



Review

Immunoregulation via Cell Density and Quorum Sensing-like Mechanisms: An Underexplored Emerging Field with Potential Translational Implications

Adrian A. Naoun ^{1,†}, Itay Raphael ^{2,*,†} and Thomas G. Forsthuber ^{1,*}

¹ Department of Molecular Microbiology and Immunology, University of Texas at San Antonio, San Antonio, TX 78249, USA

² Department of Neurological Surgery, School of Medicine, University of Pittsburgh, Pittsburgh, PA 15217, USA

* Correspondence: i.raphael@pitt.edu (I.R.); thomas.forsthuber@utsa.edu (T.G.F.)

† These authors contributed equally to this work.

Abstract: Quorum sensing (QS) was historically described as a mechanism by which bacteria detect and optimize their population density via gene regulation based on dynamic environmental cues. Recently, it was proposed that QS or similar mechanisms may have broader applications across different species and cell types. Indeed, emerging evidence shows that the mammalian immune system can also elicit coordinated responses on a population level to regulate cell density and function, thus suggesting that QS-like mechanisms may also be a beneficial trait of the immune system. In this review, we explore and discuss potential QS-like mechanisms deployed by the immune system to coordinate cellular-level responses, such as T cell responses mediated via the common gamma chain (γ c) receptor cytokines and the aryl hydrocarbon receptors (AhRs). We present evidence regarding a novel role of QS as a multifunctional mechanism coordinating CD4⁺ and CD8⁺ T cell behavior during steady state and in response to infection, inflammatory diseases, and cancer. Successful clinical therapies such as adoptive cell transfer for cancer treatment may be re-evaluated to harness the effects of the QS mechanism(s) and enhance treatment responsiveness. Moreover, we discuss how signaling threshold perturbations through QS-like mediators may result in disturbances of the complex crosstalk between immune cell populations, undesired T cell responses, and induction of autoimmune pathology. Finally, we discuss the potential therapeutic role of modulating immune-system-related QS as a promising avenue to treat human diseases.

Keywords: quorum sensing; immune system; cytokines; T cell homeostasis; macrophage



Citation: Naoun, A.A.; Raphael, I.; Forsthuber, T.G. Immunoregulation via Cell Density and Quorum Sensing-like Mechanisms: An Underexplored Emerging Field with Potential Translational Implications. *Cells* **2022**, *11*, 2442. <https://doi.org/10.3390/cells11152442>

Academic Editor: Francesca Granucci

Received: 11 June 2022

Accepted: 1 August 2022

Published: 6 August 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

The lifestyle of unicellular organisms, such as bacteria and fungi, was initially envisioned as autonomous and reclusive, whereas more sophisticated multicellular organisms were understood to operate by collectively sensing and reacting to signals in a tissue-collaborative fashion. However, it was later observed that substantial increases in bacterial population density resulted in the accumulation of soluble extracellular inducers, which, above a minimal threshold, facilitated synchronized gene expression patterns to regulate behaviors such as biofilm formation, virulence factor expression, motility, and bioluminescence [1–3]. These phenomena have emerged as a density-dependent microbial communication mechanism mediated by molecules known as autoinducers and have been termed quorum sensing (QS) [4]. Recent evidence showed that autoinducer-mediated communication is also observed in other organisms, such as fungi [5], and suggested that QS regulation or similar mechanisms may also govern the mammalian immune system [6]. While the mammalian immune system was long known to elicit synergistic responses, for example, against invading pathogens, the concept of QS-like behavior may add a new layer to our understanding of the underlying mechanisms.

The immune system has a fundamental role in protecting a host from microbial invasion [7,8]. Two arms of the immune system are engaged at different stages of infections and collaborate to defend against microbial invaders [7,9]. Initial responses are non-specific and facilitated by the innate immune system cells such as neutrophils, macrophages, and dendritic cells. Subsequently, the adaptive immune system is engaged via T and B lymphocytes [9].

These collaborative efforts include recruitment of specific immune cell populations to infected or inflamed tissues, recognition, destruction, and antigen presentation of pathogens and coordination of the proper type of immune responses, e.g., antibody class or cytokines for optimized pathogen defense [10]. However, every population of immune cells engaged in this process exhibits some degree of diversity at the level of immune cell subpopulations (e.g., M1 vs. M2 macrophages, different DC populations), or T cell receptors (TCRs) and B cell receptors (BCRs) determined by the clonality of T and B lymphocytes [11,12]. Evidently, innate and adaptive immunity cooperate on multiple levels, for instance, via recruiting adaptive immune cells by cytokines, chemokines, and the complement system provided by the innate immune system [13,14]. However, while factors such as chemokines provide strong, overarching signals recruiting immune cells to infected and/or inflamed tissues, an additional layer of spatiotemporal fine-tuning and collective coordination may be achieved by population-level QS-like cues [6,15]. Along these lines, a central role for QS is based on the capacity of immune cells such as T cells, B cells, and macrophages to sense their own cell density within a certain tissue area, as well as to sense the density of other cell populations in proximity to this region and to coordinate cell density during homeostasis and inflammatory conditions. QS in the immune system may utilize soluble mediators (e.g., cytokines, chemokines, certain metabolites) as indicators of cell density, and these mediators may act on different immune cell types directly, or indirectly via cellular intermediaries. The primary task of QS is thereby assumed to facilitate defense against microbial invasion; however, disruption of immune QS may potentially result in immune-related disorders [15].

Mammalian immunoregulation via a QS-like behavior shows parallels to the bacterial counterparts since many transcriptional alterations are also cell-density-dependent. For example, secretion of cytokines and chemokines in sufficient concentrations facilitates collective receptor-mediated responses to modulate host cell behavior, hence mimicking bacterial autoinducer detection and adaptation to dynamic environmental cues. Thus, eliciting coordinated cellular responses is imperative to regulate tissue and organ homeostasis and function, seemingly coordinated across the cellular up to the entire system levels; therefore, we posit that QS-like behavior may play an important role in immune defense and potentially immune-related dysfunction. In the following sections, we discuss evidence of QS-like behavior in the lymphoid and myeloid lineages and the application of such QS-like mechanisms to design new therapies for cancer and inflammatory diseases.

2. Quorum Sensing-like Mechanisms of T Cells and Regulation of the Adaptive Immune System

Clonally expanded T and B cell populations can be conventionally subclassified based on shared properties such as surface marker expression and cytokine profiles [16–18]. Nevertheless, on a single-cell level, functionality is rendered by a unique TCR or BCR encoding each cell's specificity and affinity for antigens, including microbial and endogenous antigenic epitopes [19–22]. After activation, B cells may differentiate into subpopulations such as germinal center B cells, plasmablasts, and plasma cells, whereas effector T cells can assume Th1, Th2, Th17, or other T cell phenotypes. Contrary to expectations, these B or T cell subpopulations seem superficially alike but may be phenotypically and functionally variable. For instance, a T cell may fall into the Th17 category based on its secretion of IL-17, but not all Th17 cells specific for a shared cognate antigen exhibit identical activation profiles. Even clonal T cells with identical TCR may exhibit different activation profiles depending on their activation state [22–24].

The parameters defining individual versus communal T or B cell behavior may be inherently attributable to the anatomical microenvironment; however, local lymphocyte collaboration should confer host benefits under physiological and inflammatory conditions. Along these lines, single-cell RNA sequencing (scRNA-seq) analyses evaluating resting and TCR-stimulated human T cells from primary, secondary, or mucosal tissue sites revealed conserved gene expression signatures within anatomical compartments [25].

Density of CD4⁺ and CD8⁺ T cells during antigen stimulation may also serve as an additional mechanism regulating the magnitude of T cell activation and/or differentiation in a local milieu. For example, in vivo studies showed that the function and phenotype of CD8⁺ T cells is dictated by the precursor cell pool present during priming [26–28]. Moreover, the trajectory of CD4⁺ T cell differentiation is modulated by the number of locally interacting cells rather than the initial cell concentration [29]. Intercellular communication failures may potentially result in immune aberrations, promoting chronic inflammatory or autoimmune conditions.

2.1. Common-Gamma Chain (γ c) Receptor and Cytokines as QS-like Mechanism for Coordinating T Lymphocyte Responses and Homeostasis

QS in bacteria is defined as an autocrine/paracrine density-dependent mechanism leading to rapid cellular function adaptation [4]. Similarly, lymphocytes have the capability to signal to each other via both autocrine and paracrine mediators, including cytokines, which then results in functional changes and coordinated function [30]. This is important, for example, to maintain the proper homeostasis of naïve and memory T cell populations in an individual [31]. Interestingly, homeostatic regulation of lymphocyte density via proliferation, survival, and metabolic reprogramming is a function reminiscent of bacterial QS.

Conceptually, homeostatic regulatory mechanisms serve to keep the total size of both the naïve and memory T cell pool relatively constant [32–34], and one of the critical mechanisms relies on signals provided via members of the common gamma chain (γ c) cytokine-receptor family and their ligands [35,36], which are commonly referred to as the γ c family of cytokines [37–39]. As a key member of this family, IL-2/IL-2R controls the differentiation and homeostasis of both pro- and anti-inflammatory T cells and is central to determining key aspects of immune regulation [40]. IL-7 and IL-7R, two other members of the γ c family of cytokines, primarily govern survival, whereas IL-15/IL-15R typically promotes basal-level proliferation [31,41]. Importantly, γ c receptors co-regulate each other, which allows rapid synchronization through these cytokines, primarily via members of the signal transducer and activator of transcription (STAT) transcription factors [42].

Following a TCR signal, naïve T cells (T cell priming) and memory T cells (recall) rapidly downregulate IL-7R α expression and upregulate CD25, the high-affinity receptor for IL-2 [42,43]. The expression of CD25 forms a highly stable IL-2R with much higher affinity for IL-2 than IL-15 [44,45]. Consequently, during activation (effector phase), memory T cells become unresponsive to IL-7 and IL-15 and heavily depend on IL-2 for survival and proliferation [31,46,47]. Thus, the γ c family of cytokine receptors and associated cytokines regulate cell numbers of naïve and memory T cells via co-regulation as well as receptor affinity to allow QS-like rapid adaptation tailored to the need of the immune system at different stages and anatomical compartments.

2.1.1. IL-2 Mediated QS-like Behavior of Effector and Regulatory T Cells

IL-2 is a type I cytokine exerting trophic activity to control T cell proliferation and expansion [48]. IL-2 is expressed by T cells upon their activation and operates analogous to a quorum sensing autoinducer promoting autocrine/paracrine T cell expansion, survival, and apoptosis in a STAT3- and STAT5-dependent manner [40,49]. IL-2/IL-2R is less involved in the resting state homeostasis of memory T cells [39,40]. Interestingly, IL-2 is also critical for regulatory T cell (Treg) expansion and survival due to its exquisite dependence on CD25 signaling [50]. Treg cell activation tangentially regulates lymphocyte homeostasis via cytokines such as IL-2, IL-10, and transforming growth factor (TGF)- β by suppressing CD4⁺

and CD8⁺ T cell proliferative activity, irrespective of antigen specificities [51,52]. Thus, it is conceivable that the tissue levels of IL-2 are influenced by local cell density and, in turn, regulate local homeostatic expansion, for example of Tregs (Figure 1).

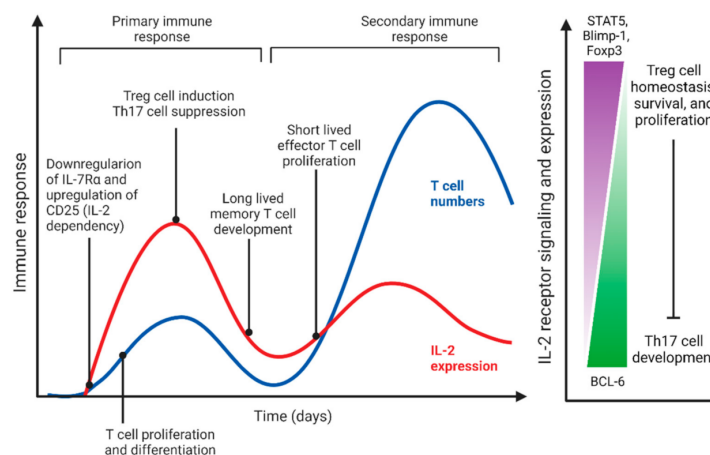


Figure 1. IL-2 as a density-dependent QS-like regulator of T cells. IL-2 and its receptor (CD25) are upregulated in T cells upon activation, leading to density-dependent changes and dose-dependent synchronization of the immune response. Low-dose IL-2 signaling leads to proliferation of early-activated T cells, their differentiation into effector cells, and development of memory T cells. High-dose IL-2 induces Treg cell activation and Th17 cell suppression through control of transcription factors and signaling.

For instance, monoclonal antibody (mAb)-mediated neutralization of circulating IL-2 or IL-2R reduces the abundance and proliferation of Foxp3-expressing CD4⁺ CD25⁺ Tregs during the neonatal period, leading to loss of immune tolerance [53,54]. Additionally, experimental knockout of the downstream transcription factor of IL-2R (CD25), STAT5, results in a loss of tolerance in mice due to a Treg density decrease. Indeed, IL-2 signaling through the high-affinity IL-2R is pivotal in Treg homeostasis, survival, as well as immunosuppression through Foxp3 and CD25 expression [55,56]. The TCR $\alpha\beta$ T cell lineage actively participates in IL-2 secretion, as Tregs remain unable to autonomously secrete the cytokine [57]. Therefore, Tregs ceaselessly ensnare exogenous IL-2 to promote their proliferation and selectively preclude the uncontrollable activation of other immune cells [58]. Notably, in mice, IL-2 deficiency and IL-2 receptor (IL-2R) α or β mutations result in lethal lymphoproliferative autoimmunity [59–62]. Evidence from humans also supports that dysregulation of Tregs can result in immune pathology, as shown by the immunodysregulation polyendocrinopathy enteropathy X-linked (IPEX) syndrome, where individuals show mutations of Foxp3 or, more rarely, CD25 [63,64].

