oxidation, and enzyme activity of Oregon hazelnuts (*Corylus avellana* L.). *Food Sci. Technol.* **2018**, *90*, 526–534. [\[CrossRef\]](http://doi.org/10.1016/j.lwt.2018.01.002)
- 25. Luo, D.; Shi, Y.J.; Song, F.H.; Zheng, W.U.; Ablat, M.; Cheng, L.I. Spatial Distribution Characteristics of Fine roots in Monoculture System of *Corylus heterophylla* × *Corylus avellana*. *For. Res.* **2019**, *32*, 81–89. [\[CrossRef\]](http://doi.org/10.13275/j.cnki.lykxyj.2019.01.011)
- 26. Burns, R.G.; DeForest, J.L.; Marxsen, J.; Sinsabaugh, R.L.; Stromberger, M.E.; Wallenstein, M.D.; Weintraub, M.N.; Zoppini, A. Soil enzymes in a changing environment: Current knowledge and future directions. *Soil Biol. Biochem.* **2013**, *58*, 216–234. [\[CrossRef\]](http://doi.org/10.1016/j.soilbio.2012.11.009)
- 27. Veres, Z.; Kotroczó, Z.; Fekete, I.; Tóth, J.A.; Lajtha, K.; Townsend, K.; Tóthmérész, B. Soil extracellular enzyme activities are sensitive indicators of detrital inputs and carbon availability. *Appl. Soil Ecol.* **2015**, *92*, 18–23. [\[CrossRef\]](http://doi.org/10.1016/j.apsoil.2015.03.006)
- 28. Milošević, T.; Milošević, N. Determination of size and shape features of hazelnuts using multivariate analysis. Acta Sci. Pol. *Hortorum Cultus* **2017**, *16*, 49–61. [\[CrossRef\]](http://doi.org/10.24326/asphc.2017.5.6)
- 29. Liao, Y.; Cao, H.X.; Xue, W.K.; Liu, X. Effects of the combination of mulching and deficit irrigation on the soil water and heat, growth and productivity of apples. Agric. *Water Manag.* **2021**, *243*, 106482. [\[CrossRef\]](http://doi.org/10.1016/j.agwat.2020.106482)
- 30. Pramanik, P.; Bandyopadhyay, K.K.; Bhaduri, D.; Bhattacharyya, R.; Aggarwal, P. Effect of mulch on soil thermal regimes—A review. *Int. J. Agric. Environ. Biotechnol.* **2015**, *8*, 645. [\[CrossRef\]](http://doi.org/10.5958/2230-732X.2015.00072.8)
- 31. Lamont, W.J. Plastics: Modifying the microclimate for the production of vegetable crops. *Horttechnology* **2005**, *15*, 477–481. [\[CrossRef\]](http://doi.org/10.21273/HORTTECH.15.3.0477)
- 32. Zhang, S.; Lovdahl, L.; Grip, H.; Tong, Y.; Yang, X.; Wang, Q. Effects of mulching and catch cropping on soil temperature, soil moisture and wheat yield on the Loess Plateau of China. *Soil Tillage Res.* **2009**, *102*, 78–86. [\[CrossRef\]](http://doi.org/10.1016/j.still.2008.07.019)
- 33. Liu, Y.; Yang, S.; Li, S.; Chen, X.; Chen, F. Growth and development of maize (*Zea mays* L.) in response to different field water management practices: Resource capture and use efficiency. *Agric. For. Meteorol.* **2010**, *150*, 606–613. [\[CrossRef\]](http://doi.org/10.1016/j.agrformet.2010.02.003)
- 34. Thakur, M.; Kumar, R. Mulching: Boosting crop productivity and improving soil environment in herbal plants. *J. Appl. Res. Med. Aromat. Plants* **2021**, *20*, 100287. [\[CrossRef\]](http://doi.org/10.1016/j.jarmap.2020.100287)
- 35. Jones, H.; Black, T.A.; Jassal, R.S.; Nesic, Z.; Johnson, M.S.; Smukler, S. Characterization of shortwave and longwave properties of several plastic film mulches and their impact on the surface energy balance and soil temperature. *Solar Energy* **2021**, *214*, 457–470. [\[CrossRef\]](http://doi.org/10.1016/j.solener.2020.11.058)
- 36. Mcmillen, M. The Effect of Mulch Type and Thickness on the Soil Surface Evaporation Rate. Available online: [https://](https://digitalcommons.calpoly.edu) digitalcommons.calpoly.edu (accessed on 1 June 2013).
