

Figure S1. Mn induced cell injury in SH-SY5Y. In order to estimate the toxic effect of Mn, we measured the LDH released into the culture medium and used CCK-8 to evaluate the cell viability, after the cells had been treated with 0, 50, 100, 200 μ M Mn for 0, 6, 12, 18, 24 h, respectively, cell viability was measured. (**A**) CCK-8 assay: after 200 μ M Mn exposure for 24 h, the viability of cells was decreased by 27.51% significantly (*p* < 0.01). (**B**) LDH release assay: after cells were exposed to 200 μ M Mn for 12, 18 and 24 h treatment period, the results showed significantly increased LDH levels in the culture medium (1.37-, 1.54- and 1.87-fold of control, respectively, *p* < 0.01) indicative of overt cytotoxicity. Data were mean ± S.D, n = 6, **p* < 0.05 and ***p* < 0.01 significantly different from the controls. However, cells treated with 200 μ M Mn within 24 h could induce significant cell damages but cell viability was above 50%.



Figure S2. Calpeptin could ameliorate Mn-induced cell injury and the over-activation of calpains. (**A**) Cells injury was detected with LDH and CCK-8 assays. In cells pretreated with 4 μ M calpeptin group, LDH release were significantly decreased compared with the cells only treated with 100 μ M Mn (22.05%, *P* < 0.05). Data showed that calpeptin reduced Mn-induced cytotoxicity, the same results were obtained for CCK-8. Data were mean ± S.D, n = 6. (**B**) [Ca²⁺]ⁱ in cells was calibrated from the measured fluorescence signals. [Ca²⁺]ⁱ was increased in 100 μ M Mn group (2.74-fold of control, *p* < 0.01), but had no obvious difference in calpeptin-pretreated groups compared with 100 μ M Mn-treated group. Calpains activity was undertaken measuring the values of absorbance. 100 μ M Mn-treated group resulted in calpains activity

increase (3.47-fold of control, p < 0.01). However, calpains activity decreased by 59.55% in cells treated with 4 µM calpeptin alone compared with the absolute controls (p < 0.05). Data were mean ± S.D., n = 6, *p < 0.05 and **p < 0.01 significantly different from the controls; *p < 0.01 and **p < 0.01 significantly different from 100 µM Mn-treated group.



Figure S3. *α*-Syn shRNA reduced *α*-Syn expression level and ameliorated cell injury during Mn exposure. (**A**) The mRNA and protein expression of *α*-Syn in LV-*α*-Syn shRNA cells were significantly decreased (48.18% and 39.14% of the normal cells, respectively, *p* < 0.01), and there was no difference between the normal cells and LV-negative shRNA cells. Expression of *α*-Syn gene and protein were normalized with β-actin gene and protein respectively (n = 4). (**B**) The protein products of *α*-Syn and β-actin. (**C**) After the normal, LV-*α*-Syn shRNA and LV-negative shRNA cells were respectively exposed to 100 µM Mn for 24 h, LDH releases and CCK-8 were performed (n = 4). LDH release in 100 µM Mn-treated LV-*α*-Syn shRNA cells decreased significantly (24.91%, *p* < 0.05) compared with the normal cells. However, the 100 µM Mn-treated LV-negative shRNA cells decreased significantly (30.81%, *p* < 0.01) versus the

untreated normal cells. As it had in CCK-8 assay, 100 μ M Mn-treated LV- α -Syn shRNA cells had a significant increase (17.43-fold, p < 0.05) compared with the normal cells, the 100 μ M Mn-treated LV-negative shRNA cells had a significant decrease (23.35%, p < 0.01) versus the untreated normal cells. Data were mean ± S.D, *p < 0.05 significantly different from the controls; *p < 0.05 and **p < 0.01 significantly different from 100 μ M Mn-treated group.



Figure S4. Co-localization fluorescence signals of VAMP-2 and α -Syn, VAMP-2 and synaptophysin in cells. (**A**) Representative colocalization of VAMP-2 (green) and α -Syn (red) in different groups. (**B**) Representative colocalization of VAMP-2 (green) and synaptophysin (red) in different groups. Scale bars: 50 µm.