Mounting evidence suggests that homeostatic regulation of effector and regulatory T cell populations can keep the development of autoimmune pathology in check under inflammatory conditions [65–67]. Recently, *in vitro* experimental studies and computer simulations supported the notion of density-dependent modulation of T cells via IL-2; thus, a QS-like mechanism mediated via IL-2 is positioned as a mediator of proliferation and population-level collective responses [68]. *In silico* modeling revealed that effector T cell density modulates the IL-2-mediated phosphorylation of STAT5 to further expand effector T cells during antigenic stimulation in a positive-feedback loop [69]. Interestingly, quantification of IL-2 kinetic rates established that transient fluctuations can result in pleiotropic pSTAT5-mediated effects enhancing Treg immunomodulation or promoting effector T cell survival. Notably, IL-2 scavenging by Tregs resulted in decreased STAT5 phosphorylation and the suppression of both weakly and strongly activated effector T cells [69]. Subtle IL-2 threshold fluctuations presumably promote complex IL-2R-mediated interactions abrogating T cell self-responsiveness or eliciting autoimmune pathogenesis.

The findings by these authors are consistent with a live-cell imaging study in microwell arrays examining the conditional differentiation of progenitor central memory T cells (pTCMs) during priming in OT-II TCR transgenic CD4⁺ T cells. Critically, the magnitude of naïve CD4⁺ T cell differentiation into memory precursors was optimal at a concentration greater than or equal to 30 cells per microwell [29]. Thus, density-dependent interactions collectively manifested due to a dynamic IL-2 and IL-6 regulatory interplay [29]. The authors speculated that novel mediator molecules potentially participate in the intricate differentiation cycle.

2.1.2. QS-like Behavior Mediated by IL-7 and IL-15 in Homeostatic Regulation of Naïve and Memory T Cells

IL-7 is an extracellular matrix-bound cytokine that is constitutively produced by cells in multiple tissues, including (i) stromal and epithelial cells in primary lymphatic organs (e.g., the bone marrow), where IL-7 is involved in lymphopoiesis; (ii) fibroblastic reticular cells in the T cell zone in secondary lymphoid organs; and (iii) cells in non-lymphoid organs including keratinocytes, mucosal epithelial cells, and even neurons [70–73]. CD8⁺ T cells surviving the contraction phase may convert into self-renewing long-lived memory cells through the action of IL-15 (reviewed in [36]). However, CD4⁺ T cells are less dependent on IL-15; therefore, additional factors may constrict the acquisition of long-lived capacities. Recent investigations have established that low clonal abundance favors naïve CD4⁺ T cell maintenance, activation, and survival of memory cell progeny [74]. Evidently, optimally diverse T cell repertoires must be homeostatically regulated to mount effective responses and facilitate long-lived memory CD4⁺ cell generation.

The IL-15R is composed of γc and a receptor chain shared between IL-15R and IL-2R known as IL-15R beta (IL-15R β). Both IL-2R and IL-15R are composed of an additional unique α chain that bestows the highest affinity for each of the cytokines: IL-2R α (CD25) for IL-2 and IL-15R α for IL-15 [75]. Importantly, IL-15R α is unique in that it is expressed by the same cells that express IL-15 and functions to tightly bind IL-15 on the surface of this cell to present it in trans to T cells [75,76]. Thus, a T cell expressing the IL-15R-signaling component receives signals in trans provided by IL-15/IL-15R α expressing neighboring cells [77,78]. Conventional memory T cells typically remain in a resting state where they chiefly respond to IL-7 and IL-15 signals to mediate their maintenance as well as long-term survival [31]. The homeostatic maintenance of the memory T cell pool is dictated by IL-7 availability and exhibits low spontaneous cell turnover [79–82].

Experimental evidence indicates that IL-7 maintains critical basal level proliferation of CD4⁺ Ag-specific memory T cells, especially under lymphopenic conditions [83]. Moreover, IL-7 alternatively induces long-term survival and reversion of effector Ag-specific CD4⁺ memory T cells to a resting state [84–87]. Altogether, local memory CD4⁺ T cells exhibit tight γc cytokine modulation, primarily via IL-7. In summary, the γc family of cytokines and receptors plays a critical role in the development, regulation, and homeostasis of T cells, operating as an autoinducer-like function endowing the adaptive immune system with a QS-like behavior by competing for γc availability and expression, mediated by members of STAT transcription factor family.

2.2. The Aryl Hydrocarbon Receptor (AhR) as a QS-like Regulator in Immune Cells

The dynamic milieu of the immune microenvironment during healthy and disease conditions affords constant cellular adaptation. Homeostatic sensors integrate complex xenobiotic, metabolic, and endogenous stimuli into specific cellular responses. The transcription factor AhR is a highly conserved member of the basic helix–loop–helix/Per-ARNT-Sim (bHLH-PAS) homology sequence along with its competitive AhR repressor (AhRR) and its co-regulator AhR nuclear translocator (ARNT) [88,89]. Notably, AhR is mechanistically a transcription factor also exerting canonical ligand activation to sense a myriad of regulators in the environment such as diet, toxins, and microbiome or endogenous factors such as oxygen level and redox potential [90]. AhR is thereby polyfunctional, acting as a

unique receptor and transcription factor activated directly through cognate ligand binding instead of upstream signal-transduction mediators [91]. Furthermore, AhR is expressed by many immune cells, including T cells, where it plays key roles in early T cell activation, differentiation, and effector functions [90,92]. Interestingly, a recent study indicated that the immune system can detect bacterial autoinducers via AhR, intercepting bacterial QS communications [93,94]. Taken together, as shown in Figure 2, AhR may allow immune cells to rapidly adapt to environmental conditions and can be viewed as a key sensor akin to a QS-like function, which may have evolved from allowing detection of microbial autoinducers by the human immune system to coordinate certain elements of its response at the population level.

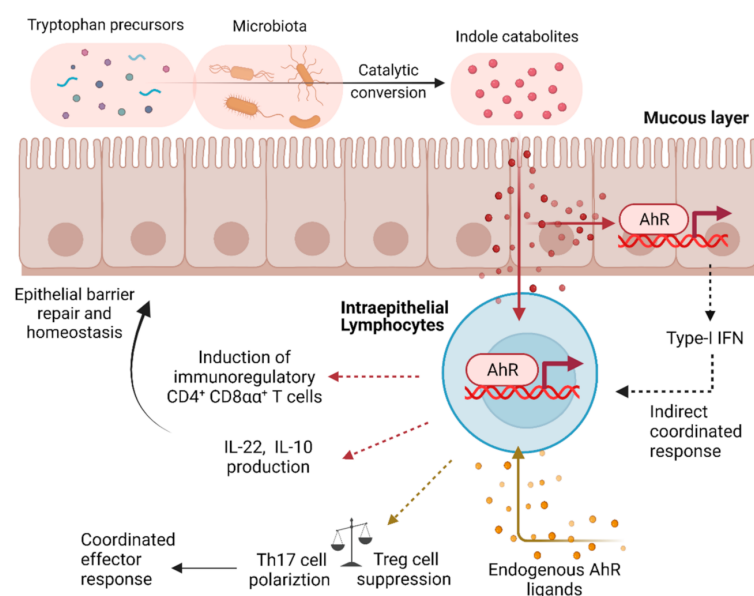


Figure 2. The aryl hydrocarbon receptor (AhR) is a QS-like regulator in immune cells. AhR expressed by immune cells (e.g., intraepithelial T cells) and non-immune cells (e.g., epithelial cells) senses exogenous and endogenous ligands, such as tryptophan metabolites produced by gut microbiota and orchestrates host transcriptional changes resulting in cytokine production and re-programming of T cells in response to environmental changes and altered metabolic demands.

The mammalian digestive tract provides a unique environment to deliver QS signals to orchestrate host and microbiota crosstalk through several mechanisms that involve the immune system [95,96]. Along these lines, tryptophan metabolites produced by gut microbiota serve as AhR ligands in the digestive system [97–99], notably promoting host homeostasis by enhancing the intestinal epithelial barrier, motility, and hormone secretion via exerted anti-inflammatory effects [100]. A recent study uncovered that *Lactobacillus reuteri* activates the AhR to reprogram intraepithelial CD4⁺ T helper cells into CD4⁺ and CD8⁺ double-positive immunoregulatory T lymphocytes [99]. Indole-mediated interplay via AhR orchestrates host transcriptional alterations resulting in IL-22 and type I-IFN signaling to promote epithelial barrier repair during acute inflammatory episodes while additionally tuning intestinal homeostasis via IL-10 secretion [101–105]. These findings are consistent with a mouse model examining AhR agonist deprivation, as IL-22 impairment triggers inflammatory bowel disease [106]. Bacterial catabolite crosstalk thereby ostensibly confers homeostatically protective roles in and via the digestive system partially through AhR signaling.

The immune system regulates cell pool thresholds in response to physiological or unphysiological conditions via expansion or contraction of leukocyte populations, thereby maintaining a delicate steady-state equilibrium. Therefore, similar to the role of AhR in coordinating immune responses to environmental and bacterial cues, it is possible that the immune system has adopted this signaling pathway to also maintain a balance

between immune pathology and immunoregulation, for instance, through coordinating the regulation between inflammatory Th17 cells and regulatory/non-inflammatory Treg cell population size [107,108]. Inherently, Treg/Th17 homeostatic dysregulations contribute to a spectrum of autoimmune, infectious, and cancerous conditions [109–115]. The underlying mechanism is not fully established; nevertheless, the AhR may play a pivotal role. IL-17/IL-22-secreting Th17 subsets express the AhR abundantly; contrarily, transcription is moderately low in Tregs and below detection in Th1, Th2, and naïve CD4⁺ T cells [116,117].

Compelling clinical studies examining ankylosing spondylitis found that the AhR agonist Semaphorin 4D induces Th17 polarization and inhibits Treg differentiation by downregulating Foxp3 expression [118]. Confirmatory transcriptome studies validated that AhR-dependent signaling regulates IL-17 and IL-22 secretion in human CD4⁺ T cells treated with Th17 inducing cytokines [119]. In a similar vein, in vivo studies examining AhR induction with a potent agonist, FICZ, revealed relapses in this experimental autoimmune encephalomyelitis (EAE) model due to exacerbated Th17 differentiation [120]. AhR activation via polycyclic aromatic hydrocarbons, prevalent in cigarette smoke, analogously exacerbates arthritis via increased Th17 polarization in mice [121]. The governing molecular pathway(s) remain incompletely understood. However, AhR dampens STAT1 and STAT5 signaling pathways, dichotomous regulators of Th17 transcriptional programs [122]. Notably, AhR and STAT3 upregulate Aiolos in Th17 polarizing microenvironments to preclude apoptosis via Bcl-2 activity [123,124]. The authors discovered that Aiolos silences the Il2 locus, thus facilitating Th17 differentiation in vitro and in vivo [124]. Altogether, we posit that AhR-mediated activity may induce homeostatic disturbances promoting autoimmune pathogenesis via Treg/Th17 cell pool imbalances in the absence of commensal organisms.

2.3. Bystander Activation of T Cells: QS-like Acute Phase Response of Specific T Cell Subsets

Bystander activation is a mechanism where T cells specific for unrelated antigens proliferate and/or are activated in various disease conditions independent of TCR signaling [125–127]. Bystander proliferation was first reported by Tough et al. in 1996 as a massive expansion of CD8⁺ T cells in the absence of detectable TCR signaling after heterologous virus infection [128]. Subsequent studies, however, showed that the extent of bystander activation may have been overestimated initially and that the majority of T cells activated by infections are virus-specific [129,130]. Nevertheless, bystander activation is thought to be a mechanism that may be beneficial or detrimental to the host depending on the circumstances [127].

Bystander activation is primarily observed by CD8⁺ memory T cells (T_{mem}) and occurs rapidly during the earliest phase of infection [127]. It is driven by proinflammatory cytokines, including type I interferons (IFNs), IL-12, IL-15, and IL-18, and appears to be mediated independently of cognate TCR signaling [127]. Bystander activation by CD4⁺ T cells is less well understood but is also centered on T_{mem} subsets and seems to be less efficient due to decreased expression of CD122 [126]. As pointed out earlier, bystander activation is primarily observed under infectious conditions, and it was first described during virus infections. However, bystander activation was also demonstrated during bacterial infections and by bacterial components such as LPS [125,126]. Indeed, Toll-like receptors (TLRs) or other innate-like receptors such as NKG2D have been reported to mediate bystander activation [125,126]. Bystander activation can result in T cell proliferation, cytokine expression, and direct cytolysis of infected cells [127] and may therefore be beneficial during infection. However, bystander activation may be detrimental in certain conditions such as chronic infections or autoimmune diseases [126].

Taken together, bystander activation shows some similarities with QS and encompasses certain aspects of it, but it seems to be centered on specific T cell subsets (e.g., T_{mem}) and to occur only during particular circumstances, such as infections and chronic disease conditions. Moreover, QS-like behavior of adaptive and innate immune cells seems to conceptually separate from bystander activation by focusing on synchronizing the behavior of

communal cell populations, whereas bystander activation seems to be focused on eliciting a (non-specific) effector response to support the earliest phase of anti-microbial immunity.

3. QS-like Regulation of Myeloid Cells and the Innate Immune System

3.1. QS as a Myeloid Lineage Modulator: Population Density of Tissue-Resident Macrophages Contributes to Spatiotemporal Regulation

Tissue-resident macrophages are phagocytes responsible for maintaining tissue homeostasis and repair, defending against pathogen invasion, and removing extracellular debris such as apoptotic cells and toxic metabolic products [131,132]. Current evidence supports that most tissue-resident macrophages may arise from embryonic precursors seeding during development as opposed to primarily deriving from hematopoietic stem cells [133–136]. Consequently, the classic paradigm postulating macrophage replenishment via circulating monocytes has shifted to incorporate embryonic precursor self-renewal based on dynamic tissue regulation.

