

## Supplementary for:

# Application of Biotechnology and Chiral Technology Methods in the Production of Ectoine Enantiomers

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## Table of contents

1. NMR spectra .....	S2
2. IR spectra.....	S18
3. Separation of (±)-ectoine enantiomers.....	S22
4. Detection of (+)-ectoine in fermentation broths of salt-tolerant marine <i>Streptomyces</i> .....	S23
4.1. <i>Streptomyces</i> strains.....	S23
4.2. HPLC chromatograms of (+)-ectoine in fermentation broths of salt-tolerant marine <i>Streptomyces</i> .....	S24
5. MS spectra of compound 7 and standard (+)-ectoine.....	S33

# 1. NMR spectra

Compound 1

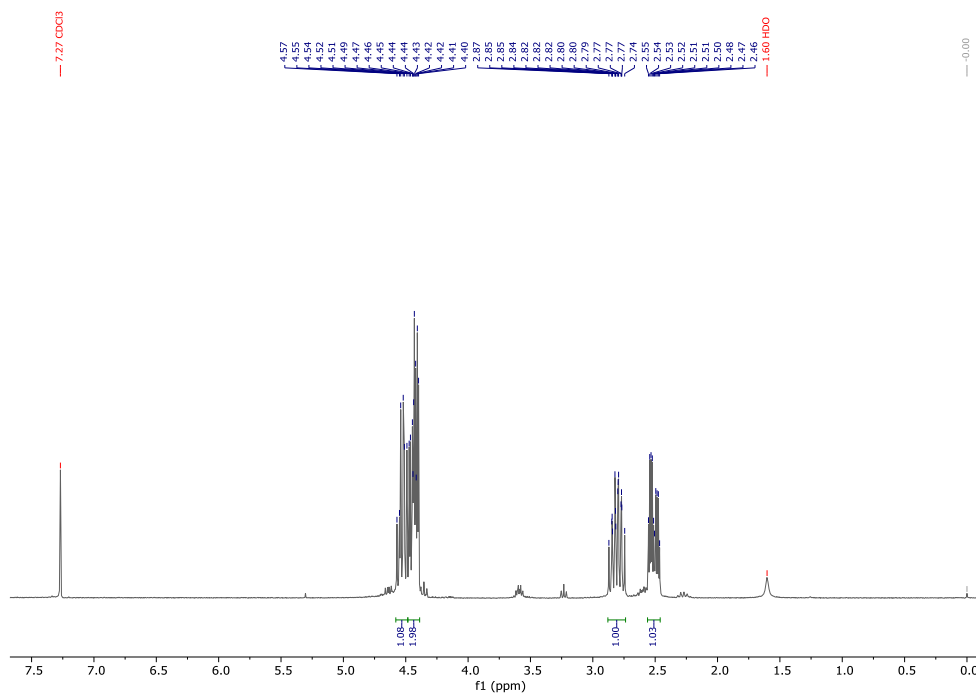
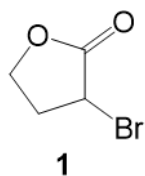


Figure S1.1.  $^1\text{H}$  NMR ( $\text{CDCl}_3$ , 300 MHz) spectra of compound 1.

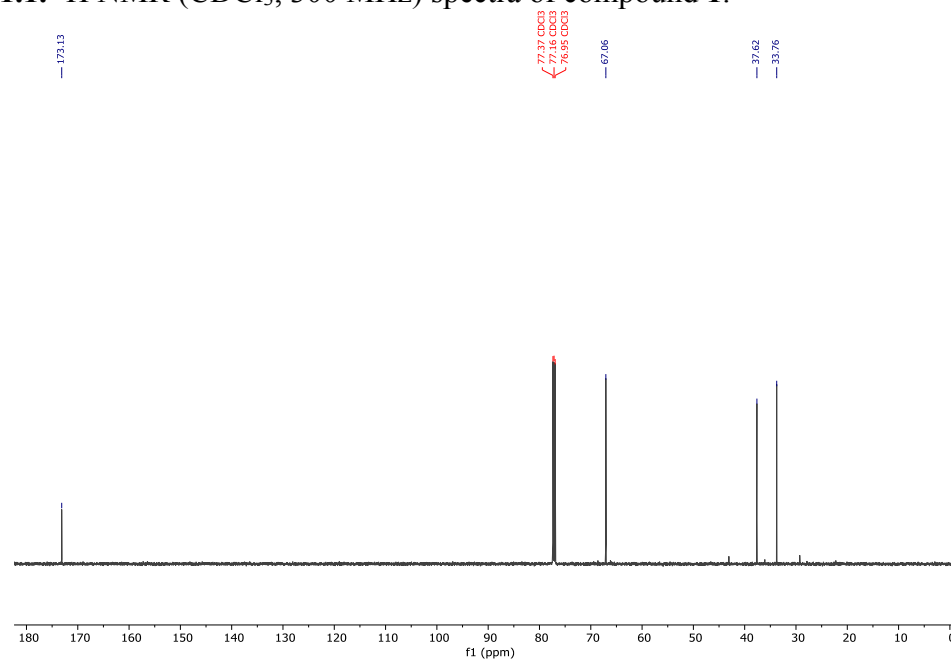
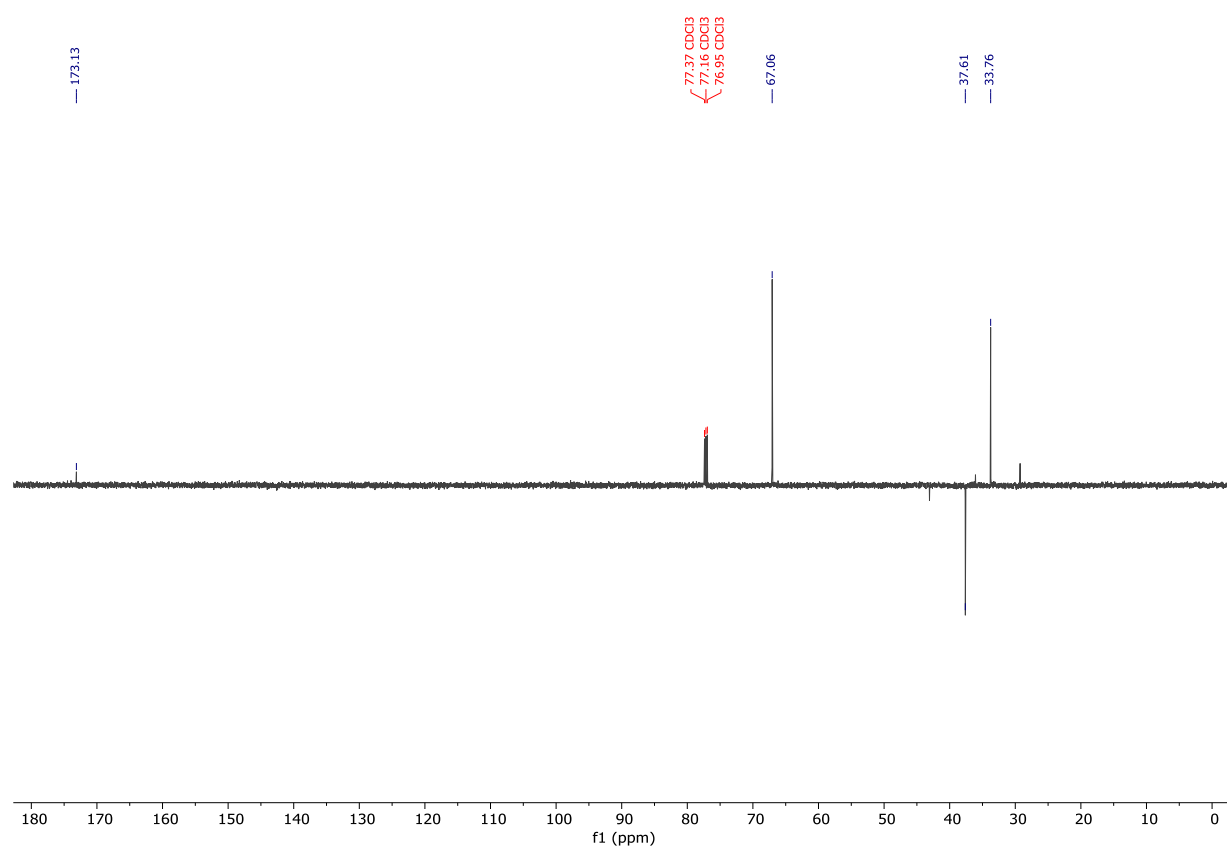
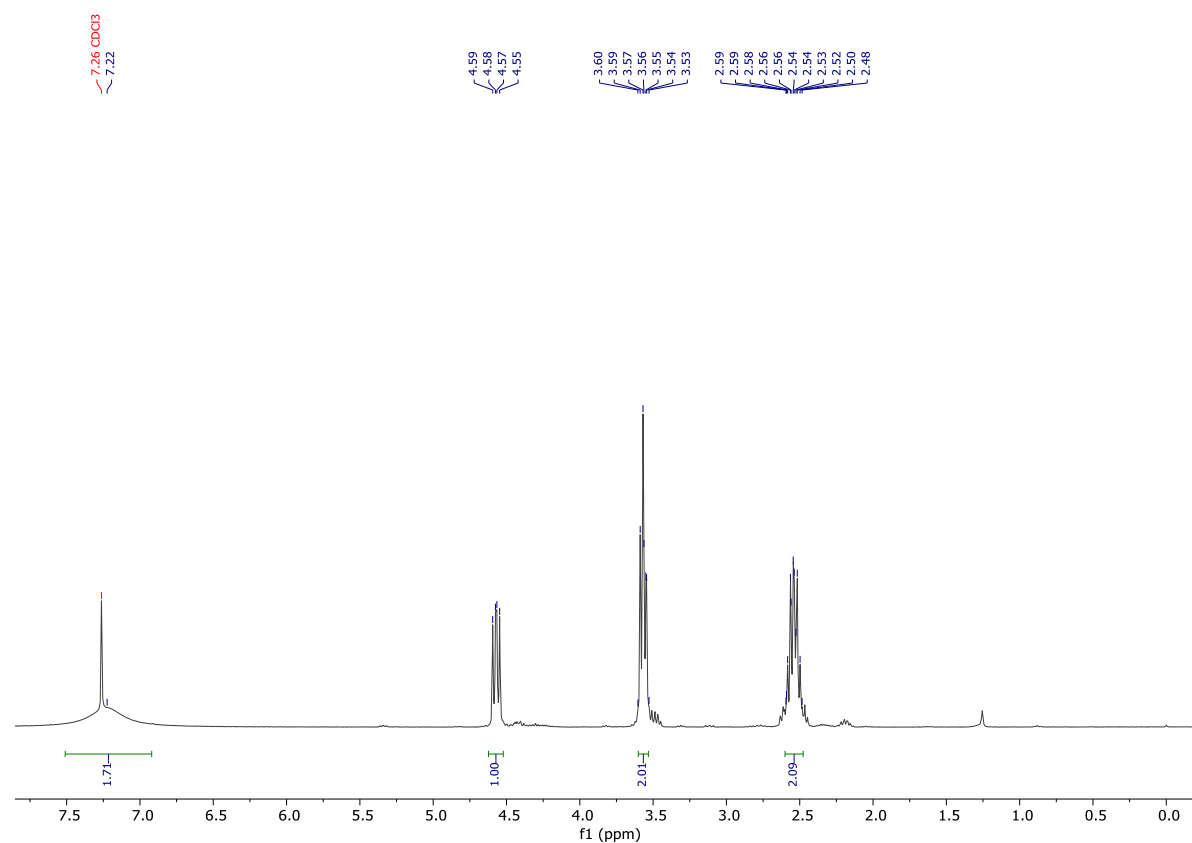
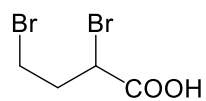


Figure S1.2.  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 75 MHz) spectra of compound 1.

