


Review

Targeting the Cation-Chloride Co-Transporter NKCC1 to Re-Establish GABAergic Inhibition and an Appropriate Excitatory/Inhibitory Balance in Selective Neuronal Circuits: A Novel Approach for the Treatment of Alzheimer's Disease

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Abstract: GABA, the main inhibitory neurotransmitter in the adult brain, depolarizes and excites immature neurons because of an initially higher intracellular chloride concentration $[Cl^-]_i$ due to the delayed expression of the chloride exporter KCC2 at birth. Depolarization-induced calcium rise via NMDA receptors and voltage-dependent calcium channels is instrumental in shaping neuronal circuits and in controlling the excitatory (E)/inhibitory (I) balance in selective brain areas. An E/I imbalance accounts for cognitive impairment observed in several neuropsychiatric disorders. The aim of this review is to summarize recent data on the mechanisms by which alterations of GABAergic signaling alter the E/I balance in cortical and hippocampal neurons in Alzheimer's disease (AD) and the role of cation-chloride co-transporters in this process. In particular, we discuss the NGF and AD relationship and how mice engineered to express recombinant neutralizing anti-NGF antibodies (AD11 mice), which develop a neurodegenerative pathology reminiscent of that observed in AD patients, exhibit a depolarizing action of GABA due to KCC2 impairment. Treating AD and other forms of dementia with bumetanide, a selective NKCC1 antagonist, contributes to re-establishing a proper E/I balance in selective brain areas, leading to amelioration of AD symptoms and the slowing down of disease progression.

Keywords: depolarizing GABA_A-mediated neurotransmission; cation-chloride co-transporters; KCC2 dysfunction; Alzheimer's disease; NGF; AD11 transgenic mice; bumetanide treatment

1. Introduction

γ -Aminobutyric acid (GABA) is the main inhibitory neurotransmitter in the adult mammalian CNS. Released from GABAergic interneurons, GABA binds to two different classes of receptors: GABA_A and GABA_B. While GABA_A receptors are integral ion channels, permeable to chloride (Cl^-) and bicarbonate (HCO_3^-), GABA_B receptors are coupled to ion channels via guanine nucleotide-binding proteins and second messengers. GABA_A receptors mediate two distinct forms of inhibition: phasic and tonic [1,2]. The first consists of fast inhibitory postsynaptic potentials (IPSPs), regulating point-to-point communication between neurons. In this case, GABA_A receptors facing presynaptic release sites are activated by a brief exposure to a high concentration of GABA released by exocytosis of presynaptic vesicles. The second consists of a persistent inhibitory conductance, which is essential for regulating the membrane potential and network excitability. In this case, "ambient" GABA spills out from the synaptic cleft and persistently activates extra synaptic

GABA_A receptors, which are characterized by high affinity for GABA and relatively poor desensitizing properties [3]. Interestingly, early in postnatal development, GABA depolarizes and excites targeted cells by an outward flux of chloride due to a higher intracellular chloride concentration $[Cl^-]_i$ that results from the differential temporal expression of the cation-chloride co-transporters NKCC1 and KCC2, which are involved in Cl^- uptake and extrusion, respectively [4,5]. The low expression of the KCC2 extruder at birth leads to Cl^- accumulation inside the neuron via NKCC1 [6].

In adulthood, GABA, via feed-forward and feed-back inhibition, shapes the spatial and temporal profile of pyramidal cell firing, thus exerting a powerful control on neuronal excitability and network oscillations crucial for information processing [7,8]. Although GABAergic interneurons constitute only 15–20% of the total neuronal population, they are nevertheless instrumental in orchestrating the activity of principal cells and to maintain normal oscillatory activity in cortical circuits [7]. Oscillations, which occur at various frequencies supporting different behavioral states and high cognitive tasks, are altered in several neuro-psychiatric diseases including neurodegenerative disorders such as AD [8].

This review will focus on recent data on GABAergic dysfunction in AD, the most common cause of dementia worldwide with serious economic and social burden. Given the projected trends in population ageing and growth, the number of people with AD and other forms of dementia is expected to increase from 57.4 million in 2019 to 152.8 in 2050 [9]. AD research has been for years concentrated on the effects of cholinergic and glutamatergic transmission [10–12], while GABAergic transmission has been often neglected [13]. However, the observation that a subset of AD patients are affected by epilepsy, a neurological condition characterized by recurrent seizures caused by an excitatory (E)/inhibitory (I) imbalance [14–16], probably secondary to deficits of GABA_A-mediated inhibitory neurotransmission, suggests the involvement of GABAergic signaling, particularly in cortical and hippocampal networks, which are the most vulnerable brain regions in AD. It is worth mentioning that AD patients carrying the risk factor apolipoprotein E4, a protein involved in lipid and cholesterol transport as well as cell repair and regulating A β deposition and neurogenesis, exhibit a deficit in GABAergic signaling. The EEG of patients with moderate AD and young people carrying the *APOE4* variant often exhibit, with respect to age-matched controls, subclinical epileptiform abnormalities, reflecting a state of neuronal hyper-excitability due to a reduced GABA_A-mediated synaptic inhibition [17,18]. Further evidence in favor of a GABAergic dysfunction in AD is the positive effects on cognitive functions exerted by antiepileptic drugs [17,19,20]. Therefore, in the last years, increasing interest in the role played by GABAergic neurotransmission in AD has been developed in both humans and animal models of the disease.

2. Alterations of GABAergic Signaling in Alzheimer's Disease and in Animal Models

A decreased level of GABA has been found in the cerebrospinal fluid of aged people and AD patients [21]. Using magnetic resonance spectroscopy, a significant reduction of GABA levels has been detected in the parietal region of AD patients [22], an effect that worsens with age [23]. Postmortem studies from AD brains have demonstrated a reduced level of glutamate decarboxylase (GAD) [23], the enzyme responsible for converting glutamate to GABA, an effect associated with a decrease in perisomatic GABAergic terminals, especially in cortical neurons adjacent to amyloid plaques [24,25]. In addition, a reduction of GABAergic interneurons containing somatostatin [22,26], calcium binding protein calretinin [27], calbindin [28], and parvalbumin [29] has been found in brain regions expressing A β and tau pathologies as well as neuroinflammation such as the neocortex and the hippocampus [30]. A decrease in somatostatin has been also found in the cerebrospinal fluid of patients with EEG abnormalities and cognitive deficits [31]. A loss of GABA_A receptors was also reported in the brain from AD patients [32,33]. Using selective antibodies, $\alpha 1$ and $\alpha 5$ subunits of GABA_A receptor were found to be reduced in the hippocampus in a region-specific way, whereas $\beta 1$, $\beta 2$, $\beta 3$, and $\gamma 2$ subunits were found to be unaffected [34]. While $\alpha 1$ subunits are expressed on synaptic GABA_A receptors, $\alpha 5$ are mainly present on

extra synaptic receptors, which are involved in tonic inhibition. A significant reduction of GABA_B receptors' immunoreactivity has been also detected in the CA1 region of the hippocampus of AD patients [35]. Furthermore, an age-dependent downregulation of currents evoked by exogenous application of GABA was found in *Xenopus oocytes* transplanted with cell membranes, isolated from human temporal cortex of AD patients and age-matched controls. These currents exhibited a faster desensitization rate and were associated with a reduction of $\alpha 1$ and $\gamma 2$ subunits and an increased expression of $\alpha 2$, $\beta 1$, and $\gamma 1$ subunits of GABA_A receptors, suggesting an AD-dependent re-modelling, resulting in atypical GABAergic signaling [36].

Mouse models carrying a specific set of mutations found in humans and mimicking various aspects of AD pathology have been very useful to better understand the molecular and cellular mechanisms underlying this devastating disorder. TgCRND8 mice, overexpressing the mutant human APP at levels approximately 5-fold higher than endogenous murine APP, which exhibit early A β deposition, show a loss of GABAergic interneurons already at 6 months [37]. In line with these results, transgenic mice of the APP/PS1 models, which overexpress the Swedish family mutated forms of the human amyloid precursor protein and various mutations in presenilin 1, exhibit a significant (50/60%) reduction in the number of GABAergic neurons co-expressing somatostatin (SOM) and neuropeptide Y (NPY), which precedes principal cells loss [38]. This results in an age-dependent increase in neuronal excitability due to E/I imbalance and in spontaneous seizures [39]. The loss of SOM/NPY⁺ GABAergic interneurons, which is associated with the accumulation of A β plaques in the extracellular space and to microglial-induced excitotoxicity, was detected also in the entorhinal cortex of six-month-old mice [40]. The E/I imbalance, due to dysfunction or loss of GABAergic interneurons in the dentate gyrus, may account for aging-related memory deficits in AD [7]. It has been also shown that the loss of GABAergic interneurons in the hippocampal hilus of mice carrying the AD risk factor apolipoprotein E4 (*APOE4* knock-in mice) leads to severe age-dependent learning and memory deficits [41–43], an effect that can be rescued by transplanting medial ganglionic eminence-derived inhibitory interneuron progenitors into the hilus [43]. Fast spiking, parvalbumin (PV)-containing interneurons pace, via feed-forward inhibition, principal cells, giving rise to gamma oscillations thought to be involved in high cognitive functions [44]. A reduced expression of PV⁺ interneurons has been found in the CA1 region of the hippocampus of 3xTg-AD mice [45] that harbor the mutant genes for amyloid precursor protein (APPS_{we}), presenilin 1 (PS1_{M146V}) and tau_{P301L}.

It is worth mentioning that AD is often associated not only to alterations of GABA_A-mediated phasic inhibition but also to modifications of the tonic one, probably consequent to the enhanced GABA release from glial cells that express GABA_A receptors [46]. In 5XFAD mice, which overexpress human *APP* with three FAD mutations (the Swedish (K670N, M671L), Florida (I716V), and London (V717I) mutations) and human *PSEN1* with two FAD mutations (M146L and L286V), the neuron–glia interaction-mediated enhancement of tonic inhibition would suppress long-term potentiation resulting in memory deficits [47]. The increase in ambient GABA would also reduce ripples oscillations, which are known to be involved in memory consolidation [48]. Other factors contributing to the enhancement of ambient GABA levels include changes in GABA uptake and clearance and expression of GABA_A receptor subunits and alterations in GABA transporters. A similar increase in tonic inhibition, mediated by the activation of extra-synaptic $\alpha 5$ subunits containing GABA_A receptors, was induced in mice by microinjecting A β _{1–42} in the CA1 region of the hippocampus, an area critical for episodic memory. This led to disruption of the E/I balance and cognitive decline [49]. Additionally, reactive astrocytes activated by A β have been shown to contribute to memory's impairment in AD by enhancing ambient GABA with consequent inhibition of synaptic function via GABA_A and GABA_B receptors [50].

AD has been found to be associated with a downregulation of GABA_B receptors. These receptors act pre-synaptically by inhibiting the release of a variety of neurotransmitters and post-synaptically by generating inhibitory K⁺ currents, which hyperpolarize the membrane

and inhibit neuronal firing [51]. As compared to controls, a significant reduction in total protein and mRNA for all GABA_B receptor subunits was detected in the hippocampus of six-month-old APP/PS1 mice in the presence of accumulating amyloid plaques. These changes would act by modifying the overall inhibitory tone, leading to behavioral deficits in spatial learning and memory [52]. Since GABA_B receptors are present not only on neurons but also on microglia, the possibility that their loss in AD leads to microglia dysfunction and to an inflammatory response cannot be excluded. In addition, using high-resolution microscopy and quantitative approaches, a significant reduction in the surface expression of GABA_B receptors in the granule cells of the DG of 12-month-old APP/PS1 mice was found. This was associated to a large amount of senile plaques and a severe impairment of cognitive functions [53]. Furthermore, the genetic loss of APP, which is known to interact with N-terminal Sushi domain of presynaptic GABA_B receptors, led to impairment of GABA_B receptor-mediated presynaptic inhibition of glutamate release by limiting presynaptic GABA_B receptor trafficking and the availability of APP for endosomal processing to A β , with consequent enhancement of A β production ([54]; Figure 1). Overall, the above-mentioned observations made in humans and in animal models of AD point to GABA and its receptors as key players in AD onset and progression. Evidence has been recently provided that, by physically interacting with the cation-chloride extruder KCC2, G-protein-coupled GABA_B receptors contribute to regulate fast synaptic GABA_A-mediated inhibition through the rapid and sustained change in the ionic driving force for the chloride-permeable GABA_A receptors [55]. Therefore, by changing chloride homeostasis and particularly by down regulating the cation-chloride extruder KCC2 in selective brain areas, it is possible in particular conditions to modify the E/I balance leading to an enhanced neuronal excitability consequent to the shift of GABA from the hyperpolarizing to the depolarizing direction as in the immature brain (Figure 1). The polarity of GABAergic neurotransmission is modulated also by the amyloid precursor protein (APP). In *in vitro* experiments from primary cortical cell cultures expressing human APP, a downregulation of KCC2 but not NKCC1 was detected. This effect was independent of the APP intracellular domain but was correlated with a reduced expression of transcriptional regulator upstream stimulating factor 1 (USF1; [56]; Figure 1). Downregulation of KCC2 led to GABA-induced rise of [Ca²⁺]_i, an effect that was prevented by the NKCC1 inhibitor bumetanide, indicating that in hAPP-expressing neurons, changes in [Cl⁻]_i may contribute to GABA-induced increase in [Ca²⁺]_i. The impact of APP on KCC2 was age-dependent, as demonstrated by the observation that the absence of APP in 3-month-old APP^{-/-} mice induced an increased expression of KCC2 but in 8-month-old animals a decrease. This was in accordance with the results of Chen et al. [57], indicating that APP can regulate KCC2 protein level by different mechanisms depending on the age examined.

