

Review

Crosstalk between Oxidative Stress and Aging in Neurodegeneration Disorders

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Abstract: The world population is aging rapidly, and increasing lifespan exacerbates the burden of age-related health issues. On the other hand, premature aging has begun to be a problem, with increasing numbers of younger people suffering aging-related symptoms. Advanced aging is caused by a combination of factors: lifestyle, diet, external and internal factors, as well as oxidative stress (OS). Although OS is the most researched aging factor, it is also the least understood. OS is important not only in relation to aging but also due to its strong impact on neurodegenerative diseases such as amyotrophic lateral sclerosis (ALS), frontotemporal dementia (FTD), Alzheimer's disease (AD), and Parkinson's disease (PD). In this review, we will discuss the aging process in relation to OS, the function of OS in neurodegenerative disorders, and prospective therapeutics capable of relieving neurodegenerative symptoms associated with the pro-oxidative condition.

Keywords: oxidative stress; free radical; aging; neurodegenerative diseases

1. Introduction

Healthy aging is characterized by a gradual breakdown of physiological systems leading to a reduction in cognitive functions and brain health, but the timing and extent of this decline vary among older people. Oxidative stress (OS) is a crucial factor in the aging process that can cause direct damage to the brain's cellular architecture, causing neurodegenerative disease.

Aging is the main lead factor for many diseases, including cancer, metabolic, cardiac and neurodegenerative diseases. Aging is linked to a loss of homeostasis, involving degradation of structural components, reduced cellular maintenance and a decrease in overall physiological function/metabolism. There are two major aging theories: the free radical theory, which postulates chronological accumulation of defects in gene expression and environmental damage. According to the free radical theory of aging by Denham Harman, 1956 [\[1\]](#page-9-0), alterations in normal metabolic and mitochondrial function are induced by production of free radicals, which cause damage, aging, and associated aging illnesses.

The second and more recent theory is the mitochondrial theory of aging by J. Miquel and colleagues in 1980 [\[2\]](#page-9-1). This theory posits that the loss of balance between free radical production and repair mechanisms is responsible for aging.

The redox network is also essential in antioxidant defense. Mitochondria serve a crucial function by turning stored energy into adenosine triphosphate (ATP) through oxidative phosphorylation and phospholipid synthesis, buffering calcium, and coordinating programmed cell death [\[3\]](#page-9-2). Free radicals such as reactive oxygen species (ROS), reactive nitrogen species (RNS), and reactive sulfur species (RSS), which are present in all cells, but which are restricted by the antioxidant systems that neutralize them, are primarily produced by oxidative phosphorylation [\[4\]](#page-9-3). Free radicals are produced naturally and modulate cellular processes such as inflammation, cell survival, and stress responses,

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as well as numerous illnesses such as cardiovascular problems, muscular dysfunction, allergies, and malignancies [\[5\]](#page-9-4).

anergies, and mangnancies [5].
Free radicals are most likely formed due to processes involving molecular oxygen that are catalyzed in the cell by oxidative enzymes and in connective tissues by trace metals such as iron, cobalt, and manganese.

The primary distinction between healthy aging and accelerated aging is the balance between free radical elevation and the body's ability to guard and fight against RNS and ROS (Figure [1\)](#page-1-0).

Figure 1. The primary distinction between healthy aging and accelerated aging is the balance between the angle of the balance between $\frac{1}{2}$ and $\frac{1}{2}$ and antioxidation to minimize oxidative stress. tween free radical generation (RNS and ROS) and antioxidation to minimize oxidative stress. **Figure 1.** The primary distinction between healthy aging and accelerated aging is the balance between

ROS production may overpower antioxidant defenses, resulting in a noxious state known tion, Ros and general degradation of normal central ranchons. This has been documented
numerous clinical studies involving mitochondrial malfunction and aging [\[6\]](#page-9-5). In the event of loss of homeostasis between free radical production and detoxification, as OS and general degradation of normal cellular functions. This has been documented in

The brain is especially prone to oxidative damage due to its high oxygen consumption, limited antioxidant defenses, and high concentration of polyunsaturated fatty acids that are easily oxidized [\[7\]](#page-9-6).

are easny oxidized [/].
Lipid peroxides (LPO) are highly reactive molecules that include malondialdehyde tion, limited antioxidant defenses, and high concentration of polyunsaturated fatty acids (MDA), 4-hydroxy-2-nonenal (HNE), acrolein, isoprostanes (IsoPs), and neuroprostanes (NeuroPs). They can disrupt proteins and DNA structure and functions $[8-10]$ $[8-10]$. Increased MDA, IsoPs, and HNE have been observed in brain tissues of Tg2576 Alzheimer's disease (AD) moder nuce and post-mortem AD brains [11,12]. Oxidance dantage to DIVA
results in formation of oxidized base adducts, including 8-hydroxyguanosine (8-OHG) and 5,6-diamino-5-formamidopyridine in brains of mild cognitive impairment (MCI) patients. Despite the brain's great potential for ROS formation, its defense system against OS remains restricted and diminishes with age, owing to low amounts of endogenous antioxidants compared to other tissues, such as liver [\[13](#page-10-0)[,14\]](#page-10-1). The limited regenerative ability of postmitotic neurons renders OS more damaging to the brain, where the damage accumulates over time, compared to other organs $[15]$. ease (AD) model mice and post-mortem AD brains [\[11](#page-9-9)[,12\]](#page-9-10). Oxidative damage to DNA such as catalase (CAT), glutathione (GSH), glutathione peroxidase (GPx), and vitamin E,

2. Other Sources of Free Radicals Contribute to OS

Even though mitochondrial dysfunction is the primary source of free radicals, there are several other sources, such as genetic (endogenous), environmental, and lifestyle causes (exogenous) [\[16\]](#page-10-3).

2.1. Environmental Factors: Radiation/UV Rays and Pollution

Environmental factors, including smoking, UV radiation, heavy metal ions, ozone, allergens, drugs or toxins, pollutants, and pesticides, may all contribute to elevated ROS production in cells [\[17,](#page-10-4)[18\]](#page-10-5). Ionizing radiation transforms hydroxyl radicals, superoxides, and organic radicals into organic hydroperoxides and hydrogen peroxide. Subsequently, peroxides react with metal ions, particularly Fe and Cu through redox reactions, with further oxidative activity at the cellular level. Several studies have demonstrated that exposure of fibroblasts to alpha particles leads to an intracellular increase in oxygen and an acceleration of peroxide formation [\[19](#page-10-6)[,20\]](#page-10-7). Ultraviolet radiation (UVA) induces oxidative processes by stimulating riboflavin, porphyrins, and NADPH-oxidase, resulting in synthesis of 8-oxo-guanine and a drop in intracellular GSH levels, with a return to normal after exposure cessation [\[21\]](#page-10-8). Heavy metals are crucial to the formation of free radicals [\[22\]](#page-10-9).

Nickel, arsenic, iron, copper, cadmium and lead can produce free radicals by Fenton or Haber-Weiss reactions [\[23,](#page-10-10)[24\]](#page-10-11), as well as through direct reactions between metal ions and cellular constituents with similar effects, such as generation of thiol-type radicals. In brain tissue, lead causes lipid peroxidation and raises GPx concentrations. By attaching to sulfhydryl groups, arsenic generates peroxides, superoxides, and nitric oxide, and inhibits antioxidant enzymes such as glutathione-S-transferase (GST), GPx, and glutathione reductase (GR) [\[25\]](#page-10-12). Free radicals produced by these reactions can cause substitutions of DNA bases, including guanine for cytosine, guanine for thymine, and cytosine for thymine [\[26\]](#page-10-13). Even in healthy individuals, ozone exposure can impair lung function by increasing inflammatory infiltration in the respiratory epithelium [\[27\]](#page-10-14).

2.2. Lifestyle Related Factors

Lifestyle factors such as smoking, drinking alcohol, diet, exercise, and frequency of exercise all contribute to OS [\[28\]](#page-10-15). Some research has shown that ROS are present at the skeletal muscle level and help to control how muscle works. Muscle fibers always produce small quantities of reactive oxygen radicals, which are elevated by muscle contraction [\[29\]](#page-10-16). These oxygen radicals have many direct and indirect effects on muscle activity (contractility, excitability, metabolism, and calcium homeostasis) and are involved in skeletal muscle fatigue during hard exercise [\[30\]](#page-10-17).

Long, exhausting exercise and overcoming limits as a phase of overtraining cause a massive response to OS. Endogenous antioxidant status is improved by moderate exercise, low-intensity training, and training for a long time.

ROS are an essential part of how cells talk to each other and how antioxidant genes are turned on and off. Nuclear factor kappa B and mitogen-activated protein kinase are upregulated by physical activity [\[31,](#page-10-18)[32\]](#page-10-19). These activate gene expression of various enzymes and proteins that maintain oxidative/antioxidant intracellular homeostasis [\[33\]](#page-10-20).

Physical exercise is the main non-drug therapy for treating chronic diseases, especially heart diseases, and lifestyle changes [\[34\]](#page-10-21). Relevant studies [\[35\]](#page-10-22) have shown that autophagy, a process that breaks down and recycles cellular organs and nutrients, is important for cardiovascular benefits of training.

Cigarette smoke has oxidants, free radicals, and organic components such as nitric oxide and nitric superoxide. These cause neutrophils and macrophages to gather in lung tissues, increasing the production of oxidants in the area [\[36](#page-10-23)[,37\]](#page-10-24).

2.3. Genetic Factors

Enzymes that are responsible for fighting damage caused by OS and these enzymes and pathways are prone to free radical damage [\[38](#page-10-25)[–42\]](#page-11-0). Oligodendrocytes are susceptible to oxidative damage due to their function in maintenance and creation of myelin and
their limited rensir mechanisms, successing that white matter may be mare succeptible. their limited repair mechanisms, suggesting that white matter may be more susceptible to oxidative activity than gray matter. Antioxidant defense enzymes in the brain, such as superoxide dismutase (SOD), CAT, GPx, and GST are essential for neutralizing toxic byproducts of oxidative phosphorylation. Allelic variants of polymorphisms encoding these antioxidants are associated with anomalies in SOD, CAT, GPx, and GST activity in
the control persons system [28,42,44] the central nervous system [\[38,](#page-10-25)[43,](#page-11-1)[44\]](#page-11-2).

dative activity than gray matter. Antioxidant defense enzymes in the brain, such as superoxide dismutase (SOD), CAT, GPx, and GST are essential for neutralizing toxic byprod-

3. Mechanistic Evolution of Neurodegenerative Diseases Caused by OS Neurodegenerative disease is an irreversible condition in which neuronal function declines over the including the incidence of neurodegenerative death. The incidence of neuroneon and the incidence of neuroneon a

Neurodegenerative disease is an irreversible condition in which neuronal function Increase the and the version condition in which neuronal runcion
declines over time, leading to neuronal death. The incidence of neurodegenerative disease is increasing every year, especially in countries with aging populations. Common neurodegenerative diseases include Alzheimer's (AD) and Parkinson's (PD) diseases. Mitochondria are the primary source of ROS and are the main cause of neurodegenerative diseases (Figure [2\)](#page-3-0).

Alzheimer's disease, Parkinson's disease, and multiple sclerosis. **Figure 2.** Illustration showing the impact of oxidative stress on amyotrophic lateral sclerosis,

comprises five multiprotein complexes that mediate interaction of these electrons with oxygen, resulting in ROS hydrogen peroxide (H₂O₂), superoxide anions (O₂-), and hydroxyl radicals (\bullet OH) [\[14,](#page-10-1)[47\]](#page-11-5). Electron transfer from NADH to ubiquinone is catalyzed by mitochondrial complex I (reduced nicotinamide adenine dinucleotide [NADH] coenzyme
O reductee) (coenzyme O), Complex II also prevides alestrons to ubiquinane (quesipate de deroxyl radicals (*Controllow) and ζ . Completing the value of the station of model and η is the upinone is catalyzed (*Controllow). Electrons from reduced ubiquinone are donated to complex III (cytochrome bc1), and subsequently, to cytochrome c (CytC). Complex IV (CytC oxidase) is involved in interactions between molecular oxygen and electrons extracted from CytC, resulting in Mitochondrial synthesis of ATP through oxidative phosphorylation, has the substantial disadvantage of producing unpaired electrons [\[45](#page-11-3)[,46\]](#page-11-4). The electron transport chain Q reductase) (coenzyme Q). Complex II also provides electrons to ubiquinone (succinate dewater production [\[48\]](#page-11-6). Complexes I, II, and III are most often linked to premature electron leakage to oxygen and are the primary source of ROS generation [\[49\]](#page-11-7).

