

## Article

# Data Mining as a Tool to Infer Chicken Carcass and Meat Cut Quality from Autochthonous Genotypes

Antonio González Ariza <sup>1,2</sup>, Francisco Javier Navas González <sup>1,3,\*</sup>, José Manuel León Jurado <sup>2</sup>,  
Ander Arando Arbulu <sup>1</sup>, Juan Vicente Delgado Bermejo <sup>1</sup> and María Esperanza Camacho Vallejo <sup>3</sup>

<sup>1</sup> Department of Genetics, Faculty of Veterinary Sciences, University of Córdoba, 14071 Córdoba, Spain

<sup>2</sup> Agropecuaria Provincial Centre, Diputación Provincial de Córdoba, 14071 Córdoba, Spain

<sup>3</sup> Institute of Agricultural Research and Training (IFAPA), Alameda del Obispo, 14004 Córdoba, Spain

\* Correspondence: franciscoj.navas@juntadeandalucia.es; Tel.: +34-651-679-262

**Simple Summary:** The present study is a meta-analysis of ninety-one research documents dealing with carcass quality characterization in autochthonous chicken genotypes. Documents were published between 2002 and 2021. Data mining methods were used to determine which variables should be considered or otherwise discarded from comprehensive carcass quality differential models to improve the study's efficiency and accuracy. Even if the impact on carcass quality of certain variables such as chicken sex, meat firmness, chewiness, L\* meat 72 h post-mortem, a\* meat 72 h post-mortem, b\* meat 72 h post-mortem, and pH 72 h post-mortem could be presumed, these should not be considered if strongly related variables are simultaneously considered too, to prevent redundancy problems. In contrast, carcass/cut weight, pH, carcass yield, slaughter age, protein, cold weight, and L\* meat must be regarded strictly due to their high potential to explain differences and correctly classify carcass cuts across chicken genotypes. The standardization of characterization methods of minority populations (with limited censuses and lacking population structure, but well-adapted to alternative systems) enhances the possibility of success of the implementation of sustainable conservation strategies through the dissemination of knowledge on local breeds and the competitivization of their distinctive products within specific market niches.

**Abstract:** The present research aims to develop a carcass quality characterization methodology for minority chicken populations. The clustering patterns described across local chicken genotypes by the meat cuts from the carcass were evaluated via a comprehensive meta-analysis of ninety-one research documents published over the last 20 years. These documents characterized the meat quality of native chicken breeds. After the evaluation of their contents, thirty-nine variables were identified. Variables were sorted into eight clusters as follows; weight-related traits, water-holding capacity, colour-related traits, histological properties, texture-related traits, pH, content of flavour-related nucleotides, and gross nutrients. Multicollinearity analyses ( $VIF \leq 5$ ) were run to discard redundancies. Chicken sex, firmness, chewiness, L\* meat 72 h post-mortem, a\* meat 72 h post-mortem, b\* meat 72 h post-mortem, and pH 72 h post-mortem were deemed redundant and discarded from the study. Data-mining chi-squared automatic interaction detection (CHAID)-based algorithms were used to develop a decision-tree-validated tool. Certain variables such as carcass/cut weight, pH, carcass yield, slaughter age, protein, cold weight, and L\* meat reported a high explanatory potential. These outcomes act as a reference guide to be followed when designing studies of carcass quality-related traits in local native breeds and market commercialization strategies.

**Keywords:** biodiversity; sustainability; local genetic resources; native breeds; chicken meat; chemical characterization; physical traits; meat cuts



**Citation:** González Ariza, A.; Navas González, F.J.; León Jurado, J.M.; Arando Arbulu, A.; Delgado Bermejo, J.V.; Camacho Vallejo, M.E. Data Mining as a Tool to Infer Chicken Carcass and Meat Cut Quality from Autochthonous Genotypes. *Animals* **2022**, *12*, 2702. <https://doi.org/10.3390/ani12192702>

Academic Editors: Stefano Paolo Marelli, Alice Cartoni Mancinelli and Avigdor Cahaner

Received: 17 August 2022

Accepted: 5 October 2022

Published: 8 October 2022

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

### 1. Introduction

Poultry production has rapidly developed worldwide [1]. Contextually, big companies are responsible for the major management of (optimized housing and feeding conditions have considerably shortened the rearing period of fast-growing chickens) and genetic improvements made to intensive farming systems in response to the current dependence on and demand for chicken meat [2].

While intensive poultry farming almost exclusively relies upon high-yielding commercial hybrid ‘only meat’ strains [3] to provide a large amount of meat for the human population on a large scale [4], indigenous chicken populations still significantly contribute to local economies, especially low-income rural livelihoods, across Asia, Africa, South America, and the South Pacific [5–7]. As a result, the replacement and hybridization of native breeds with these exotic strains, which may internationally be more commercially competitive, drastically threatens the genetic diversity of worldwide poultry populations [8].

In this regard, although the current promotion of specialized layer or meat producer genotypes on chicken farms has produced a displacement and marginalization of dual-purpose systems (supplanting native chicken breeds) [9], problems linked to meat maturity and its technological and sensory quality [10] may arise. To counteract such problems, a growing demand for poultry products from alternative production systems has brought about the opportunity to increase the importance of raising autochthonous breeds.

According to the DAD-IS (Domestic Animal Diversity Information System) FAO database [11], only 9.18% of local breeds are actually not at risk (Figure 1). Local genotypes may be the source of enhanced distinctive products, and may play a rather pivotal role in meat quality over quantity. In this regard, poultry meat quality can be understood in various ways, ranging from poultry meat nutritive value to sensory traits, among others [12].

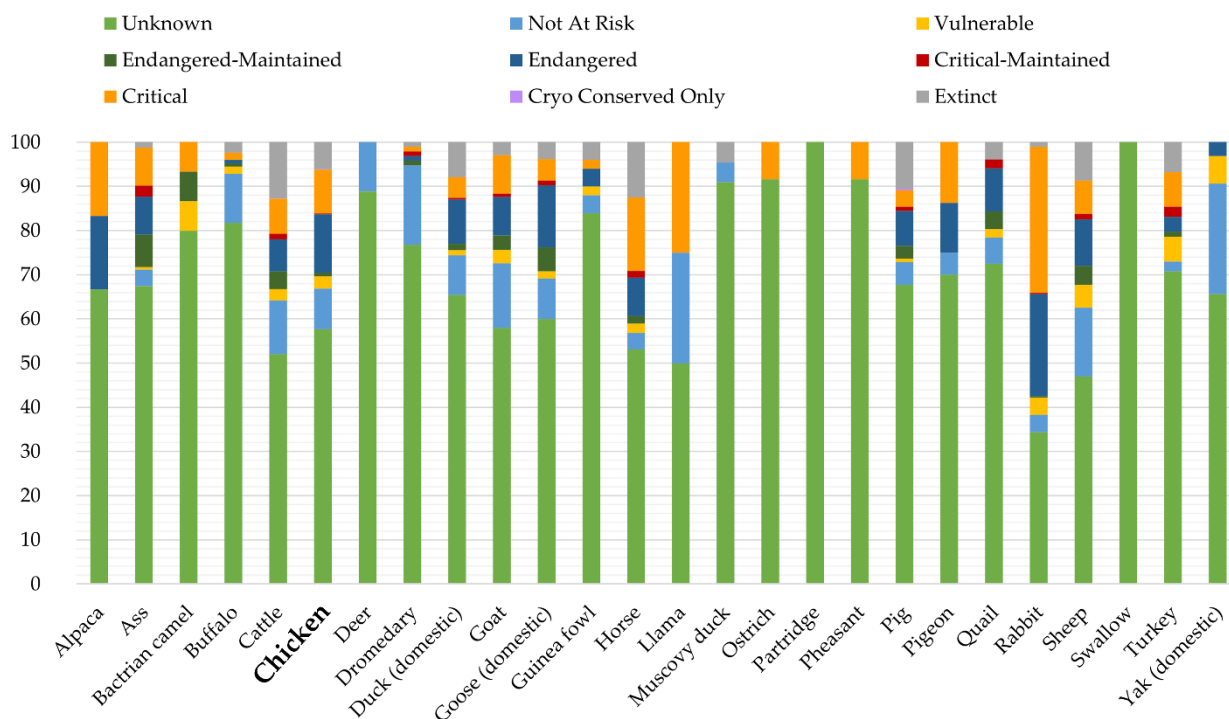


Figure 1. Classification status of breeds across species according to FAO DAD-IS (as of 2021).

The importance of such local genotypes relies on the fact that they thrive on elements of organic and free-range farming due to the suitability of their nature to adapt to their origin area [13]. This has been confirmed by the international experience of several countries, where slow-growing native chicken breeds have been able to provide good-quality meat, at a reasonable price, which is the main rationale behind the increasing demand for distinctive products [14].

On the one hand, nutritional quality comprises the content of macro- and micronutrients, unsaturated fatty acids, high-value protein, cholesterol, and other biologically active compounds. On the other hand, organoleptic quality considers sensory-related desirable traits such as meat flavour, aroma, and colour as essential traits to monitor [15]. It is the simultaneous evaluation of both which may determine the right market niche and target consumers for which each meat cut or type may be aimed [16].

Last but not the least, meat quality is subject to trends, and current consumer tastes are characterized by the challenge of obtaining low-fat meat products that preserve all the tenderness, juiciness, and good flavour and aroma of high-fat meat products [17,18]. Additionally, meat quality traits are influenced by factors of a very different nature, such as the slaughter age of birds, the feed provided to them [19], or genetic factors inherent to the genotype which the meat cuts derive from. Hence, making the appropriate choice of a specific chicken breed/variety or commercial hybrid is necessary if our aim is to maximize the expected commercial outcomes.

For these reasons, the first objective of the present study is to determine the differential clustering patterns described by the carcass and meat quality-related traits defining the cuts of meat of worldwide local chicken breeds. Second, the benefits that derive from the use of data mining are verified through the development of a functional tool to quantify the similarities and dissimilarities across carcass cuts derived from autochthonous chicken genotypes whose product quality or quantity analysis has been previously scientifically studied. The outcomes of the present study will help to tailor specific solutions to fulfill the needs of certain market niches based upon those meat cuts derived from alternative poultry farming and locally adapted breeds worldwide. Moreover, the tool that has been developed in this study may help plan the methodology for future research involving minority populations of chickens when seeking to evaluate meat quality traits in particular.

## 2. Materials and Methods

### 2.1. Systematic Review Approach Decision

The approach followed in the present systematic review has been reported to be an efficient tool in the scope of animal science specific topics [20–22]. PRISMA guidelines were discarded, given that PRISMA criteria for systematic reviews were developed in the scope of healthcare research; hence, this does not fit the diverse range and nature of the documents in which the information in regard to local breeds is made available to the public [23]. This has been supported by studies such as that by Tam et al. [24], who suggested that the adherence level of certain journals to the PRISMA statement does not significantly change whether they endorse or recommend such a guideline. Furthermore, other authors such as Haddaway et al. [25] report the limited applicability of PRISMA guidelines for reviews in conservation and environmental management.

### 2.2. Data Collection

Our data collection methodology followed the premises described in previous studies [20–22]. Two platforms ([www.google.scholar.es](http://www.google.scholar.es) and [www.sciencedirect.com](http://www.sciencedirect.com); accessed on 27 May 2022) were used for the document search [26]. The possibility to extract data from repositories was an applied inclusion criterion. In this regard, although the aforementioned repositories permit data extraction for further process and assessment, other repositories such as [www.webofscience.com/wos/woscc/basic-search](http://www.webofscience.com/wos/woscc/basic-search) and [www.ncbi.nlm.gov/pubmed/](http://www.ncbi.nlm.gov/pubmed/) (accessed on 27 May 2022) do not. Thus, this fact prompted their exclusion as information sources. Non-open access full manuscripts were accessed via the University

of Córdoba library service. The document search was performed using the subsequent keyword list h: carcass or meat quality/characterization followed each one with the words local/native/indigenous/autochthonous poultry or chicken breed, or any related term in their semantic fields [20,27]. After document collection, our study database comprehensively contained documents published from 2002 to 2021. The document search was completed by 31 December 2021 to ensure the document database comprised all documents published during 2021.

As an inclusion criterion, only those research documents which involved breeds cataloged as native in the DAD-IS database were considered for statistical analyses [28]. As a result of this selection process, 91 publications, published in English and Spanish languages, were selected to evaluate the quality of different meat pieces from different local chicken genotypes. Traits evaluated in the documents were sorted into clusters, as shown in Table 1. Unit conversion was carried out to standardize the information reported in the different documents so as to be able to quantify the quality of the different carcass pieces across all the breeds that were studied. The most widely used unit across the documents considered for each particular variable was chosen as a reference for unit conversion.