It should be stressed that cation-chloride cotransporters are very labile, and they can be disrupted in several neuropsychiatric disorders [5,58]. Rapid changes in KCC2 function can be elicited in an activity-dependent fashion and involve different post-translational regulation of transporter proteins, including their phosphorylation and regulation at the cell surface [59].

A decrease of KCC2 expression was observed in the hippocampus of mice engineered to express recombinant neutralizing anti-nerve growth factor (NGF) antibodies (AD11 mice), which develop an age-dependent neurodegenerative pathology reminiscent of that observed in AD patients ([60]; Figure 1). Although the mechanisms linking NGF deprivation to KCC2 downregulation remain to be established, this observation has nevertheless opened promising perspectives towards the development of more effective and specific treatments and finalized to restore a proper E/I balance in AD patients. In the following sections, we discuss in detail the relationship between NGF and AD and the AD11 mouse model of AD. We also discuss how treating AD and other forms of dementia with the selective KCC2 antagonist bumetanide can contribute to re-establishing a proper E/I balance in selective brain circuits, leading to amelioration of AD symptoms and the slowing down of AD progression.

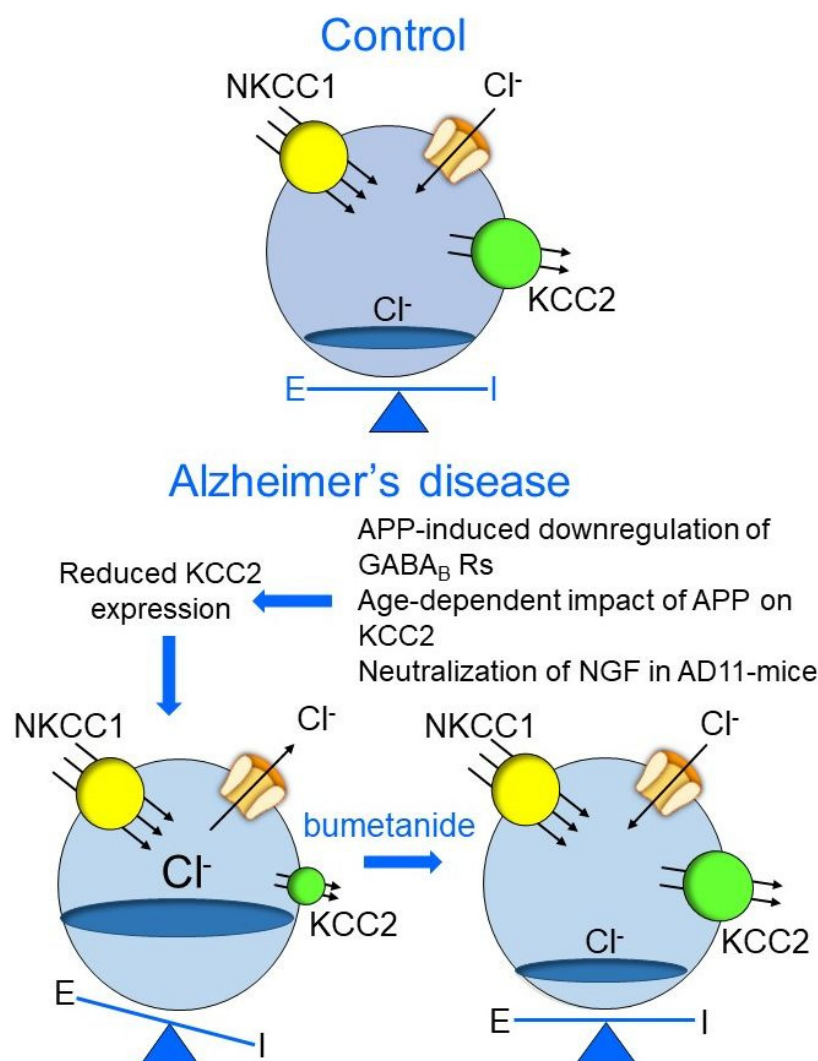


Figure 1. Altered chloride homeostasis in AD. In the adult brain (top), the intracellular levels of chloride, are maintained at very low levels by the cation-chloride importer and exporter NKCC1 and KCC2, respectively. GABA binds to its receptor and opens the channel, giving rise to an inward flux of chloride, which inhibits targeted cells by hyperpolarizing the membrane. In this way, it contributes to preserve the appropriate E/I balance in selective neuronal circuits. In AD, the APP-induced downregulation of GABA_B Rs, the age-dependent impact of APP on KCC2, or neutralization of NGF in AD11 mice allows GABA to shift from the hyperpolarizing to the depolarizing direction, as in immature neurons, giving rise to an outwardly directed flux of chloride and E/I imbalance with consequent enhancement of network excitability (down left), which is an effect that can be rescued by blocking NKCC1 with bumetanide (down right).

3. NGF and Alzheimer's Disease

3.1. NGF and the Cholinergic Hypothesis for AD

Memory loss in AD patients has been found to be associated with a selective reduction of basal forebrain cholinergic neurons (BFCNs) [61,62], which provides projections to subcortical and cortical regions, including the neocortex and the hippocampus, which are known to be involved in high cognitive functions such as learning and memory [63]. A significant loss of choline acetyltransferase (ChAT) and acetylcholinesterase (AChE) [64,65], enzymes responsible for the synthesis and breakdown of acetylcholine, respectively, can be observed in the cortex. The progressive deterioration of cholinergic function in AD has led to the so-called "cholinergic hypothesis" [66,67], which gave rise to numerous studies aimed at developing therapeutic tools for delaying or preventing cholinergic neurodegeneration.

The neurotrophin nerve growth factor (NGF), discovered in 1952 by Rita Levi-Montalcini [68], was thought to be involved in the BFCNs loss, among other epigenetic factors.

NGF is primarily involved in the growth as well as in the maintenance, proliferation, and survival of peripheral and central neurons during development and throughout adulthood [69–72].

The link between NGF and AD was hypothesized a long time ago based on the effects of NGF on BFCNs, which rely on this neurotrophin for their maintenance and survival [73,74]. mRNA encoding for both NGF and NGF protein has been detected in BFCNs targeted areas [75–78]. This neurotrophin, which is selectively internalized and retrogradely transported via axonal projections from the cortex to BFCNs [79], selectively binds to TrkA and p75^{NTR}, which are expressed on both soma and axons of BFCNs [80–82]. Respond to the exogenous administration of NGF with an increase of cholinergic phenotypical markers [73,83]. Most importantly, NGF is able to prevent BFCN death or atrophy following axotomy [74,84,85] or linked to aging [86].

3.2. NGF and the Modulation of APP Processing via TrkA and p75 Receptors

Alterations of NGF signaling have been associated not only to cholinergic dysfunction but also to altered processing of APP and tau. Indeed, an aging pathway has been shown to control the TrkA to p75^{NTR} receptor switch and amyloid β ($A\beta$) production in cells [87]. Moreover, $A\beta$ peptide binds trimers as well as monomers of p75^{NTR} and activates receptor signaling [88]. On one hand, the p75^{NTR} undergoes regulated intramembrane proteolysis by β -secretase, with the β -cleavage sites of APP and p75^{NTR} being highly homologous, releasing p75^{ICD} (intracellular domain) to transmit a signal to the nucleus in analogy to the ICD signal derived by the cleavage of APP [89]. Further direct links between NGF deficits and the activation of the AD amyloidogenic pathway have been demonstrated in cultured hippocampal neurons [90]. The withdrawal of NGF from hippocampal cells induces a rapid increase in APP levels and an overproduction of $A\beta$ peptide, which is followed by apoptotic death. Several studies clearly indicate that extensive tau hyperphosphorylation occur upon NGF deprivation in NGF-differentiated PC12 cells [91–93]. In hippocampal cells, a transient tau hyperphosphorylation, temporally related to the activation of the AD-like amyloidogenic pathway, occurs after NGF deprivation [94]. This provides evidence for a mechanism in which a discontinued or limited supply of NGF can activate a pathological pathway of APP and tau processing, triggering downstream apoptotic cell death [95]. This phenomenon is temporally and causally related with an activation of the endogenous amyloidogenic pathway, which was previously reported in hippocampal neurons undergoing cell death upon NGF withdrawal [90]. These *in vitro* data are reinforced by *in vivo* data in the AD11 anti-NGF mouse model.

3.3. AD11 Anti-NGF Mice: A Link between NGF Deprivation and Alzheimer's-like $A\beta$ and Tau Neurodegeneration

A further proof of a link between NGF deprivation and AD comes from *in vivo* studies on AD11 anti-NGF mice. The mice were obtained using the “neuroantibody approach”, according to which non-immune cells can be engineered to express genes encoding for functional immunoglobulins [96–98]. Thus, AD11 mice produce the recombinant antibody anti-NGF α D11, which is known to neutralize NGF biological activity [96,97], under the transcriptional control of the human cytomegalovirus promoter. Anti-NGF antibodies, produced in non-lymphoid cells, are expressed in almost all peripheral mouse tissues and in the brain [98]. The phenotype of these AD11 anti-NGF mice resembles the one described in the sporadic human AD, showing synaptic and behavioral deficits. Since the transgenic anti-NGF antibodies are higher in adult than in newborn mice, inhibition of NGF actions effectively occurs in adult animals only [98]. Thus, anti-NGF transgenic mice show an age-dependent neurodegenerative pathology. Indeed, after the second postnatal month, the anti-NGF antibody levels increase by at least three orders of magnitude (up to 200 ng/mL),

and neuronal NGF targets, including sympathetic, sensory, and BFCNs, become severely affected [60]. At two to three months of age, mice show a behavioral impairment [98,99] linked to a starting cholinergic deficit and accumulation of hyper-phosphorylated tau in the entorhinal cortex [100]. At six months of age, AD11 mice start to accumulate A β intracellularly in hippocampal dystrophic neurites [100,101]. Intracellular accumulation of tau spreads to other cortical regions and to the hippocampus [100]. At this age, AD11 mice show also a further reduction of cholinergic innervation and synaptic plasticity impairment [102–104]. Behavioral deficits appear to be more severe, including object recognition and object location deficits [99]. By 15 months of age, A β deposition can be found extracellularly [101], accompanied insoluble and hyper-phosphorylated tau and neurofibrillary tangles in cortical and hippocampal neurons [60,101]. They also display cortical neuronal loss, cholinergic deficit in the BFCNs, and spatial memory deficits [60,98,99]. Thus, we conclude that the link between deficits or alterations in the NGF system and AD go well beyond its long-established neurotrophic actions on BFCNs. The failure or imbalance of NGF support could be due to different causes in the overall cascade(s) of events involving NGF bioactivity: (1) decreased NGF synthesis, (2) unbalanced or altered processing, (3) alterations in receptor expression and/or activity or expression ratios, and (4) altered retrograde transport [105]. These events would be “located” upstream of the “amyloid cascade”, which is the central core of AD neurodegeneration [106], and would be part of a negative feedback loop that involves several steps (e.g., links between APP, tau, and axonal transport) [100,107,108].

This conclusion that NGF actions in the brain could be more widespread than envisaged so far calls for an important question about the targets for new targets of NGF actions in the CNS.

3.4. Microglia as New Cellular Targets of NGF Actions in the Adult CNS

What are the cellular targets for NGF actions in this negative loop scheme? This question led to investigate whether non-neuronal cells might be NGF targets in the brain. Indeed, it is known that NGF acts not only on neurons but also non-neuronal cells, including astrocytes, oligodendrocytes, microglia, immune system cells, and endothelial blood vessel cells.

Transcriptomics in the AD11 model have shown that neuroinflammation is the earliest phenotypic alteration and is already present at a presymptomatic phase at 1 month of age [100,105,109], anticipating the finding that NGF exerts a broad neuroprotection not only on TrkA expressing neuronal cells but also on the resident immune cells of the brain, i.e., microglia. Indeed, while NGF has been previously reported to be able to modulate microglial cells in culture [110], we specifically found that this neurotrophin expresses functional NGF receptors in vitro and ex vivo with activation of the downstream intracellular signaling [107,108]. Transcriptomic analysis performed after incubation with NGF showed a modulation of motility, phagocytosis, and degradation pathways. These data were confirmed by functional analysis, showing that NGF can increase microglia membrane dynamics and to activate an outward rectifying current that appears to modulate neuronal glutamatergic neurotransmission [111]. In the same study, we focused on the effects on inflammation and A β clearance. NGF was able to reduce the expression of inflammatory cytokines and to increase macro-pinocytosis of A β toxic soluble fractions. In addition, NGF-treated microglia protected neurons from A β -induced loss of spines and chemically-induced LTP deficits [111]. Thus, it can be concluded that NGF exerts a neuroprotective action not only by acting on BFCNs but also by steering microglia towards an homeostatic neuroprotective phenotype [111].

3.5. proNGF/NGF Dysmetabolism Is a Trigger for Neurodegeneration

The mechanism of neurodegeneration in AD11 mice is due to the fact that the anti NGF antibody, expressed in the transgenic mice, binds mature NGF with a 2000-fold higher affinity than that for its precursor, proNGF [112]. This creates a complete neutralization

of NGF and an excess free proNGF. Thus, the AD11 mice are a model of proNGF to NGF imbalance [113]. In line with this, we demonstrated that (i) inhibiting TrkA [114] or (ii) overexpressing uncleavable proNGF [113] likewise induce a progressive neurodegeneration. Importantly, crossing AD11 mice to a p75^{-/-}-knockout rescues the amyloid dependent neurodegeneration [114].

