



Article

# Modeling Power Generation and Energy Efficiencies in Air-Cathode Microbial Fuel Cells Based on Freter Equations

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**Featured Application:** The established mathematical model for air-cathode microbial fuel cells can be used to predict the electrical current and power generation as well as energy efficiencies at varying operating conditions.

**Abstract:** The model proposed in this study was based on the assumption that the biomass attached to the anode served as biocatalysts for microbial fuel cell (MFC) exoelectrogenesis, and this catalytic effect was quantified by the exchange current density of anode. By modifying the Freter model and combining it with the Butler–Volmer equation, this model could adequately describe the processes of electricity generation, substrate utilization, and the suspended and attached biomass concentrations, at both batch and continuous operating modes. MFC performance is affected by the operating variables such as initial substrate concentration, external resistor, influent substrate concentration, and dilution rate, and these variables were revealed to have complex interactions by data simulation. The external power generation and energy efficiency were considered as indices for MFC performance. The simulated results explained that an intermediate initial substrate concentration (about 100 mg/L under this reactor configuration) needed to be chosen to achieve maximum overall energy efficiency from substrate in the batch mode. An external resistor with the value approximately that of the internal resistance, boosted the power generation, and a resistor with several times of that of the internal resistance achieved better overall energy efficiency. At continuous mode, dilution rate significantly impacted the steady-state substrate concentration level (thus substrate removal efficiency and rate), and attached biomass could be fully developed when the influent substrate concentration was equal to or higher than 100 mg/L at any dilution rate of the tested range. Overall, this relatively simple model provided a convenient way for evaluating and optimizing the performance of MFC reactors by regulating operating parameters.

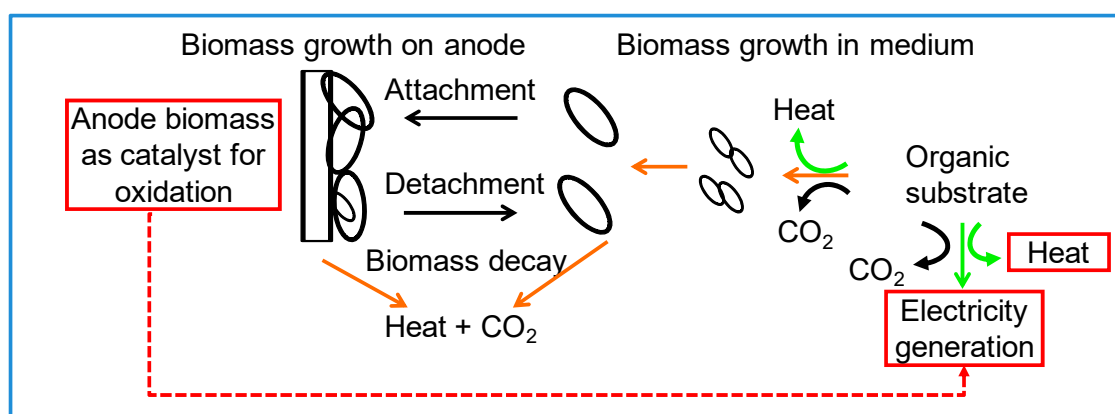
**Keywords:** exchange current; energy efficiency; heat generation; attached biomass; suspended biomass; single chamber air-cathode

## 1. Introduction

A microbial fuel cell (MFC) is a device oxidizing organic substrates in the liquid environment and releasing electrical energy to external loads. Anode attached biomass works as catalysts for substrate oxidization [1]. The catalytic microorganisms at anodes, mostly enriched from mixed microbial

consortia, are capable of handling various organic substrates [2,3], and can potentially be used in degrading organic substrates in various types of wastewater.

Due to complicated interactions between various design and operating variables in this hybrid type of reactor, it is not easy to experimentally achieve optimal conditions for power and energy generation and efficient substrate removal. Mathematical models thus provide a simple approach to investigate the effects of different variables and optimize the MFC performances. The electrochemical reactions at anode are emphasized in MFC modeling [4,5] since the anode reactions are features of MFC, and MFC performance can partly be predicted from the growth status of the anode attached bacteria [6]. The current MFC anode models are normally based on redox mediators that supposedly exist in the medium. Research progress already reveals that the outer membrane-bound cytochromes and the nanowire of bacteria conduct electricity [1], so bacteria as a whole, which are directly attached to the anode surface, can be regarded as catalysts. It can thus be assumed that only the directly anode-attached bacteria, or bacteria of a monolayer [7], contribute to electricity generation. Besides, the suspended bacteria must also be included in the model because the attached and suspended bacterial populations reach a dynamic equilibrium between each other, and because both populations utilize substrate for growth. This inclusion is especially essential when studying substrate utilization between the populations. The Freter model, which originally describes the dynamics of the suspended bacteria and wall-attached bacteria in a bioreactor [8,9], is herein adopted. The main reason for favoring this model is that it emphasizes the formation of a bacterial monolayer on a wall structure which is similar to the process of anode-bacteria attachment. The Freter model is first revised to account for the feature of MFC where electricity generation consumes substrate (Figure 1), before using it to describe the MFC bacterial populations.



**Figure 1.** A schematic of organic substrate and energy flow in a microbial fuel cell.

In this study, a dynamic model for MFC was developed based on microbial reaction, electrochemical reaction, and mass balances. Numerical simulation of the model was conducted to investigate the effect of varying operating parameters, including initial or influent substrate concentration, dilution rate, and external resistor, on the MFC microbial populations, substrate removal, power generation, and energy efficiency. Factors influencing the energy efficiency were especially emphasized in numerical simulation for both batch and continuous modes of operation.

## 2. Materials and Methods

### 2.1. MFC Design and Operation

Two identical single-chamber air-cathode MFC reactors, with a total volume of 155 mL, were fabricated from clear/extruded acrylic tubes (5 cm in internal diameter and 7.5 cm in length), covered with 0.25 cm thick plexiglass acrylic endplates at both ends. The anode was made from plain carbon cloth, while the air-cathode was made from  $0.5 \text{ mg cm}^{-2}$  Pt-containing (10% in carbon black) carbon

cloth treated with Nafion, and with polytetrafluoroethylene (PTFE) as a gas diffusion backing layer [10]. A 2.2 k $\Omega$  resistor was connected in the circuit as an external load.

Anaerobic sludge was obtained for MFC inoculation from a swine manure lagoon at the University of Minnesota Southern Research and Outreach Center in Waseca, MN. The sludge was sieved through a 1 mm mesh sieve to remove sand, gravels, and plant residues prior to use. Sodium acetate was dissolved in 50 mM phosphate buffer solution (pH 7) to prepare a medium with 40 mM acetate. Other nutrients also added to enhance and maintain the development of bacteria included (g L<sup>-1</sup>): 0.31 NH<sub>4</sub>Cl, 0.13 KCl, and mineral and vitamin solutions.

### Data Acquisition

The voltage ( $U_{ext}$ ) across the external resistor was measured by voltage probes and recorded by a CR 1000 data logger (Campbell Scientific, Inc., Logan, UT, USA). The anodic and cathodic potentials were monitored using standard Ag/AgCl electrodes (MF-2072, BASi, Inc., West Lafayette, IN, USA). The data logger supporting software, LoggerNet Version 3.4 (Campbell Scientific, Inc., Logan, UT, USA), was used to monitor and collect the voltage data of the MFC reactors.

