

Supplementary of

Environmental and Oceanographic Conditions at the Continental Margin of the Central Basin, Northwestern Ross Sea (Antarctica) Since the Last Glacial Maximum

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Supplementary S1

Grain size data

In this section the box cores are described from the grain size point of view. The grain size raw data and the assigned clusters are reported in Table S1.

BC02

Silt is the prevailing fraction (avg 76% min 53, max 86%), while the sand content ranges from 7 to 43% with a mean value of 18%. Clay content is low and ranges between 4 and 9% with a mean value of 6%. The sediment is poorly to very poorly sorted (σ_i avg: 1.94 Φ). The skewness varies from near symmetrical to positive skewed (SK avg: 0.08) and the mean diameter (Mz) varies from 4.56 to 6.47 Φ (Mz avg 5.75 Φ , coarse silt). Sediments are mainly bimodal with primary modal diameter ranging between 4 and 7 Φ , and secondary modal diameter around 2 Φ .

BC03

Silt is the prevailing fraction ranging between 56 and 88%, with a mean content of 75.5%. Medium sand content is 18% and varies from 6 and 41%. The clay content is low (between 4 and 6%; avg: 6%). The sediment is poor sorted (σ_i avg: 1.86 Φ) and positive skewness prevails (SK avg: 0.19). The mean Mz is 5.69 Φ —coarse silt (min 4.71 Φ , max 6.27 Φ). Samples are mainly bimodal with a primary mode ranging from 4 to 6 Φ . Only one sample has the primary mode at 7 Φ (24–25 cm). The secondary mode is at 1.50 Φ .

BC04

Silt is the prevalent fraction (avg 77%, min 65% max 83%), sand varies from 11 to 31% (avg 17%) and clay ranges between 3 and 7% (avg 6%). Sediment is poor to very poor sorted (from 1.58 to 2.11 Φ —avg 1.86 Φ) with a prevailing positive skewness (SK avg 0.17). Mz varies from 4.89 to 5.95 Φ (avg 5.63 Φ —coarse silt). Sediments are mainly bimodal with primary mode at 4.50–5.50 Φ and secondary mode around 2 Φ .

Table S1. Grain size data and statistical parameters. Mz, σ_i , Sk and Kg are the mean size, sorting, skewness and kurtosis according to [1], C is the 1st percentile, M is the median, >1 mm are the particles >1 mm. The assigned cluster for each level is indicated.