The negative feedback loop linking proNGF/NGF imbalance to the onset of AD neurodegeneration [107,113] received independent support by evidence linking effects of A β on proNGF processing by Cuello's group [115] and by evidence from human AD brains [116,117]. Among the most recent studies linking NGF deprivation to AD are postmortem analysis of AD brains demonstrating high levels of NGF and its precursor proNGF in the neocortex and hippocampus and low levels in the basal forebrain [118–120], suggesting a defect in NGF retrograde transport [108].

Moreover, a large body of evidence has shown that not only NGF but also its precursor proNGF, its downstream signaling pathways, as well as the proteases that are able to cut proNGF and to release the mature form of the neurotrophin are involved in the balance between the physiological and pathological phenotype. As other neurotrophins, NGF is translated as a pre-pro-protein [121]. In the trans-Golgi network, proNGF is cleaved by furin to release the mature form of NGF [122–124]. Other proteases besides furin can cleave the pro-peptide in the extracellular space [125,126]. ProNGF is not a mere precursor of NGF, but it behaves as an intramolecular chaperon to ensure the proper folding and secretion of the neurotrophin [127]. Interestingly, with respect to the mature form of NGF, proNGF can act in a synergic or opposite way [120–131] depending on relative proNGF/NGF ratio and on the type of receptors activated. This would lead to a pro-apoptotic or pro-survival outcome [122,129,131]. NGF and proNGF exert their action through the activation of three distinct receptor subtypes: TrkA, p75^{NTR}, and sortilin. TrkA is a member of tyrosine kinases receptors family; p75^{NTR} is a member of the tumor necrosis factor receptor (TNFR) superfamily [132], and sortilin is a member of the family of Vps10p-domain transmembrane receptors. Sortilin is the specific receptor of proNGF [133,134], while p75^{NTR} and TrkA can be bound by both NGF and proNGF [135]. By interacting with the co-receptor complex of the p75^{NTR} and sortilin, ProNGF can activate the cell death signaling pathway [133]. p75^{NTR} and TrkA receptors are frequently co-expressed and physically interact with each other, thus cooperating in transducing NGF signals [134]. TrkA preferentially binds to the mature form of NGF, while p75^{NTR} promotes cell death in response NGF but only in the absence of TrkA. [136–138]. In fact, although both TrkA and p75^{NTR} can individually bind neurotrophins to elicit independent signaling events, the two receptors work together to enhance trophic signaling during development and in the healthy brain by mediating at least 10-fold higher affinity of the TrkA receptor for NGF [139–142]. On the contrary, in the absence of TrkA or in association to sortilin, activation of p75^{NTR} by proNGF mediates pro-apoptotic signaling in development and/or in neurodegenerative conditions [142,143]. It has been also demonstrated that, in particular conditions, proNGF can bind trkA and p75^{NTR}, giving rise to a pro-survival outcome [116,122,144].

It is worth noting that also the proteases engaged in proNGF cleavage and in NGF degradation are involved in NGF metabolism, and their fine tuning contributes to maintaining the physiological conditions in the brain [115]. In synthesis, the mature NGF is produced via a metabolic cascade in which proNGF, zymogens, convertases, and their endogenous regulators are co-secreted from BFCN-targeted neurons in an activity-dependent manner. Upon the extracellular release of these factors, the inactive zymogen plasminogen is cleaved into its active enzyme form, plasmin, by tissue plasminogen activator (tPA) in a process regulated by the tPA inhibitor, neuroserpin. Plasmin is then responsible for the cleavage of the prodomain of proNGF and the release of mature NGF. Moreover, the degradation of the mature NGF is accomplished by the metalloproteases MMP-3 and MMP-9. Their precursors, proMMP-9 and proMMP-3, are regulated by the tissue inhibitor of metalloproteinases-1 (TIMP-1), which cleaves them into their mature forms. According to Cuello's theory, an alteration of NGF metabolism is able to explain the pathological process

in AD, solving the paradox that, in the AD brains, BFCNs degenerate, while the levels of NGF transcripts remain unchanged [111,139,140], and the protein levels of proNGF are greatly elevated [105,118,119,145–147]. The NGF dysmetabolism was confirmed both in AD and in down syndrome postmortem brains. [117,148]. Importantly, an involvement of proNGF in some aspects of the AD-like pathogenesis has been confirmed in vivo through the production and characterization of a transgenic mouse over-expressing proNGF, in which learning and memory deficits, increased A β -peptide immunoreactivity, and excitatory/inhibitory imbalance were observed [149].

Thus, after more than five decades of studies on NGF and AD, it became evident that the relationship between NGF deprivation and AD cannot be limited to the cholinergic hypothesis. However, in spite the emergence of other pathogenetic mechanisms [150], NGF is still considered a good therapeutic candidate for AD.

4. AD11 Anti-NGF Mice: A Link between NGF Deprivation, Dysfunction of GABAergic, and Cholinergic Signaling and AD

Besides neuroinflammation, a decreased expression of ChAT is one of the first signs of degeneration in AD11 brains [98,100]. Consistent with this view, we have provided evidence that, in adult AD11 mice, deficits in cholinergic signaling in the hippocampus, a key structure for learning and memory processes, are mediated by α 7 subtypes of nAChRs, which are known to be highly permeable to calcium. A β would interfere with α 7 nAChRs, which is localized on glutamatergic terminals, thus preventing nicotine-induced enhancement of synaptic efficacy at CA3-CA1 synapses, probably via a perturbed calcium homeostasis and consequent dysfunction of downstream transduction pathways [151]. Furthermore, we found that in both control and NGF-deprived mice, nicotine, via α 7- and β 2-containing nAChRs, was able at CA3-CA1 synapses to lower the threshold for the induction of long-term potentiation (LTP) and to convert short-term potentiation (STP) or weak LTP into robust LTP, an effect that in AD11 but not in control mice was prevented by GABA_A receptor antagonists [152]. The failure of nicotine to boost activity-dependent synaptic plasticity in the presence of bicuculline or gabazine strongly suggests that deficits of cholinergic signaling in AD involve not only glutamatergic but also GABAergic transmission. The latter would be able to rescue nicotine-induced enhancement of synaptic plasticity via re-arrangements of the GABAergic circuit, including the shift of GABA from the hyperpolarizing to the depolarizing direction, which, by removal the magnesium block from NMDA receptors, would facilitate their activation [152]. Thus, in cell-attach recordings that affect neither the resting membrane potential (RPM) nor the $[Cl^-]_i$, from hippocampal slices of 6-month-old AD11 mice (but not age-matched controls), pressure application of the GABA agonist isoguvacine to CA1 principal cells produced a significant increase in firing rate, which is an effect that was prevented by the GABA_A receptor antagonist gabazine [153].

In addition, while in AD11 mice, GABA-evoked single channel currents reversed at more positive values respect to controls, NMDA-evoked single channel currents, which are used as voltage sensors to measure the resting membrane potentials, reversed at the same membrane potential of control animals (Figure 2A,B). Furthermore, in gramicidin-perforated patch, to preserve the intracellular chloride gradient, the equilibrium potential of GABA_A-mediated synaptic currents (E_{GSPCS}) was more positive in AD11 mice with respect to controls (Figure 2B–D). The driving force (ΔF) equal to (RPM- E_{GSPCS}) was positive in AD11 and negative in control mice. These effects could be rescued by the NKCC1 antagonist bumetanide, suggesting that they resulted from accumulation of $[Cl^-]_i$ and the shift of GABA from the hyperpolarizing to the depolarizing direction. These results were associated with a downregulation of the KCC2 cation-chloride exporter protein [153].

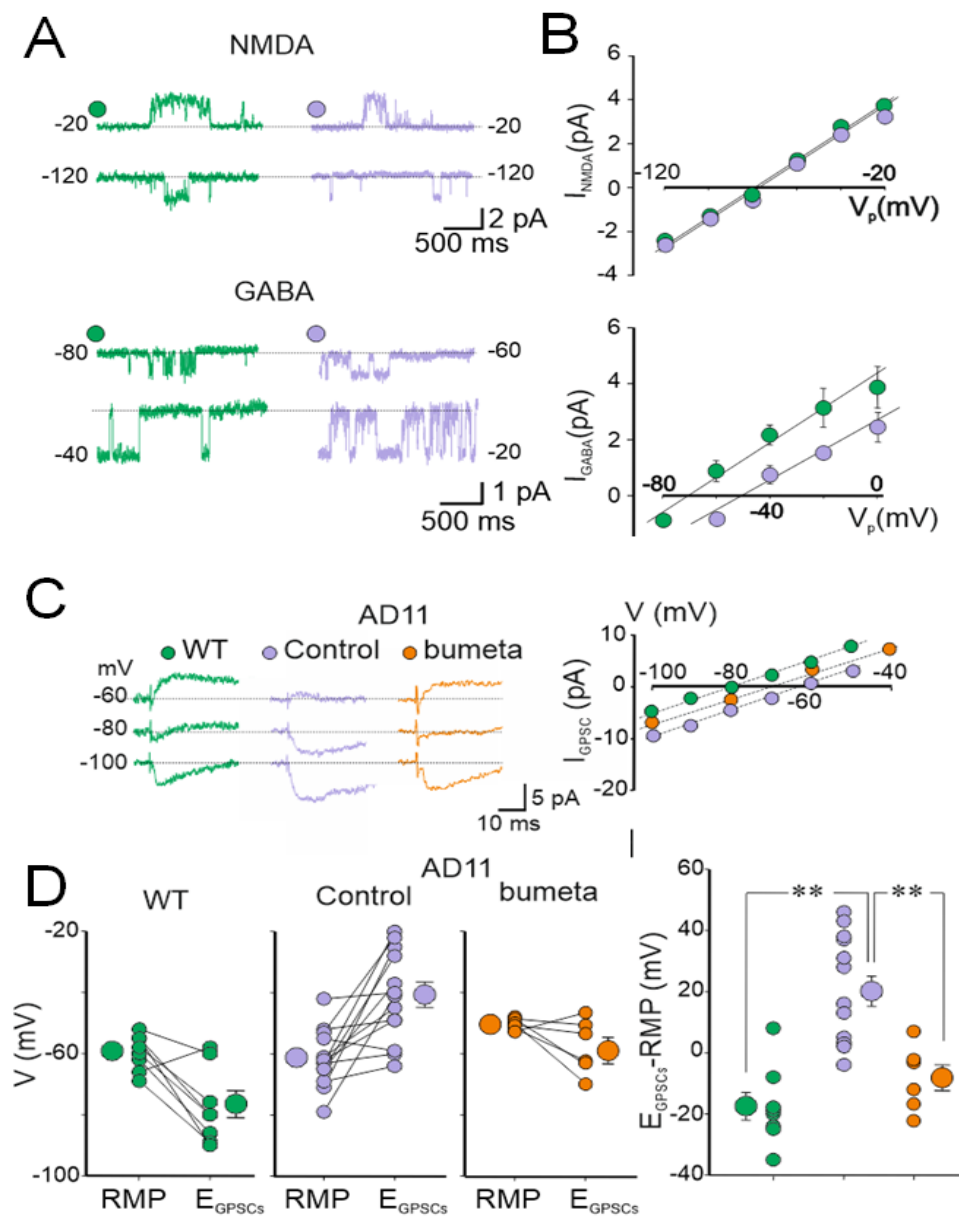


Figure 2. Depolarizing action of GABA in six-month-old AD11 mice. (A) Cell-attached recordings of single NMDA and GABA_A receptor channels obtained at two different pipette potentials in neurons from WT (green) and AD11 (violet) mice. (B) Summary plot of single NMDA (I_{NMDA}) and GABA (I_{GABA}) currents versus pipette potentials (V_p). Single NMDA currents reversed at -75.3 mV and -76.2 mV in WT and AD11 mice, respectively. Assuming a reversal of NMDA currents equal to 0 mV, we estimated a mean resting membrane potential of -75 mV and -76 mV, respectively. In contrast, single GABA_A currents reversed at -76 mV and at -56 mV in WT and AD11 mice, respectively. (C) Left: Perforated patch recordings of GPSCs evoked in CA3 principal cells by local stimulation of GABAergic interneurons in the presence of DNQX ($20 \mu\text{M}$) and D-AP-5 ($50 \mu\text{M}$) at three different holding potentials in hippocampal slices from WT (green), AD11 (violet), and AD11 mice exposed to bumetanide (yellow). Right: Synaptic currents (I_{GPSCs}) shown in A are plotted versus membrane potentials (V_m). (D) Left: individual RMPs and E_{GPSCs} values in hippocampal slices obtained from WT (green), AD11 (violet), and AD11 exposed to bumetanide (yellow). Larger symbols on the left and right refer to mean \pm SEM values. Right: plot of the driving force for GABA_A ($E_{\text{GABA-RMP}}$) in individual experiments from the three different groups of mice. ** $p < 0.001$ (Modified from [153]).

Interestingly enough, an involvement GABAergic transmission in AD11 mice has also been suggested by the transcriptomic analysis [154,155], which has demonstrated a

decreased expression of $\alpha 2$ subunit of GABA_A receptor at postnatal day 30 in the cerebral cortex and at P90 also in the hippocampus and basal forebrain region of AD11 [105,150,151], indicating a generally reduced inhibitory drive in the hippocampus and cortex, which is, at this age, a sign of a more general synaptic remodeling. The transcriptomic pattern for the GABAergic system measured in 6-month-old AD11 mice clearly shows a decreased expression of genes encoding for $\alpha 2$, δ , and $\gamma 1$ subunits of GABA_A receptors in the basal forebrain and for $\alpha 2$, $\alpha 4$, and $\alpha 5$ subunits in the hippocampus but not in the cerebral cortex. This effect is coupled to a decreased expression of the gene encoding for KCC2 (Figure 3).

