



Clearing Amyloid-Beta by Astrocytes: The Role of Rho GTPases Signaling Pathways as Potential Therapeutic Targets

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Abstract: Astrocytes, vital support cells in the central nervous system (CNS), are crucial for maintaining neuronal health. In neurodegenerative diseases such as Alzheimer's disease (AD), astrocytes play a key role in clearing toxic amyloid- β (A β) peptides. A β , a potent neuroinflammatory trigger, stimulates astrocytes to release excessive glutamate and inflammatory factors, exacerbating neuronal dysfunction and death. Recent studies underscore the role of Rho GTPases-particularly RhoA, Rac1, and Cdc42—in regulating Aβ clearance and neuroinflammation. These key regulators of cytoskeletal dynamics and intracellular signaling pathways function independently through distinct mechanisms but may converge to modulate inflammatory responses. Their influence on astrocyte structure and function extends to regulating endothelin-converting enzyme (ECE) activity, which modulates vasoactive peptides such as endothelin-1 (ET-1). Through these processes, Rho GTPases impact vascular permeability and neuroinflammation, contributing to AD pathogenesis by affecting both A β clearance and cerebrovascular interactions. Understanding the interplay between Rho GTPases and the cerebrovascular system provides fresh insights into AD pathogenesis. Targeting Rho GTPase signaling pathways in astrocytes could offer a promising therapeutic approach to mitigate neuroinflammation, enhance A β clearance, and slow disease progression, ultimately improving cognitive outcomes in AD patients.

Keywords: Alzheimer's disease; amyloid- β (A β); A β clearance; Rho GTPases; astrocytes; endothelinconverting enzyme; neprilysin

1. Introduction

1.1. Astrocytes and AB Clearance

Astrocytes, the most abundant glial cell type in the central nervous system (CNS), play a pivotal role in maintaining neuronal health and function [1]. They are in close contact with cerebral blood vessels and neurons through their foot process structures, forming a key network across the blood–brain barrier, and play an indispensable role in maintaining the homeostasis of the CNS. These cells not only provide nutritional support for neurons, but also regulate ion balance, secrete neurotrophic factors, and play an important role in removing metabolic waste [2–5]. Their diverse functions include providing structural support, regulating ion homeostasis, and participating in neurotransmitter clearance. Increasing evidence suggests that astrocytes actively contribute to the pathogenesis of multiple neurological disorders. In particular, astrocytes have an important role in waste removal processes because their foot process structures form glial limitans around cerebral blood vessels, helping to remove the accumulation of harmful substances such as amyloid β protein (A β) [6,7]. A β is one of the key pathological features of Alzheimer's disease (AD) and is usually accumulated significantly in neurodegenerative lesions [8]. In 2012, Jeffrey



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Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). et al. showed that astrocytes effectively limit the spread of A β in the brain parenchyma by regulating the fluidity of cerebrospinal fluid and brain interstitial fluid, thereby playing a vital role in regulating the metabolic balance of A β in the brain [9]. In recent years, increasing evidence has highlighted the significance of astrocytes in the clearance of A β peptides, a hallmark of AD [10–13].

1.1.1. Astrocyte Reactivity and Functions

Morphological changes: reactive astrocytes often exhibit hypertrophy and hyperplasia, accompanied by increased expression of glial fibrillary acidic protein (GFAP), a hallmark marker of astrocyte activation [14]. These morphological changes can lead to the formation of glial scars, which may impede neuronal regeneration and plasticity [15,16].

Altered gene expression: reactive astrocytes upregulate the expression of a variety of genes, including inflammatory cytokines, chemokines, and growth factors [17–20]. Some of these factors, such as interleukin-1 β (IL-1 β) and tumor necrosis factor- α (TNF- α), can exacerbate neuroinflammation and contribute to neurodegeneration [21–23]. However, other factors, such as brain-derived neurotrophic factor (BDNF) and glial cell line-derived neurotrophic factor (GDNF), may have neuroprotective effects [24,25].

Functional modifications: reactive astrocytes can exhibit altered ion channel and transporter expression, leading to changes in their ability to regulate extracellular ion concentrations and neurotransmitter levels [26–29]. They may also release neurotoxic substances, such as glutamate and reactive oxygen species (ROS), which can damage neurons [30,31].

1.1.2. Astrocyte-Mediated Aβ Clearance Mechanisms

Phagocytosis: astrocytes can directly engulf and degrade $A\beta$ through phagocytosis. This process involves the recognition of $A\beta$ by specific receptors on the astrocyte surface, followed by internalization and lysosomal degradation [32–34].

Endocytosis: astrocytes can internalize A β through endocytosis, a process that involves the formation of vesicles that transport A β into the cell [35–37]. Once inside the cell, A β can be degraded by lysosomal enzymes or transported to the perivascular space for clearance [38,39].

Perivascular clearance: astrocytes play a critical role in perivascular clearance, a process by which $A\beta$ is transported from the brain parenchyma to the cerebrospinal fluid (CSF) via the perivascular space. Astrocytes extend their endfeet, to wrap around blood vessels, forming a tight junction that regulates the exchange of substances between the blood and the brain [9]. $A\beta$ can be transported from the brain parenchyma to the perivascular space through these endfeet and then cleared into the CSF [40].