Level—Cm	Sand %	Silt %	Clay %	Mz Φ	$\sigma_i \Phi$	Sk	Kg	C μm	M μm	>1 mm Counts	Cluster
BC2-1	20.7	74.2	5.0	5.43	1.82	0.22	1.04	419	27.5	12	4
BC2-3	16.7	79.1	4.2	5.40	1.62	0.26	1.11	356	28.0	34	4
BC2-4	17.2	78.5	4.2	5.41	1.65	0.25	1.10	376	27.7	41	4
BC2-6	14.4	80.5	5.1	5.58	1.69	0.25	1.10	410	25.0	20	4
BC2-7	11.5	82.7	5.8	5.76	1.71	0.24	1.08	415	21.8	19	3
BC2-8	11.2	81.4	7.4	5.95	1.83	0.20	1.03	422	19.1	23	2
BC2-10	11.6	80.5	7.8	6.02	1.89	0.16	1.04	438	17.9	25	2
BC2-11	13.1	79.2	7.7	5.99	1.95	0.12	1.05	441	17.8	18	2
BC2-12	14.1	78.9	7.0	5.89	1.94	0.12	1.10	468	19.2	44	2
BC2-14	12.3	80.2	7.4	6.14	1.90	0.03	1.02	454	14.4	30	1
BC2-15	16.5	75.4	8.1	6.03	2.15	-0.04	1.08	502	15.2	45	1
BC2-16	15.5	77.3	7.1	6.02	2.12	-0.06	1.18	508	15.3	60	1
BC2-18	8.4	84.3	7.3	6.47	1.76	-0.04	1.09	412	11.0	51	1
BC2-19	12.6	79.5	7.9	6.24	2.08	-0.08	1.25	554	13.1	21	1
BC2-20	7.1	86.1	6.7	6.36	1.67	0.05	1.06	358	12.6	28	1
BC2-22	11.7	81.4	6.9	6.30	1.93	-0.10	1.14	464	12.0	27	1
BC2-23	7.8	86.0	6.2	6.27	1.67	0.06	1.05	348	13.5	33	1
BC2-24	11.0	79.8	9.2	6.44	1.97	-0.06	1.10	536	11.1	53	1
BC2-26	13.0	81.7	5.3	5.98	1.82	0.02	1.02	414	16.2	63	2
BC2-27	10.9	83.8	5.3	5.99	1.72	0.09	1.01	299	16.6	90	2
BC2-28	14.1	80.0	5.9	5.98	1.86	0.03	0.98	429	16.1	107	2
BC2-29	18.0	76.7	5.3	5.73	1.91	0.07	0.98	400	20.0	76	3
BC2-30	24.8	69.5	5.7	5.42	2.14	0.08	1.07	572	25.9	97	3
BC2-31	29.6	65.6	4.8	5.07	2.26	0.00	1.11	663	29.7	102	3
BC2-32	32.0	63.0	5.0	5.08	2.28	0.05	1.01	616	31.3	95	6
BC2-33	30.8	64.7	4.5	5.07	2.17	0.05	1.07	619	31.2	68	6
BC2-34	29.7	65.7	4.6	5.13	2.05	0.12	1.05	460	31.8	109	6
BC2-35	36.1	58.2	5.7	5.01	2.38	0.11	0.92	577	34.5	111	6
BC2-36	42.9	53.2	3.9	4.56	2.23	0.16	1.02	630	49.2	53	6
BC3-1	41.0	55.6	3.4	4.71	1.76	0.38	1.12	553	50.9	1	6
BC3-3	31.7	63.6	4.7	5.11	1.88	0.33	0.98	468	38.2	1	6
BC3-4	27.8	66.9	5.3	5.26	1.91	0.30	0.98	460	33.5	21	6
BC3-5	19.0	74.6	6.4	5.66	1.91	0.22	0.94	427	23.6	8	3
BC3-6	17.3	76.1	6.7	5.71	1.89	0.23	0.96	397	22.9	9	3
BC3-7	18.0	75.4	6.5	5.70	1.91	0.19	0.97	408	22.3	18	3
BC3-8	19.6	73.9	6.6	5.63	1.93	0.22	0.97	446	24.2	19	3
BC3-10	17.6	76.8	5.5	5.61	1.81	0.20	0.98	432	23.4	4	3
BC3-11	17.6	76.2	6.1	5.70	1.87	0.18	0.95	389	21.9	4	3
BC3-12	17.3	76.7	6.0	5.70	1.85	0.18	0.96	376	21.8	2	3
BC3-13	18.0	76.3	5.7	5.61	1.83	0.21	0.98	420	23.9	4	3
BC3-14	16.1	77.5	6.5	5.81	1.89	0.14	0.98	438	19.8	7	3
BC3-15	12.0	80.1	7.9	6.00	1.87	0.18	0.97	421	17.9	13	2
BC3-16	11.0	81.2	7.7	6.04	1.84	0.17	0.97	376	17.3	14	2
BC3-18	8.3	85.0	6.7	6.22	1.72	0.09	1.00	312	14.3	9	1
BC3-19	10.0	83.5	6.5	6.09	1.76	0.12	0.97	308	15.9	13	2
BC3-20	15.6	77.7	6.7	5.82	1.89	0.15	0.99	431	19.8	19	3
BC3-21	13.4	80.1	6.4	5.79	1.79	0.25	0.96	131	21.6	9	3
BC3-22	6.7	84.8	8.5	6.27	1.76	0.17	0.95	108	14.6	6	2
BC3-23	5.6	88.1	6.3	6.19	1.63	0.17	0.96	99	15.2	4	2
BC3-24	6.7	86.2	7.1	6.19	1.70	0.17	0.96	107	15.3	8	2
BC3-26	10.0	83.5	6.5	6.05	1.75	0.14	0.96	224	16.5	12	2
BC3-27	10.2	83.7	6.0	6.04	1.73	0.12	0.97	348	16.5	9	2
BC3-28	10.3	84.4	5.3	5.98	1.68	0.12	0.97	176	17.1	26	2
BC3-29	18.4	75.4	6.2	5.69	1.93	0.15	0.98	475	22.0	36	3
BC3-30	20.1	73.0	6.9	5.70	1.99	0.15	0.94	390	21.7	35	3
BC3-31	23.8	69.3	6.9	5.55	2.02	0.21	0.93	439	25.5	33	6
BC3-32	27.5	66.6	5.9	5.37	2.01	0.21	0.93	407	28.7	27	6
BC3-34	27.5	67.1	5.4	5.34	1.94	0.24	0.92	322	29.7	25	6
BC3-35	30.5	64.8	4.6	5.17	1.94	0.22	0.95	433	33.3	27	6
BC3-36	31.6	63.0	5.4	5.17	2.09	0.20	0.98	469	33.3	38	6