**Table 1.** Clusters, units, and references of the traits considered in the studies.

| Cluster                 | Trait                    | Unit  | References   |
|-------------------------|--------------------------|---|--|
| Weight-related traits   | Carcass/piece weight     | g   | [29–110]   |
|                         | Carcass yield            | %   |  |
|                         | Cold weight              | g   |  |
| Water-holding capacity  | Drip loss                | %   | [29,31,32,36,38,40–42,44,45,52,53,56–59,61–63,65,67,68,70–76,81,82,84,89,93,103–105,108,109,111–121]             |
|                         | Water-holding capacity   | %   |  |
|                         | Cooking loss             | %   |  |
| Colour-related traits   | L* meat                  | [29,31,32,34,36,38–42,44,45,47,50,52,53,57–59,61–65,68,70,71,73–76,80,82,84,93,98,99,101,103,104,111–113,115,117–119,121,122] |  |
|                         | a* meat                  |   |  |
|                         | b* meat                  |   |  |
|                         | L* meat 72 h post-mortem |   |  |
|                         | a* meat 72 h post-mortem |   |  |
| Histological properties | L* skin                  | [35,40,56,65,73,104,105,123]  |  |
|                         | a* skin                  |   |  |
|                         | b* skin                  |   |  |
| Texture-related traits  | Muscle fiber density     | fibers/mm <sup>2</sup>  | [29,32,34,36,38–42,44,45,50,52,53,56–59,62,63,65,67,70,75,76,81,82,84,93,103–105,109,111–115,117–122]            |
|                         | Muscle fiber diameter    | µm  |  |
| pH                      | Firmness                 | kg s <sup>-1</sup>  | [29–34,36,38–42,45,47,50,52,53,57–59,61–65,67,68,70–76,82,84,87,89,93,98,99,101,103–105,111–115,117–122,124–126] |
|                         | Total work               | kg mm   |  |
|                         | Shear force              | N   |  |
|                         | Hardness                 | N   |  |
|                         | Springiness              | Mm  |  |
|                         | Cohesiveness             | N   |  |
|                         | Gumminess                | N   |  |
| Chewiness               | kg mm                    |   |  |
| pH                      | pH                       | [29–34,36,38–42,45,47,50,52,53,57–59,61–65,67,68,70–76,82,84,87,89,93,98,99,101,103–105,111–115,117–122,124–126]              |  |
|                         | pH 24 h post-mortem      |   |  |
|                         | pH 72 h post-mortem      |   |  |

Table 1. Cont.

| Cluster                                | Trait       | Unit     | References  |
|--|-------------|----------|---|
| Content of flavour-related nucleotides | IMP         | mg/g     | [41,48,56,77,109,113,118]   |
|  | AMP         | mg/100 g |   |
|  | Inosine     | mg/100 g |   |
| Gross nutrients                        | Moisture    | %        | [6,30,33,34,36,38–43,47,49,52,55,56,58–61,64,65,67,68,70,72,75,77,78,80,83,84,87,89,92,93,99,100,102–105,107,109,111–116,119–121,124,125,127] |
|  | Protein     | %        |   |
|  | Fat         | %        |   |
|  | Ash         | %        |   |
|  | Collagen    | %        |   |
|  | Cholesterol | mg/100 g |   |

The information present on each document was sorted into the different study observations depending on the following factors: breed, sex, sex status, slaughtering age, and meat cuts. In regard to sex/sex status, the possibilities (levels) considered were female, male, both (when females and males were used in the documents without being reported separately), capon, and poulard.

Thirty-five different meat cuts or carcass components were studied as follows: abdominal fat, back, blood, breast, caeca, carcass (whole carcass), carcass remainder, comb, drumstick, feathers, giblet, gizzard, head, heart, intestine, liver, lungs, neck, ovary, pancreas, pelvis, proventriculus, rear, ribs, shanks, skeletal, skin, spleen, testes, thighs, thymus, trunk, viscera (whole viscera), wattles, and wings.

On the whole, 39 variables were included in the statistical analyses: sex (sex and sex status), slaughtering age, carcass/piece weight, carcass yield, cold weight, drip loss, water-holding capacity, cooking loss, L\* meat, a\* meat, b\* meat, L\* meat 72 h post-mortem, a\* meat 72 h post-mortem, b\* meat 72 h post-mortem, L\* skin, a\* skin, b\* skin, muscle fiber density, muscle fiber diameter, drip loss, water-holding capacity, cooking loss, firmness, total work, shear force, hardness, springiness, cohesiveness, gumminess, chewiness, pH, pH 24 h post-mortem, pH 72 h post-mortem, IMP, AMP, inosine, moisture, protein, fat, ash, collagen, and cholesterol.

All techniques and methodologies followed in the different research documents to collect the measurements of each particular explanatory variable were standardized and described in the research procedures present in each document. For this reason, the rationale behind the present research was not to infer about the methods provided, as reported in the literature, when standardized laboratory techniques were used; even if empirical differences may have been detected at first sight, these differences were statistically nonsignificant [128,129].

### 2.3. Data Analysis

#### 2.3.1. Multicollinearity Prevention: Preliminary Testing

Before performing the statistical analyses per se, a multicollinearity analysis was run to discard potential strong linear relationships across explanatory variables and ensure data independence. In this way, before data manipulation, redundancy problems can be detected, which limits the effects of data noise and reduces the error term of discriminant models. The multicollinearity preliminary test helps to identify unnecessary variables which should be excluded, preventing the overinflation of variance explanatory potential and type II error increase [130].

The variance inflation factor (VIF) was used to determine the occurrence of multicollinearity issues. The literature reports a recommended maximum VIF value of 5 [131]. On the other hand, tolerance ( $1 - R^2$ ) concerns the amount of variability in a certain independent variable which is not explained by the rest of the dependent variables considered (tolerance > 0.20) [132].

The multicollinearity statistics routine of the describing data package of XLSTAT software (Addinsoft Pearson Edition 2021, Addinsoft, Paris, France) was used. The following formula was used to calculate the VIF:

$$\text{VIF} = 1/(1 - R^2), \quad (1)$$

where  $R^2$  is the coefficient of determination of the regression equation.

### 2.3.2. Data-Mining Chi-Squared Automatic Interaction Detection (CHAID) Decision Tree: Splitting, Pruning and Building

The CHAID decision tree was used to classify, predict, interpret, and develop discrete categorized data tool inference. The tree routine of the Analyzing Data package of the XLSTAT software (Addinsoft Pearson Edition 2021, Addinsoft, Paris, France) was used.

In the decision tree, each internal node was built around an input variable (meat or carcass quality traits) when a significance split criterion of the chi-square test ( $p < 0.05$ ) in the so-called pre-pruning process was met.

Pre- or post-pruning methods prevent the oversizing of trees to avoid failures by seeking the addition of traits (branches) that significantly add to the overall fit [133]. Nodes that did not significantly contribute to the global prediction were discarded. After the process, the tree obtained exhaustively represents the significant relationships across the levels of the dependent variable. Additionally, CHAID is used to penalize model complexity through an adjustment of Bonferroni inequality by significance levels.

Consecutive chi-squared tests are performed during the tree-building configuration process [134]. While branches represent the test results (in a number of two or more), the leaf nodes (or terminal nodes) represent the category levels of the target variables (the piece of carcass). Classification decisions are made at each node from the first root node placed at the top of the tree. Each data record is explored along the tree until it reaches a terminal node or leaf. The correlation matrix obtained from the development of the data mining analysis was graphically depicted through the use of the web server Heatmapper ([www.heatmapper.ca](http://www.heatmapper.ca); accessed on 30 June 2022) [135].

### 2.3.3. CHAID Decision Tree Cross-Validation

Ten-fold cross-validation was performed to ensure that the set of predictors considered significantly explains the differences across dependent variable groups to validate the outcomes of the CHAID decision tree. All sample records of the training sample and the study data were used to perform the ten-fold cross-validation [133].

For ten-fold cross-validation, we created 10 random subsets of the original data, setting one fold aside which was used as a test set. Afterwards, we built a tree for the remaining folds ( $10 - 1$ ), and evaluated the tree, comparing it against the test fold. Then, we built one tree with the remaining 90% of the cases for each of the 10 subsets (subsamples). The 10% subset was treated as a test sample (subset). For a 10-fold validation, each of the 10% folds (mutually exclusive and summing up to the total observations in the sample) at once serve as a test sample and as part of the learning sample 9 times. Cross-validation compares the differences between prediction errors for a tree applied to a new potential sample (resubstitution error rate) and a training sample (cross-validation error rate).

The 'complexity parameter' (cp) was used to perform the cross-validation of the decision. The complexity parameter (cp) controls the size of the decision tree and helps to select the optimal tree size. When adding another variable to the decision tree from the current (lowest) node implies a statistical cost or increases the complexity of the discriminant model above the value of cp, then tree building stops.

The resubstitution or replacement rate refers to the proportion of misclassified original observations across the various subsets of the original tree. The resubstitution rate decreases as tree depth increases. The lowest resubstitution/replacement error rates are yielded by the largest tree. Notwithstanding, selecting trees reporting the lowest resubstitution rate

may not be the best choice due to the potential bias derived from redundant variable inclusion. Large trees add random variation to the predictions given they overfit outliers.

Ten-fold cross-validation was used to obtain a cross-validation error rate (risk). The cross-validation risk is an averaging of the risks across the 10 test samples (folds) This process was repeated for each fold, evaluating an estimate of such error. The sum of the error in the 10 portions represented the cross-validation error rate. Finally, the tree that produced the lowest cross-validation error rate and, therefore, presented the best fit, was selected. The best tree can be defined as the tree closest to the minimum. Hence, it can be used to determine the accuracy of the discriminant model for data prediction. Contextually, Albayrak [136] reports that the optimal tree depth can be identified as the shallowest tree whose cross-validation risk does not exceed the risk of the minimum cross-validation risk tree, plus one standard error of this tree's cross-validation risk. This means that the resubstitution error rate and the cross-validated error rate must be compared to choose the optimal tree as a counteracting measure of the bias derived from the overfitting of outliers.

### 3. Results

#### 3.1. Multicollinearity Prevention: Preliminary Testing

A summary of values for VIF and tolerance is reported in Table 2. Variables whose VIF values were  $\geq 5$  were discarded from further analyses. Thus, the traits removed for the following statistical analyses were sex, L\* meat 72 h post-mortem, a\* meat 72 h post-mortem, b\* meat 72 h post-mortem, firmness, chewiness, and pH 72 h post-mortem.

**Table 2.** Multicollinearity analysis of meat and carcass quality-related traits.

| Statistics/Traits      | VIF <sup>1</sup> | Tolerance (1 – R <sup>2</sup> ), |
|------------------------|------------------|----------------------------------|
| Chewiness              | 4.0515           | 0.2468                           |
| Gumminess              | 3.1989           | 0.3126                           |
| Hardness               | 2.3258           | 0.4300                           |
| Shear force            | 2.0546           | 0.4867                           |
| a* meat                | 1.8862           | 0.5302                           |
| b* skin                | 1.7745           | 0.5635                           |
| a* skin                | 1.7044           | 0.5867                           |
| Muscle fiber diameter  | 1.6223           | 0.6164                           |
| Cooking loss           | 1.6202           | 0.6172                           |
| L* skin                | 1.6152           | 0.6191                           |
| L* meat                | 1.5910           | 0.6285                           |
| Water-holding capacity | 1.5580           | 0.6418                           |
| pH                     | 1.4108           | 0.7088                           |
| Drip loss              | 1.3886           | 0.7201                           |
| pH 24 h post-mortem    | 1.3486           | 0.7415                           |
| Moisture               | 1.3462           | 0.7428                           |
| b* meat                | 1.3408           | 0.7458                           |
| Total work             | 1.2699           | 0.7875                           |
| IMP                    | 1.2534           | 0.7978                           |
| Springiness            | 1.2183           | 0.8208                           |
| Cholesterol            | 1.2101           | 0.8264                           |
| Cohesiveness           | 1.1135           | 0.8981                           |
| Collagen               | 1.1130           | 0.8985                           |
| Inosine                | 1.1058           | 0.9044                           |
| Carcass/piece weight   | 1.0949           | 0.9133                           |
| Carcass yield          | 1.0898           | 0.9176                           |
| Protein                | 1.0761           | 0.9293                           |
| AMP                    | 1.0735           | 0.9315                           |

Table 2. Cont.

| Statistics/Traits    | VIF <sup>1</sup> | Tolerance (1 – R <sup>2</sup> ), |
|----------------------|------------------|----------------------------------|
| Ash                  | 1.0463           | 0.9558                           |
| Muscle fiber density | 1.0317           | 0.9692                           |
| Cold carcass weight  | 1.0275           | 0.9732                           |
| Average age          | 1.0267           | 0.9740                           |
| Fat                  | 1.0213           | 0.9792                           |

<sup>1</sup> Interpretation thumb rule: VIF ≥ 5 (highly correlated); 5 > VIF > 1 (moderately correlated); VIF = 1 (not correlated).

3.2. Data-Mining Chi-Squared Automatic Interaction Detection (CHAID) Decision Tree: Splitting, Pruning, and Building

The data-mining CHAID decision tree obtained from the chi-square dissimilarity matrix built in this study is represented in Supplementary Figure S1. A correlation matrix across carcass and meat quality-related traits were computed and are graphically represented in Figure 2. The chi-square-based branch and node distribution suggested that observations significantly ( $p < 0.05$ ) differed across carcass meat cuts. Supplementary Tables S1 and S2 report the frequency distribution of the presence of each cut across the range of levels for the particular carcass or meat quality trait represented by the different nodes within the tree structure.