### 2.2. Model Description

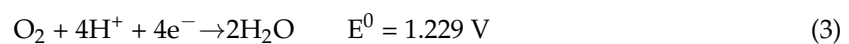
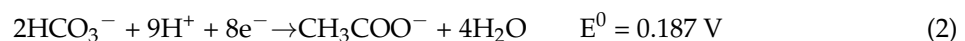
The mathematical model of microbial fuel cells to be developed simulated and predicted the electrochemical performance in batch or continuous mode of operation. A few assumptions were made to simplify the modeling of different MFC reactors. First, the anode-attached microbes were assumed to be the catalyst for the exoelectrogenic process; second, the exchange current density, or the catalytic effect, was proportional to the concentration of the attached biomass; third, the mass transfer of acetate to anode and oxygen gas to cathode, assumed to be fast enough, compared to the electrochemical processes, was not a limiting factor and was thus neglected; and fourth, the hydrolytic products of dead cells were assumed to not provide a carbon source or energy to the suspended or attached bacterial growth.

#### 2.2.1. Electrode Potentials

The electrode potential (against the standard hydrogen electrode, SHE or the normal hydrogen electrode, NHE) was dependent on the standard electrode potential and the concentration of active species according to the Nernst equation:

$$E^{o'} = E^0 + \frac{RT}{nF} \sum v_i \ln c_i \quad (1)$$

where  $E^{o'}$  was the formal potential (pH- and concentration-adjusted reduction potential),  $E^0$  was the standard electrode potential,  $v_i$  was the stoichiometric number for species  $i$  (positive for oxidized species, and negative for reduced species), and  $c_i$  was the concentration of the species  $i$ . In the pH region of this study, the Mineql+ [11] simulation indicated that bicarbonate was the major form in the carbonate equilibrium system. Thus, the anodic and cathodic reactions proceeded as follows:



The anode and cathode electrode potentials ( $E_A^{o'}$  and  $E_C^{o'}$ , respectively) based on the assumed acetate and bicarbonate concentrations could be estimated as follows:

$$E_A^{o'} = E_A^0 + \frac{RT}{nF} \sum v_i \ln c_i = -0.335 [\text{Ac}^-] = 0.04 \text{ M}; [\text{HCO}_3^-] = 10^{-4.5} \text{ M} \quad (4)$$

$$E_C^{o'} = E_C^0 + \frac{RT}{nF} \sum v_j \ln c_j = 0.805 \quad P_{\text{O}_2} = 0.2 \quad (5)$$

The experimentally measured cathodic potentials usually were much lower than the predicted values [12,13], partly due to other reactions that might have happened on the cathode surface which resulted in a mixed cathodic potential of lower values. For example, hydrogen peroxide production could happen on cathode to deviate the cathode potential to a lower value (e.g., 0.36 V) [12]. In accordance with both the literature information and the cathode potential at open-circuit operation in this study, a cathode potential of 0.51 V (NHE), rather than 0.805 V (NHE), was adopted herein. The electromotive force ( $E_{emf}$ ) was defined as the difference between the cathode and anode potentials:

$$E_{emf} = E_C^{0'} - E_A^{0'} \quad (6)$$

### 2.2.2. Exchange Current and Overpotential

Overpotential is defined as the difference of an electrode potential from its formal potential in a fuel cell. The Butler–Volmer equation, neglecting the reverse cathodic reaction [14], was used to express the relationship between the exchange current and anode overpotential:

$$I = j_{oA} A e^{\frac{\alpha_A F \eta_{act,A}}{RT}} \quad (7)$$

where  $I$  was the electric current,  $j_{oA}$  was the exchange current density of anode,  $\alpha_A$  was the transfer coefficient of the anodic reaction, and  $\eta_{act,A}$  was the anode overpotential. In a similar way, the cathode overpotential could be formulated with the reverse reaction neglected as follows:

$$I = j_{oC} A e^{\frac{(\alpha_C - 1) F \eta_{act,C}}{RT}} \quad (8)$$

where  $j_{oC}$  was the exchange current density of cathode,  $\alpha_C$  was the charge transfer coefficient of the cathode reaction, and  $\eta_{act,C}$  was the cathode overpotential. The exchange current density of cathode was consistent during the MFC operation, due to the unchanging amount of the catalyst (0.5 mg cm<sup>-2</sup> of Pt), but that of anode was proportional to the increasing catalyst load, which was the attached biomass concentration here [15]:

$$j_{oA} = j_{oAU} w \quad (9)$$

where  $j_{oAU}$  was the unit exchange current density of exoelectrogenic biomass, and  $w$  was the attached biomass concentration on anode.

### 2.2.3. Mass Balances

A model that was proposed to describe the suspended and attached microbial formation [8, 9] in an aqueous environment was modified and adopted in this study. In this model, bacteria could divide, suspend and decay in the reactor medium, and could also attach to a surface with a dynamic equilibrium between attachment and detachment. Since bacterial growth and electricity generation consumed substrate when it was available, the three components of the system, the substrate concentration ( $S$ ), the suspended biomass concentration ( $u$ ), and the attached biomass concentration ( $w$ ), which were dictated by mass balances, were expressed in three ordinary differential equations as follows:

$$\frac{dS}{dt} = D(S_{in} - S) - \gamma^{-1}(u\mu_u + \delta w\mu_w) - r_e \quad (10)$$

$$\frac{du}{dt} = u(\mu_u - D - k_u) + \beta\delta w + \delta w\mu_w(1 - G) - \alpha u(1 - W) \quad (11)$$

$$\frac{dw}{dt} = w(\mu_w G - \beta - k_w) + \delta^{-1}\alpha u(1 - W) \quad (12)$$

where  $D$  was the dilution rate,  $S_{in}$  was the influent substrate concentration,  $\gamma$  was the biomass yield of substrate,  $\mu_u$  was the suspended bacterial growth rate,  $\mu_w$  was the attached bacterial growth rate,  $\delta$  was the ratio of projected anode surface area to reactor volume,  $r_e$  was the substrate utilization rate

by electricity and heat generation,  $k_u$  was the suspended bacterial death rate,  $k_w$  was the attached bacterial death rate,  $\beta$  was the detachment rate of bacteria from wall,  $\alpha$  was the attachment rate of bacteria to anode surface,  $G$  was the fraction of daughter cells attached to anode, and  $W$  was the wall occupation fraction. Some of those parameters in the equations could be defined through further fundamental design and operating parameters:

$$D = Q/V \tag{13}$$

$$\delta = A/V \tag{14}$$

$$\mu_u = \frac{m_u S}{a_u + S} \tag{15}$$

$$\mu_w = \frac{m_w S}{a_w + S} \tag{16}$$

$$G = \frac{(1 - W)}{(1.1 - W)} \tag{17}$$

where  $Q$  was the dilution rate,  $V$  was the MFC reactor volume,  $A$  was the projected anode surface area,  $m_u$  was the maximum specific growth rate of suspended bacteria,  $a_u$  was the half saturation coefficient of suspended bacteria of the Monod kinetics,  $m_w$  was the maximum specific growth rate of attached bacteria,  $a_w$  was the half saturation coefficient of attached bacteria.