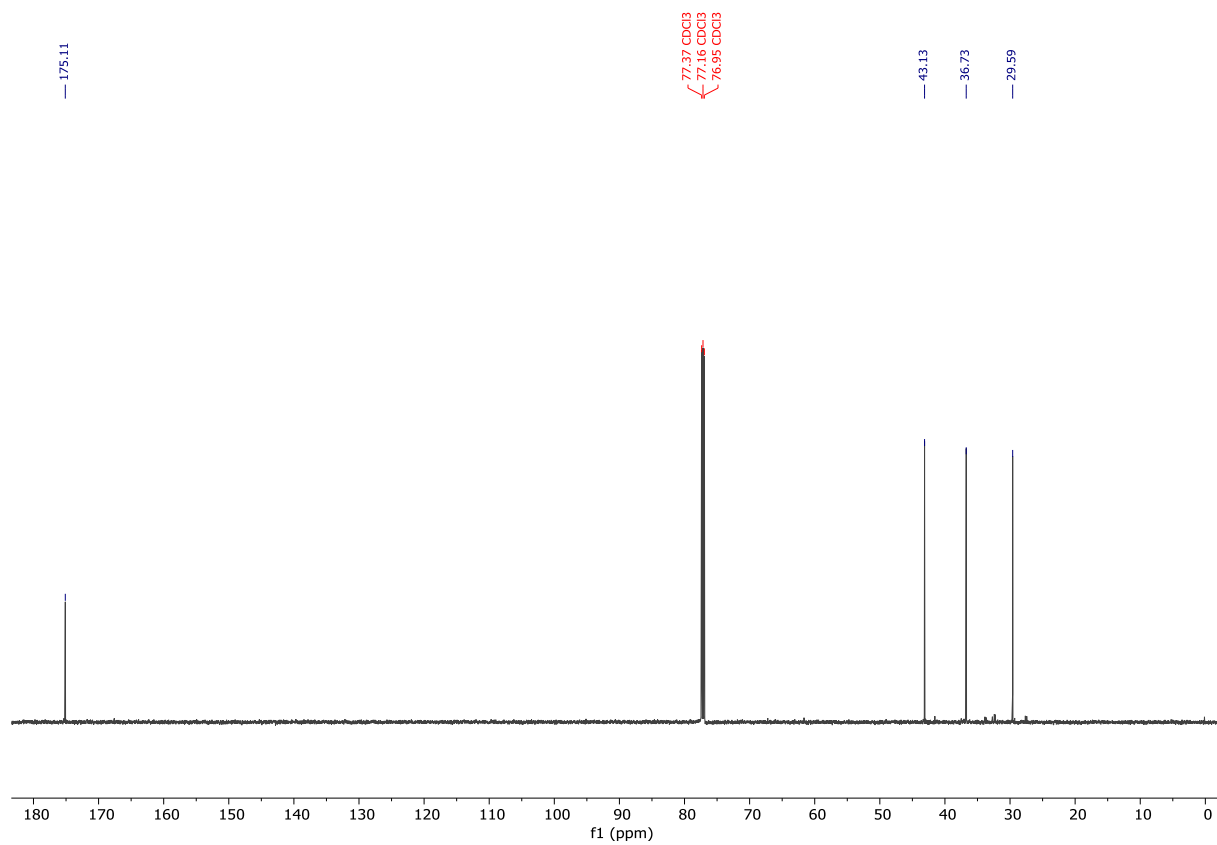


**Figure S1.3.**  $^{13}\text{C}$  APT NMR (CDCl<sub>3</sub>, 75 MHz) spectra of compound **1**.

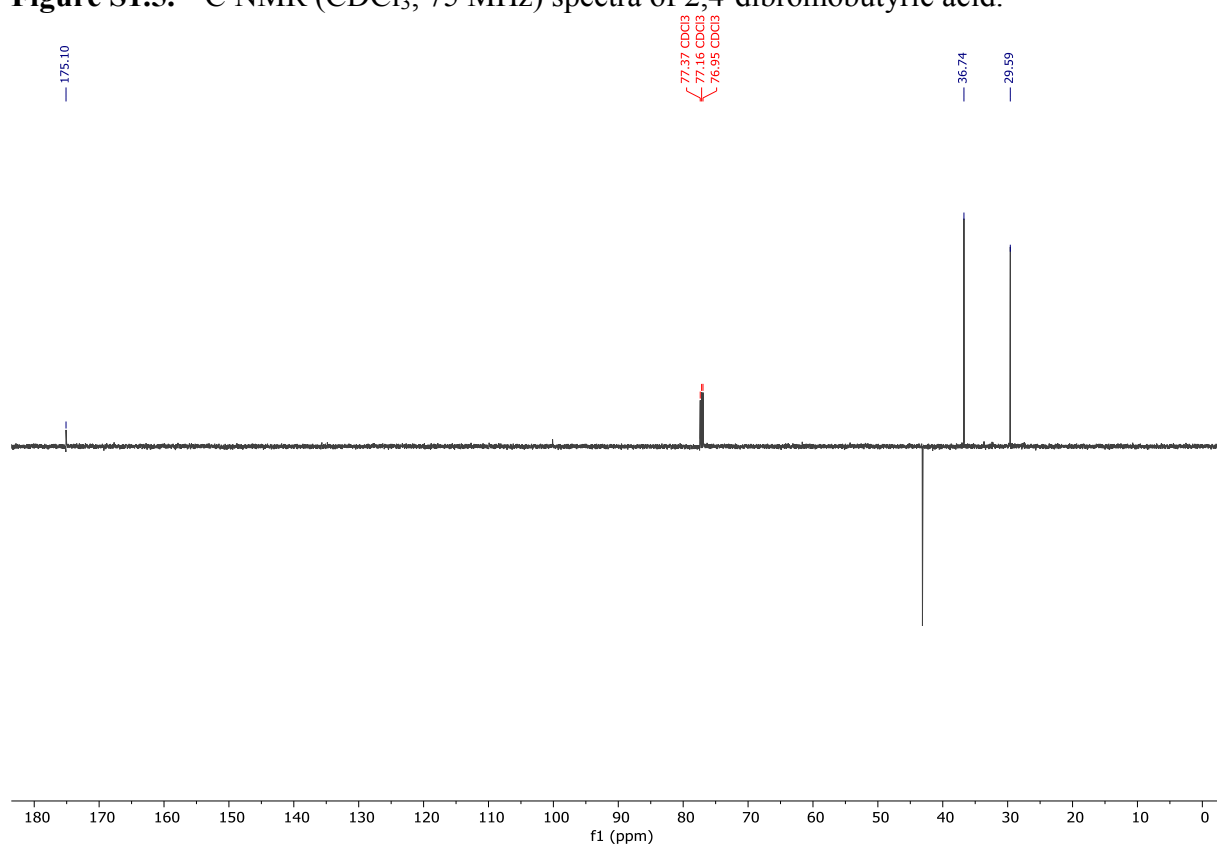
# 2,4-Dibromobutyric acid



**Figure S1.4.** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) spectra of 2,4-dibromobutyric acid.



**Figure S1.5.** <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz) spectra of 2,4-dibromobutyric acid.



**Figure S1.6.** <sup>13</sup>C APT NMR (CDCl<sub>3</sub>, 75 MHz) spectra of 2,4-dibromobutyric acid.

Compound **2**

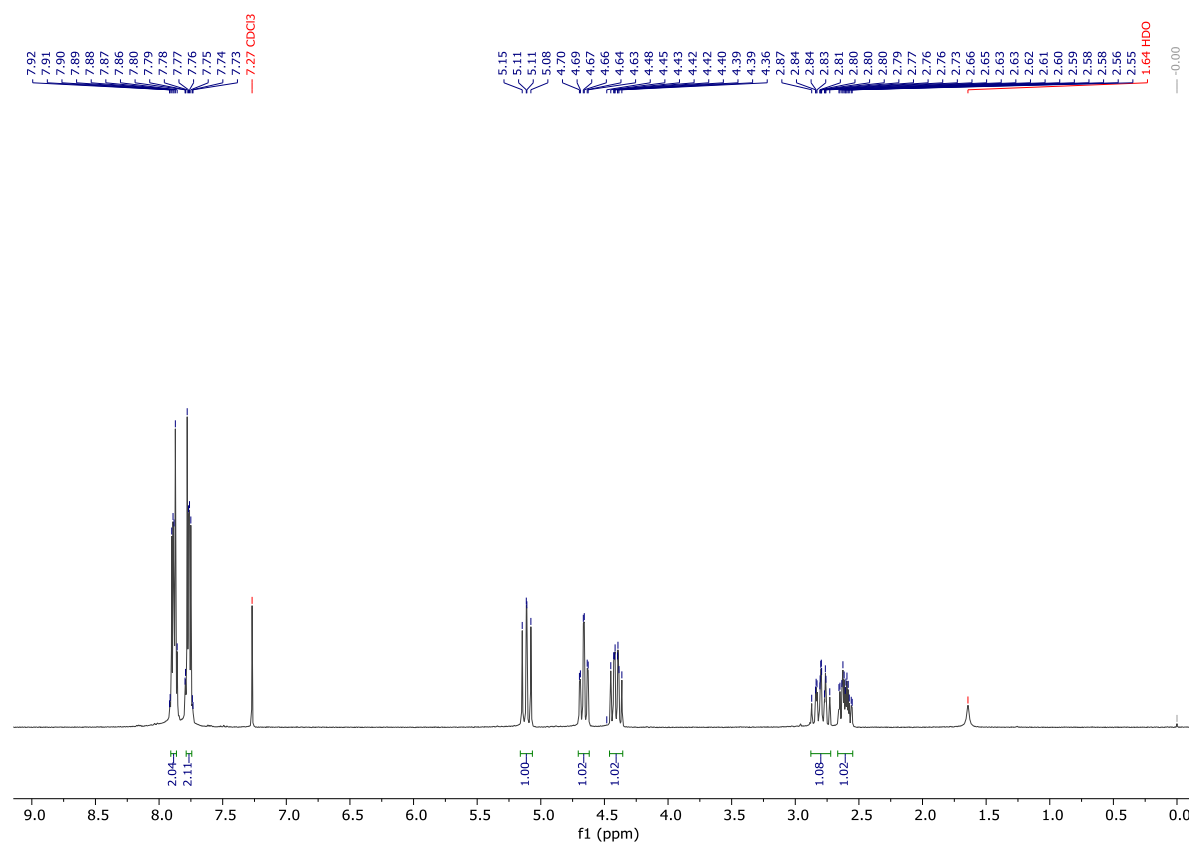
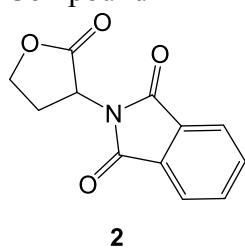
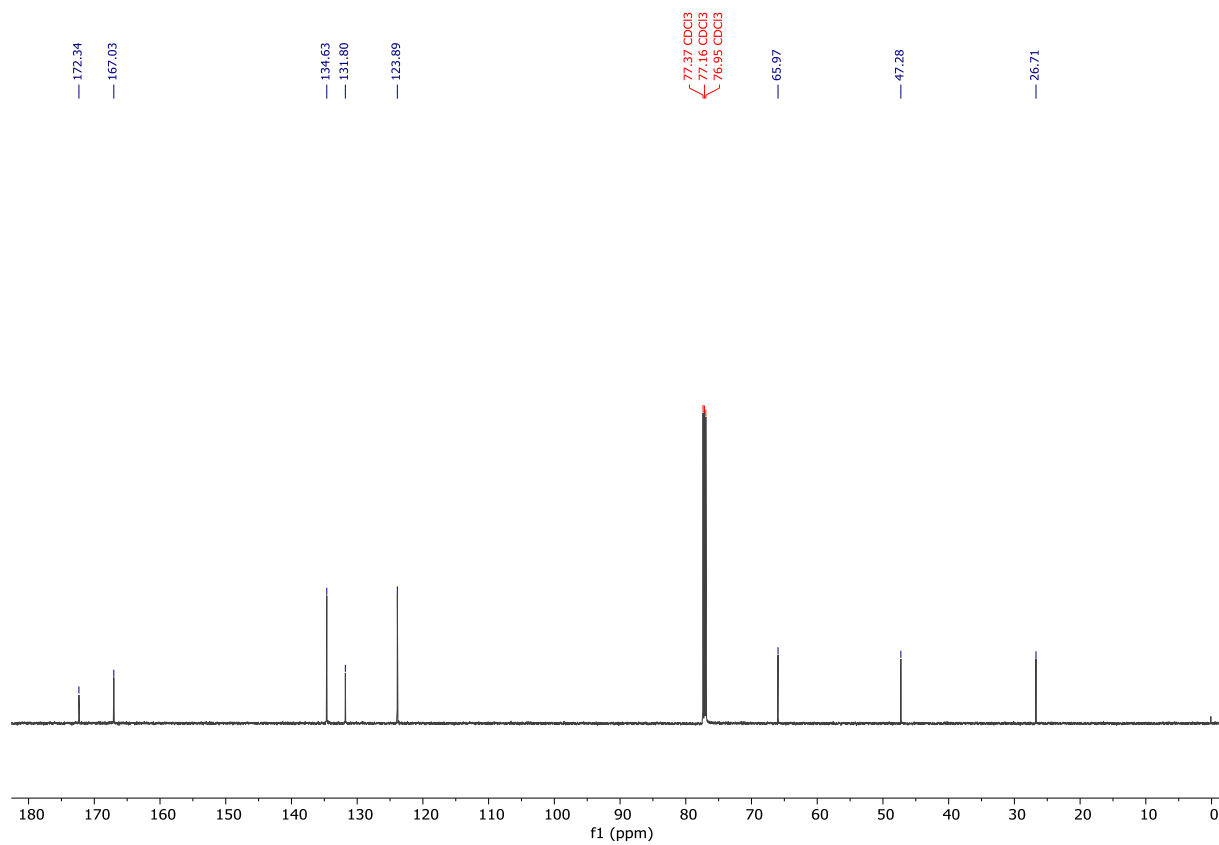
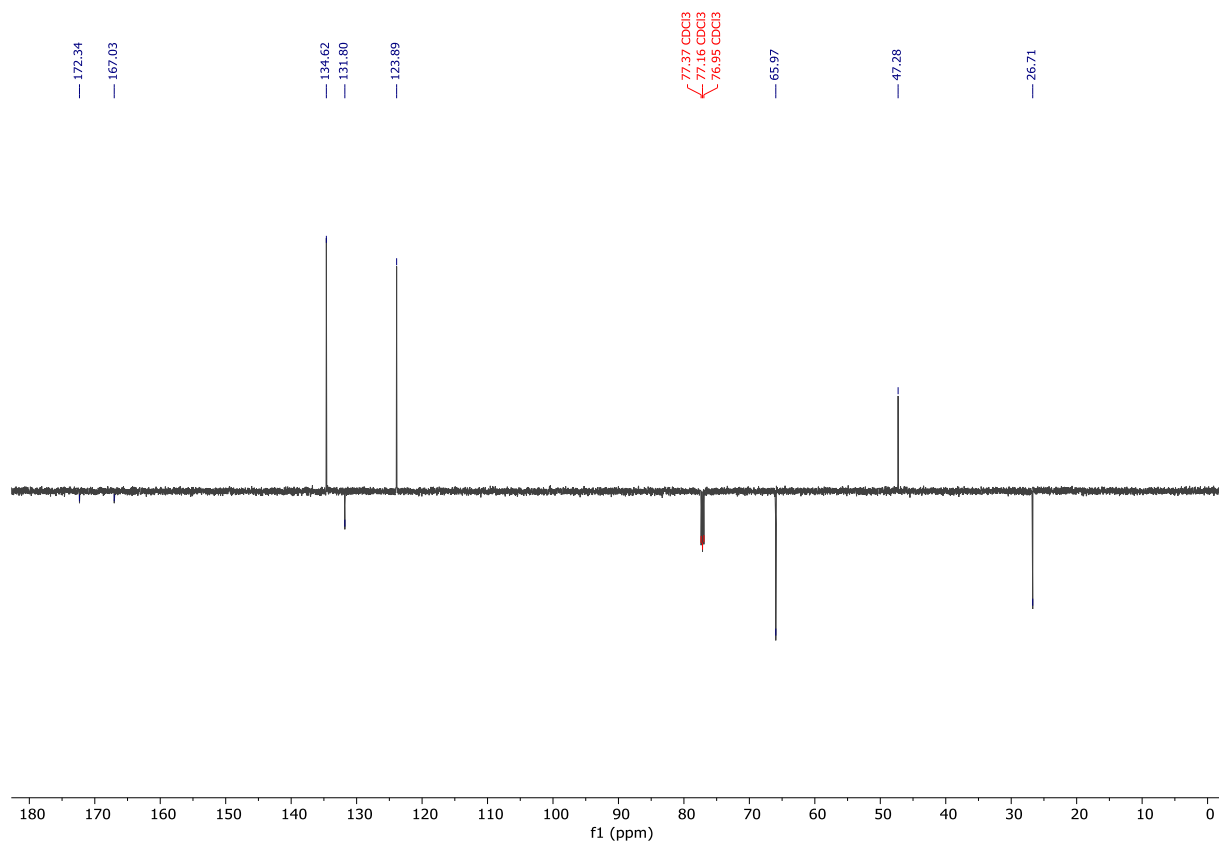


Figure S1.7. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) spectra of compound **2**.

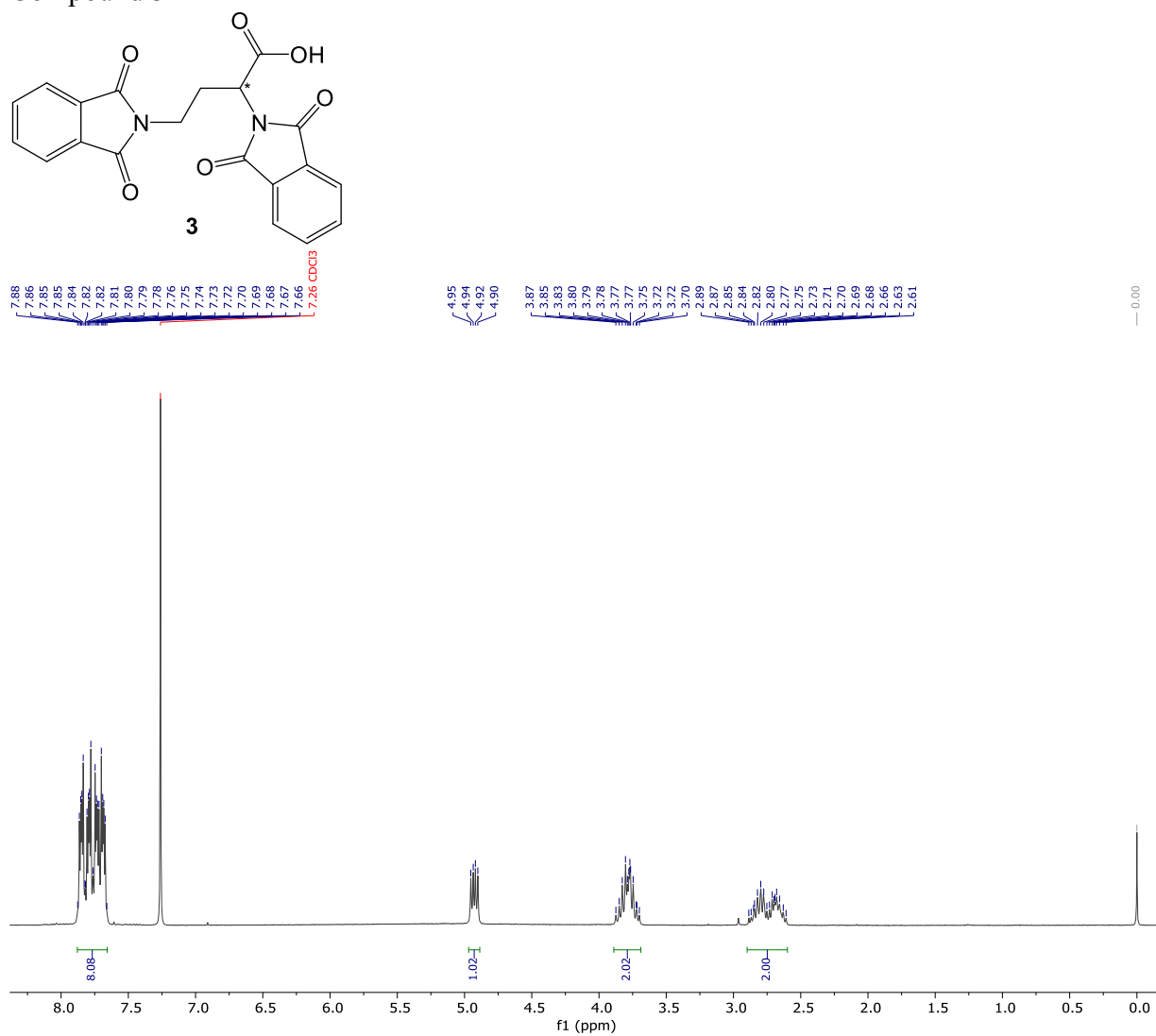


**Figure S1.8.**  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 75 MHz) spectra of compound **2**.



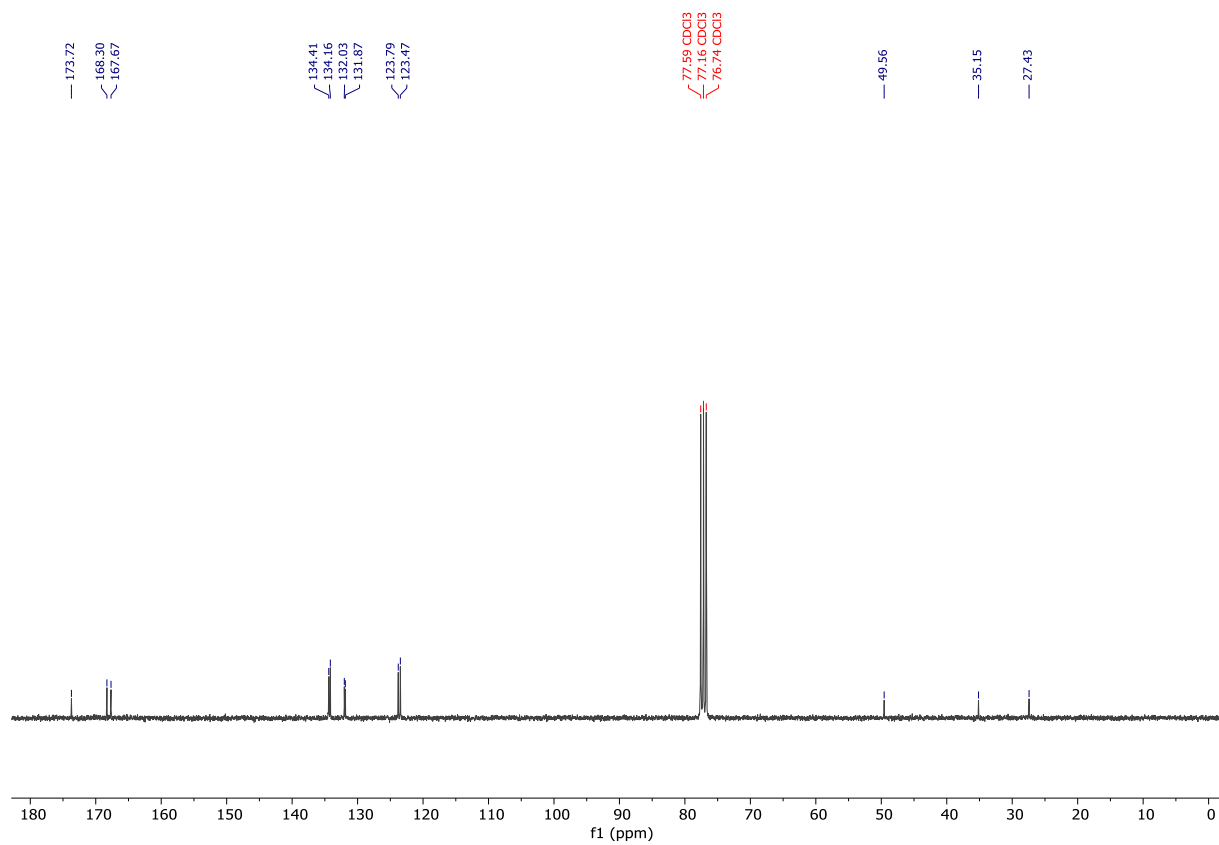
**Figure S1.9.**  $^{13}\text{C}$  APT NMR ( $\text{CDCl}_3$ , 75 MHz) spectra of compound **2**.

# Compound 3

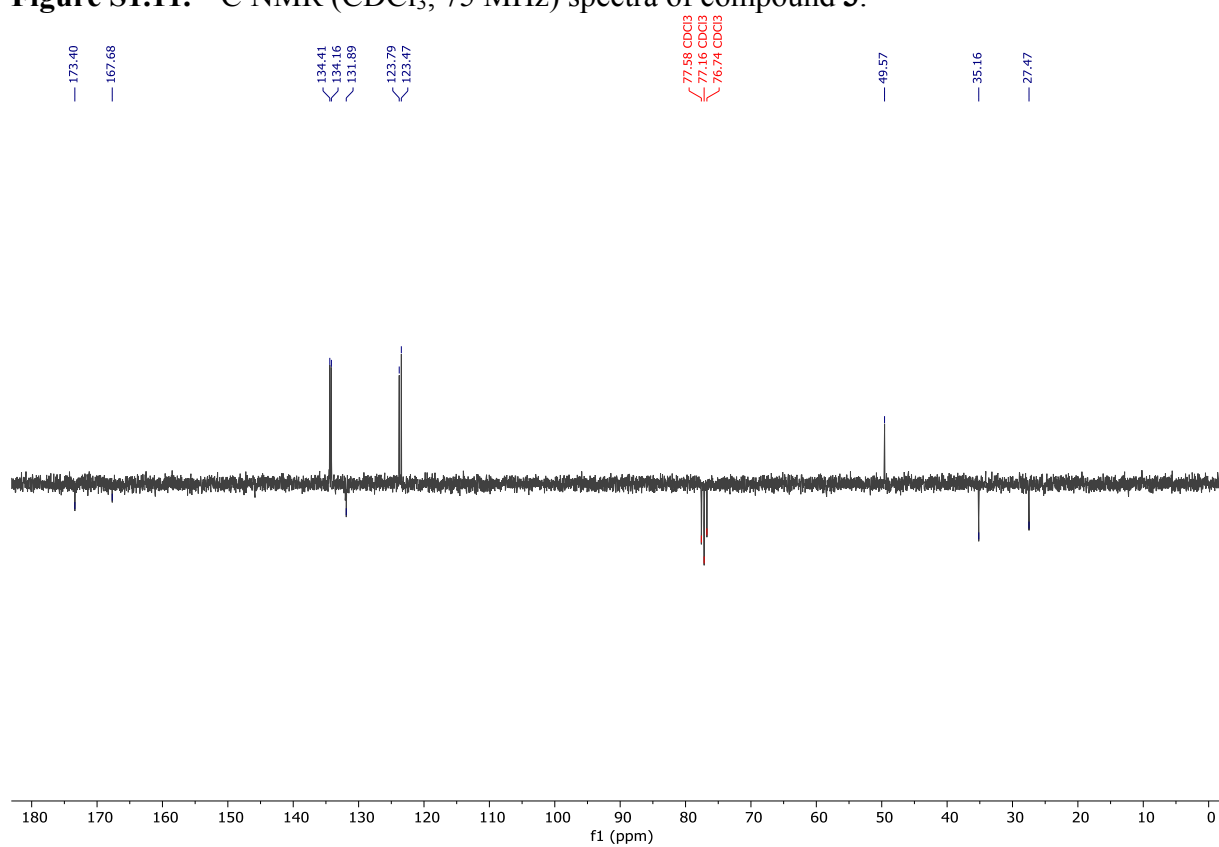


**Figure S1.10.** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) spectra of compound 3.



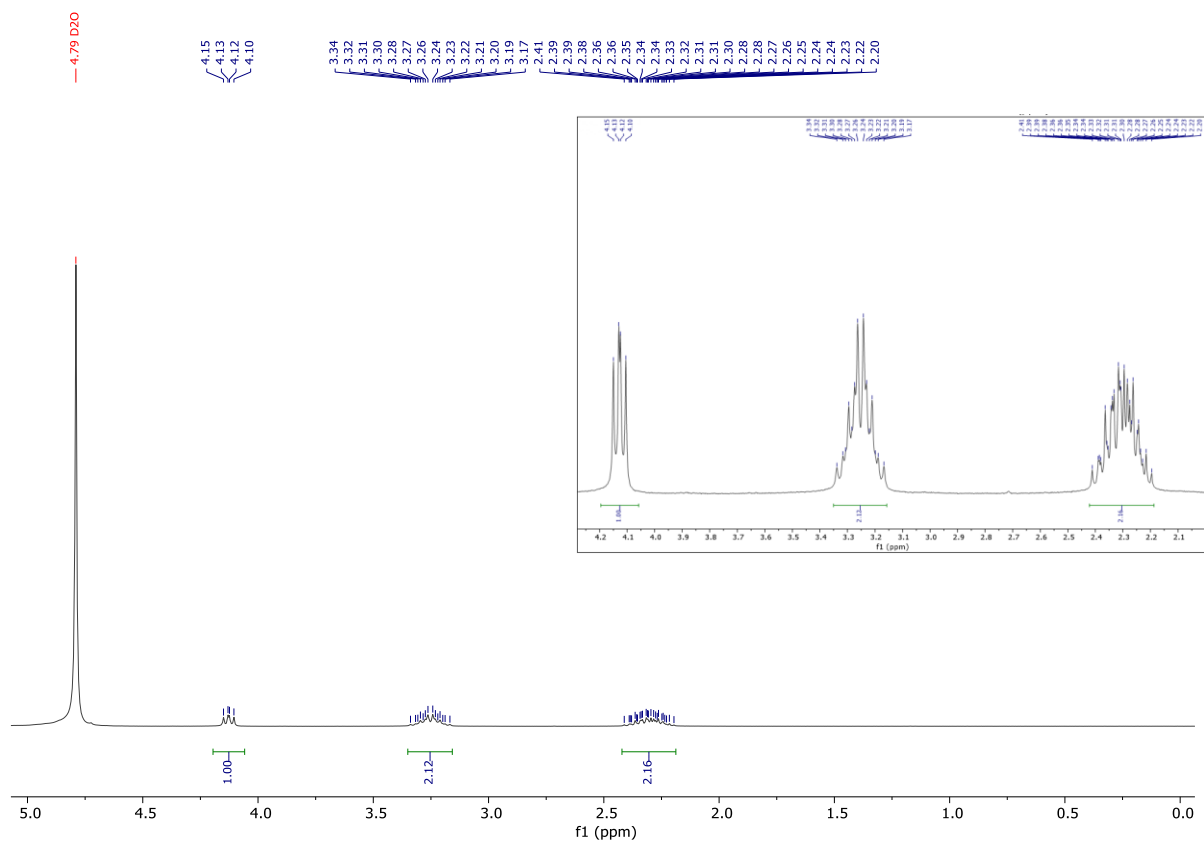
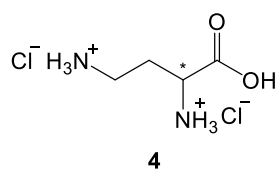