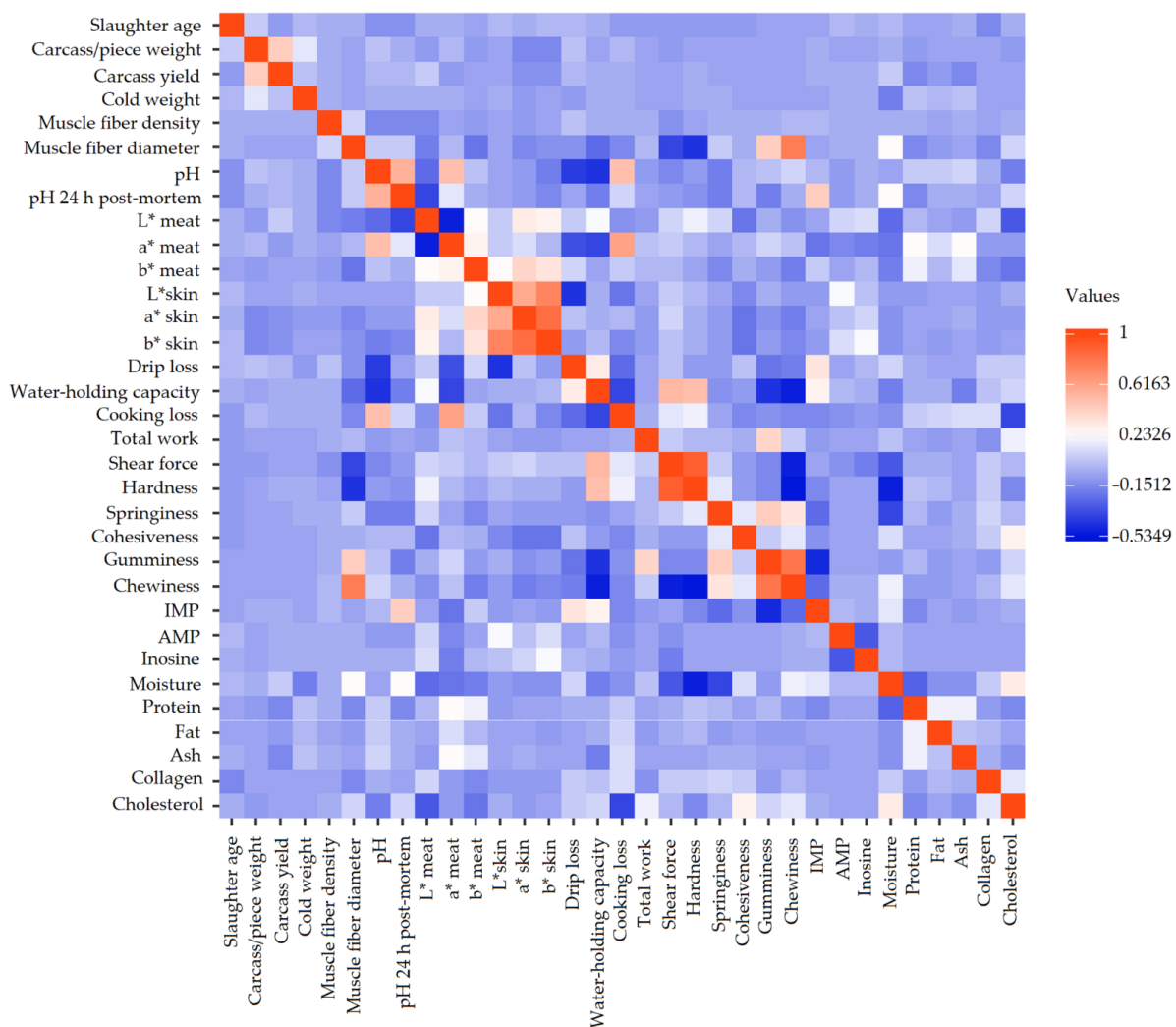
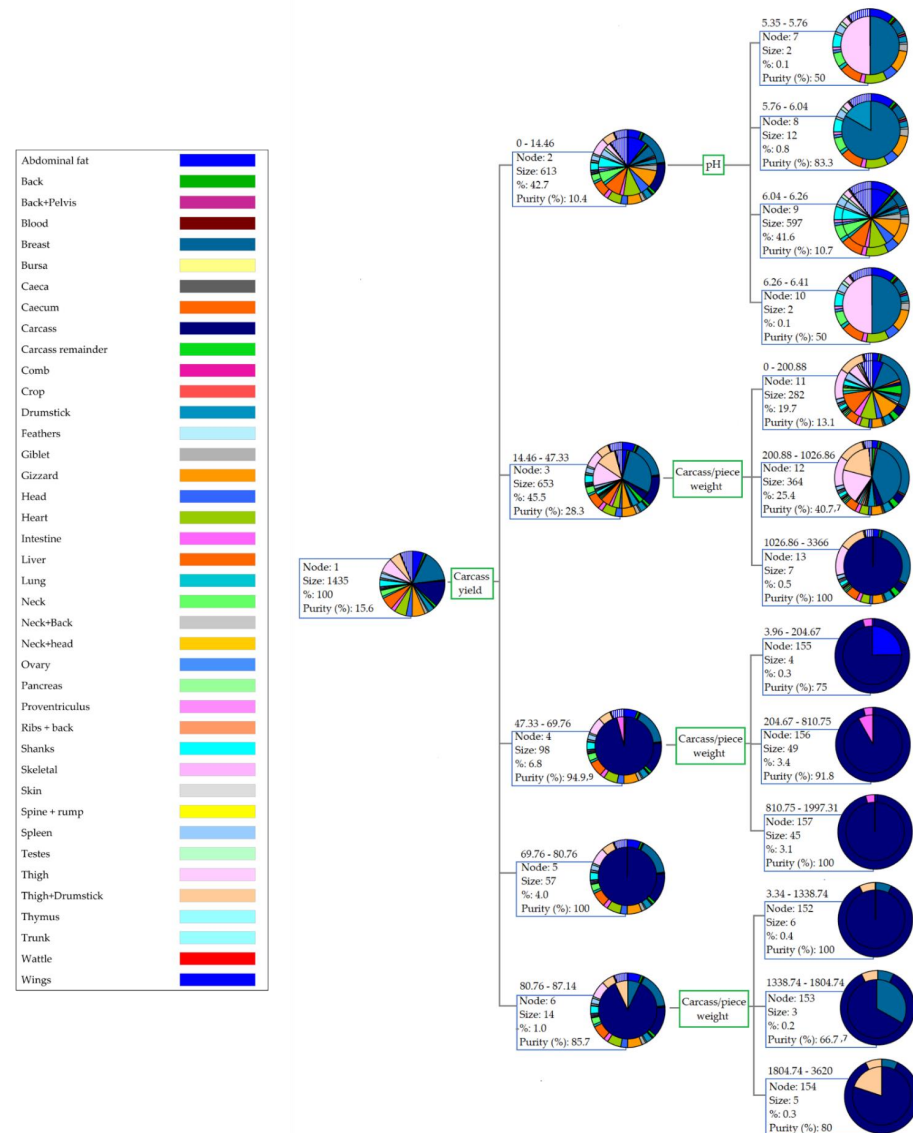


Figure 2. Correlation matrix between the different quality-related traits.



The first three branches of the CHAID decision tree are summarized in Figure 3. Five groups were depicted in the first classification (first node: 0–14.46; second node: 14.46–47.33; third node: 47.33–69.76; fourth node: 69.76–80.76; fifth node: 80.76–87.14), derived from the representativity of carcass yield for the different pieces. After this, observations were sorted into subnodes originating depending on the values for pH and carcass/piece weight in each meat cut.



**Figure 3.** Graphical representation of the first three branches of the CHAID decision tree considering meat cuts as the clustering criterion.

### 3.3. CHAID Decision Tree Cross-Validation

Finally, the validity and robustness of the results were cross-validated and the number of erroneously classified observations for each piece was computed. As reported in Table 3, the risk estimates ( $\approx 0.600$ ) and standard errors (0.013) of the model applying the cross-validation test did not differ from the results of the model without the cross-validation test. Hence, the stability of the model was guaranteed.

**Table 3.** Complexity parameter (Cp) evaluation through the comparison of model-based (resubstitution) statistics and ten-fold cross-validation error rate (risks).

| Risk (Cp)                   | Estimate | Std. Error |
|-----------------------------|----------|------------|
| Resubstitution error rate   | 0.604    | 0.013      |
| Cross-validation error rate | 0.622    | 0.013      |

#### 4. Discussion

The present research develops an updated evaluation of international research studies focusing on carcass characterization in autochthonous chicken breeds worldwide. The imbalance between the economic resources allocated to native genotypes when compared with commercial hybrid strains produces a gap in the knowledge, visualization, and impact that such local genotypes will eventually have in the research community and by extension in society [137].

Recent decades have been characterized by a progressively reducing trend in genetic diversity. Such a reduction has not only affected diversity across chicken genotypes, but also within-genotype diversity. Such a lack of diversity compromises one of the main needs that poultry production seeks to fulfil: the provision of a sufficiently diverse genetic background so as to face the adaptation to climate change and meet consumer preferences, as well as current and forthcoming market demands [138]. In this regard, to ensure such an objective is attained, breeds' long-term survival cannot be left aside, and for this the knowledge on such breeds must be deepened. The characterization of local resources and of the products which derive from them is, therefore, critical.

Local breeds have proven to be sources for products which are well appreciated in specialized market niches, although their lower productivity in comparison with selected breeds often needs to be supported by governmental incentives which sometimes barely cover production costs.

These products' distinctive features and enhanced quality may be highly valued by consumers. Such an increased popularity of breed-linked products in turn favours investments in local farmers, who act as the main preservatory agents of domestic poultry breed biodiversity [139].

As suggested in González Ariza et al. [140], redundant variables need to be discarded prior to statistical analyses, leading to CHAID decision tree building before the splitting and pruning stages. In line with this premise, on the one hand, L\* meat 72 h post-mortem, a\* meat 72 h post-mortem, b\* meat 72 h post-mortem, and pH 72 h post-mortem were deemed redundant variables and were discarded from further analyses. The basis for these redundancies may rely on the fact that measurements taken at 72 h post-mortem may be poorly representative of carcass quality, especially considering prior sampling. This finding suggests the fact that, in research dealing with the study of products derived from local chicken breeds, quality parameter measurements may not need to be taken exceeding 72 h post-mortem, as this may not report any relevant information which has not been provided by earlier measurements.

On the other hand, multicollinearity problems were also reported for the sex variable. These redundancies may be ascribed to the lack of occurrence of significant differences between females and males in the values reported for the different parameters studied. Indeed, the only empirical differences between sexes concerned the piece or carcass weight and yield variables, and were small but still nonsignificant.

Parallely, multicollinearity problems were also reported for texture-related traits such as firmness and hardness. Such a strong relationship may also be the source of misconception between the perception of consumers of these two parameters. For example, while increased hardness is always reported as an undesirable feature in meat, an increased firmness may be a sign of a better performance of meat for certain culinary preparations which involve boiling techniques, given meat does not crumble. Specifically, while firmness has been defined as the peak force exerted when a sample was compressed to a depth of

1.5 cm, using a block of wood of identical dimensions to the sample, hardness is defined as the peak force exerted when a metal probe is inserted into the sample to a depth of 1.5 cm [141].

Once redundant variables were discarded, the chewiness and gumminess traits reported the highest value (0.655) in the correlation matrix. Chewiness is the product of gumminess by springiness [142]. Hence, if values of springiness are close to 1, in general, this may be indicative of gumminess and chewiness having similar values, thus being highly correlated texture-related traits as well.

Furthermore, a high positive correlation (0.529) was found between chewiness and muscle fiber diameter. Chewiness is measured performing a sensory evaluation using a simulation of human chewing [143]. The muscle fiber diameter determines the textural characteristics of meat in a determinant moment [144]. A positive correlation between muscle fiber and texture-related characteristics has been reported in previous studies [144], with native chicken breeds presenting high values in the texture profile analysis [145,146].

The shear force/hardness pair of traits reported the second highest positive value in the correlation matrix (0.638). This may derive from the fact that the shear force trait can be defined as the force required to sever a sample of meat [147], while hardness is defined as the peak force required for the first meat compression [148].

In regard to colour-related traits, the  $b^*$  skin trait highly correlated with  $L^*$  skin and  $a^*$  skin (0.459 and 0.561, respectively). Individuals displaying dark skin pigmentation have been reported to present low  $L^*$ ,  $a^*$ , and  $b^*$  skin values. In contrast, higher values of  $L^*$ ,  $a^*$ ,  $b^*$  indexes were reported for the skin of lighter-coloured birds [149]. Parallely, when colour coordinates were measured in meat, the highest negative values in the correlation matrix were obtained between  $L^*$  and  $a^*$  values. Some authors have proposed that low  $L^*$  values in meat may most likely be ascribed to high myoglobin concentrations [150]. Additionally, the shift from the glycolytic to oxidative fiber types results in a higher concentration of muscle myoglobin and produces darker meat (higher  $a^*$  and  $b^*$  values) in the carcass [151,152].

As depicted in Figure 3, the best discriminating ability was reported for carcass yield. Certain factors such as the genotype and the environment where birds grow may interact, and such an interaction may be the source not only for large differences in the yield that meat cuts and carcass eventually reach, but also for their high variability [153].

Contextually, in local genotypes which are well-adapted to alternative organic or free-range production systems, the development of frequent extensive movements and exercise which compels animals to generate increased kinetic forces is particularly evidenced through the higher development of certain cuts such as the thighs and the drumsticks. In this regard, limb-related cuts are the parts of a bird's body which most actively participate in the successful development of kinetics, which in turn may explain the increased volumes that they reach [154]. Indeed, even with poultry being considered a species of a 'white' meat type, it may not be surprising that increased levels of myoglobin can be found in limb-related areas, which confers them a rather darker aspect derived from this mixed red/white type of fibers, which is even preserved after cooking. Furthermore, as physical exercise increases, fat deposition decreases, which is why lower values of abdominal fat are observed in animals whose life or the greatest part of their life occurs outdoors [155].

