





Review

# Green Biomass-Based Protein for Sustainable Feed and Food Supply: An Overview of Current and Future Prospective

Éva Domokos-Szabolcsy <sup>1</sup>, Seckin Reyhan Yavuz <sup>1</sup>, Edgard Picoli <sup>2</sup>, Miklós Gabor Fári <sup>1</sup>, Zoltán Kovács <sup>1</sup>, Csaba Tóth <sup>1</sup>, László Kaszás <sup>1</sup>, Tarek Alshaal <sup>1,3,\*</sup> and Nevien Elhawat <sup>1,4,†</sup>

<sup>1</sup> Department of Applied Plant Biology, University of Debrecen, Böszörményi Str. 138, 4032 Debrecen, Hungary

<sup>2</sup> Department of Plant Biology, Federal University of Viçosa, Viçosa 36570-900, Brazil

<sup>3</sup> Department of Biological and Environmental Sciences, Faculty of Home Economic, Al-Azhar University, Tanta 31732, Egypt

<sup>4</sup> Soil and Water Science Department, Faculty of Agriculture, Kafrelsheikh University, Kafr El-Sheikh 33516, Egypt

\* Correspondence: [alshaal.tarek@agr.unideb.hu](mailto:alshaal.tarek@agr.unideb.hu)

† These authors contributed equally to this work.

**Abstract:** It is necessary to develop and deploy novel protein production to allow the establishment of a sustainable supply for both humans and animals, given the ongoing expansion of protein demand to meet the future needs of the increased world population and high living standards. In addition to plant seeds, green biomass from dedicated crops or green agricultural waste is also available as an alternative source to fulfill the protein and nutrient needs of humans and animals. The development of extraction and precipitation methods (such as microwave coagulation) for chloroplast and cytoplasmic proteins, which constitute the bulk of leaf protein, will allow the production of leaf protein concentrates (LPC) and protein isolates (LPI). Obtained LPC serves as a sustainable alternative source of animal-based protein besides being an important source of many vital phytochemicals, including vitamins and substances with nutritional and pharmacological effects. Along with it, the production of LPC, directly or indirectly, supports sustainability and circular economy concepts. However, the quantity and quality of LPC largely depend on several factors, including plant species, extraction and precipitation techniques, harvest time, and growing season. This paper provides an overview of the history of green biomass-derived protein from the early green fodder mill concept by Károly Ereky to the state-of-art of green-based protein utilization. It highlights potential approaches for enhancing LPC production, including dedicated plant species, associated extraction methods, selection of optimal technologies, and best combination approaches for improving leaf protein isolation.

**Keywords:** biomass crops; RuBisCo; green-based protein; phytochemicals; vitamins; agrowastes



**Citation:** Domokos-Szabolcsy, É.; Yavuz, S.R.; Picoli, E.; Fári, M.G.; Kovács, Z.; Tóth, C.; Kaszás, L.; Alshaal, T.; Elhawat, N. Green Biomass-Based Protein for Sustainable Feed and Food Supply: An Overview of Current and Future Prospective. *Life* **2023**, *13*, 307. <https://doi.org/10.3390/life13020307>

Academic Editors: Karaffová Viera, Zuzana Ševčíková and Viera Almášiová

Received: 22 December 2022

Revised: 17 January 2023

Accepted: 18 January 2023

Published: 22 January 2023



**Copyright:** © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

Humans and animals consume protein mainly as a source of nitrogen and proteinogenic amino acids required for the synthesis of new biomolecules and as an energy source, to a lesser extent, providing four calories per gram of protein. The quality and nutritional value of protein depend heavily on its amino acid composition, particularly with regard to the nine indispensable amino acids (i.e., phenylalanine, valine, threonine, tryptophan, methionine, leucine, isoleucine, lysine, and histidine) [1].

Protein-energy malnutrition (kwashiorkor) is still the primary worldwide health problem. After World War II, the nutritional problems in developing countries received increased attention, and several low-income countries reported cases of severe kwashiorkor. Consuming protein comes first on the list of priorities. The discovery of novel protein sources, many of which came from unexpected assets, such as single-cell proteins, fish protein concentrates, and leaf protein concentrates, were prompted by this, as were efforts to add extra protein to meals [2].

The proteins from green leaves were first isolated in 1773 by the French biochemist H.M. Rouelle, although the proteins were not fully understood at the time [3]. He used a simple mortar and pestle to press the juice of the leaves. From the green juice, he obtained a green curd by heating it until his finger could no longer be held in the solution. Then the green curd was filtered off, and the brown liquid was further heated, which gave a clear curd. The isolation methods have evolved considerably since then, but the basic principles are still similar; the green juice is extracted from the leaves, the green fraction is removed, and the leaf protein is purified and concentrated as leaf protein concentrate (LPC). The majority of LPC has a biological value between 70 and 83 [4,5], which is comparable to soybean, sunflower seed, and cotton seed meals. According to studies, LPC has a real digestibility between 80 and 90, which is quite comparable to high-quality animal and vegetable proteins [6]. The main issues with storing LPC are microbial breakdown and the onset of rancidity [5].

## 2. Data from the History of Green Biomass as a Protein Source: The Forgotten Early Visions about the Establishment of the Very First Green Protein Biorefinery Factory in Hungary and England: The Green Fodder Mill Concept of Károly (Karl) Ereky

The director of the Science Museum in London, Robert Bud, published that the word “biotechnology” was coined by the mechanical engineer, agricultural economist and minister of nutrition in Hungary, Károly (Karl) Ereky in 1919 [7–10]. Ereky was a witty, ingenious and creative man. Personally, he was well versed in interdisciplinary happenings and anticipated the achievements as well trends of the development of biotech-based agricultural bio-industry [11–14].

Interestingly, Ereky started his pioneer biotech projects with the establishment of the so-called “Green Fodder Mill” factory concept in the spring of 1917. Ereky obtained finely ground green pulp from the leaves of green fodder plants, such as alfalfa, clover, and grasses. This could be wetly fractionated and/or dehydrated to provide feed and food of high protein and vitamin content, the so-called “green plasma-conservé” and “green flour”, and other commodities [15]. Experimental feeding indicated that the physically processed, stripped from cell walls, green leaves, and young shoots furnished concentrates with a “complete amino-acid composition”, “high-vitamin content”, and “100% digestible” concentrates. Ereky contended that his invention would revolutionize the industrial-scale feeding practices of pig, chicken, and dairy farms. He proved that the use of his green plasma-conservé could save about 40% of feed, especially cereal grain used in pig farms. Moreover, the feeding cycle could be shortened by 20–30%, and egg production capacity was increased to 2.5 times that of laying hen and duck, milk production was improved, and the disease resistance of horses or other domestic animals was optimal [12–14].

The third classic work of Ereky appeared in Budapest in 1925 under the title: “*The Green Fodder Mill and the Large Scale Industrial Animal Farms*” [12–14,16,17]. This book deals with his pioneering research on protein concentrate that could be extracted from the green leaves of plants, otherwise known as leaf protein concentrate (LPC). Its main purpose was the application of the principle of biotechnology in large-scale dairy, milk, and meat or fat-producing farms and “*in the service of the people’s food*” whose theoretical bases had been outlined in the former book “*Biotechnologie*” [15,18,19]. Ereky characterized his process: “*The dried plasma preserve is eminently excellent food for human beings: it contains all the vitamins, inorganic salts and complete albumen-substances in such quantities and of such quality as no other foodstuff does, and moreover easily digestible up to 100%. As in the case of the fresh pulp, the plasma preserve can be rendered more appetizing by the addition of chocolate, sugar, fruit-juice, etc. As a supplementary portion of the human digestive apparatus, the Green Pulper realizes the ancient dream of the vegetarian, and enables man to render himself independent of the consumption of flesh meat*” [16].

After many wet fractionation experiments carried out on alfalfa and grasses, some new protocols and different size green pulper machines were developed and patented in Hungary, Britain, and in some other countries between 1924 and 1928 [20–26]. From 1925, Ereky promoted the huge possibilities of protein production from plant leaves for human consumption [27].

Feeding experiments proved the value of those products of high protein and vitamin content, which means a relatively cheap mass-concentrate, and it was hoped that a new branch of the industry could be founded and developed on the basis of that method. Ereky calculated that the manufacture of 2 to 2.5 tons per hectare of cheap vegetal protein is expected in an industrial system. He stated that the introduction of the “*Ereky-process*” would increase the productivity of the unit agricultural area by a factor of 2–3 times in living weight of pork meat (i.e., 1000 kg ha<sup>-1</sup>), compared with the traditional level (300–600 kg ha<sup>-1</sup>). Similarly, he calculated that a stock of a productive dairy farm fed by a combination with the green protoplasm conservation might attain that of a traditional herd grown on an area of 150,000 ha extensive pasture land but need 12,000 irrigated fields for supplying milk to a population of one million people. He emphasized that this increment was possible only by means of the application of the principles of biotechnology, as the net “level of efficiency” of milk production was increased from 6.5% to 40%. In pig farms, a 7–8 kg dry mixture of cereal was needed for 1 kg of meat live weight, but in the future, a mixture of 3 kg of cereal supplemented with a small amount of green plasma preserve would yield the same live weight [13,14].

Ereky’s British connections began in the early 1920s, with correspondence with the Department of Agriculture of the University of Cambridge. Some of this was preserved by the pioneer of the leaf protein research and pioneer of tobacco mosaic virus (TMV) research, Norman Wingate Pirie (1907–1997), who later donated them to The Science Museum in London. Wishing to promote both science and business, in the late spring and summer of 1927, Ereky traveled to Britain and presented his principles and methods to the authorities of the Ministry of Agriculture and Ministry of Health and others from Britain, Canada and Australia [13,14,28–30].

Practical demonstration activities were also held in Britain with his smaller green pulper. These activities were followed by further pig fattening experiments lasting several months at the Lord Melchett Court (Romsey, Hampshire) organized by Nitram Ltd. (Imperial Chemical Industry, ICI [31]). These were said to prove that “*the nutritive value of one-kilogram young alfalfa juice is equivalent with 2 kg cream-free separated milk*”. The recognition received from an observer in the Ministry of Health was even more enthusiastic, “*the grinding of green stew for human consumption would be even more important than the mash of fodder grass from the point of view of supplying the population*”. Professional opinions emphasized uniformly that with the help of the “*Ereky-process*”, healing products could be developed, which were able “to reorganize the catering tasks of the population of England” [30]. More ambitious, Ereky planned to develop a mass-protein-concentrate process, which could be produced economically in the tropics and could be transported easily to large distances as a green plasma conserve [16]. During his visit to London, the representatives of the Australian government asked him to elaborate on this project. In that country, 4 million sheep were lost annually to temporary drought in semiarid areas, while abundant green fodder grew near the humid seashore [30].

On returning to Hungary, Ereky proposed in a March 1928 lecture in the presence of the Hungarian President, Miklós Horthy, that Hungarian agriculture should also be reconstructed by means of the leaf protein concentrate program based on green plasma-preserve produced by his green fodder mill system [30]. It is important to mention that the pioneer papers published by Ereky presented above were not cited by some researchers that followed him in Great Britain and in the USA and approached the wet fractionation process [32–36]. Moreover, Ereky’s British patent was cited by Norman Wingate Pirie in his earliest leaf protein papers published during the Second World War [37–39].

### 3. Green Source of Protein

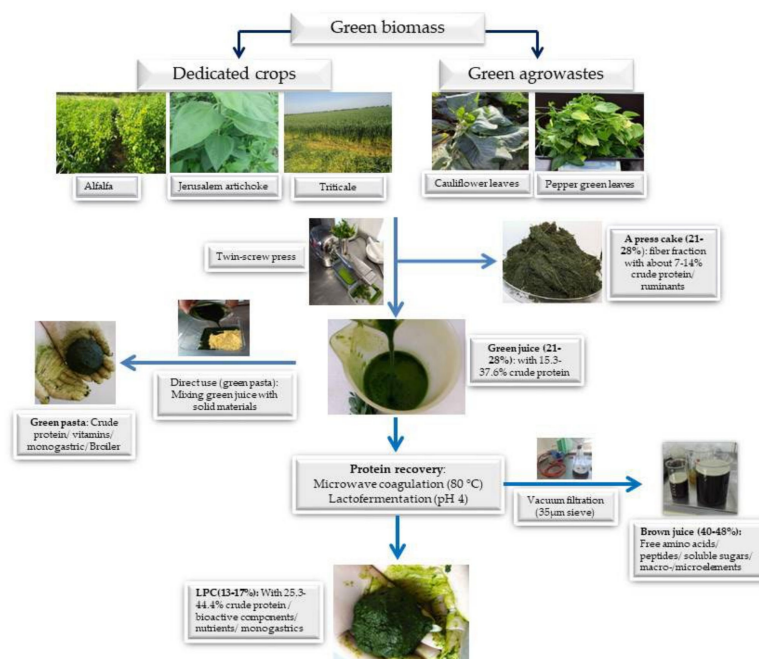
The photosynthesis process occurs mainly in the leaf thylakoid membranes and requires about 70 different proteins [40]. Leaf cytoplasmic protein represents about 20% of leaf protein, while less than 5% and 1.2% of leaf protein are located in mitochondria and cell nuclei, respectively [40]. Kromus et al. [41] identified 250–300 unique proteins and polypep-

tides in green plant extracts. Photosynthetic protein complexes in the thylakoid membrane represent the majority of the water-insoluble leaf proteins; however, water-soluble leaf proteins also exist [40,42].

Rubisco (ribulose 1,5-bisphosphate carboxylase/oxygenase) is the most prevalent protein in the world, as it is a crucial enzyme for fixing CO<sub>2</sub> during photosynthesis [40,42]. Rubisco accounts for the majority of the soluble protein component of leaves and up to 50% of their total protein content in C<sub>3</sub> plants. A modestly sized enzyme, Rubisco is hexadecameric and has a molecular weight of about 550 kDa. It is a stroma-resident protein and catalyzes the onset of photosynthetic activity. It consists of eight big (55 kDa) and eight small subunits (15 kDa); nevertheless, plants synthesize much more Rubisco in their leaves due to its poor catalytic efficiency [43].

Free amino acids, oligopeptides, and enzymes involved in the synthesis of lipids, proteins, carbohydrates, and other compounds represent the rest part of the soluble leaf proteins [40,44]. Because of this, plant leaves provide a variety of proteins that are typically consumed by cattle that are fed while grazing but may also be utilized for monogastric animal feeding. Green and white proteins are found in different proportions in green leaves, with Rubisco (referred to as fraction I protein) making up the majority of the white proteins [45].

The basic principles of leaf protein extraction are illustrated in Figure 1.



**Figure 1.** Scheme of basic idea of isolation leaf protein and its related by-products from green biomasses.

### 3.1. Dedicated Plant Species

The source of green biomass can be plants grown dedicated to this purpose or green waste as a by-product of vegetable/agricultural crop production. The most commonly used species of crops that can be targeted for green biorefining, including leaf protein concentrates (LPC) preparation, are perennial and annual legumes and grasses (Table 1). At the same time, aquatic crops, such as duckweed, also have great potential [46].

Alfalfa is a regional forage plant that is farmed to a high standard and is GMO-free throughout the European Union [47]. Further, it offers additional ecosystem support services [48], such as improved soil fertility, reduction or avoidance of nitrogen fertilizer use, and pest and disease management [49]. Alfalfa has the ability to provide large yields of dry matter and crude protein per acre in temperate regions [50]. Alfalfa can be used as a source of local protein for a variety of farm animals, which can then produce milk, meat, and eggs for human use.

Due to its availability and balanced amino acid profile, soybean is currently the most popular protein feed. Only 56% of the crude protein utilized in organic farming in Europe is indigenous to the continent [51]. Unlike soy, alfalfa requires less heat and water and can be grown in many parts of the world, including many European regions [52].

Jerusalem artichoke (JA) is a well-known crop because of its inulin-rich tubers; notwithstanding, a substantial amount of LPC can be produced from its fresh aerial biomass. The crude protein content in JA-derived LPC was about 33.3% (on a dry basis), and about 53 valuable bioactive phytochemicals were identified, including dimethoxy-tetrahydroxyflavone, dihydroxy-methoxyflavone, hymenoxin, nevadensin, Butein, kukulkanin B, and liqueritigenin [53].

Vetches (*Vicia* spp.) are legumes that are well adapted to the winter season, and therefore they are widely cultivated in West Asia, North Africa, Australia, and Turkey. Vetches can grow on a variety of soil types and are used for a variety of things, including dry matter, silage, and green manure [54]. The vetches spread because of their weak and slender stem. As a result, harvest becomes challenging, and the forage and quality of the plant decline as leaves fall off. In order to prevent spreading, vetch should be mixed with grains before being sowed. Besides increasing the forage yield, the intercropping of vetches with either grasses or cereals provides physical support facilitating its mechanical harvest. Moreover, this mixture might have an additional benefit through increasing leaf protein yield and quality [55] and should be explored.

Triticale, a hybrid of wheat and rye whose importance is rising globally and which is currently cultivated on more than 400,000 ha, is a perennial plant. When it was first cultivated, its acreage exhibited an upward tendency, had a significant decline in the late 1970s, and has been augmenting since then [56]. Its significance is due in part to its transient character but also due to the important ingredients it contains. Otherwise, triticale has an equivalent value as a plant for making bread and feed. However, in the overall assessment, it is primarily regarded as a grain for fodder. The protein content of triticale might be stated first in terms of its content value.

The crude protein content in the whole triticale plant, fertilized with the standard N-fertilizer (180 kg N ha<sup>-1</sup>), varied from 10.9 to 8.3% when green plants were harvested on 6 June and 6 July, respectively, while plants harvested on 15 August showed a crude protein content of 4.5% in their straw [57]. However, the crude protein percentage increased upon increasing the rate of applied N-fertilizer to 300 kg N ha<sup>-1</sup>.

According to Jørgensen et al. [58], about 57–74% of the total crude protein in the aboveground green biomass of triticale can be extracted by mechanical pressing into green juice from which 51–63% can be thermally coagulated into an LPC. Similar results of LPC obtained from other green biomass crops were also reported [59,60]. The efficiency of protein extraction or isolation from the green biomasses showed a dependence on plant species rather than the maturity stage/harvest time [61]. On the contrary, Solati et al. [62] found a slight decrease in leaf protein extractability from two grass species; however, this may be attributed to the last harvest at the flowering stage.

Exogenous N-application positively enhanced the protein extraction efficiency while it lowered the precipitation efficiency. However, this cannot be generalized since the crude protein is usually calculated based on the total N content, and there is a big portion of N that is not a real protein, including nitrate, amino acids, or peptides. These compounds are extractable since they are mostly water-soluble, but they cannot be thermally precipitated [58].

Triticale yielded about 730 kg LPC per hectare, which was 40% less compared to grain protein yield. This could be attributed to the low precipitation efficiency of protein in the juice; therefore, there is an urgent need to search for novel technologies to enhance protein recovery [63,64].