Secretion of A β -degrading enzymes: astrocytes can secrete a variety of enzymes, including neprilysin, insulin-degrading enzyme (IDE), and matrix metalloproteinases (MMPs), that can degrade A β . These enzymes can cleave A β into smaller, less toxic fragments, thereby reducing its neurotoxicity [12,41,42].

1.1.3. Impaired Astrocyte Function and Aβ Accumulation

In AD, astrocyte function becomes impaired, leading to reduced A β clearance and increased A β accumulation [43]. This impairment may be due to a variety of factors, including oxidative stress, inflammation, and genetic mutations [18,44,45]. As a result, A β accumulates in the brain parenchyma, forming plaques that disrupt neuronal communication and contribute to neurodegeneration.

1.1.4. Therapeutic Implications

Understanding the role of astrocytes in A β clearance has important therapeutic implications for AD. By targeting astrocyte function, it may be possible to enhance A β clearance and slow disease progression. Potential therapeutic strategies include the following: Pharmacological interventions: developing drugs that can enhance astrocyte function, such as by increasing the expression of A β -degrading enzymes or reducing inflammation [46].

Cell therapy: transplanting healthy astrocytes into the brain to replace damaged or dysfunctional astrocytes [47]. Using gene therapy to deliver genes that can enhance astrocyte function or reduce A β production [48].

In conclusion, astrocytes play a crucial role in A β clearance, and impaired astrocyte function contributes to A β accumulation and AD pathogenesis. By understanding the mechanisms underlying astrocyte-mediated A β clearance, we may be able to develop novel therapeutic strategies for AD.

1.2. Rho GTPases in Astrocyte

1.2.1. Rho GTPases in Cellular Functions

Rho GTPases, a family of small GTPases, are molecular switches that cycle between an inactive GDP-bound state and an active GTP-bound state. This dynamic regulation allows them to control a wide range of cellular processes, including cell proliferation, differentiation, migration, and adhesion [49]. They are particularly crucial in regulating the actin cytoskeleton, a complex network of proteins that provides structural support to cells and enables various cellular functions, such as cell motility and division [50].

One of the most critical roles of Rho GTPases lies in the regulation of the actin cytoskeleton, a dynamic network of actin filaments that provides structural support to cells and enables various cellular functions, such as cell motility, division, and morphogenesis. Rho GTPases influence actin dynamics through several mechanisms:

Actin polymerization: Rho GTPases, particularly Ras homolog family member A (RhoA) and Rac1, can directly interact with actin-binding proteins, such as the Arp2/3 complex, to promote actin polymerization [51,52]. This leads to the formation of actin filaments, which are essential for cell shape and motility [53].

Stress fiber formation: RhoA is a key regulator of stress fiber formation, which produces contractile bundles of actin filaments [54]. The activation of RhoA leads to the activation of Rho-associated kinase (ROCK), which phosphorylates the myosin light chain, promoting myosin–actin interactions resulting in stress fiber formation [55,56].

Lamellipodia and filopodia formation: Ras-related C3 botulinum toxin substrate 1 (Rac1) and cell division control protein 42 homolog (Cdc42), respectively, are key regulators of lamellipodia and filopodia formation [57–59]. These membrane protrusions are essential for cell migration and invasion. Activation of Rac1 and Cdc42 leads to the activation of various actin-binding proteins, such as WASP and WAVE, which promote actin polymerization and the formation of these protrusions [60,61].

Endocytosis and exocytosis: Rho GTPases also play a role in regulating endocytosis and exocytosis, processes that involve the formation and fusion of membrane-bound vesicles. For example, Rac1 and Cdc42 are involved in clathrin-mediated endocytosis, while RhoA is involved in exocytosis [62–65].

1.2.2. Rho GTPases in Diseases

Dysregulation of Rho GTPases has been implicated in a variety of human diseases, including cancer [66], cardiovascular disease, and neurodegenerative disorders. For example, aberrant activation of RhoA has been linked to tumor invasion and metastasis, while aberrant activation of Rac1 has been linked to autoimmune diseases [67,68]. In neurodegenerative diseases, such as AD and Parkinson's disease, dysregulation of Rho GTPases has been implicated in neuronal cell death and synaptic dysfunction [69,70]. Given their critical role in various cellular processes and their involvement in numerous diseases, Rho GTPases have emerged as attractive therapeutic targets. Several strategies are being explored to target Rho GTPases, including the following:

Small-molecule inhibitors: these compounds can directly inhibit Rho GTPases or their downstream effectors [71,72].

RNA interference: RNA interference can be used to knock down the expression of Rho GTPases [73]. For example, lentiviral vector-mediated shRNA targeting RhoA was applied to cultured Schwann cells to suppress RhoA expression [74].

Gene therapy: gene therapy can be used to overexpress or downregulate Rho GTPases. The designed vector, such as bacterial enzyme C3-ADP ribosyl transferase (C3), blocks RhoA from becoming active and helps axons grow and regenerate, which promotes outgrowth [75]. By targeting Rho GTPases, it may be possible to develop novel therapies for a wide range of diseases. However, further research is needed to fully understand the complex roles of Rho GTPases in cellular processes and to develop safe and effective therapies.