Additionally, elevated ROS levels trigger formation of additional reactive species, such as RNS, when $O₂$ reacts with other molecules, such as nitric oxide, to generate peroxynitrite (ONOO−). Furthermore, in addition to ROS and RNS, mitochondria generate RSSs, which are very reactive. Free oxygen radicals gradually damage proteins, lipids, and nucleic acids, resulting in inefficient or aberrant cellular functions, inflammation, and cell death [\[50\]](#page-11-8).

Mitochondrial internal components and mitochondrial DNA (mtDNA) in particular, are very vulnerable to OS-induced damage, resulting in impaired mitochondrial bioenergetics, increased ROS generation, and OS [\[51\]](#page-11-9). Enzymatic and non-enzymatic mechanisms safeguard antioxidant systems. SOD, CAT, GPx, GR, and thioredoxin (TRX) are the major enzymes involved in catalytic ROS elimination. Non-enzymatic complexes include vitamins A, C, and E, GSH, and proteins such as albumin and ceruloplasmin [\[52](#page-11-10)[,53\]](#page-11-11).

4. Major Degenerative Disorders Caused by OS

4.1. OS in Amyotrophic Lateral Sclerosis (ALS)

Despite substantial investigation, the genesis of amyotrophic lateral sclerosis (ALS) is still not fully clear. Ninety percent of ALS cases are sporadic and appear to lack a genetic foundation, whereas 10% of patients have familial ALS, mostly an autosomal dominant [\[54\]](#page-11-12). ALS has been linked to various occupational and environmental variables, such as exposure to various chemicals, metals, and pesticides, electromagnetic fields (EMFs), and lifestyle choices, including smoking and excessive exercise [\[55](#page-11-13)[–58\]](#page-11-14).

A recognized but poorly understood pathogenic ALS feature is abnormally high free radical levels and inadequate antioxidation. Undoubtedly, OS is essential for motor neuron death, but we do not know exactly when oxidative damage occurs [\[59\]](#page-11-15). OS biomarkers have been identified in brain tissue of ALS patients, cerebrospinal fluid (CSF), blood, and urine [\[60\]](#page-11-16). It is challenging to track OS biomarkers over a long period due to the short lifespans of ALS patients. Further, because of OS's random start and the present lack of tools to forecast its progression, it is difficult to be certain whether OS is a cause of ALS-associated neurodegeneration or a result of other underlying etiologic variables [\[57\]](#page-11-17). Investigations using a murine ALS model have revealed altered mitochondrial structures and nuclear factor erythroid 2-related factor 2 (Nrf2) pathway activation during early ALS stages, indicating OS involvement in the disease's early stages. OS damage is typically caused by stimulating formation of intracellular antioxidant molecules [\[61,](#page-11-18)[62\]](#page-11-19).

However, these studies used the murine mutant SOD1 ALS model, and SOD1 mutations only account for 20% of human familial ALS cases. Twenty individuals with sporadic ALS had significantly higher levels of lipid peroxidation and lower levels of the antioxidant enzymes CAT, GR, GSH, and glucose-6-phosphate dehydrogenase (G6PD) in their erythrocytes [\[63\]](#page-11-20). Progression of alterations parallel the pathophysiology of ALS, supporting OS participation in ALS development. Additionally, the aforementioned environmental and endoplasmic reticulum stress factors work together to promote pro-oxidative states that may ultimately harm motor neurons [\[64\]](#page-11-21).

The intermembrane space (IMS) of mitochondria contains a protein known as Coiled-Coil-Helix-Coiled-Coil-Helix Domain-Containing Protein 10 (CHCHD10), which has no known function [\[65\]](#page-11-22). However, it is believed to be involved in maintaining mitochondrial crista morphology and proper oxidative phosphorylation. In particular, in multiprotein complexes I, II, III, and IV, overexpression of mutant CHCHD10 harboring an allele linked to ALS alters mitochondrial structure and impairs electron transport chain function [\[66](#page-11-23)[,67\]](#page-12-0). In addition, fibroblasts from an ALS patient with a CHCHD10 mutation displayed mitochondrial ultrastructural damage and mitochondrial network fragmentation [\[66,](#page-11-23)[67\]](#page-12-0).

4.2. OS in Alzheimer's Disease (AD)

The most common neurodegenerative condition, AD, is characterized by a steady decline in behavior, cognition, and functioning that profoundly affect day-to-day activities. AD brains have higher protein, DNA, and lipid oxidation rates, as well as redox-active metals [\[51\]](#page-11-9).

Pathological indicators of AD include extracellular senile plaques (SP) and intracellular neurofibrillary tangles (NFT). Protein aggregation is sporadic, and its molecular mechanism is poorly understood. Several investigations have revealed that AD brains display elevated OS, which is crucial to disease development [\[68\]](#page-12-1). Amyloid fibril precursors are neurotoxic owing to OS generated by toxic conformer of amyloid oligomers and additional neurotoxic effects such as neuronal membrane rupture, microglia and astrocyte activation, and Ca^{2+} dyshomeostasis [\[69–](#page-12-2)[71\]](#page-12-3).

Tau protein aggregation is another hallmark of AD although in most cases, it is of late onset. Several reports have indicated that neurotoxicity caused by beta amyloid alteration is the main trigger of tau alteration to form tangles. NFTs in the early stages of AD are intracellular deposits; however, these progress to extracellular deposits in later stages. NFTs are composed of paired helical filaments (PHFs) in which the major component is tau protein [\[72–](#page-12-4)[76\]](#page-12-5).

When tau proteins are hyperphosphorylated, they self-assemble into NFTs and may be detected in neurons [\[77,](#page-12-6)[78\]](#page-12-7). According to recent data, amyloid deposition occurs 15–20 years before dementia manifests itself, and tau pathology appears thereafter [\[79](#page-12-8)[–83\]](#page-12-9).

Using multiphoton imaging, researchers identified a clear link between free radical generation and amyloid plaques in AD mouse models and human AD brain tissues, where fluorogenic free radical markers decreased following administration of a synthetic antioxidant [\[84\]](#page-12-10).

4.3. OS in Parkinson's Disease (PD)

Unlike AD, PD is clinically recognized by four cardinal motor symptoms: bradykinesia, stiffness, resting tremors, and trouble with posture and walking [\[83](#page-12-9)[–85\]](#page-12-11). In PD brains, the substantia nigra pars compacta and to a lesser extent, the globus pallidus, putamen, and caudate nucleus, exhibit selective dopaminergic neuronal loss. The nigrostriatal pathway's degenerating neurons release less of the neurotransmitter dopamine [\[86\]](#page-12-12). Lewy bodies are clusters of aberrant proteins seen in neurons of individuals with Parkinson's disease. They are components of $α$ -synuclein $(α$ -syn $)$ protein, broadly disseminated in the neurological system, but activities of which are not well understood [\[87\]](#page-12-13). α -syn fibrillation creates clumps that take up significant space within cells and ultimately result in neuronal death [\[88\]](#page-12-14). As in AD, pathophysiological pathways underlying PD are strongly related to OS [\[89\]](#page-12-15). Evidence of OS involvement may be seen in the substantia nigra of PD patients, where oxidized lipids, proteins, and DNA can be found [\[90,](#page-12-16)[91\]](#page-12-17). Additionally, the monoamine oxidase (MAO) that breaks down dopamine creates hydrogen peroxide, whereas dopamine on its own produces superoxide anion and reactive quinones. These reactive substances cause other nearby neurons, as well as dopaminergic neurons, to become cytotoxic [\[92](#page-12-18)[,93\]](#page-12-19).

4.4. OS in Multiple Sclerosis (MS)

Multiple sclerosis (MS) is a multifactorial autoimmune disease of the central nervous system (CNS), characterized by chronic inflammation, demyelination, and axon and neuron loss. Depending on the location of the demyelinating lesions, MS patients can develop almost any neurological sign or symptom, including motor, sensory, and cognitive impairment [\[94](#page-12-20)[,95\]](#page-13-0).

OS is heavily involved in several MS pathological hallmarks such as myelin destruction, axonal degeneration, and inflammation [\[96\]](#page-13-1). In MS as in other neurodegenerative diseases OS triggers activation of autophagy and microglia as well as of the neuroimmune system. Neurons, astrocytes, and oligodendrocytes produce chemicals that bind to microglial receptors in healthy settings, suppressing their activated states [\[97](#page-13-2)[–100\]](#page-13-3). When certain molecules (such as myelin CD47) are expressed less, microglial activation is increased, which may cause myelin debris to be phagocytosed and to provide neurotrophic

factors [\[101](#page-13-4)[,102\]](#page-13-5). Microglial physiological functions are transformed into harmful inflammatory insults by long-term damage, systemic inflammation, proinflammatory cytokine release, and ROS signaling [\[103\]](#page-13-6). Together, our results imply that during the onset and development of experimental autoimmune encephalomyelitis (EAE) and MS lesions, activated microglia and macrophages are directing tissue damage through their oxidative burst. Simultaneous activation of a sophisticated antioxidant response is insufficient to stop the apoptotic and degenerative processes, however.

Due to a combination of circumstances, including high levels of iron and polyunsaturated fatty acids, high iron requirements and mitochondrial activity, and restricted cell regeneration, the CNS is particularly susceptible to OS. Oxidative damage also affects the immune response that is developing in the periphery and controls MS illness outside of the CNS. First, by lowering its electrical resistance and thus changing its permeability, elevated ROS levels harm the brain endothelium [\[94](#page-12-20)[,104–](#page-13-7)[106\]](#page-13-8).

5. Significance of Non-Coding RNAs in OS

OS readily damages RNA due to its single-stranded structure and high concentration close to mitochondria, where most intracellular ROS are generated [\[107](#page-13-9)[,108\]](#page-13-10). An overabundance of ROS can cause chemical modification or even separation of RNA bases and breakage of RNA strands [\[91](#page-12-17)[,109\]](#page-13-11). Oxidatively damaged RNA accumulates in cells, leading to decreased protein synthesis, erroneous protein generation, and ultimately cell death [\[110,](#page-13-12)[111\]](#page-13-13).

Non-coding RNAs (ncRNAs), which do not encode proteins, make up the majority of RNAs in human cells [\[112](#page-13-14)[,113\]](#page-13-15). As a category of ncRNAs, regulatory ncRNAs, including long ncRNAs (lncRNAs) and small ncRNAs, which includes microRNAs (miRNAs), circular RNAs (circRNAs), and PIWI-interacting RNAs (piRNAs), are involved in regulating gene expression [\[114\]](#page-13-16). Compared to mRNAs, these regulatory ncRNAs persist relatively longer. Hence, oxidative damage can cause prolonged effects. ncRNAs of several types have been linked to various neurogenerative diseases [\[115–](#page-13-17)[118\]](#page-13-18). However, the direct relationship between ncRNA oxidation and neuronal diseases is still unclear, except in PD, in which the interaction between OS and regulatory ncRNAs has been well studied [\[119\]](#page-13-19). In contrast, regulatory small ncRNAs such as miRNAs and lncRNAs help to regulate ROS production [\[118\]](#page-13-18). Their interactions are involved in the pathophysiology of PD [\[120\]](#page-13-20).

RNA oxidation is not only a hallmark of PD but also a crucial first step in the development of the disorder [\[112](#page-13-14)[,121\]](#page-13-21). 8-oxo-7, 8-dihydroguanosine (8-oxoG) is one of many oxidative marks on RNA, and it is possibly the most common and well studied [\[122\]](#page-13-22). Guanine's vulnerability to free radicals means that it can form this base adduct, leading to mismatched bases [\[109\]](#page-13-11).

Neuronal apoptosis and autophagy are influenced by lncRNAs; thus, α-syn accumulation and degradation that restricts their function would have deleterious effects on cellular homeostasis [\[123\]](#page-14-0). For instance, free radical damage to miRNAs can cause them to misidentify their target mRNAs, which can increase expression of specific proteins [\[124\]](#page-14-1). A reduction in α-syn expression was mediated by two miRNAs in an experiment performed by Je and Kim [\[123\]](#page-14-0). High levels of α -syn and subsequent development of PD resulted from stress-induced oxidative loss of translational inhibition by these two miRNAs.