#### 2.2.4. Ohm’s Law and Kirchhoff’s Voltage Law

By applying Ohm’s law, the electrical current of the circuit, the voltage drop ( $U_m$ ) across the resistance ( $R_m$ ) of electrolyte solution of MFC medium, and the voltage drop ( $U_c$ ) across the contact resistance ( $R_c$ ) of MFC could be obtained:

$$I = U_{ext}/R_{ext} \tag{18}$$

$$U_m = \frac{Id}{A\sigma} \tag{19}$$

$$U_c = IR_c \tag{20}$$

where  $U_{ext}$  was the external voltage across the external resistor ( $R_{ext}$ ),  $d$  was the distance between two electrodes, and  $\sigma$  was the conductivity of MFC medium. By combining all the polarization losses along the circuit of MFC, the voltage conservation according to Kirchhoff’s voltage law could be denoted as follows:

$$E_{emf} = E_C^{0'} - E_A^{0'} = U_{ext} + U_c + U_m + \eta_{act, A} - \eta_{act, C} \tag{21}$$

#### 2.2.5. Power and Energy Efficiencies

The power ( $P_{ext}$ ) extracted by the external load of a resistor was the product of the external voltage and current:

$$P_{ext} = U_{ext}I \tag{22}$$

The power extracted by the electrical circuit of MFC ( $P_{cell}$ ), or termed as an MFC cell, was the product of the electromotive force and current:

$$P_{cell} = E_{emf}I \tag{23}$$

The power lost through the polarization of the activation overpotentials of anode and cathode, the contact resistance, and the medium resistance, was the power emitted in a form of heat:

$$P_{heat} = (U_c + U_m + \eta_{act, A} - \eta_{act, C})I \tag{24}$$

The rate of the substrate (acetate) loss through MFC cell ( $r_e$ ) could be calculated as follows:

$$r_e = MP_{\text{cell}} / (V\Delta H_c) \quad (25)$$

where  $\Delta H_c$  was the heat of combustion of acetic acid, and  $M$  was the relative molecular weight of acetic acid.

Besides the energy loss by heat generation in MFC, the chemical energy of substrate was also lost by providing energy for the growth of the suspended and attached bacteria. The corresponding powers for the suspended ( $P_u$ ) and attached ( $P_w$ ) bacteria could be calculated as follows:

$$P_u = \gamma^{-1} \Delta H_c \mu_u V / M \quad (26)$$

$$P_w = \gamma^{-1} \Delta H_c \delta_w \mu_w V / M \quad (27)$$

Although some studies [13] suggested that the energy efficiency was further reduced by the more positive potential (close to 0 V, NHE) of redox intermediates than that of the substrate, this energy loss was not considered because our experiment achieved anode potentials close to that of the substrate. Therefore, the MFC cell energy efficiency ( $\eta_{\text{cell}}$ ) and overall energy efficiency ( $\eta_{\text{overall}}$ ) were defined as follows:

$$\eta_{\text{cell}} = P_{\text{ext}} / P_{\text{cell}} \quad (28)$$

$$\eta_{\text{overall}} = P_{\text{ext}} / (P_{\text{cell}} + P_u + P_w) \quad (29)$$

Please note that in batch mode, the overall energy efficiency was calculated differently:

$$\eta_{\text{overall}} = \int_0^t P_{\text{ext}} dt / (\Delta H_c \Delta SV / M) \quad (30)$$

## 2.2.6. Model Parameters

Some constants and parameters involved in microbial fuel cell processes described in Equations (from (1) to (30)) were listed in Table 1. During modeling and simulation, units of those parameters and variables were not necessarily SI units, but some were adjusted to the suitable magnitudes. Microbial parameters of acetate utilizers in anaerobic conditions were adopted from the literature [16]. Some model parameters associated with electrochemical processes, including charge transfer coefficients ( $\alpha_A$  and  $\alpha_C$ ) and exchange current densities ( $j_{oA}$ ,  $j_{oAU}$  and  $j_{oC}$ ) were estimated from the polarization dataset obtained from one of two MFC reactors in this experiment. The quasi-Newton approach was used to minimize the  $\chi^2$  test statistic as an error function:

$$\chi^2 = \sum_{i=1}^n \frac{(y_i^{\text{cal}} - y_i^{\text{exp}})^2}{y_i^{\text{exp}}} \quad (31)$$

where  $n$  was the number of data points, and  $y_i^{\text{exp}}$  was the  $i$ -th experimental value, and  $y_i^{\text{cal}}$  was the  $i$ -th calculated value. The polarization curve data of the other reactor was used for validation.

**Table 1.** Constants, and design and operating parameters of a microbial fuel cell system. NHE: the normal hydrogen electrode.

Parameter Symbol	Description	Value	Units
Physical			
F	Faraday's constant	96,485	C/mol
R	Ideal gas constant	8.314	J/mol/K
T	Room temperature	298	K
$\Delta H_c$	Heat of combustion of acetic acid	-875,000	J/mol
M	Relative molecular weight of acetic acid	60	g/mol
Electrochemical			
$E_A^{0'}$	Formal reduction potential of anode	-0.335	V vs. NHE
$E_C^{0'}$	Formal reduction potential of cathode	0.51	V vs. NHE
Reactor configuration			
A	Projected surface area of electrode	0.002	m <sup>2</sup>
d	Distance between anode and cathode	0.075	m
V	Volume of MFC reactor	$1.55 \times 10^{-4}$	m <sup>3</sup>
$\delta$	Ratio of anode surface area to MFC volume	13	m <sup>2</sup> /m <sup>3</sup>
$R_c$	Contact resistance of MFC	20	$\Omega$
Operating			
D	Dilution rate	0–2	d <sup>-1</sup>
$S_{in}$	Substrate concentration	10–2400	g/m <sup>3</sup>
Q	Flow rate	$0-7.8 \times 10^{-5}$	m <sup>3</sup> /d
$\sigma$	Conductivity of MFC medium	1.1	S/m
$R_{ext}$	External resistor	2200	$\Omega$
[HCO <sub>3</sub> <sup>-</sup> ]	Bicarbonate concentration	$1 \times 10^{-4.5}$	kmol/m <sup>3</sup>
P <sub>O2</sub>	Partial oxygen pressure at cathode	0.2	atm
pH	-log <sub>10</sub> of proton concentration	7	Dimensionless
Microbial			
$\gamma$	Biomass yield from substrate	0.05	g/g
$k_u$	Death rate of suspended bacteria	0.02	d <sup>-1</sup>
$k_w$	Death rate of attached bacteria	0.02	d <sup>-1</sup>
$\alpha$	Bacterial attachment rate	0.05	d <sup>-1</sup>
$\beta$	Bacterial detachment rate	0.05	d <sup>-1</sup>
$m_u$	Maximum specific growth rate of suspended bacteria	2.4	d <sup>-1</sup>
$m_u$	Maximum specific growth rate of attached bacteria	2.4	d <sup>-1</sup>
$a_u$	Half saturation coefficient of suspended bacteria	100	g/m <sup>3</sup>
$a_w$	Half saturation coefficient of attached bacteria	100	g/m <sup>3</sup>
$w_{max}$	Maximum bacterial attachment	0.33 <sup>a</sup>	g/m <sup>2</sup>

<sup>a</sup> The maximum amount of bacteria attached on electrode was calculated from the bacteria dimensions observed in SEM images, and this value was close to the literature values of 0.47 to 0.52 g/m<sup>2</sup> [7].