Nevertheless, the total protein yield of LPC should not be the only ultimate goal. Another important factor that should be considered is the quality of the harvested protein. For instance, grains of triticale and wheat mainly contain storage proteins that are rich in glutamine, glutamate, and proline [65], while leaf protein showed higher contents of limited indispensable amino acids (e.g., lysine, cysteine, and methionine) by 20–25% than grain protein, which makes it suitable protein source for pigs, broilers, and monogastrics as soybean meal.

**Table 1.** Summary of leaf protein yields from different plant species and green agrowastes achieved by varied protein extraction techniques.

Plant Species	Crude Protein Content of Leaf Protein Concentrate (LPC) m/m%	Method of Protein Content Measurement	Protein Isolation Method	Reference
<b>Dedicated plant species</b>				
Alfalfa ( <i>Medicago sativa</i> )	40.3	Kjeldahl	Microwave coagulation	own results (unpublished data)
Alfalfa ( <i>Medicago sativa</i> )	41.0	Kjeldahl	<i>Lactobacillus salivarius</i> fermentation	[66]
Alfalfa ( <i>Medicago sativa</i> )	40.5 (Green LPC) 32.3 (White LPC)	Kjeldahl/Dumas	Thermal coagulation using two-step heating	[67]
Alfalfa ( <i>Medicago sativa</i> )	46.0	unknown	Thermal coagulation	[59]
Alfalfa ( <i>Medicago sativa</i> )	37.8–47.4	Dumas	Acid coagulation	[68]
Jerusalem artichoke ( <i>Helianthus tuberosus</i> )	33.4	Kjeldahl	Microwave coagulation	[57]
Perennial Rye grass ( <i>Lolium perenne</i> )	33.9	Kjeldahl	Thermal coagulation	[69]
Perennial Rye grass ( <i>Lolium perenne</i> , variety; Trocadero and Calvano)	24.5 (Green LPC) 22.8 (White LPC)	Kjeldahl/Dumas	Thermal coagulation using two-step heating	[67]
Ryegrass ( <i>Lolium perenne</i> )	50.7	unknown	Thermal coagulation	[59]
Red clover ( <i>Trifolium pratense</i> )	40.0	Kjeldahl	<i>Lactobacillus salivarius</i> fermentation	[66]
Red clover ( <i>Trifolium pratense</i> L., variety; Rajah and Suez)	34.6 (Green LPC) 35.6 (White LPC)	Kjeldahl/Dumas	Thermal coagulation using two-step heating	[67]
White clover ( <i>Trifolium repens</i> L., variety; Klondike and Silvester)	40.4 (Green LPC) 45.1 (White LPC)	Kjeldahl/Dumas	Thermal coagulation using two-step heating	[67]
Clover and grass mix ( <i>Trifolium pratense</i> and <i>Lolium multiflorum</i> )	40.0	Kjeldahl	<i>Lactobacillus salivarius</i> fermentation	[66]
Clover and grass mix	47.0	unknown	Thermal coagulation	[59]
<b>Green agrowastes</b>				
Green pepper ( <i>Capsicum annuum</i> )	31.2	Kjeldahl	Microwave coagulation	own results (unpublished data)
Green pepper ( <i>Capsicum annuum</i> )	26.2	Kjeldahl	Lactic acid fermentation	own results (unpublished data)
Horseradish ( <i>Armoracia rusticana</i> )	25.3	Kjeldahl	Microwave coagulation	own results (unpublished data)
Horseradish ( <i>Armoracia rusticana</i> )	24.7	Kjeldahl	Lactic acid fermentation	own results (unpublished data)

Table 1. Cont.

Plant Species	Crude Protein Content of Leaf Protein Concentrate (LPC) m/m%	Method of Protein Content Measurement	Protein Isolation Method	Reference
Forage soy ( <i>Glycine max</i> )	41.9	Kjeldahl	Microwave coagulation	own results (unpublished data)
Forage soy ( <i>Glycine max</i> )	37.1	Kjeldahl	Lactic acid fermentation	own results (unpublished data)
Triticale ( $\times$ <i>Triticosecale</i> )	41.0	Kjeldahl	Microwave coagulation	own results (unpublished data)
Triticale ( $\times$ <i>Triticosecale</i> )	34.1	Kjeldahl	Lactic acid fermentation	own results (unpublished data)
Broccoli ( <i>Brassica oleracea</i> , <i>Italica</i> )	35.3	Kjeldahl	Microwave coagulation	[70]
Broccoli ( <i>Brassica oleracea</i> , <i>Italica</i> )	39.2	Kjeldahl	Lactic acid fermentation	[70]
Cauliflower ( <i>Brassica oleracea</i> , <i>var. botrytis</i> )	44.4	Kjeldahl	Microwave coagulation	own results (unpublished data)
Cauliflower ( <i>Brassica oleracea</i> , <i>var. botrytis</i> )	43.1	Kjeldahl	Lactic acid fermentation	own results (unpublished data)
Brussels sprouts ( <i>Brassica oleracea</i> , <i>var. gemmifera</i> )	37.4	Kjeldahl	Microwave coagulation	own results (unpublished data)
Brussels sprouts ( <i>Brassica oleracea</i> , <i>var. gemmifera</i> )	34.4	Kjeldahl	Lactic acid fermentation	own results (unpublished data)
Potato haulm ( <i>Solanum tuberosum</i> )	45.3	unknown	Thermal coagulation	[71]
Sugar beet ( <i>Beta vulgaris</i> )	31.3–41.1	Dumas	Thermal coagulation	[60]
Sugar beet ( <i>Beta vulgaris</i> )	31.1–37.6 (White LPC); 43.6–47.7 (White LPC)	Kjeldahl	Thermal coagulation	[72]
Broccoli ( <i>Brassica oleracea</i> , <i>Italica</i> )	27.2 (White LPC); 30.4 White LPC	Dumas	Thermal coagulation	[73]
Kale ( <i>Brassica oleracea</i> , <i>var. Sabellica</i> )	16.7 (White LPC); 30.4 (White LPC)	Dumas	Thermal coagulation	[73]
Cassava ( <i>Manihot esculenta</i> )	40.4–45.1	Dumas	Thermal coagulation, acid precipitation and spontaneous fermentation	[74]

### 3.2. Green Agrowastes

Agro-industrial green waste and by-products could serve as readily accessible, affordable, and environmentally friendly sources of plant-based proteins, which can have a marked effect on the decrease of food waste, supporting zero waste initiative and circular economy idea [75].

By using a conventional procedure, LPC can be produced from different plant residues, such as aboveground parts, after removing the fruits (Tables 1–3). The yield of LPC varied largely according to plant species. For example, carrot top yielded 112 kg ha<sup>-1</sup>, while leaves of cucumber, potato, and tomato produced 1500 kg ha<sup>-1</sup> with a protein content ranging between 22.5% (carrot) and 50% (potato). The amino acid composition of produced LPC was similar to those obtained from other plant species.

Proteins differ in their nutritional values, although they are made of similar amino acids. The biological values of potato- and cucumber-derived proteins were 42 and 59%, respectively. During broccoli cultivation, 90% of the above-ground green of the plant is converted to agrowaste, and only 10% enters the food chain. Additionally, it can be used to produce valuable protein concentrate with important nutrients (Tables 1, 3 and 4) through valorization, including fermentation and thermo/microwave coagulation [70,73,76]. Similarly, a side stream of kale is also suitable for green protein concentration purposes [77]. Without being exhaustive, it can be mentioned the Manihot plant (*Manihot esculenta*), which also has great potential. Because of its roots, it is widely cultivated in Africa, Asia, and Latin America. However, the canopy can be used to produce a leaf protein concentrate with a crude protein content of 40–45%, an amino acid profile similar to that of soybean, and tolerable tannin levels (>1% DM) [74].



**Table 2.** Amino acid composition (m/m%) of leaf protein concentrates from selected green biomass and seed-based protein sources by pre-column derivatization method of UHPLC (own measurements).

Amino Acids	Alfalfa (LPC)	Alfalfa (Green Juice)	Jerusalem Artichoke (LPC-MW)	Green Pepper (LPC-MW)	Green Soy (LPC-MW)	Cauliflower (LPC-MW)	Soy (Seed Extracted)	Triticale (Green Juice)	Triticale (LPC-Microwave)	Triticale (LPC-Lactic Acid)
ASP	5.22	4.33	4.24	2.86	4.29	5.23	5.64	2.72	3.84	2.68
THR	2.44	1.76	1.88	1.27	1.97	2.73	1.93	1.11	1.59	1.62
SER	2.34	1.67	1.91	1.30	2.10	2.40	2.43	1.19	1.69	1.72
GLU	5.27	4.01	4.09	3.34	4.57	5.95	8.76	2.68	3.55	3.57
PRO	2.10	1.44	2.04	1.53	2.10	2.60	2.34	1.24	1.78	1.78
GLY	2.55	1.79	1.70	1.68	2.23	2.71	2.06	1.23	2.07	2.06
ALA	2.89	2.02	1.94	1.79	2.66	2.52	1.99	1.60	2.43	2.38
CYS	0.11	0.11	0.22	0.15	0.15	0.77	0.20	0.12	0.20	0.20
VAL	2.73	1.96	1.47	1.77	2.25	1.93	2.27	1.42	2.04	2.05
MET	0.25	0.21	0.61	0.32	0.67	1.11	0.31	0.29	0.41	0.42
ILE	2.20	1.58	1.25	1.41	1.71	1.99	2.14	0.97	1.47	1.47
LEU	4.37	2.96	3.21	2.74	3.84	3.83	3.70	2.03	3.26	3.26
TYR	1.53	1.11	1.35	1.29	1.82	2.33	1.50	0.97	1.51	1.53
PHE	2.74	1.83	1.99	1.73	2.42	3.43	2.44	1.31	2.24	2.26
HIS	1.11	0.74	1.28	0.67	0.92	1.03	2.47	0.52	0.80	0.82
LYS	4.15	2.33	1.90	1.94	2.76	1.94	3.92	1.44	1.91	1.89
ARG	2.10	0.22	2.07	1.72	2.67	2.78	2.61	1.54	2.18	2.12

**Table 3.** Macro- and microelements composition ( $\text{mg kg}^{-1}$ ) of selected green biomass and seed-based protein sources based on own ICP-OES measurements.

Element	Alfalfa (Green Juice)	Alfalfa (LPC)	Broccoli (Green Juice)	Soy Seed (Extracted)
Mo	7.9	4.1	1.4	1.9
Cu	11.4	23.9	2.9	13.0
Ba	6.3	12.4	6.1	20.6
B	32.8	20.2	13.5	26.2
Zn	33.6	38.2	29.8	36.3
Mn	28.4	48.4	47.7	32.3
Sr	69.0	65.9	183.0	7.4
Al	92.0	145.8	68.8	36.7
Fe	145.3	315.9	97.2	88.4
Na	411.9	91.0	5116.2	7.1
Mg	3845	2149	8759	2268
S	4310	4682	13,653	2283
P	5457	7290	3308	5721
Ca	16,050	16,017	16,266	1862
K	40,020	21,245	20,770	14,654

**Table 4.** Qualitative phytochemical analysis of selected green biomass based on own UHPLC-ESI-MS measurements.

LPC-Alfalfa		LPC-Broccoli		LPC-JA	
Chemical Name	Chemical Formula	Chemical Name	Chemical Formula	Chemical Name	Chemical Formula
4'.5.7-Trihydroxyflavanone (Naringenin)	C <sub>15</sub> H <sub>12</sub> O <sub>5</sub>	Isoliquiritigenin (2',4,4'-trihydroxychalcone)	C <sub>15</sub> H <sub>12</sub> O <sub>4</sub>	γ-Aminobutyric acid	C <sub>4</sub> H <sub>9</sub> NO <sub>2</sub>
4'.5.7-Trihydroxyflavanone 6.8-C-glucoside	C <sub>27</sub> H <sub>32</sub> O <sub>15</sub>	Quercetin (3,3',4',5,7-Pentahydroxyflavone)	C <sub>15</sub> H <sub>10</sub> O <sub>7</sub>	Quinic acid	C <sub>7</sub> H <sub>12</sub> O <sub>6</sub>
4'.7-Dihydroxyflavanone	C <sub>15</sub> H <sub>12</sub> O <sub>4</sub>	Quercetin- <i>O</i> -hexoside- <i>O</i> - hexosylhexoside isomer 1	C <sub>33</sub> H <sub>40</sub> O <sub>22</sub>	Betaine (Trimethylglycine)	C <sub>5</sub> H <sub>11</sub> NO <sub>2</sub>
Quercetin	C <sub>15</sub> H <sub>10</sub> O <sub>7</sub>	Quercetin-3- <i>O</i> -[caffeoyl-(→2)-glucosyl- (1→2)-glucoside]-7- <i>O</i> -glucoside	C <sub>42</sub> H <sub>46</sub> O <sub>25</sub>	Malic acid	C <sub>4</sub> H <sub>6</sub> O <sub>5</sub>
Quercetin-3- <i>O</i> -glucoside	C <sub>21</sub> H <sub>20</sub> O <sub>12</sub>	Quercetin- <i>O</i> -(sinapoyl) hexosylhexoside- <i>O</i> -hexoside	C <sub>44</sub> H <sub>50</sub> O <sub>26</sub>	Nicotinic acid (Niacin)	C <sub>6</sub> H <sub>5</sub> NO <sub>2</sub>
Apigenin-4'- <i>O</i> -glucuronide-7- <i>O</i> - [glucuronyl-(1→2)-glucuronide]	C <sub>33</sub> H <sub>34</sub> O <sub>23</sub>	Quercetin-3- <i>O</i> -[feruloyl-(→2)-glucosyl- (1→2)-glucoside]-7- <i>O</i> -glucoside	C <sub>43</sub> H <sub>48</sub> O <sub>25</sub>	Citric acid	C <sub>6</sub> H <sub>8</sub> O <sub>7</sub>
Apigenin- <i>O</i> -glucoside- <i>O</i> -glucuronide	C <sub>27</sub> H <sub>28</sub> O <sub>16</sub>	Quercetin- <i>O</i> -hexoside- <i>O</i> - hexosylhexoside isomer 2	C <sub>33</sub> H <sub>40</sub> O <sub>22</sub>	Neochlorogenic acid (5- <i>O</i> -Caffeoylquinic acid)	C <sub>16</sub> H <sub>18</sub> O <sub>9</sub>
Apigenin-7- <i>O</i> -[feruloyl-(→2)- [glucuronyl-(1→3)]-glucuronyl- (1→2)]glucuronide	C <sub>43</sub> H <sub>42</sub> O <sub>26</sub>	Quercetin- <i>O</i> -hexosylhexoside isomer 1	C <sub>27</sub> H <sub>30</sub> O <sub>17</sub>	Salicylic acid-2- <i>O</i> -glucoside	C <sub>13</sub> H <sub>16</sub> O <sub>8</sub>
Apigenin-4'- <i>O</i> -glucuronide-7- <i>O</i> - [feruloyl-(→2)-glucuronyl-(1→2)- glucuronide]	C <sub>43</sub> H <sub>42</sub> O <sub>26</sub>	Quercetin-di- <i>O</i> -hexoside	C <sub>27</sub> H <sub>30</sub> O <sub>17</sub>	Chlorogenic acid (3- <i>O</i> -Caffeoylquinic acid)	C <sub>16</sub> H <sub>18</sub> O <sub>9</sub>
Apigenin-7- <i>O</i> -glucuronide	C <sub>21</sub> H <sub>28</sub> O <sub>11</sub>	Quercetin- <i>O</i> -hexosylhexoside isomer 2	C <sub>27</sub> H <sub>30</sub> O <sub>17</sub>	Chrysochlorogenic acid (4- <i>O</i> -Caffeoylquinic acid)	C <sub>16</sub> H <sub>18</sub> O <sub>9</sub>
Apigenin (4'.5.7-Trihydroxyflavone)	C <sub>15</sub> H <sub>10</sub> O <sub>5</sub>	Quercetin-3- <i>O</i> -glucoside (Isoquercitrin)	C <sub>21</sub> H <sub>20</sub> O <sub>12</sub>	4- <i>O</i> -(4-Coumaroyl) quinic acid	C <sub>16</sub> H <sub>18</sub> O <sub>8</sub>
Chrysoeriol-7- <i>O</i> -glucuronide	C <sub>22</sub> H <sub>20</sub> O <sub>12</sub>	Kaempferol (3,4',5,7-Tetrahydroxyflavone)	C <sub>15</sub> H <sub>10</sub> O <sub>6</sub>	Vanillin (4-Hydroxy-3- methoxybenzaldehyde)	C <sub>8</sub> H <sub>8</sub> O <sub>3</sub>
Chrysoeriol (3'-Methoxy-4'.5.7-trihydroxyflavone)	C <sub>16</sub> H <sub>12</sub> O <sub>6</sub>	Kaempferol- <i>O</i> -hexoside- <i>O</i> - hexosylhexoside	C <sub>33</sub> H <sub>40</sub> O <sub>21</sub>	5- <i>O</i> -(4-Coumaroyl)quinic acid	C <sub>16</sub> H <sub>18</sub> O <sub>8</sub>
Chrysoeriol-glucuronyl-glucuronide	C <sub>28</sub> H <sub>28</sub> O <sub>18</sub>	Kaempferol-7- <i>O</i> -glucoside-3- <i>O</i> - sophoroside	C <sub>33</sub> H <sub>40</sub> O <sub>21</sub>	Indole-3-acetic acid	C <sub>10</sub> H <sub>9</sub> NO <sub>2</sub>
Genkwanin (4',5-Dihydroxy-7-methoxyflavone)	C <sub>16</sub> H <sub>12</sub> O <sub>5</sub>	Kaempferol- <i>O</i> - (caffeoyl)hexosylhexoside- <i>O</i> -hexoside	C <sub>42</sub> H <sub>46</sub> O <sub>24</sub>	4- <i>O</i> -(4-Coumaroyl)quinic acid cis isomer	C <sub>16</sub> H <sub>18</sub> O <sub>8</sub>

Table 4. Cont.