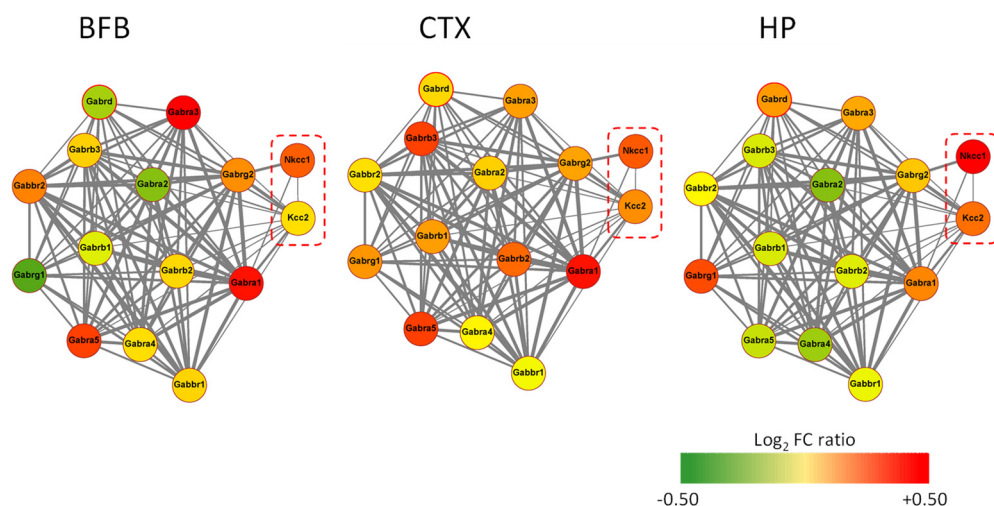


Figure 3. Interaction network in AD11 mice for NKCC1, KCC2, GABA_A, and GABA_B receptor genes. Interaction network extracted from the STRING database (<https://string-db.org/>, high interaction confidence level) for the NKCC1, KCC2 cation-chloride co-transporter genes (encircled in red), and the GABA_A, GABA_B receptor channel subunits genes in mouse. The node color corresponds to the Log₂-fold change-relative expression ratio in 6-month-old AD11 mouse compared to control mouse in three brain areas (basal forebrain, BFB; cortex, CTX; and hippocampus, HP). The thickness of grey edges between nodes corresponds to the interaction reliability provided by STRING.

It is worth noting that mice overexpressing proNGF, the precursor of mature NGF, exhibited spontaneous epileptic-like events as the result of a severe E/I imbalance, probably related to a significant reduction of parvalbumin-positive GABAergic interneurons associated with a perineuronal net depletion in the dentate gyrus [149]. However, in this case, the transcriptome analysis unveiled that genes encoding for NKCC1 and KCC2 were not significantly altered.

5. Rescuing a Proper GABAergic Signaling for the Treatment of AD by Repurposed Drugs

AD suffers the lack of effective disease-modifying drugs, which are able to directly interfere with the pathogenetic mechanisms of the neurodegenerative condition to reverse or at least stop or significantly slow down the disease trajectory both at the cellular and subcellular level and at the clinical phenotype level. The last decades have seen a large number of failures for randomized clinical trials of pharmaceutical compounds for AD and MCI conditions mainly targeting A β . More recent trials targeting tau protein and neuro-inflammation are promising but still ongoing. Complementing the traditional long-term expensive drug development process from the preclinical to clinical trial phases with other approaches, such as drug repurposing and drug repositioning, may be of great help not only to prioritize therapeutic agents for new indications potentially beneficial for patients but also to enlarge the spectrum of applications of drugs and eventually optimize the whole development process as well, with a substantial benefit for pharmaceutical industries [156].

One first example of this approach is represented by two approved drugs, namely camprostate and baclofen, that have been repurposed as potential disease-modifying com-

pounds for Parkinson's and AD, respectively, and more recently for multiple sclerosis in disease models. As for the AD, based on the fact that camprostate modulates glutamatergic transmission and E/I imbalance, while baclofen activates metabotropic GABA_B receptors, the authors show that their synergistic combination, now called PXT864, which targets glutamatergic, GABAergic, and glycinergic systems, protects against A β neurotoxicity and alleviates cognitive deficits in a transgenic mouse model of AD [157,158]. A further example is bumetanide, the FDA-approved sulfamyl diuretic, a drug broadly used to treat high blood pressure but also edema associated with heart, liver, or kidney failure. By exerting an inhibitory action on the chloride importer NKCC1, bumetanide is able to restore the inhibitory action of GABA [159]. Bumetanide has been shown to exert positive effects in several neurodevelopmental disorders such as autism, schizophrenia, *fragile X* syndrome, and epilepsy [5,58]. Interestingly, this drug was able to improve cognitive impairments in animal models of down syndrome, a condition known to be associated to an E/I imbalance and related to AD dementia [160].

An impressive study has been recently published on the repurposing of bumetanide for late-onset AD [161]. The scientists' team, led by Yadong Huang and Marina Sirota at UCSF, extracted the gene expression signature of APOE4 carriers with AD and compared them to non-demented carriers using a temporal lobe transcriptomic dataset. The signature was then compared to the cMap database to find drugs able to normalize the AD APOE4/APOE4 carriers' transcriptome. To this aim, they ranked the drugs based on their capacity to potentially reverse or flip the AD transcriptomic signature back to control baseline. The significance of fold-change gene shift was computed by a Kolmogorov–Smirnov statistics. Though cMap may not appear a suitable resource, as it is built on cancer cell lines, the strategy was successful since bumetanide ranked 4th out of 1300 compounds as the most effective flipping drug, and it was selected as the top candidate for further investigations because of its safety in long-term use and its positive effects on other brain diseases. The therapeutic potential of bumetanide for AD treatment was validated both in vitro and in vivo and statistically with real-world data. The in vitro validation in iPSC-derived neuronal cells from APOE4/APOE4 subjects confirmed the transcriptomic signatures predicted in silico by the cMap database both for gene expression and pathway analysis. Impressive in vivo validation results were obtained from APOE4/APOE4 knock-in mice and in the same animal models crossed with the J20 mice, which develop amyloid. Following bumetanide treatment, single-cell RNA-Seq showed a reversed transcriptome in many brain cell types together with a reduced amyloid burden, an electrophysiological reversal of LTP, and a recover in spatial memory deficits. Eventually the authors explored the protecting role of bumetanide for AD in a retrospective study on real-world data using electronic health records from two different and independent databases from UCSF University and Mount Sinai NY. The data showed a striking statistically significant lower prevalence of AD in people aged >65 exposed to this drug for indications different from AD: 35% lower in UCSF cohort and even 75% lower in Mount Sinai cohort as compared to controls. In agreement with previously reported data on AD11 mice [153], these results strongly suggest that the shift of GABA action from the hyperpolarizing to the depolarizing direction, consequent to a downregulation of the cation-chloride exporter, KCC2, is responsible for the E/I imbalance observed in AD patients, an effect that could be rescued by blocking the cation-chloride importer NKCC1 with bumetanide.

It is worth noting that, in agreement with [161] in rodent models of AD, it has been recently reported that memory deficiency, consequent to impaired proBDNF/BDNF conversion, which leads to a reduced GABA_A-mediated neurotransmission in the CA1 hippocampal region associated with reduced KCC2 expression and positive shift of the equilibrium potential for Cl⁻ (E_{Cl^-}), can be rescued by the intracerebroventricular administration of bumetanide, a selective antagonist of the cation-chloride importer NKCC1 [162].

6. Conclusions and Future Directions

Cation-chloride co-transporters are key regulators of intracellular Cl^- concentration. Their alterations account for several neurodevelopmental and neurodegenerative disorders, including AD. In particular, the shift of GABA from the hyperpolarizing to the depolarizing direction, due to impairment of the cation-chloride KCC2 exporter, is responsible for behavioral deficits observed in the adult AD11 mouse, an animal model of AD, in which neutralization of NGF with selective anti-NGF antibodies leads to a neurodegenerative pathology similar to that observed in AD patients [153]. In recent data obtained by analyzing in two electronic health record databases, a large cohort of patients affected by different neuro-pathologies clearly showed that treatment with bumetanide, a selective antagonist of the chloride importer NKCC1, prevents the development of APOE4-related forms of AD in individuals over the age of 65 years [161]. This suggests that AD is usually associated with a depolarizing action of GABA, probably related to a reduced expression of KCC2, which could be rescued by reducing the expression of NKCC1 with bumetanide. This opens new avenues for developing drugs that, by regulating chloride homeostasis, improve cognitive deficits associated with this devastating disorder. However, in spite of encouraging perspectives, several points need further investigation. It is unclear why NGF, primarily involved in growth, maintenance, proliferation, and survival of peripheral and central neurons during development and throughout adulthood, affects chloride homeostasis. While the cholinergic hypothesis of AD relies on the fact that NGF is involved in the maintenance and survival of these basal forebrain cholinergic neurons, the primary target of the disease, how this neurotrophin controls cation-chloride co-transporters and the direction of GABAergic signaling remains to be established.

Bumetanide has been shown to have positive effects in a wide range of pathological conditions, but its exact mechanisms of its action are still poorly understood. Although we cannot exclude the possibility that in AD patients, the blood–brain barrier (BBB) is compromised, the effectiveness of bumetanide is usually reduced by its poor pharmacokinetic properties and its low capability to cross the BBB to reach either neuronal or non-neuronal targets [163]. This limitation could be overcome with new derivatives currently under development, which are capable of better permeating the BBB [160]. Alternatively, new therapeutic tools selectively targeting the cation chloride exporter KCC2, which, in contrast to NKCC1, is exclusively expressed on central neurons [164], could be considered. By acting either on KCC2 membrane trafficking or on their intrinsic transport kinetics, these compounds will allow attenuating neuronal $[\text{Cl}^-]_i$, reinstating an appropriate Cl^- homeostasis in selective brain regions and a proper E/I balance. However, these compounds should not interfere with the well-known KCC2 structural function on dendritic spines, an effect that is independent on its intracellular chloride regulation [165].

Furthermore, in AD11 mice, gene expression experiments have unveiled a downregulation of mRNA encoding KCC2 (but not NKCC1) as well as several GABA_A receptor subunits. Therefore, as a matter of speculation, we can foresee the development of a gene therapy approach, whereby genes coding for antibodies delivered via viral vectors to particular brain regions, by selectively silencing the cation-chloride importer NKCC1, will be able to restore GABAergic function by promoting the shift of GABA from the depolarizing to the hyperpolarizing direction with consequent impairment of the E/I balance. This method has been successfully applied to develop two antibodies (mAb 12A12d e scFvA13) targeting the A β peptide and the toxic fragment of the tau protein with a high molecular and subcellular precision [166,167].