Furthermore, Chen et al. found that OS triggered modification of circRNA N6 methyladenosine (m6A). m6A-modified circRNAs can modulate expression of stress response genes (UBC and PPP2CA), which may constitute the mechanistic basis for OSinduced neurodegenerative disorders [\[125\]](#page-14-2).

The regulatory role of miRNAs in OS is closely linked to α -syn, which is responsible for inducing OS. Both microRNA-141-3p and microRNA-9-5p target the $3'$ UTR of SIRT1 mRNA. Since SIRT1 inhibits formation of α -syn aggregates, knockdown of microRNA-141-3p and microRNA-9-5p may alleviate OS and boost the viability of in vitro PD model [\[126](#page-14-3)[,127\]](#page-14-4).

SOD, CAT, and GPx are responsible for detoxifying oxidants and repairing oxidative damage. MicroRNA-137 and microRNA-494-3p aggravate OS by reducing SOD and GPx activity in PD rats treated with these miRNAs. MicroRNA-335 suppresses expression of FTH1, thereby promoting release of Fe^{2+} ions and generation of free radicals [\[128\]](#page-14-5). Downregulation of microRNA-410 expression in PD is associated with elevated ROS production [\[129\]](#page-14-6).

Regulatory lncRNAs and OS are hallmarks of PD. In PD, mitochondrial dysfunction is linked to ROS overproduction. α-syn aggregate formation can exacerbate OS by decreasing complex I activity or by activating microglia. Upregulation of beta-amyloid-cleaving enzyme-antisense (BACE1-AS), a lncRNA, was also associated with increasing levels of α syn in PD [\[130\]](#page-14-7). In addition, the lncRNA microRNA-17-92a-1 cluster host gene (MIR17HG) promotes α-syn expression. MicroRNA-153-5p cannot inhibit α-syn expression because MIR17HG acts as a sponge for microRNA-153-5p [\[131\]](#page-14-8). Inhibition of autophagy by GSK3 promotes α-syn buildup and therefore aggregation. MicroRNA-15b-5p suppressed GSK3 expression, but binding of SNHG1 to microRNA-15b-5p rescued GSK3 expression [\[132\]](#page-14-9). Cellular effects of oxidatively damaged RNA and the mechanism by which regulatory ncRNAs affect OS are just beginning to be explored.

6. Methods to Measure OS in Neurodegenerative Diseases

Although there are peripheral indicators for OS, it is difficult to detect in the human brain in vivo. The following measures have been used to quantify brain OS (Table [1\)](#page-7-0).

Table 1. Overview of the methods and biomarkers that are used to detect oxidative stress in neurodegenerative diseases.

MCI: Mild cognitive impairment; LPO: lipid peroxidation; GPx: glutathione peroxidase; GSH: glutathione; SOD: superoxide dismutase.

6.1. Peripheral Blood OS

Several surrogate OS or antioxidant activity indicators, including circulating lipid peroxides, GSH, and vitamins C and E, have been examined in peripheral blood [\[89](#page-12-15)[,133](#page-14-10)[–135\]](#page-14-17).

AD patients display reduced peripheral vitamin A, C, and E levels [\[136\]](#page-14-18) and SOD and GPx activity [\[137\]](#page-14-19). Lower plasma GSH levels are associated with more severe cognitive impairment in AD patients [\[138\]](#page-14-20). PD patients have inconsistently altered erythrocyte SOD activity [\[139,](#page-14-21)[140\]](#page-14-11). Increased SOD activity may protect against OS damage.

6.2. Magnetic Resonance Spectroscopy (MRS)

GSH is the single antioxidant assessed by ${}^{1}H$ (MRS) [\[149\]](#page-15-1). GSH content in the human brain is lower than N-acetyl aspartate, creatine, and choline, making evaluation difficult. With MRS, it is challenging to discern between GSH and glutamate resonance peaks [\[141,](#page-14-12)[150\]](#page-15-2). GSH-CH² cysteine's protons resonate at 2.93 and 2.97 ppm, overlapping with those of creatine (3.03 ppm) and aspartate (2.82 ppm) [\[151\]](#page-15-3). To detect GSH levels in the brain, spectral editing methods such as MEscher-GArwood-PRESS (MEGA-PRESS) [\[142\]](#page-14-22)

are needed to boost GSH signals so as to gather reliable nuclear signals. MEGA-PRESS combined with a 180° editing pulse in the original PRESS pulse sequence [\[142\]](#page-14-22) may distinguish GSH-cysteine signals from other signals, notably creatine signals in the brain [\[89\]](#page-12-15).

cortices [\[143,](#page-14-23)[144\]](#page-14-13). The concentration of vitamin C (ascorbic acid) in the human brain is around 1.0 mM, which may be detected by MRS [\[152\]](#page-15-4). However, measuring vitamin C using 1 H MRS is problematic owing to the similarities between the resonances of vitamin C (3.73, 4.01, and 4.50 ppm) and glutamate (3.75 ppm) [\[151\]](#page-15-3). MEGA-PRESS editing might potentially aid in measuring vitamin C levels in the human brain. Several earlier research [\[153\]](#page-15-5) used ¹H-MRS with the MEGA-PRESS to evaluate vitamin C levels in the human brain.

Recent research found that GSH levels are lower in AD patients' hippocampus and frontal

6.3. Electron Paramagnetic Resonance (EPR) Spectroscopy

Electromagnetic radiation absorption often occurs in the microwave region of the electromagnetic spectrum [\[154\]](#page-15-6). As a result, it is influenced by paramagnetic species that are present in a magnetic field. However, owing to short radical half-lives, compared to the EPR time scale, EPR spectroscopy cannot detect them directly [\[155\]](#page-15-7). To compensate for this, a stable chemical is frequently used to capture radicals to make them observable [\[156\]](#page-15-8).

EPR has been implement in detecting neurodegenerative diseases as in AD [\[145\]](#page-14-14) and as real time OS marker for post stroke patients [\[146\]](#page-14-15).

Despite years of research in EPR spectroscopy, one of the main reasons why it has not been widely employed is presumably poor sensitivity, particularly at the levels of free radicals usually encountered in biological systems. More research is required before using EPR in human clinical trials.

6.4. Positron Emission Tomography (PET)

The intracellular over-reactive state can be measured using a radiotracer for positron emission tomography (PET) [\[147\]](#page-14-16). [⁶²Cu] diacetyl-bis (N4-methylthiosemicarbazone) $({}^{62}Cu]$ ATSM) is a radiotracer widely used in PET. Evaluation of striatal OS in patients with PD using $[62Cu]$ ATSM PET was previously confirmed [\[147\]](#page-14-16). In a recent study, this tracer was utilized to visualize localized OS in PD patients that was mostly caused by mitochondrial malfunction. In this investigation, deposition of $[62Cu]$ ATSM was detected in the striatum of PD and ALS patients [\[147,](#page-14-16)[148\]](#page-15-0), indicating a localized over-reductive conditions caused by mitochondrial malfunction.

7. Approaches to Slow Aging by Fighting OS

The human body has several lines of defense against OS. Multiple antioxidant defense pathways are involved in the brain's ROS detoxification. The metalloproteins SOD, CAT, and GPx constitute the first line of antioxidant defense against ROS. Age-related accumulation of highly reactive polyunsaturated fatty acids, iron, and ROS is exacerbated by suboptimal antioxidant levels.

Apart from endogenous antioxidant defense mechanisms in aging brain and given the importance of OS in neurodegenerative diseases, manipulating ROS levels may be a viable therapeutic approach to slow neurodegeneration and reduce related symptoms.

Avoiding OS-related causes is at the top of preventative approaches. Diet, exercise, lack of sleep, sedentary behavior and circadian rhythm abnormalities are all crucial components in regulating healthy aging. The gut flora is hypersensitive to extrinsic variables linked to an unhealthy lifestyle [\[157](#page-15-9)[,158\]](#page-15-10). Preclinical studies showed that a high-fat diet (HFD) in mice changes the gut microbiome [\[159\]](#page-15-11). Pro-inflammatory bacteria such as Clostridium, Eubacterium, and Roseburia are positively correlated with hyperglycemic fluctuations in the brain [\[160\]](#page-15-12). The need for gut microbiome therapies is growing, and this area of research is continually developing. With impressive results, dietary and probiotic supplementation has been investigated as a possible therapeutic strategy for age-related disorders via changes in gut microbiota. Recent research has shown that exercise may alter

gut microbiota, adding to the potential advantages of this strategy for treating disorders related to aging [\[161\]](#page-15-13).

Nutraceuticals or natural compounds exist in food have been extensively studied worldwide due to their neuroprotective effects in vivo and in vitro, attributed to the antioxidative properties [\[162\]](#page-15-14). Nutraceuticals that show metal chelation ability and antiinflammatory properties such as curcumin [\[163](#page-15-15)[,164\]](#page-15-16), green tea polyphenols [\[164\]](#page-15-16), resveratrol [\[165\]](#page-15-17) and vitamin E [\[166](#page-15-18)[,167\]](#page-15-19) are examples of natural antioxidants from foods and other sources that show promise as therapeutic agents for OS-related neurodegenerative diseases.

8. Conclusions and Concluding Remarks

Aging is not a disease that needs to be treated but a natural process; however, premature or unhealthy aging still needs more study to understand mechanisms and leading causes. There is a controversial view that OS leads to a short life span and is linked to age-related disease and quality of life. Numerous studies have shown a significant elevation in OS and free radical concentration as a common cause of aging and accompanying neurodegenerative diseases. However, in nature, comparative studies indicate long lives of some animals despite high levels of free radicals in their tissues. This means that there are many unknown mechanisms to cope with OS. Therefore, a healthy lifestyle to avoid factors that elevate OS, as well as increased intake of natural antioxidants, can protect against OS and prevent neurodegenerative diseases before their onset.