Macrophages exhibit prolific physiological plasticity in response to endogenous and exogenous stimuli [137]. Consequently, phenotypic variation results from diverging cellular differentiation patterns partly constrained by microenvironmental parameters. Historically, macrophage polarization nomenclature comprises the classically (M1) and alternatively activated (M2) populations distinguished by unique activation markers [138]. M1 polarization is functionally associated with inflammatory (post-infectious pathogenesis) and microbicidal activity, whereas the M2 phenotype confers immunomodulatory properties mediating inflammation resolution [139]. Intricate phenotypic distributions coexist in specific tissues; therefore, regulatory mechanisms spatiotemporally exert homeostatic balance.

Compelling evidence suggests that regulatory mechanisms akin to QS serve to reestablish the steady-state concentration of macrophages following infectious episodes and injury [15]. These findings are also consistent with seminal research investigating the effects of diphtheria toxin (DT)-mediated selective depletion of liver-resident macrophages known as Kupffer cells (KCs) in a humanized mouse model [140]. The authors identified that congregating macrophages or myeloid cells acquire transcriptionally homologous tissue-resident macrophage signatures, including self-renewal capacities, within 15 days of colonization [140]. Additionally, acute acetaminophen (N-acetyl-p-aminophenol) overdose stimulated the self-renewal of tissue-resident macrophages following a marked reduction in the KC population [141]. In this case, the recruitment of circulating monocytes resulted in a transcriptionally distinct phenotype that failed to contribute to KC replenishment. The exact mechanism(s) governing the spatiotemporal regulation of tissue-resident macrophage density following threshold diminishment and the transcriptional acquisition of self-renewing phenotypes by monocyte-derived macrophages remain unsolved. However, these data collectively suggest that the steady-state concentration of macrophages is dynamically regulated in response to infection and collateral tissue damage. Consequently, it is conceivable that QS-like mechanisms could regulate tissue homeostasis of macrophage populations.

3.2. Apoptotic Metabolite Release as a Putative QS-like Mechanism Governing Macrophage Density

Groundbreaking mechanistic studies indicate that apoptotic metabolites orchestrate transcriptional alterations facilitating inflammation resolution, cell proliferation, and tissue regeneration in healthy neighboring cells to restore a homeostatic state [142]. Secretome profiling analyses identified six conserved metabolites endowing a fundamental signaling role: AMP, GMP, creatine, spermidine, glycerol 3-phosphate, and ATP [142]. The authors detected diverse apoptotic metabolites in macrophage and lymphocyte cell pellets, although caspase-mediated activation of pannexin-1 hemichannels selectively regulated molecular release under intact membrane conditions. These data collectively challenge the paradigm construing programmed cell death pathways as metabolically inert. Contrarily, the apoptotic secretome is selectively regulated to coordinate collective gene expression in local microenvironments. We postulate that signaling via apoptotic metabolites induces threshold-specific transcription programs similar to QS. Elevated apoptotic metabolite

concentrations can conceivably modulate synergistic macrophage responses in dynamic tissue parameters such as collateral tissue damage and infectious disease [143]. Moreover, metabolites that are secreted or diffuse from apoptotic cells, such as ATP and ADP, can regulate both the adaptive and innate immune system simultaneously to coordinate the immune response and immune cell homeostasis [144–148], thus endowing it with a QS-like mechanism to regulate tissue populations in response to local cell death (Figure 3).

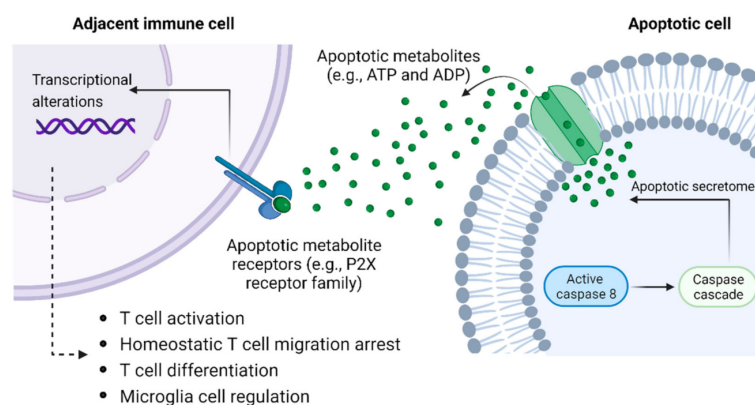


Figure 3. Apoptotic metabolites as QS-like regulators of immune cell function. Apoptotic metabolites released by cells undergoing apoptosis can induce transcriptional programs similar to QS to synchronize and coordinate T cell responses (and potentially those of other immune cells) in the proximity to regulate dynamic tissue responses to infection and tissue damage. Important known apoptotic metabolites include ATP and ADP, which signal through the P2X receptor family involved in inducing T cell activation, differentiation, and migratory changes.

In the central nervous system (CNS), microglia mechanistically operate as resident macrophages to safeguard and support neuronal functions [149]. Nevertheless, macrophages and microglial cells differ in calcium fluctuation responses, expressed biomarkers, and inflammatory profiles following traumatic events such as brain ischemia [150–153]. In the face of inflammatory or pathological insults, CNS microglia and macrophages establish the first line of defense via innate and adaptive immune components [154]. Uncontrolled microglial activation and homeostatic dysregulation contribute to CNS disorders. Indeed, unresolved chronic inflammation can result in neuronal and glial damage.

Recently, a multicolor fluorescence mapping reporter system using confocal microscopy established that expansion of microglia populations increased proximal to the site of facial nerve transection in mice; however, the pre-injury network density of microglial cells was homeostatically restored [155]. The authors also found that microglial self-renewal rates corresponded to cortex, cerebellum, and hippocampus morphological proliferation in healthy mice. We posit that the spatiotemporal regulation of microglial cells in response to injury is a QS-like mechanism, potentially guided via apoptotic metabolite release. Nonetheless, the mechanisms remain in question as the collective interaction of microglia and macrophages to restore and maintain a proper tissue density is unknown.

3.3. QS-like Density-Dependent Polarization of Macrophages

Recent evidence indicates that a density-dependent QS-like mechanism controls macrophage polarization and the magnitude of the inflammatory response [156–158]. For instance, it was reported that follicular micro-injuries caused by hair plucking trigger a concerted macrophage-mediated regeneration of resting cells in mice [157]. Subsequent molecular and genetic analyses unraveled a two-step mechanism mediated by the follicular secretion of CC-chemokine ligand 2 (CCL2) to induce the dermal recruitment of TNF-expressing macrophages to promote local regeneration [157]. Interestingly, macrophages collectively assess the magnitude of hair follicle injuries to elicit all-or-none responses [157]. Macrophage accumulation and dissemination could therefore serve as a mechanism orches-

trating local keratinocyte apoptosis to sustain a coordinated regenerative anagen phase. Along these lines, a single-cell tracking study determined that the lipopolysaccharide-induced bimodal phenotypic partitioning of primary macrophages is contingent on cell density and concerted gene expression [25]. These observations were TNF-independent, as distinct polarizations originated due to resting-state density priming, a pre-programmed response coined ‘quorum licensing’ by the authors. Hence, cell density information potentially regulates the collective activation of macrophages.

Colony-stimulating factor 1 (CSF-1) has been postulated as a central regulator of macrophage density at a steady state [6,159]. For example, stromal cells and endoneurial fibroblasts secrete CSF-1 to modulate macrophage survival, proliferation, as well as differentiation in vivo [160,161]. However, the role of CSF-1 remains controversial as the interpretation of results can be confounded by the pleiotropic effects exhibited by pharmacological or antibody-mediated CSF-1 receptor (CSF-1R) blocking. The ligands CSF-1 and interleukin-34 (IL-34) share the promiscuous CSF-1R; therefore, receptor neutralization may result in local and systemic off-target effects that may have confounded the interpretation of the results. Similarly, CSF-1R neutralization in vivo leads to physiological increases in CSF-1 levels, thus presumably interfering with the monoclonal antibody’s activity [162]. Additionally, alternative splicing and differential proteolysis render three CSF-1 homodimeric isoforms which may differ in affinity and function: a cell-surface glycoprotein (csCSF-1), a secreted glycoprotein (sgCSF-1), and a proteoglycan (spCSF-1) [163]. Notably, recent evidence revealed that spCSF-1 induces macrophage activation and neuronal damage, whereas csCSF-1 attenuates macrophage-mediated neuropathy in a mouse model for Charcot–Marie–Tooth type 1X disease [164]. Additional studies are required to further dissect the intrinsic role of each CSF-1 isoform as a QS-like cell density regulator in macrophages.

3.4. Intersection of Microbial QS with Regulation of Myeloid Cells and Tissue Macrophages

Macrophages undergo multifaceted regulation during the course of infectious diseases. In the lung microenvironment, alveolar macrophages are sentinel cells responsible for maintaining lung homeostasis, clearing cellular debris, and protecting against pathogen invasion [165,166]. However, pathogens have evolved exquisite mechanisms to evade immune surveillance, survive in hostile microenvironments, and subvert host responses. Facultative intracellular bacteria such as the pulmonary pathogen *Mycobacterium tuberculosis* (*Mtb*) can establish a permissive milieu conducive to disease persistence and systemic dissemination [167].

Apoptosis is an innate host-protective mechanism precluding *Mtb* propagation (reviewed in [168,169]); in turn, *Mtb* can differentially manipulate the timing and mode of cell death in infected alveolar macrophages [170–172]. Murine and human macrophage models showed that virulent *Mtb* strains dampen apoptosis while favoring necrotic niches associated with granuloma formation, bacterial replication, as well as uncontrolled dissemination [173,174]. Compelling evidence also indicates that human macrophage coordination at high cell density (2×10^5 cells/well) profoundly suppressed *Mycobacterium bovis* pathogenesis and growth of a live-attenuated strain known as BCG [156]. The effects were independent of mycobacterial uptake, multiplicity of infection, extracellular medium acidification, nitric oxide production, or paracrine stimulation via cytokines, as the study considered such confounding variables. We propose that QS-like collective macrophage regulation may be mediated by apoptotic metabolites and promotes cellular synergism. Likewise, TLR activity emerges as an alternative mechanism regulating QS-like responses in macrophages and dendritic cells (DCs). Viral sensors such as TLR3 provoke phenotypic alterations in DC biology, including type I IFN secretion for microenvironmental propagation [175]. A compelling study suggests that optimal DC activation is mediated by a quorum of type I IFN-secreting cells at the lymph node level [176]. The authors also found that collective DC activation is essential to mount robust innate and adaptive immune responses in the lymph node. Of note, activation of CD4⁺ T cells requires a minimum threshold of approximately 85 antigen-presenting DCs in lymph nodes [177]. Similarly, TNF

secreting populations induce concerted macrophage activation and coordinate responses at the population level in response to LPS stimulation of TLR4 [178]. Thus, TLR-mediated signaling may induce QS-like regulation to mount synergistic responses geared towards eliminating foreign invaders.

4. Clinical Implications

4.1. Exploring QS-like Regulation of T Cells for Inflammatory Diseases and GVHD

Coordinating T cell responses via γ c receptors and cytokines can act as a QS-like mechanism for the regulation of T cells under inflammatory conditions. For example, IL-2 can promote cellular responses akin to QS. Of note, IL-2 has been investigated as a viable therapeutic modality to treat underlying autoimmune pathologies, cancer, hypersensitivity responses, and allograft rejection [50,179–181].

Currently, the implications of IL-2 as a QS-like mediator have not been fully explored, yet this area may hold promise for treatment purposes. Along these lines, pharmacokinetic studies investigating mice and human responses to low-dose IL-2 (LD IL-2) have identified marked increases in Treg activation and suppressive activities [182–185]. In NOD mice, treatment for five days with LD IL-2 prevented the onset of type 1 diabetes by increasing pancreatic Treg populations [186]. Randomized placebo-controlled phase I and II clinical trials exposed six patients to daily placebos or IL-2 doses of 0.33, 1, or 3 million international units (MIU) for a 5-day course (reviewed elsewhere [50]). The results showed that IL-2 therapy facilitated a dose-dependent increase in CD4⁺ and CD8⁺ Treg cells in the absence of detrimental adverse reactions [187]. Similarly, modified IL-2 with extended in vivo half-life (termed IL-2 mutein) was shown to selectively activate and expand regulatory T cells [179].

Furthermore, clinical trials investigating the effect of LD IL-2 in active systemic lupus erythematosus (SLE) have been successful in mitigating refractory phenotypes to standard therapy [188]. Severe SLE manifestations can be attenuated with an initial IL-2 dosage of 1 MIU augmented to 3 MIU for five days; consequently, Treg populations increased while anti-dsDNA antibody concentrations were reduced [189]. The clinical feasibility of LD IL-2 has been extrapolated as a novel treatment for autoimmune alopecia areata [190]. The results of the study showed that low-dose recombinant IL-2 induced a marked increase in CD4⁺ CD25⁺ FoxP3⁺ Treg recruitment, accompanied by a decrease in effector CD8⁺ T (T_{eff}) cells from scalp biopsies [190]. Treatment resulted in severity of alopecia tool scores decreasing from 82 to a baseline of 69, therefore manifesting as partial hair regrowth in 4 out of 5 patients at six months with no reported adverse reactions [190]. These results collectively suggest a QS-like mechanism for regulating Tregs and effector T cells in autoimmune diseases.

LD IL-2 also conferred beneficial effects for the treatment of chronic graft versus host disease (GVHD) after allogeneic hematopoietic stem cell transplantation [191,192]. LD IL-2 treatment induced the homeostatic regulation of CD4⁺ T cells following transplantation as well as an increased expansion of Tregs [193,194]. In allogeneic hematopoietic stem cell transplantation patients, chronic myelogenous leukemia was significantly ameliorated upon LD IL-2 administration combined with Treg infusion [195]. LD IL-2 can alternatively elicit Treg expansion and confer a protective role in clinical manifestations of food allergy in mouse models [196]. Thus, regulation of cell density via IL-2, for example, for Tregs, may be an example of exploiting the mechanism of QS for therapeutic benefits. A critical question that has not been addressed currently is whether increased cell density translates to changes in function in these systems.