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References

1. Semyanov, A.; Walker, M.C.; Kullmann, D.M.; Silver, R.A. Tonically Active GABAA Receptors: Modulating Gain and Maintaining the Tone. *Trends Neurosci.* **2004**, *27*, 262–269. [[CrossRef](#)] [[PubMed](#)]
2. Farrant, M.; Nusser, Z. Variations on an Inhibitory Theme: Phasic and Tonic Activation of GABAA Receptors. *Nat. Rev. Neurosci.* **2005**, *6*, 215–229. [[CrossRef](#)] [[PubMed](#)]
3. Cellot, G.; Cherubini, E. Functional Role of Ambient GABA in Refining Neuronal Circuits Early in Postnatal Development. *Front. Neural Circuits* **2013**, *7*, 136. [[CrossRef](#)] [[PubMed](#)]
4. Cherubini, E.; Gaiarsa, J.L.; Ben-Ari, Y. GABA: An Excitatory Transmitter in Early Postnatal Life. *Trends Neurosci.* **1991**, *14*, 515–519. [[CrossRef](#)]
5. Ben-Ari, Y.; Khalilov, I.; Kahle, K.T.; Cherubini, E. The GABA Excitatory/Inhibitory Shift in Brain Maturation and Neurological Disorders. *Neuroscientist* **2012**, *18*, 467–486. [[CrossRef](#)]
6. Rivera, C.; Voipio, J.; Payne, J.A.; Ruusuvuori, E.; Lahtinen, H.; Lamsa, K.; Pirvola, U.; Saarma, M.; Kaila, K. The K⁺/Cl[−] Co-Transporter KCC2 Renders GABA Hyperpolarizing during Neuronal Maturation. *Nature* **1999**, *397*, 251–255. [[CrossRef](#)]
7. Palop, J.J.; Mucke, L. Amyloid- β -Induced Neuronal Dysfunction in Alzheimer’s Disease: From Synapses toward Neural Networks. *Nat. Neurosci.* **2010**, *13*, 812–818. [[CrossRef](#)]
8. Uhlhaas, P.J.; Singer, W. Neural Synchrony in Brain Disorders: Relevance for Cognitive Dysfunctions and Pathophysiology. *Neuron* **2006**, *52*, 155–168. [[CrossRef](#)]
9. Nichols, E.; Steinmetz, J.D.; Vollset, S.E.; Fukutaki, K.; Chalek, J.; Abd-Allah, F.; Abdoli, A.; Abualhasan, A.; Abu-Gharbieh, E.; Akram, T.T.; et al. Estimation of the Global Prevalence of Dementia in 2019 and Forecasted Prevalence in 2050: An Analysis for the Global Burden of Disease Study 2019. *Lancet Public Health* **2022**, *7*, e105–e125. [[CrossRef](#)]
10. Wang, R.; Reddy, P.H. Role of Glutamate and NMDA Receptors in Alzheimer’s Disease. *J. Alzheimer’s Dis.* **2017**, *57*, 1041–1048. [[CrossRef](#)]
11. Shankar, G.M.; Bloodgood, B.L.; Townsend, M.; Walsh, D.M.; Selkoe, D.J.; Sabatini, B.L. Natural Oligomers of the Alzheimer Amyloid- β Protein Induce Reversible Synapse Loss by Modulating an NMDA-Type Glutamate Receptor-Dependent Signaling Pathway. *J. Neurosci.* **2007**, *27*, 2866–2875. [[CrossRef](#)] [[PubMed](#)]
12. Mucke, L.; Selkoe, D.J. Neurotoxicity of Amyloid β -Protein: Synaptic and Network Dysfunction. *Cold Spring Harb. Perspect. Med.* **2012**, *2*, a006338. [[CrossRef](#)] [[PubMed](#)]
13. Rossor, M.N.; Emson, P.C.; Mountjoy, C.Q.; Roth, M.; Iversen, L.L. Neurotransmitters of the Cerebral Cortex in Senile Dementia of Alzheimer Type. In *The Aging Brain*; Hoyer, S., Ed.; Experimental Brain Research Supplementum; Springer: Berlin/Heidelberg, Germany, 1982; Volume 5, pp. 153–157. ISBN 978-3-642-68509-5.
14. Huang, Y.; Mucke, L. Alzheimer Mechanisms and Therapeutic Strategies. *Cell* **2012**, *148*, 1204–1222. [[CrossRef](#)] [[PubMed](#)]
15. Villette, V.; Dutar, P. GABAergic Microcircuits in Alzheimer’s Disease Models. *Curr. Alzheimer Res.* **2016**, *14*, 30–39. [[CrossRef](#)]
16. Selkoe, D.J. Early Network Dysfunction in Alzheimer’s Disease. *Science* **2019**, *365*, 540–541. [[CrossRef](#)]
17. Bakker, A.; Krauss, G.L.; Albert, M.S.; Speck, C.L.; Jones, L.R.; Stark, C.E.; Yassa, M.A.; Bassett, S.S.; Shelton, A.L.; Gallagher, M. Reduction of Hippocampal Hyperactivity Improves Cognition in Amnestic Mild Cognitive Impairment. *Neuron* **2012**, *74*, 467–474. [[CrossRef](#)]
18. Vossel, K.A.; Ranasinghe, K.G.; Beagle, A.J.; Mizuiru, D.; Honma, S.M.; Dowling, A.F.; Darwish, S.M.; Van Berlo, V.; Barnes, D.E.; Mantle, M.; et al. Incidence and Impact of Subclinical Epileptiform Activity in Alzheimer’s Disease: Subclinical Epileptiform Activity in AD. *Ann. Neurol.* **2016**, *80*, 858–870. [[CrossRef](#)]
19. Sanchez, P.E.; Zhu, L.; Verret, L.; Vossel, K.A.; Orr, A.G.; Cirrito, J.R.; Devidze, N.; Ho, K.; Yu, G.-Q.; Palop, J.J.; et al. Levetiracetam Suppresses Neuronal Network Dysfunction and Reverses Synaptic and Cognitive Deficits in an Alzheimer’s Disease Model. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, E2895–E2903. [[CrossRef](#)]
20. Vossel, K.A.; Tartaglia, M.C.; Nygaard, H.B.; Zeman, A.Z.; Miller, B.L. Epileptic Activity in Alzheimer’s Disease: Causes and Clinical Relevance. *Lancet Neurol.* **2017**, *16*, 311–322. [[CrossRef](#)]

21. Grouselle, D.; Winsky-Sommerer, R.; David, J.P.; Delacourte, A.; Dournaud, P.; Epelbaum, J. Loss of Somatostatin-like Immunoreactivity in the Frontal Cortex of Alzheimer Patients Carrying the Apolipoprotein Epsilon 4 Allele. *Neurosci. Lett.* **1998**, *255*, 21–24. [[CrossRef](#)]
22. Bai, X.; Edden, R.A.E.; Gao, F.; Wang, G.; Wu, L.; Zhao, B.; Wang, M.; Chan, Q.; Chen, W.; Barker, P.B. Decreased γ -Aminobutyric Acid Levels in the Parietal Region of Patients with Alzheimer's Disease: In Vivo GABA Levels Measurement in AD. *J. Magn. Reson. Imaging* **2015**, *41*, 1326–1331. [[CrossRef](#)] [[PubMed](#)]
23. Schwab, C.; Yu, S.; Wong, W.; McGeer, E.G.; McGeer, P.L. GAD65, GAD67, and GABAT Immunostaining in Human Brain and Apparent GAD65 Loss in Alzheimer's Disease. *J. Alzheimer's Dis.* **2013**, *33*, 1073–1088. [[CrossRef](#)] [[PubMed](#)]
24. Garcia-Marin, V. Diminished Perisomatic GABAergic Terminals on Cortical Neurons Adjacent to Amyloid Plaques. *Front. Neuroanat.* **2009**, *3*, 28. [[CrossRef](#)] [[PubMed](#)]
25. Ramos-Miguel, A.; Hercher, C.; Beasley, C.L.; Barr, A.M.; Bayer, T.A.; Falkai, P.; Leurgans, S.E.; Schneider, J.A.; Bennett, D.A.; Honer, W.G. Loss of Munc18-1 Long Splice Variant in GABAergic Terminals Is Associated with Cognitive Decline and Increased Risk of Dementia in a Community Sample. *Mol. Neurodegener.* **2015**, *10*, 65. [[CrossRef](#)] [[PubMed](#)]
26. Saiz-Sanchez, D.; Ubeda-Bañon, I.; Flores-Cuadrado, A.; Gonzalez-Rodriguez, M.; Villar-Conde, S.; Astillero-Lopez, V.; Martinez-Marcos, A. Somatostatin, Olfaction, and Neurodegeneration. *Front. Neurosci.* **2020**, *14*, 96. [[CrossRef](#)] [[PubMed](#)]
27. Tremblay, R.; Lee, S.; Rudy, B. GABAergic Interneurons in the Neocortex: From Cellular Properties to Circuits. *Neuron* **2016**, *91*, 260–292. [[CrossRef](#)]
28. Palop, J.J.; Jones, B.; Kekoni, L.; Chin, J.; Yu, G.-Q.; Raber, J.; Masliah, E.; Mucke, L. Neuronal Depletion of Calcium-Dependent Proteins in the Dentate Gyrus Is Tightly Linked to Alzheimer's Disease-Related Cognitive Deficits. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 9572–9577. [[CrossRef](#)]
29. Takahashi, H.; Brasnjevic, I.; Rutten, B.P.F.; Van Der Kolk, N.; Perl, D.P.; Bouras, C.; Steinbusch, H.W.M.; Schmitz, C.; Hof, P.R.; Dickstein, D.L. Hippocampal Interneuron Loss in an APP/PS1 Double Mutant Mouse and in Alzheimer's Disease. *Brain Struct Funct.* **2010**, *214*, 145–160. [[CrossRef](#)]
30. Zheng, J.; Li, H.-L.; Tian, N.; Liu, F.; Wang, L.; Yin, Y.; Yue, L.; Ma, L.; Wan, Y.; Wang, J.-Z. Interneuron Accumulation of Phosphorylated Tau Impairs Adult Hippocampal Neurogenesis by Suppressing GABAergic Transmission. *Cell Stem Cell* **2020**, *26*, 331–345. [[CrossRef](#)]
31. Soininen, H.; Riekkinen, P.J.; Partanen, J.; Helkala, E.-L.; Laulumaa, V.; Jolkonen, J.; Reinikainen, K. Cerebrospinal Fluid Somatostatin Correlates with Spectral EEG Variables and with Parietotemporal Cognitive Dysfunction in Alzheimer Patients. *Neurosci. Lett.* **1988**, *85*, 131–136. [[CrossRef](#)]
32. Calvo-Flores Guzmán, B.; Vinnakota, C.; Govindpani, K.; Waldvogel, H.J.; Faull, R.L.M.; Kwakowsky, A. The GABAergic System as a Therapeutic Target for Alzheimer's Disease. *J. Neurochem.* **2018**, *146*, 649–669. [[CrossRef](#)] [[PubMed](#)]
33. Kwakowsky, A.; Calvo-Flores Guzmán, B.; Pandya, M.; Turner, C.; Waldvogel, H.J.; Faull, R.L. GABA_A Receptor Subunit Expression Changes in the Human Alzheimer's Disease Hippocampus, Subiculum, Entorhinal Cortex and Superior Temporal Gyrus. *J. Neurochem.* **2018**, *145*, 374–392. [[CrossRef](#)] [[PubMed](#)]
34. Iwakiri, M.; Mizukami, K.; Ikonovic, M.D.; Ishikawa, M.; Abrahamson, E.E.; DeKosky, S.T.; Asada, T. An Immunohistochemical Study of GABA_A Receptor Gamma Subunits in Alzheimer's Disease Hippocampus: Relationship to Neurofibrillary Tangle Progression. *Neuropathology* **2009**, *29*, 263–269. [[CrossRef](#)]
35. Iwakiri, M.; Mizukami, K.; Ikonovic, M.D.; Ishikawa, M.; Hidaka, S.; Abrahamson, E.E.; DeKosky, S.T.; Asada, T. Changes in Hippocampal GABAB₁ Subunit Expression in Alzheimer's Patients: Association with Braak Staging. *Acta Neuropathol.* **2005**, *109*, 467–474. [[CrossRef](#)] [[PubMed](#)]
36. Limon, A.; Reyes-Ruiz, J.M.; Mileti, R. Loss of Functional GABA_A Receptors in the Alzheimer Diseased Brain. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, 10071–10076. [[CrossRef](#)] [[PubMed](#)]
37. Krantic, S.; Isorce, N.; Mechawar, N.; Davoli, M.A.; Vignault, E.; Albuquerque, M.; Chabot, J.-G.; Moysé, E.; Chauvin, J.-P.; Aubert, I.; et al. Hippocampal GABAergic Neurons Are Susceptible to Amyloid- β Toxicity in Vitro and Are Decreased in Number in the Alzheimer's Disease TgCRND8 Mouse Model. *J. Alzheimer's Dis.* **2012**, *29*, 293–308. [[CrossRef](#)] [[PubMed](#)]
38. Ramos, B.; Baglietto-Vargas, D.; del Rio, J.C.; Moreno-Gonzalez, I.; Santa-Maria, C.; Jimenez, S.; Caballero, C.; Lopez-Tellez, J.F.; Khan, Z.U.; Ruano, D.; et al. Early Neuropathology of Somatostatin/NPY GABAergic Cells in the Hippocampus of a PS1 \times APP Transgenic Model of Alzheimer's Disease. *Neurobiol. Aging* **2006**, *27*, 1658–1672. [[CrossRef](#)]
39. Oyelami, T.; Bondt, A.D.; den Wyngaert, I.V.; Van Hoorde, K.; Hoskens, L.; Shaban, H.; Kemp, J.A.; Drinkenburg, W.H. Age-dependent Concomitant Changes in Synaptic Function and GABAergic Pathway in the APP/PS1 Mouse Model. *Acta Neurobiol. Exp.* **2016**, *76*, 282–293. [[CrossRef](#)]
40. Moreno-Gonzalez, I.; Baglietto-Vargas, D.; Sanchez-Varo, R.; Jimenez, S.; Trujillo-Estrada, L.; Sanchez-Mejias, E.; del Rio, J.C.; Torres, M.; Romero-Acebal, M.; Ruano, D.; et al. Extracellular Amyloid- β and Cytotoxic Glial Activation Induce Significant Entorhinal Neuron Loss in Young PS1M146L/APP751SL Mice. *J. Alzheimer's Dis.* **2009**, *18*, 755–776. [[CrossRef](#)]
41. Li, G.; Bien-Ly, N.; Andrews-Zwilling, Y.; Xu, Q.; Bernardo, A.; Ring, K.; Halabisky, B.; Deng, C.; Mahley, R.W.; Huang, Y. GABAergic Interneuron Dysfunction Impairs Hippocampal Neurogenesis in Adult Apolipoprotein E4 Knockin Mice. *Cell Stem Cell* **2009**, *5*, 634–645. [[CrossRef](#)]