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References

- 1. Harman, D. Aging: A Theory Based on Free Radical and Radiation Chemistry. *J. Gerontol.* **1956**, *11*, 298–300. [\[CrossRef\]](http://doi.org/10.1093/geronj/11.3.298) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/13332224)
- 2. Miquel, J.; Economos, A.C.; Fleming, J.; Johnson, J.E., Jr. Mitochondrial Role in Cell Aging. *Exp. Gerontol.* **1980**, *15*, 575–591. [\[CrossRef\]](http://doi.org/10.1016/0531-5565(80)90010-8) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/7009178)
- 3. Smith, E.F.; Shaw, P.J.; De Vos, K.J. The Role of Mitochondria in Amyotrophic Lateral Sclerosis. *Neurosci. Lett.* **2019**, *710*, 132933. [\[CrossRef\]](http://doi.org/10.1016/j.neulet.2017.06.052) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28669745)
- 4. Sbodio, J.I.; Snyder, S.H.; Paul, B.D. Redox Mechanisms in Neurodegeneration: From Disease Outcomes to Therapeutic Opportunities. *Antioxid. Redox Signal.* **2019**, *30*, 1450–1499. [\[CrossRef\]](http://doi.org/10.1089/ars.2017.7321)
- 5. Zuo, L.; Koozechian, M.S.; Chen, L.L. Characterization of Reactive Nitrogen Species in Allergic Asthma. *Ann. Allergy Asthma Immunol.* **2014**, *112*, 18–22. [\[CrossRef\]](http://doi.org/10.1016/j.anai.2013.10.007) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/24331388)
- 6. Zuo, L.; Zhou, T.; Pannell, B.K.; Ziegler, A.C.; Best, T.M. Biological and Physiological Role of Reactive Oxygen Species—The Good, the Bad and the Ugly. *Acta Physiol.* **2015**, *214*, 329–348. [\[CrossRef\]](http://doi.org/10.1111/apha.12515)
- 7. Kausar, S.; Wang, F.; Cui, H. The Role of Mitochondria in Reactive Oxygen Species Generation and Its Implications for Neurodegenerative Diseases. *Cells* **2018**, *7*, 274. [\[CrossRef\]](http://doi.org/10.3390/cells7120274)
- 8. Taso, O.V.; Philippou, A.; Moustogiannis, A.; Zevolis, E.; Koutsilieris, M. Lipid Peroxidation Products and Their Role in Neurodegenerative Diseases. *Ann. Res. Hosp.* **2019**, *3*, 2. [\[CrossRef\]](http://doi.org/10.21037/arh.2018.12.02)
- 9. Bradley-Whitman, M.A.; Lovell, M.A. Biomarkers of Lipid Peroxidation in Alzheimer Disease (AD): An Update. *Arch. Toxicol.* **2015**, *89*, 1035–1044. [\[CrossRef\]](http://doi.org/10.1007/s00204-015-1517-6)
- 10. Ciamporcero, E.; Daga, M.; Pizzimenti, S.; Roetto, A.; Dianzani, C.; Compagnone, A.; Palmieri, A.; Ullio, C.; Cangemi, L.; Pili, R.; et al. Crosstalk between Nrf2 and YAP Contributes to Maintaining the Antioxidant Potential and Chemoresistance in Bladder Cancer. *Free Radic. Biol. Med.* **2018**, *115*, 447–457. [\[CrossRef\]](http://doi.org/10.1016/j.freeradbiomed.2017.12.005)
- 11. Perluigi, M.; Coccia, R.; Butterfield, D.A. 4-Hydroxy-2-Nonenal, a Reactive Product of Lipid Peroxidation, and Neurodegenerative Diseases: A Toxic Combination Illuminated by Redox Proteomics Studies. *Antioxid. Redox Signal.* **2012**, *17*, 1590–1609. [\[CrossRef\]](http://doi.org/10.1089/ars.2011.4406) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/22114878)
- 12. Kao, Y.-C.; Ho, P.-C.; Tu, Y.-K.; Jou, I.-M.; Tsai, K.-J. Lipids and Alzheimer's Disease. *Int. J. Mol. Sci.* **2020**, *21*, 1505. [\[CrossRef\]](http://doi.org/10.3390/ijms21041505) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/32098382)
- 13. Cheng, K.C.; Cahill, D.S.; Kasai, H.; Nishimura, S.; Loeb, L.A. 8-Hydroxyguanine, an Abundant Form of Oxidative DNA Damage, Causes G—T and A—C Substitutions. *J. Biol. Chem.* **1992**, *267*, 166–172. [\[CrossRef\]](http://doi.org/10.1016/S0021-9258(18)48474-8) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/1730583)
- 14. Cadenas, S. ROS and Redox Signaling in Myocardial Ischemia-Reperfusion Injury and Cardioprotection. *Free Radic. Biol. Med.* **2018**, *117*, 76–89. [\[CrossRef\]](http://doi.org/10.1016/j.freeradbiomed.2018.01.024) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/29373843)
- 15. Maynard, S.; Fang, E.F.; Scheibye-Knudsen, M.; Croteau, D.L.; Bohr, V.A. DNA Damage, DNA Repair, Aging, and Neurodegeneration. *Cold Spring Harb. Perspect. Med.* **2015**, *5*, a025130. [\[CrossRef\]](http://doi.org/10.1101/cshperspect.a025130) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26385091)
- 16. Sharifi-Rad, M.; Anil Kumar, N.V.; Zucca, P.; Varoni, E.M.; Dini, L.; Panzarini, E.; Rajkovic, J.; Tsouh Fokou, P.V.; Azzini, E.; Peluso, I.; et al. Lifestyle, Oxidative Stress, and Antioxidants: Back and Forth in the Pathophysiology of Chronic Diseases. *Front. Physiol.* **2020**, *11*, 694. [\[CrossRef\]](http://doi.org/10.3389/fphys.2020.00694)
- 17. Antunes dos Santos, A.; Ferrer, B.; Marques Gonçalves, F.; Tsatsakis, A.M.; Renieri, E.A.; Skalny, A.V.; Farina, M.; Rocha, J.B.T.; Aschner, M. Oxidative Stress in Methylmercury-Induced Cell Toxicity. *Toxics* **2018**, *6*, 47. [\[CrossRef\]](http://doi.org/10.3390/toxics6030047)
- 18. Mahajan, A.; Taliun, D.; Thurner, M.; Robertson, N.R.; Torres, J.M.; Rayner, N.W.; Payne, A.J.; Steinthorsdottir, V.; Scott, R.A.; Grarup, N.; et al. Fine-Mapping Type 2 Diabetes Loci to Single-Variant Resolution Using High-Density Imputation and Islet-Specific Epigenome Maps. *Nat. Genet.* **2018**, *50*, 1505–1513. [\[CrossRef\]](http://doi.org/10.1038/s41588-018-0241-6)
- 19. Spitz, D.R.; Azzam, E.I.; Jian Li, J.; Gius, D. Metabolic Oxidation/Reduction Reactions and Cellular Responses to Ionizing Radiation: A Unifying Concept in Stress Response Biology. *Cancer Metastasis Rev.* **2004**, *23*, 311–322. [\[CrossRef\]](http://doi.org/10.1023/B:CANC.0000031769.14728.bc)
- 20. Spitz, D.R.; Hauer-Jensen, M. Ionizing Radiation-Induced Responses: Where Free Radical Chemistry Meets Redox Biology and Medicine. *Antioxid. Redox Signal.* **2014**, *20*, 1407–1409. [\[CrossRef\]](http://doi.org/10.1089/ars.2013.5769)
- 21. Marchitti, S.A.; Chen, Y.; Thompson, D.C.; Vasiliou, V. Ultraviolet Radiation: Cellular Antioxidant Response and the Role of Ocular Aldehyde Dehydrogenase Enzymes. *Eye Contact Lens* **2011**, *37*, 206–213. [\[CrossRef\]](http://doi.org/10.1097/ICL.0b013e3182212642) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/21670692)
- 22. Sciskalska, M.; Zalewska, M.; Grzelak, A.; Milnerowicz, H. The Influence of the Occupational Exposure to Heavy Metals and ´ Tobacco Smoke on the Selected Oxidative Stress Markers in Smelters. *Biol. Trace Elem. Res.* **2014**, *159*, 59–68. [\[CrossRef\]](http://doi.org/10.1007/s12011-014-9984-9) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/24789476)
- 23. Koppenol, W.H. The Haber-Weiss Cycle—70 Years Later. *Redox Rep.* **2001**, *6*, 229–234. [\[CrossRef\]](http://doi.org/10.1179/135100001101536373) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/11642713)
- 24. Das, T.K.; Wati, M.R.; Fatima-Shad, K. Oxidative Stress Gated by Fenton and Haber Weiss Reactions and Its Association with Alzheimer's Disease. *Arch. Neurosci.* **2015**, *2*, e60038. [\[CrossRef\]](http://doi.org/10.5812/archneurosci.20078)
- 25. Birben, E.; Sahiner, U.M.; Sackesen, C.; Erzurum, S.; Kalayci, O. Oxidative Stress and Antioxidant Defense. *World Allergy Organ. J.* **2012**, *5*, 9–19. [\[CrossRef\]](http://doi.org/10.1097/WOX.0b013e3182439613)
- 26. Jan, A.T.; Azam, M.; Siddiqui, K.; Ali, A.; Choi, I.; Haq, Q.M.R. Heavy Metals and Human Health: Mechanistic Insight into Toxicity and Counter Defense System of Antioxidants. *Int. J. Mol. Sci.* **2015**, *16*, 29592–29630. [\[CrossRef\]](http://doi.org/10.3390/ijms161226183)
- 27. Li, J.; Zou, B.; Yeo, Y.H.; Feng, Y.; Xie, X.; Lee, D.H.; Fujii, H.; Wu, Y.; Kam, L.Y.; Ji, F.; et al. Prevalence, Incidence, and Outcome of Non-Alcoholic Fatty Liver Disease in Asia, 1999–2019: A Systematic Review and Meta-Analysis. *Lancet Gastroenterol. Hepatol.* **2019**, *4*, 389–398. [\[CrossRef\]](http://doi.org/10.1016/S2468-1253(19)30039-1)
- 28. Fisher-Wellman, K.; Bloomer, R.J. Acute Exercise and Oxidative Stress: A 30 Year History. *Dyn. Med.* **2009**, *8*, 1. [\[CrossRef\]](http://doi.org/10.1186/1476-5918-8-1)
- 29. He, F.; Li, J.; Liu, Z.; Chuang, C.-C.; Yang, W.; Zuo, L. Redox Mechanism of Reactive Oxygen Species in Exercise. *Front. Physiol.* **2016**, *7*, 486. [\[CrossRef\]](http://doi.org/10.3389/fphys.2016.00486) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/27872595)
- 30. Pingitore, A.; Lima, G.P.P.; Mastorci, F.; Quinones, A.; Iervasi, G.; Vassalle, C. Exercise and Oxidative Stress: Potential Effects of Antioxidant Dietary Strategies in Sports. *Nutrition* **2015**, *31*, 916–922. [\[CrossRef\]](http://doi.org/10.1016/j.nut.2015.02.005) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26059364)
- 31. Adams, V.; Späte, U.; Kränkel, N.; Christian, S.P.; Linke, A.; Schuler, G.; Hambrecht, R. Nuclear Factor-Kappa B Activation in Skeletal Muscle of Patients with Chronic Heart Failure: Correlation with the Expression of Inducible Nitric Oxide Synthase. *Eur. J. Cardiovasc. Prev. Rehabil.* **2003**, *10*, 273–277. [\[CrossRef\]](http://doi.org/10.1097/00149831-200308000-00009) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/14555882)
- 32. Ji, L.L.; Gomez-Cabrera, M.-C.; Vina, J. Role of Nuclear Factor kappaB and Mitogen-Activated Protein Kinase Signaling in Exercise-Induced Antioxidant Enzyme Adaptation. *Appl. Physiol. Nutr. Metab.* **2007**, *32*, 930–935. [\[CrossRef\]](http://doi.org/10.1139/H07-098) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/18059618)
- 33. Antonioni, A.; Fantini, C.; Dimauro, I.; Caporossi, D. Redox Homeostasis in Sport: Do Athletes Really Need Antioxidant Support? *Res. Sport. Med.* **2019**, *27*, 147–165. [\[CrossRef\]](http://doi.org/10.1080/15438627.2018.1563899) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30596287)