	BC3-37	27.5	67.9	4.7	5.21	1.96	0.16	1.07	475	31.1	13	6
BC4-1	26.5	69.8	3.7	4.99	1.80	0.15	1.36	479	35.9	51	5	
BC4-2	26.9	69.7	3.4	4.89	1.58	0.25	1.38	418	39.2	56	5	
BC4-3	27.5	68.7	3.8	5.03	1.91	0.14	1.31	545	35.7	67	5	
BC4-4	25.4	70.8	3.8	5.09	1.73	0.24	1.20	426	35.2	58	5	
BC4-5	30.7	65.6	3.7	4.91	1.83	0.19	1.25	491	39.0	74	5	
BC4-6	21.1	74.3	4.6	5.33	1.84	0.20	1.17	483	29.9	56	4	
BC4-7	22.8	72.6	4.6	5.30	1.94	0.16	1.20	487	30.0	44	4	
BC4-8	19.5	75.5	5.0	5.46	1.90	0.15	1.14	421	26.4	60	4	
BC4-9	27.4	68.4	4.2	5.10	2.01	0.11	1.23	610	33.5	42	4	
BC4-10	16.7	77.7	5.6	5.66	1.93	0.11	1.14	483	22.4	33	3	
BC4-11	20.2	74.6	5.2	5.50	2.05	0.05	1.20	567	24.1	20	3	
BC4-12	13.0	80.8	6.1	5.88	1.81	0.13	1.04	355	18.8	24	2	
BC4-13	13.1	81.3	5.5	5.73	1.74	0.20	1.05	429	21.7	19	3	
BC4-14	13.2	80.4	6.3	5.90	1.91	0.08	1.15	495	18.3	15	2	
BC4-15	13.2	81.3	5.5	5.77	1.77	0.14	1.09	433	20.3	17	2	
BC4-16	11.7	81.9	6.3	5.91	1.79	0.15	1.07	419	18.5	14	2	
BC4-17	12.3	81.2	6.4	5.95	1.83	0.11	1.04	450	17.7	9	2	
BC4-18	12.4	81.8	5.8	5.79	1.74	0.19	1.05	433	20.7	14	3	
BC4-19	13.8	80.5	5.6	5.73	1.77	0.18	1.07	393	21.6	27	3	
BC4-20	11.5	82.0	6.4	5.88	1.78	0.18	1.05	473	19.5	19	2	
BC4-21	14.1	79.8	6.1	5.73	1.78	0.24	1.01	294	22.6	12	3	
BC4-22	12.3	81.1	6.6	5.84	1.78	0.24	0.99	316	20.7	12	3	
BC4-23	13.9	79.3	6.7	5.82	1.86	0.20	1.01	430	20.9	12	3	
BC4-24	15.2	78.3	6.5	5.73	1.89	0.20	1.08	618	22.6	17	3	
BC4-25	11.9	81.3	6.8	5.94	1.82	0.17	1.01	414	18.7	3	2	
BC4-26	15.3	78.2	6.5	5.75	1.89	0.19	1.06	451	22.0	13	3	
BC4-27	15.2	78.3	6.6	5.76	1.86	0.20	1.02	400	21.8	17	3	
BC4-28	13.8	79.5	6.7	5.81	1.84	0.21	1.03	446	21.2	12	3	
BC4-29	15.7	78.1	6.2	5.70	1.83	0.22	1.02	414	23.0	17	3	
BC4-30	15.8	77.7	6.5	5.74	1.90	0.18	1.06	464	22.1	23	3	
BC4-31	15.3	78.2	6.5	5.79	1.90	0.16	1.06	455	21.0	19	3	
BC4-32	14.9	78.6	6.5	5.77	1.86	0.20	1.04	441	21.5	31	3	
BC4-33	20.0	74.5	5.5	5.46	1.86	0.21	1.09	456	27.0	9	4	
BC4-34	17.9	76.4	5.7	5.56	1.90	0.17	1.12	479	25.0	20	4	
BC4-35	16.0	77.6	6.5	5.73	1.89	0.20	1.04	454	22.4	26	3	
BC4-36	14.3	79.3	6.5	5.77	1.83	0.23	1.02	362	22.0	21	3	
BC4-37	11.7	82.3	6.0	5.93	1.77	0.17	0.95	309	18.4	15	2	
BC4-38	11.1	82.7	6.2	5.93	1.76	0.20	0.94	338	19.1	26	3	
BC4-39	16.0	77.0	7.0	5.80	1.90	0.21	0.95	428	21.4	24	3	
BC4-40	14.5	78.3	7.2	5.88	1.90	0.18	0.96	363	19.8	19	3	
BC4-41	12.5	82.0	5.5	5.86	1.76	0.17	0.97	342	19.5	8	3	
BC4-42	17.7	75.8	6.5	5.77	1.99	0.10	1.03	434	20.3	38	3	
BC4-43	22.2	71.8	6.0	5.56	2.08	0.09	1.04	483	23.4	58	3	
BC4-44	29.3	65.3	5.3	5.29	2.07	0.15	0.95	460	29.2	109	6	
BC4-45	24.2	70.0	5.8	5.49	2.11	0.08	1.03	545	24.5	63	3	