1.2.3. Rho GTPases in the CNS

In the central nervous system (CNS), Rho GTPases play a pivotal role in neuronal development, synaptic plasticity, and neurodegenerative diseases [76–78]. They regulate several key processes, including the following:

Axon guidance and growth cone dynamics: in the nervous system, Rho GTPases play a pivotal role in neuronal development and regeneration by regulating the cytoskeleton, with particular importance in dendrite and axon growth [79–84]. During axon guidance, Rho, Rac, and Cdc42 GTPases serve as key modulators of neuroplasticity. RhoA activation increases myosin II activity, resulting in axon retraction and growth cone collapse. Conversely, Rac1 and Cdc42 promote axon extensions by driving the formation of lamellipodia and filopodia. Through these mechanisms, Rho GTPase signaling regulates actin filament dynamics in growth cones, enabling them to respond effectively to guidance cues [85].

Rho GTPases also mediate growth cone responses to neurotrophic factors. For instance, the activation of Trk and p75 receptors can either prompt actin filament aggregation or reduce RhoA activity, thereby shaping the pseudopodia structures of growth cones [86,87]. Highly conserved in eukaryotes, Rho GTPases primarily control neuronal migration by orchestrating the assembly and actin rearrangement and microtubule cytoskeletons. Rho GTPases regulate cell polarity, adhesion, and directional migration during the formation of cortical neuron layers and the patterning of brain circuits, facilitating the development of cortical structure and establishing functional neural connections [88]. These roles make Rho GTPases indispensable as cytoskeletal regulators in nervous system development.

Dendritic spine morphology and synaptic plasticity: Rho GTPases influence the formation, maturation, and elimination of dendritic spines, which are the sites of synaptic contact between neurons. They also regulate synaptic plasticity, the ability of synapses to strengthen or weaken in response to activity. The Rho family of small GTPases are essential regulators of synaptic plasticity, influencing synaptic development through actin cytoskeleton remodeling. Key members, including Rac1, Cdc42, and RhoA, along with their effectors, play crucial roles in spine formation, spine morphology, receptor trafficking, and the processes underlying synaptic plasticity, learning, and memory. The activation of Rac1 and Cdc42 leads to an increase in immature spines. Some of these spines mature via a Rac1-dependent pathway, while others are pruned through a RhoA-dependent mechanism. This supports selective growth and balanced synaptic regulation [89].

In the active state, Rho GTPases engage with downstream effectors to orchestrate spine morphology and synapse development [90]. Studies have identified various GEFs and GAPs within the Rho family as essential for spine morphogenesis. Rho GTPases are regulated by GEFs (guanine nucleotide exchange factors), which facilitate the exchange of GDP for GTP to activate the GTPases, and by GAPs (GTPase-activating proteins), which accelerate GTP hydrolysis to inactivate them [90]. Within the spine, GEFs are pivotal in regulating the actin cytoskeleton by modulating Rho GTPases and spines, whereas RhoA restrains excessive synaptic development, maintaining a dynamic balance in excitatory synapses [13]

Neuroinflammation: Rho GTPases are involved in the inflammatory response in the CNS, regulating the activation of microglia and astrocytes. Inflammation in the central

nervous system (CNS) activates the small GTPase RhoA and its downstream effector ROCK. Activation of this pathway not only directly leads to neuronal damage and cell death, but also promotes the shrinkage and loss of neural processes and synapses [91]. In the CNS inflammatory response, the participation of astrocytes and microglia exacerbates neuronal damage, and the RhoA/ROCK signaling pathway plays a key role in regulating the functions of these glial cells and immune cells. Studies have shown that the activation of the RhoA/ROCK pathway induces neurodegeneration in the CNS [92,93].

Under neuroinflammatory conditions, heparan sulfate proteoglycans (CSPGs) released by astrocytes interact with myelin-associated inhibitor Nogo receptors, activating the RhoA/ROCK pathway and leading to axonal growth cone collapse [94]. Studies by Fujita et al. showed that the activation of RhoA/ROCK induces cytoskeletal reorganization, inhibiting neurite growth and leading to growth cone collapse [95]. In contrast, the inhibition of the RhoA/ROCK pathway promotes neurite regeneration and recovery. This was further verified by Zhang et al.'s study: by inhibiting ROCK activity with Fasudil, neuronal recovery and a significant reduction in the proliferation of reactive astrocytes after cerebral ischemia/reperfusion injury were observed [96,97].

In summary, the RhoA/ROCK pathway plays an important role in neuronal injury and neurodegeneration by regulating glial cell activation and neurotoxic phenotype transition in CNS inflammation. Inhibiting this pathway not only helps reduce neuronal damage caused by neuroinflammation but also promotes neuronal regeneration and CNS repair.

1.2.4. Rho GTPases in Regulating Astrocyte Morphology

Rho GTPases are essential regulators of astrocyte morphology, function, and reactivity. Holtje et al. found that Rho plays an inhibitory role in astrocyte neurite formation during astrocyte stellation [98]. By selectively inhibiting Rho's downstream effector ROCK with Y27632, they observed accelerated wound healing, enhanced polarized neurite formation, and increased astrocyte migration toward the lesion site, suggesting that Rho negatively regulates astrocyte neurite growth and migration responses after injury [90]. Etienne-Manneville et al. further showed that Rho influences the microtubule cytoskeleton during astrocyte migration, while Rac is essential for neurite development and maintenance in migrating astrocytes [99]. Additionally, Cdc42 is critical for forming neurites that contribute to the elongated morphology of astrocytes [99]. This implies that Rac facilitates cell elongation by directly affecting microtubule dynamics or modulating microtubule-dependent processes. Supporting this, Daub et al. showed that Rac may regulate microtubule dynamics by phosphorylating p65PAK, which inhibits the microtubule-destabilizing protein [100].

