## 1 **Supporting Information**



2 **S1** Calculated ideal surface areas of lipid mixtures



 The experimental data for the surface pressure and surface area were taken from the compression isotherms at specific pressure points of 10, 20, and 30 mN/m. To calculate the theoretical values at the three surface pressures for the lipid mixtures with differing cholesterol content in the monolayer, the isotherms of just lipids (0 % cholesterol) and pure cholesterol (100 %) were taken, and in dependency of the molar ratio of cholesterol content added. 11 Following  $\bar{A} = x_{chol} * A_{chol} + x_{lipids} * A_{lipids}$  with  $\bar{A}$  being the average molecular area of the 12 monolayer,  $A_{chol}$  and  $A_{lipids}$  are the surface areas of the pure cholesterol and pure lipid isotherm, respectively.  $x_{chol}$  is the molar ratio of cholesterol in the mixture and  $x_{lipids}$  for the lipid mixture without cholesterol. The calculated average surface areas of all three cholesterol contents are located at higher surface areas in comparison to the experimental values. This demonstrates the condensing effect of cholesterol in the monolayer of lipid mixtures.

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#### 19 **S2** Calculation of excess free energy of mixing



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**Figure S2**. Calculated excess free energies of mixing ∆G<sup>excess</sup> as a function of mole fraction of cholesterol xcholesterol 22 from monolayers with different cholesterol content at surface pressures of 10 mN/m (left), 20 mN/m (middle) and 23 35 mN/m (right).

The excess free energy of mixing is calculated by  $\Delta G^{excess} = \int_0^{\pi} A_{mix} - (x_{chol} A_{chol} +$ 0 24 25  $x_{lipids}A_{lipids}d\pi$  for not ideal mixing. Where  $x_{chol}$  is the molar ratio of cholesterol in the 26 mixture and  $x_{lipids}$  for the lipid mixture without cholesterol.  $A_{mix}$  is the surface area of the

27 mixed monolayer,  $A_{chol}$  and  $A_{lipids}$  are the surface areas of the pure cholesterol and the pure

- lipid isotherm, respectively.  $\Delta G^{excess}$  was calculated for three different surface pressures: 10, 20, and 35 mN/m. All values for the excess free energy of mixing show a tendency to be 30 negative or approximately 0 kJ/mol.  $\Delta G^{excess}$  values are negative when the monolayer is condensing, resulting from attractive forces between unlike molecules and therefore the 32 process of mixing is thermodynamically favored. High positive values of Δ*G<sup>excess</sup>* suggest the immiscibility of the monolayer mixture with a possible phase separation. The excess free energy of mixing of monolayers with MBP was not calculated because the compression isotherms start at higher surface pressures which are not suitable for calculations.
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### **S3** Comparison of different injection pressures for MBP



 **Figure S3a**. Compression isotherms of 44 % cholesterol in monolayers at different injections points (left) and time-dependent surface pressure curve with injection of MBP at 20 mN/m (right).

- The fast surface pressure increase followed by a slower pressure relaxation directly after the
- MBP injection under the lipid monolayer at 20 mN/m (Fig. 3a right) might be an
- experimental artifact as the protein was injected with a needle through the lipid film. After a
- short lag time the surface pressure gradually increased due to the MBP interaction with the
- lipid monolayer.



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- **Figure S3b**. Representative fluorescence microscopy image of a lipid monolayer with 44 % cholesterol at 25
- mN/m with MBP injected at 20 mN/m (left) and 0 mN/m (right).
- The fluorescence images in Fig. 3b depict the comparison of the two injection points of MBP
- at 20 and 0 mN/m at a surface pressure of approximately 25 mN/m. Both images show large
- domains, which are not perfectly circular anymore.

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## 55 **S4** Compressibility of monolayers







58 The compressibility  $\kappa$  of the monolayers was calculated from the isotherms of Fig. 1 and 2 with  $k = -\frac{1}{4}$  $\overline{A}$ dA  $K = -\frac{1}{4}\frac{u}{d\pi}$ . The compressibility of the monolayers with different cholesterol content shows no phase transition without MBP. But with MBP injected under the monolayers, there is a clear change in compressibility between 20 to 35 mN/m for all mixtures. This change presumably comes from the squeeze-out of the protein. The second change in compressibility at lower surface pressures (5-17 mN/m) was not examined in clearly detail. It could be a phase transition originating from cholesterol because it is most pronounced in the pure cholesterol isotherm, but could also be an artefact of the monolayer because it is a very broad transition.

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- 68 **S5** Surface area differences of monolayers with different cholesterol content and MBP.
- **Table S5**. calculated surface area difference between 20 and 35 mN/m of monolayers with different cholesterol content and MBP. content and MBP.



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 The compression isotherms of 44 % cholesterol with 0.05 mol% Rhodamine-DHPE (red) or 1 80 mol% TopFluor® Cholesterol (green) show no significant deviation from the 44 % cholesterol lipid monolayer without dye (black). Mixing the fluorescent dye to the lipid mixture should have a neglectable influence on the behavior of the monolayers.

**S7** Fluorescence microscopy image of TopFluor® Cholesterol



 **Figure S7**. Representative fluorescence microscopy image of a lipid monolayer with 44 % cholesterol and 1 mol% TopFluor® Cholesterol at ca. 10 mN/m.

- The bright areas are cholesterol-rich domains with the dye and the dark area is the
- phospholipid-rich liquid-expanded phase. The image is exactly inverted to the fluorescence

**Field Code Changed**









With MBP



- *10% cholesterol:*
- Without MBP















Without MBP









With MBP













With MBP



*60% cholesterol:*

Without MBP









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 **Figure S10.** Representative fluorescence microscopy images of lipid monolayer with 44 % cholesterol content and 0.05 mol% Rh-DHPE with a 20x magnification.

# **S11** Table of surface pressures for all fluorescence images