42. Leung, L.; Andrews-Zwilling, Y.; Yoon, S.Y.; Jain, S.; Ring, K.; Dai, J.; Wang, M.M.; Tong, L.; Walker, D.; Huang, Y. Apolipoprotein E4 Causes Age- and Sex-Dependent Impairments of Hilar GABAergic Interneurons and Learning and Memory Deficits in Mice. *PLoS ONE* **2012**, *7*, e53569. [[CrossRef](#)] [[PubMed](#)]
43. Tong, L.M.; Djukic, B.; Arnold, C.; Gillespie, A.K.; Yoon, S.Y.; Wang, M.M.; Zhang, O.; Knoferle, J.; Rubenstein, J.L.R.; Alvarez-Buylla, A.; et al. Inhibitory Interneuron Progenitor Transplantation Restores Normal Learning and Memory in ApoE4 Knock-In Mice without or with A β Accumulation. *J. Neurosci.* **2014**, *34*, 9506–9515. [[CrossRef](#)] [[PubMed](#)]
44. Hu, H.; Gan, J.; Jonas, P. Fast-Spiking, Parvalbumin⁺ GABAergic Interneurons: From Cellular Design to Microcircuit Function. *Science* **2014**, *345*, 1255263. [[CrossRef](#)] [[PubMed](#)]
45. Zallo, F.; Gardenal, E.; Verkhratsky, A.; Rodríguez, J.J. Loss of Calretinin and Parvalbumin Positive Interneurons in the Hippocampal CA1 of Aged Alzheimer's Disease Mice. *Neurosci. Lett.* **2018**, *681*, 19–25. [[CrossRef](#)]
46. Czapski, G.A.; Strosznajder, J.B. Glutamate and GABA in Microglia-Neuron Cross-Talk in Alzheimer's Disease. *Int. J. Mol. Sci.* **2021**, *22*, 1677. [[CrossRef](#)]
47. Kang, J.-Q. Epileptic Mechanisms Shared by Alzheimer's Disease: Viewed via the Unique Lens of Genetic Epilepsy. *Int. J. Mol. Sci.* **2021**, *22*, 7133. [[CrossRef](#)] [[PubMed](#)]
48. Iaccarino, H.F.; Singer, A.C.; Martorell, A.J.; Rudenko, A.; Gao, F.; Gillingham, T.Z.; Mathys, H.; Seo, J.; Kritskiy, O.; Abdurrob, F.; et al. Gamma Frequency Entrainment Attenuates Amyloid Load and Modifies Microglia. *Nature* **2016**, *540*, 230–235. [[CrossRef](#)]
49. Calvo-Flores Guzmán, B.; Kim, S.; Chawdhary, B.; Peppercorn, K.; Tate, W.P.; Waldvogel, H.J.; Faull, R.L.; Montgomery, J.; Kwakowsky, A. Amyloid-Beta1-42 -Induced Increase in GABAergic Tonic Conductance in Mouse Hippocampal CA1 Pyramidal Cells. *Molecules* **2020**, *25*, 693. [[CrossRef](#)]
50. Jo, S.; Yarishkin, O.; Hwang, Y.J.; Chun, Y.E.; Park, M.; Woo, D.H.; Bae, J.Y.; Kim, T.; Lee, J.; Chun, H.; et al. GABA from Reactive Astrocytes Impairs Memory in Mouse Models of Alzheimer's Disease. *Nat. Med.* **2014**, *20*, 886–896. [[CrossRef](#)]
51. Chu, D.C.M.; Penney, J.B.; Young, A.B. Cortical GABAB and GABAA Receptors in Alzheimer's Disease: A Quantitative Autoradiographic Study. *Neurology* **1987**, *37*, 1454. [[CrossRef](#)]
52. Salazar, A.M.; Leisgang, A.M.; Ortiz, A.A.; Murtishaw, A.S.; Kinney, J.W. Alterations of GABA B Receptors in the APP/PS1 Mouse Model of Alzheimer's Disease. *Neurobiol. Aging* **2021**, *97*, 129–143. [[CrossRef](#)] [[PubMed](#)]
53. Martín-Belmonte, A.; Aguado, C.; Alfaro-Ruiz, R.; Moreno-Martínez, A.E.; de la Ossa, L.; Martínez-Hernández, J.; Buisson, A.; Shigemoto, R.; Fukazawa, Y.; Luján, R. Density of GABAB Receptors Is Reduced in Granule Cells of the Hippocampus in a Mouse Model of Alzheimer's Disease. *Int. J. Mol. Sci.* **2020**, *21*, 2459. [[CrossRef](#)] [[PubMed](#)]
54. Dinamarca, M.C.; Raveh, A.; Schneider, A.; Fritzius, T.; Früh, S.; Rem, P.D.; Stawarski, M.; Lalanne, T.; Turecek, R.; Choo, M.; et al. Complex Formation of APP with GABAB Receptors Links Axonal Trafficking to Amyloidogenic Processing. *Nat. Commun.* **2019**, *10*, 1331. [[CrossRef](#)] [[PubMed](#)]
55. Wright, R.; Newey, S.E.; Ilie, A.; Wefelmeyer, W.; Raimondo, J.V.; Ginham, R.; McIlhinney, R.A.J.; Akerman, C.J. Neuronal Chloride Regulation via KCC2 Is Modulated through a GABA β Receptor Protein Complex. *J. Neurosci.* **2017**, *37*, 5447–5462. [[CrossRef](#)]
56. Doshina, A.; Gourgue, F.; Onizuka, M.; Opsomer, R.; Wang, P.; Ando, K.; Tasiaux, B.; Dewachter, I.; Kienlen-Campard, P.; Brion, J.-P.; et al. Cortical Cells Reveal APP as a New Player in the Regulation of GABAergic Neurotransmission. *Sci. Rep.* **2017**, *7*, 370. [[CrossRef](#)] [[PubMed](#)]
57. Chen, M.; Wang, J.; Jiang, J.; Zheng, X.; Justice, N.J.; Wang, K.; Ran, X.; Li, Y.; Huo, Q.; Zhang, J.; et al. APP Modulates KCC2 Expression and Function in Hippocampal GABAergic Inhibition. *eLife* **2017**, *6*, e20142. [[CrossRef](#)] [[PubMed](#)]
58. Cherubini, E.; Di Cristo, G.; Avoli, M. Dysregulation of GABAergic Signaling in Neurodevelopmental Disorders: Targeting Cation-Chloride Co-Transporters to Re-Establish a Proper E/I Balance. *Front. Cell. Neurosci.* **2022**, *15*, 813441. [[CrossRef](#)]
59. Fiumelli, H.; Woodin, M.A. Role of Activity-Dependent Regulation of Neuronal Chloride Homeostasis in Development. *Curr. Opin. Neurobiol.* **2007**, *17*, 81–86. [[CrossRef](#)]
60. Capsoni, S.; Ugolini, G.; Comparini, A.; Ruberti, F.; Berardi, N.; Cattaneo, A. Alzheimer-like Neurodegeneration in Aged Antinerve Growth Factor Transgenic Mice. *Proc. Natl. Acad. Sci. USA* **2000**, *97*, 6826–6831. [[CrossRef](#)]
61. Whitehouse, P.; Price, D.; Struble, R.; Clark, A.; Coyle, J.; DeLong, M.R. Alzheimer's Disease and Senile Dementia: Loss of Neurons in the Basal Forebrain. *Science* **1982**, *215*, 1237–1239. [[CrossRef](#)]
62. Bartus, R.T.; Dean, R.L.; Pontecorvo, M.J.; Flicker, C. The Cholinergic Hypothesis: A Historical Overview, Current Perspective, and Future Directions. *Ann. N. Y. Acad. Sci.* **1985**, *444*, 332–358. [[CrossRef](#)] [[PubMed](#)]
63. Giacobini, E.; Pepeu, G. Sex and Gender Differences in the Brain Cholinergic System and in the Response to Therapy of Alzheimer Disease with Cholinesterase Inhibitors. *Curr. Alzheimer Res.* **2018**, *15*, 1077–1084. [[CrossRef](#)] [[PubMed](#)]
64. Davies, P.; Maloney, A.J.F. Selective Loss of Central Cholinergic Neurons in Alzheimer's Disease. *Lancet* **1976**, *308*, 1403. [[CrossRef](#)]
65. DeKosky, D.S.T.; Harbaugh, R.E.; Schmitt, F.A.; Bakay, R.A.; Chui, H.C.; Knopman, D.S.; Reeder, T.M.; Shetter, A.G.; Senter, H.J.; Markesbery, W.R. Cortical Biopsy in Alzheimer's Disease: Diagnostic Accuracy and Neurochemical, Neuropathological, and Cognitive Correlations. *Ann. Neurol.* **1992**, *32*, 625–632. [[CrossRef](#)]
66. Bartus, R.T.; Dean, R.L.; Beer, B.; Lippa, A.S. The Cholinergic Hypothesis of Geriatric Memory Dysfunction. *Science* **1982**, *217*, 408–414. [[CrossRef](#)]
67. Coyle, J.T.; Price, D.L.; DeLong, M.R. Alzheimer's Disease: A Disorder of Cortical Cholinergic Innervation. *Science* **1983**, *219*, 1184–1190. [[CrossRef](#)]

68. Levi-Montalcini, R. Effects of Mouse Tumor Transplantation on the Nervous System. *Ann. N. Y. Acad. Sci.* **1952**, *55*, 330–344. [[CrossRef](#)]
69. Dreyfus, C.F. Effects of Nerve Growth Factor on Cholinergic Brain Neurons. *Trends Pharmacol. Sci.* **1989**, *10*, 145–149. [[CrossRef](#)]
70. Levi-Montalcini, R. The Nerve Growth Factor 35 Years Later. *Science* **1987**, *237*, 1154–1162. [[CrossRef](#)]
71. Lindsay, R.M.; Harmar, A.J. Nerve Growth Factor Regulates Expression of Neuropeptide Genes in Adult Sensory Neurons. *Nature* **1989**, *337*, 362–364. [[CrossRef](#)]
72. McAllister, A.K. Neurotrophins and Neuronal Differentiation in the Central Nervous System. *Cell Mol. Life Sci.* **2001**, *58*, 1054–1060. [[CrossRef](#)] [[PubMed](#)]
73. Mobley, W.C.; Rutkowski, J.L.; Tennekoon, G.I.; Gemski, J.; Buchanan, K.; Johnston, M.V. Nerve Growth Factor Increases Choline Acetyltransferase Activity in Developing Basal Forebrain Neurons. *Mol. Brain Res.* **1986**, *1*, 53–62. [[CrossRef](#)]
74. Hefti, F. Nerve Growth Factor Promotes Survival of Septal Cholinergic Neurons after Fimbrial Transections. *J. Neurosci.* **1986**, *6*, 2155–2162. [[CrossRef](#)] [[PubMed](#)]
75. Ayer-LeLievre, C.; Olson, L.; Ebendal, T.; Seiger, A.; Persson, H. Expression of the Beta-Nerve Growth Factor Gene in Hippocampal Neurons. *Science* **1988**, *240*, 1339–1341. [[CrossRef](#)]
76. Korsching, S.; Auburger, G.; Heumann, R.; Scott, J.; Thoenen, H. Levels of Nerve Growth Factor and Its mRNA in the Central Nervous System of the Rat Correlate with Cholinergic Innervation. *EMBO J.* **1985**, *4*, 1389–1393. [[CrossRef](#)]
77. Large, T.H.; Bodary, S.C.; Clegg, D.O.; Weskamp, G.; Otten, U.; Reichardt, L.F. Nerve Growth Factor Gene Expression in the Developing Rat Brain. *Science* **1986**, *234*, 352–355. [[CrossRef](#)]
78. Whittemore, S.R.; Ebendal, T.; Lärkfors, L.; Olson, L.; Seiger, A.; Strömberg, I.; Persson, H. Development and Regional Expression of Beta Nerve Growth Factor Messenger RNA and Protein in the Rat Central Nervous System. *Proc. Natl. Acad. Sci. USA* **1986**, *83*, 817–821. [[CrossRef](#)]
79. Seiler, M.; Schwab, M.E. Specific Retrograde Transport of Nerve Growth Factor (NGF) from Neocortex to Nucleus Basalis in the Rat. *Brain Res.* **1984**, *300*, 33–39. [[CrossRef](#)]
80. Fagan, A.M.; Garber, M.; Barbacid, M.; Silos-Santiago, I.; Holtzman, D.M. A Role for TrkA during Maturation of Striatal and Basal Forebrain Cholinergic Neurons In Vivo. *J. Neurosci.* **1997**, *17*, 7644–7654. [[CrossRef](#)]
81. Li, Y.; Holtzman, D.M.; Kromer, L.F.; Kaplan, D.R.; Chua-Couzens, J.; Clary, D.O.; Knusel, B.; Mobley, W.C. Regulation of TrkA and ChAT Expression in Developing Rat Basal Forebrain: Evidence That Both Exogenous and Endogenous NGF Regulate Differentiation of Cholinergic Neurons. *J. Neurosci.* **1995**, *15*, 2888–2905. [[CrossRef](#)]
82. Sobrieviela, T.; Clary, D.O.; Reichardt, L.F.; Brandabur, M.M.; Kordower, J.H.; Mufson, E.J. TrkA-immunoreactive Profiles in the Central Nervous System: Colocalization with Neurons Containing P75 Nerve Growth Factor Receptor, Choline Acetyltransferase, and Serotonin. *J. Comp. Neurol.* **1994**, *350*, 587–611. [[CrossRef](#)] [[PubMed](#)]
83. Gnahn, H.; Hefti, F.; Heumann, R.; Schwab, M.E.; Thoenen, H. NGF-Mediated Increase of Choline Acetyltransferase (ChAT) in the Neonatal Rat Forebrain: Evidence for a Physiological Role of NGF in the Brain? *Dev. Brain Res.* **1983**, *9*, 45–52. [[CrossRef](#)]
84. Williams, L.R.; Varon, S.; Peterson, G.M.; Wictorin, K.; Fischer, W.; Bjorklund, A.; Gage, F.H. Continuous Infusion of Nerve Growth Factor Prevents Basal Forebrain Neuronal Death after Fimbria Fornix Transection. *Proc. Natl. Acad. Sci. USA* **1986**, *83*, 9231–9235. [[CrossRef](#)] [[PubMed](#)]
85. Kromer, L.F. Nerve Growth Factor Treatment After Brain Injury Prevents Neuronal Death. *Science* **1987**, *235*, 214–216. [[CrossRef](#)]
86. Fischer, W.; Wictorin, K.; Björklund, A.; Williams, L.R.; Varon, S.; Gage, F.H. Amelioration of Cholinergic Neuron Atrophy and Spatial Memory Impairment in Aged Rats by Nerve Growth Factor. *Nature* **1987**, *329*, 65–68. [[CrossRef](#)]
87. Costantini, C.; Scrabble, H.; Puglielli, L. An Aging Pathway Controls the TrkA to P75NTR Receptor Switch and Amyloid Beta-Peptide Generation. *EMBO J.* **2006**, *25*, 1997–2006. [[CrossRef](#)]
88. Yaar, M.; Zhai, S.; Fine, R.E.; Eisenhauer, P.B.; Arble, B.L.; Stewart, K.B.; Gilchrest, B.A. Amyloid β Binds Trimers as Well as Monomers of the 75-KDa Neurotrophin Receptor and Activates Receptor Signaling. *J. Biol. Chem.* **2002**, *277*, 7720–7725. [[CrossRef](#)]
89. Frade, J.M. Nuclear Translocation of the P75 Neurotrophin Receptor Cytoplasmic Domain in Response to Neurotrophin Binding. *J. Neurosci.* **2005**, *25*, 1407–1411. [[CrossRef](#)]
90. Matrone, C.; Ciotti, M.T.; Mercanti, D.; Marolda, R.; Calissano, P. NGF and BDNF Signaling Control Amyloidogenic Route and A β Production in Hippocampal Neurons. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 13139–13144. [[CrossRef](#)]
91. Nuydens, R.; Dispersyn, G.; de Jong, M.; Van Den Kieboom, G.; Borgers, M.; Geerts, H. Aberrant Tau Phosphorylation and Neurite Retraction during NGF Deprivation in PC12 Cells. *Biochem. Biophys. Res. Commun.* **1997**, *240*, 687–691. [[CrossRef](#)]
92. Davis, P.K.; Johnson, G.V.W. The Microtubule Binding of Tau and High Molecular Weight Tau in Apoptotic PC12 Cells Is Impaired Because of Altered Phosphorylation. *J. Biol. Chem.* **1999**, *274*, 35686–35692. [[CrossRef](#)] [[PubMed](#)]
93. Shelton, S.B.; Johnson, G.V.W. Tau and HMW Tau Phosphorylation and Compartmentalization in Apoptotic Neuronal PC12 Cells. *J. Neurosci. Res.* **2001**, *66*, 203–213. [[CrossRef](#)] [[PubMed](#)]
94. Amadoro, G.; Corsetti, V.; Ciotti, M.T.; Florenzano, F.; Capsoni, S.; Amato, G.; Calissano, P. Endogenous A β Causes Cell Death via Early Tau Hyperphosphorylation. *Neurobiol. Aging* **2011**, *32*, 969–990. [[CrossRef](#)]
95. Matrone, C.; Di Luzio, A.; Meli, G.; D’Aguanno, S.; Severini, C.; Ciotti, M.T.; Cattaneo, A.; Calissano, P. Activation of the Amyloidogenic Route by NGF Deprivation Induces Apoptotic Death in PC12 Cells. *J. Alzheimer’s Dis.* **2008**, *13*, 81–96. [[CrossRef](#)] [[PubMed](#)]

