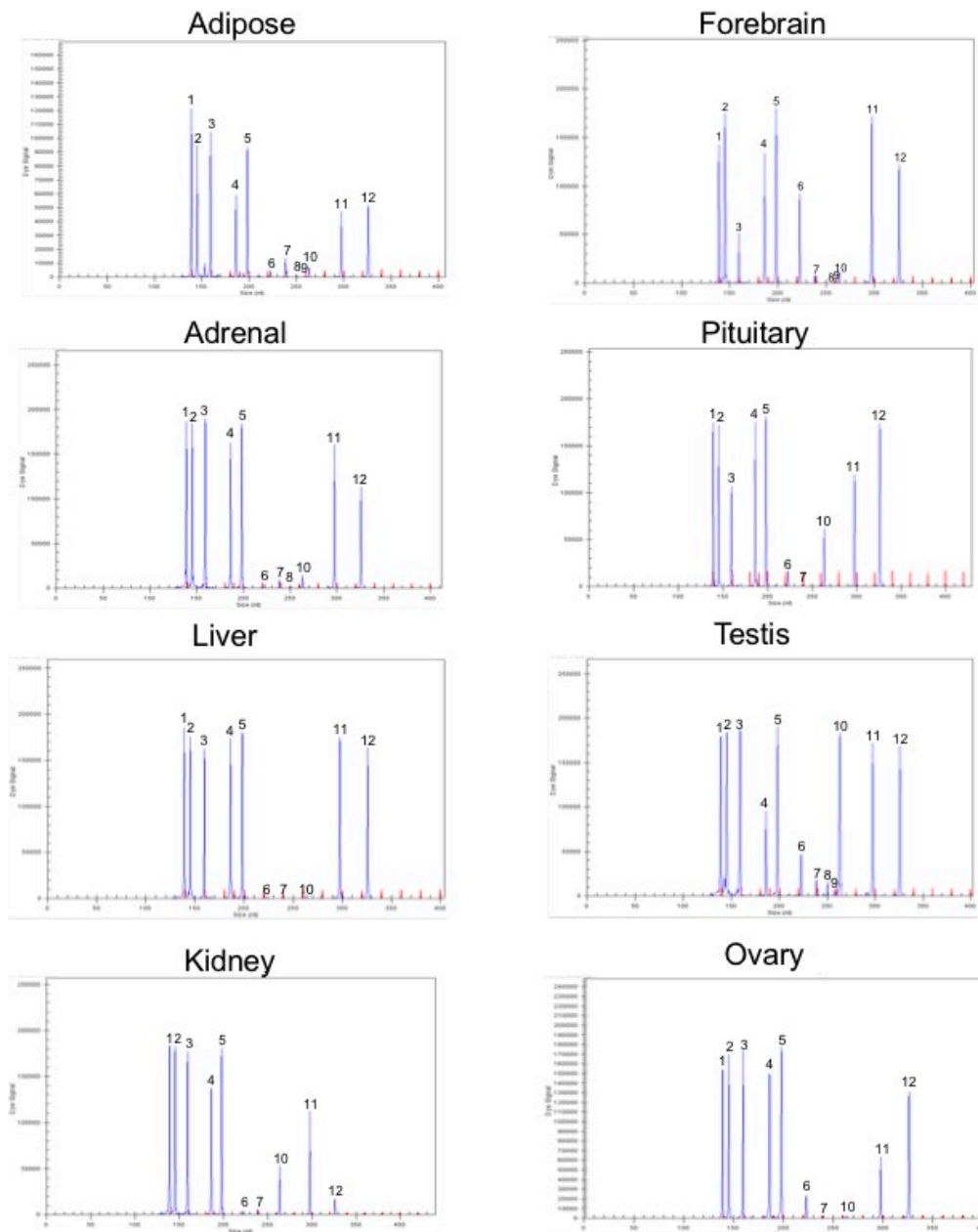
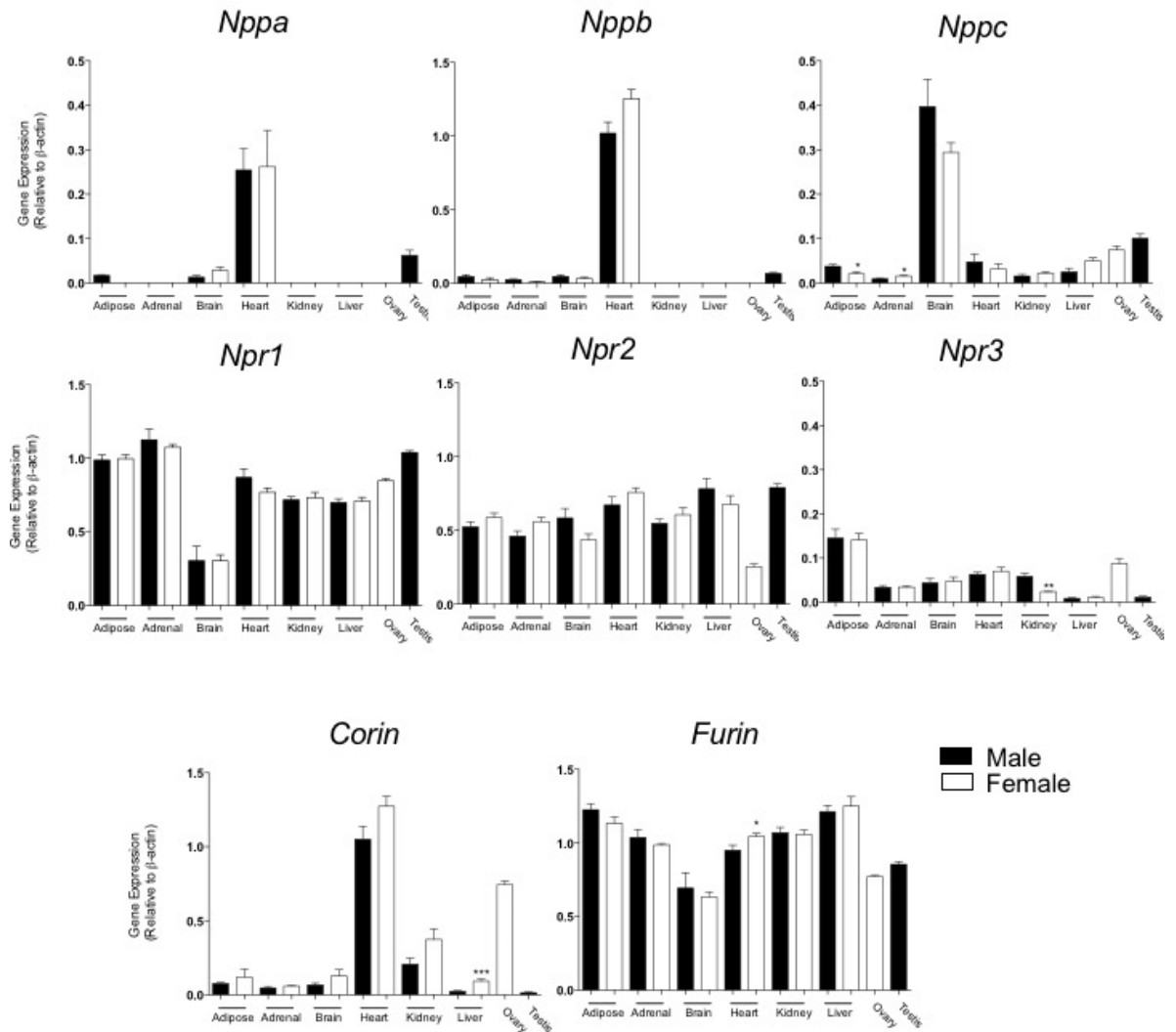


