

Supporting information of

Root bacteria recruited by *Phragmites australis* in Constructed Wetlands have the potential to enhance azo-dye phytodepuration

Supplementary Table S1. Distribution of cultivable bacteria isolated from *P. australis* roots and rhizosphere according to the phylogenetic classification at the phylum, genus and species level.

Phylum	Genus	E (80)	R (72)	Species	E (80)	R (72)					
Actinobacteria	<i>Arthrobacter</i>		1	<i>A. ureafaciens</i>		1					
				<i>M. kitamiense</i>		2					
	<i>Streptomyces</i>			5	<i>M. oxydans</i>		1				
					<i>M. maritypicum</i>		1				
					<i>S. pluricolorescens</i>		2				
					<i>S. althioticus</i>		1				
					<i>S. acidiscabies</i>		1				
Bacteroidetes	<i>Flavobacterium</i>	3	9	<i>S. caeruleatus</i>		1					
				<i>F. gyeonganense</i>	2	3					
				<i>F. johnsoniae</i>	1	5					
				<i>F. oncorhynchi</i>		1					
Firmicutes	<i>Fluviicola</i>		1	<i>F. taffensis</i>		1					
				<i>B. aryabhatai</i>	2						
	<i>Bacillus</i>		38	24	<i>B. horneckiae</i>		1				
					<i>B. litoralis</i>		1				
					<i>B. marisflavi</i>		1				
					<i>B. megaterium</i>		1				
					<i>B. niacini</i>		1				
					<i>B. oceanisediminis</i>		2				
					<i>B. pumilus</i>	34					
					<i>B. thuringiensis</i>	2	17				
					<i>Lysinibacillus</i>			8	<i>L. fusiformis</i>		6
									<i>L. parviboronicapiens</i>		1
									<i>L. varians</i>		1
					<i>Viridibacillus</i>			9	<i>V. arenosi</i>		9
Proteobacteria	<i>Caulobacter</i>	1		<i>C. segnis</i>	1						
				<i>C. neteri</i>		1					
	<i>Enterobacter</i>			4	<i>E. cancerogenus</i>		2				
					<i>E. cloacae</i>		1				
					<i>E. ludwigii</i>		1				
	<i>Ochrobactrum</i>			1	<i>O. anthropi</i>		1				
	<i>Pseudomonas</i>		24	3	<i>P. aeruginosa</i>	2					
					<i>P. brassicacearum</i>	5					
					<i>P. fluorescens</i>		1				
					<i>P. plecoglossicida</i>		1				
<i>P. stutzeri</i>						1					
<i>P. thivervalensis</i>					17						
<i>Rheinheimera</i>							1	<i>R. chironomi</i>		1	
<i>Rhizobium</i>	6			<i>A. tumefaciens</i>	6						
<i>Stenotrophomonas</i>		8	1	<i>S. rhizophila</i>		1					
				<i>S. chelatiphaga</i>	8						

Supplementary Table S2. Percentage distribution of plant growth promoting activities according to the bacterial genera isolated from the root system of *P. australis*. IAA = 3-indoleacetic acid production, ACC-d = ACC deaminase activity, Protease = protease production, EPS = exopolysaccharides production, SWIMMING =swimming lifestyle, SWARMING = swarming lifestyle.

Genus	n° strain	IAA	ACC-d	Protease	EPS	Swimming	Swarming
<i>Arthrobacter</i>	1	0	100	100	0	0	0
<i>Microbacterium</i>	4	50	100	25	25	0	0
<i>Streptomyces</i>	4	50	50	0	0	0	0
<i>Flavobacterium</i>	10	20	50	20	20	0	0
<i>Fluviicola</i>	1	0	0	0	0	100	100
<i>Bacillus</i>	12	83	58	75	8	17	33
<i>Lysinibacillus</i>	7	100	57	57	0	86	86
<i>Viridibacillus</i>	2	100	100	0	0	100	100
<i>Caulobacter</i>	1	100	100	0	0	0	0
<i>Cedecea</i>	1	100	100	0	0	0	0
<i>Enterobacter</i>	4	100	100	0	0	75	25
<i>Ochrobactrum</i>	1	100	100	0	0	0	0
<i>Pseudomonas</i>	6	100	83	50	50	17	0
<i>Rheinheimera</i>	1	100	100	100	0	0	0
<i>Stenotrophomonas</i>	3	100	33	100	33	0	0

Supplementary Table S3. Metal-tolerance phenotypes. The code of the isolates is simplified and includes only the fraction of origin (R = rhizosphere; E = endosphere) and the progressive number. The percentage in the last column is referred to the strains belonging to each detected phenotype. '+': refers to the strain tolerance to the metal at the indicated concentration.