LPC-Alfalfa		LPC-Broccoli		LPC-JA	
Chemical Name	Chemical Formula	Chemical Name	Chemical Formula	Chemical Name	Chemical Formula
Luteolin-7- <i>O</i> -glucuronide	C <sub>21</sub> H <sub>18</sub> O <sub>12</sub>	Kaempferol-3- <i>O</i> -[caffeoyl-( $\rightarrow$ 2)-glucosyl-(1 $\rightarrow$ 2)-glucoside]-7- <i>O</i> -glucoside	C <sub>42</sub> H <sub>46</sub> O <sub>24</sub>	Isoscooletin (6-Hydroxy-7-methoxycoumarin)	C <sub>10</sub> H <sub>8</sub> O <sub>4</sub>
Luteolin (3'.4'.5.7-Tetrahydroxyflavone)	C <sub>15</sub> H <sub>10</sub> O <sub>6</sub>	Kaempferol- <i>O</i> - (caffeoyl)hexosylhexoside- <i>O</i> - hexosylhexoside	C <sub>48</sub> H <sub>56</sub> O <sub>29</sub>	5- <i>O</i> -Feruloylquinic acid	C <sub>17</sub> H <sub>20</sub> O <sub>9</sub>
Tricin-7- <i>O</i> -glucuronide	C <sub>23</sub> H <sub>22</sub> O <sub>13</sub>	Kaempferol-3- <i>O</i> -[caffeoyl-( $\rightarrow$ 2)- glucosyl-(1 $\rightarrow$ 2)-glucoside]-7- <i>O</i> - [glucosyl-(1 $\rightarrow$ 4)-glucoside]	C <sub>48</sub> H <sub>56</sub> O <sub>29</sub>	Riboflavin	C <sub>17</sub> H <sub>20</sub> N <sub>4</sub> O <sub>6</sub>
Tricin-7- <i>O</i> -[feruloyl-( $\rightarrow$ 2)-glucuronyl- (1 $\rightarrow$ 2)-glucuronide]	C <sub>39</sub> H <sub>38</sub> O <sub>22</sub>	Kaempferol-3- <i>O</i> -[sinapoyl-( $\rightarrow$ 2)- glucosyl-(1 $\rightarrow$ 2)-glucoside]-7- <i>O</i> -glucoside	C <sub>44</sub> H <sub>50</sub> O <sub>25</sub>	Scopoletin (7-Hydroxy-6- methoxycoumarin)	C <sub>10</sub> H <sub>8</sub> O <sub>4</sub>
Tricin (3'.5'-Dimethoxy-4'.5.7- trihydroxyflavone)	C <sub>17</sub> H <sub>14</sub> O <sub>7</sub>	Kaempferol-3- <i>O</i> -[sinapoyl-( $\rightarrow$ 2)- glucosyl-(1 $\rightarrow$ 2)-glucoside]-7- <i>O</i> -[glucosyl-(1 $\rightarrow$ 4)-glucoside]	C <sub>50</sub> H <sub>60</sub> O <sub>30</sub>	Azelaamic acid (9-Amino-9-oxononanoic acid)	C <sub>9</sub> H <sub>17</sub> NO <sub>3</sub>
Tricin- <i>O</i> -hexoside	C <sub>22</sub> H <sub>24</sub> O <sub>12</sub>	Kaempferol-3- <i>O</i> -[feruloyl-( $\rightarrow$ 2)-glucosyl- (1 $\rightarrow$ 2)-glucoside]-7- <i>O</i> -glucoside	C <sub>43</sub> H <sub>48</sub> O <sub>24</sub>	6-Methylcoumarin	C <sub>10</sub> H <sub>8</sub> O <sub>2</sub>
4'.7-Dihydroxyflavone	C <sub>15</sub> H <sub>10</sub> O <sub>4</sub>	Kaempferol-3- <i>O</i> -[feruloyl-( $\rightarrow$ 2)- glucosyl-(1 $\rightarrow$ 2)-glucoside]-7- <i>O</i> - [glucosyl-(1 $\rightarrow$ 4)-glucoside]	C <sub>49</sub> H <sub>58</sub> O <sub>29</sub>	5- <i>O</i> -(4-Coumaroyl)quinic acid cis isomer	C <sub>16</sub> H <sub>18</sub> O <sub>8</sub>
Methoxy-tetrahydroxyflavone	C <sub>16</sub> H <sub>12</sub> O <sub>7</sub>	Kaempferol- <i>O</i> -[ <i>p</i> -coumaroyl-( $\rightarrow$ 2)- glucosyl-(1 $\rightarrow$ 2)-glucoside]-7- <i>O</i> -glucoside	C <sub>42</sub> H <sub>46</sub> O <sub>23</sub>	Indole-4-carbaldehyde	C <sub>9</sub> H <sub>7</sub> NO
Dimethoxy-hydroxyflavone	C <sub>17</sub> H <sub>14</sub> O <sub>5</sub>	Kaempferol-3,7-di- <i>O</i> -glucoside (Paeonoside)	C <sub>27</sub> H <sub>30</sub> O <sub>16</sub>	Fraxidin or Isofraxidin	C <sub>11</sub> H <sub>10</sub> O <sub>5</sub>
3'-Methoxy-4'.5.5'.7- tetrahydroxyflavone-7- <i>O</i> -glucuronide	C <sub>22</sub> H <sub>20</sub> O <sub>13</sub>	Kaempferol- <i>O</i> - (sinapoyl)hexosylhexoside- <i>O</i> - (sinapoyl)hexoside	C <sub>55</sub> H <sub>60</sub> O <sub>29</sub>	Loliolide	C <sub>11</sub> H <sub>16</sub> O <sub>3</sub>
Apigenin-8- <i>C</i> -glucoside-6- <i>C</i> -xyloside	C <sub>26</sub> H <sub>28</sub> O <sub>14</sub>	Kaempferol-di- <i>O</i> -hexoside	C <sub>27</sub> H <sub>30</sub> O <sub>16</sub>	4-Hydroxy-3- methoxycinnamaldehyde (Coniferyl aldehyde)	C <sub>10</sub> H <sub>10</sub> O <sub>3</sub>
Apigenin-6- <i>C</i> -glucoside-8- <i>C</i> -xyloside	C <sub>26</sub> H <sub>28</sub> O <sub>14</sub>	Kaempferol- <i>O</i> -(caffeoyl)hexosylhexoside	C <sub>36</sub> H <sub>36</sub> O <sub>19</sub>	7-Deoxyloganic acid isomer	C <sub>16</sub> H <sub>24</sub> O <sub>9</sub>
Alfalone (4'.7-Dimethoxy-6-hydroxyisoflavone)	C <sub>17</sub> H <sub>14</sub> O <sub>5</sub>	Kaempferol- <i>O</i> - (sinapoyl)hexosylhexoside	C <sub>38</sub> H <sub>40</sub> O <sub>20</sub>	Di- <i>O</i> -caffeoylquinic acid isomer 1	C <sub>25</sub> H <sub>24</sub> O <sub>12</sub>

Table 4. Cont.

LPC-Alfalfa		LPC-Broccoli		LPC-JA	
Chemical Name	Chemical Formula	Chemical Name	Chemical Formula	Chemical Name	Chemical Formula
Formononetin (7-Hydroxy-4'-methoxyisoflavone)	C <sub>16</sub> H <sub>12</sub> O <sub>4</sub>	Kaempferol-7- <i>O</i> -sophoroside	C <sub>27</sub> H <sub>30</sub> O <sub>16</sub>	Di- <i>O</i> -caffeoylquinic acid isomer 2	C <sub>25</sub> H <sub>24</sub> O <sub>12</sub>
Ononin (Formononetin 7- <i>O</i> -glucoside)	C <sub>22</sub> H <sub>22</sub> O <sub>9</sub>	Kaempferol- <i>O</i> -(feruloyl)hexosylhexoside	C <sub>37</sub> H <sub>38</sub> O <sub>19</sub>	Salvianolic acid derivative isomer 1	C <sub>27</sub> H <sub>22</sub> O <sub>12</sub>
Biochanin A (4'-Methylgenistein)	C <sub>16</sub> H <sub>12</sub> O <sub>5</sub>	Kaempferol- <i>O</i> -(4-coumaroyl)hexosylhexoside	C <sub>36</sub> H <sub>36</sub> O <sub>18</sub>	Butein (2',3,4,4'-Tetrahydroxychalcone)	C <sub>15</sub> H <sub>12</sub> O <sub>5</sub>
Isoliquiritigenin (2',4,4'-trihydroxychalcone)	C <sub>15</sub> H <sub>12</sub> O <sub>4</sub>	Kaempferol- <i>O</i> -(disinapoyl)hexosylhexosylhexoside- <i>O</i> -hexoside	C <sub>61</sub> H <sub>70</sub> O <sub>34</sub>	Quercetin-3- <i>O</i> -glucuronide	C <sub>21</sub> H <sub>18</sub> O <sub>13</sub>
Medicagenic acid	C <sub>30</sub> H <sub>46</sub> O <sub>6</sub>	Kaempferol- <i>O</i> -hexosylhexoside	C <sub>27</sub> H <sub>30</sub> O <sub>16</sub>	Isoquercitrin (Hirsutrin, Quercetin-3- <i>O</i> -glucoside)	C <sub>21</sub> H <sub>20</sub> O <sub>12</sub>
Medicagenic acid 28- <i>O</i> -[xylosyl-(1→4)-rhamnosyl-(1→2)-arabinosyl]ester	C <sub>46</sub> H <sub>72</sub> O <sub>18</sub>	Kaempferol-3- <i>O</i> -glucoside (Astragalin)	C <sub>21</sub> H <sub>20</sub> O <sub>11</sub>	Chrysoeriol- <i>O</i> -glucoside	C <sub>22</sub> H <sub>22</sub> O <sub>11</sub>
Medicoside H (Medicagenic acid 3- <i>O</i> -glucosyl-28- <i>O</i> -[rhamnosyl-(1→2)-arabinosyl]ester)	C <sub>47</sub> H <sub>74</sub> O <sub>19</sub>	Isorhamnetin- <i>O</i> -hexosylhexoside	C <sub>28</sub> H <sub>32</sub> O <sub>17</sub>	Salvianolic acid derivative isomer 2	C <sub>27</sub> H <sub>22</sub> O <sub>12</sub>
Medicoside G (Medicagenic acid 3,28-di- <i>O</i> -glucoside)	C <sub>42</sub> H <sub>66</sub> O <sub>16</sub>	Isorhamnetin-3- <i>O</i> -glucoside	C <sub>22</sub> H <sub>22</sub> O <sub>12</sub>	Di- <i>O</i> -caffeoylquinic acid isomer 3	C <sub>25</sub> H <sub>24</sub> O <sub>12</sub>
Medicagenic acid 3- <i>O</i> -glucuronide-28- <i>O</i> -[xylosyl-(1→4)-rhamnosyl-(1→2)-arabinosyl]ester	C <sub>52</sub> H <sub>80</sub> O <sub>24</sub>	Isorhamnetin-7- <i>O</i> -glucoside-3- <i>O</i> -sophoroside (Brassicoside)	C <sub>34</sub> H <sub>42</sub> O <sub>22</sub>	Azelaic acid	C <sub>9</sub> H <sub>16</sub> O <sub>4</sub>
Medicagenic acid rhamnosyl-pentosyl-glucuronide	C <sub>47</sub> H <sub>72</sub> O <sub>20</sub>	4',7-Dihydroxyflavanone (Liquiritigenin)	C <sub>15</sub> H <sub>12</sub> O <sub>4</sub>	Kaempferol-3- <i>O</i> -glucuronide	C <sub>21</sub> H <sub>18</sub> O <sub>12</sub>
Medicoside J (Medicagenic acid 3- <i>O</i> -glucosyl-28- <i>O</i> -[xylosyl-(1→4)-rhamnosyl-(1→2)-arabinosyl]ester)	C <sub>52</sub> H <sub>82</sub> O <sub>23</sub>	4',5,7-Trihydroxyflavanone (Naringenin)	C <sub>15</sub> H <sub>12</sub> O <sub>5</sub>	Apigenin- <i>O</i> -malonylglucoside	C <sub>24</sub> H <sub>22</sub> O <sub>13</sub>
Soyasapogenol B rhamnosyl-hexosyl-glucuronide	C <sub>48</sub> H <sub>78</sub> O <sub>18</sub>	Apigenin (4',5,7-Trihydroxyflavone)	C <sub>15</sub> H <sub>10</sub> O <sub>5</sub>	Astragaln (Kaempferol-3- <i>O</i> -glucoside)	C <sub>21</sub> H <sub>20</sub> O <sub>11</sub>
Soyasapogenol B rhamnosyl-pentosyl-glucuronide	C <sub>47</sub> H <sub>76</sub> O <sub>17</sub>	Apigenin-7- <i>O</i> -glucuronide	C <sub>21</sub> H <sub>28</sub> O <sub>11</sub>	Isorhamnetin-3- <i>O</i> -glucoside	C <sub>22</sub> H <sub>22</sub> O <sub>12</sub>

Table 4. Cont.

LPC-Alfalfa		LPC-Broccoli		LPC-JA	
Chemical Name	Chemical Formula	Chemical Name	Chemical Formula	Chemical Name	Chemical Formula
Azukisaponin II	C <sub>42</sub> H <sub>68</sub> O <sub>14</sub>	Luteolin (3',4',5,7-Tetrahydroxyflavone)	C <sub>15</sub> H <sub>10</sub> O <sub>6</sub>	Kukulkanin B (2',4',4'-Trihydroxy-3'-methoxyxchalcone)	C <sub>16</sub> H <sub>14</sub> O <sub>5</sub>
<b>Unknown saponins</b>		Neochlorogenic acid (5-O-Caffeoylquinic acid)	C <sub>16</sub> H <sub>18</sub> O <sub>9</sub>	Isorhamnetin-3-O-glucuronide	C <sub>22</sub> H <sub>20</sub> O <sub>13</sub>
unknown saponin. Aglycon: 440.32905 (C <sub>29</sub> H <sub>44</sub> O <sub>3</sub> )	C <sub>58</sub> H <sub>92</sub> O <sub>28</sub>	Chlorogenic acid (3-O-Caffeoylquinic acid)	C <sub>16</sub> H <sub>18</sub> O <sub>9</sub>	Dihydroactinidiolide	C <sub>11</sub> H <sub>16</sub> O <sub>2</sub>
unknown saponin. Aglycon: 504.34509 (C <sub>30</sub> H <sub>48</sub> O <sub>6</sub> )	C <sub>41</sub> H <sub>64</sub> O <sub>16</sub>	Chryptochlorogenic acid (4-O-Caffeoylquinic acid)	C <sub>16</sub> H <sub>18</sub> O <sub>9</sub>	Dimethoxy-tetrahydroxyflavone	C <sub>17</sub> H <sub>14</sub> O <sub>8</sub>
unknown saponin. Aglycon: 486.33452 (C <sub>29</sub> H <sub>42</sub> O <sub>3</sub> )	C <sub>42</sub> H <sub>64</sub> O <sub>16</sub>	Caffeic acid	C <sub>9</sub> H <sub>8</sub> O <sub>4</sub>	Dihydroxy-methoxyflavone	C <sub>16</sub> H <sub>12</sub> O <sub>5</sub>
unknown saponin. Aglycon: 454.34470 (C <sub>30</sub> H <sub>46</sub> O <sub>3</sub> )	C <sub>47</sub> H <sub>74</sub> O <sub>19</sub>	4-Coumaric acid	C <sub>9</sub> H <sub>8</sub> O <sub>3</sub>	Dimethoxy-trihydroxyflavone isomer 1	C <sub>17</sub> H <sub>14</sub> O <sub>7</sub>
		Sinapic acid	C <sub>11</sub> H <sub>12</sub> O <sub>5</sub>	Trihydroxy-trimethoxyflavone	C <sub>18</sub> H <sub>16</sub> O <sub>8</sub>
		Di-O-sinapoylgentiobiose	C <sub>34</sub> H <sub>42</sub> O <sub>19</sub>	Dimethoxy-trihydroxyflavone isomer 2	C <sub>17</sub> H <sub>14</sub> O <sub>7</sub>
		Tri-O-sinapoylgentiobiose	C <sub>45</sub> H <sub>52</sub> O <sub>23</sub>	Liquiritigenin (4',7-Dihydroxyflavanone)	C <sub>15</sub> H <sub>12</sub> O <sub>4</sub>
		Feruloyl-sinapoyldihexoside	C <sub>33</sub> H <sub>40</sub> O <sub>18</sub>	Hymenoxin (5,7-Dihydroxy-3',4',6,8-tetramethoxyflavone)	C <sub>19</sub> H <sub>18</sub> O <sub>8</sub>
		Di-O-sinapoylglucose	C <sub>28</sub> H <sub>32</sub> O <sub>14</sub>	Epiafzelechin trimethyl ether	C <sub>18</sub> H <sub>20</sub> O <sub>5</sub>
		Feruloyl-disinapoyldihexoside	C <sub>44</sub> H <sub>50</sub> O <sub>22</sub>	Nevadensin (5,7-Dihydroxy-4',6,8-trimethoxyflavone)	C <sub>18</sub> H <sub>16</sub> O <sub>7</sub>
		Syringaldehyde (3,5-Dimethoxy-4-hydroxybenzaldehyde)	C <sub>9</sub> H <sub>10</sub> O <sub>4</sub>		
		Glucobrassicin (3-Indolylmethyl glucosinolate)	C <sub>16</sub> H <sub>20</sub> N <sub>2</sub> O <sub>9</sub> S <sub>2</sub>		
		3-Methylsulphanylpropyl isothiocyanate	C <sub>5</sub> H <sub>9</sub> NOS <sub>2</sub>		

Table 4. Cont.

LPC-Alfalfa		LPC-Broccoli		LPC-JA	
Chemical Name	Chemical Formula	Chemical Name	Chemical Formula	Chemical Name	Chemical Formula
		4-Methoxy-3-indolylmethyl glucosinolate	$C_{17}H_{22}N_2O_{10}S_2$		
		Sulforaphane	$C_6H_{11}NOS_2$		
		Neoglucobrassicin (1-Methoxy-3-indolylmethyl glucosinolate)	$C_{17}H_{22}N_2O_{10}S_2$		
		Scopoletin (7-Hydroxy-6-methoxycoumarin)	$C_{10}H_8O_4$		
		<i>Other phytochemicals</i>			
		$\gamma$ -Aminobutyric acid	$C_4H_9NO_2$		
		Indole-4-carbaldehyde	$C_9H_7NO$		
		Abscisic acid	$C_{15}H_{20}O_4$		
		Kynurenic acid	$C_{10}H_7NO_3$		

#### 4. Biorefining of Protein from Green Biomass

Due to its nutritional value, high yields, and simplicity of extraction and preparation, LPC may be an effective and sustainable source of proteins for food and feed. However, in order to make LPC more palatable, it must be isolated, purified, and concentrated.

Several large-scale techniques have been used to extract the protein from green leaves. However, the extraction efficiency of LPC largely varies between 35 and 80%, depending on many factors.

Agents that disrupt the cellular and subcellular membranes are undoubtedly a determining element in LPC extraction. Moreover, plant species, stage of maturity, plant tissue, the presence of mucilaginous material, postharvest treatment, pH, extractant composition, flotation ratios, extraction time, and temperature [78] also influence this outcome. For example, Solati et al. [62] reported higher crude protein contents in LPC derived from legumes (i.e., white clover, red clover, or alfalfa) than grass species (i.e., ryegrass or tall fescue). Moreover, they documented higher crude protein contents in plant leaves than in plant stems. The extracted amount of LPC did not significantly differ according to the maturity stage of the five investigated plant species.

##### 4.1. Extraction of Leaf Protein

The first step in LPC production after collecting the fresh green biomass, mainly leaves and stems, is the mechanical pressing to squeeze out the protein-rich green juice from the fibers [79]. This occurs in two steps: cell walls are first disrupted using different mills or rollers to release intracellular ingredients, mainly soluble proteins and chloroplasts; secondly, the juice is collected by pressing the pulped tissues [79]. However, due to practical limitations, the two steps of green juice extraction could not take place at an industrial scale in a single unit. Therefore, attention was given in early times to developing economically efficient methods to extract green juice [3,80].