Still, yield may be conditioned by other factors such as the age of the individuals or even the breed to which animals belong. In this regard, the yield of the different cuts in the chicken carcass may change along the different stages of growth during the life of the individuals, with breast and thigh yields reaching a greater development than other cuts in older individuals (allometric growth) [156]. This may differ across breeds which have a relatively slow growth, which in turn may be the basis for the great variability found in the slaughter age of the animals.

The pH was a determinant discriminant factor in cuts weighing less than 14.46 g. pH has been related to several meat quality-related attributes including colour, water-holding capacity, tenderness, juiciness, cooking loss, shelf life, and slaughter age [29,58,157].

According to the literature, higher meat pH values are related to rather effective desirable colour retention and moisture absorption properties [158]. Additionally, lower values of pH are related to a rather sour perception of meat flavour by consumers, while higher pH values have been linked to more pleasant, sweeter tastes [159].

In most studies, pH measurement is only taken in the pectoralis major and biceps femoris muscles (breast and thigh muscles). This may be a source of bias given the pH values of thigh muscles are likely higher than those for breast muscles [160]. Muscle exercise increases the number of mitochondria in  $\alpha$ W fibers, converting them into  $\alpha$ R fibers [161]. This triggers the increase in muscle oxidative capacity to fulfill the needs of the exercise being developed [162]. Additionally, the enhancement of the aerobic catabolism of pyruvate causes a sparing of glycogen, given the oxidative pathway is the most way method to produce energy, which eventually may explain the pH differences across the different meat cuts [163].

A high discriminant potential was reported for the carcass/meat cut weight variables, which were both highly variable traits across breeds. In this context, genotype, environmental factors, and slaughter age have been reported to determine the weight that the different meat cuts reach [154,164]. The weight of the whole carcass and noble cuts, such as breast, thigh, and drumstick, have high economic and environmental importance for poultry meat production since these traits are considered the main production indicators and are the cuts which eventually reach the highest processes due to their appreciation by consumers. However, reaching good production results is conditional to the efficient use of feeds and water [165].

Countries around the world have suffered from the economic impact of the COVID-19 pandemic since 2020. This situation has been exacerbated by the recent Russia–Ukraine war conflict in 2022, as world economies may witness another rise in commodity prices and “supply chain chokeholds” [166]. The world’s largest supplier of wheat is Russia which, together with Ukraine, accounted for about 28% of the sum of global exports during the years 2015–2020 [167].

Within this global framework, autochthonous breeds characterized by specific attributes and features, such as biological breeding, sustainable production system idoneity, and their efficient use of alternative raw materials, may represent a solution to the inflation and economic instability plaguing meat production systems. In this sense, products derived from local genotypes may need to be valued and integrated into the market as quality products, taking advantage of recent trends in consumers, who progressively seek to purchase products which come from less intensive production systems [168]. In this regard, institutional support is necessary to develop investigation studies concerning local breeds, which in turn will act as a protective measurement for these genotypes, considering their ecologic value, and provide oriented market strategies based on a better and conscious valuation of sustainable products [169].

As we progressed in the valuation of the CHAID tree, the third division of the tree subnodes suggested other variables may play an important role in the classification of different meat cuts. Among them, a high discriminant potential was revealed for slaughter age. The basis for such an increase in discriminant potential may stem from the high variability reported for slaughtering age worldwide. Native genotypes are genetically and culturally integrated into the areas from which they come; hence, the determination of age of slaughter mainly affects certain characteristics of the meat that particularly adapt to the local culinary culture of the area in which specific breeds are reared [3,170].

The influence of slaughtering age also explains the high influence of age on the classification of the different pieces in the decision tree, for instance, the high positive correlation with myofiber size. Specifically, larger myofiber diameters and lower myofiber density may translate into larger cut sizes. Indeed, the number of myofibers does not increase after hatching, but meat cut growth is produced by the growth of each myofiber, which may sustain the aforementioned [170].

Simultaneously, as age increases, a decrease in lactate dehydrogenase and phosphofructokinase, which are two glycolytic enzymes, is produced in the chicken muscle. The reduced glycolytic potential produces an increase in pH in older individuals, which is more evident in noble cuts compared to the rest [171,172]. In addition to this, heavier chickens have been reported to present higher plasma glucose and, therefore, are more prone to pre-slaughter stress than lighter ones [173]. This could lead to low muscle glycogen and high pH values at the time of death in older individuals [36,174].

Among other differences between younger and older chickens, the meat of older individuals contains higher myoglobin and collagen proportions [175,176]. It is the variation in concentration in such compounds during the life of the animals which determines the higher or lower repercussion of age on colour, texture, and water-holding capacity-related traits.

Within the gross nutrient cluster, protein presented the highest discriminant potential. Indigenous chickens have been reported to usually present progressively higher protein levels as they age [177]. In this way, the high variability in the slaughter age variable also caused the protein content to vary across the genotypes that were sampled in this study. Nevertheless, the conditioning effects of other factors such as genotype and sex on protein content cannot be discarded, as suggested by the literature [177,178].

Even though the three variables classified within the weight-related traits cluster reported the least relevant discriminating cluster when compared with the rest of the clusters, a relatively high discriminating potential was observed for the cold weight variable when compared to its cluster counterparts. This variable closely relates to the initial carcass/piece weight of individuals. A loss in carcass weight is produced by the action of the cold air in forced circulation when carcasses are conserved into cooling chambers [140].

Last but not the least,  $L^*$  meat was the only variable to appear as a discriminant criterion in the first three divisions of the decision tree. Chicken meat is translucent; however, when tissues have high pH values, light scattering is weak. This means that the light path through the tissue is relatively long and the selective absorbance of light myoglobin and its derivatives increases. However, in low-pH-value tissues, light scattering is strong, the path of light through the fibers is relatively short, and the selective absorbance of light decreases. Therefore, meat translucency comprehensively and highly influences all meat colourimetry-related parameters [140].

In this context, a large number of colorimeters whose original function is to measure the colour of plastic, metal, or painted surfaces can be found in the market. Such colorimeters are erroneously used in research studies since optical problems derived from the translucency of chicken meat are not taken into account [179]. Instead, the meat colouration of every genotype is matched to the specific requirements of a particular market [180]. Furthermore, the  $L^*$  meat is also influenced by post-mortem glycolysis. Consequently, chicken nutrition, transport to the slaughterhouse, slaughter, and the refrigeration method used in each culture could contribute to  $L^*$  meat variation [179].

The present research should be taken into account when deciding which breeds should be used as control and test groups in studies aiming to determine carcass and meat cut quality in chickens. Furthermore, not only the factors that should unavoidably be considered when planning studies are proposed, but also which parameters may hold the greatest capacity to explain intergroup variability, which is eventually the source of significant differences. This enhances the efficiency of the methods used by poultry-related science and maximizes the outcomes derived from future research, which in turn is one of the milestones on which to support autochthonous breed sustainability and preservation.

## 5. Conclusions

Preliminary multicollinearity analyses suggested that meat quality parameters need not be measured after 72 h post-mortem since the information they offer can be supplemented with the rest of the variables collected at the slaughter moment. Small nonsignificant differences between males and females are responsible for the lack of effect of sex on carcass and meat cut quality. Regarding texture-related traits, multicollinearity problems

between firmness and hardness may be the source of the misconception between their perception by consumers. On the other hand, gumminess and chewiness variables are highly correlated via their connection to springiness. Native breeds generally present high texture values due to their reduced muscle fiber diameter. The strong relationship between shear force and hardness may derive from conceptual similarities. Individuals displaying dark skin pigmentation present low  $L^*$ ,  $a^*$ , and  $b^*$  skin values, as opposed to lighter-coloured skin birds. High myoglobin concentrations in local breeds are responsible for their low  $L^*$  values. Meat translucency ( $L^*$ ) is also conditioned by slaughtering stress and handling factors, and highly influences all meat colorimetry parameters. Thus, colorimeters may be erroneously used, given that translucency is not considered but directly matched with the specific requirements of particular markets. Slaughtering age, genotype, and environment interaction are the sources of carcass and meat cut yield and weight variability. Higher pH values imply a rather effective desirable colour retention, moisture absorption, and more pleasant sweeter tastes. Reduced glycolytic potential, higher plasma glucose, and proneness to pre-slaughter stress produce an increase in pH and decrease in muscle glycogen in older and heavier individuals, which is more evident in noble cuts. Slaughtering age choice conditions meat characteristics, and in turn is a manner to adapt to the local culinary culture of the area in which specific breeds are reared. Larger cut sizes derive from larger myofiber diameters but lower myofiber density, as meat cut growth is produced by the growth of each myofiber. Moreover, indigenous chickens usually present progressively higher variable protein content with age. The present tool helps to tailor efficient study plans for specific carcass and meat cut quality studies in autochthonous breeds, which in turn may act as strategy reinforcers for local genotype sustainability in the long term.

**Supplementary Materials:** The following supporting information can be downloaded at: <https://www.mdpi.com/xxx/s1>. Figure S1: Data-mining CHAID decision tree. Table S1: Frequency of each breed at different nodes of the data-mining CHAID decision tree. Table S2: Data-mining CHAID decision tree structure.

**Author Contributions:** Conceptualization, A.G.A., F.J.N.G. and J.V.D.B.; data curation, A.G.A., F.J.N.G. and J.M.L.J.; formal analysis, A.G.A., F.J.N.G., J.M.L.J. and A.A.A.; funding acquisition, J.M.L.J. and J.V.D.B.; investigation, A.G.A., F.J.N.G., J.M.L.J., A.A.A. and M.E.C.V.; methodology, A.G.A., F.J.N.G., J.M.L.J. and A.A.A.; project administration, J.V.D.B. and M.E.C.V.; resources, J.M.L.J., J.V.D.B. and M.E.C.V.; software, A.G.A., F.J.N.G. and J.M.L.J.; supervision, F.J.N.G., J.V.D.B. and M.E.C.V.; validation, F.J.N.G. and M.E.C.V.; visualization, M.E.C.V.; writing—original draft, A.G.A. and F.J.N.G.; writing—review and editing, A.G.A., F.J.N.G., J.M.L.J., A.A.A., J.V.D.B. and M.E.C.V. All authors have read and agreed to the published version of the manuscript.

**Funding:** This work was financially co-supported by the FEDER project PP.AVA.AVA201601.16.

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** All data stemming from the present research are enclosed in the tables or as Supplementary Materials. Any additional data will be made accessible from the corresponding authors upon reasonable request.

**Acknowledgments:** This work would not have been possible if it had not been for the funding of FEDER Project PP.AVA.AVA201601.16, as well as the assistance of the IFAPA, Diputación de Córdoba, and PAIDI AGR 218 research group.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Caffyn, A. Broiler battles: Contested intensive poultry unit developments in a policy void. *Land Use Policy* **2021**, *105*, 105415. [[CrossRef](#)]
2. Mcleod, A.; Thieme, O.; Mack, S. Structural changes in the poultry sector: Will there be smallholder poultry development in 2030? *Worlds Poult. Sci. J.* **2009**, *65*, 191–200. [[CrossRef](#)]

