

```
#####
```

```
#R script to analyze data
```

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```

```
###packages required
```

```
library(ggplot2)
```

```
library(multcomp)
```

```
library(openxlsx)
```

```
library(reshape2)
```

```
library(dplyr)
```

```
library(scales)
```

```
library(ggh4x)
```

```
###set working directory
```

```
setwd("C:\\Users\\charles.chappuis\\OneDrive - HESSO\\Documents\\Ra&D\\Lefort\\conf_publi")
```

```
#####
```

```
#import data sets
```

```
#####
```

```
#Olfactometry
```

```
olf <- read.xlsx("Table_S1.xlsx",sheet=1)
```

```
str(olf)
```

```
#Analytical chemistry
```

```
ana <- read.xlsx("Table_S1.xlsx",sheet=2)
```

```
str(ana)
```

```
#####
```

```
#Analysis of olfactometry data
```

```
#####
```

```
###Shorten variables and compounds names
```

```
colnames(olf) <- c("comp","mass","repl","cont","treat","main","tot")
```

```
olf$comp <- ifelse(olf$comp=="methylisonicotinate","meth",ifelse(olf$comp=="p-anisaldehyde","panis",ifelse(olf$comp=="pheromone","phero","verb")))
```

```
###add log of mass and number of activated insects and percent of activated insects choosing treated arm (attraction)
```

```
olf$act <- olf$cont+olf$treat
```

```
olf$per <- 100*olf$treat/olf$act
```

```
olf$massL <- log10(olf$mass)
```

```
olf$cont+olf$treat
```

```
###Graph attraction as a function of compounds and quantity (mass)
```

```
mySum <- olf %>%
```

```
  group_by(comp,massL) %>%
```

```
    summarise(
```

```
      n=n(),
```

```
      mean=mean(per),
```

```
      sd=sd(per)
```

```
    )
```

```
mySum$stars <- c("","","***","","","***","","**","***","","","")# different from 50% probability, see below
```

```
str(mySum)
```

```
#need statsitcal analysis below to fully graph with annotate function
```

```
p <- ggplot(mySum, aes(x=massL, y=mean, group=comp, color=comp)) +
```

```
  geom_line() +
```

```
  geom_point(size=2)+
```

```
  geom_segment(x=-0.2,y=50,xend=2.2,yend=50,linetype="dashed", color="black")+
```

```
  geom_errorbar(aes(ymin=mean-sd, ymax=mean+sd), width=0.1)+
```

```
  ylim(12.5,87.5)+
```

```
  #xlim(-0.2,2.2)+
```

```

labs(x=expression(paste("Mass loaded [",mu,"g]")), y = "Attraction [%]")+

  scale_color_manual(values=c("blue","red","black","green"),name="",breaks=c("meth", "panis",
"phero","verb"),labels = c("methyl isonicotinate", "p-anisaldehyde", "pheromone","verbenone"))+

  annotate("text",size=4,hjust=0,x=mySum$massL+0.05,y=mySum$mean+mySum$sd,label=my
Sum$stars,color=rep(c("blue","red","black","green"),each=3))+

  annotation_logticks(sides="b")

p+scale_x_continuous(breaks=c(0,log10(5),1,log10(50),2),labels=c(1,5,10,50,100),minor_breaks = c())

#ggsave("olf_mass_N.jpeg",width=1700,height=1100,device="jpeg",units="px",dpi="retina")