Screw presses recently showed higher capability in plant-based green juice extraction, as it offers the maceration of the plant cell walls along with desired pressure to squeeze out the juice with an efficient rate of about 60% [81]. Twin-screw extrusion with two propellers on separate shafts with opposite twists revealed higher extraction efficiency of green juice, up to 65% of the inherent fluid found in fresh plant materials with more than 50% of protein precipitation [82,83].

Many researchers suggested re-pressing the press cake after adding an equivalent amount of water because at least 50% of leaf protein is retained in the press cake after the first mechanical pressing [84–86]. Knuckles et al. [85] reported a 13% increase in protein recovery after re-pressing water-diluted alfalfa press cake while diluting alfalfa press cake with 5–6% water (on a dry mass basis) increased protein content in green juice by 17% [87]. Similar conclusions were reported by Colas et al. [82] in their study on green juice extraction of alfalfa green biomass using a twin-screw pump at different added water volumes to press cake. Consequently, it can be concluded that the addition of water to press cake, and re-pressing the pressed cake, may be performed if it favors protein recovery. Moreover, chopping plant tissues before pressing could help in releasing the soluble compounds [88].

However, pulping and pressing chopped plant materials in the presence of an alkali solution to bring the pH to 8.0 is recommended rather than pure water. Extraction of leaf protein in an environment with pH ranging between 7.0 and 8.0 was found to improve green juice extraction and protein recovery, where leaf protein becomes more soluble and destruction of cell walls becomes more efficient [89,90]. However, higher pH could bring risks to protein denaturation; therefore, pH should not exceed 8.0.

Another factor that cannot be ignored during the pressing of green biomass is the temperature of the plant tissues before and during the mechanical pulping. Hanna and Ogden [91] observed that heating the plant materials to above 35 °C considerably reduced the juice extraction and protein recovery, while lower temperatures below 14 °C to 3 °C showed the same effect as the room temperature.

Evidently, the efficiency of juice extraction and protein recovery from fresh plant materials depends on many factors such as pulper device, pH, chopping plant tissues,



and temperature. Moreover, the co-extraction of anti-nutritional components should be considered and evaluated.

#### 4.2. Precipitation of Leaf Protein

Isolation of protein from green juice obtained by pressing green biomass is the core step in the production of economically feasible LPC. Consequently, several techniques have been proposed to concentrate leaf protein, which can be classified into three main techniques as follows: (1) differences in solubility (on the basis of distribution coefficient), including salting, organic solvent fractionation, chromatography, crystallization, heating, and centrifugation [92]; (2) gel filtration (on the basis of molecular sizes and shapes), including size-exclusion chromatography and membrane [93]; and (3) isoelectric focusing (on the basis of isoelectric point of proteins), including ion exchange [94].

However, we summarize here the most commonly used techniques for the precipitation of leaf protein from green juices, such as thermal coagulation, fermentation, supercritical CO<sub>2</sub> extraction, pH extraction, and polyelectrolytes extraction.

##### 4.2.1. Thermal Coagulation

Recovery of leaf protein from green leaves-derived juice by heat coagulation at a temperature ranges between 60 °C and 95 °C is one of the most commonly applied approaches in the leaf protein precipitation theme [95]. Since Rubisco enzyme represents about 50% of leaf protein [96] and its denaturation temperature is 76.2 °C [97]; therefore, the optimum temperature of leaf protein coagulation is 80 °C [80]. The heat disrupts the protein structure and consequently decreases its water solubility by opening the hydrophobic sites [79]. Proteins found in green juice can be converted into coagulum by heating the green juice to 80 °C for 2–4 min. However, the total coagulum is unpalatable, particularly for human consumption, due to its green color, grassy odor, low water solubility, and digestibility. Consequently, a selective heat coagulation technique was proposed for fractionation of the leaf protein into green and white proteins, which can be directed towards non-ruminants and human consumption, respectively [95]. Green juice is primarily heated to 55 °C to isolate the green protein fraction, then after the supernatant is heated to 80 °C to separate the so-called white protein fraction. The thermal coagulation method of leaf protein is effective; however, it is characterized by many drawbacks such as being energy consuming, the LPC produced has low water solubility, and the properties of protein functional groups could be altered [97].

##### 4.2.2. Microwave Coagulation

In addition to other industrial uses, microwave as electromagnetic radiation has recently been used for protein coagulation alone or in combination with conventional thermal coagulation. It is observed that the plant protein coagulum obtained by microwave coagulation is characterized by a coherent colloidal system and dispersed macromolecular structure. The physical consistency of the coagulum is harder, which facilitates separation from the brown liquid during filtration [98,99].

##### 4.2.3. pH Precipitation

Both acidosis and alkalosis can be employed to isolate leaf protein. It is well-known that in conditions of low pH (below 4.5) or high pH (above 8.0), proteins carry positive or negative charges, respectively, which leads to the liberation of several hydrophobic free amino acids, causing a reduction in protein solubility. Usually, acid precipitation of leaf protein is performed by the addition of HCl, while NaOH or ammonia solution is utilized to increase pH to 8.0–8.5. Precipitated protein is subsequently obtained through centrifugation in the form of pellets. For example, acidifying green juice of soybean leaves to pH 3.7 precipitated about 95% of leaf protein [100]. Similarly, proteins in cassava leaves were precipitated by lowering pH to 4.0–5.0, where solubility was at the minimum [101]. Isolation of leaf protein via the pH method may decrease the activity of protease and thus

improves the carotene and lutein stability [102]. However, the disadvantages of this method are the loose structure of LPC and the accelerated oxidation of unsaturated fatty acids [64].

#### 4.2.4. Acid-Assisted Thermal Coagulation

Protein can also be denatured by acidosis as a result of changing the overall charges on the amino acids. In this method, the pH of green juice is first lowered by acid addition, followed by heating to isolated leaf proteins. Coagulation of leaf protein by this method is faster than thermal coagulation, and it yields a more compact coagulum compared to the traditional acid precipitation method. Moreover, it produces higher LPC yield in addition to being cost-effective compared to the traditional heating coagulation approach [64]. Moreover, isolated protein by acid-assisted heating method showed higher digestibility and absorption rates. Nevertheless, this method is still energy-consuming and costs more than that of other techniques [64].

#### 4.2.5. Microbial Fermentation

Isolation of leaf protein using microbial fermentation has the same basic idea as acid precipitation, where protein solubility largely depends on the pH. However, the main difference between both methods is that instead of adding acids to the green juice, inoculation green juice with acid-producing microbes such as lactic acid bacteria strains (i.e., *Lactobacillus plantarum*, *Pediococcus cerevisiae*, and *Lactobacillus salivarius*) will acidify the juice due to the naturally produced organic acids by these bacteria [64,95]. The reduction in pH of green juice after inoculation with lactic acid bacteria is mainly attributed to the production of lactic acid by the inoculant bacteria. Isolation leaf protein using *L. salivarius* achieved similar results compared to acid precipitation using sulfuric acid [103]. This method is characterized by fewer energy requirements compared to thermal coagulation and less damage to obtained protein. However, it needs a long time and requires specific equipment.

#### 4.2.6. Protein Precipitation by Flocculants

Flocculants are substances that help tiny particles aggregate, making it simpler to remove them from the liquid phase. Flocculants form large complexes with protein that can be easily isolated from the mixture at room temperature [63]. Several flocculants can be employed in leaf protein recovery, including ionic and non-ionic flocculants [104]. In a study comparing different techniques to isolate leaf proteins in alfalfa green juice, it was found that leaf protein recovery percentages were 42.7%, 42.9%, 45.0%, and 53% with cationic flocculants, acid precipitation (pH 3.5), anionic flocculants, and thermal coagulation, respectively [105]. The exploitation of many flocculants has been examined in green juices obtained from several plant species, including alfalfa, tall fescue, and ryegrass [106]. Lignosulfonates, a by-product of wood pulp production by sulfite pulping characterized by being a hydrophilic and anionic polyelectrolyte polymer, is recently used to precipitate leaf protein from green juice [63]. Compared to thermal coagulation and acid precipitation, lignosulfonates increased LPC yield from green juices of ryegrass, red clover, grass-clover combo, and spinach by 6%, 5%, 7%, and 20%, respectively. The ideal concentration of added lignosulfonates to green juice was 0.6–0.7 g per g protein [63].

#### 4.2.7. Supercritical CO<sub>2</sub> Extraction

The supercritical CO<sub>2</sub> extraction for foods and food ingredients was proposed for the first time by Zosel in 1964 [107]. CO<sub>2</sub> is a solvent that is good for the environment and works well when processing food. The merits that make supercritical CO<sub>2</sub> a typical extractant are its critical point, which is non-toxic and relatively unreactive. Under physiological and/or highly specialized experimental settings, it has been demonstrated that proteins, amino acids, and amines can interact with carbon dioxide [108]. The impact of supercritical CO<sub>2</sub> on ribonuclease was investigated as a model protein system in an earlier article [108]. Supercritical CO<sub>2</sub> may be a more practical and cost-effective way to change how proteins function; the commercial manufacturing of decaffeinated coffee is a good example of this.

Additionally, milk and soy proteins that were disseminated in an aqueous solution may be precipitated using CO<sub>2</sub> as a weak acid [109–111]. It is medically safe, inflammable, nontoxic, chemically inert, readily recyclable, and reusable. Furthermore, supercritical CO<sub>2</sub> has zero surface tension [112], which results in total and quick wetting and permits penetration of complicated structures [113]. When combined with a co-solvent such as ethanol, the supercritical CO<sub>2</sub> can be utilized to directly alter the composition of commercial whey protein components by extracting lipids and other nonpolar and polar molecules. Depletion or redistribution of nonpolar substances can alter protein structures as well. These elements might present brand-new protein capabilities.

#### 4.2.8. Ultrafiltration

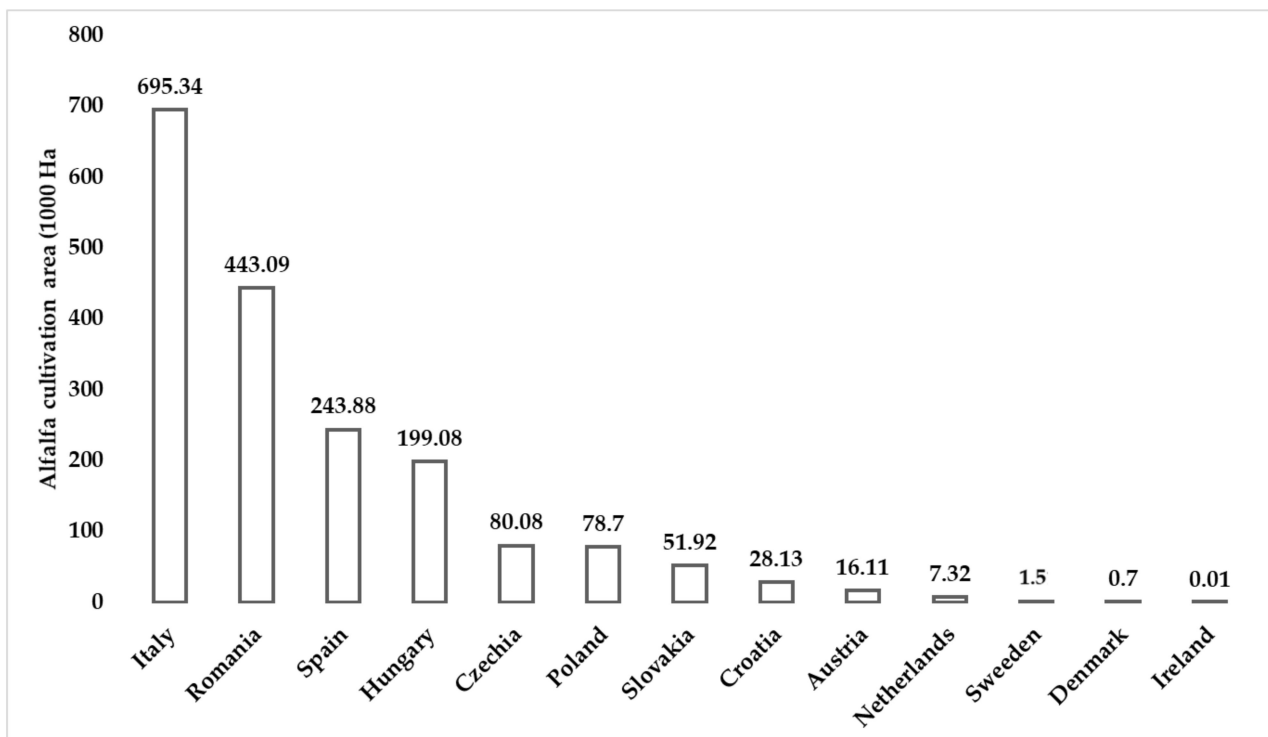
In order to reduce the energy expenditure in the LPC industry, ceramic membranes have been proposed as an alternative way to isolate leaf protein from green juices as ultrafiltration. This process yields a relatively high LPC amount with protein content ranges between 26.3 and 38.8%, while LPC produced by thermal coagulation exhibited a protein content of 40.1–46.3% [114]. However, ultrafiltration of green juice should be achieved within a short time and at low temperatures to avoid protein hydrolysis. Ref. [97] used a 10 kDa cut-off membrane to separate white protein from alfalfa green juice after eliminating green protein by thermal coagulation. Centrifugation of dissolved alfalfa green juice, which was previously frozen at –25 °C, recovered about 60% of the total N in the juice [115].

## 5. Application of Green Protein Today

### 5.1. State of Art of Green Biomass-Originated Protein in Europe

The production and supply of proteins for the agri-food sector is a recurring issue in the economic policy of the European Union. Within the plant protein utilization market, conventional feed, high-value feed, and food segments can be distinguished. Regarding the feed sector, the EU still relies on imports of protein-rich plant sources, in particular soybeans, which account for almost one-third of all protein used in animal feed [116]. Imports, however, mean vulnerability. Hence, in line with the EU's "Farm to Fork" strategy, agricultural policy reforms are stimulating local protein feed production and reducing import dependency [116].

Considering the agroecological potential of European countries, green biomass proteins offer a real alternative to soy and other seed-based proteins [117]. Green biomass includes grasses; immature cereals and legume crops, such as alfalfa; and clovers as dedicated species in Europe [95]. According to Corona et al., the area of grassland in the European Union (EU) reaches ~16.4 million Ha. Between 10 and 20% of this area could be used for alternative purposes to grazing [59]. While alfalfa is grown on ~2.5 million hectares of pure stands, including legume–grass mixtures, alfalfa is the most widely grown forage legume in almost all of Europe [118]. Currently, the global area of alfalfa is 30 million hectares, of which 25% (7.12 million hectares) is in Europe (Figure 2) [113,119].



**Figure 2.** Cultivation area of alfalfa in some EU countries in 2021 from EUROSTAT, 2022 [120].

Investigating the value and use of green biomass for the production of value-added products and platform molecules is an area of research in Europe with varying intensity from country to country. For instance, Germany, France, the Netherlands, Denmark, and Finland have developed “national protein strategies” to ensure circular and sustainable feed and food supply chains.

In Denmark, there is intensive livestock production. Pork production contributed to 6% of EU-28 production [121,122]. Sustainable pork production also requires an adequate source of feed protein. After cereals, the second most widely grown crop in Denmark is grass and green fodder, with 0.6 million hectares. A well-established selection program has resulted in grass varieties with high yield potential and clover varieties with high protein content, which are being developed for forage purposes [123]. Significant efforts have recently been made in Denmark to develop a green biomass value chain, partly with the aim of reducing dependence on soybean meal imports by using processed green forages.

Based on these, significant efforts have recently been made in Denmark to develop a green biomass value chain, partly using processed green forage, in order to reduce dependence on soybean meal imports [67,68,124]. The research and development have resulted in pilot-scale green biorefineries that demonstrate technical feasibility, such as the decentralized pilot plant at the Agricultural Research Centre in Foulum [125].

Germany is also seeking to make progress in promoting green biorefining by exploiting the surplus grassland biomass. As an example of this, a demonstration green biomass processing facility was planned, directly linked to the already existing green forage drying plant in Selbelang, Havelland. The plant was designed to process 20,000 tonnes of alfalfa and grass biomass yearly, producing platform chemicals and products [126,127].

Grassland makes up nearly 90% of Ireland’s arable land. Among these, perennial rye grass is a widely used fodder crop. Perennial rye grass can be used as green biomass for the production of leaf protein concentrate for pigs by thermal coagulation [69,128]. The protein content of the protein concentrate was 33.9%, and the amounts of lysine and cysteine amino acids were significantly lower than in soybean meal. These results indicated that soybean meal could not be completely substituted by perennial rye grass protein

concentrate. However, daily feed intake, feed conversion ratio, and average daily weight gain were better in pigs fed with protein concentrate than in the control.

In the Central-Eastern region of Europe, Hungary has under-utilized potential to develop the green biomass value chain. For instance, Eurostat data indicate that 1,835,000 tonnes of alfalfa were grown on 212,000 ha in 2021. Pioneering work of the “Green Fodder Mill” concept, a process of wet fractionation of green alfalfa biomass, was carried out by Károly (Karl) Ereky in Hungary in the 1920s [16,17,31,37]. In the 1960s, the VEPEX (Vegetable Protein Extract) program was launched, based on a technology to produce high biological value, fiber-free feed from green plants. A steam injection coagulation-based technology developed and patented was applied on an industrial scale in Tamasi and Ács (Hungary) [129]. In the early 2000s, green biorefining gained new momentum. A semi-industrial demonstration plant was built by the Tedej Co. Ltd. (Hajdúnánás-Tedej, Hungary) in cooperation with the University of Debrecen, based mainly on green biomass from lucerne. The plant was capable of producing 200 L h<sup>-1</sup> of green juice, from which ~520 kg h<sup>-1</sup> of leaf protein concentrate (36–46 m<sup>-1</sup>% crude protein) could be produced by combined heat and microwave coagulation [130]. In 2019, an industrial size green protein biorefinery factory was constructed in Hungary called PROTEOMILL, which technology has also been patented [98]. Its actual production capacity is 4000 L of green juice per hour. The fresh alfalfa green biomass is harvested from a 120–150 ha plantation belonging to the PROTEOMILL factory (Tedej Co. Ltd., Hajdúnánás-Tedej, Hungary).

France is one of the major alfalfa producers in the EU and exports the crop mainly in dried form to different countries. Along with it, they use well-known technologies such as fractionation at large scale [119]. As mentioned above, plant leaf protein can be used not only as feed but also as food. In Europe since the 1960s, the increasing proportion of plant-based dietary proteins relative to animal proteins in both consumption and production can be observed. A shift to a plant-based diet to meet the UN Sustainable Development Goals and the Paris Agreement presents the Planetary Health Diet as a benchmark for a healthy and sustainable diet for a growing population [131]. In 1993, at the suggestion of France Luzerne, the APEF (Association pour la Promotion des Extraits Foliaires en Nutrition, Paris) launched a program for the use of alfalfa leaf protein concentrate for human consumption [132]. A pilot plant was launched in 1997 with the participation of five companies, including Alfa-Laval. Then, with the approval of the EU, the FRALUPRO (“Fractionation of alfalfa juice to create a functional and functional protein ingredient for the food and non-food industry”) program was launched, with the design of the first 1200 t year<sup>-1</sup> capacity so-called Rubisco protein plant. Alfalfa leaf protein-xanthophyll concentrate (APC), which contains more than 50% total protein and 1200–2200 mg dm<sup>-3</sup> xanthophyll, is approved by EFSA as a food supplement [133,134].

