

Table S1. Primer sequences of *L. monocytogenes* and *A. flavus* genes and respective amplicons sizes used for the gene expression assay

Genes	Primers	Sequence	Concentration (μM)	Amplicon size (bp)	PCR efficiency	Reference
<i>L. monocytogenes</i>						
Reference genes						
<i>IGS</i>	IGS_f	GGCCTATAGCTCAGCTGGTTA	1.2	135	2.03	[1]
	IGS_r	GCTGAGCTAAGGCCCGTAAA	1.2			
<i>rpob</i>	Rpob_f	CCGCGATGCGAAAACAAT	0.9	69	2.04	[2]
	Rpob_r	CCWACAGAGATACGGTTATCRAATGC	0.9			
<i>16S</i>	16S_f	GATGCATAGCCGACCTGAGA	0.9	114	2.05	[3]
	16S_r	CTCCGTCAGACTTTCGTCCA	0.9			
Virulence- associated						
<i>prfA</i>	prfA_f	CTATTTGCGGTCAACTTTTAATCCT	0.9	100	2.09	[2]
	prfA_r	CCTAACTCCTGCATTGTTAAATATCC	0.9			
<i>sigB</i>	sigB_f	CCAAGAAAATGGCGATCAAGAC	1.2	166	2.13	[1]
	sigB_r	CGTTGCATCATATCTTCTAATAGCT	1.2			
<i>plcA</i>	plcA_f	CTAGAAGCAGGAATACGGTACA	1.2	115	1.94	[1]
	plcA_r	ATTGAGTAATCGTTTCTAAT	1.2			
<i>plcB</i>	plcB_f	CAGGCTACCACTGTGCATATGAA	0.9	72	2.00	[2]
	plcB_r	CCATGTCTTCYGTGCTTGATAATTG	0.9			
<i>hly</i>	hly_f	TACATTAGTGGAAGATGG	1.2	153	1.98	[2]
	hly_r	ACATTCAAGCTATTATTTACA	1.2			
<i>inlA</i>	inlA_f	AATGCTCAGGCAGCTACAMTTACA	0.9	114	2.12	[2]
	inlA_r	CGTGTCTGTTACRTTCGTTTTTCC	0.9			
<i>inlB</i>	inlB_f	AAGCAMGATTTTCATGGGAGAGT	0.9	78	2.04	[2]
	inlB_r	TTACCGTTCATCAACATCATAACTT	0.9			
<i>inlC</i>	inlC_f	ACTGGTCAGAAATGTGTGAATGA	0.9	80	2.06	[4]
	inlC_r	CCATCTGGGTCTTTGACAGT	0.9			
<i>inlJ</i>	inlJ_f	TGCGTAAATGCTCACATCCAAG	0.9	81	2.03	[4]
	inlJ_r	TTGCCCTTCAGCATCCAAGT	0.9			
Fatty acid biosynthesis						
<i>murE</i>	murE_f	GCCACAACCAACACGACAA	0.9	85	1.97	[5]
	murE_r	TCATACTCCAGACGGCTTGC	0.9			
<i>accA</i>	accA_f	GCGGTCAAAGTGAAGCCATT	0.9	94	1.99	[5]
	accA_r	CCACTTCCACCTTACCGAT	0.9			
<i>acpP</i>	acpP_f	TGAAGACGAGTTCGGAGTTGA	0.9	91	2.05	[5]
	acpP_r	TGCGTTCGCCTCTATGTACT	0.9			
Peptidoglycan biosynthesis						
<i>fapR</i>	fapR_f	CGCCGTCATCCCAATGAAA	0.9	88	2.02	[5]
	fapR_r	TGCGATGATGCGTTCCTT	0.9			
<i>A. flavus</i>						
Reference genes						
<i>18S</i>	18S_f	GCAAATTACCCAATCCCGACAC	0.2	32	1.97	[6]
	18S_r	GAATTACCGCGGCTGCTG	0.2			
<i>cal</i>	cal_f	CTTCCCGAATTCCTTACC	0.2	89	2.02	[7]
	cal_r	TCACGGATCATCTCATCGAC	0.2			
<i>Tub-β</i>	tubβ_f	CGCATGAACGTCTACTTCAACGAG	0.2	128	1.94	[8]
	tubβ_r	AGTTGTTACCAGCAGCGGACT	0.2			
Biosynthesis						
<i>aflM</i>	aflM_f	AAGTTAATGGCGGAGACG	0.2	172	2.02	[9]
	aflM_r	TCTACCTGCTCATCGGTGA	0.2			
<i>aflD</i>	aflD_f	GCGCAAGTTCACCTTTGAGA	0.2	42	1.99	[10]
	aflD_r	CCTTGGTCGCCATATCAGT	0.2			
<i>aflP</i>	aflP_f	AGCCCCGAAGACCATAAAC	0.2	170	1.91	[9]
	aflP_r	CCGAATGTCATGCTCCATC	0.2			
Regulatory						
<i>aflR</i>	aflR_f	CGGCACAGCTTGTCTGAGT	0.2	88	1.92	[10]
	aflR_r	GCATCGTCTCCACCTTCTTG	0.2			
<i>aflS</i>	aflR_f	CTGGCAAACCTGGGAATGG	0.2	63	2.05	[10]
	aflR_r	CACGAGGAAACGGAGTGATG	0.2			

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