- 34. Ren, J.; Taegtmeyer, H. Too Much or Not Enough of a Good Thing—The Janus Faces of Autophagy in Cardiac Fuel and Protein Homeostasis. *J. Mol. Cell. Cardiol.* **2015**, *84*, 223–226. [\[CrossRef\]](http://doi.org/10.1016/j.yjmcc.2015.03.001)
- 35. Wu, N.N.; Tian, H.; Chen, P.; Wang, D.; Ren, J.; Zhang, Y. Physical Exercise and Selective Autophagy: Benefit and Risk on Cardiovascular Health. *Cells* **2019**, *8*, 1436. [\[CrossRef\]](http://doi.org/10.3390/cells8111436)
- 36. Rahman, I.; Biswas, S.K.; Kode, A. Oxidant and Antioxidant Balance in the Airways and Airway Diseases. *Eur. J. Pharmacol.* **2006**, *533*, 222–239. [\[CrossRef\]](http://doi.org/10.1016/j.ejphar.2005.12.087)
- 37. Valavanidis, A.; Vlachogianni, T.; Fiotakis, C. 8-Hydroxy-2'-Deoxyguanosine (8-OHdG): A Critical Biomarker of Oxidative Stress and Carcinogenesis. *J. Environ. Sci. Health Part C* **2009**, *27*, 120–139. [\[CrossRef\]](http://doi.org/10.1080/10590500902885684)
- 38. Salminen, L.E.; Paul, R.H. Oxidative Stress and Genetic Markers of Suboptimal Antioxidant Defense in the Aging Brain: A Theoretical Review. *Rev. Neurosci.* **2014**, *25*, 805–819. [\[CrossRef\]](http://doi.org/10.1515/revneuro-2014-0046)
- 39. Pizzino, G.; Irrera, N.; Cucinotta, M.; Pallio, G.; Mannino, F.; Arcoraci, V.; Squadrito, F.; Altavilla, D.; Bitto, A. Oxidative Stress: Harms and Benefits for Human Health. *Oxid. Med. Cell. Longev.* **2017**, *2017*, e8416763. [\[CrossRef\]](http://doi.org/10.1155/2017/8416763)
- 40. Ighodaro, O.M.; Akinloye, O.A. First Line Defence Antioxidants-Superoxide Dismutase (SOD), Catalase (CAT) and Glutathione Peroxidase (GPX): Their Fundamental Role in the Entire Antioxidant Defence Grid. *Alex. J. Med.* **2018**, *54*, 287–293. [\[CrossRef\]](http://doi.org/10.1016/j.ajme.2017.09.001)
- 41. Górny, M.; Bilska-Wilkosz, A.; Iciek, M.; Hereta, M.; Kamińska, K.; Kamińska, A.; Chwatko, G.; Rogóż, Z.; Lorenc-Koci, E. Alterations in the Antioxidant Enzyme Activities in the Neurodevelopmental Rat Model of Schizophrenia Induced by Glutathione Deficiency during Early Postnatal Life. *Antioxidants* **2020**, *9*, 538. [\[CrossRef\]](http://doi.org/10.3390/antiox9060538) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/32575563)
- 42. Gusti, A.M.T.; Qusti, S.Y.; Alshammari, E.M.; Toraih, E.A.; Fawzy, M.S. Antioxidants-Related Superoxide Dismutase (SOD), Catalase (CAT), Glutathione Peroxidase (GPX), Glutathione-S-Transferase (GST), and Nitric Oxide Synthase (NOS) Gene Variants Analysis in an Obese Population: A Preliminary Case-Control Study. *Antioxidants* **2021**, *10*, 595. [\[CrossRef\]](http://doi.org/10.3390/antiox10040595) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/33924357)
- 43. Chiras, D.; Kitsos, G.; Petersen, M.B.; Skalidakis, I.; Kroupis, C. Oxidative Stress in Dry Age-Related Macular Degeneration and Exfoliation Syndrome. *Crit. Rev. Clin. Lab. Sci.* **2015**, *52*, 12–27. [\[CrossRef\]](http://doi.org/10.3109/10408363.2014.968703) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25319011)
- 44. Kowalska, M.; Wize, K.; Prendecki, M.; Lianeri, M.; Kozubski, W.; Dorszewska, J. Genetic Variants and Oxidative Stress in Alzheimer's Disease. *Curr. Alzheimer Res.* **2020**, *17*, 208–223. [\[CrossRef\]](http://doi.org/10.2174/1567205017666200224121447)
- 45. Mailloux, R.J. Teaching the Fundamentals of Electron Transfer Reactions in Mitochondria and the Production and Detection of Reactive Oxygen Species. *Redox Biol.* **2015**, *4*, 381–398. [\[CrossRef\]](http://doi.org/10.1016/j.redox.2015.02.001)
- 46. Glantzounis, G.K.; Salacinski, H.J.; Yang, W.; Davidson, B.R.; Seifalian, A.M. The Contemporary Role of Antioxidant Therapy in Attenuating Liver Ischemia-Reperfusion Injury: A Review. *Liver Transplant.* **2005**, *11*, 1031–1047. [\[CrossRef\]](http://doi.org/10.1002/lt.20504)
- 47. Panov, A.V.; Dikalov, S.I. Cardiolipin, Perhydroxyl Radicals, and Lipid Peroxidation in Mitochondrial Dysfunctions and Aging. *Oxid. Med. Cell. Longev.* **2020**, *2020*, 1323028. [\[CrossRef\]](http://doi.org/10.1155/2020/1323028)
- 48. Srinivasan, S.; Avadhani, N.G. Cytochrome c Oxidase Dysfunction in Oxidative Stress. *Free Radic. Biol. Med.* **2012**, *53*, 1252–1263. [\[CrossRef\]](http://doi.org/10.1016/j.freeradbiomed.2012.07.021)
- 49. Turrens, J.F. Mitochondrial Formation of Reactive Oxygen Species. *J. Physiol.* **2003**, *552*, 335–344. [\[CrossRef\]](http://doi.org/10.1113/jphysiol.2003.049478)
- 50. Ghosh, N.; Das, A.; Chaffee, S.; Roy, S.; Sen, C.K. Reactive Oxygen Species, Oxidative Damage and Cell Death. In *Immunity and Inflammation in Health and Disease*; Chatterjee, S., Jungraithmayr, W., Bagchi, D., Eds.; Academic Press: Cambridge, MA, USA, 2018; pp. 45–55.
- 51. Jellinger, K.A. Basic Mechanisms of Neurodegeneration: A Critical Update. *J. Cell. Mol. Med.* **2010**, *14*, 457–487. [\[CrossRef\]](http://doi.org/10.1111/j.1582-4934.2010.01010.x)
- 52. Halliwell, B.; Gutteridge, J.M.C. The Antioxidants of Human Extracellular Fluids. *Arch. Biochem. Biophys.* **1990**, *280*, 1–8. [\[CrossRef\]](http://doi.org/10.1016/0003-9861(90)90510-6) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/2191627)
- 53. Aruoma, O.I.; Neergheen, V.S.; Bahorun, T.; Jen, L.-S. Free Radicals, Antioxidants and Diabetes: Embryopathy, Retinopathy, Neuropathy, Nephropathy and Cardiovascular Complications. *Neuroembryol. Aging* **2006**, *4*, 117–137. [\[CrossRef\]](http://doi.org/10.1159/000109344)
- 54. Mejzini, R.; Flynn, L.L.; Pitout, I.L.; Fletcher, S.; Wilton, S.D.; Akkari, P.A. ALS Genetics, Mechanisms, and Therapeutics: Where Are We Now? *Front. Neurosci.* **2019**, *13*, 1310. [\[CrossRef\]](http://doi.org/10.3389/fnins.2019.01310) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31866818)
- 55. Ingre, C.; Roos, P.M.; Piehl, F.; Kamel, F.; Fang, F. Risk Factors for Amyotrophic Lateral Sclerosis. *Clin. Epidemiol.* **2015**, *7*, 181–193. [\[CrossRef\]](http://doi.org/10.2147/CLEP.S37505)
- 56. Motataianu, A.; Serban, G.; Barcutean, L.; Balasa, R. Oxidative Stress in Amyotrophic Lateral Sclerosis: Synergy of Genetic and Environmental Factors. *Int. J. Mol. Sci.* **2022**, *23*, 9339. [\[CrossRef\]](http://doi.org/10.3390/ijms23169339)
- 57. Cunha-Oliveira, T.; Montezinho, L.; Mendes, C.; Firuzi, O.; Saso, L.; Oliveira, P.J.; Silva, F.S. Oxidative Stress in Amyotrophic Lateral Sclerosis: Pathophysiology and Opportunities for Pharmacological Intervention. *Oxid. Med. Cell. Longev.* **2020**, *2020*, 5021694. [\[CrossRef\]](http://doi.org/10.1155/2020/5021694)
- 58. Carrì, M.T.; Valle, C.; Bozzo, F.; Cozzolino, M. Oxidative Stress and Mitochondrial Damage: Importance in Non-SOD1 ALS. *Front. Cell. Neurosci.* **2015**, *9*, 41. [\[CrossRef\]](http://doi.org/10.3389/fncel.2015.00041)
- 59. Agar, J.; Durham, H. Relevance of Oxidative Injury in the Pathogenesis of Motor Neuron Diseases. *Amyotroph. Lateral Scler. Other Mot. Neuron Disord.* **2003**, *4*, 232–242. [\[CrossRef\]](http://doi.org/10.1080/14660820310011278)
- 60. Singh, A.; Kukreti, R.; Saso, L.; Kukreti, S. Oxidative Stress: A Key Modulator in Neurodegenerative Diseases. *Molecules* **2019**, *24*, 1583. [\[CrossRef\]](http://doi.org/10.3390/molecules24081583)
- 61. Kraft, A.D.; Resch, J.M.; Johnson, D.A.; Johnson, J.A. Activation of the Nrf2–ARE Pathway in Muscle and Spinal Cord during ALS-like Pathology in Mice Expressing Mutant SOD1. *Exp. Neurol.* **2007**, *207*, 107–117. [\[CrossRef\]](http://doi.org/10.1016/j.expneurol.2007.05.026)
- 62. Velde, C.V.; McDonald, K.K.; Boukhedimi, Y.; McAlonis-Downes, M.; Lobsiger, C.S.; Hadj, S.B.; Zandona, A.; Julien, J.-P.; Shah, S.B.; Cleveland, D.W. Misfolded SOD1 Associated with Motor Neuron Mitochondria Alters Mitochondrial Shape and Distribution Prior to Clinical Onset. *PLoS ONE* **2011**, *6*, e22031. [\[CrossRef\]](http://doi.org/10.1371/journal.pone.0022031) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/21779368)
- 63. Babu, G.N.; Kumar, A.; Chandra, R.; Puri, S.K.; Singh, R.L.; Kalita, J.; Misra, U.K. Oxidant–Antioxidant Imbalance in the Erythrocytes of Sporadic Amyotrophic Lateral Sclerosis Patients Correlates with the Progression of Disease. *Neurochem. Int.* **2008**, *52*, 1284–1289. [\[CrossRef\]](http://doi.org/10.1016/j.neuint.2008.01.009) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/18308427)
- 64. Keon, M.; Musrie, B.; Dinger, M.; Brennan, S.E.; Santos, J.; Saksena, N.K. Destination Amyotrophic Lateral Sclerosis. *Front. Neurol.* **2021**, *12*, 596006. [\[CrossRef\]](http://doi.org/10.3389/fneur.2021.596006)
- 65. Anderson, C.J.; Bredvik, K.; Burstein, S.R.; Davis, C.; Meadows, S.M.; Dash, J.; Case, L.; Milner, T.A.; Kawamata, H.; Zuberi, A.; et al. ALS/FTD Mutant CHCHD10 Mice Reveal a Tissue-Specific Toxic Gain-of-Function and Mitochondrial Stress Response. *Acta Neuropathol.* **2019**, *138*, 103–121. [\[CrossRef\]](http://doi.org/10.1007/s00401-019-01989-y)
- 66. Bannwarth, S.; Ait-El-Mkadem, S.; Chaussenot, A.; Genin, E.C.; Lacas-Gervais, S.; Fragaki, K.; Berg-Alonso, L.; Kageyama, Y.; Serre, V.; Moore, D.G.; et al. A Mitochondrial Origin for Frontotemporal Dementia and Amyotrophic Lateral Sclerosis through *CHCHD10* Involvement. *Brain* **2014**, *137*, 2329–2345. [\[CrossRef\]](http://doi.org/10.1093/brain/awu138)
- 67. Genin, E.C.; Bannwarth, S.; Ropert, B.; Lespinasse, F.; Mauri-Crouzet, A.; Augé, G.; Fragaki, K.; Cochaud, C.; Donnarumma, E.; Lacas-Gervais, S.; et al. CHCHD10 and SLP2 Control the Stability of the PHB Complex: A Key Factor for Motor Neuron Viability. *Brain* **2022**, *145*, 3415–3430. [\[CrossRef\]](http://doi.org/10.1093/brain/awac197) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/35656794)