We posit that dysregulation of one T cell subset (i.e., Treg cells) results in disturbances of other T cell subsets, for example, by promoting effector T cell activities in inflammatory/autoimmune diseases. Density dysregulation may therefore be a key feature in a spectrum of adverse immunological conditions. The exact pathophysiological mechanism(s) remain incompletely resolved; however, it appears that mediators, such as IL-2, can regulate collective responses in a QS-like mechanism and that their disruption may foster tissue pathology. Accordingly, threshold parameters in IL-2, for example, will dictate

their immunomodulatory potential as a novel strategy to treat pathological inflammatory conditions. In addition, other QS-like mediators, such as AhR agonists, may have similar or even more potent effects on T cell regulation. Thus, we propose that exploration of QS-like modulation in immune cell populations may hold as of yet untapped potential.

4.2. QS-like Modulation of T Cells for Cancer Immunotherapy

Oncogene expression and tumor suppressor deactivation are among the mechanisms mediating carcinogenesis, an uncontrolled cellular proliferation eluding cell-regulatory mechanisms [197]. Surgical excision of tumors is typically the primary modality for solid cancers, yet surgical or other interventions such as radiation or chemotherapy are often futile in advanced cancer stages, where most malignant tumors have broken through organ confines, invaded other tissues, and metastasized [198]. Immunotherapies have emerged as promising adjunct or salvage alternatives for cancers that are difficult to treat with conventional medical therapies, such as metastatic malignant melanoma. Ultimately, immunotherapies could selectively target cancer cells while mitigating residual side effects associated with traditional treatment regimens. For example, checkpoint inhibition, adoptive cell therapy, and bivalent antibodies are clinically successful immunotherapies in combating otherwise incurable malignancies.

In contrast to harnessing inflammatory conditions with LD IL-2, high-dose IL-2 (HD IL-2) formulations were used to combat malignant tumors, for example in melanoma and renal cancer patients [199–201]. In malignant melanoma patients, drug administration resulted in a durable overall remission in 16% of the cases with reported toxicities such as hypotension and secondary to underlying capillary leak [202]. Moreover, patients exhibited a tumor regression rate of 20% in renal cell carcinoma trials [203]. Collectively, these data indicated that HD IL-2 promoted a polyclonal expansion of effector T cell subsets to ameliorate anergy and mediate tumor regression. Of note, experimental evidence indicates that exogenous HD IL-2 abrogated CD25⁺ FoxP3⁺ Treg-induced immunosuppression [204]. Commensurate increases in effector T cell population density circumvent Treg attenuation; thus, the QS-like effects of IL-2 may emerge as a viable concept in immuno-oncology.

4.3. Implications of QS for Treatment of Infectious or Inflammatory Diseases via Regulation of Macrophage and Myeloid Cells

Therapeutically, the feasibility of exploiting QS-like signaling for host-directed therapies to optimize antimicrobial immunity, ameliorate immunopathology, and collateral tissue damage has yet to be determined. However, QS-like regulation of density-dependent macrophage responses may hold untapped potential for improving difficult-to-treat infectious diseases, such as multidrug-resistant tuberculosis. Macrophages may be instructed to collectively polarize, thus resulting in enhanced pathogen neutralization. Further research with diverse pathogens may reveal novel mechanisms in macrophage biology, as, for example, apoptosis metabolite thresholds are unknown for specific microenvironments. Furthermore, compelling preliminary data suggest that local administration of select metabolite cocktails dampens inflammatory arthritis and lung graft rejection in mouse models [142]. Taken together, research to harness macrophage QS may have the potential to lead to a new generation of host-directed therapies.

5. Concluding Remarks

Spatiotemporal modulation of adaptive and innate immune cell density following infectious episodes, acute injury, and immunopathology has profound consequences for immune responsiveness and homeostasis. QS-like signaling has the potential to guide collective tissue immune responses, for example, via density-dependent synergistic interactions. Furthermore, the mammalian digestive tract poses a unique QS-sensing portal orchestrating host–commensal crosstalk.

To further elucidate the biological complexities of QS, complementary approaches will benefit from integrating biological models with bioinformatics and omics studies in

combination with computational models. The application of these approaches to QS in bacteria has revealed novel insights, for example to decipher biofilm formation [205,206]. Similarly, theoretical considerations and mathematical modeling of QS for effector and regulatory T cells have provided novel insights and broadened our conceptual understanding of immune QS and QS-based decision making [207–209]. It is hoped that these approaches accelerate our comprehension of QS in the immune system and facilitate discovery of regulatory mechanisms and autoinducers.

We posit that QS-like mechanisms endow a regulatory role in immune homeostasis and communal effector responses. Therefore, we anticipate that cell pool dysregulations may etiologically contribute to immune aberrations. Compelling evidence revealed that AhR-mediated signaling induces robust polarization of the inflammatory T_H17 subset while selectively downregulating Foxp3 expression to preclude Treg differentiation. Subsequently, Treg/T_H17 cell pool imbalances might conceivably foster autoimmune pathogenesis. Future studies will demonstrate the clinical feasibility of host-directed therapies tailored to QS-like mechanisms in human disease conditions.

Author Contributions: Conceptualization, A.A.N., I.R. and T.G.F.; Writing—original draft preparation, A.A.N. and I.R.; Writing—review and editing, A.A.N., I.R. and T.G.F.; Supervision, T.G.F. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by grants NS117742 and AI144731 from the National Institute of Health (T.G.F.) and support from the Jesse H. & Mary Jones Gibbs Endowed Chair (T.G.F.). I.R. was supported by a fellowship from UPMC Children’s Hospital of Pittsburgh and The Walter L. Copeland Foundation.

Acknowledgments: Figures were created using BioRender.com.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Ng, W.L.; Bassler, B.L. Bacterial quorum-sensing network architectures. *Annu. Rev. Genet.* **2009**, *43*, 197–222. [[CrossRef](#)] [[PubMed](#)]
2. Hoang, H.H.; Gurich, N.; Gonzalez, J.E. Regulation of motility by the ExpR/Sin quorum-sensing system in *Sinorhizobium meliloti*. *J. Bacteriol.* **2008**, *190*, 861–871. [[CrossRef](#)] [[PubMed](#)]
3. Rutherford, S.T.; Bassler, B.L. Bacterial quorum sensing: Its role in virulence and possibilities for its control. *Cold Spring Harb Perspect. Med.* **2012**, *2*, a012427. [[CrossRef](#)] [[PubMed](#)]
4. Mukherjee, S.; Bassler, B.L. Bacterial quorum sensing in complex and dynamically changing environments. *Nat. Rev. Microbiol.* **2019**, *17*, 371–382. [[CrossRef](#)] [[PubMed](#)]
5. Padder, S.A.; Prasad, R.; Shah, A.H. Quorum sensing: A less known mode of communication among fungi. *Microbiol. Res.* **2018**, *210*, 51–58. [[CrossRef](#)]
6. Antonioli, L.; Blandizzi, C.; Pacher, P.; Williams, M.; Hasko, G. Quorum sensing in the immune system. *Nat. Rev. Immunol.* **2018**, *18*, 537–538. [[CrossRef](#)]
7. Brodin, P.; Davis, M.M. Human immune system variation. *Nat. Rev. Immunol.* **2017**, *17*, 21–29. [[CrossRef](#)]
8. Parkin, J.; Cohen, B. An overview of the immune system. *Lancet* **2001**, *357*, 1777–1789. [[CrossRef](#)]
9. Netea, M.G.; Dominguez-Andres, J.; Barreiro, L.B.; Chavakis, T.; Divangahi, M.; Fuchs, E.; Joosten, L.A.B.; van der Meer, J.W.M.; Mhlanga, M.M.; Mulder, W.J.M.; et al. Defining trained immunity and its role in health and disease. *Nat. Rev. Immunol.* **2020**, *20*, 375–388. [[CrossRef](#)]
10. Iwasaki, A.; Medzhitov, R. Control of adaptive immunity by the innate immune system. *Nat. Immunol.* **2015**, *16*, 343–353. [[CrossRef](#)]
11. Boehm, T. Design principles of adaptive immune systems. *Nat. Rev. Immunol.* **2011**, *11*, 307–317. [[CrossRef](#)] [[PubMed](#)]
12. Liston, A.; Humblet-Baron, S.; Duffy, D.; Goris, A. Human immune diversity: From evolution to modernity. *Nat. Immunol.* **2021**, *22*, 1479–1489. [[CrossRef](#)] [[PubMed](#)]
13. Gasteiger, G.; Rudensky, A.Y. Interactions between innate and adaptive lymphocytes. *Nat. Rev. Immunol.* **2014**, *14*, 631–639. [[CrossRef](#)]
14. Vivier, E.; Malissen, B. Innate and adaptive immunity: Specificities and signaling hierarchies revisited. *Nat. Immunol.* **2005**, *6*, 17–21. [[CrossRef](#)]
15. Antonioli, L.; Blandizzi, C.; Pacher, P.; Williams, M.; Hasko, G. Rethinking Communication in the Immune System: The Quorum Sensing Concept. *Trends Immunol.* **2019**, *40*, 88–97. [[CrossRef](#)]
16. Raphael, I.; Nalawade, S.; Eagar, T.N.; Forsthuber, T.G. T cell subsets and their signature cytokines in autoimmune and inflammatory diseases. *Cytokine* **2015**, *74*, 5–17. [[CrossRef](#)]

17. Raphael, I.; Joern, R.R.; Forsthuber, T.G. Memory CD4(+) T Cells in Immunity and Autoimmune Diseases. *Cells* **2020**, *9*, 531. [[CrossRef](#)]
18. Negron, A.; Robinson, R.R.; Stuve, O.; Forsthuber, T.G. The role of B cells in multiple sclerosis: Current and future therapies. *Cell Immunol.* **2019**, *339*, 10–23. [[CrossRef](#)] [[PubMed](#)]
19. van Bladel, D.A.G.; van den Brand, M.; Rijntjes, J.; Pamidimarri Naga, S.; Haacke, D.; Luijks, J.; Hebeda, K.M.; van Krieken, J.; Groenen, P.; Scheijen, B. Clonality assessment and detection of clonal diversity in classic Hodgkin lymphoma by next-generation sequencing of immunoglobulin gene rearrangements. *Mod Pathol.* **2021**, *35*, 757–766. [[CrossRef](#)]
20. Gascoigne, N.R.; Rybakin, V.; Acuto, O.; Brzostek, J. TCR Signal Strength and T Cell Development. *Annu. Rev. Cell Dev. Biol.* **2016**, *32*, 327–348. [[CrossRef](#)]
21. Niiro, H.; Clark, E.A. Regulation of B-cell fate by antigen-receptor signals. *Nat. Rev. Immunol.* **2002**, *2*, 945–956. [[CrossRef](#)] [[PubMed](#)]
22. Adams, N.M.; Grassmann, S.; Sun, J.C. Clonal expansion of innate and adaptive lymphocytes. *Nat. Rev. Immunol.* **2020**, *20*, 694–707. [[CrossRef](#)] [[PubMed](#)]
23. Hesse, M.D.; Karulin, A.Y.; Boehm, B.O.; Lehmann, P.V.; Tary-Lehmann, M. A T cell clone's avidity is a function of its activation state. *J. Immunol.* **2001**, *167*, 1353–1361. [[CrossRef](#)]
24. Chiffelle, J.; Genolet, R.; Perez, M.A.; Coukos, G.; Zoete, V.; Harari, A. T-cell repertoire analysis and metrics of diversity and clonality. *Curr. Opin. Biotechnol.* **2020**, *65*, 284–295. [[CrossRef](#)] [[PubMed](#)]
25. Szabo, P.A.; Levitin, H.M.; Miron, M.; Snyder, M.E.; Senda, T.; Yuan, J.; Cheng, Y.L.; Bush, E.C.; Dogra, P.; Thapa, P.; et al. Single-cell transcriptomics of human T cells reveals tissue and activation signatures in health and disease. *Nat. Commun.* **2019**, *10*, 4706. [[CrossRef](#)]
26. Obar, J.J.; Khanna, K.M.; Lefrancois, L. Endogenous naive CD8+ T cell precursor frequency regulates primary and memory responses to infection. *Immunity* **2008**, *28*, 859–869. [[CrossRef](#)]
27. Badovinac, V.P.; Haring, J.S.; Harty, J.T. Initial T cell receptor transgenic cell precursor frequency dictates critical aspects of the CD8(+) T cell response to infection. *Immunity* **2007**, *26*, 827–841. [[CrossRef](#)]
28. Marzo, A.L.; Klonowski, K.D.; Le Bon, A.; Borrow, P.; Tough, D.F.; Lefrancois, L. Initial T cell frequency dictates memory CD8+ T cell lineage commitment. *Nat. Immunol.* **2005**, *6*, 793–799. [[CrossRef](#)]
29. Polonsky, M.; Rimer, J.; Kern-Perets, A.; Zaretsky, I.; Miller, S.; Bornstein, C.; David, E.; Kopelman, N.M.; Stelzer, G.; Porat, Z.; et al. Induction of CD4 T cell memory by local cellular collectivity. *Science* **2018**, *360*, eaaj1853. [[CrossRef](#)]
30. Altan-Bonnet, G.; Mukherjee, R. Cytokine-mediated communication: A quantitative appraisal of immune complexity. *Nat. Rev. Immunol.* **2019**, *19*, 205–217. [[CrossRef](#)]
31. Schluns, K.S.; Lefrancois, L. Cytokine control of memory T-cell development and survival. *Nat. Rev. Immunol.* **2003**, *3*, 269–279. [[CrossRef](#)] [[PubMed](#)]
32. Mackall, C.L.; Hakim, F.T.; Gress, R.E. Restoration of T-cell homeostasis after T-cell depletion. *Semin. Immunol.* **1997**, *9*, 339–346. [[CrossRef](#)] [[PubMed](#)]
33. Bell, E.B.; Sparshott, S.M. The peripheral T-cell pool: Regulation by non-antigen induced proliferation? *Semin. Immunol.* **1997**, *9*, 347–353. [[CrossRef](#)] [[PubMed](#)]
34. Sprent, J.; Tough, D.F. Lymphocyte life-span and memory. *Science* **1994**, *265*, 1395–1400. [[CrossRef](#)]
35. Surh, C.D.; Sprent, J. Homeostasis of naive and memory T cells. *Immunity* **2008**, *29*, 848–862. [[CrossRef](#)]
36. Jameson, S.C. Maintaining the norm: T-cell homeostasis. *Nat. Rev. Immunol.* **2002**, *2*, 547–556. [[CrossRef](#)]
37. Masse, G.X.; Corcuff, E.; Decaluwe, H.; Bommhardt, U.; Lantz, O.; Buer, J.; Di Santo, J.P. gamma(c) cytokines provide multiple homeostatic signals to naive CD4(+) T cells. *Eur. J. Immunol.* **2007**, *37*, 2606–2616. [[CrossRef](#)]
38. Kinter, A.L.; Godbout, E.J.; McNally, J.P.; Sereti, I.; Roby, G.A.; O'Shea, M.A.; Fauci, A.S. The common gamma-chain cytokines IL-2, IL-7, IL-15, and IL-21 induce the expression of programmed death-1 and its ligands. *J. Immunol.* **2008**, *181*, 6738–6746. [[CrossRef](#)]
39. Overwijk, W.W.; Schluns, K.S. Functions of gammaC cytokines in immune homeostasis: Current and potential clinical applications. *Clin. Immunol.* **2009**, *132*, 153–165. [[CrossRef](#)]
40. Boyman, O.; Sprent, J. The role of interleukin-2 during homeostasis and activation of the immune system. *Nat. Rev. Immunol.* **2012**, *12*, 180–190. [[CrossRef](#)]
41. Ross, S.H.; Cantrell, D.A. Signaling and Function of Interleukin-2 in T Lymphocytes. *Annu. Rev. Immunol.* **2018**, *36*, 411–433. [[CrossRef](#)] [[PubMed](#)]
42. Rochman, Y.; Spolski, R.; Leonard, W.J. New insights into the regulation of T cells by gamma(c) family cytokines. *Nat. Rev. Immunol.* **2009**, *9*, 480–490. [[CrossRef](#)] [[PubMed](#)]
43. Di Santo, J.P.; Aifantis, I.; Rosmaraki, E.; Garcia, C.; Feinberg, J.; Fehling, H.J.; Fischer, A.; von Boehmer, H.; Rocha, B. The common cytokine receptor gamma chain and the pre-T cell receptor provide independent but critically overlapping signals in early alpha/beta T cell development. *J. Exp. Med.* **1999**, *189*, 563–574. [[CrossRef](#)] [[PubMed](#)]
44. Di Santo, J.P.; Kuhn, R.; Muller, W. Common cytokine receptor gamma chain (gamma c)-dependent cytokines: Understanding in vivo functions by gene targeting. *Immunol. Rev.* **1995**, *148*, 19–34. [[CrossRef](#)] [[PubMed](#)]
45. Yamane, H.; Paul, W.E. Cytokines of the gamma(c) family control CD4+ T cell differentiation and function. *Nat. Immunol.* **2012**, *13*, 1037–1044. [[CrossRef](#)] [[PubMed](#)]