96. Cattaneo, A.; Rapposelli, B.; Calissano, P. Three Distinct Types of Monoclonal Antibodies After Long-Term Immunization of Rats with Mouse Nerve Growth Factor. *J. Neurochem.* **1988**, *50*, 1003–1010. [[CrossRef](#)] [[PubMed](#)]
97. Molnar, M.; Tongiorgi, E.; Avignone, E.; Gonfloni, S.; Ruberti, F.; Domenici, L.; Cattaneo, A. The Effects of Anti-Nerve Growth Factor Monoclonal Antibodies on Developing Basal Forebrain Neurons Are Transient and Reversible: Transient Effects of Early Postnatal NGF Neutralization. *Eur. J. Neurosci.* **1998**, *10*, 3127–3140. [[CrossRef](#)] [[PubMed](#)]
98. Ruberti, F.; Capsoni, S.; Comparini, A.; Di Daniel, E.; Franzot, J.; Gonfloni, S.; Rossi, G.; Berardi, N.; Cattaneo, A. Phenotypic Knockout of Nerve Growth Factor in Adult Transgenic Mice Reveals Severe Deficits in Basal Forebrain Cholinergic Neurons, Cell Death in the Spleen, and Skeletal Muscle Dystrophy. *J. Neurosci.* **2000**, *20*, 2589–2601. [[CrossRef](#)] [[PubMed](#)]
99. De Rosa, R.; Garcia, A.A.; Braschi, C.; Capsoni, S.; Maffei, L.; Berardi, N.; Cattaneo, A. Intranasal Administration of Nerve Growth Factor (NGF) Rescues Recognition Memory Deficits in AD11 Anti-NGF Transgenic Mice. *Proc. Natl. Acad. Sci. USA* **2005**, *102*, 3811–3816. [[CrossRef](#)]
100. Capsoni, S.; Brandi, R.; Arisi, I.; D’Onofrio, M.; Cattaneo, A. A Dual Mechanism Linking NGF/ProNGF Imbalance and Early Inflammation to Alzheimer’s Disease Neurodegeneration in the AD11 Anti-NGF Mouse Model. *CNS Neurol. Disord.-Drug Targets* **2011**, *10*, 635–647. [[CrossRef](#)]
101. Capsoni, S.; Giannotta, S.; Cattaneo, A. β -Amyloid Plaques in a Model for Sporadic Alzheimer’s Disease Based on Transgenic Anti-Nerve Growth Factor Antibodies. *Mol. Cell. Neurosci.* **2002**, *21*, 15–28. [[CrossRef](#)]
102. Pesavento, E.; Capsoni, S.; Domenici, L.; Cattaneo, A. Acute Cholinergic Rescue of Synaptic Plasticity in the Neurodegenerating Cortex of Anti-Nerve-Growth-Factor Mice: Decreased Synaptic Plasticity in Anti-NGF Mice. *Eur. J. Neurosci.* **2002**, *15*, 1030–1036. [[CrossRef](#)] [[PubMed](#)]
103. Origlia, N.; Capsoni, S.; Domenici, L.; Cattaneo, A. Time Window in Cholinomimetic Ability to Rescue Long-Term Potentiation in Neurodegenerating Anti-Nerve Growth Factor Mice. *J. Alzheimer’s Dis.* **2006**, *9*, 59–68. [[CrossRef](#)] [[PubMed](#)]
104. Houeland, G.; Romani, A.; Marchetti, C.; Amato, G.; Capsoni, S.; Cattaneo, A.; Marie, H. Transgenic Mice with Chronic NGF Deprivation and Alzheimer’s Disease-Like Pathology Display Hippocampal Region-Specific Impairments in Short- and Long-Term Plasticities. *J. Neurosci.* **2010**, *30*, 13089–13094. [[CrossRef](#)] [[PubMed](#)]
105. Mufson, E.J.; Counts, S.E.; Ginsberg, S.D.; Mahady, L.; Perez, S.E.; Massa, S.M.; Longo, F.M.; Ikonovic, M.D. Nerve Growth Factor Pathobiology during the Progression of Alzheimer’s Disease. *Front. Neurosci.* **2019**, *13*, 533. [[CrossRef](#)] [[PubMed](#)]
106. Selkoe, D.J. The Origins of Alzheimer Disease: A Is for Amyloid. *JAMA* **2000**, *283*, 1615–1617. [[CrossRef](#)]
107. Capsoni, S.; Cattaneo, A. On the Molecular Basis Linking Nerve Growth Factor (NGF) to Alzheimer’s Disease. *Cell. Mol. Neurobiol.* **2006**, *26*, 617–631. [[CrossRef](#)]
108. Cattaneo, A.; Capsoni, S.; Paoletti, F. Towards Non Invasive Nerve Growth Factor Therapies for Alzheimer’s Disease. *J. Alzheimer’s Dis.* **2008**, *15*, 255–283. [[CrossRef](#)]
109. D’Onofrio, M.; Arisi, I.; Brandi, R.; Di Mambro, A.; Felsani, A.; Capsoni, S.; Cattaneo, A. Early Inflammation and Immune Response MRNAs in the Brain of AD11 Anti-NGF Mice. *Neurobiol. Aging* **2011**, *32*, 1007–1022. [[CrossRef](#)]
110. De Simone, R.; Ambrosini, E.; Carnevale, D.; Ajmone-Cat, M.A.; Minghetti, L. NGF Promotes Microglial Migration through the Activation of Its High Affinity Receptor: Modulation by TGF- β . *J. Neuroimmunol.* **2007**, *190*, 53–60. [[CrossRef](#)]
111. Rizzi, C.; Tiberi, A.; Giustizieri, M.; Marrone, M.C.; Gobbo, F.; Carucci, N.M.; Meli, G.; Arisi, I.; D’Onofrio, M.; Marinelli, S.; et al. NGF Steers Microglia toward a Neuroprotective Phenotype. *Glia* **2018**, *66*, 1395–1416. [[CrossRef](#)]
112. Paoletti, F.; Covaceuszach, S.; Konarev, P.V.; Gonfloni, S.; Malerba, F.; Schwarz, E.; Svergun, D.I.; Cattaneo, A.; Lamba, D. Intrinsic Structural Disorder of Mouse ProNGF. *Proteins Struct. Funct. Bioinform.* **2009**, *75*, 990–1009. [[CrossRef](#)] [[PubMed](#)]
113. Tiveron, C.; Fasulo, L.; Capsoni, S.; Malerba, F.; Marinelli, S.; Paoletti, F.; Piccinin, S.; Scardigli, R.; Amato, G.; Brandi, R. ProNGF\NGF Imbalance Triggers Learning and Memory Deficits, Neurodegeneration and Spontaneous Epileptic-like Discharges in Transgenic Mice. *Cell Death Differ.* **2013**, *20*, 1017–1030. [[CrossRef](#)] [[PubMed](#)]
114. Capsoni, S.; Tiveron, C.; Vignone, D.; Amato, G.; Cattaneo, A. Dissecting the Involvement of Tropomyosin-Related Kinase A and P75 Neurotrophin Receptor Signaling in NGF Deficit-Induced Neurodegeneration. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 12299–12304. [[CrossRef](#)] [[PubMed](#)]
115. Cuello, A.C.; Pentz, R.; Hall, H. The Brain NGF Metabolic Pathway in Health and in Alzheimer’s Pathology. *Front. Neurosci.* **2019**, *13*, 62. [[CrossRef](#)] [[PubMed](#)]
116. Fahnestock, M.; Shekari, A. ProNGF and Neurodegeneration in Alzheimer’s Disease. *Front. Neurosci.* **2019**, *13*, 129. [[CrossRef](#)]
117. Pentz, R.; Iulita, M.F.; Ducatenzeiler, A.; Bennett, D.A.; Cuello, A.C. The Human Brain NGF Metabolic Pathway Is Impaired in the Pre-Clinical and Clinical Continuum of Alzheimers Disease. *Mol. Psychiatry* **2020**, *26*, 6023–6037. [[CrossRef](#)]
118. Fahnestock, M.; Michalski, B.; Xu, B.; Coughlin, M.D. The Precursor Pro-Nerve Growth Factor Is the Predominant Form of Nerve Growth Factor in Brain and Is Increased in Alzheimer’s Disease. *Mol. Cell. Neurosci.* **2001**, *18*, 210–220. [[CrossRef](#)]
119. Fahnestock, M.; Scott, S.A.; Jetté, N.; Weingartner, J.A.; Crutcher, K.A. Nerve Growth Factor mRNA and Protein Levels Measured in the Same Tissue from Normal and Alzheimer’s Disease Parietal Cortex. *Mol. Brain Res.* **1996**, *42*, 175–178. [[CrossRef](#)]
120. Mufson, E.J.; Conner, J.M.; Kordower, J.H. Nerve Growth Factor in Alzheimer’s Disease: Defective Retrograde Transport to Nucleus Basalis. *Neuroreport* **1995**, *6*, 1063–1066. [[CrossRef](#)]
121. Shooter, E.M. Early Days of the Nerve Growth Factor Proteins. *Annu. Rev. Neurosci.* **2001**, *24*, 601–629. [[CrossRef](#)]
122. Fahnestock, M.; Yu, G.; Coughlin, M.D. ProNGF: A Neurotrophic or an Apoptotic Molecule? *Prog. Brain Res.* **2004**, *146*, 101–110. [[PubMed](#)]

