

Supplementary Material

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1 Datasets

This section describe the datasets analysed by MOMIC and table ?? summarises type of data used as input and their sizes.

GWES: Microarray: We have used data from the Gene Expression Omnibus (GEO) database [1]. The first dataset, GSE48350 contains microarray data from normal controls (aged 20-99 years) and Alzheimer’s disease cases, from 4 brain regions: hippocampus, entorhinal cortex, superior frontal cortex, post-central gyrus. Changes in expression of synaptic and immune related genes were analyzed, investigating age-related changes and AD-related changes, and region-specific patterns of change. These AD cases were processed simultaneously with the control cases (young and aged) included in GSE11882 (GSE11882 dataset contains data exclusively from normal control brains). The second dataset, GSE15222, is also related to genetic control of human brain transcript expression in Alzheimer’s disease.

GWES: RNASeq. We have produced a synthetic dataset simulating astrocytes from AD cases and controls, illustrating the diversity of data layers that can be integrated using the tools provided by MOMIC.

GWAS: We have prepared a dataset using data from the HapMap Project [3]. This preparation consists on updating the datasets to rsID, removing SNPs with minor allele frequency and adding fake case/control phenotypes. The goal of the International HapMap Project is to determine the common patterns of DNA sequence variation in the human genome and to make this information freely available in the public domain.

Data from the International Genomics of Alzheimer’s Project (IGAP) [2] have been used to illustrate the GWAS Meta-analysis protocol. IGAP is a large two-stage study based upon genome-wide association studies (GWAS) on individuals of European ancestry. In stage 1, IGAP used genotyped and imputed data on 7,055,881 single nucleotide polymorphisms (SNPs) to meta-analyse four previously-published GWAS datasets consisting of 17,008 Alzheimer’s disease cases and 37,154 controls (The European Alzheimer’s disease Initiative – EADI the Alzheimer Disease Genetics Consortium – ADGC The Cohorts for Heart and Aging Research in Genomic Epidemiology consortium – CHARGE The Genetic and Environmental Risk in AD consortium – GERAD). In stage 2,

Table 1: Sizes of input datasets

Protocol	Dataset	Type	Size
GWAS	1KG	bed	1.2 GB
GWES Micorarray	GSE15222	csv (expression matrix)	17 MB
GWES Micorarray	GSE48350	CEL (raw data)	1.3 GB
GWES RNASeq	Synthetic data	FASTQ	7.2 GB
Proteomics	BLSA	csv (intensity matrix)	11 MB

11,632 SNPs were genotyped and tested for association in an independent set of 8,572 Alzheimer’s disease cases and 11,312 controls. Finally, a meta-analysis was performed combining results from stages 1 & 2.

Proteomics: The dataset used for the proteomics testing is the anonymized Baltimore Longitudinal Study of Aging (BLSA) [4]. This study is a clinical research program on human aging that began in 1958. Volunteers of different ages join the study when they are healthy, and have follow-up visits for life. Visits last for multiple days. Participants are evaluated for many physical elements as well as for brain function. Physical tests are given. Information on mood, personality, and social aspects of life is also collected. This program has contributed more than any other research project to our understanding of aging.

2 Protocol diagrams

MOMIC currently compiles protocols for whole genome SNP data (GWAS), mRNA expression (both from arrays and RNAseq experiments) and protein data. Along with enrichment analysis and methods for combining heterogeneous data at different molecular levels. Figures 1 to 4 reveal the collection of notebooks designed for each protocol.

References

- [1] E. Clough and T. Barrett. The gene expression omnibus database. In *Statistical genomics*, pages 93–110. Springer, 2016.
- [2] J.-C. L. et al. Meta-analysis of 74,046 individuals identifies 11 new susceptibility loci for alzheimer’s disease. 45:1452–1458, 2013.
- [3] R. A. Gibbs, J. W. Belmont, P. Hardenbol, T. D. Willis, F. Yu, H. Yang, L.-Y. Ch’ang, W. Huang, B. Liu, Y. Shen, et al. The international hapmap project. 2003.
- [4] N. W. Shock. Normal human aging: The baltimore longitudinal study of aging. 1984.

Quality Control & Data Pre-processing	Notebook: Task1_fromGEOtoMatrix_template.ipynb Description: Downloads a GEO dataset from NCBI using GEOquery package, converts it to an expression matrix and clinical dataset, and pre-process the obtained data. Input: GSExxxx GEO GSE name Output: objects.RData expression and clinical datasets
	Notebook: Task1_fromAffyRawCELtoMatrix_template.ipynb Description: Transforms raw .CEL files from Affymetrix platform to an expression matrix. Pre-process it using method from affy and gcrma R packages [ref]. Input: GSExxxx GEO GSE name Output: objects.RData expression and clinical datasets
	Notebook: Task1_fromAgilentRawtoMatrix_template.ipynb Description: Transforms raw data from Agilent platform to an expression matrix and pre-process it with limma R package [ref]. There is no Agilent data provided but the notebook states all the necessary steps. Input: path/to/Agilentdata Path to Agilent data Output: MAList_object.RData Curated expression data
	Notebook: Task1_Data_Preprocessing_template.ipynb Description: Use this template in case you have data different from Affymetrix, Agilent or GEO. This template performs background correction from limma, quantile normalization with preprocessCore R, logarithm transformation and correction for batch effect. Input: raw expression data Output: exprdata_qced.RData Quality expression dataset
Differential Analysis	Notebook: Task2_DifferentialExpression_template-compact.ipynb Task2_DifferentialExpression_template-stepbystep.ipynb Description: Performs DE analysis with Limma and custom scripts. Inputs: objects.RData expression and clinical curated datasets Outputs: limma_condlvscond2_annot List of differential expressed genes annotated (depending on platform)