Supplementary S2

In this section we report a summary of the ecological and paleoenvironment meaning of the micropaleontological proxies used in this study.

Diatoms

The group composed of *Chaetoceros* resting spore (CRS) is used as a proxy for stratified and low salinity water related with the glacial retreat and/or ice melting. CRS is also associated with high productivity and/or upwelling [1–3]. *Eucampia antarctica* var. *recta* (the symmetric and polar forms, hereafter *E. antarctica*) lives at the ice edge [4] and is used as a proxy for meltwater from glacier and/or iceberg which release iron and promote *E. antarctica* growth [5–8]. *Thalassiosira antarctica* flourishes during the summer and forms resting spores when sea ice returns [9]. In the western Ross Sea [10] reported that *T. antarctica* was associated with the proximity of the ice shelf or sea ice edge. *Fragilariaopsis curta* is used as a proxy for qualitative sea ice extension [3,11,12].

F. obliquecostata is related to cold water near the ice edge [10,13]. *F. kerguelensis* dominates in open ocean zones where sea ice is absent during the summer [9] and it is related to warm water intrusion [14,15] in the Ross Sea; it is used as an indicator of CDW [16]. The “warm water group” species, is composed of *Thalassiosira lentiginosa*, *T. oliverana*, and *T. gracilis*, and indicates warm water intrusion and/or open ocean condition [14].