These findings collectively suggest that Rho, Rac, and Cdc42 have distinct roles in astrocyte morphology and migration, with Rho acting as a negative regulator, while Rac and Cdc42 contribute to neurite elongation and stability by modulating the microtubule cytoskeleton.

1.2.5. Rho GTPases in Regulating Astrocyte Function and Reactivity

Rho GTPases also regulate a variety of astrocyte functions, including the following:

Gliotransmitter release: astrocytes release gliotransmitters, such as glutamate and ATP, which can modulate neuronal activity. Rho GTPases have been identified as key regulators of exocytosis and may play a role in modulating the exocytosis of these gliotransmitters [101,102].

Water and ion homeostasis: astrocytes play a crucial role in maintaining water and ion homeostasis in the brain [103,104]. Rho GTPases regulate the expression and activity of ion channels and transporters, which are essential for this function [105,106].

Neuroinflammation: as mentioned earlier, Rho GTPases are involved in the inflammatory response in the CNS, including the activation of astrocytes. Inhibition of the RhoA/ROCK pathway can significantly reduce reactive gliosis, reduce the over-activation of astrocytes, and induce their expression of pro-survival genes [107–110]. Profilin 1 (PFN1), as one of the downstream effectors of RhoA/ROCK, may play a neuroprotective role by affecting the polarization of microglia. Ermei et al. indicated that knocking down PFN1 can promote the neuroprotective polarization of microglia, which may be achieved through the inhibition of RhoA/ROCK [111,112]. In addition, NF- κ B, a downstream effector of ROCK, plays an important role in the neurotoxic phenotype conversion of microglia. Zhang et al. found that inhibiting the RhoA/ROCK/NF- κ B pathway can prevent microglia from polarizing to neurotoxic subtypes, promoting their transformation into neuroprotective phenotypes, and help them recover from brain damage under inflammatory conditions [113]. This activation can lead to the release of inflammatory cytokines and chemokines, which may contribute to neurodegeneration.

In response to injury or disease, astrocytes undergo a process known as reactive astrogliosis. This process involves changes in astrocyte morphology, gene expression, and function. Rho GTPases play a critical role in regulating reactive astrogliosis [114]. For example, the activation of RhoA can promote the formation of glial scars, which can limit tissue damage but can also hinder neuronal regeneration [115]. In contrast, loss of Rac1 can promote neurogenesis and synaptogenesis [116]. Rho GTPases are essential regulators of astrocyte function and reactivity. By understanding the role of Rho GTPases in astrocytes, we can gain insights into the mechanisms underlying neurodegenerative diseases and develop novel therapeutic strategies. Targeting Rho GTPases may provide a promising approach to modulate astrocyte function and promote neuroprotection.

1.3. Rho GTPases in $A\beta$ Clearance

Given that Rho GTPases are dysregulated in AD, several studies have investigated the relationship between Rho GTPases, amyloid precursor protein (APP) synthesis, and A β production across various cell lines. For instance, in primary hippocampal neurons from mice, the inhibition of Rac1 negatively regulates APP gene synthesis [117] and reduces A β 42 production by altering γ -secretase substrate selectivity, leading to increased processing of Notch1 instead of APP [118].

The dysregulation of Rho GTPase signaling has been linked to impaired A β clearance and the progression of AD. For instance, age-related changes in Rho GTPase activity can contribute to decreased astrocytic and microglial function, leading to diminished A β clearance and increased deposition of neurotoxic aggregates. Moreover, genetic and pharmacological modulation of Rho GTPases has shown promise in preclinical models, suggesting that maybe targeting these signaling pathways could enhance A β clearance and provide therapeutic benefits in AD [119–121].

Thus, in the following sections, we discuss the mechanisms by which astrocyteassociated Rho GTPases facilitate $A\beta$ clearance from the brain, particularly focusing on the role of Rho GTPases in $A\beta$ clearance enzymes in astrocytes. Additionally, we explore the potential of astrocyte Rho GTPases as therapeutic targets for disease modification in AD.

2. Aβ Clearance Enzymes in Astrocytes

In AD, the accumulation of $A\beta$ is a hallmark pathological feature, with astrocytes playing a crucial role in its clearance through various mechanisms. Astrocytes first help reduce $A\beta$ deposition in the brain by transferring $A\beta$ from the brain parenchyma to the perivascular space, a process dependent on the functional integrity of the neurovascular unit [122]. Additionally, $A\beta$ can be transported across the blood–brain barrier (BBB) and cleared out of the brain, helping to mitigate its neurotoxicity [123,124]. Astrocytes also contribute by clearing $A\beta$ from the brain's lymphatic system, thus reducing $A\beta$ -associated damage through their clearance functions [9,125]. In particular, astrocytes degrade $A\beta$ by secreting proteases, such as neprilysin (NEP) and ECEs (ECE-1 and ECE-2) as $A\beta$ -degrading enzymes. These enzymes cleave $A\beta$ peptides at specific sites, inactivating or transforming them to reduce their toxicity and accumulation [126,127].