Figure 1: GWES Microarray protocol

Quality Control	Notebook: Task1_QC_raw_data_template.ipynb Description: Performs quality checks of raw reads using FastQC software. User can decide whether to continue with the downstream or remove sequences with low quality analysis based on this QC results. Inputs: sample_info.txt Sample metadata *.fastq.gz Reads Outputs: *.zip, *.html FastQC results multiqc_report.html
Alignment & Read Quantification	Notebook: Task2.1_Alignment_and_ReadQuantification_template.ipynb Description: Aligns the reads to the reference genome with STAR Inputs: *.fastq.gz Reads Outputs: sampleName.bam Aligned sorted bam file sampleNameLog.final.out Alignment statistics sampleName.ReadsPerGen.out.tab Gene based read counts
	Notebook: Task2.2_ReadQuantification_to_DESeqDataSet_template.ipynb Description: Transforms the results of STAR quantification into a DESeq2::DESeqDataSet object Inputs: sampleName.ReadsPerGen.out.tab Read counts sample_info.txt Sample metadata Outputs: DESeq_object DESeq2:: DESeqDataSet
Differential Analysis	Notebook: Task4_DifferentialAnalysis_compact_template.ipynb Task4_DifferentialAnalysis_step_by_step_template.ipynb Description: Performs DE analysis with DESeq2 Inputs: DESeq_object DESeq2:: DESeqDataSet Outputs: DEG.results.annot List of differential expressed genes annotated using biomaRt

Figure 2: GWES RNASeq protocol

Data Pre-processing	Notebook: Task1_DataPreprocessing_template.ipynb Description: Curates metadata and protein data Input: <code>metadata.txt</code> Metadata <code>MaxQuantCorrIntensityUniprot.csv</code> Protein data Output: <code>protein_objects.RData</code> curated datasets
Differential Analysis	Notebook: Task2_DifferentialExpression_template.ipynb Description: Performs DE analysis with DEqMS R package Inputs: <code>proteins_objects.RData</code> curated datasets Outputs: <code>cond1vscond2_annot</code> List of differential expressed proteins annotated

Figure 3: Proteomics protocol

Pre-Quality Control	<p>Notebook: <code>Task1_PreQC_Build_template.ipynb</code></p> <p>Description: Prepares a working dataset in PLINK v1.9 binary format with all SNPs identified by the rs number and coordinates based on the genome build GRCh37/hg19. Use this template for changing builds.</p> <p>Input: <code>database.b36</code> Working dataset in build 36 - bfile (bim,bam,fam)</p> <p>Output: <code>database.b37</code> Working dataset in build 37 - bfile</p>
QC & Data Pre-processing	<p>Notebook: <code>Task2.1_Quality_Control_template.ipynb</code></p> <p>Description: Implements a quality control process in PLINK aimed at removing individuals and markers with particularly high error rates and filtering out population stratification errors.</p> <p>Input: <code>database.b37</code> Working dataset in build 37 - bfile</p> <p>Output: <code>database.b37.IBD</code> Clean working dataset - bfile <code>indepSNP.prune.in</code> List of non (highly) correlated SNPs</p> <p>Notebook: <code>Task2.2_Population_Stratification_template.ipynb</code></p> <p>Description: Implements a quality control process in PLINK aimed at removing individuals and markers with particularly high error rates and filtering out population stratification errors.</p> <p>Input: <code>database.b37.IBD</code> Clean dataset - bfile <code>indepSNP.prune.in</code> List of non (highly) correlated SNPs</p> <p>Output: <code>database.b37.Qced</code> Clean/QCed dataset - bfile <code>covar_mds.txt</code> Covariates file</p>
Imputation	<p>Notebook: <code>Task3_Imputation_template.ipynb</code></p> <p>Description: Implements genotype imputation with the Michigan Imputation Server, using the minimac 3 algorithm, the HRC reference panel and the SHAPEIT tool for haplotype phasing. Will Rayner's toolbox to prepare the data.</p> <p>Input: <code>database.b37.Qced</code> Clean/QCed dataset - bfile</p> <p>Output: <code>chr22.dose.for.assoc.fam</code> Fam dataset with updated phenotype <code>chri.dose.rsq.DS.vcf.gz</code> Genotype dosages</p>
Association	<p>Notebook: <code>Task4_Assoc_template.ipynb</code></p> <p>Description: Performs a case control association study using PLINK.</p> <p>Input: <code>chr22.dose.for.assoc.fam</code> Fam dataset with updated sex and phenotype <code>chri.dose.rsq.DS.vcf.gz</code> Genotype dosages <code>covar_mds.txt</code> Covariates file</p> <p>Output: <code>dataset.b37.imputed.assoc.dosage.clean.rs.200kb.annot</code> Annotated association results</p>
Visualisation	<p>Notebook: <code>Task5_Visualisation.ipynb</code></p> <p>Description: Displays GWAS results, plotting p-values that indicate the significance of the difference in frequency of the allele tested between cases and controls.</p> <p>Input: <code>dataset.b37.imputed.assoc.dosage.clean.rs.200kb.annot</code> Annotated association results</p> <p>Output: manhattans and QQ plots to standard output</p>
Gene-wise Statistics	<p>Notebook: <code>Task6_Gene-wise_Statistics_template.ipynb</code></p> <p>Description: Performs gene-wise statistic with MAGMA.</p> <p>Input: <code>dataset.b37.imputed.assoc.dosage.clean.rs.200kb.annot</code> Annotated association results</p> <p>Output: <code>dataset.b37.imputed.assoc.dosage.maf0.01.LOC.50kb.genes.annot</code> Annotated association results aggregated to genes.</p>

Figure 4: GWAS protocol