3. Sokołowicz, Z.; Krawczyk, J.; Świątkiewicz, S. 4. Quality of Poultry Meat from Native Chicken Breeds—A Review. *Ann. Anim. Sci.* **2016**, *16*, 347–368. [[CrossRef](#)]
4. Hedman, H.D.; Vasco, K.A.; Zhang, L. A review of antimicrobial resistance in poultry farming within low-resource settings. *Animals* **2020**, *10*, 1264. [[CrossRef](#)] [[PubMed](#)]
5. Berthouly-Salazar, C.; Rognon, X.; Nhu Van, T.; Gély, M.; Vu Chi, C.; Tixier-Boichard, M.; Bed'Hom, B.; Bruneau, N.; Verrier, E.; Maillard, J.-C. Vietnamese chickens: A gate towards Asian genetic diversity. *BMC Genet.* **2010**, *11*, 1–11. [[CrossRef](#)]
6. Nguyen Van, D.; Moula, N.; Moyses, E.; Do Duc, L.; Vu Dinh, T.; Farnir, F. Productive performance and egg and meat quality of two indigenous poultry breeds in Vietnam, Ho and Dong Tao, fed on commercial feed. *Animals* **2020**, *10*, 408. [[CrossRef](#)]
7. Toalombo Vargas, P.A.; Navas González, F.J.; Landi, V.; León Jurado, J.M.; Delgado Bermejo, J.V. Sexual dimorphism and breed characterization of Creole hens through biometric canonical discriminant analysis across Ecuadorian agroecological areas. *Animals* **2020**, *10*, 32. [[CrossRef](#)]
8. González Ariza, A.; Navas González, F.J.; Arando Arbulu, A.; León Jurado, J.M.; Barba Capote, C.J.; Camacho Vallejo, M.E. Non-Parametrical Canonical Analysis of Quality-Related Characteristics of Eggs of Different Varieties of Native Hens Compared to Laying Lineage. *Animals* **2019**, *9*, 153. [[CrossRef](#)]
9. Muth, P.C.; Ghaziani, S.; Klaiber, I.; Valle Zárate, A. Are carcass and meat quality of male dual-purpose chickens competitive compared to slow-growing broilers reared under a welfare-enhanced organic system? *Org. Agric.* **2018**, *8*, 57–68. [[CrossRef](#)]
10. Bianchi, M.; Petracci, M.; Cavani, C. The influence of genotype, market live weight, transportation, and holding conditions prior to slaughter on broiler breast meat color. *Poult. Sci.* **2006**, *85*, 123–128. [[CrossRef](#)]
11. FAO. *Domestic Animal Diversity Information System (DAD-IS): Risk Status of Animal Genetic Resources*; FAO: Rome, Italy, 2022.
12. Suliman, G.M.; Alowaimer, A.N.; Al-Mufarrej, S.I.; Hussein, E.O.S.; Fazea, E.H.; Naiel, M.A.E.; Alhotan, R.A.; Swelum, A.A. The effects of clove seed (*Syzygium aromaticum*) dietary administration on carcass characteristics, meat quality, and sensory attributes of broiler chickens. *Poult. Sci.* **2021**, *100*, 100904. [[CrossRef](#)] [[PubMed](#)]
13. González Ariza, A.; Arando Arbulu, A.; Navas González, F.J.; Ruíz Morales, F.d.A.; León Jurado, J.M.; Barba Capote, C.J.; Camacho Vallejo, M.E. Sensory Preference and Professional Profile Affinity Definition of Endangered Native Breed Eggs Compared to Commercial Laying Lineages' Eggs. *Animals* **2019**, *9*, 920. [[CrossRef](#)] [[PubMed](#)]
14. Arando, A.; González-Ariza, A.; Lupi, T.; Nogales, S.; León, J.; Navas-González, F.; Delgado, J.; Camacho, M. Comparison of non-linear models to describe the growth in the Andalusian turkey breed. *Ital. J. Anim. Sci.* **2021**, *20*, 1156–1167. [[CrossRef](#)]
15. Jin, Y.; Cui, H.; Yuan, X.; Liu, L.; Liu, X.; Wang, Y.; Ding, J.; Xiang, H.; Zhang, X.; Liu, J. Identification of the main aroma compounds in Chinese local chicken high-quality meat. *Food Chem.* **2021**, *359*, 129930. [[CrossRef](#)] [[PubMed](#)]
16. González Ariza, A.; Nogales Baena, S.; Lupi, T.M.; Arando Arbulu, A.; Navas González, F.J.; León Jurado, J.M.; Delgado Bermejo, J.V.; Camacho Vallejo, M.E. Characterisation of biological growth curves of different varieties of an endangered native hen breed kept under free range conditions. *Ital. J. Anim. Sci.* **2021**, *20*, 806–813. [[CrossRef](#)]
17. Selamat, J.; Zaidy, N.A.; Zakaria, N.S.; Juhari, N.H.; Murugesu, S. Comparison of Physicochemical Characteristics and Sensory Attributes of Four Different Chicken Breeds from the Genuine and Selected Local Market. *J. Food Qual.* **2022**, *2022*, 1419937. [[CrossRef](#)]
18. Walley, K.; Parrott, P.; Custance, P.; Meledo-Abraham, P.; Bourdin, A. A review of French consumers purchasing patterns, perceptions and decision factors for poultry meat. *Worlds Poult. Sci. J.* **2015**, *71*, 5–14. [[CrossRef](#)]
19. Gratta, F.; Birolo, M.; Sacchetto, R.; Radaelli, G.; Xiccato, G.; Ballarin, C.; Bertotto, D.; Piccirillo, A.; Petracci, M.; Maertens, L. Effect of feed restriction timing on live performance, breast myopathy occurrence, and muscle fiber degeneration in 2 broiler chicken genetic lines. *Poult. Sci.* **2019**, *98*, 5465–5476. [[CrossRef](#)] [[PubMed](#)]
20. González Ariza, A.; Arando Arbulu, A.; Navas González, F.J.; Nogales Baena, S.; Delgado Bermejo, J.V.; Camacho Vallejo, M.E. The Study of Growth and Performance in Local Chicken Breeds and Varieties: A Review of Methods and Scientific Transference. *Animals* **2021**, *11*, 2492. [[CrossRef](#)]
21. Iglesias Pastrana, C.; Navas González, F.J.; Ciani, E.; Barba Capote, C.J.; Delgado Bermejo, J.V. Effect of research impact on emerging camel husbandry, welfare and social-related awareness. *Animals* **2020**, *10*, 780. [[CrossRef](#)]
22. McLean, A.K.; Gonzalez, F.J.N. Can scientists influence donkey welfare? Historical perspective and a contemporary view. *J. Equine Vet. Sci.* **2018**, *65*, 25–32. [[CrossRef](#)]
23. Page, M.J.; Moher, D.; McKenzie, J.E. Introduction to PRISMA 2020 and implications for research synthesis methodologists. *Res. Synth. Methods* **2022**, *13*, 156–163. [[CrossRef](#)] [[PubMed](#)]
24. Tam, W.W.; Lo, K.K.; Khalechelvam, P. Endorsement of PRISMA statement and quality of systematic reviews and meta-analyses published in nursing journals: A cross-sectional study. *BMJ Open* **2017**, *7*, e01390. [[CrossRef](#)]
25. Haddaway, N.R.; Macura, B.; Whaley, P.; Pullin, A.S. ROSES RepOrting standards for Systematic Evidence Syntheses: Pro forma, flow-diagram and descriptive summary of the plan and conduct of environmental systematic reviews and systematic maps. *Environ. Evid.* **2018**, *7*, 1–8. [[CrossRef](#)]
26. Gehanno, J.-F.; Rollin, L.; Darmoni, S. Is the coverage of Google Scholar enough to be used alone for systematic reviews. *BMC Med. Inform. Decis. Mak.* **2013**, *13*, 1–5. [[CrossRef](#)]
27. Schlosser, R.W.; Wendt, O.; Bhavnani, S.; Nail-Chiwetalu, B. Use of information-seeking strategies for developing systematic reviews and engaging in evidence-based practice: The application of traditional and comprehensive Pearl Growing. A review. *Int. J. Lang. Commun. Disord.* **2006**, *41*, 567–582. [[CrossRef](#)] [[PubMed](#)]

28. FAO. *Domestic Animal Diversity Information System (DAD-IS): Browse by Species and Country*; FAO: Rome, Italy, 2022.
29. Choo, Y.; Kwon, H.; Oh, S.; Um, J.; Kim, B.; Kang, C.; Lee, S.; An, B. Comparison of growth performance, carcass characteristics and meat quality of Korean local chickens and silky fowl. *Asian Australas. J. Anim. Sci.* **2014**, *27*, 398. [[CrossRef](#)] [[PubMed](#)]
30. Zidane, A.; Ababou, A.; Metlef, S.; Niar, A.; Bouderoua, K. Growth and meat quality of three free-range chickens and commercial broiler under the same breeding conditions. *Acta Sci. Anim. Sci.* **2018**, *40*, 39663. [[CrossRef](#)]
31. Jaspal, M.H.; Ali, S.; Rajput, N.; Naeem, M.; Talpur, F.N.; Rehman, I. Fatty acid profiling and comparative evaluation of carcass cut up yield, meat quality traits of Cobb Sasso, commercial broiler and native aseel chicken. *Pure Appl. Biol.* **2020**, *9*, 56–65. [[CrossRef](#)]
32. Yang, L.; Wang, X.; He, T.; Xiong, F.; Chen, X.; Chen, X.; Jin, S.; Geng, Z. Association of residual feed intake with growth performance, carcass traits, meat quality, and blood variables in native chickens. *J. Anim. Sci.* **2020**, *98*, 121. [[CrossRef](#)]
33. Puspita, U.E.; Saragih, H.; Hartatik, T.; Daryono, B.S. Body Weight Gain and Carcass Quality of the Hybrid Chicken Derived from the Crossing between Female F1 Kampung Super and Male F1 Kampung-Broiler. *J. Trop. Biodivers. Biotech.* **2021**, *6*, 60934. [[CrossRef](#)]
34. Franco, D.; Rois, D.; Vázquez, J.A.; Purriños, L.; González, R.; Lorenzo, J.M. Breed effect between Mos rooster (Galician indigenous breed) and Sasso T-44 line and finishing feed effect of commercial fodder or corn. *Poult. Sci.* **2012**, *91*, 487–498. [[CrossRef](#)] [[PubMed](#)]
35. Koomkrong, N.; Theerawatanasirikul, S.; Boonkaewwan, C.; Jaturasitha, S.; Kayan, A. Breed-related number and size of muscle fibres and their response to carcass quality in chickens. *Ital. J. Anim. Sci.* **2015**, *14*, 4145. [[CrossRef](#)]
36. Zanetti, E.; De Marchi, M.; Dalvit, C.; Molette, C.; Réminon, H.; Cassandro, M. Carcass characteristics and qualitative meat traits of three Italian local chicken breeds. *Br. Poult. Sci.* **2010**, *51*, 629–634. [[CrossRef](#)]
37. Iqbal, S.; Pampori, Z.; Hasin, D. Carcass and egg characteristics of indigenous chicken of Kashmir (*Kashmir favorella*). *Indian J. Anim. Res.* **2009**, *43*, 194–196.
38. Jaturasitha, S.; Kayan, A.; Wicke, M. Carcass and meat characteristics of male chickens between Thai indigenous compared with improved layer breeds and their crossbred. *Arch. Anim. Breed.* **2008**, *51*, 283–294. [[CrossRef](#)]
39. Motsepe, R.; Mabelebele, M.; Norris, D.; Brown, D.; Ngambi, J.; Ginindza, M. Carcass and meat quality characteristics of South African indigenous chickens. *Indian J. Anim. Res.* **2016**, *50*, 580–587. [[CrossRef](#)]
40. Devatkal, S.K.; Vishnuraj, M.R.; Kulkarni, V.V.; Kotaiah, T. Carcass and meat quality characterization of indigenous and improved variety of chicken genotypes. *Poult. Sci.* **2018**, *97*, 2947–2956. [[CrossRef](#)]
41. Kaewkot, C.; Ruangsuriya, J.; Kreuzer, M.; Jaturasitha, S. Carcass and meat quality of crossbreds of Thai indigenous chickens and Rhode Island Red layer chickens as compared with the purebreds and with broilers. *Anim. Prod. Sci.* **2019**, *60*, 454–463. [[CrossRef](#)]
42. Mueller, S.; Kreuzer, M.; Siegrist, M.; Mannale, K.; Messikommer, R.E.; Gangnat, I.D. Carcass and meat quality of dual-purpose chickens (Lohmann Dual, Belgian Malines, Schweizerhuhn) in comparison to broiler and layer chicken types. *Poult. Sci.* **2018**, *97*, 3325–3336. [[CrossRef](#)]
43. Haunshi, S.; Sunitha, R.; Shanmugam, M.; Padhi, M.; Niranjana, M. Carcass characteristics and chemical composition of breast and thigh muscles of native chicken breeds. *Indian J. Poult. Sci.* **2013**, *48*, 219–222.
44. Pripwai, N.; Pattanawong, W.; Punyatong, M.; Teltathum, T. Carcass characteristics and meat quality of Thai inheritance chickens. *J. Agric. Sci.* **2014**, *6*, 182. [[CrossRef](#)]
45. Cassandro, M.; De Marchi, M.; Penasa, M.; Rizzi, C. Carcass characteristics and meat quality traits of the Padovana chicken breed, a commercial line, and their cross. *Ital. J. Anim. Sci.* **2015**, *14*, 3848. [[CrossRef](#)]
46. Jatoi, A.; Iqbal, M.; Sahota, A.; Akram, M.; Javed, K.; Mehmood, S.; Hussain, J.; Ishaq, H. Carcass characteristics and organ development in four different varieties of native Aseel chicken of Pakistan. *Pak. J. Sci.* **2015**, *67*, 127–132.
47. Liu, F.; Niu, Z. Carcass quality of different meat-typed chickens when achieve a common physiological body weight. *Int. J. Poult. Sci.* **2008**, *7*, 319–322. [[CrossRef](#)]
48. Jung, S.; Bae, Y.S.; Kim, H.J.; Jayasena, D.D.; Lee, J.H.; Park, H.B.; Heo, K.N.; Jo, C. Carnosine, anserine, creatine, and inosine 5'-monophosphate contents in breast and thigh meats from 5 lines of Korean native chicken. *Poult. Sci.* **2013**, *92*, 3275–3282. [[CrossRef](#)]
49. Nematbakhsh, S.; Selamat, J.; Idris, L.H.; Abdull Razis, A.F. Chicken Authentication and Discrimination via Live Weight, Body Size, Carcass Traits, and Breast Muscle Fat Content Clustering as Affected by Breed and Sex Varieties in Malaysia. *Foods* **2021**, *10*, 1575. [[CrossRef](#)]
50. Rajkumar, U.; Muthukumar, M.; Haunshi, S.; Niranjana, M.; Raju, M.; Rama Rao, S.; Chatterjee, R. Comparative evaluation of carcass traits and meat quality in native Aseel chickens and commercial broilers. *Br. Poult. Sci.* **2016**, *57*, 339–347. [[CrossRef](#)]
51. Biazen, A.; Mengistu, U.; Negassi, A.; Getenet, A.; Solomon, A.; Tadelles, D. Comparative Growth Performance, Carcass Characteristics and Meat Quality of Local Horro and Exotic Cockerels of Tropical Origin Fed Growers Diet. *Open J. Anim. Sci.* **2021**, *11*, 62–83. [[CrossRef](#)]
52. Jaturasitha, S.; Leangwunta, V.; Leotaragul, A.; Phongphaew, A.; Apichartsrunkoon, T.; Simasathitkul, N.; Veerasilp, T.; Worachai, L.; ter Meulen, U. A comparative study of Thai native chicken and broiler on productive performance, carcass and meat quality. *Dtsch. Trop.* **2002**, *146*, 1–9.
53. Khan, U.; Hussain, J.; Mahmud, A.; Khalique, A.; Mehmood, S.; Badar, I.; Usman, M.; Jaspal, M.; Ahmad, S. Comparative study on carcass traits, meat quality and taste in broiler, broiler breeder and aseel chickens. *Braz. J. Poult. Sci.* **2019**, *21*. [[CrossRef](#)]