- 68. Fontana, I.C.; Zimmer, A.R.; Rocha, A.S.; Gosmann, G.; Souza, D.O.; Lourenco, M.V.; Ferreira, S.T.; Zimmer, E.R. Amyloid-β Oligomers in Cellular Models of Alzheimer's Disease. *J. Neurochem.* **2020**, *155*, 348–369. [\[CrossRef\]](http://doi.org/10.1111/jnc.15030)
- 69. Fernandez-Perez, E.J.; Peters, C.; Aguayo, L.G. Membrane Damage Induced by Amyloid Beta and a Potential Link with Neuroinflammation. *Curr. Pharm. Des.* **2016**, *22*, 1295–1304. [\[CrossRef\]](http://doi.org/10.2174/138161282210160304111702)
- 70. Meraz-Ríos, M.A.; Lira-De León, K.I.; Campos-Peña, V.; De Anda-Hernández, M.A.; Mena-López, R. Tau Oligomers and Aggregation in Alzheimer's Disease. *J. Neurochem.* **2010**, *112*, 1353–1367. [\[CrossRef\]](http://doi.org/10.1111/j.1471-4159.2009.06511.x)
- 71. Calvo-Rodriguez, M.; Bacskai, B.J. Mitochondria and Calcium in Alzheimer's Disease: From Cell Signaling to Neuronal Cell Death. *Trends Neurosci.* **2021**, *44*, 136–151. [\[CrossRef\]](http://doi.org/10.1016/j.tins.2020.10.004)
- 72. Liu, Z.; Li, T.; Li, P.; Wei, N.; Zhao, Z.; Liang, H.; Ji, X.; Chen, W.; Xue, M.; Wei, J. The Ambiguous Relationship of Oxidative Stress, Tau Hyperphosphorylation, and Autophagy Dysfunction in Alzheimer's Disease. *Oxid. Med. Cell. Longev.* **2015**, *2015*, e352723. [\[CrossRef\]](http://doi.org/10.1155/2015/352723) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26171115)
- 73. Rojas Quijano, F.A.; Morrow, D.; Wise, B.M.; Brancia, F.L.; Goux, W.J. Prediction of Nucleating Sequences from Amyloidogenic Propensities of Tau-Related Peptides. *Biochemistry* **2006**, *45*, 4638–4652. [\[CrossRef\]](http://doi.org/10.1021/bi052226q) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/16584199)
- 74. Goedert, M. Tau Protein and the Neurofibrillary Pathology of Alzheimer's Disease. *Trends Neurosci.* **1993**, *16*, 460–465. [\[CrossRef\]](http://doi.org/10.1016/0166-2236(93)90078-Z) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/7507619)
- 75. Weaver, C.L.; Espinoza, M.; Kress, Y.; Davies, P. Conformational Change as One of the Earliest Alterations of Tau in Alzheimer's Disease. *Neurobiol. Aging* **2000**, *21*, 719–727. [\[CrossRef\]](http://doi.org/10.1016/S0197-4580(00)00157-3)
- 76. Grabowski, T.J.; Cho, H.S.; Vonsattel, J.P.G.; Rebeck, G.W.; Greenberg, S.M. Novel Amyloid Precursor Protein Mutation in an Iowa Family with Dementia and Severe Cerebral Amyloid Angiopathy. *Ann. Neurol.* **2001**, *49*, 697–705. [\[CrossRef\]](http://doi.org/10.1002/ana.1009)
- 77. Hansson, O. Biomarkers for Neurodegenerative Diseases. *Nat. Med.* **2021**, *27*, 954–963. [\[CrossRef\]](http://doi.org/10.1038/s41591-021-01382-x)
- 78. Braak, H.; Thal, D.R.; Ghebremedhin, E.; Del Tredici, K. Stages of the Pathologic Process in Alzheimer Disease: Age Categories from 1 to 100 Years. *J. Neuropathol. Exp. Neurol.* **2011**, *70*, 960–969. [\[CrossRef\]](http://doi.org/10.1097/NEN.0b013e318232a379)
- 79. Fleisher, A.S.; Chen, K.; Quiroz, Y.T.; Jakimovich, L.J.; Gomez, M.G.; Langois, C.M.; Langbaum, J.B.; Ayutyanont, N.; Roontiva, A.; Thiyyagura, P. Florbetapir PET Analysis of Amyloid-β Deposition in the Presenilin 1 E280A Autosomal Dominant Alzheimer's Disease Kindred: A Cross-Sectional Study. *Lancet Neurol.* **2012**, *11*, 1057–1065. [\[CrossRef\]](http://doi.org/10.1016/S1474-4422(12)70227-2)
- 80. O'Brien, J.T.; Herholz, K. Amyloid Imaging for Dementia in Clinical Practice. *BMC Med.* **2015**, *13*, 163. [\[CrossRef\]](http://doi.org/10.1186/s12916-015-0404-6)
- 81. Gunasingh Masilamoni, J.; Philip Jesudason, E.; Dhandayuthapani, S.; Ashok, B.S.; Vignesh, S.; Jebaraj, W.C.E.; Paul, S.F.; Jayakumar, R. The Neuroprotective Role of Melatonin against Amyloid *β* Peptide Injected Mice. *Free Radic. Res.* **2008**, *42*, 661–673. [\[CrossRef\]](http://doi.org/10.1080/10715760802277388)
- 82. Smith, D.G.; Cappai, R.; Barnham, K.J. The Redox Chemistry of the Alzheimer's Disease Amyloid Beta Peptide. *Biochim. Biophys. Acta* **2007**, *1768*, 1976–1990. [\[CrossRef\]](http://doi.org/10.1016/j.bbamem.2007.02.002) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/17433250)
- 83. Jankovic, J. Parkinson's Disease: Clinical Features and Diagnosis. *J. Neurol. Neurosurg. Psychiatry* **2008**, *79*, 368–376. [\[CrossRef\]](http://doi.org/10.1136/jnnp.2007.131045) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/18344392)
- 84. Blandini, F.; Nappi, G.; Tassorelli, C.; Martignoni, E. Functional Changes of the Basal Ganglia Circuitry in Parkinson's Disease. *Prog. Neurobiol.* **2000**, *62*, 63–88. [\[CrossRef\]](http://doi.org/10.1016/S0301-0082(99)00067-2) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/10821982)
- 85. Agosta, F.; Weiler, M.; Filippi, M. Propagation of Pathology through Brain Networks in Neurodegenerative Diseases: From Molecules to Clinical Phenotypes. *CNS Neurosci. Ther.* **2015**, *21*, 754–767. [\[CrossRef\]](http://doi.org/10.1111/cns.12410) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/26031656)
- 86. Jansen van Rensburg, Z.; Abrahams, S.; Bardien, S.; Kenyon, C. Toxic Feedback Loop Involving Iron, Reactive Oxygen Species, α-Synuclein and Neuromelanin in Parkinson's Disease and Intervention with Turmeric. *Mol. Neurobiol.* **2021**, *58*, 5920–5936. Available online: <https://link.springer.com/article/10.1007/s12035-021-02516-5> (accessed on 3 January 2023). [\[CrossRef\]](http://doi.org/10.1007/s12035-021-02516-5)
- 87. Kim, G.H.; Kim, J.E.; Rhie, S.J.; Yoon, S. The Role of Oxidative Stress in Neurodegenerative Diseases. *Exp. Neurobiol.* **2015**, *24*, 325. [\[CrossRef\]](http://doi.org/10.5607/en.2015.24.4.325)
- 88. Jenner, P. Oxidative Stress in Parkinson's Disease. *Ann. Neurol.* **2003**, *53*, S26–S38. [\[CrossRef\]](http://doi.org/10.1002/ana.10483)
- 89. Fahn, S.; Cohen, G. The Oxidant Stress Hypothesis in Parkinson's Disease: Evidence Supporting It. *Ann. Neurol.* **1992**, *32*, 804–812. [\[CrossRef\]](http://doi.org/10.1002/ana.410320616)
- 90. Delcambre, S.; Nonnenmacher, Y.; Hiller, K. Dopamine Metabolism and Reactive Oxygen Species Production. In *Mitochondrial Mechanisms of Degeneration and Repair in Parkinson's Disease*; Buhlman, L.M., Ed.; Springer International Publishing: Cham, Switzerland, 2016; pp. 25–47. ISBN 978-3-319-42139-1.
- 91. Nunomura, A.; Moreira, P.I.; Castellani, R.J.; Lee, H.; Zhu, X.; Smith, M.A.; Perry, G. Oxidative Damage to RNA in Aging and Neurodegenerative Disorders. *Neurotox. Res.* **2012**, *22*, 231–248. [\[CrossRef\]](http://doi.org/10.1007/s12640-012-9331-x)
- 92. Li, Z.; Chen, X.; Liu, Z.; Ye, W.; Li, L.; Qian, L.; Ding, H.; Li, P.; Aung, L.H.H. Recent Advances: Molecular Mechanism of RNA Oxidation and Its Role in Various Diseases. *Front. Mol. Biosci.* **2020**, *7*, 184. [\[CrossRef\]](http://doi.org/10.3389/fmolb.2020.00184)
- 93. Jellinger, K.A. Recent Advances in Our Understanding of Neurodegeneration. *J. Neural Transm.* **2009**, *116*, 1111–1162. [\[CrossRef\]](http://doi.org/10.1007/s00702-009-0240-y) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/19707851)
- 94. Pegoretti, V.; Swanson, K.A.; Bethea, J.R.; Probert, L.; Eisel, U.L.M.; Fischer, R. Inflammation and Oxidative Stress in Multiple Sclerosis: Consequences for Therapy Development. *Oxid. Med. Cell. Longev.* **2020**, *2020*, e7191080. [\[CrossRef\]](http://doi.org/10.1155/2020/7191080) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/32454942)
- 95. Compston, A. Genetic Epidemiology of Multiple Sclerosis. *J. Neurol. Neurosurg. Psychiatry* **1997**, *62*, 553–561. [\[CrossRef\]](http://doi.org/10.1136/jnnp.62.6.553) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/9219738)
- 96. Adamczyk-Sowa, M.; Galiniak, S.; Żyracka, E.; Grzesik, M.; Naparło, K.; Sowa, P.; Bartosz, G.; Sadowska-Bartosz, I. Oxidative Modification of Blood Serum Proteins in Multiple Sclerosis after Interferon Beta and Melatonin Treatment. *Oxid. Med. Cell. Longev.* **2017**, *2017*, e7905148. [\[CrossRef\]](http://doi.org/10.1155/2017/7905148) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/29181127)
- 97. Hernangómez, M.; Carrillo-Salinas, F.J.; Mecha, M.; Correa, F.; Mestre, L.; Loría, F.; Feliú, A.; Docagne, F.; Guaza, C. Brain Innate Immunity in the Regulation of Neuroinflammation: Therapeutic Strategies by Modulating CD200-CD200R Interaction Involve the Cannabinoid System. *Curr. Pharm. Des.* **2014**, *20*, 4707–4722. [\[CrossRef\]](http://doi.org/10.2174/1381612820666140130202911)
- 98. Nair, A.; Frederick, T.J.; Miller, S.D. Astrocytes in Multiple Sclerosis: A Product of Their Environment. *Cell. Mol. Life Sci.* **2008**, *65*, 2702–2720. [\[CrossRef\]](http://doi.org/10.1007/s00018-008-8059-5) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/18516496)
- 99. Kawabori, M.; Yenari, M.A. Inflammatory Responses in Brain Ischemia. *Curr. Med. Chem.* **2015**, *22*, 1258–1277. [\[CrossRef\]](http://doi.org/10.2174/0929867322666150209154036) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25666795)
- 100. Degan, D.; Ornello, R.; Tiseo, C.; Carolei, A.; Sacco, S.; Pistoia, F. The Role of Inflammation in Neurological Disorders. *Curr. Pharm. Des.* **2018**, *24*, 1485–1501. [\[CrossRef\]](http://doi.org/10.2174/1381612824666180327170632)
- 101. Kasschau, M.; Sherman, K.; Haider, L.; Frontario, A.; Shaw, M.; Datta, A.; Bikson, M.; Charvet, L. A Protocol for the Use of Remotely-Supervised Transcranial Direct Current Stimulation (TDCS) in Multiple Sclerosis (MS). *J. Vis. Exp.* **2015**, *106*, e53542.