2.1. NEP

NEP is a key Aβ-degrading enzyme primarily located in hippocampal neurons in the CA1-3 region [128,129] and in reactive astrocytes, but is found less frequently in mi-

croglia [130,131]. Studies have shown that elevated NEP levels in AD patients can reduce A β 42 [132]. Furthermore, Kim et al. found that exercise-induced hormones significantly increased NEP release in astrocytes, effectively reducing A β levels [11]. NMDA antagonists inhibit NEP's role in A β degradation, reducing the ability of astrocytes to manage exogenous A β [133]. Other studies have shown that astrocyte transplantation can promote A β clearance, while NEP inhibitors can negatively affect A β clearance efficiency [10]. However, NEP plays a key role in the clearance of extracellular A β , while ECEs primarily degrade A β intracellularly [134]. Rho GTPases are molecular switches that relay extracellular signals into the cell, where they initiate intracellular events.

2.2. ECE-1

Multiple studies provide evidence that ECEs in astrocytes assist A β clearance and may protect against AD. Eckman et al. demonstrated that ECE-1 and ECE-2 can degrade A β in vitro and in vivo, with reduced ECE activity leading to increased amyloid plaque formation. This was one of the first studies to suggest ECE's role in regulating A β levels in the brain [135]. Iwata et al. confirmed that decreased ECE activity results in A β accumulation in animal models [136]. Subsequent research by Padilla et al. and Palmer et al. emphasized astrocyte ECE's essential role in clearing A β from brain capillaries, highlighting astrocyte ECEs as key to A β breakdown, particularly in the perivascular space [137,138].

ECE-1 is a membrane-bound protein that plays a critical role in mediating vasoconstriction during inflammatory responses or tissue injury. It achieves this by converting large precursor molecules of endothelin-1 (ET-1) into their biologically active forms. ECE-1 is primarily localized in endosomal compartments and has been shown to exhibit activation in endothelial cells in response to A β [138,139].

The presence of $A\beta$ oligomers is particularly concerning, as these aggregates can disrupt cerebral blood flow in capillaries. This disruption is mediated through the production of ROS, which subsequently triggers the release of ET-1 [140]. Elevated levels of ET-1 can have detrimental effects on neurovascular function and exacerbate neuroinflammatory processes within the brain. Specifically, overexpression of ET-1 in astrocytes has been linked to heightened neuroinflammatory damage, worsening the overall pathological state [141]. Interestingly, research has indicated that inflammatory responses lead to the activation of reactive astrocytes, particularly in conditions like focal multiple sclerosis lesions, which are identified as primary sources of ET-1. In contrast, astrocytes that remain unaffected do not display significant ET-1 activity [142] (Figure 1). This differential expression underscores the role of reactive astrocytes in the context of neuroinflammation and highlights the complex interplay between neuroinflammatory factors and glial cell responses.



Figure 1. Schematic diagram of astrocyte ECE-1 in $A\beta$ clearance. The upward arrow indicates an increase.

Further investigations into inflammatory mediators, such as interleukin-1 beta (IL-1 β), revealed that these cytokines can significantly upregulate ET-1 production in astrocytes. Conversely, compounds like resveratrol showed potential in inhibiting ET-1 production, suggesting promising therapeutic avenues to mitigate neuroinflammation and associated vascular dysfunction [143]. By exploring the pathways involving ECE-1, researchers may uncover new strategies for treating neurodegenerative diseases characterized by inflammation and impaired blood flow.

2.3. ECE-2

ECE-2 is predominantly expressed in neurons but is also found in specific populations of astrocytes and microglia [144,145]. This enzyme plays a crucial role in the metabolism of neuropeptides and has been implicated in various neurobiological processes. Studies utilizing ECE-2 knockout mice demonstrated that the absence of this enzyme leads to significantly increased levels of A β in the brain. This accumulation of A β is closely associated with cognitive impairments, resulting in notable deficits in memory and learning [139,146]. Interestingly, in patients diagnosed with AD, research showed that the levels of ECE-2 mRNA are elevated. This increase may be a compensatory response to the accumulation of $A\beta$ and reduced cerebral blood flow often observed in AD [145]. The relationship between ECE-2 and $A\beta$ levels is complex. While ECE-2 deficiency appears to contribute to elevated A β levels, it is essential to consider that other contributing factors may play a more significant role in this process. For instance, the pathological environment created by neuroinflammation and vascular dysfunction in AD may influence the dynamics of A β accumulation independently of ECE-2 level [147–150]. Thus, while the relationship between ECE-2 and A β accumulation is noteworthy, it is crucial to recognize that the mechanisms underlying A β deposition in AD are multifactorial.

Future research should aim to clarify the precise role of ECE-2 in the context of neurodegenerative diseases and explore potential therapeutic interventions that target this enzyme to mitigate cognitive decline associated with $A\beta$ accumulation. Understanding these intricate pathways could pave the way for novel strategies in the treatment of AD.