54. Promket, D.; Ruangwittayanusorn, K. The comparatives of growth and carcass performance of the Thai native chicken between economic selection (Chee KKU12) and natural selection (Chee N). *Vet. Integr. Sci.* **2021**, *19*, 247–257. [[CrossRef](#)]
55. Tor, M.; Estany, J.; Villalba, D.; Molina, E.; Cubiló, D. Comparison of carcass composition by parts and tissues between cocks and capons. *Anim. Res.* **2002**, *51*, 421–431. [[CrossRef](#)]
56. Sarsenbek, A.; Wang, T.; Zhao, J.; Jiang, W. Comparison of carcass yields and meat quality between Baicheng-You chickens and Arbor Acres broilers. *Poult. Sci.* **2013**, *92*, 2776–2782. [[CrossRef](#)] [[PubMed](#)]
57. Franco, D.; Rois, D.; Vázquez, J.A.; Lorenzo, J. Comparison of growth performance, carcass components, and meat quality between Mos rooster (Galician indigenous breed) and Sasso T-44 line slaughtered at 10 months. *Poult. Sci.* **2012**, *91*, 1227–1239. [[CrossRef](#)] [[PubMed](#)]
58. Wattanachant, S.; Benjakul, S.; Ledward, D.A. Composition, color, and texture of Thai indigenous and broiler chicken muscles. *Poult. Sci.* **2004**, *83*, 123–128. [[CrossRef](#)] [[PubMed](#)]
59. Jaturasitha, S.; Srikanchai, T.; Kreuzer, M.; Wicke, M. Differences in carcass and meat characteristics between chicken indigenous to northern Thailand (Black-boned and Thai native) and imported extensive breeds (Bresse and Rhode Island Red). *Poult. Sci.* **2008**, *87*, 160–169. [[CrossRef](#)]
60. Kwiecień, M.; Kasperek, K.; Tomaszewska, E.; Muszyński, S.; Jeżewska-Witkowska, G.; Winiarska-Mieczan, A.; Grela, E.; Kamińska, E. Effect of breed and caponisation on the growth performance, carcass composition, and fatty acid profile in the muscles of Greenleg Partridge and Polbar breeds. *Braz. J. Poult. Sci.* **2018**, *20*, 583–594. [[CrossRef](#)]
61. Pateiro, M.; Rois, D.; Lorenzo, J.M.; Vázquez, J.A.; Franco, D. Effect of breed and finishing diet on growth performance, carcass and meat quality characteristics of Mos young hens. *Span. J. Agric. Res.* **2018**, *16*, e0402. [[CrossRef](#)]
62. Puchała, M.; Krawczyk, J.; Sokołowicz, Z.; Utnik-Banaś, K. Effect of breed and production system on physicochemical characteristics of meat from multi-purpose hens. *Ann. Anim. Sci.* **2015**, *15*, 247–261. [[CrossRef](#)]
63. Tougan, P.; Dahouda, M.; Ahounou, G.; Salifou, C.; Kpodekon, M.; Mensah, G.; Kossou, D.; Amenou, C.; Kogbeto, C.; Thewis, A. Effect of breeding mode, type of muscle and slaughter age on technological meat quality of local poultry population of *Gallus gallus* species of Benin. *Int. J. Biosci.* **2013**, *3*, 1–17.
64. Miguel, J.; Ciria, J.; Asenjo, B.; Calvo, J. Effect of caponisation on growth and on carcass and meat characteristics in Castellana Negra native Spanish chickens. *Animal* **2008**, *2*, 305–311. [[CrossRef](#)] [[PubMed](#)]
65. Calik, J.; Poltowicz, K.; Swiatkiewicz, S.; Krawczyk, J.; Nowak, J. Effect of caponization on meat quality of Greenleg Partridge cockerels. *Ann. Anim. Sci.* **2015**, *15*, 541. [[CrossRef](#)]
66. Durán, A.M. The effect of caponization on production indices and carcass and meat characteristics in free-range Extremeña Azul chickens. *Span. J. Agric. Res.* **2004**, *2*, 211–216. [[CrossRef](#)]
67. Jiang, R.; Zhao, G.; Chen, J.; Zheng, M.; Zhao, J.; Li, P.; Hu, J.; Wen, J. Effect of dietary supplemental nicotinic acid on growth performance, carcass characteristics and meat quality in three genotypes of chicken. *J. Anim. Physiol. Anim. Nutr.* **2011**, *95*, 137–145. [[CrossRef](#)]
68. Sosnowka-Czajka, E.; Skomorucha, I.; Muchacka, R. Effect of organic production system on the performance and meat quality of two purebred slow-growing chicken breeds. *Ann. Anim. Sci.* **2017**, *17*, 1197. [[CrossRef](#)]
69. Jiang, M.; Fan, W.; Xing, S.; Wang, J.; Li, P.; Liu, R.; Li, Q.; Zheng, M.; Cui, H.; Wen, J. Effects of balanced selection for intramuscular fat and abdominal fat percentage and estimates of genetic parameters. *Poult. Sci.* **2017**, *96*, 282–287. [[CrossRef](#)]
70. Franco, D.; Pateiro, M.; Rois, D.; Vázquez, J.A.; Lorenzo, J.M.; Rodriguez, J. Effects of caponization on growth performance, carcass and meat quality of Mos breed capons reared in free-range production system. *Ann. Anim. Sci.* **2016**, *16*, 909–929. [[CrossRef](#)]
71. Guo, X.; Nan, H.; Shi, D.; Zhou, J.; Wan, Y.; Zhou, B.; Geng, Z.; Chen, X.; Jiang, R. Effects of caponization on growth, carcass, and meat characteristics and the mRNA expression of genes related to lipid metabolism in roosters of a Chinese indigenous breed. *Czech J. Anim. Sci.* **2015**, *60*, 327–333. [[CrossRef](#)]
72. Wang, D.; Huang, H.; Zhou, L.; Li, W.; Zhou, H.; Hou, G.; Liu, J.; Hu, L. Effects of dietary supplementation with turmeric rhizome extract on growth performance, carcass characteristics, antioxidant capability, and meat quality of Wenchang broiler chickens. *Ital. J. Anim. Sci.* **2015**, *14*, 3870. [[CrossRef](#)]
73. Zhao, J.; Zhao, G.; Jiang, R.; Zheng, M.; Chen, J.; Liu, R.; Wen, J. Effects of diet-induced differences in growth rate on metabolic, histological, and meat-quality properties of 2 muscles in male chickens of 2 distinct broiler breeds. *Poult. Sci.* **2012**, *91*, 237–247. [[CrossRef](#)] [[PubMed](#)]
74. Khatun, H.; Faruqe, S.; Mostafa, M.G. Effects of different dietary energy and protein levels on the performance and carcass characteristics of native hilly chicken during growing phase in confinement. *Asian Australas. J. Biosci. Biotechnol.* **2021**, *6*, 1–9. [[CrossRef](#)]
75. Cheng, F.-Y.; Huang, C.; Wan, T.-C.; Liu, Y.-T.; Lin, L.; Lou Chyr, C.-Y. Effects of free-range farming on carcass and meat qualities of black-feathered Taiwan native chicken. *Asian Australas. J. Anim. Sci.* **2008**, *21*, 1201–1206. [[CrossRef](#)]
76. Bughio, E.; Hussain, J.; Mahmud, A.; Khalique, A. Effects of production system and feeding regimen on carcass and meat quality traits of Naked Neck chicken. *S. Afr. J. Anim. Sci.* **2021**, *51*, 250–261. [[CrossRef](#)]
77. Chen, J.; Zhao, G.; Zheng, M.; Wen, J.; Yang, N. Estimation of genetic parameters for contents of intramuscular fat and inosine-5'-monophosphate and carcass traits in Chinese Beijing-You chickens. *Poult. Sci.* **2008**, *87*, 1098–1104. [[CrossRef](#)] [[PubMed](#)]
78. Yousif, I.; Binda, B.; Elamin, K.; Malik, H.; Babiker, M. Evaluation of carcass characteristics and meat quality of indigenous fowl ecotypes and exotic broiler strains raised under hot climate. *Glob. J. Anim. Sci.* **2014**, *2*, 365–371.

79. Rajkumar, U.; Prince, L.; Haunshi, S.; Paswan, C.; Reddy, B. Evaluation of Vanaraja female line chicken for growth, production, carcass and egg quality traits. *Indian J. Anim. Sci.* **2020**, *90*, 603–609.
80. Cerolini, S.; Vasconi, M.; Sayed, A.A.; Iaffaldano, N.; Mangiagalli, M.G.; Pastorelli, G.; Moretti, V.M.; Zaniboni, L.; Mosca, F. Free-range rearing density for male and female Milanino chickens: Carcass yield and qualitative meat traits. *J. Appl. Poult. Res.* **2019**, *28*, 1349–1358. [[CrossRef](#)]
81. Molee, A.; Kuadsantia, P.; Kaewnakian, P. Gene effects on body weight, carcass yield, and meat quality of Thai indigenous chicken. *J. Poult. Sci.* **2018**, *55*, 94–102. [[CrossRef](#)]
82. Bungsisawat, P.; Tumwasorn, S.; Loongyai, W.; Nakthong, S.; Sopannarath, P. Genetic parameters of some carcass and meat quality traits in Betong chicken (KU line). *Agric. Nat. Resour.* **2018**, *52*, 274–279. [[CrossRef](#)]
83. Peters, S.O.; Idowu, O.M.; Agaviezor, B.O.; Egbede, R.O.; Fafiolu, A.O. Genotype and sex effect on gastrointestinal nutrient content, microflora and carcass traits in Nigerian native chickens. *Int. J. Poult. Sci.* **2010**, *9*, 731–737. [[CrossRef](#)]
84. Franco, D.; Rois, D.; Vázquez, J.A.; Lorenzo, J. Growth performance, carcass morphology and meat quality of meat from roosters 1 slaughtered at eight months affected by genotype and finishing feeding. *Span. J. Agric. Res.* **2013**, *11*, 382–393. [[CrossRef](#)]
85. Nolte, T.; Jansen, S.; Weigend, S.; Moerlein, D.; Halle, I.; Link, W.; Hummel, J.; Simianer, H.; Sharifi, A.R. Growth performance of local chicken breeds, a high-performance genotype and their crosses fed with regional faba beans to replace soy. *Animals* **2020**, *10*, 702. [[CrossRef](#)]
86. Keambou, T.; Mboumba, S.; Touko, B.; Bembide, C.; Mezui, T.; Tedongmo, A.; Manjeli, Y. Growth performances, carcass and egg characteristics of the local chicken and its first generation reciprocal crossbreds with an exotic strain in Cameroon. *Adv. Anim. Vet. Sci.* **2015**, *3*, 507–513. [[CrossRef](#)]
87. Paredes, M.; Vásquez, B. Growth, carcass characteristics, weight of internal organs and meat proximate composition of six genotypes in chickens reared in Andean region of northern Peruvian. *Sci. Agropecu.* **2020**, *11*, 365–374. [[CrossRef](#)]
88. Tsudzuki, M.; Onitsuka, S.; Akiyama, R.; Iwamizu, M.; Goto, N.; Nishibori, M.; Takahashi, H.; Ishikawa, A. Identification of quantitative trait loci affecting shank length, body weight and carcass weight from the Japanese cockfighting chicken breed, Oh-Shamo (Japanese Large Game). *Cytogenet. Genome Res.* **2007**, *117*, 288–295. [[CrossRef](#)] [[PubMed](#)]
89. Magala, H.; Kugonza, D.; Kwizera, H.; Kyarisiima, C. Influence of management system on growth and carcass characteristics of Ugandan local chickens. *J. Anim. Sci. Adv.* **2012**, *2*, 558–567.
90. Kasperek, K.; Drabik, K.; Miachalak, K.; Pietras-Ożga, D.; Winiarczyk, S.; Zięba, G.; Batkowska, J. The Influence of Sex on the Slaughter Parameters and Selected Blood Indices of Greenleg Partridge, Polish Native Breed of Hens. *Animals* **2021**, *11*, 517. [[CrossRef](#)]
91. Haunshi, S.; Paswan, C.; Prince, L.; Chatterjee, R. Inheritance of growth traits and impact of selection on carcass and egg quality traits in Vanashree, an improved indigenous chicken. *Trop. Anim. Health Prod.* **2021**, *53*, 1–8. [[CrossRef](#)]
92. Zhao, J.; Chen, J.; Zhao, G.; Zheng, M.; Jiang, R.; Wen, J. Live performance, carcass composition, and blood metabolite responses to dietary nutrient density in two distinct broiler breeds of male chickens. *Poult. Sci.* **2009**, *88*, 2575–2584. [[CrossRef](#)]
93. Obrzut, J.; Krawczyk, J.; Calik, J.; Świątkiewicz, S.; Pietras, M.; Utnik-Banaś, K. Meat quality of poulards obtained from three conserved breeds of hens. *Ann. Anim. Sci.* **2018**, *18*, 261. [[CrossRef](#)]
94. Liu, L.; Dou, T.; Li, Q.; Rong, H.; Tong, H.; Xu, Z.; Huang, Y.; Gu, D.; Chen, X.; Ge, C. Myostatin mRNA expression and its association with body weight and carcass traits in Yunnan Wuding chicken. *Genet. Mol. Res.* **2016**, *15*, gmr15048967. [[CrossRef](#)] [[PubMed](#)]
95. Pavlovski, Z.; Škrbić, Z.; Lukić, M.; Vitorović, D.; Petričević, V.; Milošević, N. Naked Neck chicken of Serbian and foreign origin: Carcass characteristic. *Biotechnol. Anim. Husb.* **2009**, *25*, 1023–1032. [[CrossRef](#)]
96. Pavlovski, Z.; Škrbić, Z.; Lukić, M.; Vitorović, D.; Petričević, V. Naked neck: Autochthonous breed of chicken in Serbia: Carcass characteristics. *Biotechnol. Anim. Husb.* **2009**, *25*, 1–10. [[CrossRef](#)]
97. Lariviere, J.; Farnir, F.; Dettleux, J.; Michaux, C.; Verleyen, V.; Leroy, P. Performance, breast morphological and carcass traits in the Ardennaise chicken breed. *Int. J. Poult. Sci.* **2009**, *8*, 452–456. [[CrossRef](#)]
98. Raach-Moujahed, A.; Haddad, B. Performance, livability, carcass yield and meat quality of Tunisian local poultry and fast-growing genotype (Arbor Acres) fed standard diet and raised outdoor access. *J. Anim. Prod. Adv.* **2013**, *3*, 75–85. [[CrossRef](#)]
99. Amorim, A.; Rodrigues, S.; Pereira, E.; Teixeira, A. Physicochemical composition and sensory quality evaluation of capon and rooster meat. *Poult. Sci.* **2016**, *95*, 1211–1219. [[CrossRef](#)]
100. Zhang, L.; Zhu, Q.; Liu, Y.; Gilbert, E.R.; Li, D.; Yin, H.; Wang, Y.; Yang, Z.; Wang, Z.; Yuan, Y. Polymorphisms in the perilipin gene may affect carcass traits of Chinese meat-type chickens. *Asian Australas. J. Anim. Sci.* **2015**, *28*, 763. [[CrossRef](#)] [[PubMed](#)]
101. Tasoniero, G.; Cullere, M.; Baldan, G.; Dalle Zotte, A. Productive performances and carcass quality of male and female Italian Padovana and Polverara slow-growing chicken breeds. *Ital. J. Anim. Sci.* **2018**, *17*, 530–539. [[CrossRef](#)]
102. Dalle Zotte, A.; Gleeson, E.; Franco, D.; Cullere, M.; Lorenzo, J.M. Proximate composition, amino acid profile, and oxidative stability of slow-growing indigenous chickens compared with commercial broiler chickens. *Foods* **2020**, *9*, 546. [[CrossRef](#)]
103. Mosca, F.; Zaniboni, L.; Stella, S.; Kuster, C.; Iaffaldano, N.; Cerolini, S. Slaughter performance and meat quality of Milanino chickens reared according to a specific free-range program. *Poult. Sci.* **2018**, *97*, 1148–1154. [[CrossRef](#)] [[PubMed](#)]
104. Węglarz, A.; Andres, K.; Wojtyśiak, D. Slaughter value and meat quality in two strains of polish crested cockerels. *Ital. J. Anim. Sci.* **2020**, *19*, 813–821. [[CrossRef](#)]