- 102. Gitik, M.; Liraz-Zaltsman, S.; Oldenborg, P.-A.; Reichert, F.; Rotshenker, S. Myelin Down-Regulates Myelin Phagocytosis by Microglia and Macrophages through Interactions between CD47 on Myelin and SIRPα (Signal Regulatory Protein-α) on Phagocytes. *J. Neuroinflamm.* **2011**, *8*, 24. [\[CrossRef\]](http://doi.org/10.1186/1742-2094-8-24)
- 103. Perry, V.H. Contribution of Systemic Inflammation to Chronic Neurodegeneration. *Acta Neuropathol.* **2010**, *120*, 277–286. [\[CrossRef\]](http://doi.org/10.1007/s00401-010-0722-x) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/20644946)
- 104. Rodgers, J.M.; Miller, S.D. Cytokine Control of Inflammation and Repair in the Pathology of Multiple Sclerosis. *Yale J. Biol. Med.* **2012**, *85*, 447. [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/23239947)
- 105. Carson, M.J.; Doose, J.M.; Melchior, B.; Schmid, C.D.; Ploix, C.C. CNS Immune Privilege: Hiding in Plain Sight. *Immunol. Rev.* **2006**, *213*, 48–65. [\[CrossRef\]](http://doi.org/10.1111/j.1600-065X.2006.00441.x)
- 106. Holman, D.W.; Klein, R.S.; Ransohoff, R.M. The Blood–Brain Barrier, Chemokines and Multiple Sclerosis. *Biochim. Biophys. Acta Mol. Basis Dis.* **2011**, *1812*, 220–230. [\[CrossRef\]](http://doi.org/10.1016/j.bbadis.2010.07.019) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/20692338)
- 107. Tanaka, M.; Song, H.; Küpfer, P.A.; Leumann, C.J.; Sonntag, W.E. An Assay for RNA Oxidation Induced Abasic Sites Using the Aldehyde Reactive Probe. *Free Radic. Res.* **2011**, *45*, 237–247. [\[CrossRef\]](http://doi.org/10.3109/10715762.2010.535529) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/21062214)
- 108. Pieper, A.A.; Verma, A.; Zhang, J.; Snyder, S.H. Poly (ADP-Ribose) Polymerase, Nitric Oxide and Cell Death. *Trends Pharmacol. Sci.* **1999**, *20*, 171–181. [\[CrossRef\]](http://doi.org/10.1016/S0165-6147(99)01292-4)
- 109. Nunomura, A.; Perry, G. RNA and Oxidative Stress in Alzheimer's Disease: Focus on MicroRNAs. *Oxid. Med. Cell. Longev.* **2020**, *2020*, e2638130. [\[CrossRef\]](http://doi.org/10.1155/2020/2638130)
- 110. Moreira, P.I.; Nunomura, A.; Nakamura, M.; Takeda, A.; Shenk, J.C.; Aliev, G.; Smith, M.A.; Perry, G. Nucleic Acid Oxidation in Alzheimer Disease. *Free Radic. Biol. Med.* **2008**, *44*, 1493–1505. [\[CrossRef\]](http://doi.org/10.1016/j.freeradbiomed.2008.01.002)
- 111. Djebali, S.; Davis, C.A.; Merkel, A.; Dobin, A.; Lassmann, T.; Mortazavi, A.; Tanzer, A.; Lagarde, J.; Lin, W.; Schlesinger, F.; et al. Landscape of Transcription in Human Cells. *Nature* **2012**, *489*, 101–108. [\[CrossRef\]](http://doi.org/10.1038/nature11233)
- 112. Zhang, P.; Wu, W.; Chen, Q.; Chen, M. Non-Coding RNAs and Their Integrated Networks. *J. Integr. Bioinform.* **2019**, *16*, 20190027. [\[CrossRef\]](http://doi.org/10.1515/jib-2019-0027)
- 113. Johnson, R.; Noble, W.; Tartaglia, G.G.; Buckley, N.J. Neurodegeneration as an RNA Disorder. *Prog. Neurobiol.* **2012**, *99*, 293–315. [\[CrossRef\]](http://doi.org/10.1016/j.pneurobio.2012.09.006) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/23063563)
- 114. Salta, E.; De Strooper, B. Noncoding RNAs in Neurodegeneration. *Nat. Rev. Neurosci.* **2017**, *18*, 627–640. [\[CrossRef\]](http://doi.org/10.1038/nrn.2017.90) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28855739)
- 115. Abdelhamid, R.F.; Ogawa, K.; Beck, G.; Ikenaka, K.; Takeuchi, E.; Yasumizu, Y.; Jinno, J.; Kimura, Y.; Baba, K.; Nagai, Y.; et al. PiRNA/PIWI Protein Complex as a Potential Biomarker in Sporadic Amyotrophic Lateral Sclerosis. *Mol. Neurobiol.* **2022**, *59*, 1693–1705. [\[CrossRef\]](http://doi.org/10.1007/s12035-021-02686-2) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/35015250)
- 116. Tai, Y.; Chen, J.; Tao, Z.; Ren, J. Non-Coding RNAs: New Players in Mitophagy and Neurodegeneration. *Neurochem. Int.* **2022**, *152*, 105253. [\[CrossRef\]](http://doi.org/10.1016/j.neuint.2021.105253) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/34864089)
- 117. Zhang, H.; Liu, X.; Liu, Y.; Liu, J.; Gong, X.; Li, G.; Tang, M. Crosstalk between Regulatory Non-Coding RNAs and Oxidative Stress in Parkinson's Disease. *Front. Aging Neurosci.* **2022**, *14*, 975248. [\[CrossRef\]](http://doi.org/10.3389/fnagi.2022.975248) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/36016854)
- 118. Konovalova, J.; Gerasymchuk, D.; Parkkinen, I.; Chmielarz, P.; Domanskyi, A. Interplay between MicroRNAs and Oxidative Stress in Neurodegenerative Diseases. *Int. J. Mol. Sci.* **2019**, *20*, 6055. [\[CrossRef\]](http://doi.org/10.3390/ijms20236055) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31801298)
- 119. Cervinkova, B.; Krcmova, L.K.; Sestakova, V.; Solichova, D.; Solich, P. A Fully Validated Bioanalytical Method Using an UHPLC– MS/MS System for Quantification of DNA and RNA Oxidative Stress Biomarkers. *Anal Bioanal. Chem.* **2017**, *409*, 3611–3621. [\[CrossRef\]](http://doi.org/10.1007/s00216-017-0301-2) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28341989)
- 120. Lushchak, V.I.; Duszenko, M.; Gospodaryov, D.V.; Garaschuk, O. Oxidative Stress and Energy Metabolism in the Brain: Midlife as a Turning Point. *Antioxidants* **2021**, *10*, 1715. [\[CrossRef\]](http://doi.org/10.3390/antiox10111715)
- 121. Zhang, X.; Li, L. The Significance of 8-OxoGsn in Aging-Related Diseases. *Aging Dis.* **2020**, *11*, 1329–1338. [\[CrossRef\]](http://doi.org/10.14336/AD.2019.1021)
- 122. Yan, L.L.; Simms, C.L.; McLoughlin, F.; Vierstra, R.D.; Zaher, H.S. Oxidation and Alkylation Stresses Activate Ribosome-Quality Control. *Nat. Commun.* **2019**, *10*, 5611. [\[CrossRef\]](http://doi.org/10.1038/s41467-019-13579-3)
- 123. Je, G.; Kim, Y.-S. Mitochondrial ROS-Mediated Post-Transcriptional Regulation of α-Synuclein through MiR-7 and MiR-153. *Neurosci. Lett.* **2017**, *661*, 132–136. [\[CrossRef\]](http://doi.org/10.1016/j.neulet.2017.09.065) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28986122)
- 124. Chen, N.; Tang, J.; Su, Q.; Chou, W.-C.; Zheng, F.; Guo, Z.; Yu, G.; Shao, W.; Li, H.; Wu, S. Paraquat-Induced Oxidative Stress Regulates N6-Methyladenosine (M6A) Modification of Circular RNAs. *Environ. Pollut.* **2021**, *290*, 117816. [\[CrossRef\]](http://doi.org/10.1016/j.envpol.2021.117816) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/34425375)
- 125. Zheng, Y.; Dong, L.; Liu, N.; Luo, X.; He, Z. Mir-141-3p Regulates Apoptosis and Mitochondrial Membrane Potential via Targeting Sirtuin1 in a 1-Methyl-4-Phenylpyridinium In Vitro Model of Parkinson's Disease. *BioMed Res. Int.* **2020**, *2020*, e7239895. [\[CrossRef\]](http://doi.org/10.1155/2020/7239895) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/33204711)
- 126. Ge, H.; Yan, Z.; Zhu, H.; Zhao, H. MiR-410 Exerts Neuroprotective Effects in a Cellular Model of Parkinson's Disease Induced by 6-Hydroxydopamine via Inhibiting the PTEN/AKT/MTOR Signaling Pathway. *Exp. Mol. Pathol.* **2019**, *109*, 16–24. [\[CrossRef\]](http://doi.org/10.1016/j.yexmp.2019.05.002)
- 127. Li, X.; Si, W.; Li, Z.; Tian, Y.; Liu, X.; Ye, S.; Huang, Z.; Ji, Y.; Zhao, C.; Hao, X.; et al. MiR-335 Promotes Ferroptosis by Targeting Ferritin Heavy Chain 1 in In Vivo and In Vitro Models of Parkinson's Disease. *Int. J. Mol. Med.* **2021**, *47*, 61. [\[CrossRef\]](http://doi.org/10.3892/ijmm.2021.4894)
- 128. Li, Y.; Fang, J.; Zhou, Z.; Zhou, Q.; Sun, S.; Jin, Z.; Xi, Z.; Wei, J. Downregulation of LncRNA BACE1-AS Improves Dopamine-Dependent Oxidative Stress in Rats with Parkinson's Disease by Upregulating MicroRNA-34b-5p and Downregulating BACE1. *Cell Cycle* **2020**, *19*, 1158–1171. [\[CrossRef\]](http://doi.org/10.1080/15384101.2020.1749447)
- 129. Zhang, J.; Yang, Y.; Zhou, C.; Zhu, R.; Xiao, X.; Zhou, B.; Wan, D. LncRNA MiR-17-92a-1 Cluster Host Gene (MIR17HG) Promotes Neuronal Damage and Microglial Activation by Targeting the MicroRNA-153-3p/Alpha-Synuclein Axis in Parkinson's Disease. *Bioengineered* **2022**, *13*, 4493–4516. [\[CrossRef\]](http://doi.org/10.1080/21655979.2022.2033409)
- 130. Xie, N.; Qi, J.; Li, S.; Deng, J.; Chen, Y.; Lian, Y. Upregulated LncRNA Small Nucleolar RNA Host Gene 1 Promotes 1-Methyl-4-Phenylpyridinium Ion-Induced Cytotoxicity and Reactive Oxygen Species Production through MiR-15b-5p/GSK3β Axis in Human Dopaminergic SH-SY5Y Cells. *J. Cell. Biochem.* **2019**, *120*, 5790–5801. [\[CrossRef\]](http://doi.org/10.1002/jcb.27865)
- 131. Rodrigo, R.; Libuy, M.; Feliú, F.; Hasson, D. Oxidative Stress-Related Biomarkers in Essential Hypertension and Ischemia-Reperfusion Myocardial Damage. *Dis. Mark.* **2013**, *35*, 773–790. [\[CrossRef\]](http://doi.org/10.1155/2013/974358)
- 132. Cherubini, A.; Ruggiero, C.; Polidori, M.C.; Mecocci, P. Potential Markers of Oxidative Stress in Stroke. *Free Radic. Biol. Med.* **2005**, *39*, 841–852. [\[CrossRef\]](http://doi.org/10.1016/j.freeradbiomed.2005.06.025)
- 133. Boskovic, M.; Vovk, T.; Kores Plesnicar, B.; Grabnar, I. Oxidative Stress in Schizophrenia. *Curr. Neuropharmacol.* **2011**, *9*, 301–312. 134. Varesi, A.; Chirumbolo, S.; Campagnoli, L.I.M.; Pierella, E.; Piccini, G.B.; Carrara, A.; Ricevuti, G.; Scassellati, C.; Bonvicini, C.; Pascale, A. The Role of Antioxidants in the Interplay between Oxidative Stress and Senescence. *Antioxidants* **2022**, *11*, 1224.
- [\[CrossRef\]](http://doi.org/10.3390/antiox11071224) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/35883714) 135. Michiels, C.; Raes, M.; Toussaint, O.; Remacle, J. Importance of Se-Glutathione Peroxidase, Catalase, and Cu/Zn-SOD for Cell Survival against Oxidative Stress. *Free Radic. Biol. Med.* **1994**, *17*, 235–248. [\[CrossRef\]](http://doi.org/10.1016/0891-5849(94)90079-5) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/7982629)