Collectively, these studies affirm that ECE-1 and ECE-2, particularly within astrocytes, significantly contribute to $A\beta$ degradation, potentially reducing the risk of amyloid plaque formation in the brain.

3. Rho GTPase Family and Their Signaling Pathways Regulating $A\beta$ Clearance Enzymes

The Rho GTPase family may effectively regulate the activity of astrocyte ECEs through multiple signaling pathways, affecting the function of the endothelin system. Astrocytes regulate the expression and release of ECEs by releasing proinflammatory factors and oxidative stress molecules, changing the level of endothelin, and playing a key role in the pathological process of the nervous system [151–153]. The following are the specific mechanisms and pathological effects of Rho GTPase family members regulating astrocyte ECE and its related signaling pathways.

3.1. RhoA/ROCK

The RhoA/ROCK signaling pathway affects the morphology, adhesion, and activation state of astrocytes by regulating the remodeling of the cytoskeleton. When RhoA activates ROCK, the contraction and reorganization of the cytoskeleton are enhanced, making astrocytes more active under inflammatory or oxidative stress conditions, promoting the release of ECEs [154]. Research by Minamino et al. showed that the enhanced ECE activity can catalyze the production of more endothelin and activate surrounding endothelin receptors, regulate local vascular tension, and aggravate inflammatory responses and ECE-1 may promote the occurrence and development of atherosclerosis through the autocrine and paracrine mechanisms of endothelin-1 and blocking ECE-1 can effectively reduce this promoting effect [155].

Studies have shown that inhibiting ROCK can reduce reactive gliosis and increase the expression of astrocyte pro-survival genes [93,98,156,157], and this regulation is essential for the health of the nervous system. Kimura et al. showed that upregulation of the RhoA/ROCK pathway is closely related to a series of pathological processes in ischemic stroke and spinal cord injury [158], including neuronal apoptosis, neuroinflammation, BBB dysfunction, astrogliosis, and axonal growth inhibition. Animal models and clinical trials have demonstrated that that ROCK inhibitors, such as Fasudil and VX-210, can reduce apoptosis, neuroinflammation, oxidative stress, and axonal growth inhibition in ischemic stroke and spinal cord injury [159–161].

In addition, inhibiting the RhoA/ROCK pathway may have deleterious effects on neuroinflammation, BBB dysfunction, neuronal apoptosis, astrogliosis, and axonal injury after ischemic stroke. Wen et al. demonstrated that the inhibition of the RhoA/ROCK pathway with Y-27632 can significantly improve cerebral ischemia/reperfusion injury [162].

These findings provide a new perspective for understanding the dual role of the RhoA/ROCK signaling pathway in neuropathology and offer potential intervention targets for the future treatment of ischemic injury.

3.2. Rac1

The production of brain ET-1, which increases in brain disorders, is involved in the pathophysiological response of the nervous system. Barker et al. have concluded that the brain of AD patients has an increased amount of ET-1 in the temporal cortex of the brain [163]. Rac1 plays an important role in oxidative stress response, mainly by regulating the production of ROS [164]. Rac1 promotes the generation of ROS by activating NADPH oxidase, increasing the oxidative stress level of astrocytes [165–167], which may affect the expression and secretion of ECEs [168,169]. At the onset of neuroinflammation, Rac1-mediated ROS generation is vital, as it activates and boosts pro-inflammatory signaling ECE activity, and increases endothelin production [170,171].

Therefore, in neurodegenerative diseases such as AD, the activation of Rac1 may be closely related to the generation of ROS and the upregulation of ECEs in astrocytes, leading to increased endothelin levels, aggravated neuroinflammation, and oxidative stress, thus causing further damage to neurons.

3.3. Cdc42

Cell division control protein 42 homolog (Cdc42) plays an important role in astrocyte migration and morphological regulation, especially by controlling the cytoskeleton, and regulating the formation of cell pseudopods and cell extensibility, enhancing the migration ability of astrocytes [172,173]. The activity of Cdc42 can enhance the migration and reactivity of astrocytes [99]. Especially with brain trauma, astrocytes will migrate to the damaged area and release ECEs to regulate local blood vessels and assist in repair. In models of neural injury [174,175] and in patients with stroke, traumatic brain injury, and neurodegenerative diseases, such as AD, brain levels of ET-1 are significantly elevated [176–178].

Immunohistochemical studies showed that ET-1 in the damaged brain is mainly produced by brain microvascular endothelial cells and reactive astrocytes [179,180]. Studies have shown that factors such as TNF- α , IL-1 β , thrombin, and hypoxia can induce brain microvascular endothelial cells and astrocytes to secrete ET-1, and ET-1 itself can also stimulate astrocytes to further produce ET-1 [181–184]. However, excessive release of ECEs may lead to excessive vasoconstriction, further triggering ischemia and delaying the tissue recovery process. In brain trauma models, the activation of Cdc42 promotes the migration of astrocytes to the injured area and induces vascular responses through the activation of the endothelin system to support the repair of the injured area [185–188].

In summary, Rho GTPase family members RhoA, Rac1, and Cdc42 regulate the activity of ECEs and the endothelin system in astrocytes through multiple signaling pathways, affecting vascular regulation and inflammatory response. These regulatory mechanisms show specific patterns in different pathological states, revealing the potential target value of Rho GTPase signaling in neuropathological processes (Figure 2).