## 5.2. State of Art of Green Biomass-Originated Protein in America

The urge to shift from animal to plant-based foods observed in Europe is also experienced in America [135–139]. In the globalized world market, other continents are expected to face similar challenges. This is due to an increasing world population [140,141] and to meet a demand for lower energy and labor inputs to food production. Success in this matter means sustaining our feeding needs and consists of an intricate web involving psychological [142], political [136], environmental [143,144], cultural, economic, and personal issues [136,139,141,142,145].

The international trade market is either an active or passive player, suffering and leading the outcomes that run through agricultural and livestock products [136,140,143]. For instance, events such as the COVID-19 pandemic [146]; others such as the Russia–Ukraine conflict; and shades of political, cultural, and ideological beliefs may particularly influence our capacity to produce and consume food. Consequently, caution and consciousness are essential for success, locally and worldwide.

Plant- and meat-based products have always been part of the human diet [147]. Further, we are constantly faced with new food ingredients and the possible reuse of agricultural byproducts that can minimize waste and pollution [148]. A compiled guidance for

reliable and environment-friendly feeding habits is observed [136,140,149], and it includes the change to plant-based food protein, new food ingredients, and the due regulation of food production [135,136,141]. The use of plant varieties that are less demanding on energy, minerals, and other inputs while, ideally, presenting more nutritive, productive potential, and increased acceptance is encouraged. This comprises the continuous efforts of plant breeding and dealing with climate change and the decrease in water resources that, despite imperative, are not covered here.

There are commercial perspectives and patent interest in plant-based protein food (Table 5). Nevertheless, it seems that the interest in this market overrules the attention to nutritional, healthy, and other science-related issues with plant-based foods. It appears to be, at least, a delay in the compass among the information and actions in the trade sector, research initiatives, and consumer preferences. Detailed market analysis on plant-based foods and protein and products are available (Table 5), whereas information on the protein quality and amino acid composition may be recent or incipient [149]. Despite the possible bias in the World Wide Web information and the need for datamining expertise, we face an evolving area of merchandise and advertisement on plant-based foods.

The focus on the market and advertisement on plant-based food and protein is surprising! This leads us to question “the state of market of green biomass and protein” in addition to the “state of art of green biomass originated protein in America”. We observe attractive “market polls” and promising niche of plant-based protein (Table 5). This covers conflicting information on the internet approaching opinions, polls, markets, etc. This will easily confuse the reader and the consumer, so we are compelled to ask: what is the role of science in this trend? Scientific articles cover objectives from the opinion, psychological and healthy issues of the consumer to the approach of protein extraction protocols.

Patents dealing with the processing of plant-based foods (Table 5) as the endorsement of the positive contribution of plant-based diets to human health [150,151] are available. This figure ought to be higher since Arbach et al. [152], for instance, found 113 patent applications related to plant-based beverages. Scientific research may provide new and improved plant varieties and species through selection, breeding, and biotechnological tools that afford biomass and protein as food for humans [148,153–156].

Despite globalized and scattered information, plant-based foods and plant (and other sources of) protein [135,136,138] are deemed to be at the forefront of our interest. There are recent approaches to the transition to plant-based protein [136,139,145,157,158], signaling efforts of the scientific community to keep up with this trend.

A bibliometric analysis on “the disposition of reducing meat consumption” was conducted by gathering information from 1994 to 2020 [158]. According to the authors, none of the consulted articles of the eight most productive journals focused on marketing. This was attributed to the lack of interest in the subject, although consumer behavior regarding reducing meat consumption was identified as a gap and relevant topic for marketing research. Although the reduction in animal-based protein was identified earlier, the outcomes from a transition to plant-based protein alternatives have been explored only recently. Sha and Xiong [159] also recognized an increase in scientific publications in the last decade approaching meat alternatives and analogs.

The need for a filter and northernizing the information completes the canvas of our state-of-the-art of green mass protein production in America. In this brief, we sought articles and information derived or associated with data gathered in the American continent. The adoption of plant-based protein is expected for several reasons, as pointed out in Moreira et al.’s [158] analysis, focused on North American, European, and Oceania countries. Plant protein and plant-based meat are increasing, concomitantly with product diversity, regardless of the lack of understanding of the consumer view [157]. This intriguing observation let out of the block a missing engine that is propelling this system, and it is based not only on the consumer.

**Table 5.** Internet search engine (selected) results with keywords “Plant-based foods” and “Plant-based protein”.

Heading of the Source	Address (Internet)/Reference (Patent)	Main Information (Alleged)/Abstract
10 Best Sources of Plant-Based Protein by Whitney E. RD	<a href="https://www.house-foods.com/eat-happy/10-best-sources-of-plant-based-protein-by-whitney-e.-rd">https://www.house-foods.com/eat-happy/10-best-sources-of-plant-based-protein-by-whitney-e.-rd</a> accessed on 28 November 2022	Protein sources for a plant-based diet (Personal information)
Australian Plant Proteins: Optimized faba bean protein extraction	<a href="https://www.csiro.au/en/work-with-us/funding-programs/sme/csiro-kick-start/app">https://www.csiro.au/en/work-with-us/funding-programs/sme/csiro-kick-start/app</a> accessed on 28 November 2022	Upcoming of plant-based protein products. Disclosure of ongoing research (institutional)
Plant-Based Foods & Proteins Summit Americas	<a href="https://bridge2food.com/summits/americas/">https://bridge2food.com/summits/americas/</a> accessed on 28 November 2022	Ongoing and upcoming courses and meetings on innovation, business, and industry data on plant-based food and products. Disclosure of information on innovation, market and training
Go Plant-Based with Pistachios	<a href="https://americanpistachios.org/nutrition-and-health/the-plant-based-athlete">https://americanpistachios.org/nutrition-and-health/the-plant-based-athlete</a> accessed on 28 November 2022 <a href="https://americanpistachios.org/sites/default/files/inline-files/GoPlantBasedWithPistachiosFactSheet_112416.pdf">https://americanpistachios.org/sites/default/files/inline-files/GoPlantBasedWithPistachiosFactSheet_112416.pdf</a> accessed on 28 November 2022	Advertisement and information of a organized society of a protein-rich plant source (pistachio)
Plant-Based Protein Market— Global and Canadian Market Analysis	<a href="https://nrc.canada.ca/en/research-development/research-collaboration/programs/plant-based-protein-market-global-canadian-market-analysis">https://nrc.canada.ca/en/research-development/research-collaboration/programs/plant-based-protein-market-global-canadian-market-analysis</a> accessed on 30 November 2022	Executive summary of plant-based protein market, advertisement
Plant-Based Protein Market—Market Insights on Plant-Based Protein covering sales outlook, demand forecast & up-to-date key trends	<a href="https://www.futuremarketinsights.com/reports/plant-based-protein-market">https://www.futuremarketinsights.com/reports/plant-based-protein-market</a> accessed on 30 November 2022	Plant-based protein market analysis/report, prospective advertisement, and information for financial investment
Plant-based Protein Market Forecast, 2021–2031	<a href="https://www.transparencymarketresearch.com/plantbased-protein-market.html">https://www.transparencymarketresearch.com/plantbased-protein-market.html</a> accessed on 30 November 2022	Plant-based protein market analysis/report, prospective advertisement, and information for financial investment
Plant-based Protein Market by Type (Soy Protein, Wheat Protein, Pea Protein, Potato Protein, Rice Protein, Corn Protein), Crop Type (GMO), Source Process (Organic), Application (Food and Beverages, Animal Feed, Nutritional Supplements)—Global Forecast to 2028	<a href="https://www.meticulousresearch.com/product/plant-based-protein-market-5031">https://www.meticulousresearch.com/product/plant-based-protein-market-5031</a> accessed on 30 November 2022	Plant-based protein market analysis/report, prospective advertisement, and information for financial investment

Table 5. Cont.

Heading of the Source	Address (Internet)/Reference (Patent)	Main Information (Alleged)/Abstract
Plant Based Protein Market Worth \$23.4 Billion By 2028—Exclusive Report by Meticulous Research®	<a href="https://www.globenewswire.com/en/news-release/2022/01/03/2360111/0/en/Plant-Based-Protein-Market-Worth-23-4-Billion-By-2028-Exclusive-Report-by-Meticulous-Research.html">https://www.globenewswire.com/en/news-release/2022/01/03/2360111/0/en/Plant-Based-Protein-Market-Worth-23-4-Billion-By-2028-Exclusive-Report-by-Meticulous-Research.html</a> accessed on 30 November 2022	Plant-based protein market analysis/report, prospective advertisement, and information for financial investment
Plant-based proteins: A growth industry in Canada’s backyard	<a href="https://www.edc.ca/en/blog/canada-plant-based-protein-growth.html">https://www.edc.ca/en/blog/canada-plant-based-protein-growth.html</a> accessed on 17 January 2023	Plant-based protein market analysis/report, prospective advertisement, and information for financial investment
Plant-based Proteins Market-Market Study on Plant-based Proteins: Popularity of Pea & Wheat Proteins to Rise Faster Than Others	<a href="https://www.persistencemarketresearch.com/market-research/plantbased-protein-market.asp">https://www.persistencemarketresearch.com/market-research/plantbased-protein-market.asp</a> accessed on 17 January 2023	Plant-based protein market analysis/report, prospective advertisement, and information for financial investment
Plant-based Foods Market to Hit \$162 Billion in Next Decade, Projects Bloomberg Intelligence	<a href="https://www.bloomberg.com/company/press/plant-based-foods-market-to-hit-162-billion-in-next-decade-projects-bloomberg-intelligence/">https://www.bloomberg.com/company/press/plant-based-foods-market-to-hit-162-billion-in-next-decade-projects-bloomberg-intelligence/</a> accessed on 28 November 2022	Plant-based protein market analysis/report, prospective advertisement, and information for financial investment
Plant Based Protein Market, By Source (Soybeans, Wheat, Pea, Others), By Type (Isolates, Concentrates, Textured), By Form (Dry Form, Wet Form), By Application, and By Region Forecast to 2030	<a href="https://www.emergenresearch.com/industry-report/plant-based-protein-market">https://www.emergenresearch.com/industry-report/plant-based-protein-market</a> accessed on 30 November 2022	Plant-based protein market analysis/report, prospective advertisement, and information for financial investment
Alternative Proteins Market	<a href="https://www.datamintelligence.com/research-report/alternative-proteins-market">https://www.datamintelligence.com/research-report/alternative-proteins-market</a> accessed on 30 November 2022	Plant-based protein market analysis/report, prospective advertisement, and information for financial investment
Alternative proteins: The race for market share is on	<a href="https://www.mckinsey.com/~media/McKinsey/Industries/Agriculture/Our%20Insights/Alternative%20proteins%20The%20race%20for%20market%20share%20is%20on/Alternative-proteins-The-race-for-market-share-is-on.pdf">https://www.mckinsey.com/~media/McKinsey/Industries/Agriculture/Our%20Insights/Alternative%20proteins%20The%20race%20for%20market%20share%20is%20on/Alternative-proteins-The-race-for-market-share-is-on.pdf</a> accessed on 30 November 2022	Plant-based protein market analysis/report, prospective advertisement, and information for financial investment



Table 5. Cont.

Heading of the Source	Address (Internet)/Reference (Patent)	Main Information (Alleged)/Abstract
Plant Based Protein Supplements Market Size, Share & Trends Analysis Report By Raw Material (Soy, Spirulina, Pumpkin Seed, Wheat, Hemp, Rice, Pea, Others), By Product, By Distribution Channel, By Application, By Region, And Segment Forecasts, 2022–2030	<a href="https://www.grandviewresearch.com/industry-analysis/plant-based-protein-supplements-market">https://www.grandviewresearch.com/industry-analysis/plant-based-protein-supplements-market</a> accessed on 17 January 2023	Plant-based protein market analysis/report, prospective advertisement, and information for financial investment
Vegan Protein Market: Global Industry Analysis and Trends (2022–2029) Key Trends, Technology Trends, Market Share and Size	<a href="https://www.maximizemarketresearch.com/market-report/global-vegan-protein-market/87218/">https://www.maximizemarketresearch.com/market-report/global-vegan-protein-market/87218/</a> accessed on 30 November 2022	Plant-based protein market analysis/report, prospective advertisement, and information for financial investment
Increased Usage of Plant Based Protein for Various Applications is Anticipated to Accelerate the Overall Growth of the Market Further	<a href="https://www.databridgemarketresearch.com/press-release/global-plant-based-protein-market">https://www.databridgemarketresearch.com/press-release/global-plant-based-protein-market</a> accessed on 28 November 2022	Plant-based protein market analysis/report, prospective advertisement, and information for financial investment
Brazil's Future Farm launches its US expansion with the mission to democratize plant-based meat	<a href="https://www.fooddive.com/news/brazils-future-farm-launches-its-us-expansion-with-the-mission-to-democrat/602655/">https://www.fooddive.com/news/brazils-future-farm-launches-its-us-expansion-with-the-mission-to-democrat/602655/</a> accessed on 28 November 2022	Plant-based meat advertisement/interview
Latin America & Caribbean: Green finance state of the market 2019 I	<a href="https://www.climatebonds.net/resources/reports/latin-america-caribbean-green-finance-state-market-2019">https://www.climatebonds.net/resources/reports/latin-america-caribbean-green-finance-state-market-2019</a> accessed on 26 November 2022	Green bonds market analysis/report, prospective advertisement and information for financial investment
Plant Protein Primer, Exploring the Landscape of Plant Protein Sources for Applications in Plant-Based Meat, Eggs, and Dairy	<a href="https://gfi.org/wp-content/uploads/2021/02/2021-02-23_Plant_Protein_Primer_GFI.pdf">https://gfi.org/wp-content/uploads/2021/02/2021-02-23_Plant_Protein_Primer_GFI.pdf</a> accessed on 28 November 2022	Presentation, plant species for protein extraction, market analysis
Plant-based Protein Market by Source (Soy, Wheat, and Pea), Type (Isolates, Concentrates, and Textured), Application (Dairy Alternatives, Meat Alternatives, and Performance Nutrition, Animal Feed), and Region (North America, Europe, Asia Pacific, South America, Middle East and Africa), Global trends and forecast from 2019 to 2028	<a href="https://exactitudeconsultancy.com/reports/1246/plant-based-protein-market/">https://exactitudeconsultancy.com/reports/1246/plant-based-protein-market/</a> accessed on 28 November 2022	Plant-based protein market analysis/report, prospective advertisement, and information for financial investment
The future of plant-based food, according to industry leaders	<a href="https://www.veganfoodandliving.com/vegan-business/the-future-of-plant-based-food/">https://www.veganfoodandliving.com/vegan-business/the-future-of-plant-based-food/</a> accessed on 30 November 2022	Plant-based food and protein market analysis/report, prospective advertisement, and information for financial investment

Table 5. Cont.

Heading of the Source	Address (Internet)/Reference (Patent)	Main Information (Alleged)/Abstract
Plant-based proteins: building a sustainable future	<a href="https://impact.economist.com/perspectives/sites/default/files/plant_based_protein_eiu_infographic.pdf">https://impact.economist.com/perspectives/sites/default/files/plant_based_protein_eiu_infographic.pdf</a> accessed on 30 November 2022	Infographic on Plant-based food and protein market analysis/report, prospective advertisement, and information for financial investment
Plant-Based Innovation For Latin America: Beyond Burgers	<a href="https://www.mintel.com/blog/food-market-news/plant-based-innovation-for-latin-america-beyond-burgers">https://www.mintel.com/blog/food-market-news/plant-based-innovation-for-latin-america-beyond-burgers</a> accessed on 2 December 2022	
NotCo becomes Chile's newest unicorn	<a href="https://www.leadersleague.com/fr/news/notco-becomes-chile-s-newest-unicorn">https://www.leadersleague.com/fr/news/notco-becomes-chile-s-newest-unicorn</a> accessed on 17 January 2023	Financial information of Plant-based food company.
Creating a Sustainable Food Future	<a href="https://research.wri.org/wrr-food">https://research.wri.org/wrr-food</a> accessed on 30 November 2022	Folder on sustainable food production from World Resources Institute
You Heard it Here first: The Plant-Based Revolution	<a href="https://www.mintel.com/blog/food-market-news/you-heard-it-here-first-predicting-the-plant-based-revolution">https://www.mintel.com/blog/food-market-news/you-heard-it-here-first-predicting-the-plant-based-revolution</a> accessed on 2 December 2022	Data on plant-based food trend
Fazenda do futuro	<a href="https://www.fazendafuturo.io/pt-br">https://www.fazendafuturo.io/pt-br</a> accessed on 17 January 2023	Home site of Future Farm Company
NotCo	<a href="https://notco.com/br/">https://notco.com/br/</a> accessed on 17 January 2023	Home site of NotCo Company
Beyond Meat	<a href="https://www.beyondmeat.com/en-US/">https://www.beyondmeat.com/en-US/</a> accessed on 17 January 2023	Home site of Beyond Meat Company
The New Live Geen Co.	<a href="https://thenewbutchers.com.br/nossos-produtos/">https://thenewbutchers.com.br/nossos-produtos/</a> accessed on 17 January 2023	Home site of The Live Green Company
	<a href="https://www.thelivegreenco.com/">https://www.thelivegreenco.com/</a> accessed on 17 January 2023	Home site of NotCo Company

Table 5. Cont.

Heading of the Source	Address (Internet)/Reference (Patent)	Main Information (Alleged)/Abstract
Patents	Address (internet)/Reference (patent)	Main information (alleged)/Abstract
Protein Compositions for Plant-Based Food Products and Methods for Making	WO 2021/119498, June, 2021. accessed on 28 November 2022	Disclosed is a method for making protein emulsions for use in making products such as meat substitutes, meat extenders, egg substitutes, dairy analogs, etc., as well as methods for using the emulsion(s) to make various meat substitutes, egg substitutes, dairy analogs etc. Vegetable protein crumbles for use as meat substitutes are also disclosed, either alone or in combination with the emulsion(s).
Protein-Rich microalgal biomass compositions of optimized sensory quality	US 10,119,947 B2, November, 2018. accessed on 28 November 2022	The invention relates to a method for determining the organoleptic quality of protein-rich microalgal biomass composition, comprising the determination of the content of 11 volatile organic compounds, wherein the 11 volatile organic compounds are pentanal, hexanal, 1-octen-2-one), 3,5-octadien-2-one, nonanal, 2-nonenal, (E, E)-2,4-nonadienal and hexanoic acid.