46. Xue, H.H.; Kovanen, P.E.; Pise-Masison, C.A.; Berg, M.; Radovich, M.F.; Brady, J.N.; Leonard, W.J. IL-2 negatively regulates IL-7 receptor alpha chain expression in activated T lymphocytes. *Proc. Natl. Acad. Sci. USA* **2002**, *99*, 13759–13764. [[CrossRef](#)]
47. Kovanen, P.E.; Leonard, W.J. Cytokines and immunodeficiency diseases: Critical roles of the gamma(c)-dependent cytokines interleukins 2, 4, 7, 9, 15, and 21, and their signaling pathways. *Immunol. Rev.* **2004**, *202*, 67–83. [[CrossRef](#)]
48. Lucey, D.R.; Clerici, M.; Shearer, G.M. Type 1 and type 2 cytokine dysregulation in human infectious, neoplastic, and inflammatory diseases. *Clin. Microbiol. Rev.* **1996**, *9*, 532–562. [[CrossRef](#)]
49. Benczik, M.; Gaffen, S.L. The interleukin (IL)-2 family cytokines: Survival and proliferation signaling pathways in T lymphocytes. *Immunol. Investig.* **2004**, *33*, 109–142. [[CrossRef](#)]
50. Ye, C.; Brand, D.; Zheng, S.G. Targeting IL-2: An unexpected effect in treating immunological diseases. *Signal Transduct. Target* **2018**, *3*, 2. [[CrossRef](#)]
51. Piccirillo, C.A.; Thornton, A.M. Cornerstone of peripheral tolerance: Naturally occurring CD4+CD25+ regulatory T cells. *Trends Immunol.* **2004**, *25*, 374–380. [[CrossRef](#)] [[PubMed](#)]
52. Wen, Z.; Shimojima, Y.; Shirai, T.; Li, Y.; Ju, J.; Yang, Z.; Tian, L.; Goronzy, J.J.; Weyand, C.M. NADPH oxidase deficiency underlies dysfunction of aged CD8+ Tregs. *J. Clin. Investig.* **2016**, *126*, 1953–1967. [[CrossRef](#)] [[PubMed](#)]
53. Setoguchi, R.; Hori, S.; Takahashi, T.; Sakaguchi, S. Homeostatic maintenance of natural Foxp3(+) CD25(+) CD4(+) regulatory T cells by interleukin (IL)-2 and induction of autoimmune disease by IL-2 neutralization. *J. Exp. Med.* **2005**, *201*, 723–735. [[CrossRef](#)] [[PubMed](#)]
54. Bayer, A.L.; Yu, A.; Adeegbe, D.; Malek, T.R. Essential role for interleukin-2 for CD4(+)CD25(+) T regulatory cell development during the neonatal period. *J. Exp. Med.* **2005**, *201*, 769–777. [[CrossRef](#)]
55. Fontenot, J.D.; Rudensky, A.Y. A well adapted regulatory contrivance: Regulatory T cell development and the forkhead family transcription factor Foxp3. *Nat. Immunol.* **2005**, *6*, 331–337. [[CrossRef](#)]
56. Malek, T.R. The biology of interleukin-2. *Annu. Rev. Immunol.* **2008**, *26*, 453–479. [[CrossRef](#)]
57. Almeida, A.R.; Zaragoza, B.; Freitas, A.A. Indexation as a novel mechanism of lymphocyte homeostasis: The number of CD4+CD25+ regulatory T cells is indexed to the number of IL-2-producing cells. *J. Immunol.* **2006**, *177*, 192–200. [[CrossRef](#)]
58. Busse, D.; de la Rosa, M.; Hobiger, K.; Thurley, K.; Flossdorf, M.; Scheffold, A.; Hofer, T. Competing feedback loops shape IL-2 signaling between helper and regulatory T lymphocytes in cellular microenvironments. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 3058–3063. [[CrossRef](#)]
59. Sadlack, B.; Merz, H.; Schorle, H.; Schimpl, A.; Feller, A.C.; Horak, I. Ulcerative colitis-like disease in mice with a disrupted interleukin-2 gene. *Cell* **1993**, *75*, 253–261. [[CrossRef](#)]
60. Sadlack, B.; Löhler, J.; Schorle, H.; Klebb, G.; Haber, H.; Sickel, E.; Noelle, R.J.; Horak, I.; Horak, I. Generalized autoimmune disease in interleukin-2-deficient mice is triggered by an uncontrolled activation and proliferation of CD4+ T cells. *Eur. J. Immunol.* **1995**, *25*, 3053–3059. [[CrossRef](#)]
61. Suzuki, H.; Ündig, T.; Furlonger, C.; Wakeham, A.; Timms, E.; Matsuyama, T.; Schmits, R.; Simard, J.J.; Ohashi, P.S.; Griesser, H.; et al. Deregulated T cell activation and autoimmunity in mice lacking interleukin-2 receptor beta. *Science* **1995**, *268*, 1472–1476. [[CrossRef](#)] [[PubMed](#)]
62. Willerford, D.M.; Chen, J.; Ferry, J.A.; Davidson, L.; Ma, A.; Alt, F.W. Interleukin-2 receptor alpha chain regulates the size and content of the peripheral lymphoid compartment. *Immunity* **1995**, *3*, 521–530. [[CrossRef](#)]
63. Caudy, A.A.; Reddy, S.T.; Chatila, T.; Atkinson, J.P.; Verbsky, J.W. CD25 deficiency causes an immune dysregulation, polyendocrinopathy, enteropathy, X-linked-like syndrome, and defective IL-10 expression from CD4 lymphocytes. *J. Allergy Clin. Immunol.* **2007**, *119*, 482–487. [[CrossRef](#)] [[PubMed](#)]
64. Bennett, C.L.; Christie, J.; Ramsdell, F.; Brunkow, M.E.; Ferguson, P.J.; Whitesell, L.; Kelly, T.E.; Saulsbury, F.T.; Chance, P.F.; Ochs, H.D. The immune dysregulation, polyendocrinopathy, enteropathy, X-linked syndrome (IPEX) is caused by mutations of FOXP3. *Nat. Genet.* **2001**, *27*, 20–21. [[CrossRef](#)] [[PubMed](#)]
65. Josefowicz, S.Z.; Niec, R.E.; Kim, H.Y.; Treuting, P.; Chinen, T.; Zheng, Y.; Umetsu, D.T.; Rudensky, A.Y. Extrathymically generated regulatory T cells control mucosal TH2 inflammation. *Nature* **2012**, *482*, 395–399. [[CrossRef](#)]
66. Haribhai, D.; Williams, J.B.; Jia, S.; Nickerson, D.; Schmitt, E.G.; Edwards, B.; Ziegelbauer, J.; Yassai, M.; Li, S.H.; Relland, L.M.; et al. A requisite role for induced regulatory T cells in tolerance based on expanding antigen receptor diversity. *Immunity* **2011**, *35*, 109–122. [[CrossRef](#)]
67. Haribhai, D.; Lin, W.; Edwards, B.; Ziegelbauer, J.; Salzman, N.H.; Carlson, M.R.; Li, S.H.; Simpson, P.M.; Chatila, T.A.; Williams, C.B. A central role for induced regulatory T cells in tolerance induction in experimental colitis. *J. Immunol.* **2009**, *182*, 3461–3468. [[CrossRef](#)]
68. Butler, T.C.; Kardar, M.; Chakraborty, A.K. Quorum sensing allows T cells to discriminate between self and nonself. *Proc. Natl. Acad. Sci. USA* **2013**, *110*, 11833–11838. [[CrossRef](#)]
69. Feinerman, O.; Jentsch, G.; Tkach, K.E.; Coward, J.W.; Hathorn, M.M.; Sneddon, M.W.; Emonet, T.; Smith, K.A.; Altan-Bonnet, G. Single-cell quantification of IL-2 response by effector and regulatory T cells reveals critical plasticity in immune response. *Mol. Syst. Biol.* **2010**, *6*, 437. [[CrossRef](#)]
70. Heufler, C.; Topar, G.; Grasseger, A.; Stanzl, U.; Koch, F.; Romani, N.; Namen, A.E.; Schuler, G. Interleukin 7 is produced by murine and human keratinocytes. *J. Exp. Med.* **1993**, *178*, 1109–1114. [[CrossRef](#)]

