

**Table S1.** Bacterial strains, mutants and plasmids used in this study.

Strain/Plasmid	Relevant characteristics <sup>a</sup>	Reference/Source <sup>b</sup>
<b>Strain</b>		
<i>E. coli</i> TOP10	F-, mcrA, Δ(mrr-hsdRMS-crBC) Φ80 lacZΔM15 ΔlacX74 recA1 araD139 Δ(araleu)7697 galU galK rpsL (StrR) endA1 nupG	Invitrogen, Carlsbad, USA
<i>E. coli</i> ER2925	ara-14 leuB6 fhuA31 lacY1 tsx78 glnV44 galK2 galT22 mcrA dcm-6 hisG4 rfbD1 R(zgb210::Tn10)TetS endA1 rpsL136 dam13::Tn9 xylA-5 mtl-1 thi-1 mcrB1 hsdR2	NEB, Hertfordshire, UK
<i>P. savastanoi</i> pv. <i>nerii</i> ( <i>Psn23</i> )	Wild type	LPVM collection [30]
<i>Psn23_ΔiaaM</i>	<i>iaaM</i> in-frame deletion mutant of <i>Psn23</i>	[17]
<i>Psn23_ΔiaaL</i>	<i>iaaL</i> in-frame deletion mutant of <i>Psn23</i>	[17]
<i>Psn23_ΔmatE</i>	<i>matE</i> in-frame deletion mutant of <i>Psn23</i>	This study
<i>Psn23_pT3-matE</i>	Gm <sup>R</sup> , lacZ, mcs, <i>hrpA</i> _promoter+ <i>matE</i>	This study
<i>Psn23_D182A</i>	<i>matE</i> replaced mutant (Asp182Ala) of <i>Psn23</i>	This study
<i>Psn23_Y200A</i>	<i>matE</i> replaced mutant (Tyr200Ala) of <i>Psn23</i>	This study
<i>Psn23_T17035A</i>	<i>matE</i> replaced mutant (Thr170Ala, Thr173Ala and Thr175Ala) of <i>Psn23</i>	This study
<b>Plasmid</b>		
<i>pK18-ΔhrpA</i>	pK18mobsacB derivative, in-frame deletion of the <i>hrpA</i> gene (273 bp), Km <sup>R</sup>	[17]
<i>pK18-ΔmatE</i>	pK18mobsacB derivative, in-frame deletion of the <i>matE</i> gene (1113 bp), Km <sup>R</sup>	This study
<i>pK18-matE</i> (D182A)	pK18mobsacB derivative, <i>matE</i> gene replaced in amino acid position 182 (Asp → Ala), Km <sup>R</sup>	This study
<i>pK18-matE</i> (Y200A)	pK18mobsacB derivative, <i>matE</i> gene replaced in amino acid position 200 (Tyr → Ala), Km <sup>R</sup>	This study
<i>pK18-matE</i> (T17035A)	pK18mobsacB derivative, <i>matE</i> gene replaced in amino acid positions 170, 173 and 175 (Thr → Ala), Km <sup>R</sup>	This study
<i>pLPVM-T3A</i>	Gm <sup>R</sup> , lacZ, mcs, <i>hrpA</i> _promoter+ <i>gfp</i>	[33]
<i>pLPVM-matE</i>	Gm <sup>R</sup> , lacZ, mcs, <i>hrpA</i> _promoter+ <i>matE</i>	[33]

<sup>a</sup> Gm<sup>R</sup>, gentamicin resistance; Km<sup>R</sup>, kanamycin resistance. <sup>b</sup> LPVM Laboratorio di Patologia Vegetale Molecolare (University of Florence).

**Table S2.** Primers used in this study.

<b>Primer name</b>	<b>Primer sequence (5'-3')<sup>a</sup></b>
matE_PstI_For	TTT <u>CTGCAGT</u> CCAGAACACAGACATT
matE_cross_Rev	CCGGATCCACTGAAACTTGCTATGCCAAGAATCATC
matE_cross_For	AAGTTTCAGTGGATCCGGCGAGATCCATTCAATAGG
matE_EcoRI_Rev	TTTGAATTC <u>TAATCGTGT</u> TGTTTCAGAG
pT3_matE_BamHI_For	TTTGGATCCATGGTAGTTATCAA
pT3_matE_KpnI_Rev	AAAGGTACCTTATGAGTACTCCTGT
pK18_matE_PstI_For	TTTCTGCAGATGGTAGTTATCAA
pK18_matE_EcoRI_Rev	TTTGAATTC <u>TTATGAGT</u> ACTCCTGT
matE_D182A_For	GTTGCCATCGCAGCCCCGCTGCTTATTG
matE_D182A_Rev	CAATAAGCAGCGGGGCTGCGATGGCAAC
matE_Y200A_For	CGGCATCGCCGCCCTGATATCGAG
matE_Y200A_Rev	CTCGATATCAGGGCGGGCATGCCG
matE_T17035A_For	GTGGGCCCTGCTGGCGGGCGGGCGGC
matE_T17035A_Rev	GCCGCCCGCCAGCAGGGCCCCAC
hrpA_RT_For	GCAGGGTATCAACAGCGTCAAG
hrpA_RT_Rev	CCGTTCTCTCGTTCGCAAGT
hrpRS_RT_For	ACCCGCAGAGTGAAGAAC
hrpRS_RT_Rev	CGCTTGAGTGACTGTTGAATC
iaaM_RT_For	TTCACTGCCTCACGGATAGCG
iaaM_RT_Rev	CGACTGGATGGTGGTGGGAAG
iaaL_RT_For	ACCTCAGCAGCGGCGTAAAG
iaaL_RT_Rev	TCGTCCGTGTGTATGGCAGTTC
iaaH_RT_For	TGATGATGCCGATATTGTC
iaaH_RT_Rev	AAGGTGGTGATTGATGATG
matE_RT_For	CATCGCAGCCATTACG
matE_RT_Rev	AGCCTGAAGAACCTGTC

<sup>a</sup> The nucleotides underlined refer to digestion cutting sites for molecular cloning.