Marinangeli et al. [160] compiled the importance, advantages, and market of plant-based protein, addressing the need for a regulatory landscape and formulation guidance for these innovative products. For instance, the requirement for standards for essential amino acids and digestibility approaches of these plant-based products according to life stage and health are needed. Oppositely, the choice of beef and plant-based foods were not affected by the nutrition facts panels or ingredient lists [161]. There are differences in the willingness for the preference and nutritional perception of these foods based on the assortment of general consumers and nutrition professionals [157]. This highlights the importance of information as a promotor for the consumption and adjustments of the composition [156], organoleptic properties, and other traits [135,159] of plant-based foods and proteins.

The negative relationship between predisposition changes in behavior for the adoption of a sustainable diet and the lack of information on plant-based diets [142] pinches objectives for future research. Focus themes such as refining the positive health effects, affordable products, improving behavior, and the disclosure of amino acid and digestibility standards of the plant-based proteins are other research inputs. The belief that these products contribute to good health [150,151] and are friendlier to the environment [136] may lead consumers to be increasingly amenable to plant-based diets and plant-based meat alternatives. Accordingly, economic modeling suggests positive effects of the adoption of plant-based beef alternatives such as reduced greenhouse gas emissions, reduced carbon footprint, and exports of agriculture products. On the other hand, it may result in the disruption of cattle and beef processing sectors throughout the agriculture economy, which will face a complex ethical and political transition [145].

Despite the origin (species, plant sample, and the use of raw or processed samples) of plant-based protein products, there is a niche for the development and commerce of alternative foods derived from plant biomass. The innovative capacity and speed of product development of the rising food companies contribute to the landscape. In Latin America, there are intents and actions recognized as a laboratory and a showcase for inhibiting the consumption of (ultra) processed food [162]. Nevertheless, there is an increase in the consumption of industrialized food and other sources of animal protein [140] due to the increase in population income and cheaper nutritional sources [136,162,163].

Consumers are recently challenged with new plant-based products that illustrate the innovation capacity and the speed of action expected from South America's entrepreneurs. "Future Farm" brand, a plant-based meat company in Brazilian and abroad markets. It is also working on and planning the release of plant-based milk, cheese, and chicken products. Chile's NotCo is an important food company founded in 2015 that is investing in plant-based lactic products and is considered one of the fastest-growing Food Companies in LATAM. There are other South American brands, such as "The Live Green Co." and "The New Butchers", and North American brands, such as "Beyond Meat" [135] and others that are investing in plant-based food products (Table 5).

De Marchi et al. [139] reported similar characteristics of meat-based (MBB) and plant-based burgers (PBB), such as color, pH, gross composition, total fatty acid profile, and protein, although there were more carbohydrates and fiber content (PBB) and significant differences in amino acids, polyunsaturated fatty acids, cholesterol, and minerals. Soy and pea protein and beet are listed as ingredients for the plant-based protein PBB. Other interesting plant-based products are beverages enriched with plant proteins. Despite Arbach et al.'s [152] review, which basically reports the use of seed-protein in these beverages that benefited from lower prices and environmental damage, there is also an application for green juices and plant biomass [164].

It seems that seed-protein-based ingredients are the first option, although there is an open window for other plant-biomass ingredients. There are efforts to use green juice from plant biomass as possible raw material for plant-based beverages [165]. There are some promising initiatives of plants that should be revisited as protein sources, such as the use of cassava leaves [166] that, probably to HCN content in leaf tissues, seem to be

abandoned. Alternative ingredients, either non-seed or non-animal origin, for these new food and non-food products are also available [159]. The use of ingredients from other plant species may contribute to equalizing meat-based and plant-based product characteristics. Biofabric products are alternatives that can contribute to an increased bioaccessibility and bioavailability of green plant biomass [137,138,148,164,167]. This includes the use of biofabrics to extract protein from plant biomass [148,164].

Regardless of pioneer initiatives [139,156,167], a further contribution from academic research could be the development of protein from green biomass as a potential alternative to seed-based protein. It should be noted that there are other applications for plant-based materials, such as fuel production [164,168,169], that are not discussed here. The key to successful application lies in the processing possibilities and in the applied plant species/varieties [138,148].

In summary, there are gaps in the chains among commerce, consumers, industry, and producer, and the development and research on plant-based protein products ought to be fulfilled to achieve the benefits of these food alternatives. The high technology, investments, and productivity of countries such as the US, the tradition in agribusiness, the territory availability of countries such as Brazil, and entrepreneur initiatives and companies, indicate that protein and other products derived from plant biomass is a fertile areas in American countries.

### 5.3. State of Art of Green Biomass-Originated Protein in Africa

Poverty and food shortage, especially a protein-rich diet, are the driving factors of malnutrition and its detrimental consequences in Africa [170]. Green biorefinery may serve as a substituent for food insecurity as it helps to fulfill food and feed needs for Africa. Processing green biomass into acceptable products for direct human consumption may have nutritional, economic, and environmental merits [171].

According to several researches being conducted, there are some particular plants to use as biomass for a sustainable protein supply in Africa. The most prominent ones are *Moringa oleifera*, *Manihot esculenta*, *Glyricidia sepium*, and *Leucaena leucocephala*.

*Moringa oleifera* is a fast-growing tree from the family of Moringaceae, which is highly drought-resistant. It has a high profile of nutritional composition and is most widely cultivated in Africa. *Moringa oleifera* leaf protein concentrate is a promising source of protein for most of the developing countries in Africa. LPC obtained from *Moringa oleifera* is nutritionally valuable [170].

According to Sodamade's research [170], *Moringa oleifera*'s leaf protein concentrate's moisture content is  $9.00 \text{ mg } 100 \text{ g}^{-1}$ , ash content is  $6.00 \text{ mg } 100 \text{ g}^{-1}$ , crude fat is  $2.43 \text{ mg } 100 \text{ g}^{-1}$ , crude fiber is  $5.43 \text{ mg } 100 \text{ g}^{-1}$ , carbohydrate content is  $38.21 \text{ mg } 100 \text{ g}^{-1}$ , and crude protein content is  $39.13 \text{ mg } 100 \text{ g}^{-1}$ . This remarkable amount of crude protein in the plant means that *Moringa oleifera* leaf protein concentrate may be evaluated as nutritionally valuable and a healthy ingredient to improve protein needed in the human diet and animal feed. Its functional properties, such as water absorption capacity, fat absorption capacity, emulsion capacity, and foaming stability, are also significant [170].

*Manihot esculenta*, also known as cassava or manioc, is a tuberous edible plant of the Euphorbiaceae family. Since cassava flour, bread, tapioca, an alcoholic beverage, and laundry starch are derived from its tuberous roots, it is cultivated in many plantations in Africa. Moreover, 250 million Africans make use of the starchy root crop *Manihot esculenta* as an important part of their diet [172].

Cassava plays an important role in agriculture, especially in sub-Saharan African countries. Cassava leaves are consumed as a major source of dietary protein for all of Central Africa, most of East Africa, and even some parts of West Africa [173].

Both the leaves and roots of the cassava plant are nutritionally valuable, and they offer the potential as a feed source. The root of cassava is mostly a carbohydrate resource that contains 60–65% moisture, 20–31% carbohydrate, and 1–2% crude protein. It contains a relatively low content of vitamins and minerals [174].

Cassava leaves are potential biomass that is affluent in protein with a balanced content of amino acids. Thus the leaves represent valuable biomass for the extraction of proteins. According to Gundersen's study [74], between 21% and 26% (*w/w*) of leaf crude protein can be recovered in the leaf protein concentrates. After the drying process, the product contains 40–45% crude protein with an amino acid notable profile that can be compared with soybean. Its level of tannins is tolerable in the case of animal feed purposes [74].

The cassava leaf protein concentrate's (LPC) amino acid profile is fairly similar for the starting leaf material and the press cake. Except for aspartic and glutamic acid, for most amino acids, the content is barely lower in the produced green juice. In research that compared the amino acids profiles of cassava LPCs obtained by heat coagulation and acid precipitation with the soybean reference, it was found that cassava LPCs' content of methionine, leucine, and valine is higher than the soybean reference [74].

On the other hand, Cassava has some antinutritional elements. Cassava leaves' cyanogenic glucosides potential is 5 to 20 times higher than the cyanogenic potential of its roots. Although there is a risk of intoxication held by the consumption of cassava leaves, during processing, the risk is decreased due to the capacity of the leaves to release cyanogens quickly [173]. The releasable HCN amount existing in the dried protein product is around 150–250 ppm. However, this amount is still higher than 10–50 ppm which is considered safe for food and feed purposes by the food safety authorities [74].

In sub-Saharan Africa, *Glyricidia sepium* and *Leucaena leucocephala* are also notable plants for biorefinery purposes. They have foliage production ability for all-year-round. Moreover, they are rich in protein, minerals, and vitamins [175].

*Glyricidia sepium* is a tropical forage plant from the Fabaceae family. Its leaves are considered to contain high protein content and are suitable for producing protein-rich forage with its high nutritive value. It is a plant that shows a wide distribution and variation in productivity. *Leucaena leucocephala* is a fast-growing evergreen plant from the Fabaceae family. Its young leaves and seeds may be used as a vegetable in human nutrition [176].

In sub-Saharan Africa, *Glyricidia sepium* and *Leucaena leucocephala* leaves mount up all year round. The leaves have rich protein, minerals, vitamins, and amino acid content. Therefore, *Glyricidia sepium* and *Leucaena leucocephala* are convenient for producing leaf protein concentrates. According to the research carried out by Agbede and Aletor [176], *Leucaena leucocephala* leaves involve lower crude protein and higher crude fiber than *Glyricidia sepium* leaves, but their ash values are equal. Their crude protein in the LPCs showed amino acids with a good balance. Their LPCs have similar amino acid profiles, but the values of *Glyricidia* LPC (G. LPC) are generally a little higher than *Leucaena* LPC (L. LPC), except for proline and methionine. Some of the amino acid compositions ( $\text{g } 100 \text{ g}^{-1}$  sample) of leaf protein concentrate that were measured during their research are lysine 5.99 in L. LPC and 6.60 in G. LPC; histidine 2.11 in L. LPC and 2.51 in G. LPC; arginine 5.54 in L. LPC and 6.30 in G. LPC; Threonine 4.61 in L. LPC and 5.08 in G. LPC; methionine 2.25 in L. LPC and 2.05 in G. LPC. Due to this high nutritional amino acid concentration, *Glyricidia* and *Leucaena* LPCs are comparable with whole egg amino acids profile [176].

In conclusion, with its relatively high protein content and excellent amino acid profile, *Glyricidia* or *Leucaena* LPC may be a successful substitute for the soybean, which is a more expensive protein source and a sustainable alternative for supplying affordable food in Africa [175].

**Funding:** This work has been implemented with the TKP2021-EGA-20 and TKP2020-NKA-04 support provided by the National Research, Development and Innovation Fund of Hungary. The present work is supported by the 2C5SBMGA0045-2021-1.2.4-TÉT *Plant species targeting for green biorefining purposes in Brazil and the Carpathian Basin, their processing technologies and product development possibilities* entitled project. Éva Domokos-Szabolcsy was supported by the János Bolyai Research Scholarship of the Hungarian Academy of Sciences. Zoltán Kovács was supported by the National Research, Development and Innovation Fund (Hungary) Science Patronage Program via MEC\_R 141362 *Green biorefining concept and presentation of Hungarian innovations at a Brazilian agricultural exhibition* entitled project.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

- Wu, G. Amino Acids: Metabolism, Functions, and Nutrition. *Amino Acids* **2009**, *37*, 1–17. [[CrossRef](#)] [[PubMed](#)]
- Hambraeus, L. Protein and Amino Acids in Human Nutrition. In *Reference Module in Biomedical Sciences*; Elsevier: Amsterdam, The Netherlands, 2014; ISBN 978-0-12-801238-3.
- Pirie, N.W. *Leaf Protein: And Its By-Products in Human and Animal Nutrition*; Cambridge University Press: Cambridge, UK, 1987; ISBN 978-0-521-33030-5.
- Parrish, G.K.; Kroger, M.; Weaver, J.C.; Furia, T. The Prospects of Leaf Protein as a Human Food-and a Close Look at Alfalfa. *Crit. Rev. Food Technol.* **1974**, *5*, 1–13. [[CrossRef](#)]
- Kinsella, J. Evaluation of Plant Leaf Protein as a Source of Food Protein. *Chem. Ind.* **1970**, *17*, 550–554. [[PubMed](#)]
- Pirie, N.W. Protein Nutritional Quality. *Baroda J. Nutr.* **1975**, *2*, 43–48.
- Bud, R. History of “Biotechnology”. *Nature* **1989**, *337*, 10. [[CrossRef](#)]
- Bud, R. Janus-Faced Biotechnology: An Historical Perspective. *TIBTECH* **1989**, *7*, 230–233. [[CrossRef](#)]
- Bud, R. Biotechnology in the Twentieth Century. *Soc. Stud. Sci.* **1991**, *21*, 415–457. [[CrossRef](#)]
- Bud, R. *The Uses of Life: A History of Biotechnology*, 1st ed.; Cambridge University Press: Cambridge, UK, 1994; ISBN 978-0-521-47699-7.
- Fári, M.G.; Menezes, E.A.; Bud, R.; Kralovanszky, P.U. Horticultural Biotechnology, as Efficient Means for Developing Countries Suggested by K. Ereky. *Acta Hort.* **2001**, *560*, 557–564. [[CrossRef](#)]
- Fári, M.G.; Kralovanszky, U.P. The Founding Father of Biotechnology: Károly (Karl) Ereky. *Int. J. Hortic. Sci.* **2006**, *12*, 9–12. [[CrossRef](#)]
- Fári, M.G.; Popp, J. (Eds.) *Biotechnology-Anno 1917–1919. Károly Ereky’s Vision about the Application of Life Sciences. (Ereky Károly Viziója az Élettudomány Alkalmazásáról, in Hungarian)*; Szaktudás Publishing House: Budapest, Hungary, 2015; ISBN 978-615-5224-65-2.
- Fári, M.G.; Popp, J. (Eds.) *Biotechnology-Anno 1920–1938 and Today. In Károly Ereky’s Program About Solving the Protein Problem and Today’s Tasks, (Biotechnológia anno 1920–1938 és ma-Ereky Károly programja a fehérjeprobléma megoldásáról és napjaink feladatai, in Hungarian)*; Szaktudás Publishing House: Budapest, Hungary, 2016; ISBN 978-615-5224-67-6.
- Ereky, K. *Biotechnologie der Fleisch-, Fett-und Milcherzeugung im Landwirtschaftlichen Grossbetriebe für Naturwissenschaftlich Gebildete Landwirte, Verfasst*; Paul Parey: Singhofen, Germany, 1919.
- Ereky, K. *Green Fodder Pulp Mill and Large-Scale Animal Husbandry Enterprises: (A Zöldtakarmánymalom és a Nagy Istállóüzemek, in Hungarian)*; Athenaeum Irodalmi és Nyomdai Részvény-Társulat: Budapest, Hungary, 1925.
- Hammond, J. Agricultural Physiology. In *Science Progress in the Twentieth Century (1919–1933)*; Sage Publications Ltd.: London, UK, 1929; Volume 24, pp. 231–238.
- Ereky, K. Die Steigerungsmöglichkeiten der landwirtschaftlichen Lebensmittelproduktion. *Naturwissenschaften* **1920**, *8*, 1033–1038. [[CrossRef](#)]
- Ereky, K. De Factoren Tot Vermeerdering Der Landbouwproducten. *U. H. Duitsch.* **1921**, *3*, 314–326.
- Ereky, K. Process for the Manufacture and for the Preservation of Green Fodder Pulp or Other Plant Pulp and Dry Product Made Therefrom. British Patent No. 270,629, 28 September 1926.
- Ereky, K. Eljárás Zöldnövény-Pép Előállítására És Konzerválására. Hungarian Patent No. 92,680, 7 May 1926.
- Ereky, K. Procédé Pour Préparer et Conserver de la Pulpe D’herbages. EU Patent No. 623,150, 14 October 1926.
- Ereky, K. Verfahren zur Herstellung von Grünpflanzenbrei Und Konservierung Desselben. Austrian Patent No. 118,622, 25 July 1926.
- Ereky, K. Improvements in Machines Suitable for the Manufacture of Green Fodder Pulp. British Patent No. 291,752, 27 September 1927.
- Ereky, K. Gép Főleg Zöldnövényepép Előállítására. Hungarian Patent No. 95,006, 8 June 1927.
- Ereky, K. Pulp Manufacture (Production de Pulpe). Canadian Patent No. 282,415, 14 August 1928.
- Ereky, K. New Methods of Preparing Green Vegetables for the Table. *Food Manuf.* **1927**, *2*, 207.
- Ereky, K. *Grünbreimühle und Grünkonservenfabrikation*; Königliche Ungarische Universität Druckerei: Budapest, Hungary, 1926; p. 26.
- Ereky, K. *Green Pulper and Green Mill*; Atheneum Books: New York, NY, USA, 1926.
- Ereky, K. *Reconstruction of the Hungarian Agriculture: (A Magyar Mezőgazdaság Rekonstrukciója)*; Mezőgazdasági Könyvtár: Budapest, Hungary, 1928. (in Hungarian)
- Duckham, A.N. Grass and Fodder Crop Conservation in Transportable Form. Memorandum. Agricultural Economics in the Empire. Report of a Committee Appointed by the Empire. *Mark. Board* **1928**, *8*, 1–43.
- Goodall, C. Improvements Relating to the Treatment of Grass and Other Vegetable Substances. British Patent No. 457,789, 7 December 1935.
- Smith, H.A. Process for the Manufacture of a Feed Material from Lettuce. U.S. Patent No. 2,190,176, 19 August 1937.
- Slade, R.E.; Birkinshaw, J.H. Improvements in, or Relating to the Utilization of Grass and Other Green Crops. British Patent GB511525A, 18 February 1938.
- Slade, R.E.; Branscombe, D.J.; Gount, W.E. Improvement in or Relating to the Production of Food from Plant Leaves. British Patent No. 577,172, 8 May 1940.
- Slade, R.E.; Braitscombe, D.J.; McGowan, J.C. Protein extraction. *Chem. Ind.* **1946**, *25*, 194–197.
- Pirie, N.W. The Direct Use of Leaf Protein in Human Nutrition. *Chem. Ind.* **1942**, *61*, 45–48.
- Pirie, N.W. Green Leaves as a Source of Proteins and Other Nutrients. *Nature* **1942**, *149*, 251. [[CrossRef](#)]
- Pirie, N.W. Some practical aspects of leaf protein manufacture. *Food Manuf.* **1942**, *17*, 283–286.