**Figure S1.11.**  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 75 MHz) spectra of compound **3**.

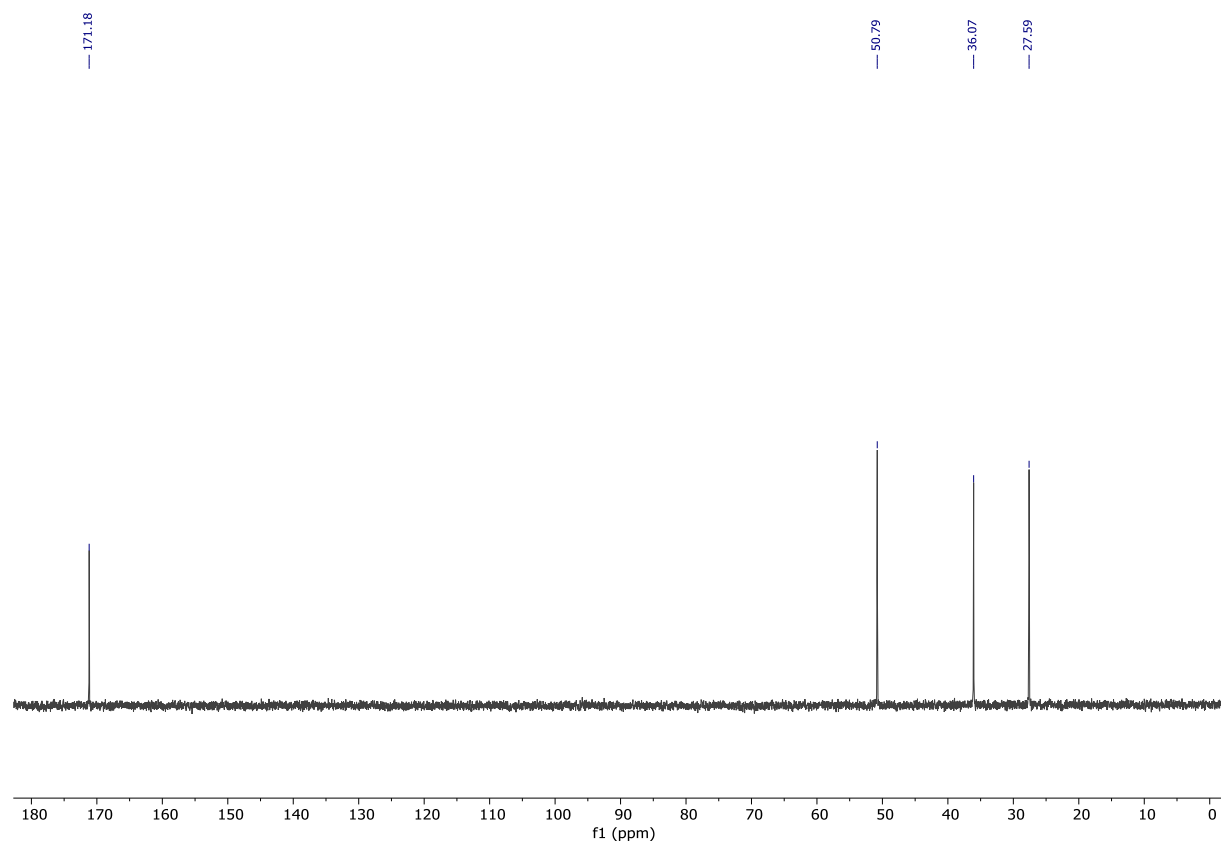


**Figure S1.12.**  $^{13}\text{C}$  APT NMR ( $\text{CDCl}_3$ , 75 MHz) spectra of compound **3**.

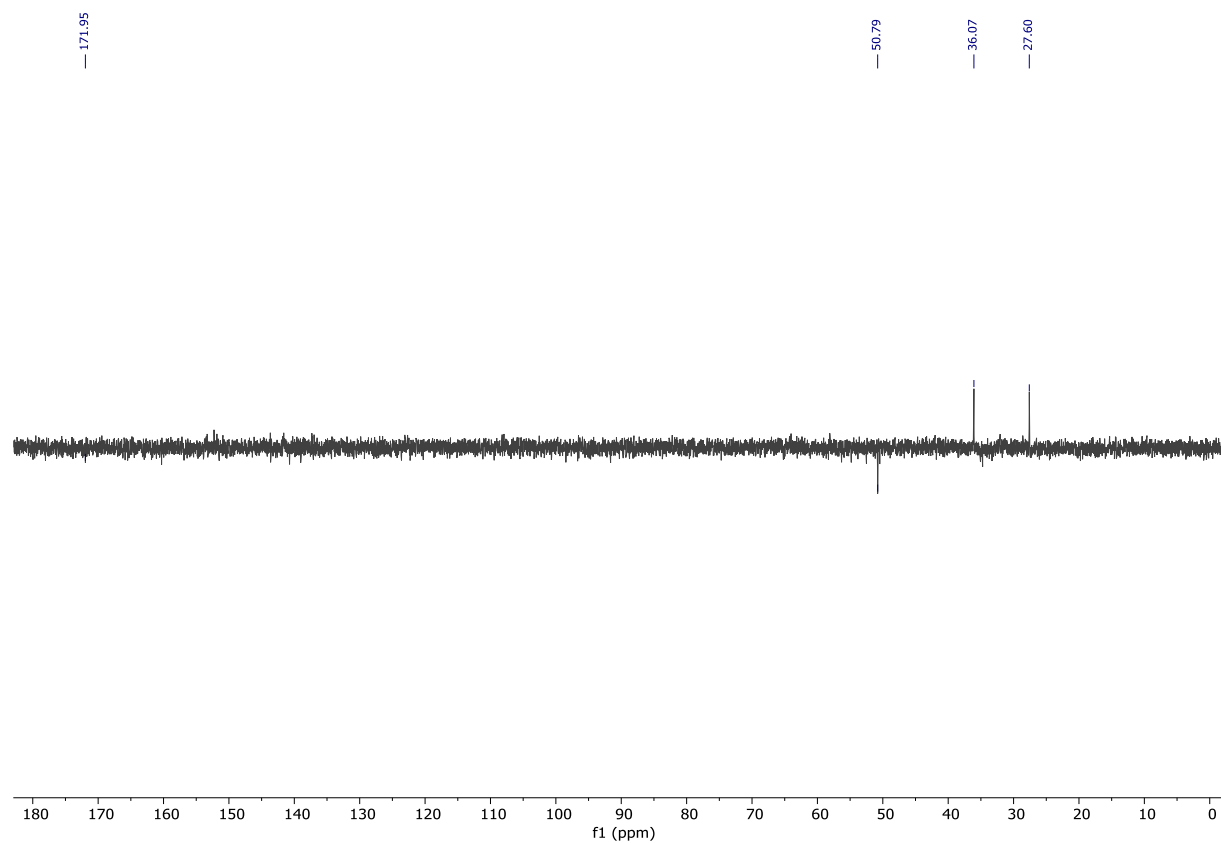
# Compound 4



**Figure S1.13.**  $^1\text{H}$  NMR ( $\text{CDCl}_3$ , 300 MHz) spectra of compound **4**.

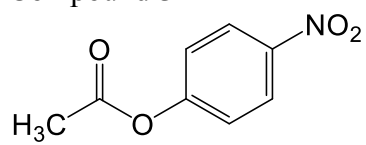


**Figure S1.14.**  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 75 MHz) spectra of compound **4**.

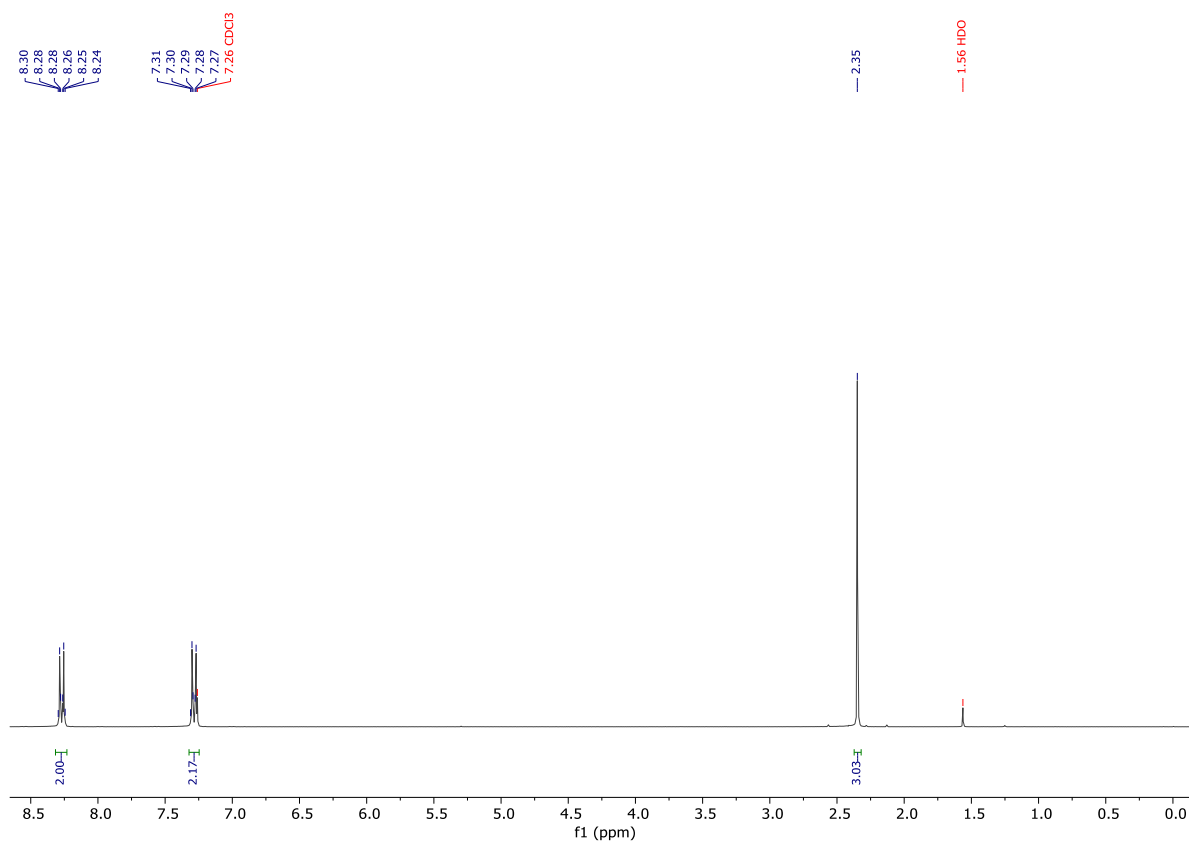


**Figure S1.15.**  $^{13}\text{C}$  APT NMR ( $\text{CDCl}_3$ , 75 MHz) spectra of compound **4**.

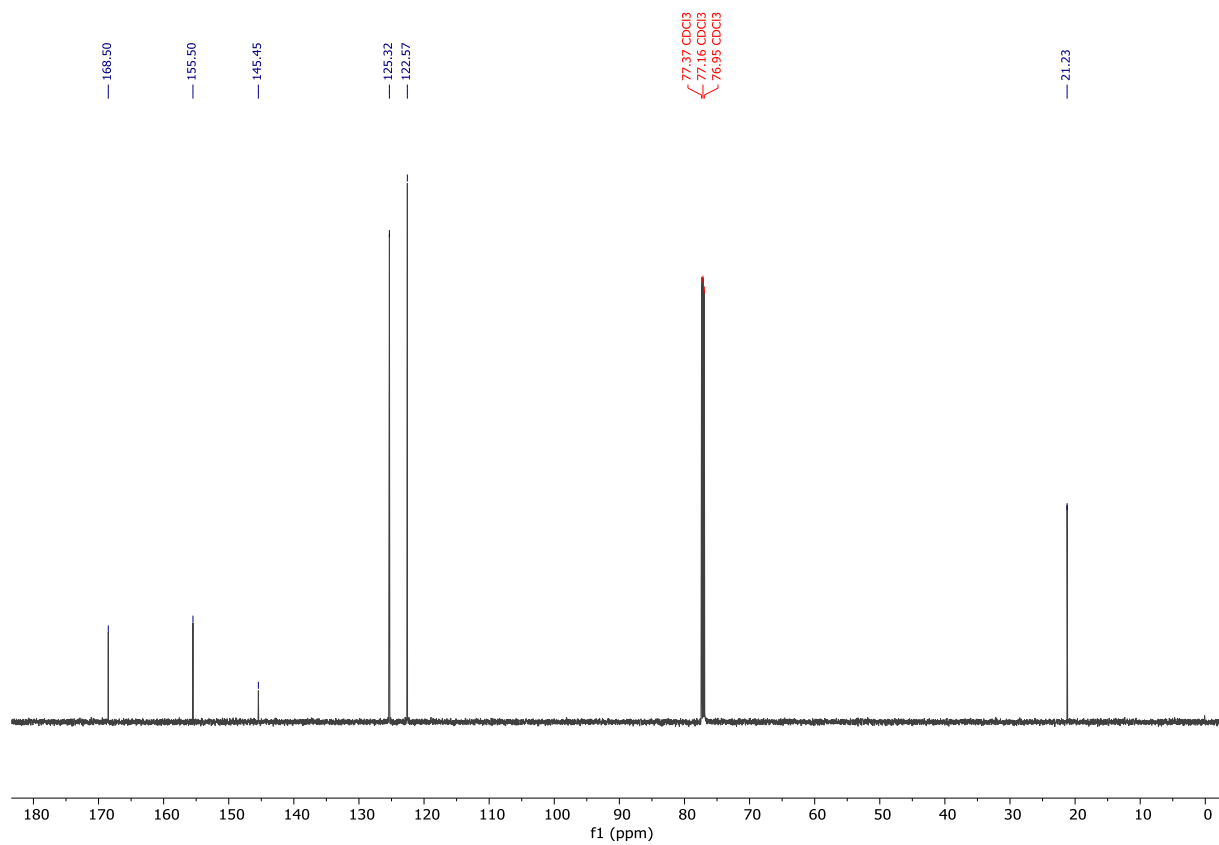
Compound **5**



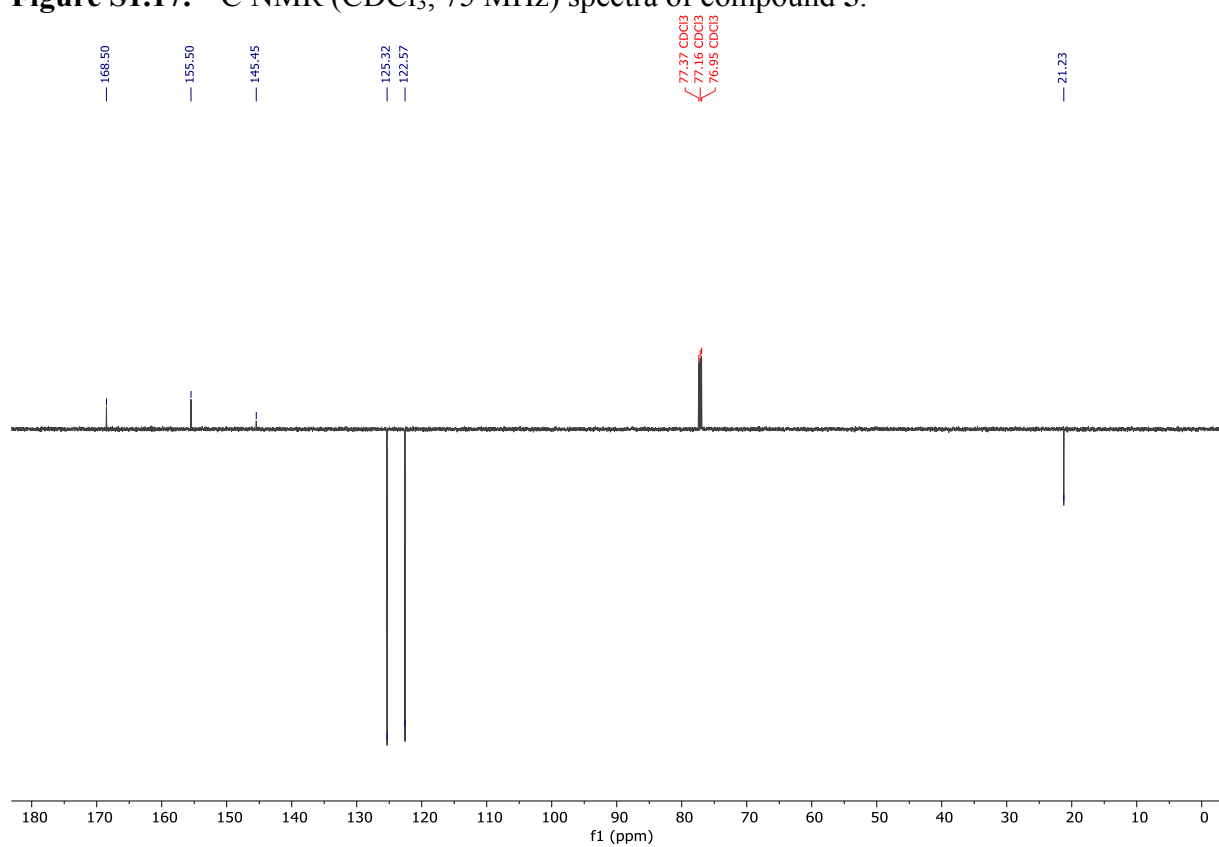
**5**



**Figure S1.16.** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) spectra of compound **5**.

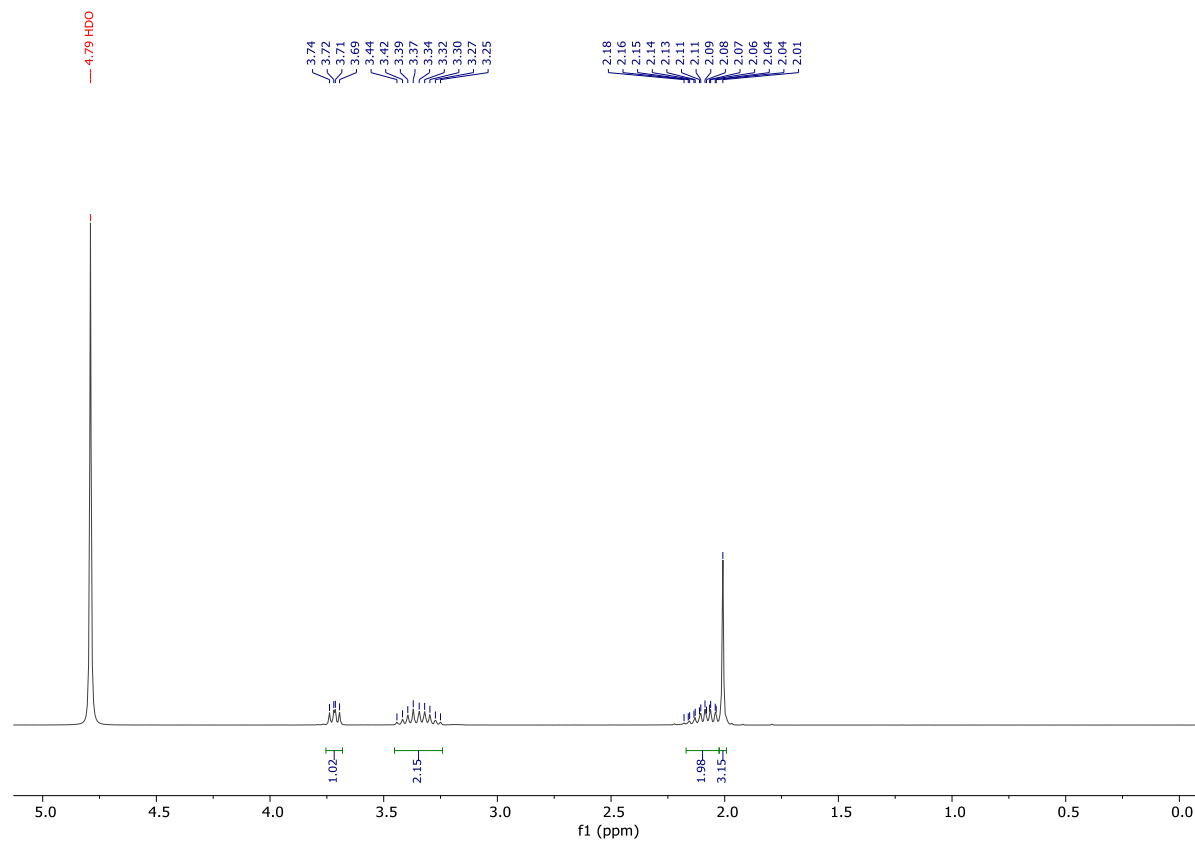
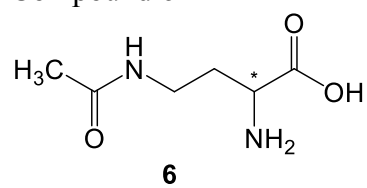


**Figure S1.17.** <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz) spectra of compound **5**.

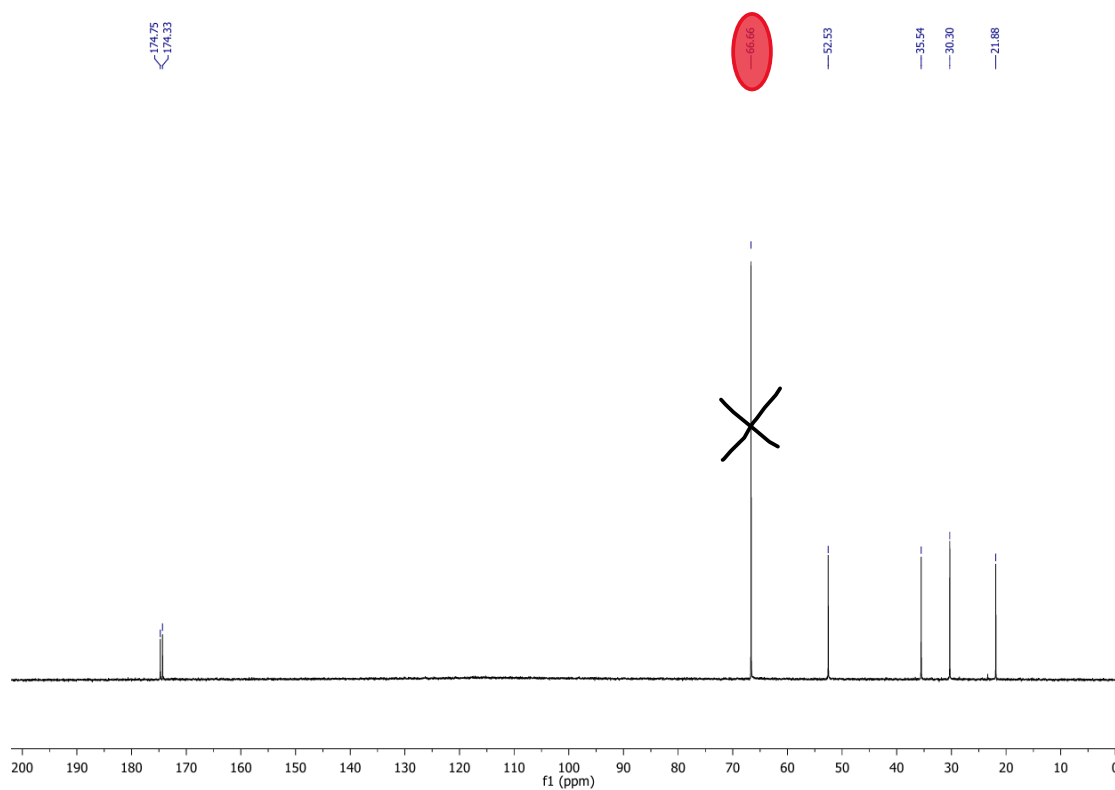