- 136. Lin, C.-H.; Lane, H.-Y. Plasma Glutathione Levels Decreased with Cognitive Decline among People with Mild Cognitive Impairment (MCI): A Two-Year Prospective Study. *Antioxidants* **2021**, *10*, 1839. [\[CrossRef\]](http://doi.org/10.3390/antiox10111839)
- 137. Delfino, R.J.; Staimer, N.; Tjoa, T.; Polidori, A.; Arhami, M.; Gillen, D.L.; Kleinman, M.T.; Vaziri, N.D.; Longhurst, J.; Zaldivar, F. Circulating Biomarkers of Inflammation, Antioxidant Activity, and Platelet Activation Are Associated with Primary Combustion Aerosols in Subjects with Coronary Artery Disease. *Environ. Health Perspect.* **2008**, *116*, 898–906. [\[CrossRef\]](http://doi.org/10.1289/ehp.11189)
- 138. Sussulini, A.; Hauser-Davis, R.A. Metallomics Applied to the Study of Neurodegenerative and Mental Diseases. In *Metallomics: The Science of Biometals*; Springer: Berlin/Heidelberg, Germany, 2018; pp. 21–37.
- 139. Crotty, G.F.; Ascherio, A.; Schwarzschild, M.A. Targeting Urate to Reduce Oxidative Stress in Parkinson Disease. *Exp. Neurol.* **2017**, *298*, 210–224. [\[CrossRef\]](http://doi.org/10.1016/j.expneurol.2017.06.017)
- 140. Das, T.K.; Javadzadeh, A.; Dey, A.; Sabesan, P.; Theberge, J.; Radua, J.; Palaniyappan, L. Antioxidant Defense in Schizophrenia and Bipolar Disorder: A Meta-Analysis of MRS Studies of Anterior Cingulate Glutathione. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry* **2019**, *91*, 94–102. [\[CrossRef\]](http://doi.org/10.1016/j.pnpbp.2018.08.006)
- 141. Shukla, D.; Mandal, P.K.; Tripathi, M.; Vishwakarma, G.; Mishra, R.; Sandal, K. Quantitation of In Vivo Brain Glutathione Conformers in Cingulate Cortex among Age-Matched Control, MCI, and AD Patients Using MEGA-PRESS. *Hum. Brain Mapp.* **2020**, *41*, 194–217. [\[CrossRef\]](http://doi.org/10.1002/hbm.24799)
- 142. Mandal, P.K.; Saharan, S.; Tripathi, M.; Murari, G. Brain Glutathione Levels–a Novel Biomarker for Mild Cognitive Impairment and Alzheimer's Disease. *Biol. Psychiatry* **2015**, *78*, 702–710. [\[CrossRef\]](http://doi.org/10.1016/j.biopsych.2015.04.005)
- 143. Shih, Y.-Y.; Büchert, M.; Chung, H.-W.; Hennig, J.; von Elverfeldt, D. Vitamin C Estimation with Standard ¹H Spectroscopy Using a Clinical 3T MR System: Detectability and Reliability within the Human Brain. *J. Magn. Reson. Imaging* **2008**, *28*, 351–358. [\[CrossRef\]](http://doi.org/10.1002/jmri.21466)
- 144. Terpstra, M.; Gruetter, R. ¹H NMR Detection of Vitamin C in Human Brain In Vivo. *Magn. Reson. Med.* **2004**, *51*, 225–229. [\[CrossRef\]](http://doi.org/10.1002/mrm.10715)
- 145. Sepkhanova, I.; Drescher, M.; Meeuwenoord, N.J.; Limpens, R.W.; Koning, R.I.; Filippov, D.V.; Huber, M. Monitoring Alzheimer Amyloid Peptide Aggregation by EPR. *Appl. Magn. Reson.* **2009**, *36*, 209–222. [\[CrossRef\]](http://doi.org/10.1007/s00723-009-0019-1) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/19946595)
- 146. Nikolova, G.; Komsiyska, D.; Karamalakova, Y.; Petkov, Y.; Ivanov, V.; Manolova, T.; Gadjeva, V. Real Time Oxidative Stress Markers of Patients with Post-Stroke Depression: EPR Study. *Bulg. Chem. Commun.* **2018**, *50*, 64–68.
- 147. Ikawa, M.; Okazawa, H.; Kudo, T.; Kuriyama, M.; Fujibayashi, Y.; Yoneda, M. Evaluation of Striatal Oxidative Stress in Patients with Parkinson's Disease Using [62Cu]ATSM PET. *Nucl. Med. Biol.* **2011**, *38*, 945–951. [\[CrossRef\]](http://doi.org/10.1016/j.nucmedbio.2011.02.016) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/21982566)
- 148. Ikawa, M.; Okazawa, H.; Tsujikawa, T.; Matsunaga, A.; Yamamura, O.; Mori, T.; Hamano, T.; Kiyono, Y.; Nakamoto, Y.; Yoneda, M. Increased Oxidative Stress Is Related to Disease Severity in the ALS Motor Cortex: A PET Study. *Neurology* **2015**, *84*, 2033–2039. [\[CrossRef\]](http://doi.org/10.1212/WNL.0000000000001588) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25904686)
- 149. Harris, A.D.; Saleh, M.G.; Edden, R.A. Edited 1H Magnetic Resonance Spectroscopy In Vivo: Methods and Metabolites. *Magn. Reson. Med.* **2017**, *77*, 1377–1389. [\[CrossRef\]](http://doi.org/10.1002/mrm.26619)
- 150. Matsuzawa, D.; Hashimoto, K. Magnetic Resonance Spectroscopy Study of the Antioxidant Defense System in Schizophrenia. *Antioxid. Redox Signal.* **2011**, *15*, 2057–2065. [\[CrossRef\]](http://doi.org/10.1089/ars.2010.3453)
- 151. Yahya, A. Metabolite Detection by Proton Magnetic Resonance Spectroscopy Using PRESS. *Prog. Nucl. Magn. Reson. Spectrosc.* **2009**, *55*, 183–198. [\[CrossRef\]](http://doi.org/10.1016/j.pnmrs.2009.04.001)
- 152. Johnson, C.C.; Guy, A.W. Nonionizing Electromagnetic Wave Effects in Biological Materials and Systems. *Proc. IEEE* **1972**, *60*, 692–718. [\[CrossRef\]](http://doi.org/10.1109/PROC.1972.8728)
- 153. Samuni, A.; Carmichael, A.J.; Russo, A.; Mitchell, J.B.; Riesz, P. On the Spin Trapping and ESR Detection of Oxygen-Derived Radicals Generated inside Cells. *Proc. Natl. Acad. Sci. USA* **1986**, *83*, 7593–7597. [\[CrossRef\]](http://doi.org/10.1073/pnas.83.20.7593)
- 154. Hawkins, C.L.; Davies, M.J. Detection and Characterisation of Radicals in Biological Materials Using EPR Methodology. *Biochim. Et Biophys. Acta Gen. Subj.* **2014**, *1840*, 708–721. [\[CrossRef\]](http://doi.org/10.1016/j.bbagen.2013.03.034)
- 155. Mrakic-Sposta, S.; Gussoni, M.; Montorsi, M.; Porcelli, S.; Vezzoli, A. Assessment of a Standardized ROS Production Profile in Humans by Electron Paramagnetic Resonance. *Oxid. Med. Cell. Longev.* **2012**, *2012*, e973927. [\[CrossRef\]](http://doi.org/10.1155/2012/973927)
- 156. Askarova, S.; Umbayev, B.; Masoud, A.-R.; Kaiyrlykyzy, A.; Safarova, Y.; Tsoy, A.; Olzhayev, F.; Kushugulova, A. The Links between the Gut Microbiome, Aging, Modern Lifestyle and Alzheimer's Disease. *Front. Cell. Infect. Microbiol.* **2020**, *10*, 104. [\[CrossRef\]](http://doi.org/10.3389/fcimb.2020.00104)
- 157. Poroyko, V.A.; Carreras, A.; Khalyfa, A.; Khalyfa, A.A.; Leone, V.; Peris, E.; Almendros, I.; Gileles-Hillel, A.; Qiao, Z.; Hubert, N.; et al. Chronic Sleep Disruption Alters Gut Microbiota, Induces Systemic and Adipose Tissue Inflammation and Insulin Resistance in Mice. *Sci. Rep.* **2016**, *6*, 35405. [\[CrossRef\]](http://doi.org/10.1038/srep35405)
- 158. Abraham, D.; Feher, J.; Scuderi, G.L.; Szabo, D.; Dobolyi, A.; Cservenak, M.; Juhasz, J.; Ligeti, B.; Pongor, S.; Gomez-Cabrera, M.C.; et al. Exercise and Probiotics Attenuate the Development of Alzheimer's Disease in Transgenic Mice: Role of Microbiome. *Exp. Gerontol.* **2019**, *115*, 122–131. [\[CrossRef\]](http://doi.org/10.1016/j.exger.2018.12.005)
- 159. Carbajo-Pescador, S.; Porras, D.; García-Mediavilla, M.V.; Martínez-Flórez, S.; Juarez-Fernández, M.; Cuevas, M.J.; Mauriz, J.L.; González-Gallego, J.; Nistal, E.; Sánchez-Campos, S. Beneficial Effects of Exercise on Gut Microbiota Functionality and Barrier Integrity, and Gut-Liver Crosstalk in an In Vivo Model of Early Obesity and Non-Alcoholic Fatty Liver Disease. *Dis. Model. Mech.* **2019**, *12*, dmm039206. [\[CrossRef\]](http://doi.org/10.1242/dmm.039206)
- 160. Ramli, N.Z.; Yahaya, M.F.; Tooyama, I.; Damanhuri, H.A. A Mechanistic Evaluation of Antioxidant Nutraceuticals on Their Potential against Age-Associated Neurodegenerative Diseases. *Antioxidants* **2020**, *9*, 1019. [\[CrossRef\]](http://doi.org/10.3390/antiox9101019)
- 161. Khan, M.S.; Muhammad, T.; Ikram, M.; Kim, M.O. Dietary Supplementation of the Antioxidant Curcumin Halts Systemic LPS-Induced Neuroinflammation-Associated Neurodegeneration and Memory/Synaptic Impairment via the JNK/NF-*κ*B/Akt Signaling Pathway in Adult Rats. *Oxid. Med. Cell. Longev.* **2019**, *2019*, e7860650. [\[CrossRef\]](http://doi.org/10.1155/2019/7860650)
- 162. Dong, H.; Xu, L.; Wu, L.; Wang, X.; Duan, W.; Li, H.; Li, C. Curcumin Abolishes Mutant TDP-43 Induced Excitability in a Motoneuron-like Cellular Model of ALS. *Neuroscience* **2014**, *272*, 141–153. [\[CrossRef\]](http://doi.org/10.1016/j.neuroscience.2014.04.032)
- 163. Zhao, J.; Xu, L.; Liang, Q.; Sun, Q.; Chen, C.; Zhang, Y.; Ding, Y.; Zhou, P. Metal Chelator EGCG Attenuates Fe(III)-Induced Conformational Transition of α-Synuclein and Protects AS-PC12 Cells against Fe(III)-Induced Death. *J. Neurochem.* **2017**, *143*, 136–146. [\[CrossRef\]](http://doi.org/10.1111/jnc.14142)
- 164. Wan Nasri, W.N.; Makpol, S.; Mazlan, M.; Tooyama, I.; Wan Ngah, W.Z.; Damanhuri, H.A. Tocotrienol Rich Fraction Supplementation Modulate Brain Hippocampal Gene Expression in APPswe/PS1dE9 Alzheimer's Disease Mouse Model. *J. Alzheimer's Dis.* **2019**, *70*, S239–S254. [\[CrossRef\]](http://doi.org/10.3233/JAD-180496) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30507571)
- 165. Yu, L.; Tu, Y.; Jia, X.; Fang, K.; Liu, L.; Wan, L.; Xiang, C.; Wang, Y.; Sun, X.; Liu, T.; et al. Resveratrol Protects Against Pulmonary Arterial Hypertension in Rats via Activation of Silent Information Regulator 1. *Cell. Physiol. Biochem.* **2017**, *42*, 55–67. [\[CrossRef\]](http://doi.org/10.1159/000477115) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28494457)
- 166. Chow, C.K. Vitamin E and Oxidative Stress. *Free Radic. Biol. Med.* **1991**, *11*, 215–232. [\[CrossRef\]](http://doi.org/10.1016/0891-5849(91)90174-2)
- 167. Sacheck, J.M.; Blumberg, J.B. Role of Vitamin E and Oxidative Stress in Exercise. *Nutrition* **2001**, *17*, 809–814. [\[CrossRef\]](http://doi.org/10.1016/S0899-9007(01)00639-6) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/11684385)

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