40. Fiorentini, R.; Galoppini, C. The Proteins from Leaves. *Plant Food Hum. Nutr.* **1983**, *32*, 335–350. [[CrossRef](#)]
41. Kromus, S.; Kamm, B.; Kamm, M.; Fowler, P.; Narodoslowsky, M. Green Biorefineries: The Green Biorefinery Concept—Fundamentals and Potential. In *Biorefineries—Industrial Processes and Products*; John Wiley & Sons, Ltd.: Hoboken, NJ, USA, 2005; pp. 253–294, ISBN 978-3-527-61984-9.
42. Tamayo Tenorio, A.; Kyriakopoulou, K.E.; Suarez-Garcia, E.; Van den Berg, C.; Van der Goot, A.J. Understanding Differences in Protein Fractionation from Conventional Crops, and Herbaceous and Aquatic Biomass—Consequences for Industrial Use. *Trends Food Sci. Technol.* **2018**, *71*, 235–245. [[CrossRef](#)]
43. Nishimura, K.; Ogawa, T.; Ashida, H.; Yokota, A. Molecular Mechanisms of RuBisCO Biosynthesis in Higher Plants. *Plant Biotechnol.* **2008**, *25*, 285–290. [[CrossRef](#)]
44. Ellis, R.J. Chloroplast Proteins and Their Synthesis. In *Plant Proteins*; Butterworths: London, UK, 1977.
45. Andersson, I.; Backlund, A. Structure and Function of Rubisco. *Plant Physiol. Biochem.* **2008**, *46*, 275–291. [[CrossRef](#)] [[PubMed](#)]
46. Nieuwland, M.; Geerdink, P.; Engelen-Smit, N.P.; Van der Meer, I.M.; America, A.H.; Mes, J.J.; Kootstra, A.M.J.; Henket, J.T.; Mulder, W.J. Isolation and Gelling Properties of Duckweed Protein Concentrate. *ACS Food Sci. Technol.* **2021**, *1*, 908–916. [[CrossRef](#)]
47. Guide to Genetically Modified Alfalfa. Available online: <https://www.worc.org/publication/guide-to-genetically-modified-alfalfa/> (accessed on 20 December 2022).
48. Reid, W.V.; Mooney, H.A.; Cropper, A.; Capistrano, D.; Carpenter, S.R.; Chopra, K.; Dasgupta, P.; Dietz, T.; Duraiappah, A.K.; Hassan, R.; et al. *Ecosystems and Human Well-Being—Synthesis: A Report of the Millennium Ecosystem Assessment*; Island Press: Washington, DC, USA, 2005; ISBN 978-1-59726-040-4.
49. Wiggering, H.; Finckh, M.R.; Heß, J. *Fachforum Leguminosen: Wissenschaft, Wirtschaft, Gesellschaft—Ökosystemleistungen von Leguminosen Wettbewerbsfähig Machen: Forschungsstrategie der Deutschen Agrarforschungsallianz*; Stand 07/2012; Deutsche Agrarforschungsallianz (DAFA): Braunschweig, Germany, 2012; ISBN 978-3-86576-092-0.
50. Wilkins, R.J.; Jones, R. Alternative Home-Grown Protein Sources for Ruminants in the United Kingdom. *Anim. Feed. Sci. Technol.* **2000**, *85*, 23–32. [[CrossRef](#)]
51. Früh, B.; Schlatter, B.; Isensee, A.; Maurer, V.; Willer, H. *Report on Organic Protein Availability and Demand in Europe = Deliverable 1.2 of the CORE Organic Project (ICOPP)*; Research Institute of Organic Agriculture (FiBL): Frick, Switzerland, 2014.
52. Li, X.; Brummer, E.C. Applied Genetics and Genomics in Alfalfa Breeding. *Agronomy* **2012**, *2*, 40–61. [[CrossRef](#)]
53. Kaszás, L.; Alshaal, T.; El-Ramady, H.; Kovács, Z.; Koroknai, J.; Elhawat, N.; Nagy, É.; Cziáky, Z.; Fári, M.; Domokos-Szabolcsy, É. Identification of Bioactive Phytochemicals in Leaf Protein Concentrate of Jerusalem Artichoke (*Helianthus Tuberosus* L.). *Plants* **2020**, *9*, 889. [[CrossRef](#)]
54. Açıkgöz, E. *Yem bitkileri*; Uludağ Üniversitesi Ziraat Fakültesi Tarla Bitkileri Bölümü: Bursa, Turkey, 2001; ISBN 975-564-124-6.
55. Anil, L.; Park, J.; Phipps, R.H.; Miller, F.A. Temperate Intercropping of Cereals for Forage: A Review of the Potential for Growth and Utilization with Particular Reference to the UK. *Grass Forage Sci.* **1998**, *53*, 301–317. [[CrossRef](#)]
56. De Zumelzú, D.M.; Costero, B.; Cavaleri, P.; Maich, R. Selection Responses for Some Agronomic Traits in Hexaploid Triticale. *Agriscientia* **2002**, *19*, 45–50.
57. Kaszás, L.; Alshaal, T.; Kovács, Z.; Koroknai, J.; Elhawat, N.; Nagy, É.; El-Ramady, H.; Fári, M.; Domokos-Szabolcsy, É. Refining High-Quality Leaf Protein and Valuable Co-Products from Green Biomass of Jerusalem Artichoke (*Helianthus Tuberosus* L.) for Sustainable Protein Supply. *Biomass Conv. Bioref.* **2020**, *12*, 2149–2164. [[CrossRef](#)]
58. Jørgensen, H.; Thomsen, S.T.; Schjoerring, J.K. The Potential for Biorefining of Triticale to Protein and Sugar Depends on Nitrogen Supply and Harvest Time. *Ind. Crops Prod.* **2020**, *149*, 112333. [[CrossRef](#)]
59. Corona, A.; Parajuli, R.; Ambye-Jensen, M.; Hauschild, M.Z.; Birkved, M. Environmental Screening of Potential Biomass for Green Biorefinery Conversion. *J. Clean. Prod.* **2018**, *189*, 344–357. [[CrossRef](#)]
60. Tamayo Tenorio, A.; Gieteling, J.; De Jong, G.A.H.; Boom, R.M.; Van der Goot, A.J. Recovery of Protein from Green Leaves: Overview of Crucial Steps for Utilisation. *Food Chem.* **2016**, *203*, 402–408. [[CrossRef](#)]
61. Santamaría-Fernández, M.; Karkov Ytting, N.; Lübeck, M. Influence of the Development Stage of Perennial Forage Crops for the Recovery Yields of Extractable Proteins Using Lactic Acid Fermentation. *J. Clean. Prod.* **2019**, *218*, 1055–1064. [[CrossRef](#)]
62. Solati, Z.; Jørgensen, U.; Eriksen, J.; Søgaard, K. Dry Matter Yield, Chemical Composition and Estimated Extractable Protein of Legume and Grass Species during the Spring Growth: Protein Extractability in Legumes and Grasses. *J. Sci. Food Agric.* **2017**, *97*, 3958–3966. [[CrossRef](#)]
63. La Cour, R.; Schjoerring, J.K.; Jørgensen, H. Enhancing Protein Recovery in Green Biorefineries by Lignosulfonate-Assisted Precipitation. *Front. Sustain. Food Syst.* **2019**, *3*, 112. [[CrossRef](#)]
64. Zhang, W.; Grimi, N.; Jaffrin, M.Y.; Ding, L.; Tang, B. A Short Review on the Research Progress in Alfalfa Leaf Protein Separation Technology. *J. Chem. Technol. Biotechnol.* **2017**, *92*, 2894–2900. [[CrossRef](#)]
65. Siegert, W.; Boguhn, J.; Maurer, H.P.; Weiss, J.; Zuber, T.; Möhring, J.; Rodehutschord, M. Effect of Nitrogen Fertilisation on the Amino Acid Digestibility of Different Triticale Genotypes in Caecotomised Laying Hens: Effect of Nitrogen Fertilisation on the Amino Acid. *J. Sci. Food Agric.* **2017**, *97*, 144–150. [[CrossRef](#)]
66. Santamaría-Fernández, M.; Molinuevo-Salces, B.; Kiel, P.; Steinfeldt, S.; Uellendahl, H.; Lübeck, M. Lactic Acid Fermentation for Refining Proteins from Green Crops and Obtaining a High Quality Feed Product for Monogastric Animals. *J. Clean. Prod.* **2017**, *162*, 875–881. [[CrossRef](#)]
67. Damborg, V.K.; Jensen, S.K.; Weisbjerg, M.R.; Adamsen, A.P.; Stødkilde, L. Screw-Pressed Fractions from Green Forages as Animal Feed: Chemical Composition and Mass Balances. *Anim. Feed. Sci. Technol.* **2020**, *261*, 114401. [[CrossRef](#)]



68. Hansen, M.; Andersen, C.A.; Jensen, P.R.; Hobbey, T.J. Scale-Up of Alfalfa (*Medicago sativa*) Protein Recovery Using Screw Presses. *Foods* **2022**, *11*, 3229. [CrossRef]
69. Ravindran, R.; Koopmans, S.; Sanders, J.P.M.; McMahon, H.; Gaffey, J. Production of Green Biorefinery Protein Concentrate Derived from Perennial Ryegrass as an Alternative Feed for Pigs. *Clean Technol.* **2021**, *3*, 656–669. [CrossRef]
70. Domokos-Szabolcsy, É.; Elhawat, N.; Domingos, G.J.; Kovács, Z.; Koroknai, J.; Bodó, E.; Fári, M.G.; Alshaal, T.; Bákonyi, N. Comparison of Wet Fractionation Methods for Processing Broccoli Agricultural Wastes and Evaluation of the Nutri-Chemical Values of Obtained Products. *Foods* **2022**, *11*, 2418. [CrossRef] [PubMed]
71. Hanczakowski, P.; Skraba, B.; Młodkowski, M. Nutritive Value of Leaf-Protein Concentrate from Potato Haulm for Rats and Chicks. *Anim. Feed. Sci. Technol.* **1981**, *6*, 413–419. [CrossRef]
72. Jwanny, E.W.; Montanari, L.; Fantozzi, P. Protein Production for Human Use from Sugarbeet: Byproducts. *Bioresour. Technol.* **1993**, *43*, 67–70. [CrossRef]
73. Prade, T.; Muneer, F.; Berndtsson, E.; Nynäs, A.-L.; Svensson, S.-E.; Newson, W.R.; Johansson, E. Protein Fractionation of Broccoli (*Brassica oleracea*, Var. *Italica*) and Kale (*Brassica oleracea*, Var. *Sabellica*) Residual Leaves—A Pre-Feasibility Assessment and Evaluation of Fraction Phenol and Fibre Content. *Food Bioprod. Process.* **2021**, *130*, 229–243. [CrossRef]
74. Gundersen, E.; Christiansen, A.H.C.; Jørgensen, K.; Lübeck, M. Production of Leaf Protein Concentrates from Cassava: Protein Distribution and Anti-Nutritional Factors in Biorefining Fractions. *J. Clean. Prod.* **2022**, *379*, 134730. [CrossRef]
75. Hadidi, M.; Jafarzadeh, S.; Forough, M.; Garavand, F.; Alizadeh, S.; Salehabadi, A.; Khaneghah, A.M.; Jafari, S.M. Plant Protein-Based Food Packaging Films; Recent Advances in Fabrication, Characterization, and Applications. *Trends Food Sci. Technol.* **2022**, *120*, 154–173. [CrossRef]
76. Berndtsson, E.; Andersson, R.; Johansson, E.; Olsson, M.E. Side Streams of Broccoli Leaves: A Climate Smart and Healthy Food Ingredient. *IJERPH* **2020**, *17*, 2406. [CrossRef]
77. Berndtsson, E.; Nynäs, A.-L.; Newson, W.; Langton, M.; Andersson, R.; Johansson, E.; Olsson, M.E. 21. The Underutilised Side Streams of Broccoli and Kale—Valorisation via Proteins and Phenols. In *Sustainable Governance and Management of Food Systems*; Vinnari, E., Vinnari, M., Eds.; Wageningen Academic Publishers: Turku, Finland, 2019; pp. 153–159.
78. Baraniak, B.M.; Waleriańczyk, E. Flocculation. In *Encyclopedia of Food Sciences and Nutrition*, 2nd ed.; Caballero, B., Finglas, P., Toldra, F., Eds.; Academic Press: Oxford, UK, 2003; pp. 2531–2535. ISBN 978-0-12-227055-0.
79. Bals, B.D.; Dale, B.E.; Balan, V. Recovery of Leaf Protein for Animal Feed and High-Value Uses. In *Biorefinery Co-Products*; John Wiley & Sons, Ltd.: Hoboken, NJ, USA, 2012; pp. 179–197. ISBN 978-0-470-97669-2.
80. Pirie, N.W.; Pirie, N.W. *Leaf Protein: Its Agronomy, Preparation, Quality and Use*; IBP Handbook No. 20; International Biological Programme: London, UK, 1971; ISBN 978-0-632-08350-3.
81. Arlabosse, P.; Blanc, M.; Kerfaï, S.; Fernandez, A. Production of Green Juice with an Intensive Thermo-Mechanical Fractionation Process. Part I: Effects of Processing Conditions on the Dewatering Kinetics. *Chem. Eng. J.* **2011**, *168*, 586–592. [CrossRef]
82. Colas, D.; Doumeng, C.; Pontalier, P.Y.; Rigal, L. Green Crop Fractionation by Twin-Screw Extrusion: Influence of the Screw Profile on Alfalfa (*Medicago sativa*) Dehydration and Protein Extraction. *Chem. Eng. Process. Process Intensif.* **2013**, *72*, 1–9. [CrossRef]
83. Colas, D.; Doumeng, C.; Pontalier, P.Y.; Rigal, L. Twin-Screw Extrusion Technology, an Original Solution for the Extraction of Proteins from Alfalfa (*Medicago sativa*). *Food Bioprod. Process.* **2013**, *91*, 175–182. [CrossRef]
84. Byers, M.; Sturrock, J.W. The Yields of Leaf Protein Extracted by Large-Scale Processing of Various Crops. *J. Sci. Food Agric.* **1965**, *16*, 341–355. [CrossRef]
85. Knuckles, B.E.; Bickoff, E.M.; Kohler, G.O. PRO-XAN Process. Methods for Increasing Protein Recovery from Alfalfa. *J. Agric. Food Chem.* **1972**, *20*, 1055–1057. [CrossRef]
86. Morrison, J.E.; Pirie, N.W. The Large-Scale Production of Protein from Leaf Extracts. *J. Sci. Food Agric.* **1961**, *12*, 1–5. [CrossRef]
87. Edwards, R.H.; De Fremery, D.; Mackey, B.E.; Kohler, G.O. Factors Affecting Juice Extraction and Yield of Leaf Protein Concentrate from Ground Alfalfa. *Trans. ASAE* **1978**, *21*, 55–59. [CrossRef]
88. King, C.; McEniry, J.; O’Kiely, P.; Richardson, M. The Effects of Hydrothermal Conditioning, Detergent and Mechanical Pressing on the Isolation of the Fibre-Rich Press-Cake Fraction from a Range of Grass Silages. *Biomass Bioenergy* **2012**, *42*, 179–188. [CrossRef]
89. Sari, Y.W.; Mulder, W.J.; Sanders, J.P.M.; Bruins, M.E. Towards Plant Protein Refinery: Review on Protein Extraction Using Alkali and Potential Enzymatic Assistance. *Biotechnol. J.* **2015**, *10*, 1138–1157. [CrossRef]
90. Sari, Y.W.; Syafitri, U.; Sanders, J.P.M.; Bruins, M.E. How Biomass Composition Determines Protein Extractability. *Ind. Crops Prod.* **2015**, *70*, 125–133. [CrossRef]
91. Hanna, M.A.; Ogden, R.L. Expression of Alfalfa Juice. Available online: <https://pubs.acs.org/doi/pdf/10.1021/jf60232a070> (accessed on 20 December 2022).
92. Arulvasu, C.; Shakthi, S.K.S.; Babu, G.; Radhakrishnan, N. Purification and Identification of Bioactive Protein from Leaves of *Datura innoxia* P. Mil. *Biomed. Prev. Nutr.* **2014**, *4*, 143–149. [CrossRef]
93. Liu, Q.H.; Zhang, T.W.; Tian-Cai, L.I. Review on Extraction, Purification and Application of Leaf Protein. *Guizhou Agric. Sci.* **2011**, *32*, 468–471.
94. Bals, B.; Dale, B.E. Economic Comparison of Multiple Techniques for Recovering Leaf Protein in Biomass Processing. *Biotechnol. Bioeng.* **2011**, *108*, 530–537. [CrossRef]
95. Santamaría-Fernández, M.; Lübeck, M. Production of Leaf Protein Concentrates in Green Biorefineries as Alternative Feed for Monogastric Animals. *Anim. Feed. Sci. Technol.* **2020**, *268*, 114605. [CrossRef]