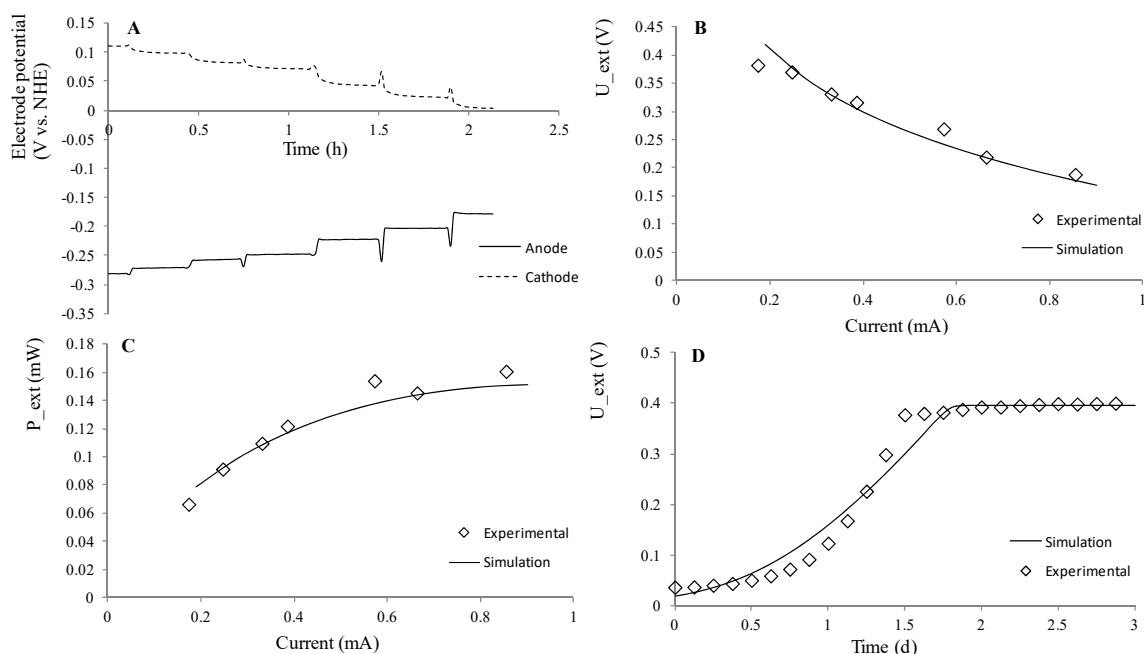
### 3. Results and Discussion

#### 3.1. Parameter Estimation

Two identically-made MFC reactors with respect to reactor configuration and electrode materials were inoculated with the same anaerobic sludge and operated in parallel. After MFC were fully inoculated and operated for 30 d, polarization curves of MFC was obtained by varying the external resistor from 2200 to 220  $\Omega$ , with the electrode potentials recorded (Figure 2A). The resultant electrochemical parameters estimated by fitting polarization curves of electrodes were listed in Table 2. The simulation results based on the estimated parameters were in good agreement with the experimental data of both reactors, as shown in Figure 2B,C. The anodic exchange current density of 62.5 mA m<sup>-2</sup> was in the same order of a literature value of 29 mA m<sup>-2</sup> [17]. The cathodic exchange current density (0.975 mA m<sup>-2</sup>) observed in this study was higher than those of non-catalyzed and bio-catalyzed cathodes [18,19] but lower than another Pt-based cathode [20]. To further validate the model, the external voltage ( $U_{ext}$ ) during inoculation stage was simulated based on the listed parameters except the unit exchange current, which was fitted from the experimental data at the inoculation stage (0.129 mA mg<sup>-2</sup> biomass) and had a difference by 32% from 0.189 mA mg<sup>-1</sup> which



was fitted from polarization curves. This difference could be a result of the change of the activity or catalytic effect of the attached biomass during the inoculation and 30 d operation. The simulated data and the experimental data of the inoculation stage generally showed good agreement with each other (Figure 2D).



**Figure 2.** Electrochemical parameters estimation and validation; (A) the electrode potential during polarization experiment; (B,C) model validation for experimental data from another identical microbial fuel cell (MFC) reactor (B for  $U_{ext}$ , and C for  $P_{ext}$ ); and (D) comparison of the simulated and experimental  $U_{ext}$  during inoculation stage.

**Table 2.** Estimated electrochemical parameters for MFC in this study.

Estimated Parameters	Description	Value	Units
$\alpha_A$	Charge transfer coefficient, anode	0.318	Dimensionless
$\alpha_C$	Charge transfer coefficient, cathode	0.694	Dimensionless
$j_{oA}$	Exchange current density, anode	62.5	$\text{mA}/\text{m}^2$
$j_{oAU}$	Unit exchange current, anode	0.189	$\text{mA}/\text{mg}$
$j_{oC}$	Exchange current density, cathode	0.975	$\text{mA}/\text{m}^2$

### 3.2. Simulation of Batch Mode MFC

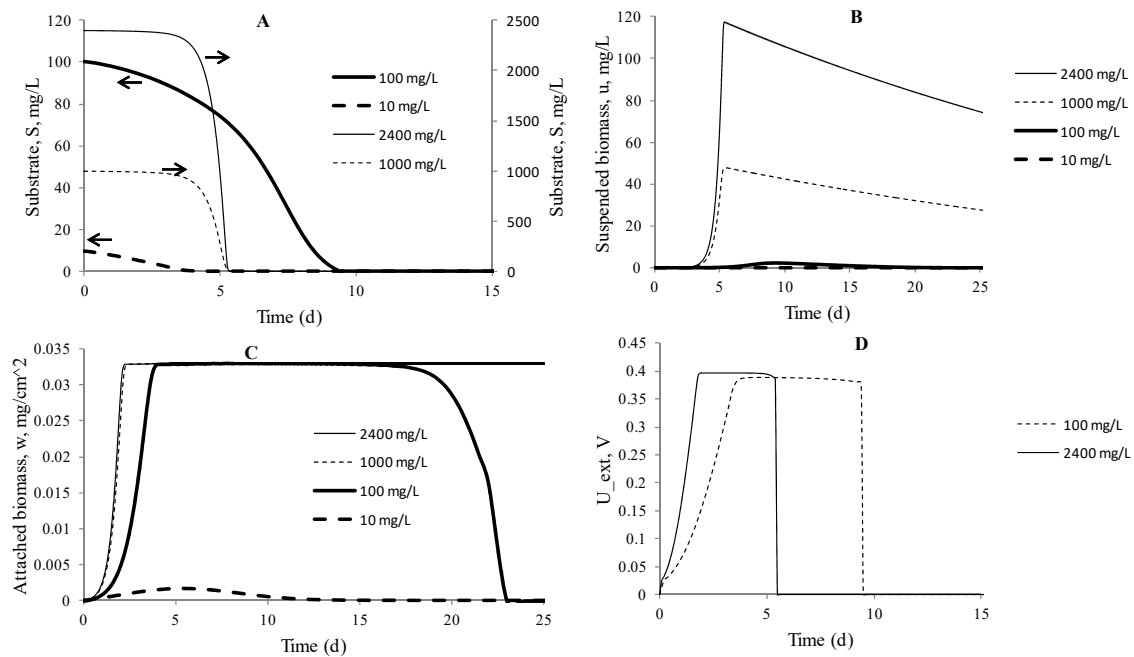
#### 3.2.1. Effect of Initial Substrate Concentration

Simulation of the proposed model was performed through Matlab on a personal computer to illustrate the processes of substrate utilization, biomass concentration evolution, and voltage generation across an external resistor. Since most MFC studies in literature were carried out in batch mode, this simulation assumed the medium dilution rate  $D = 0$  with initial substrate (HAc) concentrations of 10, 100, 1000, and 2400  $\text{mg L}^{-1}$ , consecutively.