Silicoflagellates

Open ocean group: including *Stephanocha. speculum* var. *speculum*, and var. *monospicata*, var. *bispicata*; it is found in the present-day permanently in open ocean waters or close to the winter sea ice limit [17]. *S. speculum* var. *coronata* is found in the present-day sea ice dominated waters [18], *S. speculum* var. *minuta* (included in this work along with var. *coronata* because the two characters co-occur) is a cold-water indicator, typically increasing in relative abundance at higher latitudes [19–21].

Foraminifera

Neogloboquadrina pachyderma (Ehrenberg) is the only planktic foraminifera able to live in polar oceans, generally feeding on bacteria and diatoms. This species can also survive in brine channels within sea ice under hyper-saline and low temperature conditions [22–24]. The encrusted adults are related to warm-season while non-encrusted juvenile forms are related to autumn–winter season [25]. The co-presence of both growth stages of *N. pachyderma* suggests the accumulation of tests from different seasons, as already observed in a core located in the same area [26]. *Cibicides* spp., *Globocassidulina biora*, *G. subglobosa*, *Nonionella* spp., and *Trifarina earlandi* are commonly correlated to diverse sub ice shelf paleoenvironments, from a proximal to a distal position in respect to the grounding line sector (Bart et al., 2016 [27–31]). *Alabaminella weddellensis* and *Epistominella exigua* are small taxa generally occurring in the deep Atlantic and upper slope bathyal Southern Ocean under the influence of the AABW [32,33]. Furthermore, they are considered opportunistic species (r-strategist) adapted to fluctuating food supply in areas where sea ice hinders the seasonal accumulation of phytodetritus [34–36]. *E. exigua* may indicate the occasional influence of the CDW. In fact, it is thought that this species may be related to these warmer waters, as suggested by [30,37]). *Miliammina earlandi*, agglutinated taxon, is typically associated with the influence of corrosive HSSW in open water conditions [30,38].

Supplementary S3

In this section we report foraminifera data used in this study.

depth (cm) BC03	plankton/g	benthos/g	fragmentation %	<i>Alabaminella weddellensis</i>	<i>Astromion antarcticus</i>	<i>Astromion echolsi</i>	<i>Cassidinella carinata</i>	<i>Cibicides lobatulus</i>	<i>Cibicides refugens</i>	<i>Discorbis</i> sp.	<i>Epistominella exigua</i>	<i>Fissurina marginata</i>	<i>Fuskenkonia fusiformis</i>	<i>Globocassidinella biora</i>	<i>Globocassidinella subglobosa</i>	<i>Heroniallenia kempfi</i>	<i>Lagenia</i> spp.	<i>Melonis hardaeum</i>	<i>Miliammina carlundi</i>	<i>Nonionella bradii</i>	<i>Nonionella iridea</i>	<i>Oolina</i> spp.	<i>Patellina cornuta</i>	<i>Pullenia subcarinata</i>	<i>Stainforthia concava</i>	<i>Trifarina earlandi</i>	<i>Trifarina eurlandi</i>	
1	12		57.9																									
3	1		60.0																									
4		1																										
8		1																										
12		3	21.4																									
16		6	29.0																									
20			2																									
22	1	3	27.3																									
24	1	4	41.9																									
25	2	7	47.3																									
28	24	32	28.4	9		2		1		1	21	3						1	4	2	5	3			6	22		
31	818	1318	27.6	43		12	5	71	2	7	123	24	1	35	34	6	7	1	22	75	2	2	2	4	41			
34	673	1227	31.4	26	1	3	3	50	1	4	81	13		32	9	4	3	2	3	44	1		2	23				
36.5	500	215	20.5	12		4	4	27	5	2	56	23		43	25	2	8		4	14	8			1	39			
depth (cm) BC04	plankton/g	benthos/g	fragmentation %	<i>Alabaminella weddellensis</i>	<i>Cyclaminina orbicularis</i>	<i>Epistominella exigua</i>	<i>Fissurina marginata</i>	<i>Globocassidinella biora</i>	<i>Globocassidinella subglobosa</i>	<i>Legena</i> spp.	<i>Miliammina carlundi</i>	<i>Oolina</i> spp.	<i>Trifarina earlandi</i>	<i>Triloculina tricarinata</i>														
1					1																							
6		0	50.0																									
11		2	46.2																									
15		1	25.0																									
21	1	1					1				1																	
25	26	4	51.3				1				5	1																
31	206	14	37.6		3		1	13	4			3																
35	145	11	63.1	3		11				15	10			1														
41	2		53.8																									
45			100.0																									

Figure S1.