96. Martin, A.H.; Nieuwland, M.; De Jong, G.A.H. Characterization of Heat-Set Gels from RuBisCO in Comparison to Those from Other Proteins. *J. Agric. Food Chem.* **2014**, *62*, 10783–10791. [CrossRef] [PubMed]
97. Lamsal, B.P.; Koegel, R.G.; Gunasekaran, S. Some Physicochemical and Functional Properties of Alfalfa Soluble Leaf Proteins. *LWT-Food Sci. Technol.* **2007**, *40*, 1520–1526. [CrossRef]
98. Fári, M.G.; Domokos-Szabolcsy, É. Method for Producing Plant Protein Coagulum. Hungarian Patent WO/2019/150144, 8 August 2019.
99. Fári, M.G.; Domokos-Szabolcsy, É. Növényifehéreje-Koagulum Előállítására Szolgáló Eljárás. Hungarian Patent P1800041/40, 2018.
100. Betschart, A.; Kinsella, J.E. Extractability and Solubility of Leaf Protein. *J. Agric. Food Chem.* **1973**, *21*, 60–65. [CrossRef] [PubMed]
101. Coldebella, P.F.; Gomes, S.D.; Evarini, J.A.; Cereda, M.P.; Coelho, S.R.M.; Coldebella, A. Evaluation of Protein Extraction Methods to Obtain Protein Concentrate from Cassava Leaf. *Eng. Agric.* **2013**, *33*, 1223–1233. [CrossRef]
102. Horigome, T.; Cho, Y.S.; Ohshima, M. A Study on the Hypocholesterolemic Activity of Various Leaf Proteins of Italian Ryegrass (*Lolium multiflorum* Lam.) in the Rat. *Ital. J. Food Sci.* **1990**, *2*, 227–233.
103. Santamaria-Fernandez, M. Protein Recovery and Biogas Production From Organically Grown Biomass: A Green Biorefinery Concept. Master's Thesis, Aarhus University, Aarhus, Denmark, 2015.
104. Knuckles, B.E.; Edwards, R.H.; Kohler, G.O.; Whitney, L.F. Flocculants in the Separation of Green and Soluble White Protein Fractions from Alfalfa. *J. Agric. Food Chem.* **1980**, *28*, 32–36. [CrossRef]
105. Baraniak, B. The Effect of Flocculant Applied in the Process of Fractionating Alfalfa Juice on the Chemical Composition of the Obtained Protein Concentrates. *Anim. Feed. Sci. Technol.* **1990**, *31*, 305–311. [CrossRef]
106. Bray, W.J.; Humphries, C. Preparation of White Leaf Protein Concentrate Using a Polyanionic Flocculant. *J. Sci. Food Agric.* **1979**, *30*, 171–176. [CrossRef]
107. Zosel, A. Der Schubmodul von Hochpolymeren als Funktion von Druck und Temperatur. *Kolloid-Z. U. Z. Polymere* **1964**, *199*, 113–125. [CrossRef]
108. Weder, J.K.P. Effect of Supercritical Carbon Dioxide on Proteins. *Z. Lebensm. Unters. Forch.* **1980**, *171*, 95–100. [CrossRef]
109. Thiering, R.; Hofland, G.; Foster, N.; Witkamp, G.J.; Van De Wielen, L. Carbon Dioxide Induced Soybean Protein Precipitation: Protein Fractionation, Particle Aggregation, and Continuous Operation. *Biotechnol. Prog.* **2001**, *17*, 513–521. [CrossRef]
110. Thiering, R.; Hofland, G.; Foster, N.; Witkamp, G.J.; Van de Wielen, L. Fractionation of Soybean Proteins with Pressurized Carbon Dioxide as a Volatile Electrolyte. *Biotechnol. Bioeng.* **2001**, *73*, 1–11. [CrossRef]
111. Tomasula, P.M.; Yee, W.C. Enriched Fractions of Alpha-Lactalbumin ( $\alpha$ -LA) and Beta-Lactoglobulin ( $\beta$ -LG) from Whey Protein Concentrate Using Carbon Dioxide. Functional Properties in Aqueous Solution 1. *J. Food Process. Preserv.* **2001**, *25*, 267–282. [CrossRef]
112. McHugh, M.A.; Krukonis, V.J. *Supercritical Fluid Extraction*, 2nd ed.; Butterworth-Heinemann: Boston, MA, USA, 1993; ISBN 978-0-08-051817-6.
113. Zhang, J.; Burrows, S.; Gleason, C.; Matthews, M.A.; Drews, M.J.; LaBerge, M.; An, Y.H. Sterilizing *Bacillus Pumilus* Spores Using Supercritical Carbon Dioxide. *J. Microbiol. Methods* **2006**, *66*, 479–485. [CrossRef]
114. Kromus, S.; Wachter, B.; Koschuh, W.; Mandl, M.; Krotscheck, C.; Narodoslawsky, M. The Green Biorefinery Austria—Development of an Integrated System for Green Biomass Utilization. *Chem. Biochem. Eng. Q.* **2004**, *18*, 8–12.
115. Hernández, M.T.; Centeno, C.; Martínez, C.; Hernández, A. Effect of Solvent Extraction on the Nitrogen Compounds in Alfalfa Protein Concentrates. *J. Agric. Food Chem.* **1995**, *43*, 3065–3069. [CrossRef]
116. Karlsson, J.O.; Parodi, A.; Van Zanten, H.H.E.; Hansson, P.-A.; Rööös, E. Halting European Union Soybean Feed Imports Favours Ruminants over Pigs and Poultry. *Nat. Food* **2020**, *2*, 38–46. [CrossRef]
117. Nynäs, A.-L. White Proteins from Green Leaves in Food Applications. In *Introductory Paper at the Faculty of Landscape Architecture, Horticulture and Crop Production Science*; Department of Plant Breeding, Swedish University of Agricultural Sciences: Uppsala, Sweden, 2018.
118. Julier, B.; Gastal, F.; Louarn, G.; Badenhausser, I.; Annicchiarico, P.; Crocq, G.; Le Chatelier, D.; Guillemot, E.; Emile, J.C. Lucerne (Alfalfa) in European Cropping Systems. In *Legumes in Cropping Systems*; Murphy-Bokern, D., Stoddard, F.L., Watson, C.A., Eds.; CABI: Oxfordshire, UK, 2017; pp. 168–192. ISBN 978-1-78064-498-1.
119. Kamm, B.; Schönicke, P.; Hille, C. Green Biorefinery—Industrial Implementation. *Food Chem.* **2016**, *197*, 1341–1345. [CrossRef]
120. Database-Eurostat. Available online: <https://ec.europa.eu/eurostat/web/main/data/database> (accessed on 20 December 2022).
121. Bonou, A.; Colley, T.A.; Hauschild, M.Z.; Olsen, S.I.; Birkved, M. Life Cycle Assessment of Danish Pork Exports Using Different Cooling Technologies and Comparison of Upstream Supply Chain Efficiencies between Denmark, China and Australia. *J. Clean. Prod.* **2020**, *244*, 118816. [CrossRef]
122. Corona, A.; Ambye-Jensen, M.; Vega, G.C.; Hauschild, M.Z.; Birkved, M. Techno-Environmental Assessment of the Green Biorefinery Concept: Combining Process Simulation and Life Cycle Assessment at an Early Design Stage. *Sci. Total Environ.* **2018**, *635*, 100–111. [CrossRef] [PubMed]
123. Termansen, M.; Gylling, M.; Jørgensen, U.; Hermansen, J.; Knudsen, M.T.; Adamsen, A.P.S.; Ambye-Jensen, M.; Vestby, M.; Jensen, S.K.; Andersen, H.E.; et al. *Green Biomass—Protein Production through Biorefining*; DCA-Danish Centre for Food and Agriculture: Tjele, Denmark, 2017.
124. Fernandez, M.S. A Novel Green Biorefinery Concept: Protein Refining by Lactic Acid Fermentation and Biogas Production from Green Biomass. Ph.D. Thesis, Aalborg University, Aalborg, Denmark, 2018; pp. 1–185.

125. DCA. Green Protein. Available online: <https://dca.au.dk/en/knowledge-sharing/bioeconomy-and-biobased-production/biobase/green-protein> (accessed on 20 December 2022).
126. Kamm, B.; Hille, C.; Schönicke, P.; Dautzenberg, G. Green Biorefinery Demonstration Plant in Havelland (Germany). *Biofuels Bioprod. Biorefining* **2010**, *4*, 253–262. [CrossRef]
127. Kamm, B. Introduction of Biomass and Biorefineries. In *The Role of Green Chemistry in Biomass Processing and Conversion*; Wiley & Sons, Inc.: Hoboken, NJ, USA, 2013; pp. 1–26. ISBN 978-0-470-64410-2.
128. Carton, O.; Maguire, M.F.; Craig, J. The Nutritive Value of Preserved Grass Juice for Growing Pigs. *Ir. J. Agric. Res.* **1983**, *22*, 95–104.
129. Holló, J.; Kralovánszky, U.P. Biotechnology in Hungary. *Adv. Biochem. Eng. Biotechnol.* **2000**, *69*, 151–173. [CrossRef]
130. Bódi, L.; Fári, M.G. Eljárás Magasabb Beltartalmi Értékű Lucernaszárítmányokelőállítására. Hungarian Patent Application No. P0,203,889, 11 November 2002.
131. Päivärinta, E.; Itkonen, S.; Pellinen, T.; Lehtovirta, M.; Erkkola, M.; Pajari, A.-M. Replacing Animal-Based Proteins with Plant-Based Proteins Changes the Composition of a Whole Nordic Diet—A Randomised Clinical Trial in Healthy Finnish Adults. *Nutrients* **2020**, *12*, 943. [CrossRef]
132. Zanin. APEF, Association Pour La Promotion Des Extraits Foliaires En Nutrition | Feedipedia. 1998. Available online: <https://www.feedipedia.org/node/18347> (accessed on 20 December 2022).
133. EFSA. Opinion on the Safety of ‘Alfalfa Protein Concentrate’ as Food—EFSA. Available online: <https://www.efsa.europa.eu/en/efsajournal/pub/997> (accessed on 20 December 2022).
134. Grela, E. Production Technology, Chemical Composition and Use of Alfalfa Protein-Xanthophyll Concentrate as Dietary Supplement. *J. Food Process. Technol.* **2014**, *5*, 1000373. [CrossRef]
135. Fellet, M. A Fresh Take on Fake Meat. *ACS Cent. Sci.* **2015**, *1*, 347–349. [CrossRef]
136. Henchion, M.; Hayes, M.; Mullen, A.; Fenelon, M.; Tiwari, B. Future Protein Supply and Demand: Strategies and Factors Influencing a Sustainable Equilibrium. *Foods* **2017**, *6*, 53. [CrossRef]
137. Bessada, S.M.F.; Barreira, J.C.M.; Oliveira, M.B.P.P. Pulses and Food Security: Dietary Protein, Digestibility, Bioactive and Functional Properties. *Trends Food Sci. Technol.* **2019**, *93*, 53–68. [CrossRef]
138. Schweiggert-Weisz, U.; Eisner, P.; Bader-Mittermaier, S.; Osen, R. Food Proteins from Plants and Fungi. *Curr. Opin. Food Sci.* **2020**, *32*, 156–162. [CrossRef]
139. De Marchi, M.; Costa, A.; Pozza, M.; Goi, A.; Manuelian, C.L. Detailed Characterization of Plant-Based Burgers. *Sci. Rep.* **2021**, *11*, 2049. [CrossRef]
140. Grau, H.R.; Aide, M. Globalization and Land-Use Transitions in Latin America. *Ecol. Soc.* **2008**, *13*, 16. [CrossRef]
141. Kearney, J. Food Consumption Trends and Drivers. *Phil. Trans. R. Soc. B* **2010**, *365*, 2793–2807. [CrossRef]
142. Lourenco, C.E.; Nunes-Galbes, N.M.; Borgheresi, R.; Cezarino, L.O.; Martins, F.P.; Liboni, L.B. Psychological Barriers to Sustainable Dietary Patterns: Findings from Meat Intake Behaviour. *Sustainability* **2022**, *14*, 2199. [CrossRef]
143. Goldstein, B.; Moses, R.; Sammons, N.; Birkved, M. Potential to Curb the Environmental Burdens of American Beef Consumption Using a Novel Plant-Based Beef Substitute. *PLoS ONE* **2017**, *12*, e0189029. [CrossRef]
144. Wyckhuys, K.A.G.; Aebi, A.; Bijleveld van Lexmond, M.F.I.J.; Bojaca, C.R.; Bonmatin, J.-M.; Furlan, L.; Guerrero, J.A.; Mai, T.V.; Pham, H.V.; Sanchez-Bayo, F.; et al. Resolving the Twin Human and Environmental Health Hazards of a Plant-Based Diet. *Environ. Int.* **2020**, *144*, 106081. [CrossRef]
145. Mason-D’Croz, D.; Barnhill, A.; Bernstein, J.; Bogard, J.; Dennis, G.; Dixon, P.; Fanzo, J.; Herrero, M.; McLaren, R.; Palmer, J.; et al. Ethical and Economic Implications of the Adoption of Novel Plant-Based Beef Substitutes in the USA: A General Equilibrium Modelling Study. *Lancet Planet. Health* **2022**, *6*, e658–e669. [CrossRef]
146. Janssen, M.; Chang, B.P.I.; Hristov, H.; Pravst, I.; Profeta, A.; Millard, J. Changes in Food Consumption During the COVID-19 Pandemic: Analysis of Consumer Survey Data From the First Lockdown Period in Denmark, Germany, and Slovenia. *Front. Nutr.* **2021**, *8*, 635859. [CrossRef]
147. FAO. *The State of Food Insecurity in the World: Addressing Food Insecurity in Protracted Crises*; FAO: Rome, Italy, 2010; ISBN 978-92-5-106610-2.
148. De Schouwer, F.; Claes, L.; Vandekerckhove, A.; Verduyck, J.; De Vos, D.E. Protein-Rich Biomass Waste as a Resource for Future Biorefineries: State of the Art, Challenges, and Opportunities. *ChemSusChem* **2019**, *12*, 1272–1303. [CrossRef]
149. Ložnjak Švarc, P.; Jensen, M.B.; Langwagen, M.; Poulsen, A.; Trolle, E.; Jakobsen, J. Nutrient Content in Plant-Based Protein Products Intended for Food Composition Databases. *J. Food Compos. Anal.* **2022**, *106*, 104332. [CrossRef]
150. Song, M.; Fung, T.T.; Hu, F.B.; Willett, W.C.; Longo, V.D.; Chan, A.T.; Giovannucci, E.L. Association of Animal and Plant Protein Intake With All-Cause and Cause-Specific Mortality. *JAMA Intern. Med.* **2016**, *176*, 1453. [CrossRef] [PubMed]
151. Kim, H.; Caulfield, L.E.; Garcia-Larsen, V.; Steffen, L.M.; Coresh, J.; Rebholz, C.M. Plant-Based Diets Are Associated With a Lower Risk of Incident Cardiovascular Disease, Cardiovascular Disease Mortality, and All-Cause Mortality in a General Population of Middle-Aged Adults. *JAMA* **2019**, *8*, e012865. [CrossRef] [PubMed]
152. Arbach, C.T.; Alves, I.A.; Serafini, M.R.; Stephani, R.; Perrone, Í.T.; De Carvalho da Costa, J. Recent Patent Applications in Beverages Enriched with Plant Proteins. *NPJ Sci. Food* **2021**, *5*, 28. [CrossRef] [PubMed]
153. Sullivan, J.G.; Bliss, F.A. Expression of Enhanced Seed Protein Content in Inbred Backcross Lines of Common Bean. *J. Am. Soc. Hortic. Sci.* **1983**, *108*, 787–791. [CrossRef]

154. Hallauer, A.R. Evolution of Plant Breeding. *Crop Breed. Appl. Biotechnol.* **2011**, *11*, 197–206. [CrossRef]
155. Babu, R.; Prasanna, B.M. Molecular Breeding for Quality Protein Maize (QPM). In *Genomics of Plant Genetic Resources*; Tuberosa, R., Graner, A., Frison, E., Eds.; Springer: Dordrecht, The Netherlands, 2014; pp. 489–505. ISBN 978-94-007-7574-9.
156. Rojas Conzuelo, Z.; Robyr, R.; Kopf-Bolanz, K.A. Optimization of Protein Quality of Plant-Based Foods Through Digitalized Product Development. *Front. Nutr.* **2022**, *9*, 902565. [CrossRef]
157. Estell, M.; Hughes, J.; Grafenauer, S. Plant Protein and Plant-Based Meat Alternatives: Consumer and Nutrition Professional Attitudes and Perceptions. *Sustainability* **2021**, *13*, 1478. [CrossRef]
158. Moreira, M.N.B.; Da Veiga, C.P.; Da Veiga, C.R.P.; Reis, G.G.; Pascuci, L.M. Reducing Meat Consumption: Insights from a Bibliometric Analysis and Future Scopes. *Future Foods* **2022**, *5*, 100120. [CrossRef]
159. Sha, L.; Xiong, Y.L. Plant Protein-Based Alternatives of Reconstructed Meat: Science, Technology, and Challenges. *Trends Food Sci. Technol.* **2020**, *102*, 51–61. [CrossRef]
160. Marinangeli, C.P.F.; Mansilla, W.D.; Shoveller, A.-K. Navigating Protein Claim Regulations in North America for Foods Containing Plant-Based Proteins. *CFW* **2018**, *63*, 207–216. [CrossRef]
161. Tonsor, G.T.; Lusk, J.L.; Schroeder, T.C. Market Potential of New Plant-based Protein Alternatives: Insights from Four US Consumer Experiments. *Appl. Econ. Perspect. Policy* **2022**, aexp.13253. [CrossRef]
162. Fundação Getúlio Vargas. *Food Industry in Brazil*; FGV Projetos: Rio de Janeiro, Brazil, 2016; ISBN 978-85-64878-44-0.
163. Henchion, M.; McCarthy, M.; Resconi, V.C.; Troy, D. Meat Consumption: Trends and Quality Matters. *Meat Sci.* **2014**, *98*, 561–568. [CrossRef]
164. Xiu, S.; Shahbazi, A. Development of Green Biorefinery for Biomass Utilization: A Review. *Trends Ren. Energy* **2015**, *1*, 4–15. [CrossRef]
165. Paiva, M.J.A.; Borges, L.L.R.; Costa, N.A.; Vieira, E.N.R.; Picoli, E.A.T.; Domokos-Szabolcsy, E.; Pereira, G.C. Atividade Antioxidante Do Extrato de Pennisetum Purpureum (Schum: Poaceae) Cv. BRS Capiaçú Aos 90 Dias Pós Rebrotas. In *Simpósio Integrado de Inovação Em Tecnologia de Alimentos*; Departamento de Ciência e Tecnologia de Alimentos, UFV, Viçosa-MG: Viçosa, Brazil, 2022.
166. Lancaster, P.A.; Brooks, J.E. Cassava Leaves as Human Food. *Econ. Bot.* **1983**, *37*, 331–348. [CrossRef]
167. Gorissen, S.H.M.; Crombag, J.J.R.; Senden, J.M.G.; Waterval, W.A.H.; Bierau, J.; Verdijk, L.B.; Van Loon, L.J.C. Protein Content and Amino Acid Composition of Commercially Available Plant-Based Protein Isolates. *Amino Acids* **2018**, *50*, 1685–1695. [CrossRef]
168. Li, S.-Y.; Ng, I.-S.; Chen, P.T.; Chiang, C.-J.; Chao, Y.-P. Biorefining of Protein Waste for Production of Sustainable Fuels and Chemicals. *Biotechnol. Biofuels* **2018**, *11*, 256. [CrossRef]
169. USDE. Sustainable Aviation Fuel: Review of Technical Pathways Report. Available online: <https://www.energy.gov/eere/bioenergy/downloads/sustainable-aviation-fuel-review-technical-pathways-report> (accessed on 20 December 2022).
170. Sodamade, A.; Bolaji, O.S.; Adeboye, O.O. Proximate Analysis, Mineral Contents and Functional Properties of *Moringa oleifera* Leaf Protein Concentrate. *J. Appl. Chem.* **2013**, *4*, 47–51.
171. Moure, A.; Sineiro, J.; Domínguez, H.; Parajó, J.C. Functionality of Oilseed Protein Products: A Review. *Food Res. Int.* **2006**, *39*, 945–963. [CrossRef]
172. Sayre, R.; Beeching, J.R.; Cahoon, E.B.; Egesi, C.; Fauquet, C.; Fellman, J.; Fregene, M.; Gruissem, W.; Mallowa, S.; Manary, M.; et al. The BioCassava Plus Program: Biofortification of Cassava for Sub-Saharan Africa. *Annu. Rev. Plant Biol.* **2011**, *62*, 251–272. [CrossRef]
173. Bokanga, M. Processing OF Cassava Leaves for Human Consumption. *Acta Hortic.* **1994**, *375*, 203–208. [CrossRef]
174. Tewe Olumide, O. The Global Cassava Development Strategy and Implementation Plan. Available online: <https://www.fao.org/3/y0169e/y0169e00.htm> (accessed on 21 December 2022).
175. Agbede, J.; Aletor, V. Comparative Evaluation of Weaning Foods from Glyricidia and Leucaena Leaf Protein Concentrates and Some Commercial Brands in Nigeria. *J. Sci. Food Agric.* **2004**, *84*, 21–30. [CrossRef]
176. Agbede, J.O.; Aletor, V.A. Chemical Characterization and Protein Quality Evaluation of Leaf Protein Concentrates from *Glyricidia sepium* and *Leucaena leucocephala*. *Int. J. Food Sci. Technol.* **2004**, *39*, 253–261. [CrossRef]

**Disclaimer/Publisher’s Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.