The simulation results are shown in Figure 3. Initial substrate concentrations of 1000 and 2400  $\text{mg L}^{-1}$  did not show obvious decrease in the first 4 d (Figure 3A), but substrate was consumed dramatically after this period. MFCs started with the lower substrate concentrations behaved differently from the higher strength cases, as a continuous and gradual decrease was observed. The substrate in the case of 100  $\text{mg L}^{-1}$  lasted the longest, i.e., 9 d, which almost doubled that in the higher strength cases. These results indicated that it was not always that more concentrated substrate



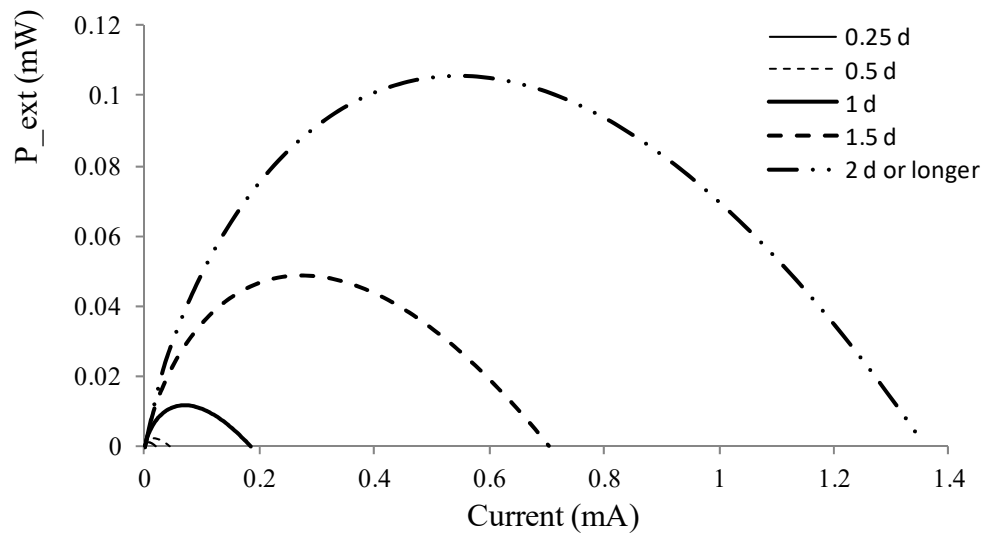
could sustain the longest reaction period, and a moderate substrate concentration should be chosen for keeping MFC from starvation. Trends of substrate utilization were generally the work of suspended bacteria, which could accumulate to a high level and subsequently, consumed substrate at a high rate that rapidly reduced substrate concentration (Figure 3B). For example, in the initial substrate concentrations of 100 and 2400 mg L<sup>-1</sup>, the peak suspended biomass concentrations were 2.5 and 117 mg L<sup>-1</sup>, respectively. The latter was 47 times higher than the former, which consumed substrate at a higher rate due to two reasons: the higher level of biomass concentration and the enhanced specific substrate utilization rate indicated by the Monod equation (Equations (15) and (16)).



**Figure 3.** Simulated time-course profiles of process variables at different initial substrate concentration. (A) the substrate concentration; (B) suspended biomass concentration; (C) attached biomass concentration, and (D) the external voltage  $U_{ext}$ .

The attached biomass concentrations (Figure 3C) accumulated to their plateau values (0.033 mg cm<sup>-2</sup>) for all initial substrate concentrations (in 2 d for 1000 and 2400 mg L<sup>-1</sup>, and in 4 d for 100 mg L<sup>-1</sup>), except for the one with 10 mg L<sup>-1</sup> substrate, indicating that there might exist a lower limit for substrate concentration. Thus, a substrate concentration lower than 10 mg L<sup>-1</sup> should be avoided during inoculation stage. The higher the initial substrate concentration, the longer the attached biomass would sustain: both lasted more than 25 d for 1000 and 2400 mg L<sup>-1</sup>, but only 23 d for 100 mg L<sup>-1</sup> before the biomass disappeared. The electricity generation (Figure 3D) was simulated in a form of external voltage ( $U_{ext}$ ) for the initial substrate concentrations of 100 and 2400 mg L<sup>-1</sup>, because both of them fully developed the attached biomass, and were also the typical levels for the low and high substrate concentrations, respectively. During the first few days of anode inoculation, the growing voltage was caused by the growing attached biomass and eventually reached a plateau value of about 0.39 V. Since the substrate lasted longer in the 100 mg L<sup>-1</sup> MFC than in the 2400 mg L<sup>-1</sup> MFC, the electricity generation also lasted longer by 4 more days. Energy generated was 25.1 and 40 J for 2400 mg L<sup>-1</sup> MFC and for 100 mg L<sup>-1</sup> MFC, with overall energy efficiencies extracted from acetate ( $\eta_{overall}$ ) of 0.46% and 17.7%, respectively. Therefore, based on electricity energy generation and overall energy efficiency, the MFC with 100 mg L<sup>-1</sup> of substrate substantially outperformed the one with 2400 mg L<sup>-1</sup>. Controlling the initial substrate concentration was thus demonstrated as a way of regulating the suspended biomass concentration, and was helpful in harvesting more energy from the substrate for electricity.

The growing amount of the attached biomass not only increased the external voltage ( $U_{ext}$ ), but also improved the external power, evidenced by the gradual expansion of the polarization curves against the time in Figure 4. These simulations were based on the high initial substrate concentration of  $2400 \text{ mg L}^{-1}$  in order to avoid a noticeable substrate decrease in 2 d. The pattern of the curve expansion was similar to that which was observed in the literature [4,6], illustrating the importance of the attached biomass on power generation.



