

Editorial

# Mesenchymal Stromal Cells “Think” Globally, but Act Locally

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In this Special Issue of *Cells*, entitled “Immunomodulation by Mesenchymal Stem Cells 2020”, you can find five excellent papers on the role of mesenchymal stem/stromal cells (MSCs) in immunomodulation, which also includes regenerative processes, such as wound healing.

MSCs are fibroblast-like multipotent progenitor cells with immunosuppressive properties *in vitro* and *in vivo* [1]. MSCs are increasingly being used in clinical trials, e.g., for chronic graft-versus-host disease following haematopoietic stem cell transplantation [2]. Although the precise mechanism of action (MOA) of these cells in various clinical applications is not fully understood, recent progress has been made in the functional characterisation of MSCs and their extracellular vesicles, especially in the perinatal field [3–6], which can easily be translated to adult MSCs.

MSCs cannot only suppress immune responses, but also induce them, e.g., humoral immunity. In this Special Issue, Bikorimana et al. describe that an immunoproteasome complex expressing MSCs could produce antibodies in response to ovalbumin stimulation, which has an antitumour function *in vivo* [7]. Of note, humoral immunity by MSCs requires the presence of phagocytes providing efferocytosis [7]. Another focus of this Special Issue lies in adipose tissue-derived MSCs (ASCs). A very good example for a local action of MSCs is given by Kuca-Warnawin et al., who point out that ASCs from ankylosing spondylitis patients could—when co-cultured with peripheral blood mononuclear cells—enhance the generation of anti-inflammatory regulatory T cells (Treg) and could downregulate interferon  $\gamma$  expression [8]. Two more papers on ASCs from Karen Bieback’s group also address Treg generation and its MOA. Fiori et al. report that ASCs generally inhibit CD4+ T cell proliferation through the local action of indoleamine 2,3-dioxygenase (IDO), but could stimulate the differentiation between CD127+ and FoxP3+ Treg at the same time. Importantly, ASCs only had these effects when co-cultured with unstimulated PBMC [9]. In an important comparison of MSCs from different sources, Torres Crigna et al. found ASCs to be superior in their T-cell antiproliferative effect to MSCs from bone marrow or cord blood [10]. Interestingly, only conditioned medium, but not EV from ASCs, could exert similar effects. Again, IDO was identified to be responsible for this type of immunomodulation. Furthermore, human ASCs could not inhibit murine lymphocyte proliferation, bringing into question interspecies experimental set-ups for the functional testing of MSCs in general [10]. The final addition to this Special Issue of *Cells* is a comprehensive review by Raquel Guillamat-Prats about the role of MSCs in wound healing, with a focus on diabetes and fibrosis. Of note, MSCs and their derivatives have different functions in different steps of the wound repair process [11].

In this Special Issue of *Cells*, we have learned about the critical role of IDO in the mechanism of MSC action and how important the fine-tuning of local MSC actions is for an effective therapeutic approach. Further work is needed to elaborate how MSC derivatives can be used in the clinic.

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## References

1. Liu, S.; Liu, F.; Zhou, Y.; Jin, B.; Sun, Q.; Guo, S. Immunosuppressive Property of MSCs Mediated by Cell Surface Receptors. *Front. Immunol.* **2020**, *11*, 1076. [[CrossRef](#)]
2. Doglio, M.; Crossland, R.E.; Alho, A.C.; Penack, O.; Dickinson, A.M.; Stary, G.; Lacerda, J.F.; Eissner, G.; Inngjerdingen, M. Cell-based therapy in prophylaxis and treatment of chronic graft-versus-host disease. *Front. Immunol.* **2022**, *13*, 1045168. [[CrossRef](#)]
3. Papait, A.; Silini, A.R.; Gazouli, M.; Malvicini, R.; Muraca, M.; O'Driscoll, L.; Pacienza, N.; Toh, W.S.; Yannarelli, G.; Ponsaerts, P.; et al. Perinatal derivatives: How to best validate their immunomodulatory functions. *Front. Bioeng. Biotechnol.* **2022**, *10*, 981061.
4. Silini, A.R.; Ramuta, T.Z.; Pires, A.S.; Banerjee, A.; Dubus, M.; Gindraux, F.; Kerdjoudj, H.; Maciulatis, J.; Weidinger, A.; Wolbank, S.; et al. Methods and criteria for validating the multimodal functions of perinatal derivatives when used in oncological and antimicrobial applications. *Front. Bioeng. Biotechnol.* **2022**, *10*, 958669.
5. Flores, A.I.; Pipino, C.; Jerman, U.D.; Liarte, S.; Gindraux, F.; Kreft, M.E.; Nicolas, F.J.; Pandolfi, A.; Tratnjek, L.; Giebel, B.; et al. Perinatal derivatives: How to best characterize their multimodal functions in vitro. Part C: Inflammation, angiogenesis, and wound healing. *Front. Bioeng. Biotechnol.* **2022**, *10*, 965006. [[CrossRef](#)]
6. Pozzobon, M.; D'Agostino, S.; Roubelakis, M.G.; Cargnoni, A.; Gramignoli, R.; Wolbank, S.; Gindraux, F.; Bollini, S.; Kerdjoudj, H.; Fenelon, M.; et al. General consensus on multimodal functions and validation analysis of perinatal derivatives for regenerative medicine applications. *Front. Bioeng. Biotechnol.* **2022**, *10*, 961987. [[CrossRef](#)]
7. Bikorimana, J.P.; Abusarah, J.; Salame, N.; El-Hachem, N.; Shammaa, R.; Rafei, M. Humoral Immunity to Allogeneic Immunoproteasome-Expressing Mesenchymal Stromal Cells Requires Efferocytosis by Endogenous Phagocytes. *Cells* **2022**, *11*, 596. [[CrossRef](#)]
8. Kuca-Warnawin, E.; Janicka, I.; Bonek, K.; Kontny, E. Modulatory Impact of Adipose-Derived Mesenchymal Stem Cells of Ankylosing Spondylitis Patients on T Helper Cell Differentiation. *Cells* **2021**, *10*, 280. [[CrossRef](#)]
9. Fiori, A.; Uhlig, S.; Kluter, H.; Bieback, K. Human Adipose Tissue-Derived Mesenchymal Stromal Cells Inhibit CD4+ T Cell Proliferation and Induce Regulatory T Cells as Well as CD127 Expression on CD4+CD25+ T Cells. *Cells* **2021**, *10*, 58.
10. Torres Crigna, A.; Uhlig, S.; Elvers-Hornung, S.; Kluter, H.; Bieback, K. Human Adipose Tissue-Derived Stromal Cells Suppress Human, but Not Murine Lymphocyte Proliferation, via Indoleamine 2,3-Dioxygenase Activity. *Cells* **2020**, *9*, 2419. [[CrossRef](#)]
11. Guillamat-Prats, R. The Role of MSC in Wound Healing, Scarring and Regeneration. *Cells* **2021**, *10*, 1729. [[CrossRef](#)]

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