71. Fry, T.J.; Mackall, C.L. The many faces of IL-7: From lymphopoiesis to peripheral T cell maintenance. *J. Immunol.* **2005**, *174*, 6571–6576. [[CrossRef](#)] [[PubMed](#)]
72. Link, A.; Vogt, T.K.; Favre, S.; Britschgi, M.R.; Acha-Orbea, H.; Hinz, B.; Cyster, J.G.; Luther, S.A. Fibroblastic reticular cells in lymph nodes regulate the homeostasis of naive T cells. *Nat. Immunol.* **2007**, *8*, 1255–1265. [[CrossRef](#)]
73. Hofmeister, R.; Khaled, A.R.; Benbernou, N.; Rajnavolgyi, E.; Muegge, K.; Durum, S.K. Interleukin-7: Physiological roles and mechanisms of action. *Cytokine Growth Factor Rev.* **1999**, *10*, 41–60. [[CrossRef](#)]
74. Hataye, J.; Moon, J.; Khoruts, A.; Reilly, C.; Jenkins, M.K. Naive and memory CD4+ T cell survival controlled by clonal abundance. *Science* **2006**, *312*, 114–116. [[CrossRef](#)] [[PubMed](#)]
75. Ma, A.; Koka, R.; Burkett, P. Diverse functions of IL-2, IL-15, and IL-7 in lymphoid homeostasis. *Annu. Rev. Immunol.* **2006**, *24*, 657–679. [[CrossRef](#)] [[PubMed](#)]
76. Schluns, K.S.; Stoklasek, T.; Lefrancois, L. The roles of interleukin-15 receptor alpha: Trans-presentation, receptor component, or both? *Int. J. Biochem. Cell. Biol.* **2005**, *37*, 1567–1571. [[CrossRef](#)]
77. Anderson, D.M.; Kumaki, S.; Ahdieh, M.; Bertles, J.; Tometsko, M.; Loomis, A.; Giri, J.; Copeland, N.G.; Gilbert, D.J.; Jenkins, N.A.; et al. Functional characterization of the human interleukin-15 receptor alpha chain and close linkage of IL15RA and IL2RA genes. *J. Biol. Chem.* **1995**, *270*, 29862–29869. [[CrossRef](#)]
78. Colpitts, S.L.; Stonier, S.W.; Stoklasek, T.A.; Root, S.H.; Aguila, H.L.; Schluns, K.S.; Lefrancois, L. Transcriptional regulation of IL-15 expression during hematopoiesis. *J. Immunol.* **2013**, *191*, 3017–3024. [[CrossRef](#)]
79. Seddon, B.; Tomlinson, P.; Zamoyska, R. Interleukin 7 and T cell receptor signals regulate homeostasis of CD4 memory cells. *Nat. Immunol.* **2003**, *4*, 680–686. [[CrossRef](#)]
80. Kondrack, R.M.; Harbertson, J.; Tan, J.T.; McBreen, M.E.; Surh, C.D.; Bradley, L.M. Interleukin 7 regulates the survival and generation of memory CD4 cells. *J. Exp. Med.* **2003**, *198*, 1797–1806. [[CrossRef](#)]
81. Purton, J.F.; Tan, J.T.; Rubinstein, M.P.; Kim, D.M.; Sprent, J.; Surh, C.D. Antiviral CD4+ memory T cells are IL-15 dependent. *J. Exp. Med.* **2007**, *204*, 951–961. [[CrossRef](#)] [[PubMed](#)]
82. Robertson, J.M.; MacLeod, M.; Marsden, V.S.; Kappler, J.W.; Marrack, P. Not all CD4+ memory T cells are long lived. *Immunol. Rev.* **2006**, *211*, 49–57. [[CrossRef](#)] [[PubMed](#)]
83. Lenz, D.C.; Kurz, S.K.; Lemmens, E.; Schoenberger, S.P.; Sprent, J.; Oldstone, M.B.; Homann, D. IL-7 regulates basal homeostatic proliferation of antiviral CD4+T cell memory. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 9357–9362. [[CrossRef](#)]
84. Swain, S.L.; Hu, H.; Huston, G. Class II-independent generation of CD4 memory T cells from effectors. *Science* **1999**, *286*, 1381–1383. [[CrossRef](#)] [[PubMed](#)]
85. Swain, S.L. CD4 T-cell memory can persist in the absence of class II. *Philos. Trans. R. Soc. London. Ser. B Biol. Sci.* **2000**, *355*, 407–411. [[CrossRef](#)] [[PubMed](#)]
86. Murali-Krishna, K.; Lau, L.L.; Sambhara, S.; Lemonnier, F.; Altman, J.; Ahmed, R. Persistence of memory CD8 T cells in MHC class I-deficient mice. *Science* **1999**, *286*, 1377–1381. [[CrossRef](#)]
87. Li, J.; Huston, G.; Swain, S.L. IL-7 promotes the transition of CD4 effectors to persistent memory cells. *J. Exp. Med.* **2003**, *198*, 1807–1815. [[CrossRef](#)]
88. Hahn, M.E.; Karchner, S.; Shapiro, M.A.; Perera, S.A. Molecular evolution of two vertebrate aryl hydrocarbon (dioxin) receptors (AHR1 and AHR2) and the PAS family. *J. Biochem.* **2003**. [[CrossRef](#)]
89. Kikuchi, Y.; Ohsawa, S.; Mimura, J.; Ema, M.; Takasaki, C.; Sogawa, K.; Fujii-Kuriyama, Y. Heterodimers of bHLH-PAS protein fragments derived from AhR, AhRR, and Arnt prepared by co-expression in *Escherichia coli*: Characterization of their DNA binding activity and preparation of a DNA complex. *J. Biochem.* **2003**, *134*, 83–90. [[CrossRef](#)]
90. Rothhammer, V.; Quintana, F.J. The aryl hydrocarbon receptor: An environmental sensor integrating immune responses in health and disease. *Nat. Rev. Immunol.* **2019**, *19*, 184–197. [[CrossRef](#)]
91. Entschladen, F.; Lindquist, J.; Serfling, E.; Thiel, G.; Kieser, A.; Giehl, K.; Ehrhardt, C.; Feller, S.M.; Ullrich, O.; Schaper, F.; et al. Signal transduction-receptors, mediators, and genes. *Cell Commun. Signal* **2009**, *2*, mr3.
92. Shinde, R.; McGaha, T.L. The Aryl Hydrocarbon Receptor: Connecting Immunity to the Microenvironment. *Trends Immunol.* **2018**, *39*, 1005–1020. [[CrossRef](#)] [[PubMed](#)]
93. Minton, K. Intercepting bacterial communications. *Nat. Rev. Immunol.* **2020**, *20*, 138–139. [[CrossRef](#)]
94. Moura-Alves, P.; Puyskens, A.; Stinn, A.; Klemm, M.; Guhlich-Bornhof, U.; Dorhoi, A.; Furkert, J.; Kreuchwig, A.; Protze, J.; Lozza, L.; et al. Host monitoring of quorum sensing during *Pseudomonas aeruginosa* infection. *Science* **2019**, *366*. [[CrossRef](#)]
95. Perdew, G.H.; Babbs, C.F. Production of Ah receptor ligands in rat fecal suspensions containing tryptophan or indole-3-carbinol. *Nutr. Cancer* **1991**, *16*, 209–218. [[CrossRef](#)] [[PubMed](#)]
96. Wattenberg Lw Fau - Loub, W.D.; Loub, W.D. Inhibition of polycyclic aromatic hydrocarbon-induced neoplasia by naturally occurring indoles. *Cancer Res.* **1978**, *38*, 1410–1413.
97. Cheng, Y.; Jin, U.H.; Allred, C.D.; Jayaraman, A.; Chapkin, R.S.; Safe, S. Aryl Hydrocarbon Receptor Activity of Tryptophan Metabolites in Young Adult Mouse Colonocytes. *Drug Metab. Dispos.* **2015**, *43*, 1536–1543. [[CrossRef](#)] [[PubMed](#)]
98. Hubbard, T.D.; Murray, I.A.; Bisson, W.H.; Lahoti, T.S.; Gowda, K.; Amin, S.G.; Patterson, A.D.; Perdew, G.H. Adaptation of the human aryl hydrocarbon receptor to sense microbiota-derived indoles. *Sci. Rep.* **2015**, *5*, 12689. [[CrossRef](#)] [[PubMed](#)]

99. Cervantes-Barragan, L.; Chai, J.N.; Tianero, M.D.; Di Luccia, B.; Ahern, P.P.; Merriman, J.; Cortez, V.S.; Caparon, M.G.; Donia, M.S.; Gilfillan, S.; et al. *Lactobacillus reuteri* induces gut intraepithelial CD4(+)CD8alphaalpha(+) T cells. *Science* **2017**, *357*, 806–810. [[CrossRef](#)]
100. Roager, H.M.; Licht, T.R. Microbial tryptophan catabolites in health and disease. *Nat. Commun.* **2018**, *9*, 3294. [[CrossRef](#)]
101. Zelante, T.; Iannitti, R.G.; Cunha, C.; De Luca, A.; Giovannini, G.; Pieraccini, G.; Zecchi, R.; D'Angelo, C.; Massi-Benedetti, C.; Fallarino, F.; et al. Tryptophan catabolites from microbiota engage aryl hydrocarbon receptor and balance mucosal reactivity via interleukin-22. *Immunity* **2013**, *39*, 372–385. [[CrossRef](#)] [[PubMed](#)]
102. Swimm, A.; Giver, C.R.; DeFilipp, Z.; Rangaraju, S.; Sharma, A.A.-O.; Ulezko Antonova, A.; Sonowal, R.; Capaldo, C.; Powell, D.; Qayed, M.; et al. Indoles derived from intestinal microbiota act via type I interferon signaling to limit graft-versus-host disease. *Blood* **2018**, *132*, 2506–2519. [[CrossRef](#)]
103. Qiu, J.; Guo, X.; Chen, Z.M.; He, L.; Sonnenberg, G.F.; Artis, D.; Fu, Y.X.; Zhou, L. Group 3 innate lymphoid cells inhibit T-cell-mediated intestinal inflammation through aryl hydrocarbon receptor signaling and regulation of microflora. *Immunity* **2013**, *39*, 386–399. [[CrossRef](#)]
104. Qiu, J.; Heller, J.J.; Guo, X.; Chen, Z.M.; Fish, K.; Fu, Y.X.; Zhou, L. The aryl hydrocarbon receptor regulates gut immunity through modulation of innate lymphoid cells. *Immunity* **2012**, *36*, 92–104. [[CrossRef](#)]
105. Lindemans, C.A.; Calafiore, M.; Mertelsmann, A.M.; O'Connor, M.H.; Dudakov, J.A.; Jenq, R.R.; Velardi, E.; Young, L.F.; Smith, O.M.; Lawrence, G.; et al. Interleukin-22 promotes intestinal-stem-cell-mediated epithelial regeneration. *Nature* **2015**, *528*, 560–564. [[CrossRef](#)] [[PubMed](#)]
106. Lamas, B.; Richard, M.L.; Leducq, V.; Pham, H.P.; Michel, M.L.; Da Costa, G.; Bridonneau, C.; Jegou, S.; Hoffmann, T.W.; Natividad, J.M.; et al. CARD9 impacts colitis by altering gut microbiota metabolism of tryptophan into aryl hydrocarbon receptor ligands. *Nat. Med.* **2016**, *22*, 598–605. [[CrossRef](#)]
107. Lee, G.R. The Balance of Th17 versus Treg Cells in Autoimmunity. *Int. J. Mol. Sci.* **2018**, *19*, 730. [[CrossRef](#)]
108. Zi, C.; He, L.; Yao, H.; Ren, Y.; He, T.; Gao, Y. Changes of Th17 cells, regulatory T cells, Treg/Th17, IL-17 and IL-10 in patients with type 2 diabetes mellitus: A systematic review and meta-analysis. *Endocrine* **2022**, *76*, 263–272. [[CrossRef](#)]
109. Lan, Y.T.; Wang, Z.L.; Tian, P.; Gong, X.N.; Fan, Y.C.; Wang, K. Treg/Th17 imbalance and its clinical significance in patients with hepatitis B-associated liver cirrhosis. *Diagn Pathol.* **2019**, *14*, 114. [[CrossRef](#)]
110. Kleinewietfeld, M.; Hafler, D.A. The plasticity of human Treg and Th17 cells and its role in autoimmunity. *Semin. Immunol.* **2013**, *25*, 305–312. [[CrossRef](#)]
111. Yan, J.B.; Luo, M.M.; Chen, Z.Y.; He, B.H. The Function and Role of the Th17/Treg Cell Balance in Inflammatory Bowel Disease. *J. Immunol. Res.* **2020**, *2020*, 8813558. [[CrossRef](#)] [[PubMed](#)]
112. Wang, D.; Zhang, Z.; Si, Z.; Yang, Y.; Li, S.; Xue, Y. Dapagliflozin reverses the imbalance of T helper 17 and T regulatory cells by inhibiting SGK1 in a mouse model of diabetic kidney disease. *FEBS Open Bio* **2021**, *11*, 1395–1405. [[CrossRef](#)] [[PubMed](#)]
113. Elson, C.O.; Cong, Y.; Weaver, C.T.; Schoeb, T.R.; McClanahan, T.K.; Fick, R.B.; Kastelein, R.A. Monoclonal anti-interleukin 23 reverses active colitis in a T cell-mediated model in mice. *Gastroenterology* **2007**, *132*, 2359–2370. [[CrossRef](#)]
114. Tao, L.; Liu, H.; Gong, Y. Role and mechanism of the Th17/Treg cell balance in the development and progression of insulin resistance. *Mol. Cell. Biochem.* **2019**, *459*, 183–188. [[CrossRef](#)] [[PubMed](#)]
115. Kleczynska, W.; Jakiela, B.; Plutecka, H.; Milewski, M.; Sanak, M.; Musial, J. Imbalance between Th17 and regulatory T-cells in systemic lupus erythematosus. *Folia Histochem. Cytobiol.* **2011**, *49*, 646–653. [[CrossRef](#)] [[PubMed](#)]
116. Stockinger, B.; Di Meglio, P.; Gialitakis, M.; Duarte, J.H. The aryl hydrocarbon receptor: Multitasking in the immune system. *Annu. Rev. Immunol.* **2014**, *32*, 403–432. [[CrossRef](#)]
117. Veldhoen, M.; Hirota, K.; Westendorf, A.M.; Buer, J.; Dumoutier, L.; Renauld, J.C.; Stockinger, B. The aryl hydrocarbon receptor links TH17-cell-mediated autoimmunity to environmental toxins. *Nature* **2008**, *453*, 106–109. [[CrossRef](#)]
118. Xie, J.; Wang, Z.; Wang, W. Semaphorin 4D Induces an Imbalance of Th17/Treg Cells by Activating the Aryl Hydrocarbon Receptor in Ankylosing Spondylitis. *Front. Immunol.* **2020**, *11*, 2151. [[CrossRef](#)]
119. McAleer, J.P.; Fan, J.; Roar, B.; Primerano, D.A.; Denvir, J. Cytokine Regulation in Human CD4 T Cells by the Aryl Hydrocarbon Receptor and Gq-Coupled Receptors. *Sci. Rep.* **2018**, *8*, 10954. [[CrossRef](#)]
120. Quintana, F.J.; Basso, A.S.; Iglesias, A.H.; Korn, T.; Farez, M.F.; Bettelli, E.; Caccamo, M.; Oukka, M.; Weiner, H.L. Control of T(reg) and T(H)17 cell differentiation by the aryl hydrocarbon receptor. *Nature* **2008**, *453*, 65–71. [[CrossRef](#)]
121. Talbot, J.; Peres, R.S.; Pinto, L.G.; Oliveira, R.D.R.; Lima, K.A.; Donate, P.B.; Silva, J.R.; Ryffel, B.; Cunha, T.M.; Alves-Filho, J.C.; et al. Smoking-induced aggravation of experimental arthritis is dependent of aryl hydrocarbon receptor activation in Th17 cells. *Arthritis Res. Ther.* **2018**, *20*, 119. [[CrossRef](#)] [[PubMed](#)]
122. Kimura, A.; Naka, T.; Nohara, K.; Fujii-Kuriyama, Y.; Kishimoto, T. Aryl hydrocarbon receptor regulates Stat1 activation and participates in the development of Th17 cells. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 9721–9726. [[CrossRef](#)] [[PubMed](#)]
123. Romero, F.; Martínez, A.C.; Camonis, J.; Rebollo, A. Aiolos transcription factor controls cell death in T cells by regulating Bcl-2 expression and its cellular localization. *EMBO J* **1999**, *18*, 3419–3430. [[CrossRef](#)] [[PubMed](#)]
124. Quintana, F.J.; Jin, H.; Burns, E.J.; Nadeau, M.; Yeste, A.; Kumar, D.; Rangachari, M.; Zhu, C.; Xiao, S.; Seavitt, J.; et al. Aiolos promotes TH17 differentiation by directly silencing Il2 expression. *Nat. Immunol.* **2012**, *13*, 770–777. [[CrossRef](#)] [[PubMed](#)]
125. Whiteside, S.K.; Snook, J.P.; Williams, M.A.; Weis, J.J. Bystander T Cells: A Balancing Act of Friends and Foes. *Trends Immunol.* **2018**, *39*, 1021–1035. [[CrossRef](#)]