References (S1)

1. Folk, R. L.; Ward, W. C. Brazos River bar: a study in the significance of grain size parameters, *J. Sediment. Petrol.*, **1957**, *27*, 3–26.

References (S2)

1. Leventer, A. Sediment trap diatom assemblages from the northern Antarctic Peninsula region. *Deep-Sea Res.* **1991**, *38*, 1127–1143.
2. Crosta, X.; Pichon, J.J.; Labracherie, M. Distribution of Chaetoceros resting spores in modern peri-Antarctic sediments. *Mar. Micropaleontol.* **1997**, *29*, 283–299
3. Armand, L. K. The use of diatom transfer functions in estimating sea-surface temperature and sea-ice in cores from the southeast Indian Ocean, Ph.D. thesis, Australian National University, Canberra, Australia, **1997**
4. Thomas, E. R.; Allen, C. S.; Etourneau, J.; King, A. C. F.; Severi, M.; Winton, V. H. L.; Mueller, J.; Crosta, X.; Peck, V. L. Antarctic Sea Ice Proxies from Marine and Ice Core Archives Suitable for Reconstructing Sea Ice over the Past 2000 Years. *Geosciences*, **2019**, *9*, 506
5. Burckle, L. H.; Cooke, D. W. Late Pleistocene *Eucampia antarctica* abundance stratigraphy in the Atlantic sector of the Southern Ocean. *Micropaleontology* **1983**, *29* (1), 6–10
6. Burckle, L. H. Ecology and paleoecology of the marine diatom *Eucampia antarctica* (Castr.) Mangin. *Mar. Micropaleontol.* **1984**, *9*, 77–86
7. Armand, L.K.; Crosta, X.; Queguiner, B.; Mossner, J.; Garcia, N. Diatoms preserved in surface sediments of the northeastern Kerguelen Plateau. *Deep Sea Res Part II Top Stud Ocean*, **2008** *55* (507), 677–692.
8. Minzoni, R. T.; Anderson, J. B.; Fernandez, R.; Wellner, J. S. Marine record of Holocene climate, ocean, and cryosphere interactions: Herbert Sound, James Ross Island, Antarctica, *Quat. Sci. Rev.*, **2015**, *129*, 239–259
9. Campagne, P.; Crosta, X.; Schmidt, S.; Houssais, M. N.; Ther, O.; Massé, G. Sedimentary response to sea ice and atmospheric variability over the instrumental period off Adélie Land, East Antarctica, *Biogeosciences*, **2016**, *13*, 4205–4218