**Figure S1.18.** <sup>13</sup>C APT NMR (CDCl<sub>3</sub>, 75 MHz) spectra of compound **5**.

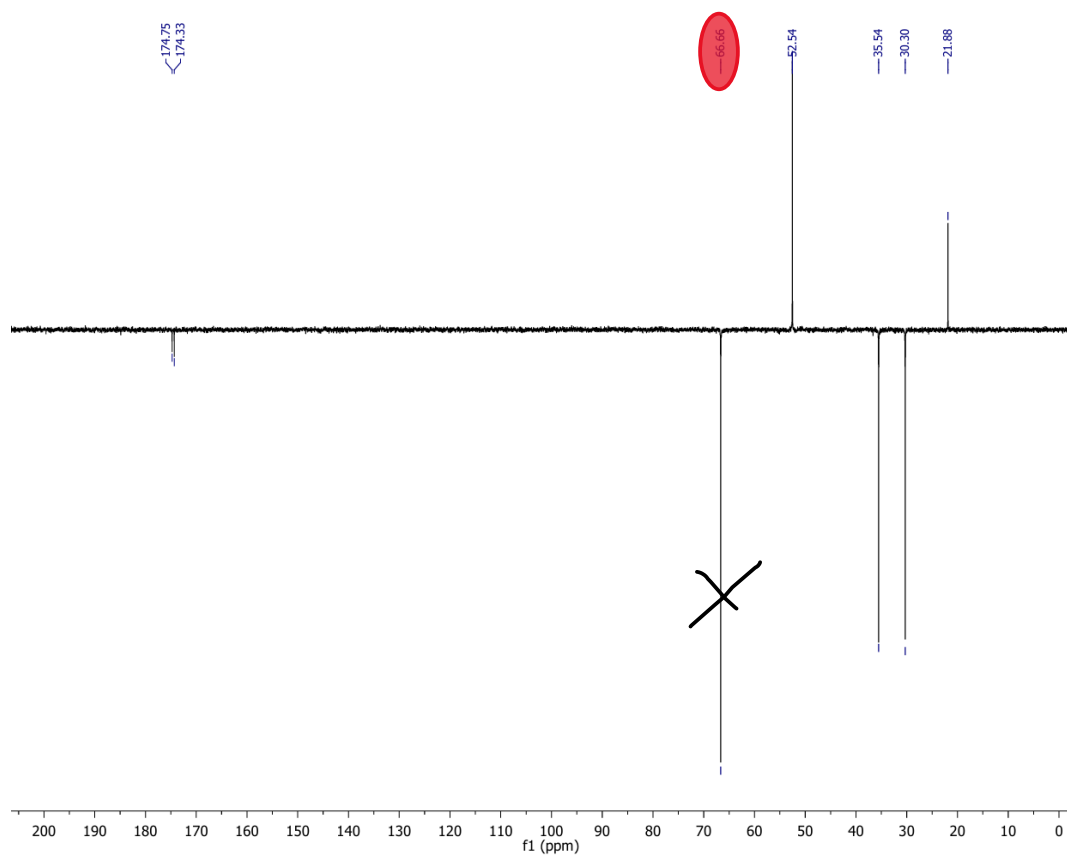
Compound **6**



**Figure S1.19.**  $^1\text{H}$  NMR ( $\text{D}_2\text{O}_3$ , 300 MHz) spectra of compound **6**.

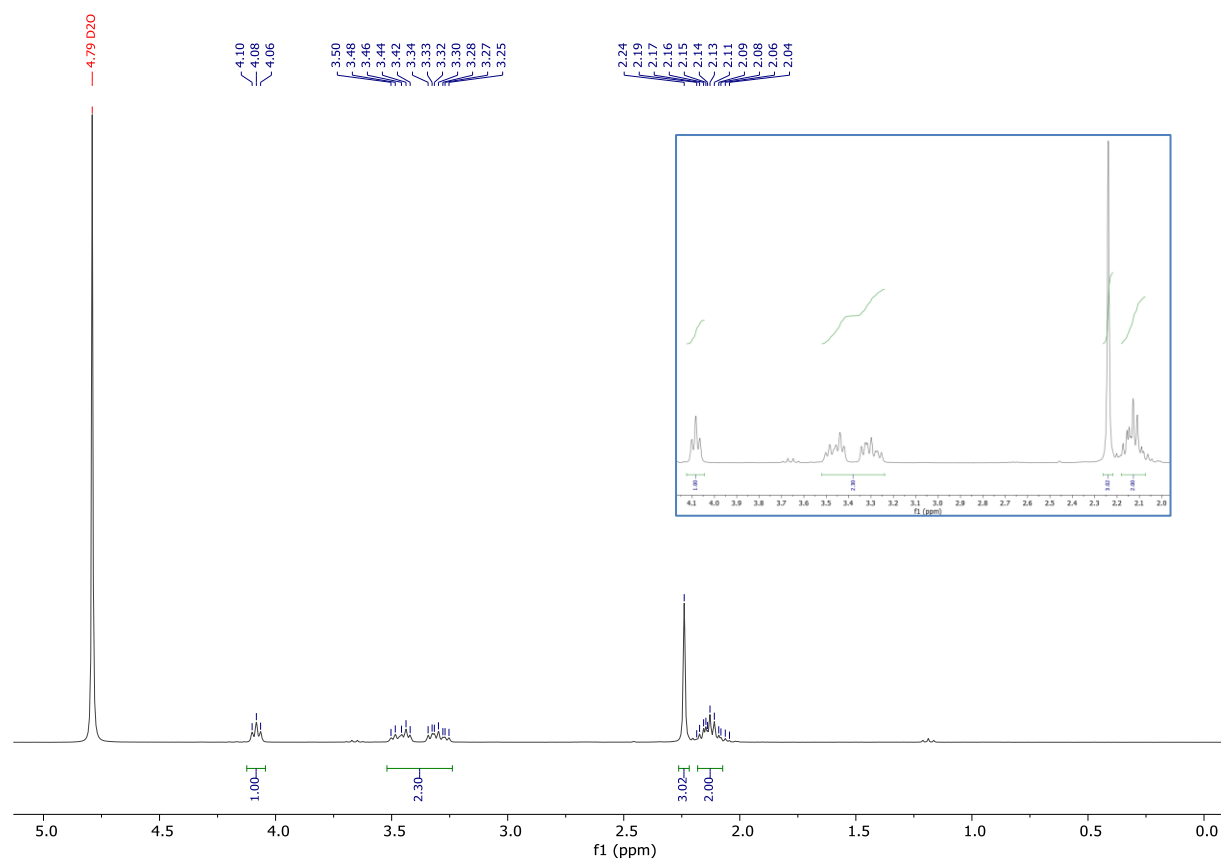
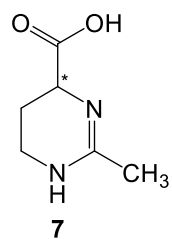


**Figure S1.20.**  $^{13}\text{C}$  NMR ( $\text{D}_2\text{O}$ +dioxane- $d_8$ , 75 MHz) spectra of compound **6**.



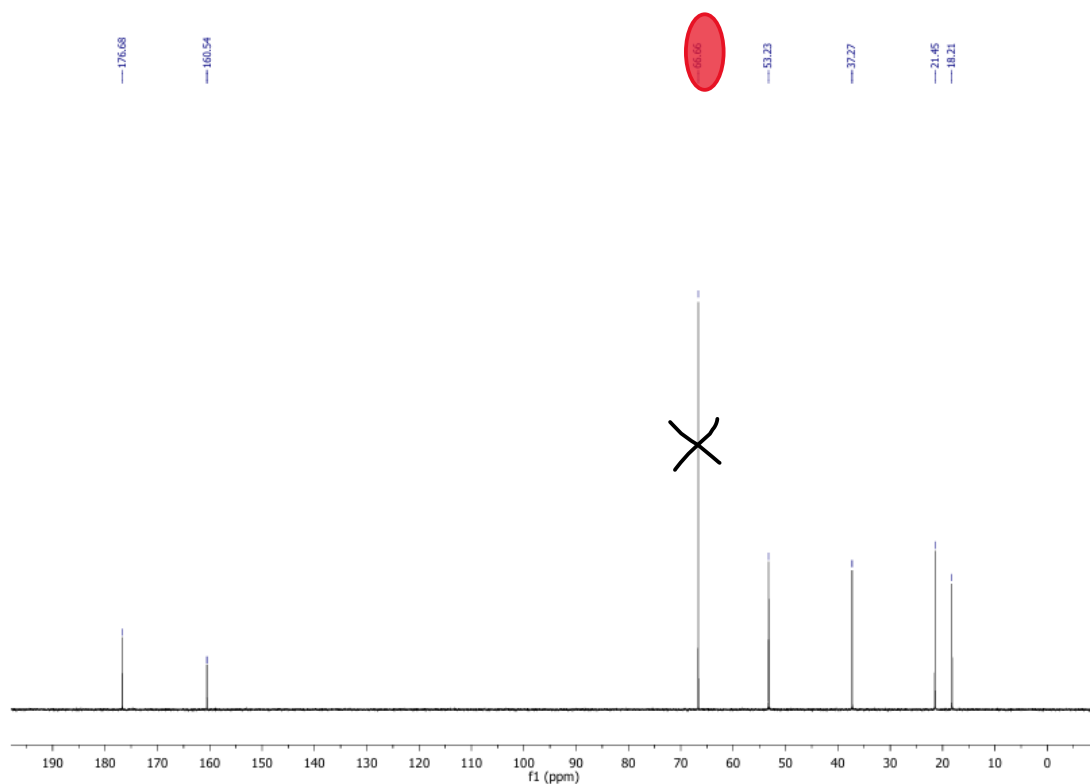
**Slika D1.21.**  $^{13}\text{C}$  APT NMR ( $\text{D}_2\text{O}$ +dioxane- $d_8$ , 75 MHz) spectra of compound **6**.

Compound 7

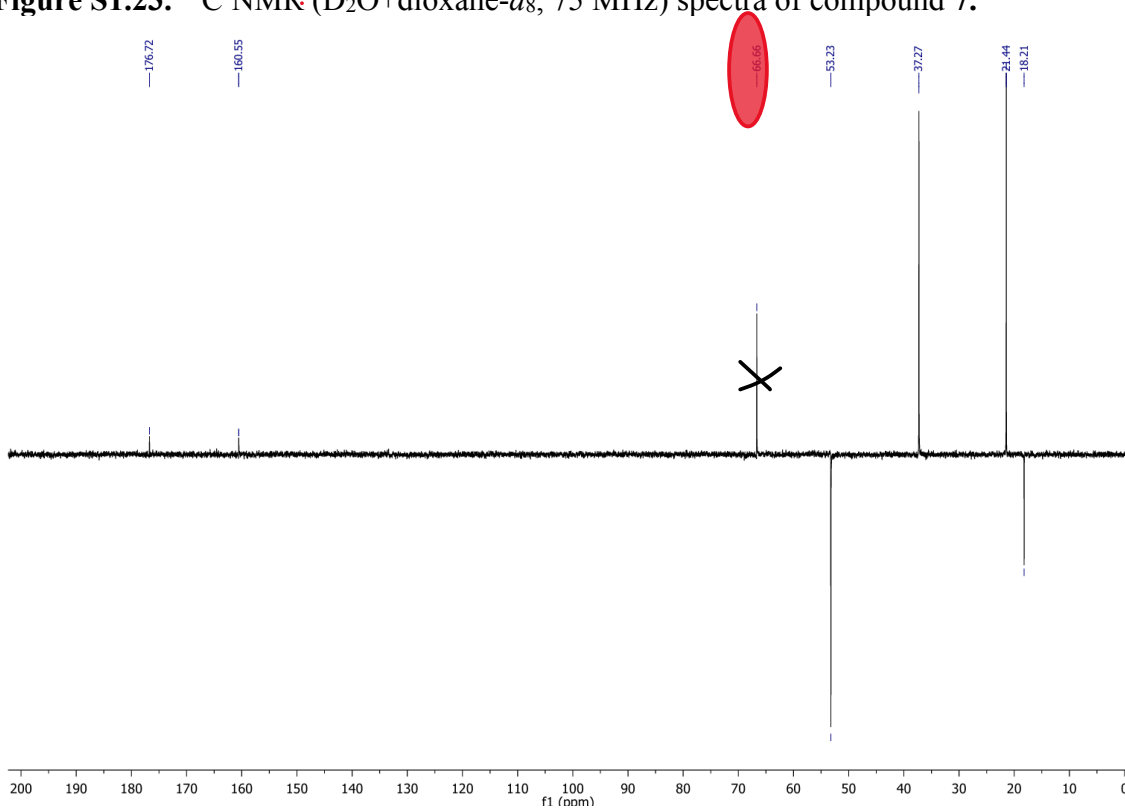


**Figure S1.22.**  $^1\text{H}$  NMR ( $\text{D}_2\text{O}$ , 300 MHz) spectra of compound 7.





**Figure S1.23.**  $^{13}\text{C}$  NMR ( $\text{D}_2\text{O}$ +dioxane- $d_8$ , 75 MHz) spectra of compound **7**.



**Figure S1.24.**  $^{13}\text{C}$  APT NMR ( $\text{D}_2\text{O}$ +dioxane- $d_8$ , 75 MHz) spectra of compound **7**.

## 2. IR spectra

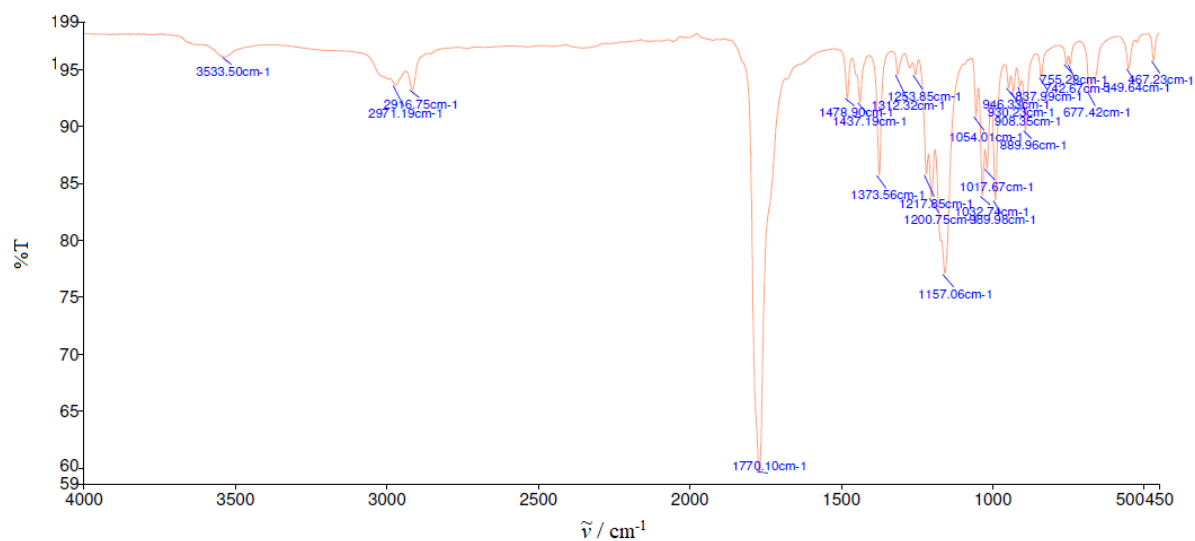


Figure S2.1. IR spectra of compound 1.