```

###Statistical analysis using GLM with binomial distribution and logit link function.

```

str(olf)

Tolf <- data.frame(prod=c(),repl=c(),ma=c(), att=c())

i="meth"

for(i in levels(as.factor(olf$comp))){

  olfi <- olf[olf$comp==i,]

  suA <- c(rep(1,sum(olfi$treat[olfi$mass==1])),rep(0,sum(olfi$cont[olfi$mass==1])))

  replA <-
  c(rep(1,olfi$treat[olfi$mass==1&olfi$repl==1]),rep(2,olfi$treat[olfi$mass==1&olfi$repl==2]),rep(3,olfi
$treat[olfi$mass==1&olfi$repl==3]),rep(1,olfi$cont[olfi$mass==1&olfi$repl==1]),rep(2,olfi$cont[olfi$
mass==1&olfi$repl==2]),rep(3,olfi$cont[olfi$mass==1&olfi$repl==3]))

  suB <- c(rep(1,sum(olfi$treat[olfi$mass==10])),rep(0,sum(olfi$cont[olfi$mass==10])))

  replB <-
  c(rep(1,olfi$treat[olfi$mass==10&olfi$repl==1]),rep(2,olfi$treat[olfi$mass==10&olfi$repl==2]),rep(3,olfi
$treat[olfi$mass==10&olfi$repl==3]),rep(1,olfi$cont[olfi$mass==10&olfi$repl==1]),rep(2,olfi$cont[
olfi$mass==10&olfi$repl==2]),rep(3,olfi$cont[olfi$mass==10&olfi$repl==3]))

  suC <- c(rep(1,sum(olfi$treat[olfi$mass==100])),rep(0,sum(olfi$cont[olfi$mass==100])))

  replC <-
  c(rep(1,olfi$treat[olfi$mass==100&olfi$repl==1]),rep(2,olfi$treat[olfi$mass==100&olfi$repl==2]),rep(
3,olfi$treat[olfi$mass==100&olfi$repl==3]),rep(1,olfi$cont[olfi$mass==100&olfi$repl==1]),rep(2,olfi$
cont[olfi$mass==100&olfi$repl==2]),rep(3,olfi$cont[olfi$mass==100&olfi$repl==3]))

  prodi <- rep(i,sum(c(length(suA),length(suB),length(suC))))

  mai <- c(rep(1,length(suA)),rep(10,length(suB)),rep(100,length(suC)))

  repli <- c(replA,replB,replC)

  res <- data.frame(comp=prodi,repl=repli,ma=mai,att=c(suA,suB,suC))

```

```

Tolf <- rbind(Tolf,res)
}
str(Tolf)
Tolf$Cma <- as.factor(Tolf$ma)
Tolf$Cprod <- as.factor(Tolf$comp)
Tolf$Crepl <- as.factor(Tolf$repl)
mod <- glm(att~Cma*Cprod*Crepl,family=quasibinomial,data=Tolf)
summary(mod)

#effect of replicates
modB <- glm(att~Cma*Cprod+Crepl,family=quasibinomial,data=Tolf)
summary(modB)
anova(mod,modB,test = "Chisq")
modC <- glm(att~Cma*Cprod,family=quasibinomial,data=Tolf)
summary(modC)
anova(modB,modC,test = "Chisq") ###replicates has no significant effect, data can be pooled.

#relevel to determine the level of significance for data point
mod <- glm(att~relevel(Cma,"1")*relevel(Cprod,"verb"),family=quasibinomial,data=Tolf)
summary(mod)

mySum$stars <- c("", "", "***", "", "", "***", "", "*", "***", "", "", "")# different from 50% probability

###Activation rates
mySumA <- olf %>%
  group_by(comp,massL) %>%
    summarise(
      n=n(),
      mean=mean(act),

```

```

sd=sd(act)

)

str(mySumA)

#####

#Analysis of release rates

#####

###Shorten variables and compounds names

colnames(ana) <- c("comp","repl","mass","conc","massE","rel","comment")

ana$comp <- ifelse(ana$comp=="methylisonicotinate","meth",ifelse(ana$comp=="p-
anisaldehyde","panis",ifelse(ana$comp=="pheromone","phero","verb")))

ana <- na.omit(ana)

###add log of mass

ana$massL <- log10(ana$mass)

ana$relL <- log10(ana$rel)

ana$compF <- as.factor(ana$comp)

###Graph release rate as a function of compounds and quantity (mass)

p <- ggplot(ana, aes(x=massL, y=relL, group=comp, color=comp)) +
  geom_point(size=2)+
  labs(x=expression(paste("Mass loaded [",mu,"g]")), y = "Release rate [ng/min]")+
  scale_color_manual(values=c("blue","red","black","green"),name="",breaks=c("meth",
"panis", "phero","verb"),labels = c("methyl isonicotinate", "p-anisaldehyde",
"pheromone","verbenone"))+
  theme(legend.text=element_text(size=8))+
  #xlim(0,2)+

```

```
    annotate("text",size=3,x=c(rep(1.75,4)),y=c(2.89,2.68,2.38,1.6),label=c("a","a","b","c"),color=
c("blue","green","red","black"))+
```

```
    geom_abline(intercept=c(0.943,0.445,-
0.402,0.925),slope=c(rep(1.052,4)),color=c("blue","red","black","green"))+#parameters from linear
model below
```

```
    annotation_logticks(sides="bl")+
```

```
    scale_x_continuous(breaks=c(0,log10(5),1,log10(50),2),labels=c(1,5,10,50,100),minor_breaks
= c())+
```

```
    scale_y_continuous(breaks=c(log10(0.5),0,log10(5),1,log10(50),2,log10(500),3),labels=c(0.5,1
,5,10,50,100,500,1000),minor_breaks = c())
```