123. Lee, R.; Kermani, P.; Teng, K.K.; Hempstead, B.L. Regulation of Cell Survival by Secreted Proneurotrophins. *Science* **2001**, *294*, 1945–1948. [[CrossRef](#)] [[PubMed](#)]
124. Mouri, A.; Nomoto, H.; Furukawa, S. Processing of Nerve Growth Factor: The Role of Basic Amino Acid Clusters in the pro-Region. *Biochem. Biophys. Res. Commun.* **2007**, *353*, 1056–1062. [[CrossRef](#)] [[PubMed](#)]
125. Bruno, M.A.; Cuello, A.C. Activity-Dependent Release of Precursor Nerve Growth Factor, Conversion to Mature Nerve Growth Factor, and Its Degradation by a Protease Cascade. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 6735–6740. [[CrossRef](#)] [[PubMed](#)]
126. Seidah, N.G.; Benjannet, S.; Pareek, S.; Savaria, D.; Hamelin, J.; Goulet, B.; Laliberté, J.; Lazure, C.; Chrétien, M.; Murphy, R.A. Cellular Processing of the Nerve Growth Factor Precursor by the Mammalian Pro-Protein Convertases. *Biochem. J.* **1996**, *314*, 951–960. [[CrossRef](#)] [[PubMed](#)]
127. Suter, U.; Heymach, J.V., Jr.; Shooter, E.M. Two Conserved Domains in the NGF Propeptide Are Necessary and Sufficient for the Biosynthesis of Correctly Processed and Biologically Active NGF. *EMBO J.* **1991**, *10*, 2395–2400. [[CrossRef](#)] [[PubMed](#)]
128. Arisi, I.; Cattaneo, A.; Rosato, V. Parameter Estimate of Signal Transduction Pathways. *BMC Neurosci.* **2006**, *7*, S6. [[CrossRef](#)]
129. Hempstead, B.L. Dissecting the Diverse Actions of Pro- and Mature Neurotrophins. *Curr. Alzheimer Res.* **2006**, *3*, 19–24. [[CrossRef](#)]
130. Twiss, J.L.; Chang, J.H.; Schanen, N.C. Pathophysiological Mechanisms for Actions of the Neurotrophins. *Brain Pathol.* **2006**, *16*, 320–332. [[CrossRef](#)]
131. Huang, E.J.; Reichardt, L.F. Neurotrophins: Roles in Neuronal Development and Function. *Annu. Rev. Neurosci.* **2001**, *24*, 677–736. [[CrossRef](#)]
132. Pezet, S.; McMahon, S.B. Neurotrophins: Mediators and Modulators of Pain. *Annu. Rev. Neurosci.* **2006**, *29*, 507–538. [[CrossRef](#)] [[PubMed](#)]
133. Nykjaer, A.; Lee, R.; Teng, K.K.; Jansen, P.; Madsen, P.; Nielsen, M.S.; Jacobsen, C.; Kliemann, M.; Schwarz, E.; Willnow, T.E. Sortilin Is Essential for ProNGF-Induced Neuronal Cell Death. *Nature* **2004**, *427*, 843–848. [[CrossRef](#)] [[PubMed](#)]
134. Schweigreiter, R. The Dual Nature of Neurotrophins. *Bioessays* **2006**, *28*, 583–594. [[CrossRef](#)] [[PubMed](#)]
135. Masoudi, R.; Ioannou, M.S.; Coughlin, M.D.; Pagadala, P.; Neet, K.E.; Clewes, O.; Allen, S.J.; Dawbarn, D.; Fahnestock, M. Biological Activity of Nerve Growth Factor Precursor Is Dependent upon Relative Levels of Its Receptors. *J. Biol. Chem.* **2009**, *284*, 18424–18433. [[CrossRef](#)]
136. Coulson, E.J.; Reid, K.; Baca, M.; Shipham, K.A.; Hulett, S.M.; Kilpatrick, T.J.; Bartlett, P.F. Chopper, a New Death Domain of the P75 Neurotrophin Receptor That Mediates Rapid Neuronal Cell Death. *J. Biol. Chem.* **2000**, *275*, 30537–30545. [[CrossRef](#)]
137. Huang, E.J.; Reichardt, L.F. Trk Receptors: Roles in Neuronal Signal Transduction. *Annu. Rev. Biochem.* **2003**, *72*, 609–642. [[CrossRef](#)]
138. Yoon, S.O.; Casaccia-Bonnel, P.; Carter, B.; Chao, M.V. Competitive Signaling between TrkA and P75 Nerve Growth Factor Receptors Determines Cell Survival. *J. Neurosci.* **1998**, *18*, 3273–3281. [[CrossRef](#)]
139. Berg, M.M.; Sternberg, D.W.; Hempstead, B.L.; Chao, M.V. The Low-Affinity P75 Nerve Growth Factor (NGF) Receptor Mediates NGF-Induced Tyrosine Phosphorylation. *Proc. Natl. Acad. Sci. USA* **1991**, *88*, 7106–7110. [[CrossRef](#)]
140. Hempstead, B.L.; Martin-Zanca, D.; Kaplan, D.R.; Parada, L.F.; Chao, M.V. High-Affinity NGF Binding Requires Coexpression of the Trk Proto-Oncogene and the Low-Affinity NGF Receptor. *Nature* **1991**, *350*, 678–683. [[CrossRef](#)]
141. Landreth, G.E.; Shooter, E.M. Nerve Growth Factor Receptors on PC12 Cells: Ligand-Induced Conversion from Low-to High-Affinity States. *Proc. Natl. Acad. Sci. USA* **1980**, *77*, 4751–4755. [[CrossRef](#)]
142. Gargano, N.; Levi, A.; Alema, S. Modulation of Nerve Growth Factor Internalization by Direct Interaction between P75 and TrkA Receptors. *J. Neurosci. Res.* **1997**, *50*, 1–12. [[CrossRef](#)]
143. Mahadeo, D.; Kaplan, L.; Chao, M.V.; Hempstead, B.L. High Affinity Nerve Growth Factor Binding Displays a Faster Rate of Association than P140trk Binding. Implications for Multi-Subunit Polypeptide Receptors. *J. Biol. Chem.* **1994**, *269*, 6884–6891. [[CrossRef](#)]
144. Conroy, J.N.; Coulson, E.J. High-Affinity TrkA and P75 Neurotrophin Receptor Complexes: A Twisted Affair. *J. Biol. Chem.* **2022**, *298*, 101568. [[CrossRef](#)] [[PubMed](#)]
145. Goedert, M.; Fine, A.; Hunt, S.P.; Ullrich, A. Nerve Growth Factor mRNA in Peripheral and Central Rat Tissues and in the Human Central Nervous System: Lesion Effects in the Rat Brain and Levels in Alzheimer's Disease. *Mol. Brain Res.* **1986**, *1*, 85–92. [[CrossRef](#)]
146. Jette, N.; Cole, M.S.; Fahnestock, M. NGF mRNA Is Not Decreased in Frontal Cortex from Alzheimer's Disease Patients. *Mol. Brain Res.* **1994**, *25*, 242–250. [[CrossRef](#)]
147. Pedraza, C.E.; Podlesniy, P.; Vidal, N.; Arévalo, J.C.; Lee, R.; Hempstead, B.; Ferrer, I.; Iglesias, M.; Espinet, C. Pro-NGF Isolated from the Human Brain Affected by Alzheimer's Disease Induces Neuronal Apoptosis Mediated by P75NTR. *Am. J. Pathol.* **2005**, *166*, 533–543. [[CrossRef](#)]
148. Pentz, R.; Iulita, M.F.; Ducatenzeiler, A.; Videla, L.; Benejam, B.; Carmona-Iragui, M.; Blesa, R.; Lleó, A.; Fortea, J.; Cuello, A.C. Nerve Growth Factor (NGF) Pathway Biomarkers in Down Syndrome Prior to and after the Onset of Clinical Alzheimer's Disease: A Paired CSF and Plasma Study. *Alzheimer's Dement.* **2021**, *17*, 605–617. [[CrossRef](#)]
149. Fasulo, L.; Brandi, R.; Arisi, I.; La Regina, F.; Berretta, N.; Capsoni, S.; D'Onofrio, M.; Cattaneo, A. ProNGF Drives Localized and Cell Selective Parvalbumin Interneuron and Perineuronal Net Depletion in the Dentate Gyrus of Transgenic Mice. *Front. Mol. Neurosci.* **2017**, *10*, 20. [[CrossRef](#)]

150. Chen, X.-Q.; Mobley, W.C. Alzheimer Disease Pathogenesis: Insights from Molecular and Cellular Biology Studies of Oligomeric A β and Tau Species. *Front. Neurosci.* **2019**, *13*, 659. [[CrossRef](#)]
151. Sola, E.; Capsoni, S.; Rosato-Siri, M.; Cattaneo, A.; Cherubini, E. Failure of Nicotine-Dependent Enhancement of Synaptic Efficacy at Schaffer-Collateral CA1 Synapses of AD11 Anti-Nerve Growth Factor Transgenic Mice. *Eur. J. Neurosci.* **2006**, *24*, 1252–1264. [[CrossRef](#)]
152. Rosato-Siri, M.; Cattaneo, A.; Cherubini, E. Nicotine-Induced Enhancement of Synaptic Plasticity at CA3-CA1 Synapses Requires GABAergic Interneurons in Adult Anti-NGF Mice. *J. Physiol.* **2006**, *576*, 361–377. [[CrossRef](#)] [[PubMed](#)]
153. Lagostena, L.; Rosato-Siri, M.; D’Onofrio, M.; Brandi, R.; Arisi, I.; Capsoni, S.; Franzot, J.; Cattaneo, A.; Cherubini, E. In the Adult Hippocampus, Chronic Nerve Growth Factor Deprivation Shifts GABAergic Signaling from the Hyperpolarizing to the Depolarizing Direction. *J. Neurosci.* **2010**, *30*, 885–893. [[CrossRef](#)] [[PubMed](#)]
154. Arisi, I.; D’Onofrio, M.; Brandi, R.; Felsani, A.; Capsoni, S.; Drovandi, G.; Felici, G.; Weitschek, E.; Bertolazzi, P.; Cattaneo, A. Gene Expression Biomarkers in the Brain of a Mouse Model for Alzheimer’s Disease: Mining of Microarray Data by Logic Classification and Feature Selection. *J. Alzheimer’s Dis.* **2011**, *24*, 721–738. [[CrossRef](#)] [[PubMed](#)]
155. Arisi, I.; D’Onofrio, M.; Brandi, R.; Cattaneo, A.; Bertolazzi, P.; Cumbo, F.; Felici, G.; Guerra, C. Time Dynamics of Protein Complexes in the AD11 Transgenic Mouse Model for Alzheimer’s Disease like Pathology. *BMC Neurosci.* **2015**, *16*, 28. [[CrossRef](#)] [[PubMed](#)]
156. Ballard, C.; Aarsland, D.; Cummings, J.; O’Brien, J.; Mills, R.; Molinuevo, J.L.; Fladby, T.; Williams, G.; Doherty, P.; Corbett, A.; et al. Drug Repositioning and Repurposing for Alzheimer Disease. *Nat. Rev. Neurol.* **2020**, *16*, 661–673. [[CrossRef](#)] [[PubMed](#)]
157. Chumakov, I.; Nabirotkin, S.; Cholet, N.; Milet, A.; Boucard, A.; Toulorge, D.; Pereira, Y.; Gaudens, E.; Traoré, S.; Fouquier, J.; et al. Combining Two Repurposed Drugs as a Promising Approach for Alzheimer’s Disease Therapy. *Sci. Rep.* **2015**, *5*, 7608. [[CrossRef](#)] [[PubMed](#)]
158. Boussicault, L.; Laffaire, J.; Schmitt, P.; Rinaudo, P.; Callizot, N.; Nabirotkin, S.; Hajj, R.; Cohen, D. Combination of Acamprosate and Baclofen (PXT864) as a Potential New Therapy for Amyotrophic Lateral Sclerosis. *J. Neurosci. Res.* **2020**, *98*, 2435–2450. [[CrossRef](#)]
159. Ben-Ari, Y. NKCC1 Chloride Importer Antagonists Attenuate Many Neurological and Psychiatric Disorders. *Trends Neurosci.* **2017**, *40*, 536–554. [[CrossRef](#)]
160. Savardi, A.; Borgogno, M.; Narducci, R.; La Sala, G.; Ortega, J.A.; Summa, M.; Armirotti, A.; Bertorelli, R.; Contestabile, A.; De Vivo, M.; et al. Discovery of a Small Molecule Drug Candidate for Selective NKCC1 Inhibition in Brain Disorders. *Chem* **2020**, *6*, 2073–2096. [[CrossRef](#)]
161. Taubes, A.; Nova, P.; Zalocusky, K.A.; Kosti, I.; Bicak, M.; Zilberter, M.Y.; Hao, Y.; Yoon, S.Y.; Oskotsky, T.; Pineda, S.; et al. Experimental and Real-World Evidence Supporting the Computational Repurposing of Bumetanide for APOE4-Related Alzheimer’s Disease. *Nat. Aging* **2021**, *1*, 932–947. [[CrossRef](#)]
162. Bie, B.; Wu, J.; Lin, F.; Naguib, M.; Xu, J. Suppression of Hippocampal GABAergic Transmission Impairs Memory in Rodent Models of Alzheimer’s Disease. *Eur. J. Pharmacol.* **2022**, *917*, 174771. [[CrossRef](#)] [[PubMed](#)]
163. Virtanen, M.A.; Uvarov, P.; Hübner, C.A.; Kaila, K. NKCC1, an Elusive Molecular Target in Brain Development: Making Sense of the Existing Data. *Cells* **2020**, *9*, 2607. [[CrossRef](#)] [[PubMed](#)]
164. Gagnon, M.; Bergeron, M.J.; Lavertu, G.; Castonguay, A.; Tripathy, S.; Bonin, R.P.; Perez-Sanchez, J.; Boudreau, D.; Wang, B.; Dumas, L.; et al. Chloride Extrusion Enhancers as Novel Therapeutics for Neurological Diseases. *Nat. Med.* **2013**, *19*, 1524–1528. [[CrossRef](#)] [[PubMed](#)]
165. Blaesse, P.; Airaksinen, M.S.; Rivera, C.; Kaila, K. Cation-Chloride Cotransporters and Neuronal Function. *Neuron* **2009**, *61*, 820–838. [[CrossRef](#)]
166. Amadoro, G.; Corsetti, V.; Sancesario, G.M.; Lubrano, A.; Melchiorri, G.; Bernardini, S.; Calissano, P.; Sancesario, G. Cerebrospinal Fluid Levels of a 20–22 KDa NH2 Fragment of Human Tau Provide a Novel Neuronal Injury Biomarker in Alzheimer’s Disease and Other Dementias. *J. Alzheimer’s Dis.* **2014**, *42*, 211–226. [[CrossRef](#)]
167. Scopa, C.; Marrocco, F.; Latina, V.; Ruggeri, F.; Corvaglia, V.; La Regina, F.; Ammassari-Teule, M.; Middei, S.; Amadoro, G.; Meli, G.; et al. Impaired Adult Neurogenesis Is an Early Event in Alzheimer’s Disease Neurodegeneration, Mediated by Intracellular A β Oligomers. *Cell Death Differ.* **2020**, *27*, 934–948. [[CrossRef](#)]