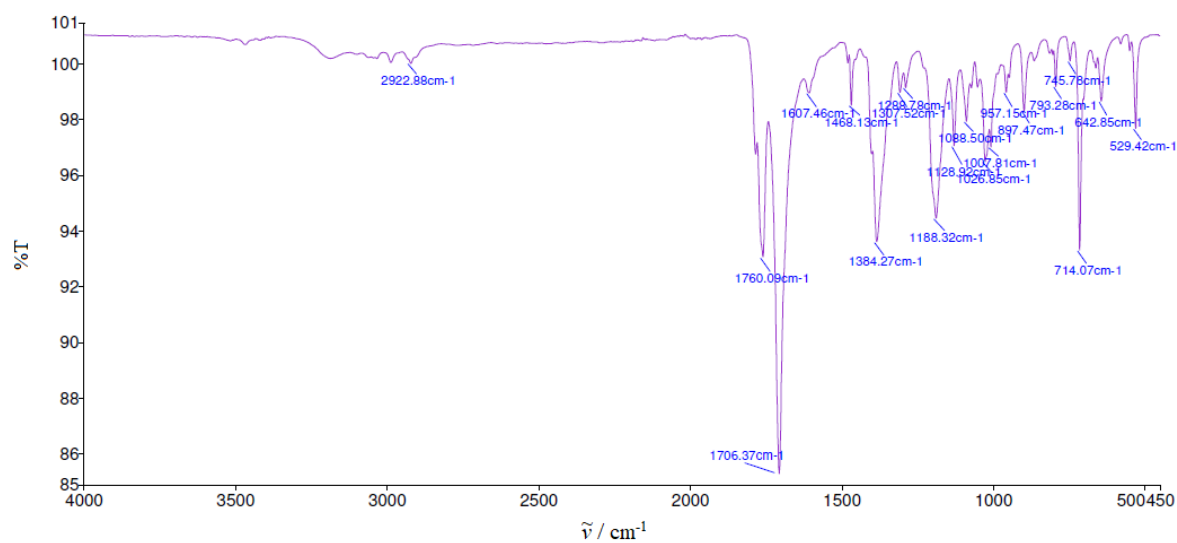
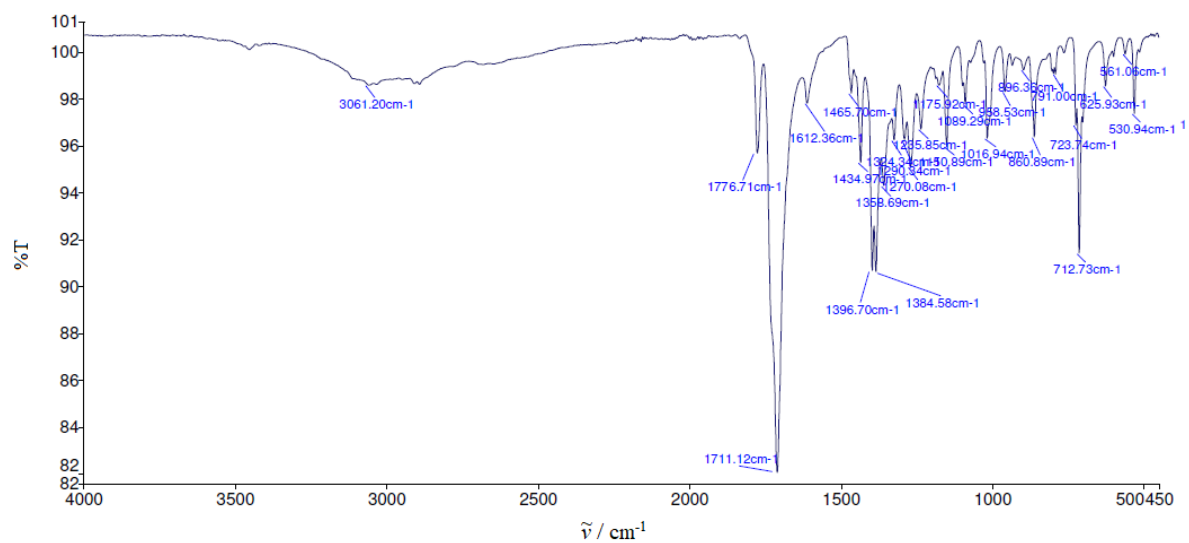
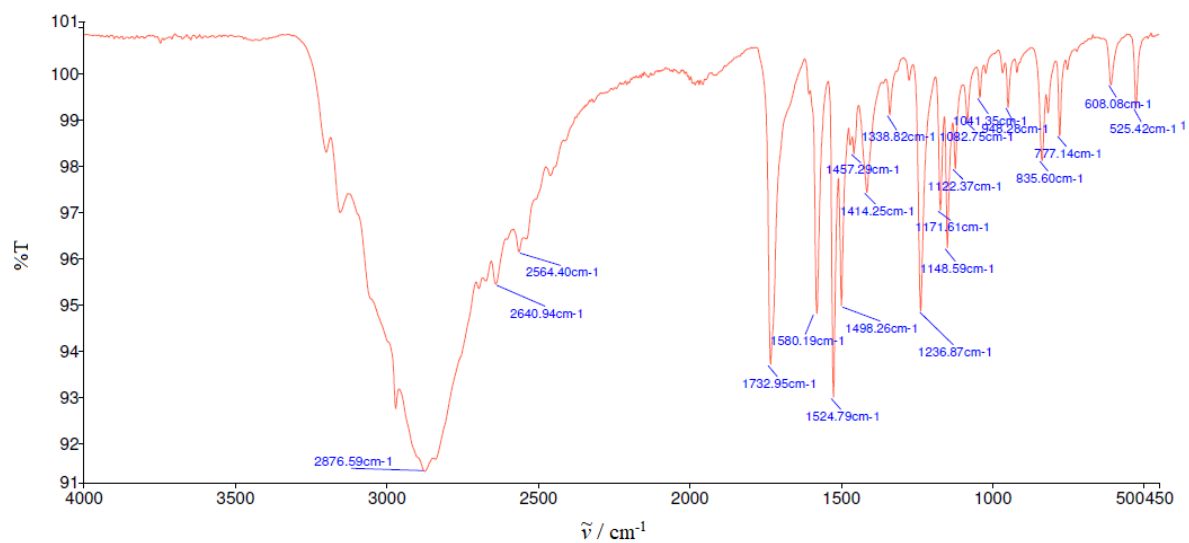


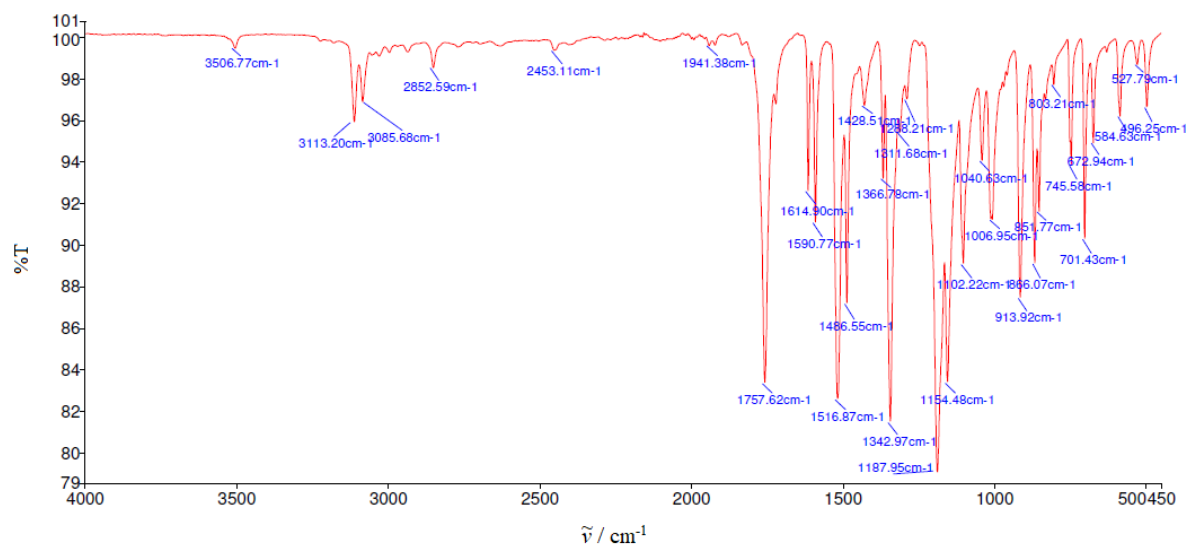
Figure S2.2. IR spectra of compound 2.



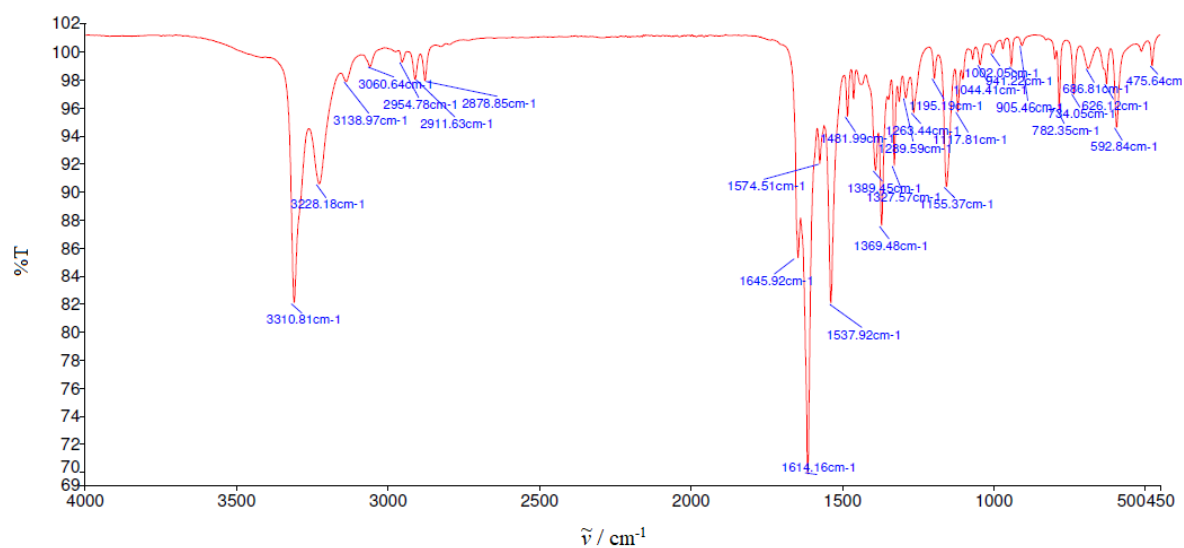
**Figure S2.3.** IR spectra of compound 3.



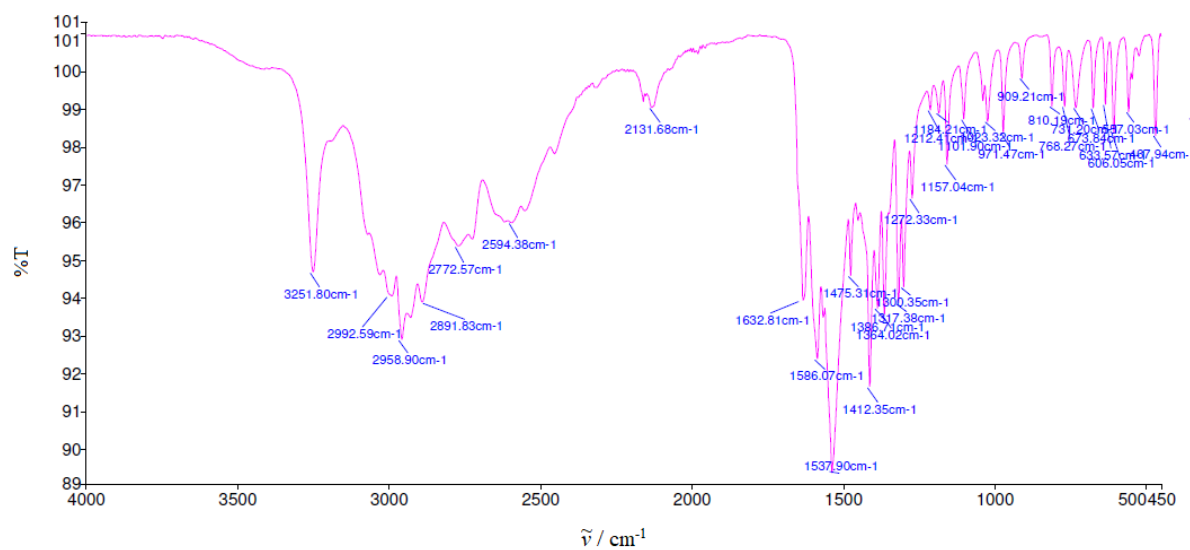
**Figure S2.4.** IR spectra of compound 4.



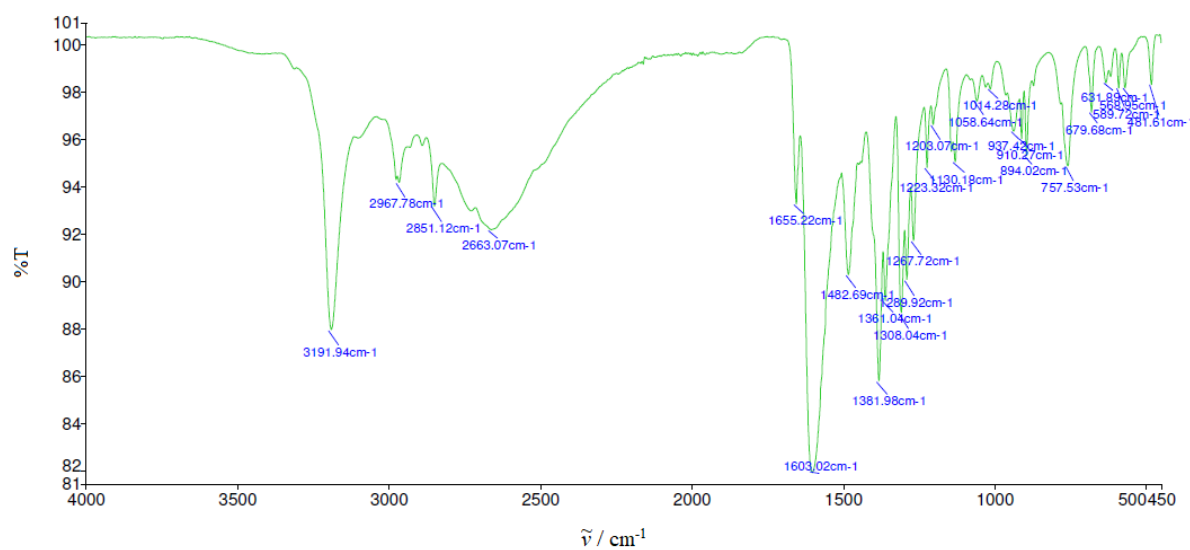
**Figure S2.5.** IR spectra compound **5**.



**Figure S2.6.** IR spectra of complex Cu(II) and compound **6**.



**Figure S2.7.** IR spectra of compound 6.



**Figure S2.8.** IR spectra of compound 7.

### 3. Separation of (±)-ectoine enantiomers

The solvents used for analyses were HPLC purity: MeOH, ACN, Milli-Q H<sub>2</sub>O. All parameters used are listed in Table S3.1. UV detection was performed at 210 nm. The concentrations of (±)-ectoine used were 0.25 mg/mL. The injection volumes were 10 µL. Commercially available (+)-ectoine was used as a standard.

**Table S1.** Methods and conditions for (±)-ectoine enantioseparation.

Method	Column	Mobile phase*	Flow rate [mL/min]	T [°C]
1	(R,R)-WhelkO1	MeOH/ 0.1 % AcOH	1	25
2	(R,R)-WhelkO1	MeOH:H <sub>2</sub> O = 1:1 / 0.1 % AcOH	1	25
3	(R,R)-WhelkO1	MeOH:H <sub>2</sub> O = 7:3 / 0.1 % AcOH	1	25
4	(R,R)-WhelkO1	ACN:H <sub>2</sub> O = 7:3	1	25
5	(R,R)-WhelkO1	ACN:H <sub>2</sub> O = 15:85	1	25
6	Chiralcel OJ-RH	ACN:H <sub>2</sub> O = 5:95	1	25
7	Chiralcel OJ-RH	MeOH:H <sub>2</sub> O = 1:1/0.1 % AcOH	1	25
8	Chiralcel OJ-RH	ACN:H <sub>2</sub> O = 15:85	1	25
9	Chiralcel OJ-RH	ACN:H <sub>2</sub> O = 15:85/0.1 % AcOH	1	25
10	Chiralcel OJ-RH	ACN:H <sub>2</sub> O = 1:9	1	25
11	Chiralcel OJ-RH	ACN:H <sub>2</sub> O = 3:7	1	25
12	Chiralcel OJ-RH	ACN:H <sub>2</sub> O = 7:3	1	25
13	Chiralcel OD-RH	ACN:H <sub>2</sub> O = 15:85	1	25
14	Chiralcel OD-RH	MeOH/0.1 % AcOH	1	25
15	Chirallica PST-4	MeOH/0.1 % AcOH	1	25
16	Chiral-T	MeOH	1	30
17	Chiral-T	MeOH/0.1 % FA	1	25
18	Chiral-T	MeOH/0.01 % FA	1	25
19	Chiral-T	MeOH 0.01 % TEA	1	25
20	Chiral-T	MeOH:H <sub>2</sub> O = 9:1	1	30
21	Chiral-T	MeOH:H <sub>2</sub> O = 9:1	1	35
22	Chiral-T	MeOH:H <sub>2</sub> O = 9:1	1	40
23	Chiral-T	MeOH:H <sub>2</sub> O = 7.5:2.5	0.5	30
24	Chiral-T	MeOH:H <sub>2</sub> O = 7.5:2.5	0.5	40
25	Chiral-T	MeOH:H <sub>2</sub> O = 7.5:2.5	1	40
26	Chiral-T	MeOH:H <sub>2</sub> O = 7.5:2.5	1.2	40
27	Chiral-T	MeOH:H <sub>2</sub> O = 1:1	0.5	40
28	Chiral-T	MeOH:H <sub>2</sub> O = 1:1	0.75	40
29	Chiral-T	MeOH:H <sub>2</sub> O = 1:1	1	30
30	Chiral-T	MeOH:H <sub>2</sub> O = 1:1	1	35
31	Chiral-T	MeOH:H <sub>2</sub> O = 1:1	1	40
32	Chiral-T	MeOH:H <sub>2</sub> O = 1:1	1	45
33	Chiral-T	MeOH:NH <sub>4</sub> COOH (25 mM, pH = 3.0) = 7:3	1	40
34	Chiral-T	MeOH:NH <sub>4</sub> COOH (25 mM, pH = 3.0) = 1:1	1	40
35	Chiral-T	ACN:H <sub>2</sub> O = 8:2	1	30