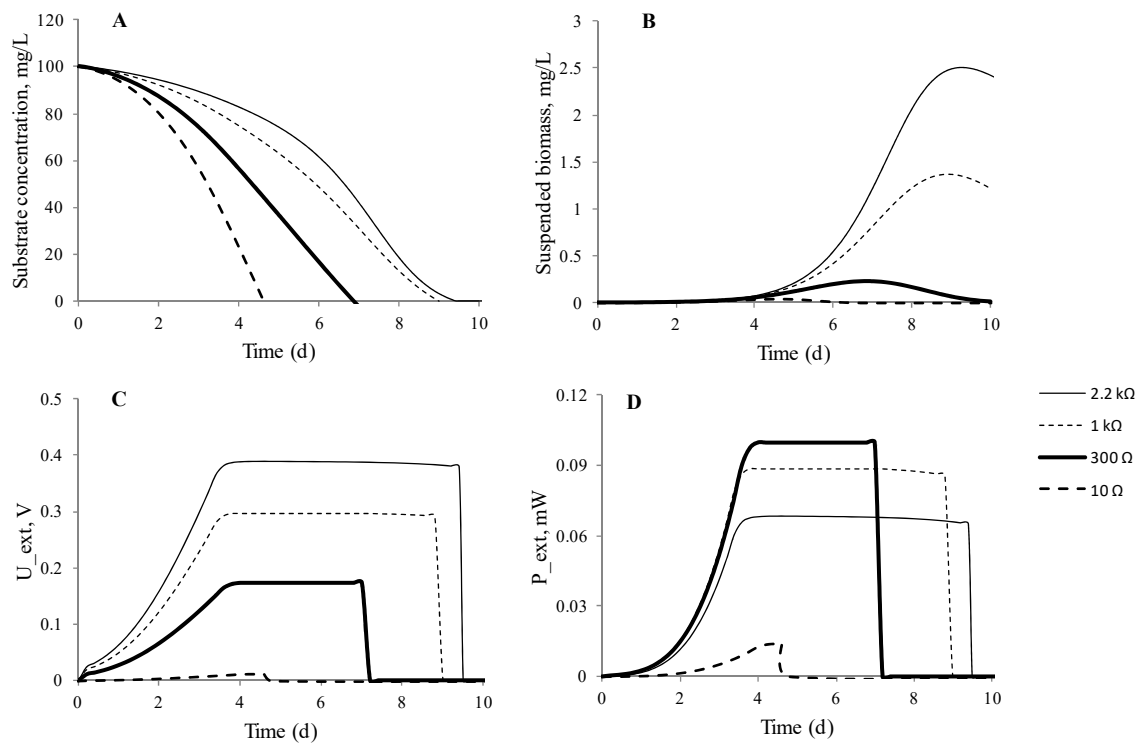
**Figure 4.** Simulated polarization curves at different moments of inoculation. Attached bacteria concentration reached a plateau value after about 2 d, and the polarization curves did not change much thereafter.

### 3.2.2. Effect of External Resistor

From the comparisons of external voltage and overall energy efficiency, it could be seen that under this experimental condition, a  $100 \text{ mg L}^{-1}$  of initial substrate worked best among the four tested substrate concentrations. The next step of this study simulated the impact of the external resistor on the batch mode MFC voltage and power generation as well as overall energy efficiency. In MFC study, the internal resistance ( $R_{in}$ ) is used for indicating the overall resistance of a reactor, which is a useful index because varying an external resistor to the value of internal resistance generates the maximum external power density [21]. Depending on the reactor design and operation, the values of  $R_{in}$  might differ but usually was within a range of several hundreds of Ohms, which was also in accordance with the observations ( $332 \Omega$  in the experimental study; and  $471 \Omega$  in the simulation study) in MFC reactors in this study. In order to see the effect of external resistors on MFC energy and power generation, a range of resistor covering the values of internal resistance was chosen for simulation, i.e., 10, 300, 1000, and  $2200 \Omega$ .

The simulation results were presented in Figure 5. The smaller the value of the external resistor was, the more rapidly the substrate was consumed (Figure 5A), which was also observed in two-chamber MFC reactors for organic matter removal [22,23]. The main reason was that the increased heat generation, which depicted the part of energy loss through overpotentials (Equation (24)), consumed the substrate at a higher rate as a result of the increased current with the smaller external resistor [24]. When the resistor reduced to  $10 \Omega$ , a major part of the substrate was lost via heat, so that there was not enough substrate to fully support the suspended (Figure 5B) and attached microbial growth. Therefore, the  $10\text{-}\Omega$  resistor caused an apparently impaired power generation performance due to the failure of the microbial growth. On the other hand, the small resistor was beneficial for organic substrate removal because of its higher substrate utilization rate, being more appropriate for treating high strength wastewater, such as swine wastewater [25]. A greater external resistor yielded

a higher external voltage (Figure 5C), and took a longer substrate consumption time. Simulation of external power generation (Figure 5D) confirmed that 300  $\Omega$ , which was the closest among the three to the internal resistance observed in this study, generated the largest power. A similar trend was also observed in a previous experimental study [23]. However, this value of resistor sustained a shorter reaction time by 2 to 3 d compared to that of 1000 and 2200  $\Omega$ . Overall, the 1000  $\Omega$  resistor harvested the most energy from substrate, achieving overall energy efficiency of 20.7% (Table 3). Energy efficiency between 15% and 20% achieved in experimental studies was considered the best cases [26,27], while a value of around 5% was more frequently observed for general cases [27,28].



**Figure 5.** Simulated batch performance of MFC with 100 mg L<sup>-1</sup> initial substrate at different external resistors. (A) substrate concentration; (B) suspended biomass concentration; (C) external voltage; and (D) power generation.

**Table 3.** Simulated peak power and energy recovery from an external resistor, and overall energy efficiency ( $\eta_{overall}$ ) at different external resistors.

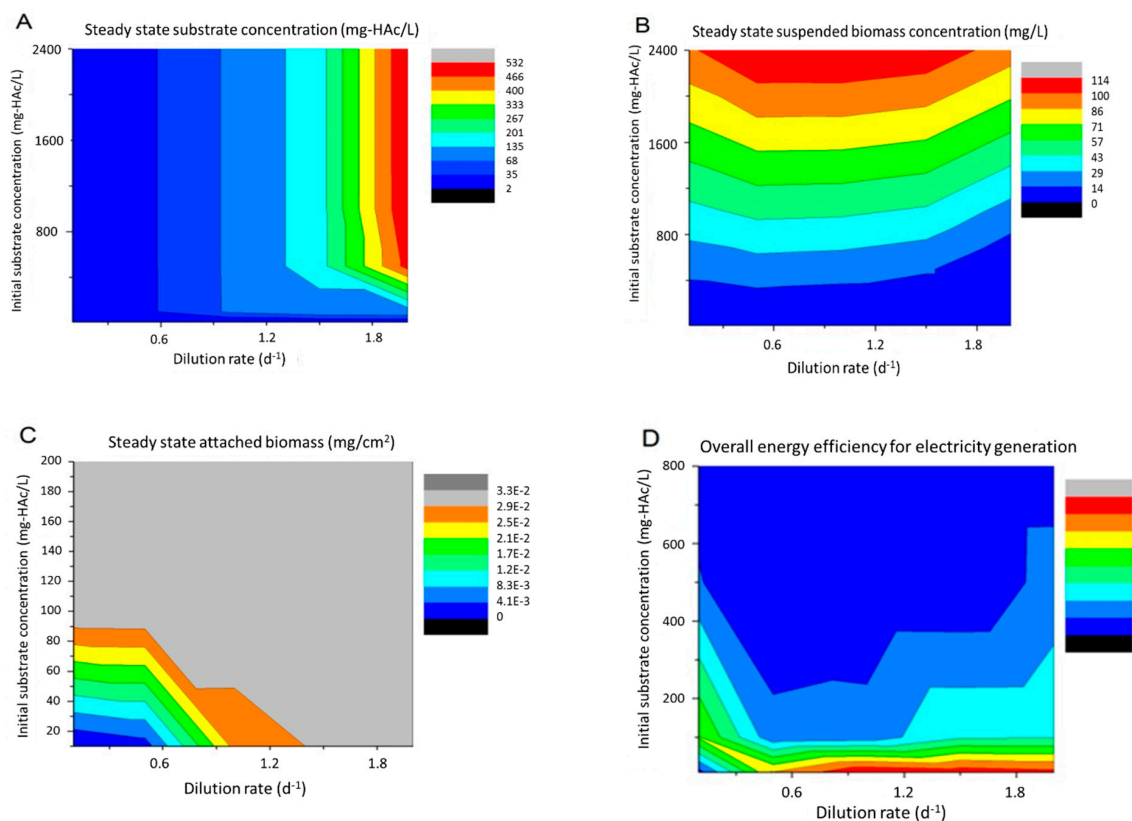
External Resistor	Peak Power	Energy Recovered	$\eta_{overall}$
10 $\Omega$	0.0142 mW	1.96 J	0.87%
300 $\Omega$	0.0998 mW	37.1 J	16.4%
1000 $\Omega$	0.0882 mW	47.3 J	20.7%
2200 $\Omega$	0.0685 mW	40.0 J	17.7%