Gene	Forward primer (5'-3'), reverse primer (3'-5')	Product Size (bp)
<i>Nppa</i>	AGGTGACACTATAGAATACGGTACCGAAGATAACAGCC GTACGACTCACTATAGGGACAGAGTGGGAGAGGCAAGAC	256
<i>Nppb</i>	AGGTGACACTATAGAATACACCCAAAAAGAGTCCTTCG GTACGACTCACTATAGGGAAAAGAGACCCAGGCAGAGTCA	249
<i>Nppc</i>	AGGTGACACTATAGAATAAATACAAAGCGGCAACAAG GTACGACTCACTATAGGGACGTTGGAGGTGTTCCAGAT	221
<i>Npr1</i>	AGGTGACACTATAGAATACTTGAATTCTGAAGCAGC GTACGACTCACTATAGGGACTGGACATAGAGCAGGAGCC	158
<i>Npr2</i>	AGGTGACACTATAGAATACCTTGATGTCTTTGGGGAGA GTACGACTCACTATAGGGAGATTTGGGGTTCTCGGTAT	186
<i>Npr3</i>	AGGTGACACTATAGAATATCTGCTGTCTCTGTCCCTT GTACGACTCACTATAGGGACTGGTTTTGAAGGGCATCAT	235
<i>Furin</i>	AGGTGACACTATAGAATAGGCTTTCATGACAACCCATT GTACGACTCACTATAGGGAGGTCAGCGTCCCATAGTTGT	137
<i>Corin</i>	AGGTGACACTATAGAATAGAATCTTCCATTCCGCAA GTACGACTCACTATAGGGATATCAATGAGGCAAATGGCA	263
<i>β-Actin</i>	AGGTGACACTATAGAATAGTACCACCATGTACCCAGGC GTACGACTCACTATAGGGAGTACTTGCCTCAGGAGGAG	144
<i>cFos</i>	AGGTGACACTATAGAATACTGTCCGGTTCCTTCTATGC GTACGACTCACTATAGGGAAGTACAGGTGACCACGGGAG	151
<i>cJun</i>	AGGTGACACTATAGAATATAACAGTGGGTGCCAACTCA GTACGACTCACTATAGGGATGTCGCAACCAGTCAAGTTC	165
<i>Egr1</i>	AGGTGACACTATAGAATAGGTGGAGACGAGTTATCCCA GTACGACTCACTATAGGGAAGGCCACTGACTAGGCTGAA	172
<i>Nr5a1</i>	AGGTGACACTATAGAATACCCAGAGGATACCATGAGA GTACGACTCACTATAGGGAGATAAATACCAGCCCAGCCA	242
<i>Nr0b1</i>	AGGTGACACTATAGAATATCCTGTACCGCAGCTATGTG GTACGACTCACTATAGGGACCACCTGTGGATCCTTGAGT	214
<i>β-Actin</i>	AGGTGACACTATAGAATAGTACCACCATGTACCCAGGC GTACGACTCACTATAGGGAGTACTTGCCTCAGGAGGAG	144
<i>Gnrhr</i>	AGGTGACACTATAGAATACTTGATACAGGGCAAGCTCC GTACGACTCACTATAGGGAGCACCTTCCTTGAGAGC	207
<i>KanR</i>	AGGTGACACTATAGAATAATCATCAGCATTGCATTCCGATTCTGTTTG GTACGACTCACTATAGGGAATCCGACTCGTCCAACATC	325