\*volume ratio v/v

## 4. Detection of (+)-ectoine in fermentation broths of salt-tolerant marine *Streptomyces*

### 4.1. *Streptomyces* strains

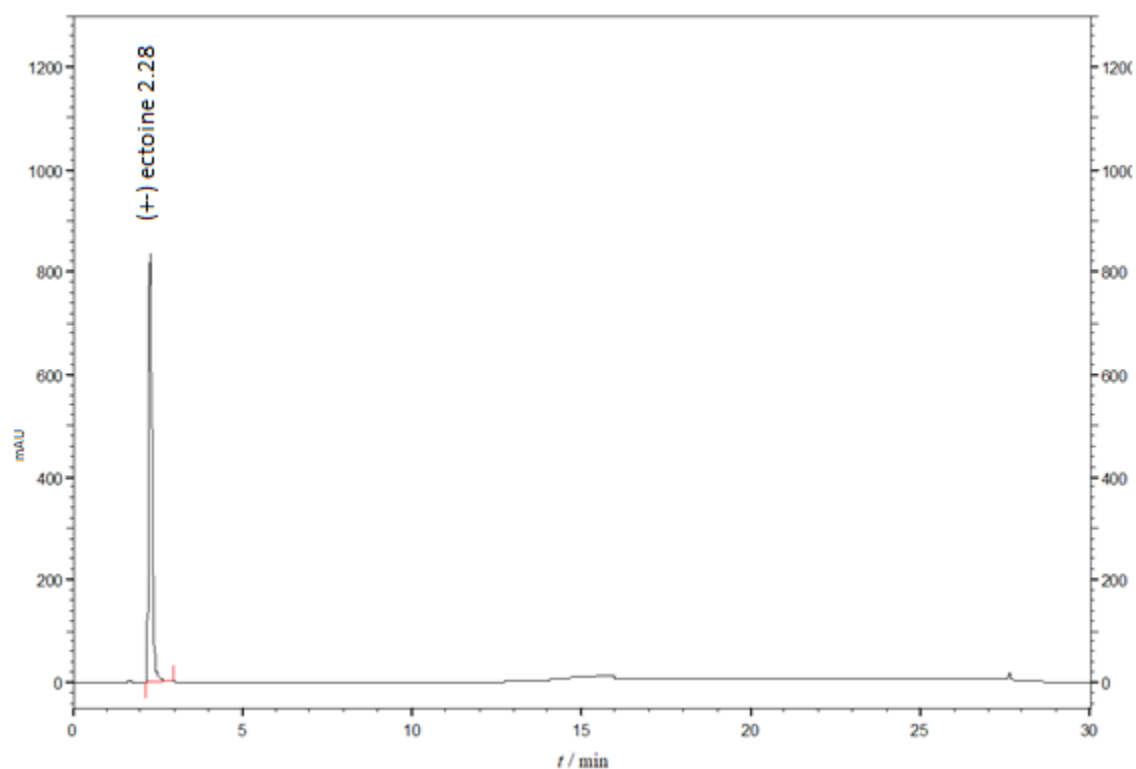
**Table S2.** *Streptomyces* strains isolated from marine invertebrates and sediments used in this study.

Strain	Closest match <sup>a</sup>	Identity (%)	(Gene Bank Acc. No.) <sup>b</sup>	Type of sample	Sampling site	Sampling date
<b>BC81</b>	<i>Streptomyces hydrogenans</i> strain Kris3	100	PP784949	Sponge – <i>Geodia cydonium</i>	Adriatic Sea, Croatia, Dugi otok, Dragove 44°07'10.8"N 14°55'43.5"E	14.07.2019
<b>BC121</b>	<i>Streptomyces hydrogenans</i> strain Kris3	100	PP784953	Tunicate – <i>Phallusia mammillata</i>	Adriatic Sea, Croatia, Zadar, Tankerkomerc Marina 44°07'08.8"N 15°13'41.9"E	20.02.2020
<b>BC123</b>	<i>Streptomyces</i> sp. strain BSP1	100	PP784954	Tunicate – <i>Phallusia mammillata</i>		
<b>BC126</b>	<i>Streptomyces diastaticus</i> subsp. <i>diastaticus</i> strain YNF17-1	100	PP784955	Tunicate – <i>Phallusia mammillata</i>	Adriatic Sea, Croatia, Zaton, Šepurine 44°12'42.3"N 15°09'31.3"E	
<b>BC131</b>	<i>Streptomyces anulatus</i> strain NBC 01759	100	PP784956	Mollusca – <i>Mytilus galloprovincialis</i>	Adriatic Sea, Croatia, Zadar, Uvala Bregdeti Marina 44°05'58.4"N 15°14'56.3"E	18.02.2020
<b>BC104</b>	<i>Streptomyces diastaticus</i> subsp. <i>diastaticus</i> strain YNF17-1	100	PP784950	Sediment	Adriatic Sea, Croatia, Zaton, Šepurine 44°12'42.3"N 15°09'31.3"E	23.05.2019
<b>BC114</b>	<i>Streptomyces albidoflavus</i> strain R-53649	100	PP784951	Sediment	Adriatic Sea, Croatia, Zadar, Kolovare 44°06'15.5"N 15°14'06.6"E	23.05.2019
<b>BC115</b>	<i>Streptomyces hydrogenans</i> strain Kris3	100	PP784952	Sediment		
<b>BC156</b>	<i>Streptomyces</i> sp. strain HBUM206428	100	PP784957	Sediment	Adriatic Sea, Croatia, Zaton, Šepurine 44°12'42.3"N 15°09'31.3"E	21.02.2020
<b>BC160</b>	<i>Streptomyces</i> sp. strain BSP1	100	PP784958	Sediment		
<b>BC167</b>	<i>Streptomyces</i> sp. JC498	100	PP784959	Sediment		

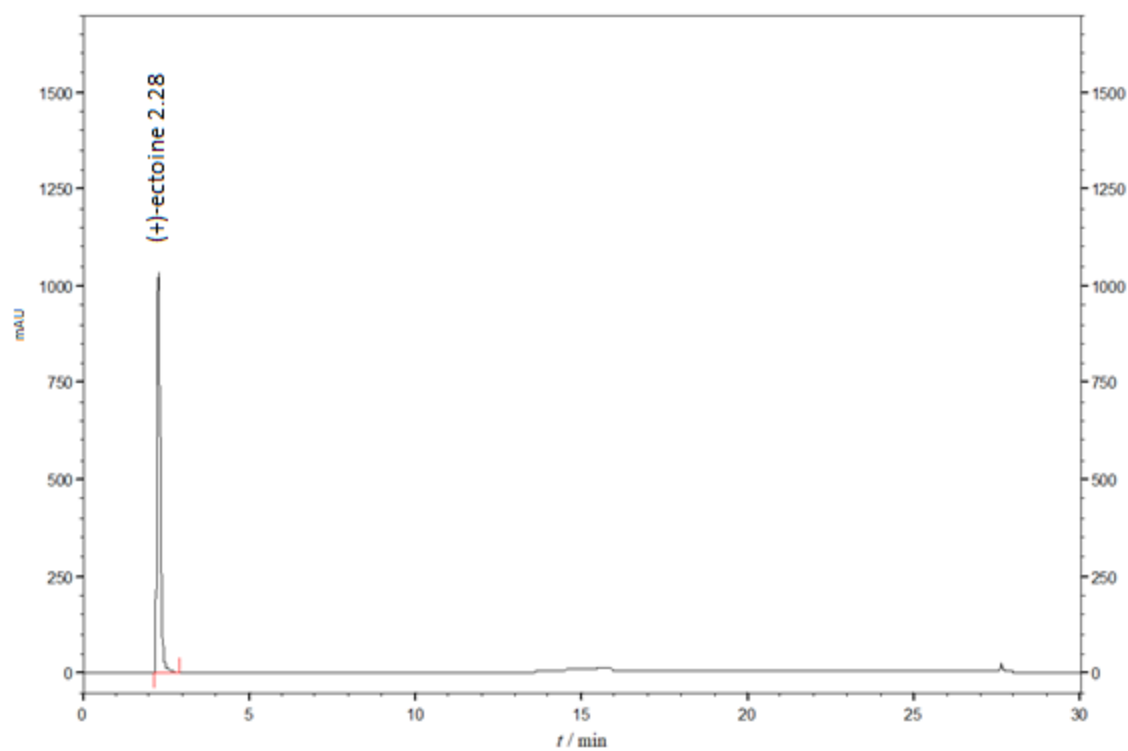
<sup>a</sup> Based on partial 16S rRNA sequence alignment (> 900 bp) with sequences from NCBI BLAST.

<sup>b</sup> The sequences were submitted to GeneBank (Accession number will be added in proof)

#### 4.2. HPLC chromatograms of (+)-ectoine in fermentation broths of salt-tolerant marine *Streptomyces*

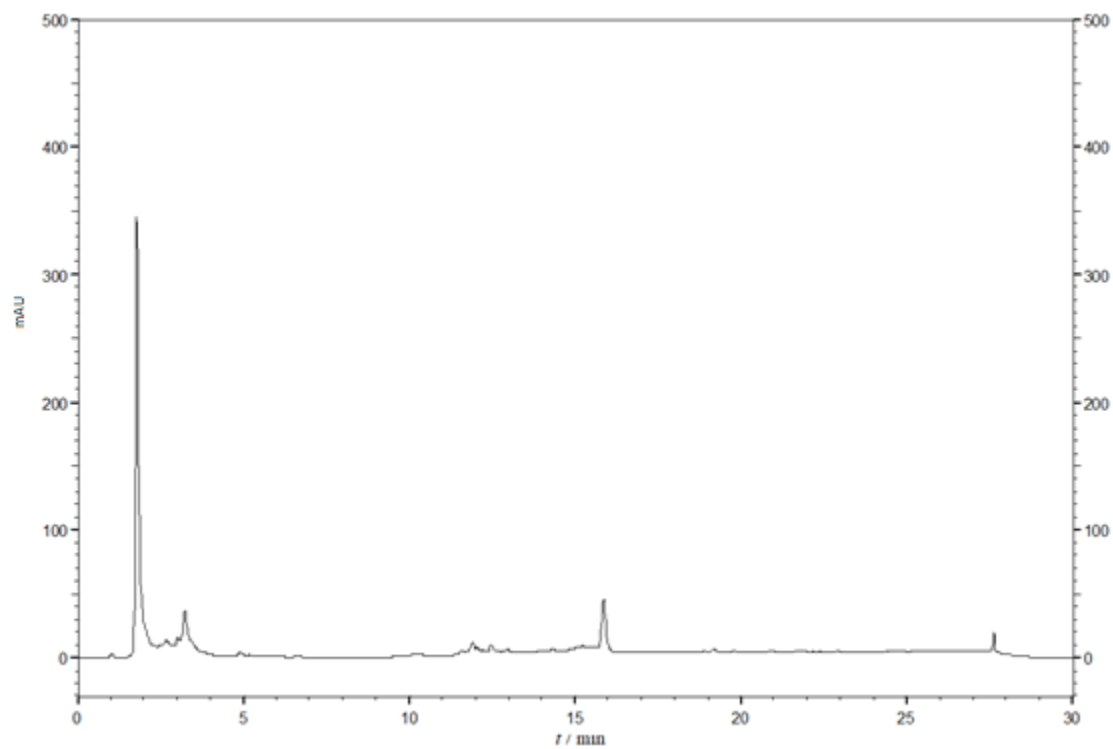


**Figure S4.2.1.** RP-HPLC chromatogram of ( $\pm$ )-ectoine at 220 nm.

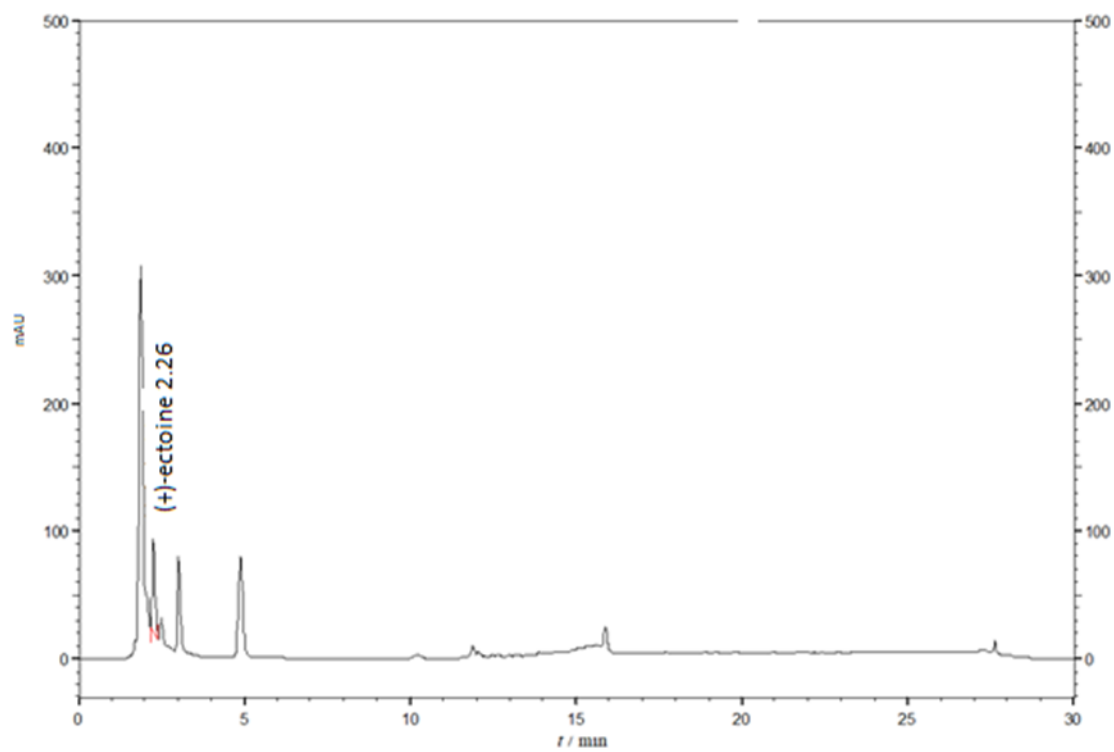


**Figure S4.2.2.** RP-HPLC chromatogram of (+)-ectoine at 220 nm.

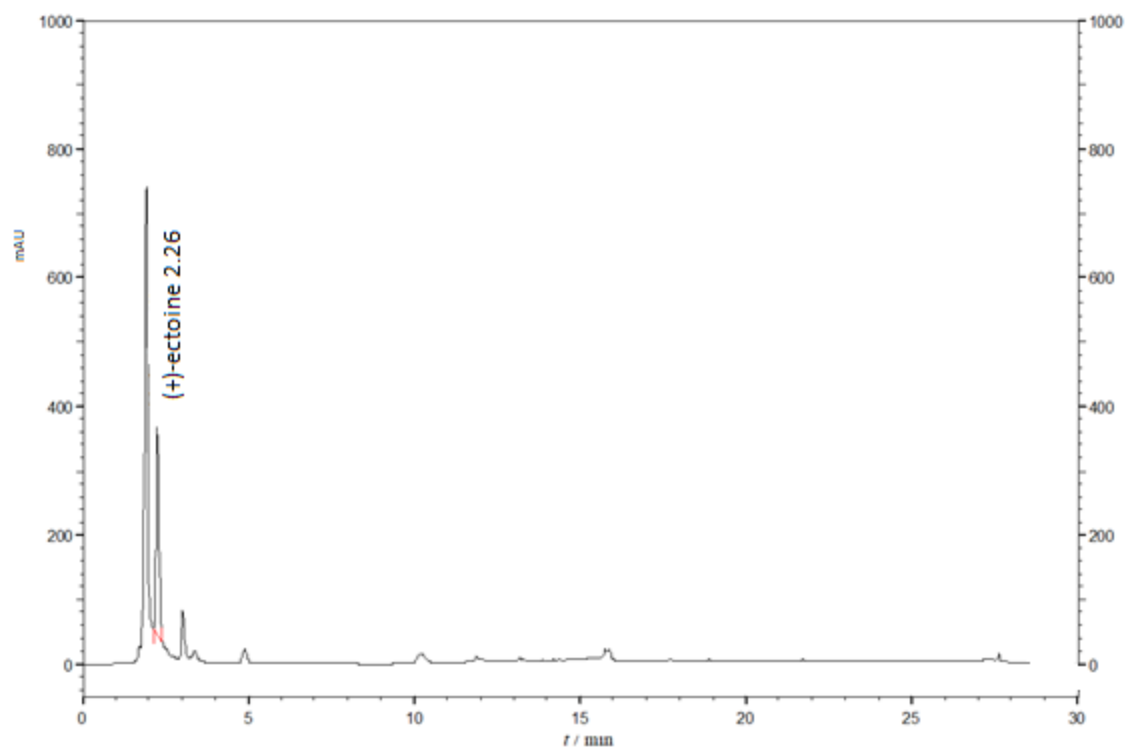




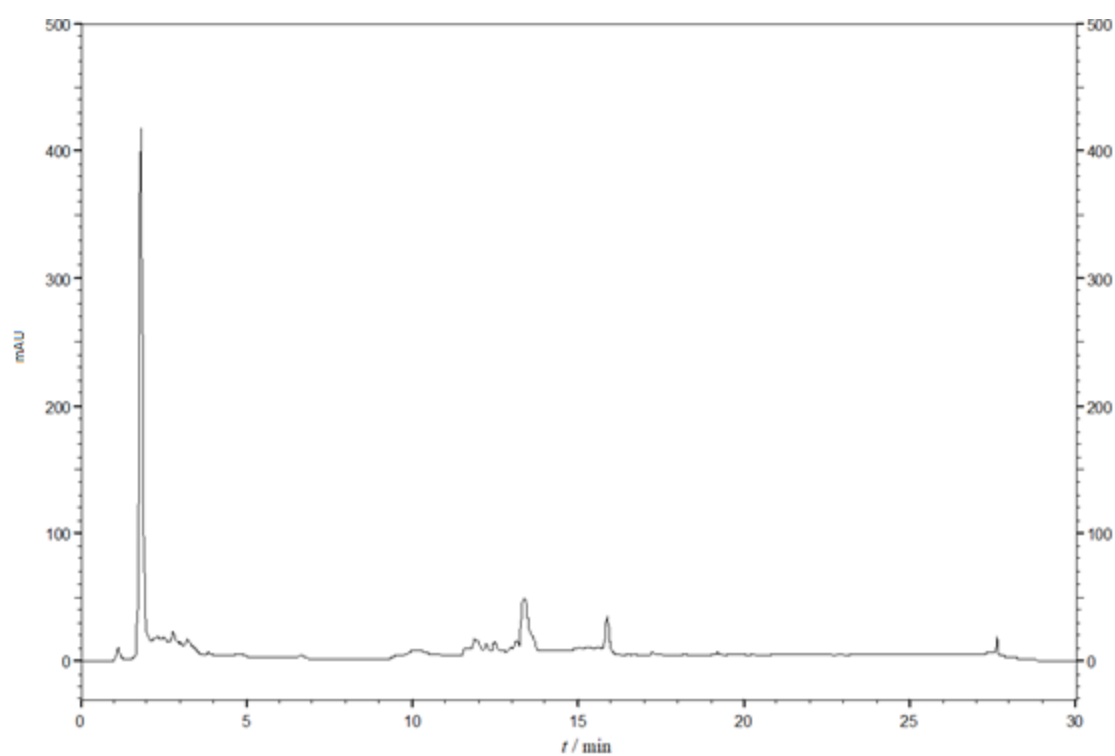
**Figure S4.2.3.** RP-HPLC chromatogram of sample **1** at 220 nm.