Phenotype	Code of the isolates	Metal Tolerance									% of isolates
		ZnCl ₂ (mM)			NiCl ₂ (mM)			CdCl ₂ (mM)			
		0.5	1	2	0.5	1	2	0.05	0.5	1	
1	R8, R25, R31, R32, R34, R67	+	+	+	+	+	+	+	+	+	12
2	R17, R20, R78	+	+	+	+	+		+	+	+	6
3	R3, R6, R50	+	+	+	+	+	+	+	+		6
4	R15	+	+		+	+	+	+	+		2
5	R22, R68	+	+	+	+	+		+	+		4
6	E2, E6, E8, E14, E16, E21, E28, E33, E73, R4, R28, R65, R71, R75	+	+	+	+	+	+	+			28
7	E13, R76, R79	+	+	+	+	+		+			6
8	R26, R69	+	+		+	+	+	+			4
9	R2	+	+		+	+		+			2
10	R16, R23, R38, R39, R47, R49, R52, R64	+	+	+	+	+	+				16
11	E15, R10, R40	+	+	+	+	+					6
12	R33	+	+		+	+	+				2
13	R7	+			+	+	+	+			2
14	E27	+	+		+	+					2
15	R77	+						+			2

Supplementary Table S4 (next page). Characterization of the bacterial collection.

In vitro plant growth promotion (PGP) tests: IAA = IAA production; ACC-d = ACC deaminase activity; Prot. = protease production. *In vivo* PGP test under hydroponic condition with *L. esculentum* plants: % germin. = germination percentage; root-l = root length; shoot-l = shoot length; 2ary roots = production of secondary roots; SVI = seedling vigor index; root dw = root dry weight. Ability to bind the root apparatus: Swimming = swimming lifestyle; Swarming = swarming lifestyle; EPS exopolysaccharides release. BPA tolerance. *In vitro* decolorization ability of RB5 = Reactive Black 5.

The + indicates that the bacterium is positive to the test. The stars indicate statistically significant differences (* $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$). R indicates the resistance to the antibiotic.

Strain	Closest Described Relative	IAA	ACC-d	Prot.	Hydroponic Experiment					Swimming	Swarming	EPS	BPA	RB5
					% Germin.	Root-l	Shoot-l	2ary Roots	SVI					
CWMP-8E2	<i>Pseudomonas thivervalensis</i>	+	+	+								+	+	
CWMP-8E6	<i>Flavobacterium johnsoniae</i>	+	+	+								+	+	
CWMP-8E8	<i>Stenotrophomonas chelatiphaga</i>	+	+	+								+	+	
CWMP-8E13	<i>Bacillus thuringiensis</i>			+									+	
CWMP-8E14	<i>Flavobacterium gyeonganense</i>		+		*			**					+	
CWMP-8E15	<i>Bacillus aryabhatai</i>	+	+	+										
CWMP-8E16	<i>Flavobacterium gyeonganense</i>												+	
CWMP-8E21	<i>Pseudomonas thivervalensis</i>	+	+	+				**				+	+	
CWMP-8E27	<i>Bacillus aryabhatai</i>	+	+	+									+	
CWMP-8E28	<i>Pseudomonas brassicacearum</i>	+	+	+								+	+	
CWMP-8E33	<i>Bacillus pumilus</i>	+		+								+		
CWMP-8E42	<i>Caulobacter segnis</i>	+	+					**					+	
CWMP-8E73	<i>Stenotrophomonas chelatiphaga</i>	+		+										
CWMP-8R1	<i>Bacillus niacini</i>	+											+	
CWMP-8R2	<i>Microbacterium kitamiense</i>		+					**	*	*			+	1%
CWMP-8R3	<i>Bacillus thuringiensis</i>	+	+	+							+	+	+	24%
CWMP-8R4	<i>Flavobacterium oncorhynchi</i>		+										+	17%
CWMP-8R6	<i>Flavobacterium johnsoniae</i>		+										+	7%
CWMP-8R7	<i>Bacillus megaterium</i>	+	+	+	**	*	***		***	*			+	14%
CWMP-8R8	<i>Enterobacter cancerogenus</i>	+	+			*	***	**	**	*	+	+	+	32%
CWMP-8R9	<i>Streptomyces althioticus</i>							***					+	