Figure 2. Rho GTPase family's role with endothelin-converting enzyme 1 in astrocytes. The upward arrow represents an increase.

4. Conclusions

The role of Rho GTPases in the central nervous system has received increasing attention, especially their regulatory role in astrocyte function and the mechanisms of neurodegenerative diseases. Astrocytes are the most abundant glial cells in the central nervous system and play an important role in maintaining homeostasis, providing metabolic support to neurons, and clearing neurotoxic substances such as A β .

The accumulation of $A\beta$ is a major feature of AD and is directly related to the significant neurodegenerative process. The complex interactions between Rho GTPases, astrocyte ECE activity, and $A\beta$ clearance reveal the high complexity of astrocyte responses in neurodegenerative diseases. Studying the dynamic regulatory mechanisms of these signaling pathways may not only deepen the understanding of astrocyte function, but also provide novel therapeutic strategies to enhance $A\beta$ clearance.

Studying how Rho GTPases regulate the activity of astrocytes will not only help to reveal these mechanisms, but also may provide potential therapeutic targets for AD and similar diseases. RhoA, Rac1, and Cdc42 in the Rho GTPase family are the focus of research, which significantly affects the morphology and function of astrocytes. These GTPases maintain the structural integrity and plasticity of astrocytes by regulating cytoskeletal dynamics. The RhoA/ROCK signaling pathway has been shown to promote astrocyte activation, enhance the secretion of proinflammatory factors, and regulate the activity of ECE under inflammatory conditions. Enhanced ECE activity leads to increased ET-1 levels, which, as a potent vasoconstrictor involved in neuroinflammation, can aggravate neuronal damage and promote the progression of neurodegenerative diseases. Rac1 indirectly affects the clearance of $A\beta$ by regulating the generation of ROS in astrocytes, affecting oxidative stress and the expression level of ECE. The accumulation of ROS also enhances the inflammatory response, forming a feedback loop that further weakens the neuroprotective function of astrocytes. In contrast, Cdc42 plays a key role in the migration of astrocytes and the response to injury. Its ability to regulate the formation of cellular processes promotes the migration of astrocytes to the site of injury and enhances the efficiency of A β clearance.

Future studies should focus on the refined analysis of the signal specificity of Rho GTPases in different pathological states and their regulatory networks, and explore strategies for the combined regulation of RhoA, Rac1, and Cdc42 to achieve optimal therapeutic effects. This will provide an important scientific basis for the development of targeted intervention methods for AD and other neurodegenerative diseases and open new directions for innovative therapies based on regulating astrocyte function.