105. Shakila, S.; Bhaskar Reddy, G.V.; Amaravathi, P. Studies on carcass and meat quality characteristics of Rajasri chicken. *J. Entomol. Zool. Stud.* **2020**, *8*, 1345–1349.
106. Pathak, P.; Dubey, P.; Dash, S.; Chaudhary, M. Studies on growth and carcass traits of Aseel and Kadaknath chicken. *Indian J. Poult. Sci.* **2015**, *50*, 327–328.
107. Hussain, M.; Mahmud, A.; Hussain, J.; Qaisrani, S.; Mehmood, S.; Rehman, A. Subsequent effect of dietary lysine regimens fed in the starter phase on the growth performance, carcass traits and meat chemical composition of aseel chicken in the grower phase. *Braz. J. Poult. Sci.* **2018**, *20*, 455–462. [[CrossRef](#)]
108. Tougan, U.; Dahouda, M.; Salifou, C.; Ahounou, S.; Kpodekon, M.; Mensah, G.; Kossou, D.; Amenou, C.; Kogbeto, C.; Thewis, A. Variability of carcass traits of local poultry populations of *Gallus gallus* species of Benin. *Int. J. Poult. Sci.* **2013**, *12*, 473. [[CrossRef](#)]
109. Tang, H.; Gong, Y.; Wu, C.; Jiang, J.; Wang, Y.; Li, K. Variation of meat quality traits among five genotypes of chicken. *Poult. Sci.* **2009**, *88*, 2212–2218. [[CrossRef](#)]
110. Toalombo, P.; Villafuerte, A.; Fiallos, L.; Andino, P.; Damián, P.; Duchi, N.; Trujillo, V.; Hidalgo, L. Polyphenols of Thyme (*Thymus vulgaris*) and ginger (*Zingiber officinale*) in the feeding of local hens. *Actas Iberoam. Conserv. Anim.* **2017**, *10*, 88–93.
111. Chuaynukool, K.; Wattanachant, S.; Siripongvutikorn, S.; Yai, H. Chemical and physical properties of raw and cooked spent hen, broiler and Thai indigenous chicken muscles in mixed herbs acidified soup (Tom Yum). *J. Food Technol.* **2007**, *5*, 180–186.
112. El-Attrouny, M.M.; Iraqi, M.M.; Sabike, I.I.; Abdelatty, A.M.; Moustafa, M.M.; Badr, O.A. Comparative evaluation of growth performance, carcass characteristics and timed series gene expression profile of GH and IGF-1 in two Egyptian indigenous chicken breeds versus Rhode Island Red. *J. Anim. Breed. Genet.* **2021**, *138*, 463–473. [[CrossRef](#)]
113. Jung, Y.-K.; Jeon, H.-J.; Jung, S.; Choe, J.-H.; Lee, J.-H.; Heo, K.-N.; Kang, B.-S.; Jo, C.-R. Comparison of quality traits of thigh meat from Korean native chickens and broilers. *Food Sci. Anim. Resour.* **2011**, *31*, 684–692. [[CrossRef](#)]
114. Amorim, A.; Rodrigues, S.; Pereira, E.; Valentim, R.; Teixeira, A. Effect of caponisation on physicochemical and sensory characteristics of chickens. *Animal* **2016**, *10*, 978–986. [[CrossRef](#)] [[PubMed](#)]
115. Ramella, M.V.; Rodríguez, J.M.L.; Losada, D.R.; Arias, A.; Justo, J.R.; Moure, M.P.; Pedrouso, M.D.L.; Chico, D. Effect of finishing diet on carcass characteristics and meat quality of Mos cockerel. *Span. J. Agric. Res.* **2021**, *19*, 601.
116. Mosca, F.; Kuster, C.; Stella, S.; Farina, G.; Madeddu, M.; Zaniboni, L.; Cerolini, S. Growth performance, carcass characteristics and meat composition of Milanino chickens fed on diets with different protein concentrations. *Br. Poult. Sci.* **2016**, *57*, 531–537. [[CrossRef](#)] [[PubMed](#)]
117. Jin, S.; Yang, L.; Zang, H.; Xu, Y.; Chen, X.; Chen, X.; Liu, P.; Geng, Z. Influence of free-range days on growth performance, carcass traits, meat quality, lymphoid organ indices, and blood biochemistry of Wannan Yellow chickens. *Poult. Sci.* **2019**, *98*, 6602–6610. [[CrossRef](#)] [[PubMed](#)]
118. Escobedo del Bosque, C.I.; Altmann, B.A.; Ciulu, M.; Halle, I.; Jansen, S.; Nolte, T.; Weigend, S.; Mörlein, D. Meat quality parameters and sensory properties of one high-performing and two local chicken breeds fed with *Vicia faba*. *Foods* **2020**, *9*, 1052. [[CrossRef](#)] [[PubMed](#)]
119. Gnanaraj, P.T.; Sundaram, A.S.; Rajkumar, K.; Babu, R.N. Proximate composition and meat quality of three indian native chicken breeds. *Indian J. Anim. Res.* **2020**, *54*, 1584–1589. [[CrossRef](#)]
120. Jeong, H.S.; Utama, D.T.; Kim, J.; Barido, F.H.; Lee, S.K. Quality comparison of retorted Samgyetang made from white semi-broilers, commercial broilers, Korean native chickens, and old laying hens. *Asian Australas. J. Anim. Sci.* **2020**, *33*, 139. [[CrossRef](#)]
121. Chumngoen, W.; Tan, F.-J. Relationships between descriptive sensory attributes and physicochemical analysis of broiler and Taiwan native chicken breast meat. *Asian Australas. J. Anim. Sci.* **2015**, *28*, 1028. [[CrossRef](#)]
122. Elkhazen, A.; LARBI, M.; M’hamdi, N.; Haddad, B. Comparison of meat quality of local poultry and Arbors acres reared in two farming systems in Tunisia. *J. New Sci.* **2016**, *34*, 1922–1929.
123. Zhao, G.; Cui, H.; Liu, R.; Zheng, M.; Chen, J.; Wen, J. Comparison of breast muscle meat quality in 2 broiler breeds. *Poult. Sci.* **2011**, *90*, 2355–2359. [[CrossRef](#)] [[PubMed](#)]
124. Rajkumar, U.; Haunshi, S.; Paswan, C.; Raju, M.; Rao, S.R.; Chatterjee, R. Characterization of indigenous Aseel chicken breed for morphological, growth, production, and meat composition traits from India. *Poult. Sci.* **2017**, *96*, 2120–2126. [[CrossRef](#)] [[PubMed](#)]
125. Youssao, I.; Alkoiret, I.; Dahouda, M.; Assogba, M.; Idrissou, N.; Kayang, B.; Yapi-Gnaoré, V.; Assogba, H.; Houinsou, A.; Ahounou, S. Comparison of growth performance, carcass characteristics and meat quality of Benin indigenous chickens and Label Rouge (T55×SA51). *Afr. J. Biotechnol.* **2012**, *11*, 15569–15579.
126. Batool, T.; Farooq, S.; Roohi, N.; Mahmud, A.; Usman, M.; Ghayas, A.; Ahmad, S. Effect of different dietary lysine regimens on meat quality attributes in varieties of indigenous Aseel chicken. *Kafkas Univ. Vet. Fak. Derg.* **2018**, *24*, 639–645.
127. Tanim, S.; Phasuk, Y.; Aggrey, S.E.; Duangjinda, M. Gene expression of fatty acid binding protein genes and its relationship with fat deposition of Thai native crossbreed chickens. *Anim. Biosci.* **2021**, *34*, 751–758. [[CrossRef](#)] [[PubMed](#)]
128. Lorenzen, C.; Calkins, C.; Green, M.; Miller, R.; Morgan, J.; Wasser, B. Efficacy of performing Warner–Bratzler and slice shear force on the same beef steak following rapid cooking. *Meat Sci.* **2010**, *85*, 792–794. [[CrossRef](#)]
129. Arantes-Pereira, L.; Vargas, F.C.; Balieiro, J.C.; Bittante, A.M.Q.; Sobral, P.J. Reproducibility and correlation between meat shear force measurements by Warner–Bratzler machine and a Texturometer. *Int. J. Food Stud.* **2016**, *5*, 193. [[CrossRef](#)]
130. González Ariza, A.; Arando Arbulu, A.; Navas González, F.J.; Delgado Bermejo, J.V.; Camacho Vallejo, M.E. Discriminant Canonical Analysis as a Validation Tool for Multivariety Native Breed Egg Commercial Quality Classification. *Foods* **2021**, *10*, 632. [[CrossRef](#)]