- 37. Kader, M.A.; Nakamura, K.; Senge, M.; Mojid, M.A.; Kawashima, S. Soil hydro-thermal regimes and water use efficiency of rain-fed soybean (*Glycine max*) as affected by organic mulches. *Agric. Water Manag.* **2019**, *223*, 105707. [\[CrossRef\]](http://doi.org/10.1016/j.agwat.2019.105707)
- 38. Massaccesi, L.; Rondoni, G.; Tosti, G.; Conti, E.; Agnelli, A. Soil functions are affected by transition from conventional to organic mulch-based cropping system. *Appl. Soil Ecol.* **2020**, *153*, 103639. [\[CrossRef\]](http://doi.org/10.1016/j.apsoil.2020.103639)
- 39. Ni, X.; Song, W.; Zhang, H.; Yang, X.; Wang, L. Effects of Mulching on Soil Properties and Growth of Tea Olive (*Osmanthus fragrans*). *PLoS ONE* **2016**, *11*, e0158228. [\[CrossRef\]](http://doi.org/10.1371/journal.pone.0158228)
- 40. Al-Shammary, A.A.G.; Kouzani, A.; Gyasi-Agyei, Y.; Gates, W.; Rodrigo-Comino, J. Effects of solarisation on soil thermal-physical properties under different soil treatments: A review. *Geoderma* **2020**, *363*, 114137. [\[CrossRef\]](http://doi.org/10.1016/j.geoderma.2019.114137)
- 41. Pires, L.F.; Borges, J.A.; Rosa, J.A.; Cooper, M.; Heck, R.J.; Passoni, S.; Roque, W.L. Soil structure changes induced by tillage systems. *Soil Tillage Res.* **2017**, *165*, 66–79. [\[CrossRef\]](http://doi.org/10.1016/j.still.2016.07.010)
- 42. Nzeyimana, I.; Hartemink, A.E.; Ritsema, C.; Stroosnijder, L.; Lwanga, E.H.; Geissen, V. Mulching as a strategy to improve soil properties and reduce soil erodibility in coffee farming systems of Rwanda. *CATENA* **2017**, *149*, 43–51. [\[CrossRef\]](http://doi.org/10.1016/j.catena.2016.08.034)
- 43. Zanella, A.; Ponge, J.F.; Guercini, S.; Rumor, C.; Nold, F.; Sambo, P.; Gobbi, V.; Schimmer, C.; Chaabane, C.; Mouchard, M.L. Humusica 2, article 16: Techno humus systems and recycling of waste. *Appl. Soil Ecol.* **2017**, *122*, 220–236. [\[CrossRef\]](http://doi.org/10.1016/j.apsoil.2017.09.037)
- 44. Moreno, M.M.; Moreno, A. Effect of different biodegradable and polyethylene mulches on soil properties and production in a tomato crop. *Sci. Hortic.* **2008**, *116*, 256–263. [\[CrossRef\]](http://doi.org/10.1016/j.scienta.2008.01.007)
- 45. Steinmetz, Z.; Wollmann, C.; Schaefer, M.; Buchmann, C.; David, J.; Troeger, J.; Munoz, K.; Froer, O.; Schaumann, G.E. Plastic mulching in agriculture. Trading short-term agronomic benefits for long-term soil degradation? *Sci. Total Environ.* **2016**, *550*, 690–705. [\[CrossRef\]](http://doi.org/10.1016/j.scitotenv.2016.01.153)
- 46. Pal, P.K.; Mahajan, M. Tillage system and organic mulch influence leaf biomass, steviol glycoside yield and soil health under sub-temperate conditions. *Ind. Crops Prod.* **2017**, *104*, 33–44. [\[CrossRef\]](http://doi.org/10.1016/j.indcrop.2017.04.012)
- 47. Study on the Soil Ecological Effects and Physiological Response in Different Groundcover Pear Tree. Available online: [https:](https://kns.cnki.net/kns8) [//kns.cnki.net/kns8](https://kns.cnki.net/kns8) (accessed on 1 January 2013).
- 48. Study on Soil Ecological Effect of Different Mulching Measures in Tea Garden. Available online: <https://kns.cnki.net/kns8> (accessed on 12 June 2006).