p

```
#ggsave("releaserate_mass_N.jpeg",width=1700,height=1100,device="jpeg",units="px",dpi="retina")
```

```
###statistical analysis with linear models
```

```
mod <- lm(relL~massL*compF,data=ana)
```

```
summary(mod) #no interaction terms significant, slopes are not significantly different among the
compounds
```

```
modN <- lm(relL~massL+compF,data=ana)
```

```
summary(modN)
```

```
anova(mod, modN)
```

```
modNN <- lm(relL~massL,data=ana)
```

```
summary(modNN)
```

```
anova(modN, modNN)
```

```
#relevels
```

```
modN <- lm(relL~massL+relevel(compF,"phero"),data=ana)
```

```
summary(modN)
```

```
10^c(0.939,0.925,0.445,-0.402)#retransformed data to obtain ng/min for the intercept
```

```
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```
#Analysis of release rates and behavioral data
```

```
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```

```
anaS <- ana %>%
```

```
  group_by(comp,massL) %>%
```

```
  summarise(
```

```
    n=n(),
```

```
    mean=mean(relL, na.rm=T),
```

```
    sd=sd(relL, na.rm=T)
```

```
  )
```

```
str(anaS)
```

```
10^c(-0.352,0.609,1.68)
```

```
10^c(2.09,-0.352)
```

```
10^c(-0.352,3,2.61)
```

```
anaS$attr <- mySum$mean
```

```
anaS$attrSD <- mySum$sd
```

```
anaS$stars <- mySum$stars
```

```
p <- ggplot(anaS, aes(x=mean, y=attr, group=comp, color=comp)) +
```

```
  geom_line() +
```

```
  geom_point(size=2)+
```

```
  geom_segment(x=-1,y=50,xend=3.5,yend=50,linetype="dashed", color="black")+
```

```
  geom_errorbar(aes(ymin=attr-attrSD, ymax=attr+attrSD), width=0.2)+
```

```
  ylim(12.5,87.5)+
```

```
  #xlim(-0.5,3.5)+
```

```
  labs(x=expression("Release rate [ng/min]"), y = "Attraction [%]")+
```

```
  scale_color_manual(values=c("blue","red","black","green"),name="",breaks=c("meth", "panis",  
"phero","verb"),labels = c("methyl isonicotinate", "p-anisaldehyde", "pheromone","verbenone"))+
```

```

theme(legend.text=element_text(size=8),axis.text.x = element_text(size = 6),)+
  annotate("text",size=4,hjust=0,x=anaS$mean+0.1,y=anaS$attr+anaS$attrSD,label=anaS$stars,
color=rep(c("blue","red","black","green"),each=3))+
  annotation_logticks(sides="b")+
  scale_x_continuous(breaks=c(log10(0.5),0,log10(5),1,log10(50),2,log10(500),3),labels=c(0.5,1
,5,10,50,100,500,1000),minor_breaks = c())

```

p

```

ggsave("olfactometer_releaserate_N.jpeg",width=1700,height=1100,device="jpeg",units="px",dpi="r
etina")

```

```

anaS$Rmean <- 10^anaS$mean
anaS$gPhase <- anaS$Rmean/0.15
anaS$test <- anaS$Rmean/122

```

```
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#P-value that a proportion is significantly different from 50% with a pool of 75 thrips

```
#####
```

```
Nthrips <- 75
```

```
i=1
```

```
pval <- data.frame(cat=c(),val=c())
```

```
for(i in c(1:(Nthrips-1))){
```

```
  veca <- c(rep(1,i),rep(0,Nthrips-i))
```

```
  cata <- rep(i,Nthrips)
```

```
  vecb <- c(rep(1,i+1),rep(0,Nthrips-(i+1)))
```

```
  catb <- rep(i+1,Nthrips)
```

```
  ro <- data.frame(cat=c(cata,catb),resp=c(veca,vecb))
```

```
  ro$cat <- as.factor(ro$cat)
```

```
  mod <- glm(resp~cat, family=quasibinomial, data=ro)
```



```

sumM <- summary(mod)

pv <- coef(sumM)[7]

pvali <- data.frame(cat=i,val=pv)

pval <- rbind(pval,pvali)
}

pval$prop <- 100*pval$cat/Nthrips

p <- ggplot(pval, aes(x=prop, y=val)) +
  geom_line() +
  geom_point(size=1)+
  geom_segment(x=0,y=0.05,xend=100,yend=0.05,linetype="dashed",color="black")+
  labs(x="Attraction [%]", y="Pvalue")+
  scale_x_continuous(breaks = seq(0,100,12.5),labels = seq(0,100,12.5))

p

ggsave("stat_problem.jpeg",width=1400,height=1000,device="jpeg",units="px",dpi="retina")

```