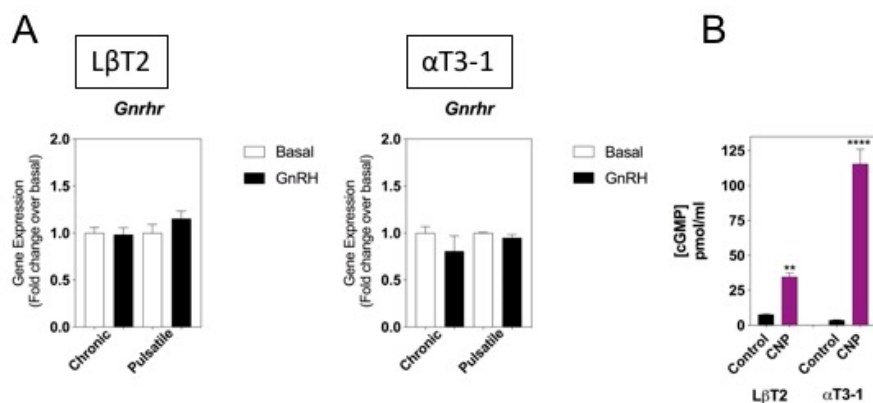
Supplemental Table S1: Primer sequences for multiplex RT-qPCR assays for natriuretic peptide genes, and gonadotrope transcription factor genes. Murine mRNA sequences were obtained from NCBI Nucleotide (<https://www.ncbi.nlm.nih.gov/nucleotide>), were imported into express Designer Software (Beckman Coulter).



Supplemental Figure S1: Representative electropherograms of multiplex RT-qPCR analyses of expression for natriuretic peptides (*Nppa*, *Nppb*, *Nppc*), natriuretic peptide receptors (*Npr1*, *Npr2*, *Npr3*), and proconvertase enzymes (*Furin* and *Corin*) (blue peaks). Total RNA was extracted from adipose, adrenal, liver, kidney, forebrain, pituitary, testis and ovary. Genes were detected in order of size corresponding to size standards run alongside the products (140nt-420nt) (red peaks) and quantified by capillary electrophoresis.



Supplemental Figure S2: Multiplex RT-qPCR data of natriuretic peptide gene expression from murine adipose, adrenal, liver, kidney, forebrain, pituitary, testis and ovary. Tissue was collected from n=5 to n=8, 12 week-old C57/B6 males or females and total RNA extracted.



Supplemental Figure S3 A) Effect of continuous or pulsatile GnRH stimulation on *Gnhr* expression in L β T2 and α T3-1 cell lines. Cells were treated with 0 or 100nM GnRH, for either 4hr continuously, or as 5 min pulses every hour for 4hr, before extracting RNA and performing multiplex RT-qPCR for *Gnhr* (as part of the same multiplex described in Figure 4). Data shown are means \pm SEM (n= 6 to 9 individual RNA extractions) of relative gene expression (normalized to

ActB). **B)** Total cGMP accumulation in L β T2 and α T3-1 cells treated with 0 or 100nM CNP for 1h in physiological saline solution containing 1mM IBMX, before measuring with a commercially available cGMP-EIA kit (R&D Systems) as described previously [8]. Data shown are means \pm SEM representative of three independent experiments, each performed in triplicate. **P<0.01, ****P<0.0001, significantly different from Control cells.