CWMP-8R10	<i>Viridibacillus arenosi</i>	+	+		*	**	***		***	**	+	+		+	9%
CWMP-8R12	<i>Streptomyces acidiscabies</i>		+			***	***		***					+	
CWMP-8R15	<i>Bacillus horneckiae</i>		+	+			**		*	*	+	+			29%
CWMP-8R16	<i>Lysinibacillus fusiformis</i>	+	+				**				+	+		+	11%
CWMP-8R17	<i>Enterobacter cloacae</i>	+	+								+	+		+	37%
CWMP-8R19	<i>Bacillus oceanisediminis</i>	+	+	+			*	*						+	
CWMP-8R20	<i>Cedecea neteri</i>	+	+				*	*	*					+	10%
CWMP-8R22	<i>Enterobacter cancerogenus</i>	+	+											+	35%
CWMP-8R23	<i>Lysinibacillus fusiformis</i>	+		+		***	***	**	*		+	+		+	18%
CWMP-8R26	<i>Flavobacterium gyeonganense</i>	+	+											+	13%
CWMP-8R28	<i>Flavobacterium johnsoniae</i>				*	*								+	5%
CWMP-8R31	<i>Microbacterium kitamiense</i>		+				*							+	6%
CWMP-8R32	<i>Ochrobactrum anthrop</i>	+	+					*	*					+	27%
CWMP-8R33	<i>Arthrobacter ureafaciens</i>		+	+										+	50%
CWMP-8R38	<i>Lysinibacillus varians</i>	+	+	+										+	22%
CWMP-8R39	<i>Lysinibacillus parviboronicapiens</i>	+		+		**			*		+	+		+	19%
CWMP-8R40	<i>Viridibacillus arenosi</i>	+	+			**	***		**		+	+		+	7%
CWMP-8R47	<i>Fluviicola taffensis</i>						***							+	20%
CWMP-8R49	<i>Lysinibacillus fusiformis</i>	+	+	+			***		*		+	+		+	18%
CWMP-8R50	<i>Pseudomonas plecoglossicida</i>	+	+	+										+	12%
CWMP-8R52	<i>Streptomyces pluricologrescens</i>	+			*		***		***					+	
CWMP-8R57	<i>Rheinheimera chironomi</i>	+	+	+		***	***		***					+	
CWMP-8R64	<i>Lysinibacillus fusiformis</i>	+	+				***				+	+		+	74%
CWMP-8R65	<i>Flavobacterium johnsoniae</i>						***							+	22%
CWMP-8R68	<i>Bacillus marisflavi</i>	+		+		*	***		***			+		+	10%
CWMP-8R69	<i>Pseudomonas stutzeri</i>	+				*	***		**					+	30%
CWMP-8R72	<i>Bacillus litoralis</i>	+	+											+	
CWMP-8R76	<i>Flavobacterium gyeonganense</i>			+		***	***		***					+	16%
CWMP-8R77	<i>Bacillus oceanisediminis</i>	+				*	***		***					+	
CWMP-8R79	<i>Stenotrophomonas rhizophila</i>	+		+		**	***		**					+	32%
CWMP-8R80	<i>Streptomyces caeruleatus</i>	+	+				**		*					+	

Supplementary Table S5. Effect of bacterial consortium on *Juncus acutus* plants grown in soil. The stars indicate statistically significant differences in comparison to the non bacterized plants (* $0.01 \leq p < 0.05$).

Treatment	Root			Shoot			Chlorophyll (mg/g of leaf)
	Length (cm)	Fresh Weight (g)	Dry Weight (g)	Length (cm)	Fresh Weight (g)	Dry Weight (g)	
NC	22	0,78	0,24	35	3,30	0,85	0,23
MIX	23	1,37 *	0,32 *	36	4,45	1,10	0,36 *

NC: negative control

Supplementary Figure S1. Difference in root apparatus development of (a) bacterized and (b) non bacterized tomato seedlings.