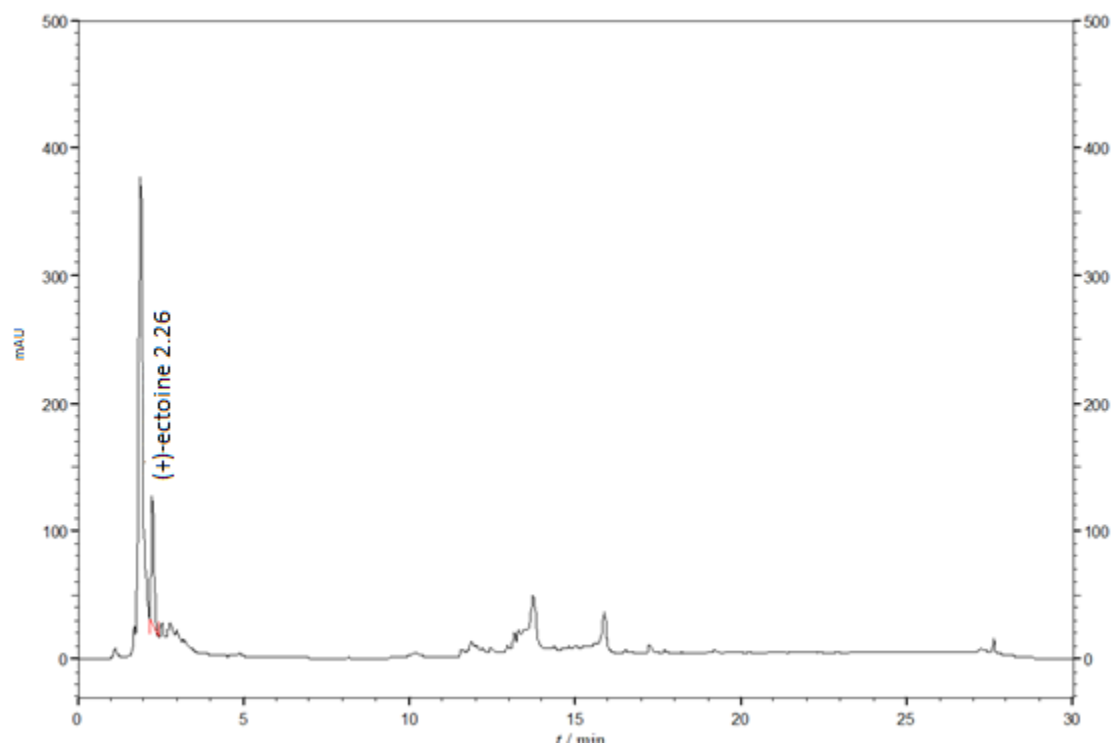
**Figure S4.2.4.** RP-HPLC chromatogram of sample **2** at 220 nm.



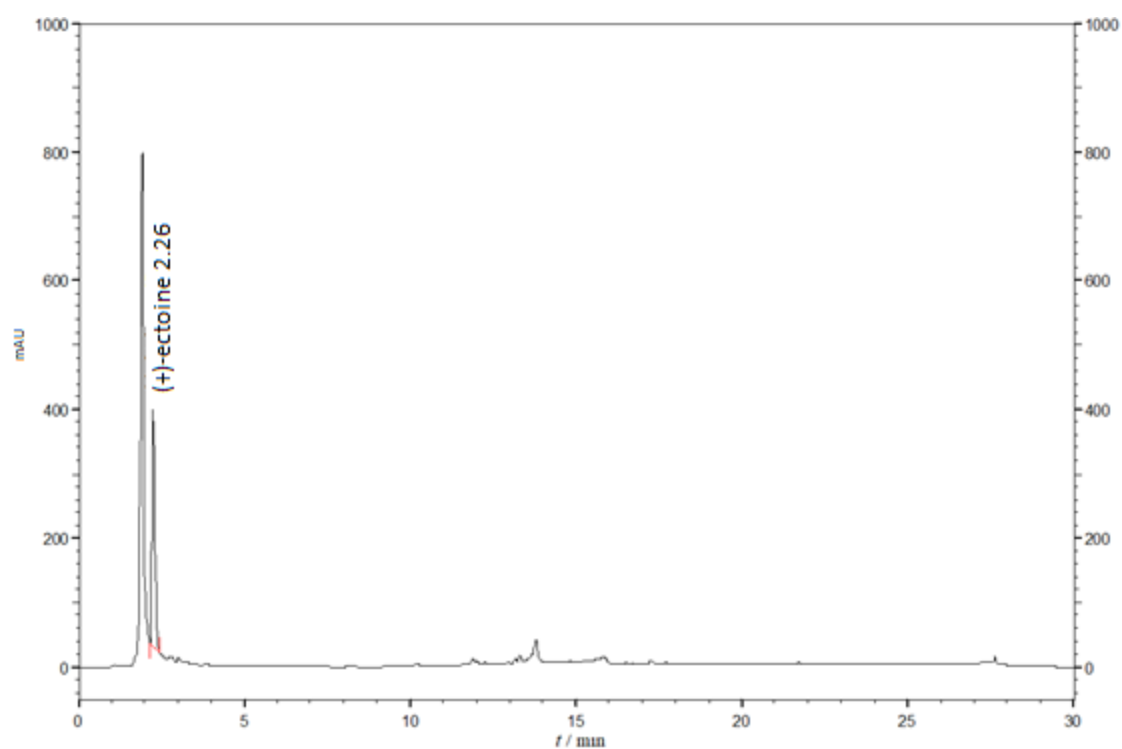
**Figure S4.2.5.** RP-HPLC chromatogram of sample **3** at 220 nm.



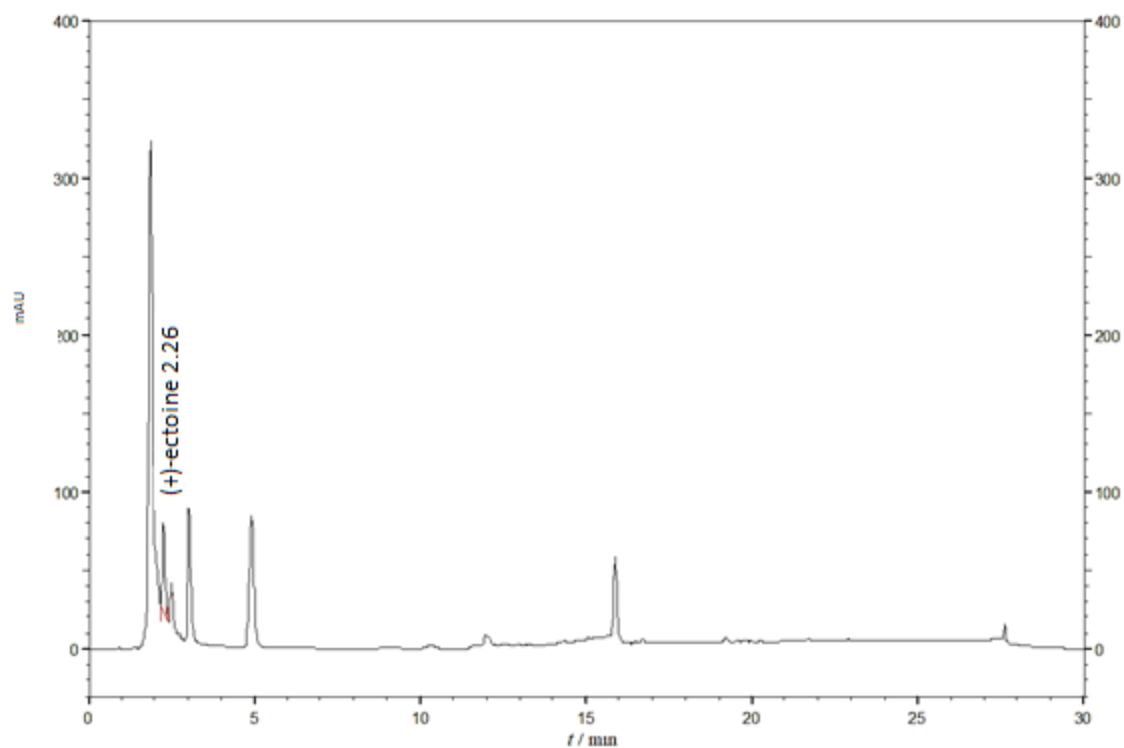
**Figure S4.2.6.** RP-HPLC chromatogram of sample **4** at 220 nm.



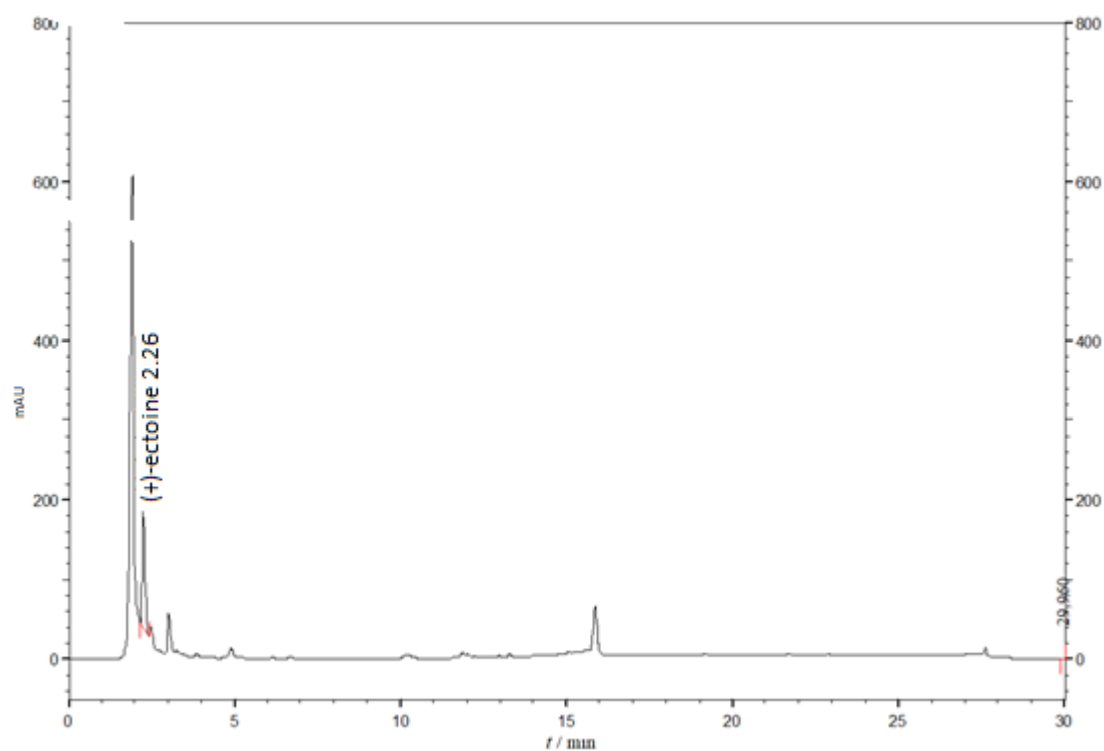
**Figure S4.2.7.** RP-HPLC chromatogram of sample **5** at 220 nm.



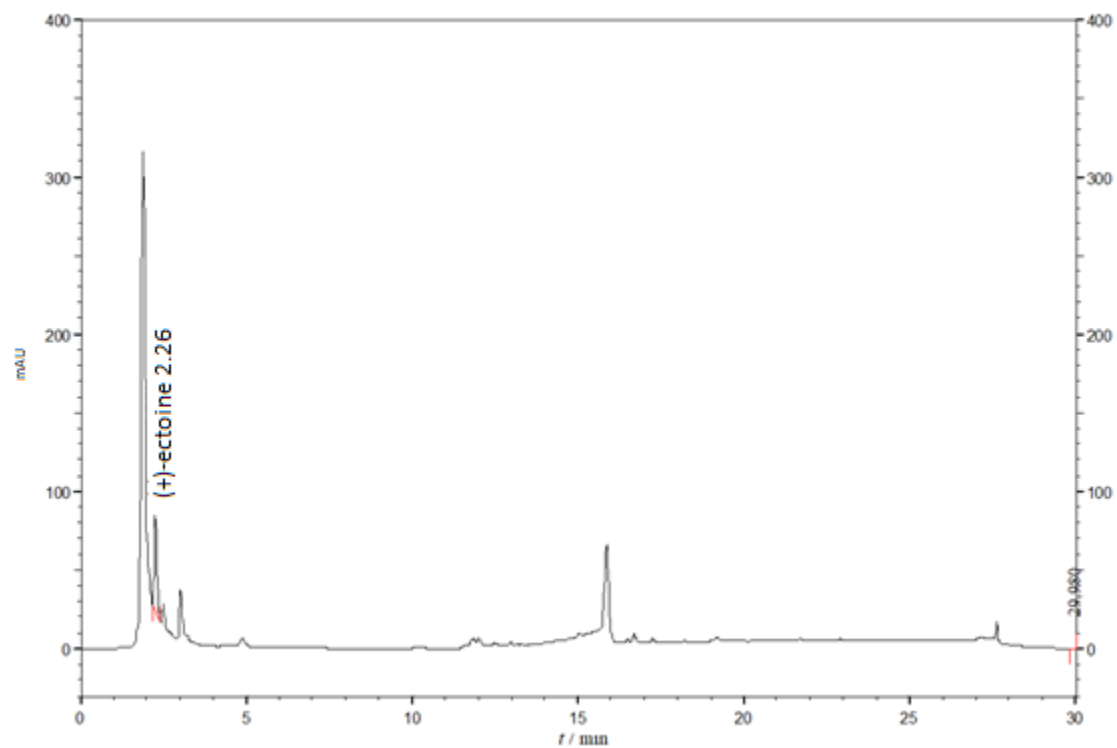
**Figure S4.2.8.** RP-HPLC chromatogram of sample **6** at 220 nm.



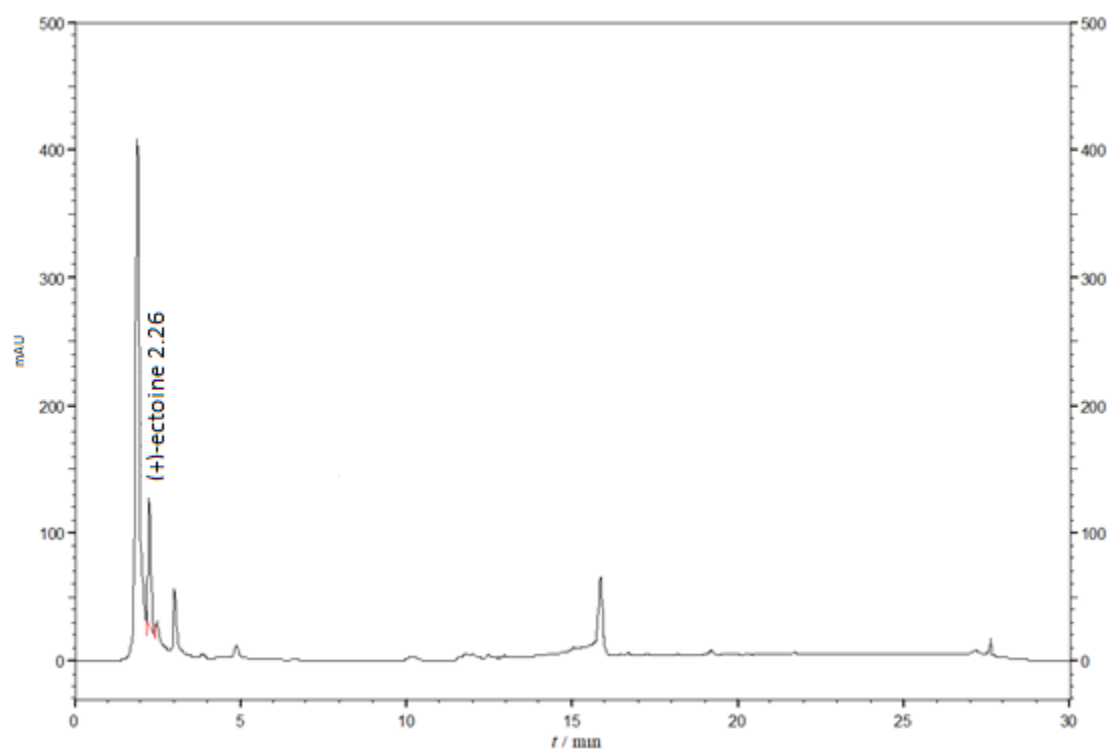
**Figure S4.2.9.** RP-HPLC chromatogram of sample 7 at 220 nm.



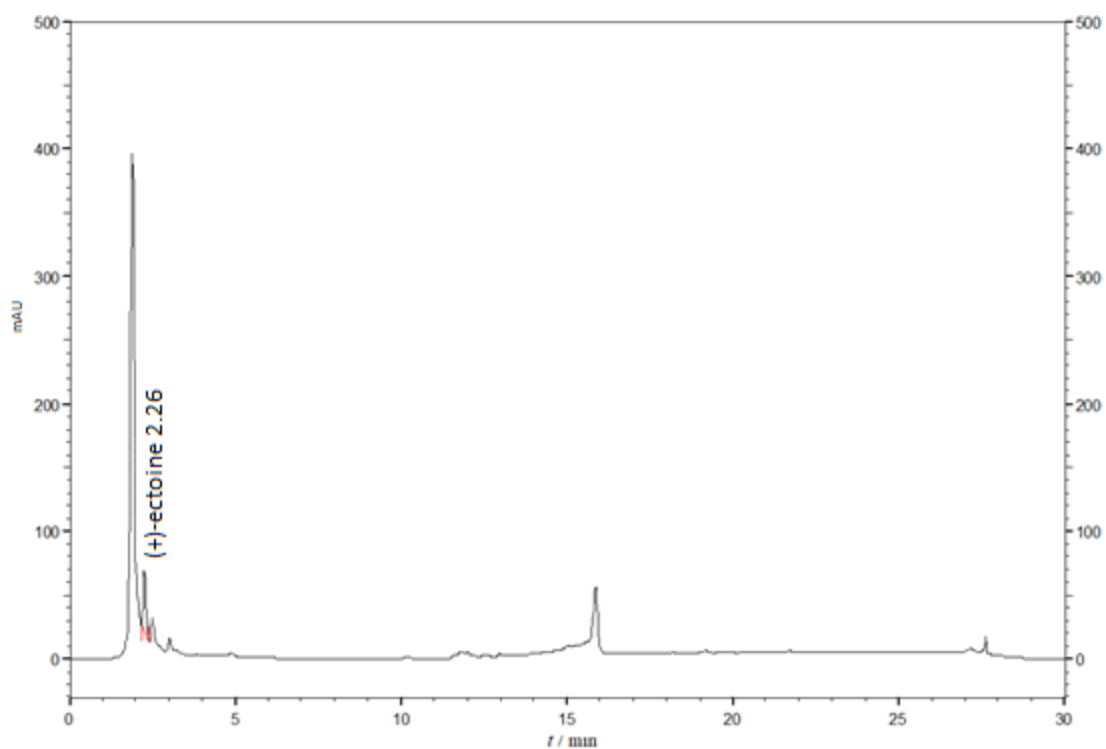
**Figure S4.2.10.** RP-HPLC chromatogram of sample 8 at 220 nm.



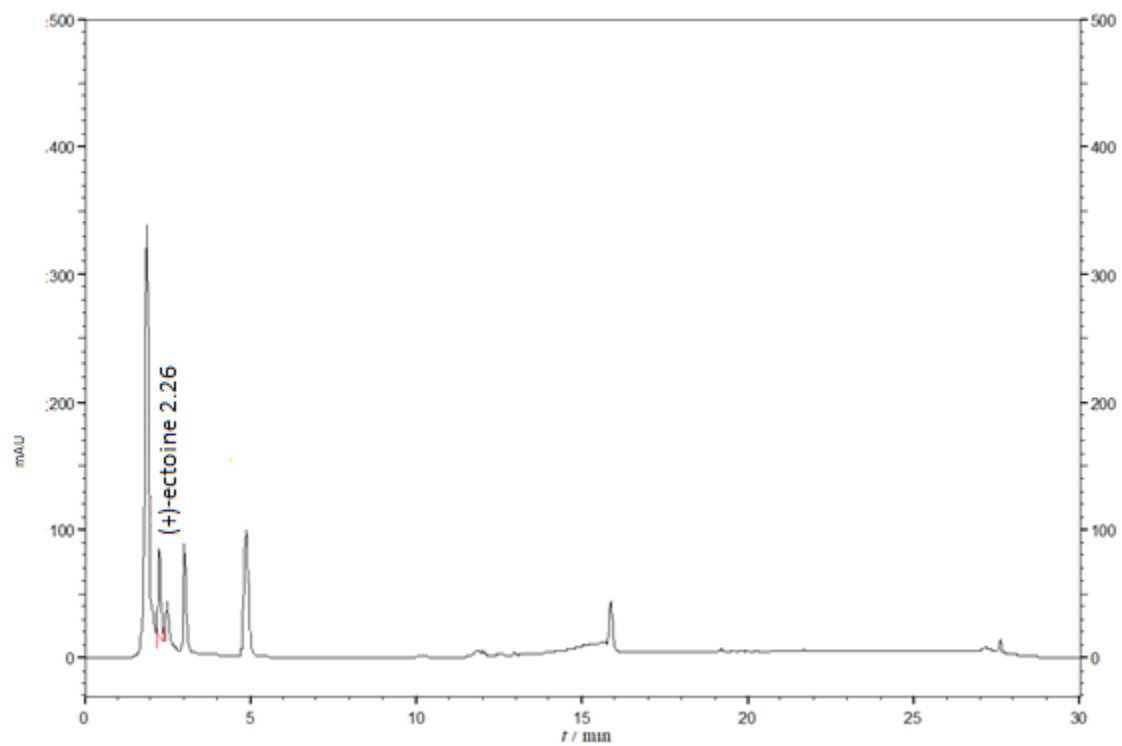
**Figure S4.2.11.** RP-HPLC chromatogram of sample **9** at 220 nm.