### 3.3. Simulation of Continuous Mode MFC

#### 3.3.1. Effect of Influent Substrate Concentration and Dilution Rate

When MFC reactors were operated in continuous mode, the operating variables that might have pronounced effects were dilution rate ( $D$ ) and influent substrate concentration ( $S_{in}$ ). These two variables controlled the substrate inflow rate and biomass washout rate, and therefore would eventually impact MFC power generation and energy efficiency at steady state. So in the simulation, the dilution rate was varied between 0.1 to 2 d<sup>-1</sup>. The upper limit was chosen to be less than the maximum microbial specific growth rate (2.4 d<sup>-1</sup>) to prevent biomass washout. The influent substrate

(HAc) concentration was between 10 to 2400 mg L<sup>-1</sup>. The results, including the substrate concentration, suspended biomass concentration, attached biomass concentration and overall energy efficiency, were plotted in Figure 6. Please note that the external power ( $P_{ext}$ ) was not considered in the simulation of varying  $S_0$  and  $D$ , because it was found that when assuming the attached biomass was fully grown at steady state, the external power was a function of the external resistor, rather than a function of these two operating variables. It should be noted that little effort in MFC studies of varying these operating parameters was found in continuous mode, so literature data for comparison were not available.



**Figure 6.** Process variables and MFC performance predicted at steady state under different dilution rates and influent substrate concentrations. (A) steady state substrate concentration; (B) suspended biomass concentration; (C) attached biomass concentration, and (D) the overall energy efficiency  $\eta_{ext}$ .

At a given level of the influent substrate concentration, the steady state substrate concentration went up with the increasing dilution rate, basically due to the increased substrate inflow to the reactor (Figure 6A). When the influent substrate concentration was higher than 500 mg L<sup>-1</sup>, the concentration did not have much impact on the steady state substrate concentration at a given dilution rate. But when the influent concentration was lower than 500 mg L<sup>-1</sup> and the dilution rate was greater than 1.5 d<sup>-1</sup>, the steady state concentration increased along with the influent concentration. The suspended biomass concentration obviously increased with the influent substrate concentration in a linear manner at any dilution rate (Figure 6B). The dilution rate between 0.5 and 1 d<sup>-1</sup> resulted in a maximum suspended biomass at a given influent substrate concentration, because a lower rate caused substrate limitation and a higher rate could partially wash out suspended biomass. The attached biomass was more stable and could generally achieve its maximum in most cases (Figure 6C), except for the cases of  $S_{in}$ , which were much lower than 100 mg L<sup>-1</sup> and were too low to provide enough carbon sources for supporting the bacterial growth and attachment. As to the overall energy efficiency, it could be concluded that a higher influent substrate concentration reduced the efficiency (Figure 6D). The only exception was the case with about 10 mg L<sup>-1</sup> of influent substrate and 0.1 d<sup>-1</sup> of dilution rate, under which condition the attached biofilm was not fully mature, due to substrate deficiency,

so the MFC did not generate electricity. Considering the tradeoff between higher electrical power generation (reflected by the attached biomass concentration, which was the catalyst) and acceptable overall energy efficiency, it was recommended to operate MFC with about 100 mg L<sup>-1</sup> of influent substrate concentration. Meanwhile, at this substrate concentration, the choice of a lower dilution rate, e.g., 0.1 d<sup>-1</sup>, would help achieve lower effluent substrate and better removal efficiency, and the choice of a larger dilution rate, e.g., 2 d<sup>-1</sup>, might otherwise help generate more electric energy.

### 3.3.2. Effect of External Resistor

The next simulation was implemented for changing electrical currents by varying  $R_{ext}$  at the operating conditions of  $S_{in}$  and  $D$  listed in Table 4. The relationship between  $R_{ext}$  and current  $I$  was depicted in the inset of Figure 7A. These results were useful for determining a suitable range of the external resistor for better substrate removal, power generation or overall energy recovery. As shown in Figure 7A and Table 4, all cases resulted in a maximum power generation ( $P_{ext, max}$ ) of 0.106 mW at  $R_{ext}$  of 352  $\Omega$ , except for the case of the lowest substrate loading ( $S_{in} = 100$  mg L<sup>-1</sup> and  $D = 0.1$ ), which had a highest power of 0.091 mW at  $R_{ext}$  of 1007  $\Omega$ . For the lowest substrate loading, a further decrease of  $R_{ext}$  distributed more organic substrate utilization for heat generation and would finally lead to a system failure without electrical generation due to substrate limitation for biomass growth. On the contrary, all the MFCs with enough substrate generated the same external powers at given  $R_{ext}$ . Figure 7B plotted the results of MFC cell efficiency ( $\eta_{cell}$ , defined in Equation (28)), which showed that MFC cell efficiency decreased with the increasing circuit current (or the decreasing external resistor) because of the increased heat loss through the cell (Equation (24)). Again, the curve of the lowest substrate loading differed from the others and was interrupted at  $R_{ext}$  less than 928  $\Omega$  due to insufficient substrate available for biomass growth.