10. Cunningham, W. L.; Leventer, A. Distribution of diatom assemblages in surface sediments of the Ross Sea, Antarctica: relation to modern oceanographic conditions. *Antarctic Science*, **1998**, *10* (2), 134-146.
11. X. Crosta, D.; Denis, O. Ther, Sea ice seasonality during the Holocene, Adélie Land, East Antarctica, *Marine Micropaleontology*, **2008**, *66*, 222-232.
12. Esper O.; Gersonde R. New tools for the reconstruction of Pleistocene Antarctic Sea Ice. *Palaeogeogr. Palaeoclim. Palaeoeco.* **2014**, *399*, 260-283.
13. Cunningham, W.L.; Leventer, A.; Andrews, J.T.; Jennings, A.E.; Licht, K.J. Late Pleistocene–Holocene marine conditions in the Ross Sea, Antarctica: evidence from the diatom. *The Holocene*, **1999**, *9*, 129–139
14. Crosta, X.; Romero, O.; Armand, L.K.; Pichon, J.J. The biogeography of major diatom taxa in Southern Ocean sediments: 2. Open ocean related species. *Palaeogeogr. Palaeoclim. Palaeoeco.* **2005**, *223*, 66-92
15. Pike, J.; Allen, C. S.; Leventer, A.; Stickley, C. E.; Pudsey, C. J. Comparison of contemporary and fossil diatom assemblages from the western Antarctic Peninsula shelf. *Mar. Micropaleontol.*, **2008**, *67*, 274-287
16. Mezgec, K.; Stenni, B.; Crosta, X.; Masson-Delmotte, V.; Baroni, C.; Braida, M.; Ciardini, V.; Colizza, E.; Melis, R.; Salvatore, M. C.; Severi, M.; Scarchilli, C.; Traversi, R.; Udisti, R.; Frezzotti, M. Holocene sea ice variability driven by wind and polynya efficiency in the Ross Sea. *Nat. Commun.* **2017**, *8*, 1334
17. Malinverno, E. Extant morphotypes of *Distephanus speculum* (Silicoflagellata) from the Australian sector of the Southern Ocean: Morphology, morphometry and biogeography. *Mar. Micropal.* **2010**, *77*, 154–174.
18. Malinverno, E.; Maffioli, P.; Gariboldi, K. Latitudinal distribution of extant fossilizable phytoplankton in the Southern Ocean: Planktonic provinces, hydrographic fronts and palaeoecological perspectives. *Mar. Micropal.* **2016**, *123*, 41–58.
19. Bukry, D. Coccolith and silicoflagellate stratigraphy, Deep Sea Drilling Project Leg 18, eastern North Pacific. In: Kulm, L.D., von Huene, R., et al. (Eds.), *Initial Reports of the Deep Sea Drilling Project 1973*, *18*, 817-831.
20. Bukry, D. Silicoflagellate and coccolith stratigraphy, Deep Sea Drilling Project, Leg 29. In: Kennet, J.P., Houtz, R.E., et al. (Eds.), *Initial Reports of the Deep Sea Drilling Project 1975*, *29*, 845-872.
21. Bukry, D. Silicoflagellate stratigraphy of offshore California and Baja California, Deep Sea Drilling Project Leg 63. In: Yeats, R. S., Haq, B. U., et al., Init. Repts. DSDP, *63*: Washington (U.S. Govt. Printing Office), **1981**, 595-610.
22. Spindler, M.; Dieckmann, G. S. Distribution and abundance of the planktic foraminifer *N. pachyderma* in the sea ice of the Weddell Sea (Antarctica). *Polar Biol.* **1986**, *5*, 185-191
23. Dieckmann, G. S.; Spindler, M.; Lange, M. A.; Ackley, S. F.; Eicken, H. Antarctic sea ice: a habitat for the foraminifer *Neogloboquadrina pachyderma*. *J. Foraminiferal Res* **1991**, *21*, 182-189.
24. Bergami, C.; Capotondi, L.; Langone, L.; Giglio, F.; Ravaioli, M. Distribution of living planktonic foraminifera in the Ross Sea and the Pacific sector of the Southern Ocean (Antarctica). *Mar. Micropaleontol.* **2009**, *73*, 37-48.
25. Mikis, A.; Hendry, K.R.; Pike, J.; Schmidt, D.N.; Edgar, K.M.; Peck, V.; Peeters, F.J.C.; Leng, M.J.; Meredith, M.P.; Todd, C.L.; Stammerjohn, S.; Ducklow, H. Temporal variability in foraminiferal morphology and geochemistry at the West Antarctic Peninsula: a sediment trap study. *Biogeosciences* **2019**, *16*, 3267-3282