126. Pacheco, Y.; Acosta-Ampudia, Y.; Monsalve, D.M.; Chang, C.; Gershwin, M.E.; Anaya, J.M. Bystander activation and autoimmunity. *J. Autoimmun.* **2019**, *103*, 102301. [[CrossRef](#)]
127. Maurice, N.J.; Taber, A.K.; Prlc, M. The Ugly Duckling Turned to Swan: A Change in Perception of Bystander-Activated Memory CD8 T Cells. *J. Immunol.* **2021**, *206*, 455–462. [[CrossRef](#)]
128. Tough, D.F.; Borrow, P.; Sprent, J. Induction of bystander T cell proliferation by viruses and type I interferon in vivo. *Science* **1996**, *272*, 1947–1950. [[CrossRef](#)]
129. Zarozinski, C.C.; Welsh, R.M. Minimal bystander activation of CD8 T cells during the virus-induced polyclonal T cell response. *J. Exp. Med.* **1997**, *185*, 1629–1639. [[CrossRef](#)]
130. Murali-Krishna, K.; Altman, J.D.; Suresh, M.; Sourdive, D.J.; Zajac, A.J.; Miller, J.D.; Slansky, J.; Ahmed, R. Counting antigen-specific CD8 T cells: A reevaluation of bystander activation during viral infection. *Immunity* **1998**, *8*, 177–187. [[CrossRef](#)]
131. Montoya, D.; Cruz, D.; Teles, R.M.; Lee, D.J.; Ochoa, M.T.; Krutzik, S.R.; Chun, R.; Schenk, M.; Zhang, X.; Ferguson, B.G.; et al. Divergence of macrophage phagocytic and antimicrobial programs in leprosy. *Cell Host. Microbe* **2009**, *6*, 343–353. [[CrossRef](#)] [[PubMed](#)]
132. Hussell, T.; Bell, T.J. Alveolar macrophages: Plasticity in a tissue-specific context. *Nat. Rev. Immunol.* **2014**, *14*, 81–93. [[CrossRef](#)] [[PubMed](#)]
133. Williams, M.; De Kleer, I.; Henri, S.; Post, S.; Vanhoutte, L.; De Prijck, S.; Deswarte, K.; Malissen, B.; Hammad, H.; Lambrecht, B.N. Alveolar macrophages develop from fetal monocytes that differentiate into long-lived cells in the first week of life via GM-CSF. *J. Exp. Med.* **2013**, *210*, 1977–1992. [[CrossRef](#)]
134. Hoeffel, G.; Wang, Y.; Greter, M.; See, P.; Teo, P.; Malleret, B.; Leboeuf, M.; Low, D.; Oller, G.; Almeida, F.; et al. Adult Langerhans cells derive predominantly from embryonic fetal liver monocytes with a minor contribution of yolk sac-derived macrophages. *J. Exp. Med.* **2012**, *209*, 1167–1181. [[CrossRef](#)] [[PubMed](#)]
135. Christian Schulz, E.G.P.; Chorro, L.; Szabo-Rogers, H.; Cagnard, N.; Kierdorf, K.; Prinz, M.; Wu, B.; Eirik, S.; Jacobsen, W.; Frampton, J.; et al. A Lineage of Myeloid Cells Independent of Myb and Hematopoietic Stem Cells. *Science* **2012**, *336*, 86–90. [[CrossRef](#)] [[PubMed](#)]
136. Ginhoux, F.; Greter, M.; Leboeuf, M.; Nandi, S.; See, P.; Gokhan, S.; Mehler, M.F.; Conway, S.J.; Ng, L.G.; Stanley, E.R.; et al. Fate mapping analysis reveals that adult microglia derive from primitive macrophages. *Science* **2010**, *330*, 841–845. [[CrossRef](#)]
137. Mosser, D.M.; Edwards, J.P. Exploring the full spectrum of macrophage activation. *Nat. Rev. Immunol.* **2008**, *8*, 958–969. [[CrossRef](#)]
138. Benoit, M.; Desnues, B.; Mege, J.L. Macrophage polarization in bacterial infections. *J. Immunol.* **2008**, *181*, 3733–3739. [[CrossRef](#)]
139. Orecchioni, M.; Ghosheh, Y.; Pramod, A.B.; Ley, K. Macrophage Polarization: Different Gene Signatures in M1(LPS+) vs. Classically and M2(LPS-) vs. Alternatively Activated Macrophages. *Front. Immunol.* **2019**, *10*, 1084. [[CrossRef](#)]
140. Scott, C.L.; Zheng, F.; De Baetselier, P.; Martens, L.; Saeys, Y.; De Prijck, S.; Lippens, S.; Abels, C.; Schoonoghe, S.; Raes, G.; et al. Bone marrow-derived monocytes give rise to self-renewing and fully differentiated Kupffer cells. *Nat. Commun.* **2016**, *7*, 10321. [[CrossRef](#)]
141. Zigmund, E.; Samia-Grinberg, S.; Pasmanik-Chor, M.; Brazowski, E.; Shibolet, O.; Halpern, Z.; Varol, C. Infiltrating monocyte-derived macrophages and resident kupffer cells display different ontogeny and functions in acute liver injury. *J. Immunol.* **2014**, *193*, 344–353. [[CrossRef](#)] [[PubMed](#)]
142. Medina, C.B.; Mehrotra, P.; Arandjelovic, S.; Perry, J.S.A.; Guo, Y.; Morioka, S.; Barron, B.; Walk, S.F.; Ghesquiere, B.; Krupnick, A.S.; et al. Metabolites released from apoptotic cells act as tissue messengers. *Nature* **2020**, *580*, 130–135. [[CrossRef](#)] [[PubMed](#)]
143. Baxter, A.A.; Poon, I.K. Apoptotic cells secrete metabolites to regulate immune homeostasis. *Immunol. Cell Biol.* **2020**, *98*, 355–357. [[CrossRef](#)] [[PubMed](#)]
144. Dunkle, A.; He, Y.W. Apoptosis and autophagy in the regulation of T lymphocyte function. *Immunol. Res* **2011**, *49*, 70–86. [[CrossRef](#)]
145. Birk, A.V.; Broekman, M.J.; Gladek, E.M.; Robertson, H.D.; Drosopoulos, J.H.; Marcus, A.J.; Szeto, H.H. Role of extracellular ATP metabolism in regulation of platelet reactivity. *J. Lab. Clin. Med.* **2002**, *140*, 166–175. [[CrossRef](#)]
146. Yip, L.; Woehrl, T.; Corriden, R.; Hirsh, M.; Chen, Y.; Inoue, Y.; Ferrari, V.; Insel, P.A.; Junger, W.G. Autocrine regulation of T-cell activation by ATP release and P2 × 7 receptors. *FASEB J.* **2009**, *23*, 1685–1693. [[CrossRef](#)]
147. Winoto, A. Cell death in the regulation of immune responses. *Curr. Opin. Immunol.* **1997**, *9*, 365–370. [[CrossRef](#)]
148. Ganeshan, K.; Chawla, A. Metabolic regulation of immune responses. *Annu. Rev. Immunol.* **2014**, *32*, 609–634. [[CrossRef](#)]
149. van Rossum, D.; Hanisch, U.K. Microglia. *Metab. Brain Dis.* **2004**, *19*, 393–411. [[CrossRef](#)]
150. Amici, S.A.; Dong, J.; Guerau-de-Arellano, M. Molecular Mechanisms Modulating the Phenotype of Macrophages and Microglia. *Front. Immunol.* **2017**, *8*, 1520. [[CrossRef](#)]
151. DePaula-Silva, A.B.; Gorbea, C.; Doty, D.J.; Libbey, J.E.; Sanchez, J.M.S.; Hanak, T.J.; Cazalla, D.; Fujinami, R.S. Differential transcriptional profiles identify microglial- and macrophage-specific gene markers expressed during virus-induced neuroinflammation. *J. Neuroinflamm.* **2019**, *16*, 152. [[CrossRef](#)] [[PubMed](#)]
152. Eichhoff, G.; Brawek, B.; Garaschuk, O. Microglial calcium signal acts as a rapid sensor of single neuron damage in vivo. *Biochim. Biophys. Acta* **2011**, *1813*, 1014–1024. [[CrossRef](#)]
153. Zarruk, J.G.; Greenhalgh, A.D.; David, S. Microglia and macrophages differ in their inflammatory profile after permanent brain ischemia. *Exp. Neurol.* **2018**, *301*, 120–132. [[CrossRef](#)] [[PubMed](#)]

154. Yin, J.; Valin, K.L.; Dixon, M.L.; Leavenworth, J.W. The Role of Microglia and Macrophages in CNS Homeostasis, Autoimmunity, and Cancer. *J. Immunol. Res.* **2017**, *2017*, 5150678. [[CrossRef](#)] [[PubMed](#)]
155. Tay, T.L.; Mai, D.; Dautzenberg, J.; Fernandez-Klett, F.; Lin, G.; Sagar, Datta, M.; Drougard, A.; Stempf, T.; Ardura-Fabregat, A.; et al. A new fate mapping system reveals context-dependent random or clonal expansion of microglia. *Nat. Neurosci.* **2017**, *20*, 793–803. [[CrossRef](#)]
156. Boechat, N.; Bouchonnet, F.; Bonay, M.; Grodet, A.; Pelicic, V.; Gicquel, B.; Hance, A.J. Culture at high density improves the ability of human macrophages to control mycobacterial growth. *J. Immunol.* **2001**, *166*, 6203–6211. [[CrossRef](#)] [[PubMed](#)]
157. Chen, C.C.; Wang, L.; Plikus, M.V.; Jiang, T.X.; Murray, P.J.; Ramos, R.; Guerrero-Juarez, C.F.; Hughes, M.W.; Lee, O.K.; Shi, S.; et al. Organ-level quorum sensing directs regeneration in hair stem cell populations. *Cell* **2015**, *161*, 277–290. [[CrossRef](#)]
158. Muldoon, J.J.; Chuang, Y.; Bagheri, N.; Leonard, J.N. Macrophages employ quorum licensing to regulate collective activation. *Nat. Commun.* **2020**, *11*, 878. [[CrossRef](#)]
159. Postat, J.; Bousso, P. Quorum Sensing by Monocyte-Derived Populations. *Front. Immunol.* **2019**, *10*, 2140. [[CrossRef](#)]
160. Xu, D.; Andrew, R.G.R.; Hapel, J.; Melissa, G.; Dominguez, R.; Russell, G.; Sara Kapp, V.S.; Stanley, E.R. Targeted disruption of the mouse colony-stimulating factor 1 receptor gene results in osteopetrosis, mononuclear phagocyte deficiency, increased primitive progenitor cell frequencies, and reproductive defects. *Blood* **2002**, *99*, 111–120.
161. Groh, J.; Weis, J.; Zieger, H.; Stanley, E.R.; Heuer, H.; Martini, R. Colony-stimulating factor-1 mediates macrophage-related neural damage in a model for Charcot-Marie-Tooth disease type 1X. *Brain* **2012**, *135*, 88–104. [[CrossRef](#)] [[PubMed](#)]
162. Lenzo, J.C.; Turner, A.L.; Cook, A.D.; Vlahos, R.; Anderson, G.P.; Reynolds, E.C.; Hamilton, J.A. Control of macrophage lineage populations by CSF-1 receptor and GM-CSF in homeostasis and inflammation. *Immunol. Cell Biol.* **2012**, *90*, 429–440. [[CrossRef](#)] [[PubMed](#)]
163. Nandi, S.; Akhter, M.P.; Seifert, M.F.; Dai, X.M.; Stanley, E.R. Developmental and functional significance of the CSF-1 proteoglycan chondroitin sulfate chain. *Blood* **2006**, *107*, 786–795. [[CrossRef](#)]
164. Groh, J.; Basu, R.; Stanley, E.R.; Martini, R. Cell-Surface and Secreted Isoforms of CSF-1 Exert Opposing Roles in Macrophage-Mediated Neural Damage in Cx32-Deficient Mice. *J. Neurosci. Off. J. Soc. Neurosci.* **2016**, *36*, 1890–1901. [[CrossRef](#)] [[PubMed](#)]
165. Martin, F.P.; Jacqueline, C.; Poschmann, J.; Roquilly, A. Alveolar Macrophages: Adaptation to Their Anatomic Niche during and after Inflammation. *Cells* **2021**, *10*, 2720. [[CrossRef](#)] [[PubMed](#)]
166. Ahmad, F.; Rani, A.; Alam, A.; Zarin, S.; Pandey, S.; Singh, H.; Hasnain, S.E.; Ehtesham, N.Z. Macrophage: A Cell With Many Faces and Functions in Tuberculosis. *Front. Immunol.* **2022**, *13*, 747799. [[CrossRef](#)] [[PubMed](#)]
167. Silva, M.T.; Pestana, N.T. The in vivo extracellular life of facultative intracellular bacterial parasites: Role in pathogenesis. *Immunobiology* **2013**, *218*, 325–337. [[CrossRef](#)]
168. Srinivasan, L.; Ahlbrand, S.; Briken, V. Interaction of Mycobacterium tuberculosis with host cell death pathways. *Cold Spring Harb. Perspect. Med.* **2014**, *4*, a022459. [[CrossRef](#)]
169. Behar, S.M.; Martin, C.; Booty, M.; Nishimura, T.; Zhao, X.; Gan, H.X.; Divangahi, M.; Remold, H.G.; Remold, H.G. Apoptosis is an innate defense function of macrophages against Mycobacterium tuberculosis. *Mucosal Immunol.* **2011**, *4*, 279–287. [[CrossRef](#)]
170. Keane, J.; Remold, H.; Kornfeld, H. Virulent Mycobacterium tuberculosis strains evade apoptosis of infected alveolar macrophages. *J. Immunol.* **2000**, *164*, 2016–2020. [[CrossRef](#)]
171. Danelishvili, L.; McGarvey, J.; Li, Y.; Bermudez, L.E. Mycobacterium tuberculosis infection causes different levels of apoptosis and necrosis in human macrophages and alveolar epithelial cells. *Cell Microbiol.* **2003**, *5*, 649–660. [[CrossRef](#)] [[PubMed](#)]
172. Afriyie-Asante, A.; Dabla, A.; Dagenais, A.; Berton, S.; Smyth, R.; Sun, J. Mycobacterium tuberculosis Exploits Focal Adhesion Kinase to Induce Necrotic Cell Death and Inhibit Reactive Oxygen Species Production. *Front. Immunol.* **2021**, *12*, 742370. [[CrossRef](#)] [[PubMed](#)]
173. Lerner, T.R.; Borel, S.A.-O.X.; Greenwood, D.A.-O.X.; Repnik, U.; Russell, M.A.-O.; Herbst, S.; Jones, M.A.-O.; Collinson, L.M.; Griffiths, G.; Gutierrez, M.A.-O. Mycobacterium tuberculosis replicates within necrotic human macrophages. *J. Cell Biol.* **2017**, *216*, 583–594. [[CrossRef](#)] [[PubMed](#)]
174. Deng, J.H.; Chen, H.Y.; Huang, C.; Yan, J.M.; Yin, Z.; Zhang, X.L.; Pan, Q. Accumulation of EBI3 induced by virulent Mycobacterium tuberculosis inhibits apoptosis in murine macrophages. *Pathog Dis.* **2019**, *77*, ftz007. [[CrossRef](#)] [[PubMed](#)]
175. Longhi, M.P.; Trumpfheller, C.; Idoyaga, J.; Caskey, M.; Matos, I.; Kluger, C.; Salazar, A.M.; Colonna, M.; Steinman, R.M. Dendritic cells require a systemic type I interferon response to mature and induce CD4+ Th1 immunity with poly IC as adjuvant. *J. Exp. Med.* **2009**, *206*, 1589–1602. [[CrossRef](#)]
176. Bardou, M.; Postat, J.; Loaec, C.; Lemaitre, F.; Ronteix, G.; Garcia, Z.; Bousso, P. Quorum sensing governs collective dendritic cell activation in vivo. *EMBO J.* **2021**, *40*, e107176. [[CrossRef](#)]
177. Celli, S.; Day, M.; Müller, A.J.; Molina-Paris, C.; Lythe, G.; Bousso, P. How many dendritic cells are required to initiate a T-cell response? *Blood* **2012**, *120*, 3945–3948. [[CrossRef](#)]
178. Xue, Q.; Lu, Y.; Eisele, M.R.; Sulistijo, E.S.; Khan, N.; Fan, R.; Miller-Jensen, K. Analysis of single-cell cytokine secretion reveals a role for paracrine signaling in coordinating macrophage responses to TLR4 stimulation. *Sci. Signal* **2015**, *8*, ra59. [[CrossRef](#)]
179. Peterson, L.B.; Bell, C.J.M.; Howlett, S.K.; Pekalski, M.L.; Brady, K.; Hinton, H.; Sauter, D.; Todd, J.A.; Umana, P.; Ast, O.; et al. A long-lived IL-2 mutein that selectively activates and expands regulatory T cells as a therapy for autoimmune disease. *J. Autoimmun.* **2018**, *95*, 1–14. [[CrossRef](#)]
180. Mullard, A. Restoring IL-2 to its cancer immunotherapy glory. *Nat. Rev. Drug Discov.* **2021**, *20*, 163–165. [[CrossRef](#)]