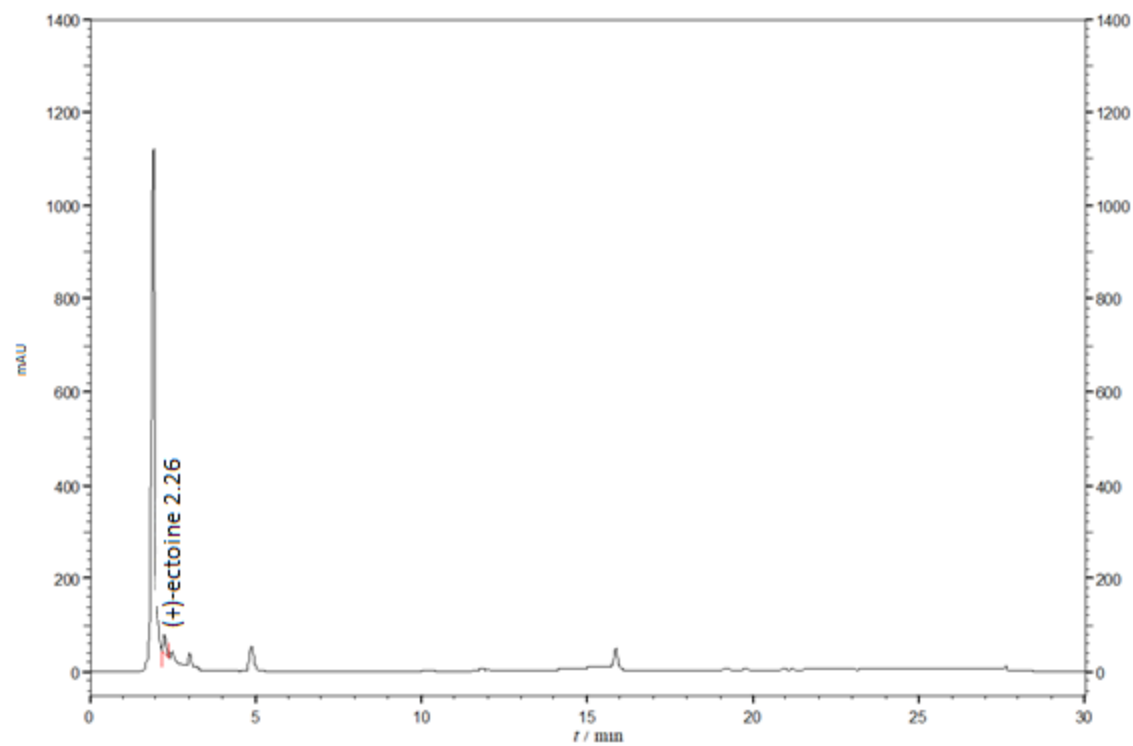
**Figure S4.2.12.** RP-HPLC chromatogram of sample **10** at 220 nm.



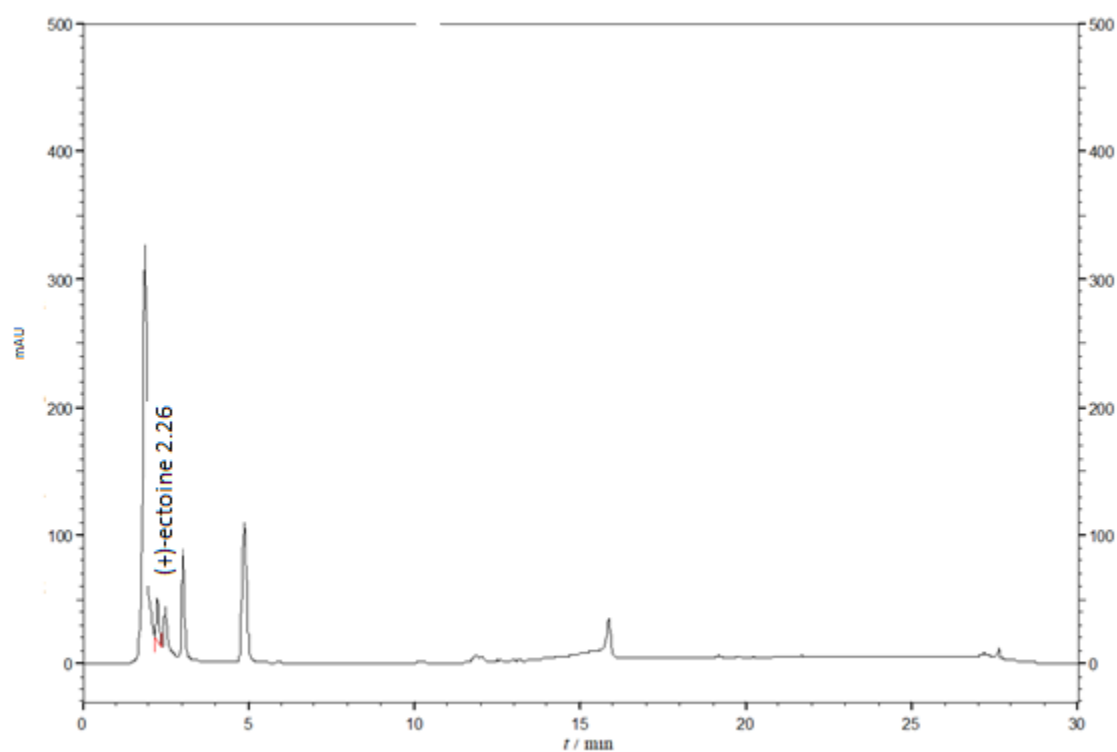
**Figure S4.2.13.** RP-HPLC chromatogram of sample **11** at 220 nm.



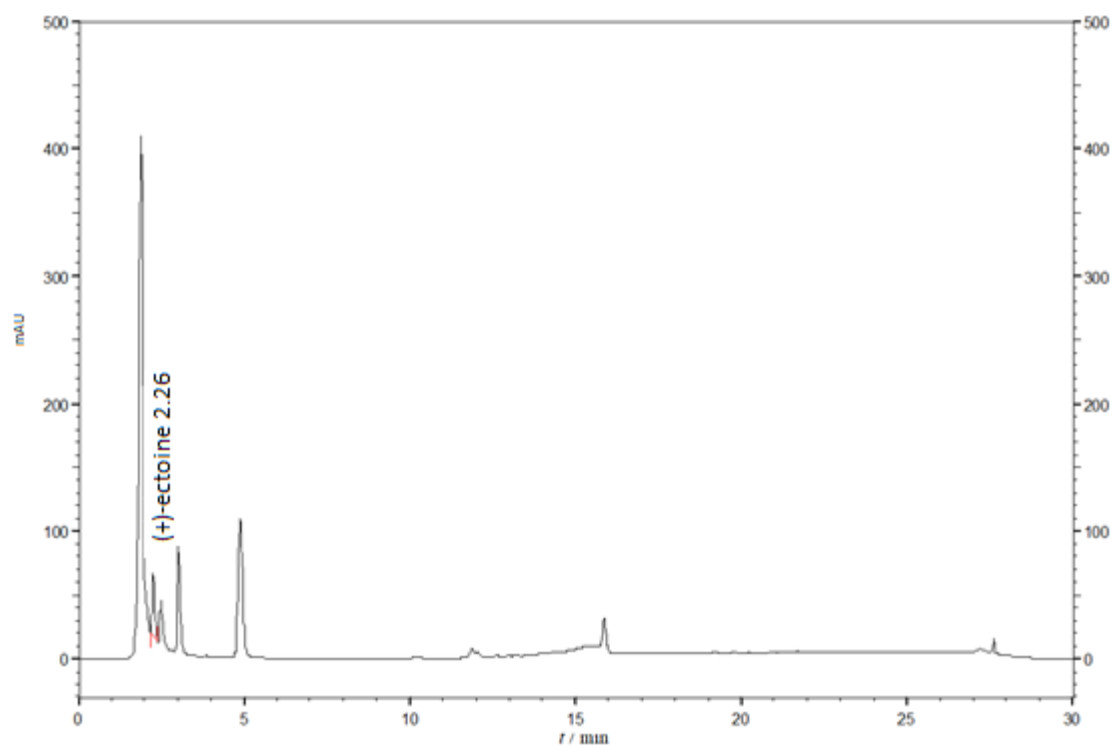
**Figure S4.2.14.** RP-HPLC chromatogram of sample **12** at 220 nm.



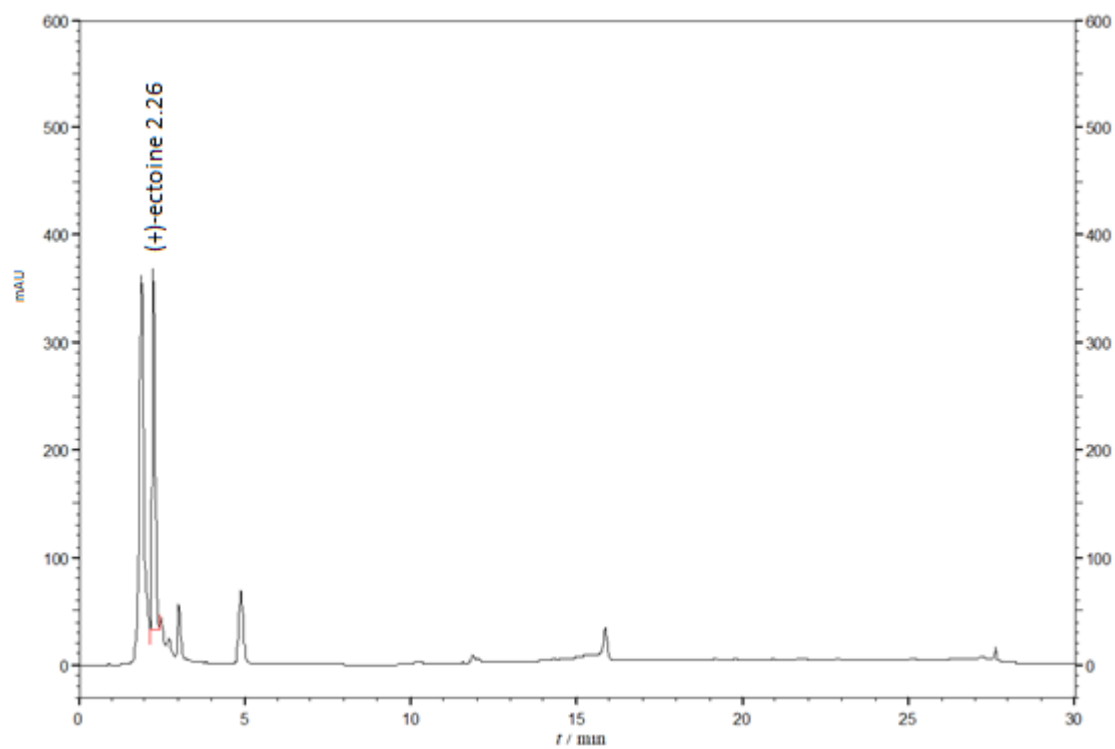
**Figure S4.2.15.** RP-HPLC chromatogram of sample **13** at 220 nm.



**Figure S4.2.16.** RP-HPLC chromatogram of sample **14** at 220 nm.



**Figure S4.2.17.** RP-HPLC chromatogram of sample **15** at 220 nm.



**Figure S4.2.18.** RP-HPLC chromatogram of sample **16** at 220 nm.



## 5. MS spectra of compound 7 and standard (+)-ectoine

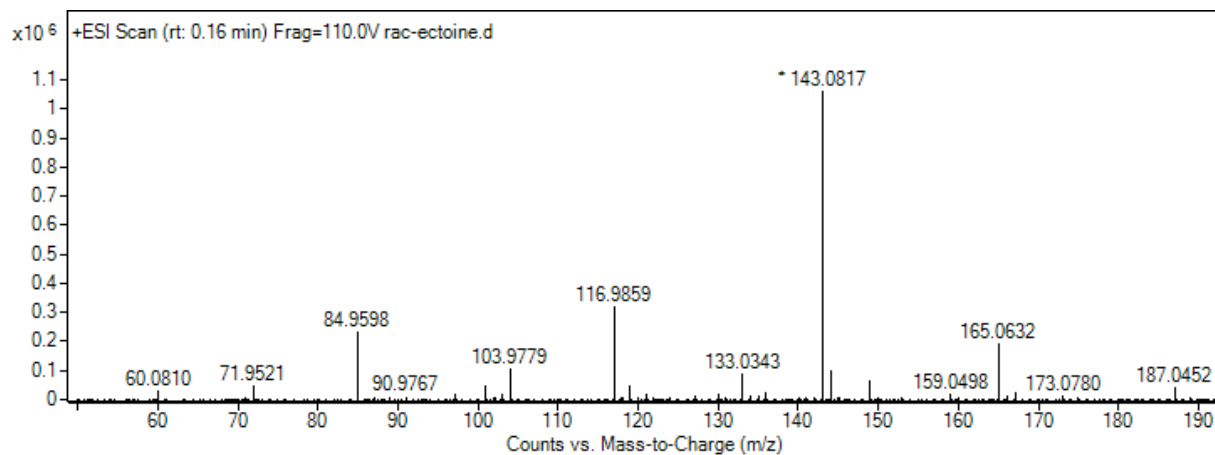


Figure S5.1. HRMS spectrum of compound 7 [(±)-ectoine].

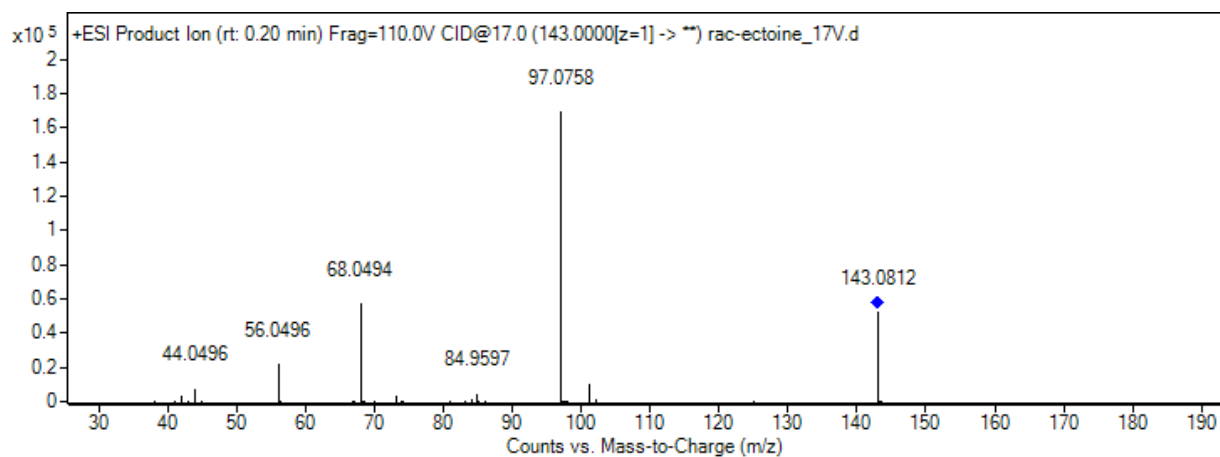


Figure S5.2. MS/MS Fragmentation of product ion  $m/z$  143 (compound 7).

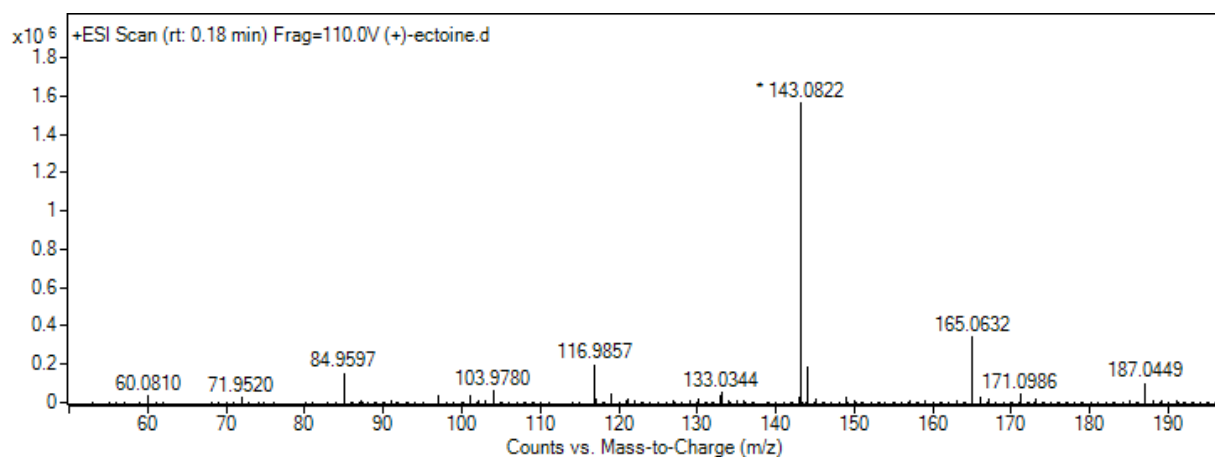
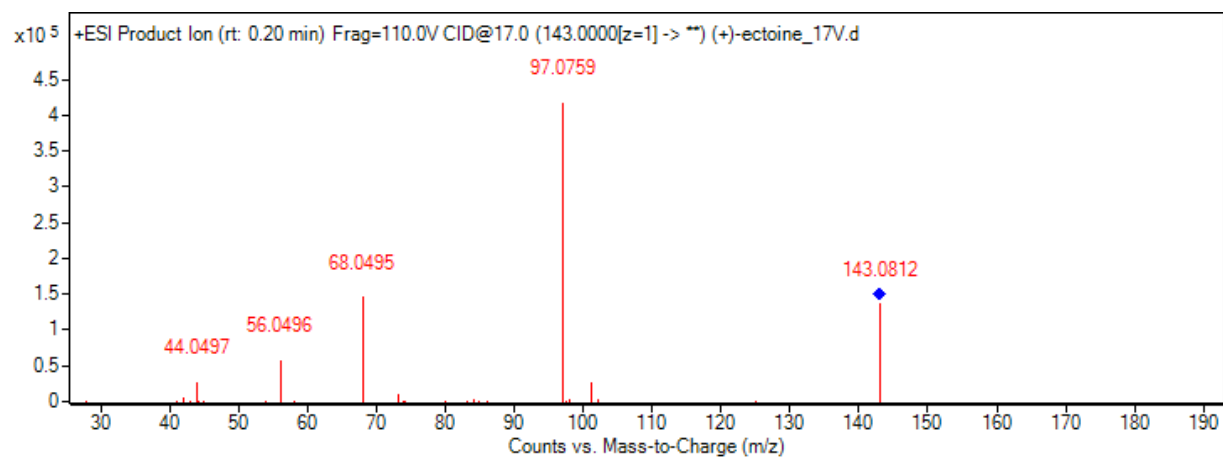


Figure S5.3. HRMS spectrum of standard (+)-ectoine.



**Figure S5.4.** MS/MS Fragmentation of product ion  $m/z$  143 (standard, (+)-ectoine).