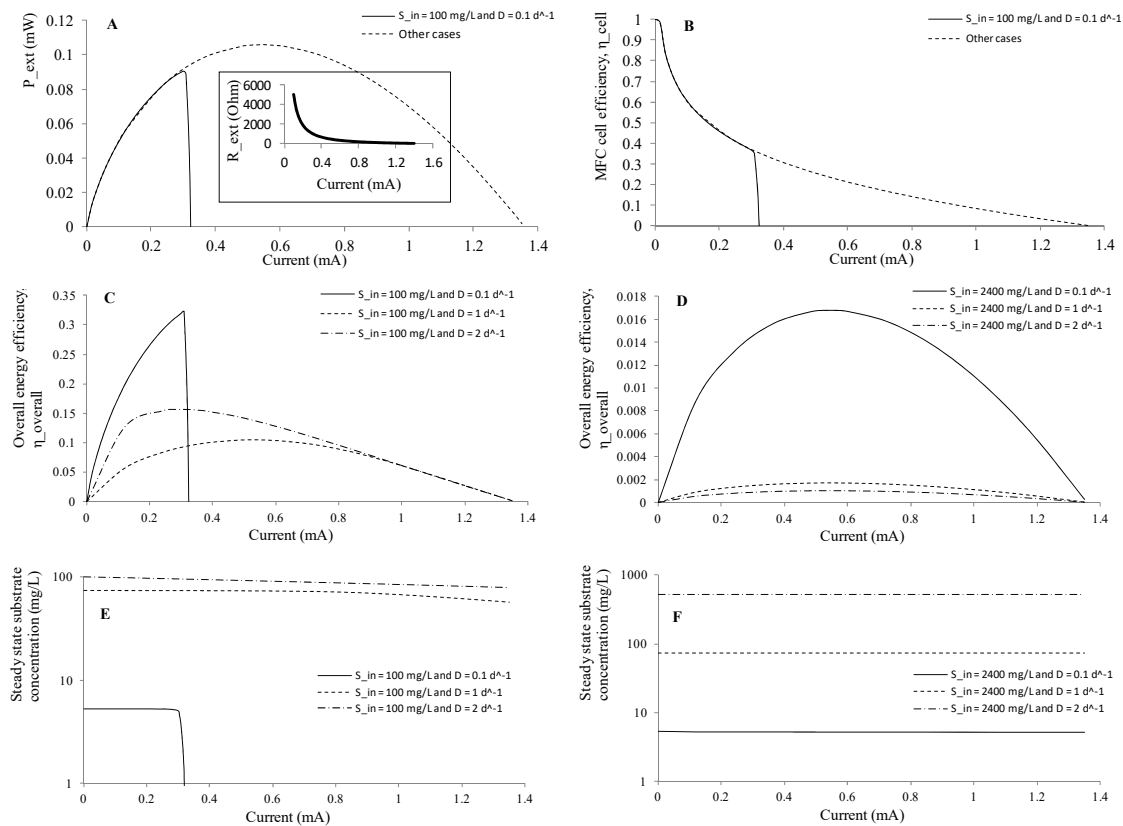
**Table 4.** Simulated MFC performance: substrate removal, power generation and overall energy efficiency.

$S_{in}$ mg/L	$D$ d <sup>-1</sup>	Steady State $S$ mg/L	$S$ Removal %	$P_{ext, max}$ mW	$\eta_{overall, max}$	$R_{in}$ <sup>a</sup> $\Omega$	$R_{ext, P}$ <sup>b</sup> $\Omega$	$R_{ext, \eta}$ <sup>c</sup> $\Omega$
100	0.1	0–5.26	>94.7	0.091	0.322	471	1007	928
	1	57.6–73.5	26.5–42.4	0.106	0.105		352	405
	2	79–100	<21.0	0.106	0.156		352	1017
2400	0.1	5.17–5.26	100	0.106	0.0167	471	352	352
	1	73.9	96.9	0.106	0.0017		352	352
	2	531	77.9	0.106	0.0011		352	352

<sup>a</sup> The internal resistance was estimated from the transiently simulated polarization curves in the electrical current range between 0.18 to 0.82 mA. <sup>b</sup> This was the value of the external resistor where the  $P_{ext, max}$  was achieved. <sup>c</sup> This was the value of the external resistor where the  $\eta_{overall, max}$  was achieved.

Figure 7C,D showed the overall energy efficiencies for the influent substrate concentrations of 100 and 2400 mg L<sup>-1</sup>, respectively. For the lower substrate cases, the maximum overall energy efficiencies ( $\eta_{overall, max}$ ) were between 0.105 and 0.322, and were achieved at different external resistors depending on dilution rates (928  $\Omega$  at  $D = 0.1$  d<sup>-1</sup>; 405  $\Omega$  at  $D = 1$  d<sup>-1</sup>; and 1017  $\Omega$  at  $D = 2$  d<sup>-1</sup>). For the higher substrate cases, the maximum overall energy efficiencies were between 0.0011 and 0.0167, and were achieved at  $R_{ext} = 352$   $\Omega$  at which the  $P_{ext, max}$  values were concurrently obtained. Therefore, the lower substrate concentration reactor had one or two orders of magnitude of higher overall efficiency than that of the higher substrate concentration. This substantial difference was a result of more proliferated biomass growth at higher substrate concentrations and the biomass in turn consumed more substrate. Please also note that the internal resistance  $R_{in}$  estimated by polarization curves was 471  $\Omega$ , and was independent of these operating parameters. The steady state substrate concentration ( $S$ ) and substrate removal were almost staying unchanging at different electrical current (Figure 7E,F), but depended on the influent substrate concentration and dilution rate, as discussed earlier. Removal efficiencies greater

than 94.7% or about 100% were achieved at  $S_{in} = 100 \text{ mg L}^{-1}$  and  $D = 0.1 \text{ d}^{-1}$ , and  $S_{in} = 2400 \text{ mg L}^{-1}$  and  $D = 0.1 \text{ d}^{-1}$ , respectively.



**Figure 7.** Simulated MFC performance at steady state under different electrical currents (or external resistors), dilution rates and influent substrate concentrations. (A) steady state power density; (B) MFC cell energy efficiency; (C) overall energy efficiency at  $S_{in} = 100 \text{ mg L}^{-1}$ ; and (D)  $S_{in} = 2400 \text{ mg L}^{-1}$ ; (E) effluent substrate concentration at  $S_{in} = 100 \text{ mg L}^{-1}$ ; and (F)  $S_{in} = 2400 \text{ mg L}^{-1}$ .

#### 4. Conclusions

The model proposed in this study was based on the assumption that the anode attached to the bacterial monolayer served as biocatalysts for MFC exoelectrogenesis. By modifying the Freter model and combining it with the Butler–Volmer equation, this model could adequately describe the processes of electricity generation, substrate utilization, and suspended and attached biomass growth, in both batch and continuous operating mode. MFC performances were impacted by various operating variables such as initial substrate concentration, external resistor, influent substrate concentration, and dilution rate, and their interactions, which were revealed by data simulation to be complicated. The simulation results explained that in batch mode, an intermediate initial substrate concentration ( $S_0$  about  $100 \text{ mg L}^{-1}$  at this reactor configuration) was appropriate to achieve maximum overall energy efficiency. With the  $S_0$  of  $100 \text{ mg L}^{-1}$ , an external resistor (about  $300 \Omega$ ) with the value of around the internal resistance ( $R_{in}$  of about  $471 \Omega$ ) could boost the power generation, and a resistor of about twice (about  $1000 \Omega$ ) that value achieved better overall energy efficiency. However, a small external resistor should be adopted for MFC reactors aiming to rapidly remove organic substrates. In continuous mode and at  $R_{ext} = 2200 \Omega$ , the anode-attached biomass could fully grow when the influent substrate concentration was equal to or higher than  $100 \text{ mg L}^{-1}$  at any dilution rates within the tested range. The maximum external power of  $0.106 \text{ mW}$  could be achieved at  $R_{ext} = 352 \Omega$  when there was enough substrate to sustain the power generation. An influent substrate concentration of  $100 \text{ mg L}^{-1}$  achieved the overall energy efficiency greater than 10%. The high substrate concentration, e.g.,  $2400 \text{ mg L}^{-1}$ ,



encouraged substantial biomass growth and thus reduced the overall energy efficiency to less than 2%. For the low influent substrate concentration ( $100 \text{ mg L}^{-1}$ ), the maximum power generation and overall energy efficiency could not be concurrently achieved, and operating MFC at an external resistor  $R_{ext}$  close to the internal resistance ( $R_{in}$ ) or two of  $R_{in}$  was recommended to obtain optimal values for both indices. Overall, this relatively simple model provided a convenient way for evaluating and optimizing performance of MFC reactors by regulating operating parameters.

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