- 49. Olander, L.P.; Vitousek, P.M. Regulation of soil phosphatase and chitinase activityby N and P availability. *Biogeochemistry* **2000**, *49*, 175–190. [\[CrossRef\]](http://doi.org/10.1023/A:1006316117817)
- 50. Qu, Y.; Feng, B.L. Straw mulching improved yield of field buckwheat (*Fagopyrum*) by increasing water-temperature use and soil carbon in rain-fed farmland. *Acta Ecol. Sin.* **2020**, *3*, 10–12. [\[CrossRef\]](http://doi.org/10.1016/j.chnaes.2020.11.008)
- 51. Li, F.; Kong, Q.; Zhang, Q.; Wang, H.; Luo, T. Spent mushroom substrates affect soil humus composition, microbial biomass and functional diversity in paddy fields. *Appl. Soil Ecol.* **2020**, *149*, 103489. [\[CrossRef\]](http://doi.org/10.1016/j.apsoil.2019.103489)
- 52. Farooq, M.; Flower, K.C.; Jabran, K.; Wahid, A.; Siddique, K. Crop yield and weed management in rainfed conservation agriculture. *Soil Tillage Res.* **2011**, *117*, 172–183. [\[CrossRef\]](http://doi.org/10.1016/j.still.2011.10.001)
- 53. Marble, S.C.; Koeser, A.K.; Hasing, G. A Review of Weed Control Practices in Landscape Planting Beds: Part II—Chemical Weed Control Methods. *Hortscience* **2015**, *50*, 857–862. [\[CrossRef\]](http://doi.org/10.21273/HORTSCI.50.6.857)
- 54. Ranaivoson, L.; Naudin, K.; Ripoche, A.; Rabeharisoa, L.; Corbeels, M. Is mulching an efficient way to control weeds? Effects of type and amount of crop residue in rainfed rice based cropping systems in Madagascar. *Field Crops Res.* **2018**, *217*, 20–31. [\[CrossRef\]](http://doi.org/10.1016/j.fcr.2017.11.027)
- 55. Taak, P.; Koul, B.; Chopra, M.; Sharma, K. Comparative Assessment of Mulching and Herbicide Treatments for Weed Management in Stevia rebaudiana (Bertoni) Cultivation. Available online: <https://www.sciencedirect.com/science> (accessed on 10 June 2020).
- 56. Namaghi, M.N.; Davarynejad, G.H.; Ansary, H.; Nemati, H.; Feyzabady, A.Z. Effects of mulching on soil temperature and moisture variations, leaf nutrient status, growth and yield of pistachio trees (*Pistacia vera* L). *Sci. Hortic.* **2018**, *241*, 115–123. [\[CrossRef\]](http://doi.org/10.1016/j.scienta.2018.06.092)
- 57. Wang, H.; Wang, C.; Zhao, X.; Wang, F. Mulching increases water-use efficiency of peach production on the rainfed semiarid Loess Plateau of China. *Agric. Water Manag.* **2015**, *154*, 20–28. [\[CrossRef\]](http://doi.org/10.1016/j.agwat.2015.02.010)
- 58. Chen, Y.; Wen, X.; Sun, Y.; Zhang, J.; Wu, W.; Liao, Y. Mulching practices altered soil bacterial community structure and improved orchard productivity and apple quality after five growing seasons. *Sci. Hortic.* **2014**, *172*, 248–257. [\[CrossRef\]](http://doi.org/10.1016/j.scienta.2014.04.010)
- 59. Suo, G.D.; Xie, Y.S.; Zhang, Y.; Luo, H. Long-term effects of different surface mulching techniques on soil water and fruit yield in an apple orchard on the Loess Plateau of China. *Sci. Hortic.* **2019**, *246*, 643–651. [\[CrossRef\]](http://doi.org/10.1016/j.scienta.2018.11.028)
- 60. Jin, S.; Wang, Y.; Shi, L.; Guo, X.; Zhang, J. Effects of pruning and mulching measures on annual soil moisture, yield, and water use efficiency in jujube (*Ziziphus jujube* Mill.) plantations. *Glob. Ecol. Conserv.* **2018**, *15*, e00406. [\[CrossRef\]](http://doi.org/10.1016/j.gecco.2018.e00406)
- 61. Basile, B.; Solari, L.I.; Dejong, T.M. Intra-canopy variability of fruit growth rate in peach trees grafted on rootstocks with different vigour-control capacity. *J. Hortic. Sci. Biotechnol.* **2007**, *82*, 243–256. [\[CrossRef\]](http://doi.org/10.1080/14620316.2007.11512226)
- 62. Dai, Z.; Hu, J.; Fan, J.; Fu, W.; Wang, H.; Hao, M. No-tillage with mulching improves maize yield in dryland farming through regulating soil temperature, water and nitrate-N. *Agric. Ecosyst. Environ.* **2021**, *309*, 107228. [\[CrossRef\]](http://doi.org/10.1016/j.agee.2020.107288)
- 63. Zhang, Y.Q.; Wen, Y.; Bai, Q.; Ma, Z.; Ye, H.L.; Su, S.C. Spatio-temporal effects of canopy microclimate on fruit yield and quality of Sapindus mukorossi Gaertn. *Sci. Hortic.* **2019**, *251*, 136–149. [\[CrossRef\]](http://doi.org/10.1016/j.scienta.2019.02.074)
- 64. Kar, G.; Kumar, A.; Martha, M. Water use efficiency and crop coefficients of dry season oilseed crops. *Agric. Water Manag.* **2007**, *87*, 73–82. [\[CrossRef\]](http://doi.org/10.1016/j.agwat.2006.06.002)