26. Melis, R.; Capotondi, L.; Torricella, F.; Ferretti, P.; Geniram, A.; Hong, J.K.; Khun, G.; Khim, B.K.; Kim, S.; Malinverno, E.; Yoo, K.C.; Colizza, E. Last Glacial Maximum to Holocene paleoceanography of the northwestern Ross Sea inferred from sediment core geochemistry and micropaleontology at Hallett Ridge. *J. Micropal.* **in press**
27. Bart, P. J.; Coquereau, L.; Warny, S.; Majewski, W. In situ foraminifera in grounding zone diamict: a working hypothesis. *Antarct. Sci.* **2016**, *28*, 313-321.
28. McGlannan, A.J.; Bart, P.J.; Chow, J.M.; DeCesare, M. On the influence of post- LGM ice shelf loss and grounding zone sedimentation on West Antarctic ice sheet stability. *Mar. Geol.* **2017**, *392*, 151-169.
29. Majewski, W.; Bart, P. J.; McGlannan, A. J. Foraminiferal assemblages from ice-proximal paleo-settings in the Whales Deep Basin, Eastern Ross Sea, Antarctica. *Palaeogeogr. Palaeocl.* **2018**, *493*, 64-81
30. Majewski, W.; Prothro, L. O.; Simkins, L. M.; Demianiuk, E. J.; Anderson, J. B. Foraminiferal patterns in deglacial sediment in the western Ross Sea, Antarctica: life near grounding lines. *Paleocean. and Paleoocl.* **2020**, *35*
31. Prothro, L.O.; Simkins, L.M.; Majewski, W.; Anderson, J.B. Glacial retreat patterns and processes determined from integrated sedimentology and geomorphology records. *Mar. Geol.* **2018**, *395*, 104-119.
32. Mackensen, A.; Grobe, H.; Kuhn, G.; Fütterer, D. K. Benthic foraminiferal assemblages from the eastern Weddell Sea between 68 and 73 °S: distribution, ecology and fossilization potential. *Mar. Micropaleontol.* **1990**, *16*, 241-283.
33. Mackensen, A.; Fütterer, D. K.; Grobe, H.; Schmiedl, G. Benthic foraminiferal assemblages from the eastern South Atlantic Polar Front region between 35 and 57 S: Distribution, ecology and fossilization potential. *Mar. Micropaleontol.* **1993**, *22*, 33–69.
34. Mackensen, A.; Grobe, H.; Hubberten, H. W.; and Kuhn, G. Benthic foraminiferal assemblages and the d13C-signal in the Atlantic sector of the Southern Ocean: Glacial-to-interglacial contrasts, in: Carbon cycling in the glacial ocean: Constraints on the ocean's role in global change, edited by: Zahn, R., Kaminski, M., Labeyrie, L., and Pedersen, T., NATO ASI series, Springer, Berlin **1994**, *17*, 105-144.
35. Ishman, S. E.; Szymcek, P. Foraminiferal distributions in the former Larsen-A Ice Shelf and Prince Gustav Channel region, eastern Antarctic Peninsula margin: a baseline for Holocene paleoenvironmental change, in: Antarctic Peninsula Climate Variability: Historical and Paleoenvironmental Perspectives, Antarctic Research Series, *79*, edited by: Domack, E., Leventer, A.; Burnet, A.; Bindschadler, R.; Convey, P.; Kirby, M. *American Geophysical Union* **2003**, 239–260
36. Jorissen, F. J.; Fontanier, C.; Thomas, E. Paleoceanographical proxies based on deep-sea benthic foraminiferal assemblage characteristics, in Proxies in Late Cenozoic. Paleoceanography, Developments in Marine Geology, *1*, edited by: Hillaire-Marcel, C.; De Vernal, A., Elsevier Sc. **2007**, 263-325.

37. Majewski, W.; Wellner, J. S.; Anderson, J. B. Environmental connotations of benthic foraminiferal assemblages from coastal West Antarctica. *Mar. Micropaleontol.* **2016**, *124*, 1–15
38. Melis, R.; Salvi, G. Late Quaternary foraminiferal assemblages from western Ross Sea (Antarctica) in relation to the main glacial and marine lithofacies. *Marine Micropaleontol.* **2009**, *70*, 39–53.