131. Rogerson, P.A. *Data Reduction: Factor Analysis and Cluster Analysis*; Sage: London, UK, 2001; pp. 192–197.
132. Nanda, M.A.; Seminar, K.B.; Nandika, D.; Maddu, A. Discriminant analysis as a tool for detecting the acoustic signals of termites *Coptotermes curvignathus* (Isoptera: Rhinotermitidae). *Int. J. Technol.* **2018**, *9*, 840–851. [[CrossRef](#)]
133. González Ariza, A.; Arando Arbulu, A.; León Jurado, J.M.; Navas González, F.J.; Delgado Bermejo, J.V.; Camacho Vallejo, M.E. Discriminant Canonical Tool for Differential Biometric Characterization of Multivariety Endangered Hen Breeds. *Animals* **2021**, *11*, 2211. [[CrossRef](#)]
134. Breimann, L.; Friedman, J.H.; Olshen, R.A.; Stone, C.J. *Classification and Regression Trees*; Taylor & Francis: New York, NY, USA, 2017.
135. Babicki, S.; Arndt, D.; Marcu, A.; Liang, Y.; Grant, J.R.; Maciejewski, A.; Wishart, D.S. Heatmapper: Web-enabled heat mapping for all. *Nucleic Acids Res.* **2016**, *44*, 147–153. [[CrossRef](#)] [[PubMed](#)]
136. Albayrak, A.S. Classification of domestic and foreign commercial banks in Turkey based on financial performances using linear discriminant analysis, logistic regression and artificial neural network models. *Suleyman Demirel Univ. J. Fac. Econ. Adm. Sci.* **2009**, *14*, 113–139.
137. Leroy, G.; Baumung, R.; Notter, D.; Verrier, E.; Wurzinger, M.; Scherf, B. Stakeholder involvement and the management of animal genetic resources across the world. *Livest. Sci.* **2017**, *198*, 120–128. [[CrossRef](#)]
138. Biscarini, F.; Nicolazzi, E.L.; Stella, A.; Boettcher, P.J.; Gandini, G. Challenges and opportunities in genetic improvement of local livestock breeds. *Front. Genet.* **2015**, *6*, 33. [[CrossRef](#)]
139. Lordelo, M.; Cid, J.; Cordovil, C.M.; Alves, S.P.; Bessa, R.J.; Carolino, I. A comparison between the quality of eggs from indigenous chicken breeds and that from commercial layers. *Poult. Sci.* **2020**, *99*, 1768–1776. [[CrossRef](#)]
140. González Ariza, A.; Navas González, F.J.; Arando Arbulu, A.; León Jurado, J.M.; Delgado Bermejo, J.V.; Camacho Vallejo, M.E. Variability of Meat and Carcass Quality from Worldwide Native Chicken Breeds. *Foods* **2022**, *11*, 1700. [[CrossRef](#)]
141. Fitzgerald, E.; Buckley, J. Effect of total and partial substitution of sodium chloride on the quality of Cheddar cheese. *J. Dairy Sci.* **1985**, *68*, 3127–3134. [[CrossRef](#)]
142. Wee, M.S.M.; Goh, A.T.; Stieger, M.; Forde, C.G. Correlation of instrumental texture properties from textural profile analysis (TPA) with eating behaviours and macronutrient composition for a wide range of solid foods. *Food Funct.* **2018**, *9*, 5301–5312. [[CrossRef](#)]
143. Xu, H.; Dong, X.; Zuo, R.; Mai, K.; Ai, Q. Response of juvenile Japanese seabass (*Lateolabrax japonicus*) to different dietary fatty acid profiles: Growth performance, tissue lipid accumulation, liver histology and flesh texture. *Aquaculture* **2016**, *461*, 40–47. [[CrossRef](#)]
144. Johnston, I.A.; Alderson, R.; Sandham, C.; Dingwall, A.; Mitchell, D.; Selkirk, C.; Nickell, D.; Baker, R.; Robertson, B.; Whyte, D. Muscle fibre density in relation to the colour and texture of smoked Atlantic salmon (*Salmo salar* L.). *Aquaculture* **2000**, *189*, 335–349. [[CrossRef](#)]
145. Jeon, H.J.; Choe, J.H.; Jung, Y.K.; Kruk, Z.A.; Lim, D.G.; Jo, C.R. Comparison of the chemical composition, textural characteristics, and sensory properties of North and South Korean native chickens and commercial broilers. *Food Sci. Anim. Resour.* **2010**, *30*, 171–178. [[CrossRef](#)]
146. Jin, S.; Jayasena, D.; Jo, C.; Lee, J. The breeding history and commercial development of the Korean native chicken. *Worlds Poult. Sci. J.* **2017**, *73*, 163–174. [[CrossRef](#)]
147. Holman, B.W.B.; Hopkins, D.L. The use of conventional laboratory-based methods to predict consumer acceptance of beef and sheep meat: A review. *Meat Sci.* **2021**, *181*, 108586. [[CrossRef](#)]
148. Zheng, L.; Teng, F.; Wang, N.; Zhang, X.-N.; Regenstein, J.M.; Liu, J.-S.; Li, Y.; Wang, Z.-J. Addition of Salt Ions before Spraying Improves Heat- and Cold-Induced Gel Properties of Soy Protein Isolate (SPI). *Appl. Sci.* **2019**, *9*, 1076. [[CrossRef](#)]
149. Marelli, S.P.; Zaniboni, L.; Strillacci, M.G.; Madeddu, M.; Cerolini, S. Morphological Characterization of Two Light Italian Turkey Breeds. *Animals* **2022**, *12*, 571. [[CrossRef](#)] [[PubMed](#)]
150. Neethling, N.E.; Suman, S.P.; Sigge, G.O.; Hoffman, L.C.; Hunt, M.C. Exogenous and endogenous factors influencing color of fresh meat from ungulates. *Meat Muscle Biol.* **2017**, *1*, 253. [[CrossRef](#)]
151. Moon, S.; Yang, H.; Park, G.; Joo, S. The relationship of physiological maturity and marbling judged according to Korean grading system to meat quality traits of Hanwoo beef females. *Meat Sci.* **2006**, *74*, 516–521. [[CrossRef](#)]
152. Gagaoua, M.; Picard, B.; Monteils, V. Associations among animal, carcass, muscle characteristics, and fresh meat color traits in Charolais cattle. *Meat Sci.* **2018**, *140*, 145–156. [[CrossRef](#)]
153. Uhlířová, L.; Tůmová, E.; Chodová, D.; Vlčková, J.; Ketta, M.; Volek, Z.; Skřivanová, V. The effect of age, genotype and sex on carcass traits, meat quality and sensory attributes of geese. *Asian Australas. J. Anim. Sci.* **2018**, *31*, 421–428. [[CrossRef](#)]
154. Tong, H.; Lu, J.; Zou, J.; Wang, Q.; Shi, S. Effects of stocking density on growth performance, carcass yield, and immune status of a local chicken breed. *Poult. Sci.* **2012**, *91*, 667–673. [[CrossRef](#)]
155. Wang, K.; Shi, S.; Dou, T.; Sun, H. Effect of a free-range raising system on growth performance, carcass yield, and meat quality of slow-growing chicken. *Poult. Sci.* **2009**, *88*, 2219–2223. [[CrossRef](#)]
156. Tilki, M.; Saatci, M.; Kirmizibayrak, T.; Aksoy, A. Effect of age on growth and carcass composition of Native Turkish Geese. *Arch. Geflügelkd.* **2005**, *69*, 77–83.
157. Park, S.-Y.; Byeon, D.-S.; Kim, G.-W.; Kim, H.-Y. Carcass and retail meat cuts quality properties of broiler chicken meat based on the slaughter age. *J. Anim. Sci. Technol.* **2021**, *63*, 180. [[CrossRef](#)] [[PubMed](#)]

158. Husak, R.; Sebranek, J.; Bregendahl, K. A survey of commercially available broilers marketed as organic, free-range, and conventional broilers for cooked meat yields, meat composition, and relative value. *Poult. Sci.* **2008**, *87*, 2367–2376. [[CrossRef](#)] [[PubMed](#)]
159. Siekmann, L.; Meier-Dinkel, L.; Janisch, S.; Altmann, B.; Kaltwasser, C.; Sürle, C.; Krischek, C. Carcass Quality, Meat Quality and Sensory Properties of the Dual-Purpose Chicken Lohmann Dual. *Foods* **2018**, *7*, 156. [[CrossRef](#)] [[PubMed](#)]
160. Shi, K.; Zhao, Q.; Shao, M.; Duan, Y.; Li, D.; Lu, Y.; Tang, Y.; Feng, C. Untargeted Metabolomics Reveals the Effect of Selective Breeding on the Quality of Chicken Meat. *Metabolites* **2022**, *12*, 367. [[CrossRef](#)] [[PubMed](#)]
161. Castellini, C.; Dal Bosco, A.; Mugnai, C.; Pedrazzoli, M. Comparison of two chicken genotypes organically reared: Oxidative stability and other qualitative traits of the meat. *Ital. J. Anim. Sci.* **2006**, *5*, 29–42. [[CrossRef](#)]
162. Petersen, J.; Henckel, P.; Maribo, H.; Oksbjerg, N.; Sørensen, M. Muscle metabolic traits, post mortem-pH-decline and meat quality in pigs subjected to regular physical training and spontaneous activity. *Meat Sci.* **1997**, *46*, 259–275. [[CrossRef](#)]
163. Castellini, C.; Mugnai, C.; Dal Bosco, A. Meat quality of three chicken genotypes reared according to the organic system. *Ital. J. Food Sci.* **2002**, *14*, 411–412.
164. Youssao, A.; Senou, M.; Dahouda, M.; Kpodekon, T.; Djenontin, J.; Idrissou, N.; Bonou, G.; Tougan, U.; Ahounou, S.; Assogba, H. Genetic improvement of local chickens by crossing with the Label Rouge (T55XSA51): Carcass Characteristic, Organoleptic Qualities and Heterosis Effects. *Int. J. Poult. Sci.* **2009**, *8*, 626–633. [[CrossRef](#)]
165. Poltowicz, K. Effect of slaughter age on performance and meat quality of slow-growing broiler chickens. *Ann. Anim. Sci.* **2012**, *12*, 621. [[CrossRef](#)]
166. Mbah, R.E.; Wasum, D.F. Russian-Ukraine 2022 War: A review of the economic impact of Russian-Ukraine crisis on the USA, UK, Canada, and Europe. *Adv. Soc. Sci. Res. J.* **2022**, *9*, 144–153. [[CrossRef](#)]
167. Glauben, T.; Svanidze, M.; Götz, L.; Prehn, S.; Jamali Jaghdani, T.; Đurić, I.; Kuhn, L. The War in Ukraine, Agricultural Trade and Risks to Global Food Security. *Intereconomics* **2022**, *57*, 157–163. [[CrossRef](#)]
168. Coutinho, P.; Simões, M.; Pereira, C.; Paiva, T. Sustainable local exploitation and innovation on meat products based on the autochthonous bovine breed jarmelista. *Sustainability* **2021**, *13*, 2515. [[CrossRef](#)]
169. Hu, Y.; Cheng, H.; Tao, S. Environmental and human health challenges of industrial livestock and poultry farming in China and their mitigation. *Environ. Int.* **2017**, *107*, 111–130. [[CrossRef](#)]
170. Li, J.; Yang, C.; Peng, H.; Yin, H.; Wang, Y.; Hu, Y.; Yu, C.; Jiang, X.; Du, H.; Li, Q.; et al. Effects of Slaughter Age on Muscle Characteristics and Meat Quality Traits of Da-Heng Meat Type Birds. *Animals* **2020**, *10*, 69. [[CrossRef](#)]
171. Baéza, E.; Arnould, C.; Jlali, M.; Chartrin, P.; Gigaud, V.; Mercierand, F.; Durand, C.; Meteau, K.; Le Bihan-Duval, E.; Berri, C. Influence of increasing slaughter age of chickens on meat quality, welfare, and technical and economic results. *J. Anim. Sci.* **2012**, *90*, 2003–2013. [[CrossRef](#)]
172. McGilchrist, P.; Greenwood, P.; Pethick, D.; Gardner, G. Selection for increased muscling in Angus cattle did not increase the glycolytic potential or negatively impact pH decline, retail colour stability or mineral content. *Meat Sci.* **2016**, *114*, 8–17. [[CrossRef](#)]
173. Gao, J.; Lin, H.; Song, Z.; Jiao, H. Corticosterone alters meat quality by changing pre-and postslaughter muscle metabolism. *Poult. Sci.* **2008**, *87*, 1609–1617. [[CrossRef](#)]
174. Xie, X.; Meng, Q.; Cui, Z.; Ren, L. Effect of cattle breed on meat quality, muscle fiber characteristics, lipid oxidation and fatty acids in China. *Asian Australas. J. Anim. Sci.* **2012**, *25*, 824. [[CrossRef](#)]
175. Tougan, P.U.; Dahouda, M.; Salifou, C.F.A.; Ahounou, S.G.A.; Kpodekon, M.T.; Mensah, G.A.; Thewis, A.; Karim, I. Conversion of chicken muscle to meat and factors affecting chicken meat quality: A review. *Int. J. Agron. Agric. Res.* **2013**, *3*, 1–20.
176. Lonergan, S.; Deeb, N.; Fedler, C.; Lamont, S. Breast meat quality and composition in unique chicken populations. *Poult. Sci.* **2003**, *82*, 1990–1994. [[CrossRef](#)] [[PubMed](#)]
177. Panpipat, W.; Chaijan, M.; Karnjanapratum, S.; Keawtong, P.; Tansakul, P.; Panya, A.; Phonsatta, N.; Aoumtes, K.; Quan, T.H.; Petcharat, T. Quality Characterization of Different Parts of Broiler and Ligor Hybrid Chickens. *Foods* **2022**, *11*, 1929. [[CrossRef](#)]
178. Katemala, S.; Molee, A.; Thumanu, K.; Yongsawatdigul, J. Meat quality and Raman spectroscopic characterization of Korat hybrid chicken obtained from various rearing periods. *Poult. Sci.* **2021**, *100*, 1248–1261. [[CrossRef](#)] [[PubMed](#)]
179. Swatland, H. How pH causes paleness or darkness in chicken breast meat. *Meat Sci.* **2008**, *80*, 396–400. [[CrossRef](#)]
180. Fletcher, D. Poultry meat quality. *Worlds Poult. Sci. J.* **2002**, *58*, 131–145. [[CrossRef](#)]