181. Tahvildari, M.; Dana, R. Low-Dose IL-2 Therapy in Transplantation, Autoimmunity, and Inflammatory Diseases. *J. Immunol.* **2019**, *203*, 2749–2755. [[CrossRef](#)] [[PubMed](#)]
182. Churlaud, G.; Abbara, C.; Vinot, P.A.; Fourcade, G.; Ritvo, P.G.; Lorenzon, R.; Rosenzweig, M.; Diquet, B.; Klatzmann, D. Pharmacodynamics of regulatory T cells in mice and humans treated with low-dose IL-2. *J. Allergy Clin. Immunol.* **2018**, *142*, 1344–1346.e3. [[CrossRef](#)]
183. Yu, A.; Snowwhite, I.; Vendrame, F.; Rosenzweig, M.; Klatzmann, D.; Pugliese, A.; Malek, T.R. Selective IL-2 responsiveness of regulatory T cells through multiple intrinsic mechanisms supports the use of low-dose IL-2 therapy in type 1 diabetes. *Diabetes* **2015**, *64*, 2172–2183. [[CrossRef](#)] [[PubMed](#)]
184. Klatzmann, D.; Abbas, A.K. The promise of low-dose interleukin-2 therapy for autoimmune and inflammatory diseases. *Nat. Rev. Immunol.* **2015**, *15*, 283–294. [[CrossRef](#)] [[PubMed](#)]
185. Saadoun, D.; Rosenzweig, M.; Joly, F.; Six, A.; Carrat, F.; Thibault, V.; Sene, D.; Cacoub, P.; Klatzmann, D. Regulatory T-cell responses to low-dose interleukin-2 in HCV-induced vasculitis. *N. Engl. J. Med.* **2011**, *365*, 2067–2077. [[CrossRef](#)]
186. Baeyens, A.; Perol, L.; Fourcade, G.; Cagnard, N.; Carpentier, W.; Woytschak, J.; Boyman, O.; Hartemann, A.; Piaggio, E. Limitations of IL-2 and rapamycin in immunotherapy of type 1 diabetes. *Diabetes* **2013**, *62*, 3120–3131. [[CrossRef](#)]
187. Rosenzweig, M.; Churlaud, G.; Mallone, R.; Six, A.; Derian, N.; Chaara, W.; Lorenzon, R.; Long, S.A.; Buckner, J.H.; Afonso, G.; et al. Low-dose interleukin-2 fosters a dose-dependent regulatory T cell tuned milieu in T1D patients. *J. Autoimmun.* **2015**, *58*, 48–58. [[CrossRef](#)]
188. Humrich, J.Y.; von Spee-Mayer, C.; Siegert, E.; Alexander, T.; Hiepe, F.; Radbruch, A.; Burmester, G.R.; Riemekasten, G. Rapid induction of clinical remission by low-dose interleukin-2 in a patient with refractory SLE. *Ann. Rheum. Dis.* **2015**, *74*, 791–792. [[CrossRef](#)]
189. He, J.; Zhang, X.; Wei, Y.; Sun, X.; Chen, Y.; Deng, J.; Jin, Y.; Gan, Y.; Hu, X.; Jia, R.; et al. Low-dose interleukin-2 treatment selectively modulates CD4(+) T cell subsets in patients with systemic lupus erythematosus. *Nat. Med.* **2016**, *22*, 991–993. [[CrossRef](#)]
190. Castela, E.; Le Duff, F.; Butori, C.; Tichioni, M.; Hofman, P.; Bahadoran, P.; Lacour, J.P.; Passeron, T. Effects of low-dose recombinant interleukin 2 to promote T-regulatory cells in alopecia areata. *JAMA Derm.* **2014**, *150*, 748–751. [[CrossRef](#)]
191. Kennedy-Nasser, A.A.; Ku, S.; Castillo-Caro, P.; Hazrat, Y.; Wu, M.F.; Liu, H.; Melenhorst, J.; Barrett, A.J.; Ito, S.; Foster, A.; et al. Ultra low-dose IL-2 for GVHD prophylaxis after allogeneic hematopoietic stem cell transplantation mediates expansion of regulatory T cells without diminishing antiviral and antileukemic activity. *Clin. Cancer Res.* **2014**, *20*, 2215–2225. [[CrossRef](#)] [[PubMed](#)]
192. Koreth, J.; Matsuoka, K.; Kim, H.T.; McDonough, S.M.; Bindra, B.; Alyea, E.P., 3rd; Armand, P.; Cutler, C.; Ho, V.T.; Treister, N.S.; et al. Interleukin-2 and regulatory T cells in graft-versus-host disease. *N. Engl. J. Med.* **2011**, *365*, 2055–2066. [[CrossRef](#)] [[PubMed](#)]
193. Kim, N.; Jeon, Y.W.; Nam, Y.S.; Lim, J.Y.; Im, K.I.; Lee, E.S.; Cho, S.G. Therapeutic potential of low-dose IL-2 in a chronic GVHD patient by in vivo expansion of regulatory T cells. *Cytokine* **2016**, *78*, 22–26. [[CrossRef](#)] [[PubMed](#)]
194. Matsuoka, K.; Koreth, J.; Kim, H.T.; Bascug, G.; McDonough, S.; Kawano, Y.; Murase, K.; Cutler, C.; Ho, V.T.; Alyea, E.P.; et al. Low-dose interleukin-2 therapy restores regulatory T cell homeostasis in patients with chronic graft-versus-host disease. *Sci. Transl. Med.* **2013**, *5*, 179ra143. [[CrossRef](#)]
195. Zorn, E.; Mohseni, M.; Kim, H.; Porcheray, F.; Lynch, A.; Bellucci, R.; Canning, C.; Alyea, E.P.; Soiffer, R.J.; Ritz, J. Combined CD4+ donor lymphocyte infusion and low-dose recombinant IL-2 expand FOXP3+ regulatory T cells following allogeneic hematopoietic stem cell transplantation. *Biol. Blood Marrow Transpl.* **2009**, *15*, 382–388. [[CrossRef](#)]
196. Bonnet, B.; Vigneron, J.; Levacher, B.; Vazquez, T.; Pitoiset, F.; Brimaud, F.; Churlaud, G.; Klatzmann, D.; Bellier, B. Low-Dose IL-2 Induces Regulatory T Cell-Mediated Control of Experimental Food Allergy. *J. Immunol.* **2016**, *197*, 188–198. [[CrossRef](#)]
197. Sarkar, S.; Horn, G.; Moulton, K.; Oza, A.; Byler, S.; Kokolus, S.; Longacre, M. Cancer development, progression, and therapy: An epigenetic overview. *Int. J. Mol. Sci.* **2013**, *14*, 21087–21113. [[CrossRef](#)]
198. Kaye, S.B. New antimetabolites in cancer chemotherapy and their clinical impact. *Br. J. Cancer* **1998**, *78* (Suppl. 3), 1–7. [[CrossRef](#)]
199. Ahmadzadeh, M.; Rosenberg, S.A. IL-2 administration increases CD4+ CD25(hi) Foxp3+ regulatory T cells in cancer patients. *Blood* **2006**, *107*, 2409–2414. [[CrossRef](#)]
200. Klapper, J.A.; Downey, S.G.; Smith, F.O.; Yang, J.C.; Hughes, M.S.; Kammula, U.S.; Sherry, R.M.; Royal, R.E.; Steinberg, S.M.; Rosenberg, S. High-dose interleukin-2 for the treatment of metastatic renal cell carcinoma: A retrospective analysis of response and survival in patients treated in the surgery branch at the National Cancer Institute between 1986 and 2006. *Cancer* **2008**, *113*, 293–301. [[CrossRef](#)]
201. Buchbinder, E.I.; Dutcher, J.P.; Daniels, G.A.; Curti, B.D.; Patel, S.P.; Holtan, S.G.; Miletello, G.P.; Fishman, M.N.; Gonzalez, R.; Clark, J.I.; et al. Therapy with high-dose Interleukin-2 (HD IL-2) in metastatic melanoma and renal cell carcinoma following PD1 or PDL1 inhibition. *J. Immunother. Cancer* **2019**, *7*, 49. [[CrossRef](#)]
202. Davar, D.; Ding, F.; Saul, M.; Sander, C.; Tarhini, A.A.; Kirkwood, J.M.; Tawbi, H.A. High-dose interleukin-2 (HD IL-2) for advanced melanoma: A single center experience from the University of Pittsburgh Cancer Institute. *J. Immunother. Cancer* **2017**, *5*, 74. [[CrossRef](#)] [[PubMed](#)]
203. Gaffen, S.L.; Liu, K.D. Overview of interleukin-2 function, production and clinical applications. *Cytokine* **2004**, *28*, 109–123. [[CrossRef](#)] [[PubMed](#)]

204. Moon, B.I.; Kim, T.H.; Seoh, J.Y. Functional Modulation of Regulatory T Cells by IL-2. *PLoS ONE* **2015**, *10*, e0141864. [[CrossRef](#)]
205. Dockery, J.D.; Keener, J.P. A mathematical model for quorum sensing in *Pseudomonas aeruginosa*. *Bull. Math. Biol.* **2001**, *63*, 95–116. [[CrossRef](#)]
206. Vital-Lopez, F.G.; Reifman, J.; Wallqvist, A. Biofilm Formation Mechanisms of *Pseudomonas aeruginosa* Predicted via Genome-Scale Kinetic Models of Bacterial Metabolism. *PLoS Comput. Biol.* **2015**, *11*, e1004452. [[CrossRef](#)]
207. Almeida, A.R.; Amado, I.F.; Reynolds, J.; Berges, J.; Lythe, G.; Molina-París, C.; Freitas, A.A. Quorum-Sensing in CD4(+) T Cell Homeostasis: A Hypothesis and a Model. *Front. Immunol.* **2012**, *3*, 125. [[CrossRef](#)] [[PubMed](#)]
208. Reynolds, J.; Amado, I.F.; Freitas, A.A.; Lythe, G.; Molina-París, C. A mathematical perspective on CD4(+) T cell quorum-sensing. *J. Biol.* **2014**, *347*, 160–175. [[CrossRef](#)]
209. Schrom, E.C., 2nd; Levin, S.A.; Graham, A.L. Quorum sensing via dynamic cytokine signaling comprehensively explains divergent patterns of effector choice among helper T cells. *PLoS Comput. Biol.* **2020**, *16*, e1008051. [[CrossRef](#)]