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References

- 1. Wei, D.C.; Morrison, E.H. Histology, Astrocytes. In *StatPearls*; StatPearls Publishing: Treasure Island, FL, USA, 2024.
- 2. Molofsky, A.V.; Deneen, B. Astrocyte development: A Guide for the Perplexed. Glia 2015, 63, 1320–1329. [CrossRef] [PubMed]
- 3. Nedergaard, M.; Ransom, B.; Goldman, S.A. New roles for astrocytes: Redefining the functional architecture of the brain. *Trends Neurosci.* **2003**, *26*, 523–530. [CrossRef] [PubMed]
- 4. Petzold, G.C.; Murthy, V.N. Role of astrocytes in neurovascular coupling. Neuron 2011, 71, 782–797. [CrossRef] [PubMed]
- 5. Hirrlinger, J.; Nimmerjahn, A. A perspective on astrocyte regulation of neural circuit function and animal behavior. *Glia* 2022, 70, 1554–1580. [CrossRef]
- Ferrari-Souza, J.P.; Ferreira, P.C.L.; Bellaver, B.; Tissot, C.; Wang, Y.T.; Leffa, D.T.; Brum, W.S.; Benedet, A.L.; Ashton, N.J.; De Bastiani, M.A.; et al. Astrocyte biomarker signatures of amyloid-beta and tau pathologies in Alzheimer's disease. *Mol. Psychiatry* 2022, 27, 4781–4789. [CrossRef]
- Zysk, M.; Beretta, C.; Naia, L.; Dakhel, A.; Pavenius, L.; Brismar, H.; Lindskog, M.; Ankarcrona, M.; Erlandsson, A. Amyloid-beta accumulation in human astrocytes induces mitochondrial disruption and changed energy metabolism. *J. Neuroinflammation* 2023, 20, 43. [CrossRef]
- 8. Zhang, Y.; Chen, H.; Li, R.; Sterling, K.; Song, W. Amyloid beta-based therapy for Alzheimer's disease: Challenges, successes and future. *Signal Transduct. Target. Ther.* **2023**, *8*, 248. [CrossRef]
- 9. Iliff, J.J.; Wang, M.; Liao, Y.; Plogg, B.A.; Peng, W.; Gundersen, G.A.; Benveniste, H.; Vates, G.E.; Deane, R.; Goldman, S.A.; et al. A paravascular pathway facilitates CSF flow through the brain parenchyma and the clearance of interstitial solutes, including amyloid beta. *Sci. Transl. Med.* 2012, *4*, 147ra111. [CrossRef]
- 10. Pihlaja, R.; Koistinaho, J.; Kauppinen, R.; Sandholm, J.; Tanila, H.; Koistinaho, M. Multiple cellular and molecular mechanisms are involved in human Abeta clearance by transplanted adult astrocytes. *Glia* **2011**, *59*, 1643–1657. [CrossRef]
- Kim, E.; Kim, H.; Jedrychowski, M.P.; Bakiasi, G.; Park, J.; Kruskop, J.; Choi, Y.; Kwak, S.S.; Quinti, L.; Kim, D.Y.; et al. Irisin reduces amyloid-beta by inducing the release of neprilysin from astrocytes following downregulation of ERK-STAT3 signaling. *Neuron* 2023, 111, 3619–3633.e3618. [CrossRef]
- 12. Yamamoto, N.; Ishikuro, R.; Tanida, M.; Suzuki, K.; Ikeda-Matsuo, Y.; Sobue, K. Insulin-signaling Pathway Regulates the Degradation of Amyloid beta-protein via Astrocytes. *Neuroscience* **2018**, *385*, 227–236. [CrossRef] [PubMed]
- 13. Kiraly, D.D.; Eipper-Mains, J.E.; Mains, R.E.; Eipper, B.A. Synaptic plasticity, a symphony in GEF. *ACS Chem Neurosci.* **2010**, *1*, 348–365. [CrossRef] [PubMed]
- Wilhelmsson, U.; Bushong, E.A.; Price, D.L.; Smarr, B.L.; Phung, V.; Terada, M.; Ellisman, M.H.; Pekny, M. Redefining the concept of reactive astrocytes as cells that remain within their unique domains upon reaction to injury. *Proc. Natl. Acad. Sci. USA* 2006, 103, 17513–17518. [CrossRef] [PubMed]
- 15. Xie, C.; Shen, X.; Xu, X.; Liu, H.; Li, F.; Lu, S.; Gao, Z.; Zhang, J.; Wu, Q.; Yang, D.; et al. Astrocytic YAP Promotes the Formation of Glia Scars and Neural Regeneration after Spinal Cord Injury. *J. Neurosci.* **2020**, *40*, 2644–2662. [CrossRef]
- 16. Middeldorp, J.; Hol, E.M. GFAP in health and disease. *Prog Neurobiol.* 2011, 93, 421–443. [CrossRef]
- 17. Schlotterose, L.; Cossais, F.; Lucius, R.; Hattermann, K. Breaking the circulus vitiosus of neuroinflammation: Resveratrol attenuates the human glial cell response to cytokines. *Biomed Pharmacother.* **2023**, *163*, 114814. [CrossRef]
- LaRocca, T.J.; Cavalier, A.N.; Roberts, C.M.; Lemieux, M.R.; Ramesh, P.; Garcia, M.A.; Link, C.D. Amyloid beta acts synergistically as a pro-inflammatory cytokine. *Neurobiol. Dis.* 2021, 159, 105493. [CrossRef]
- 19. Kim, H.; Leng, K.; Park, J.; Sorets, A.G.; Kim, S.; Shostak, A.; Embalabala, R.J.; Mlouk, K.; Katdare, K.A.; Rose, I.V.L.; et al. Reactive astrocytes transduce inflammation in a blood-brain barrier model through a TNF-STAT3 signaling axis and secretion of alpha 1-antichymotrypsin. *Nat. Commun.* **2022**, *13*, 6581. [CrossRef]
- Chapouly, C.; Tadesse Argaw, A.; Horng, S.; Castro, K.; Zhang, J.; Asp, L.; Loo, H.; Laitman, B.M.; Mariani, J.N.; Straus Farber, R.; et al. Astrocytic TYMP and VEGFA drive blood-brain barrier opening in inflammatory central nervous system lesions. *Brain* 2015, 138, 1548–1567. [CrossRef]
- 21. Meraz-Rios, M.A.; Toral-Rios, D.; Franco-Bocanegra, D.; Villeda-Hernandez, J.; Campos-Pena, V. Inflammatory process in Alzheimer's Disease. *Front. Integr. Neurosci.* 2013, 7, 59. [CrossRef]
- Ali, J.; Khan, A.; Park, J.S.; Tahir, M.; Ahmad, W.; Choe, K.; Kim, M.O. Neuroprotective Effects of N-methyl-(2S, 4R)-trans-4hydroxy-L-proline (NMP) against Amyloid-beta-Induced Alzheimer's Disease Mouse Model. *Nutrients* 2023, 15, 4986. [CrossRef] [PubMed]

- 23. Carrero, I.; Gonzalo, M.R.; Martin, B.; Sanz-Anquela, J.M.; Arevalo-Serrano, J.; Gonzalo-Ruiz, A. Oligomers of beta-amyloid protein (Abeta1-42) induce the activation of cyclooxygenase-2 in astrocytes via an interaction with interleukin-1beta, tumour necrosis factor-alpha, and a nuclear factor kappa-B mechanism in the rat brain. *Exp. Neurol.* **2012**, *236*, 215–227. [CrossRef] [PubMed]
- de Pins, B.; Cifuentes-Diaz, C.; Farah, A.T.; Lopez-Molina, L.; Montalban, E.; Sancho-Balsells, A.; Lopez, A.; Gines, S.; Delgado-Garcia, J.M.; Alberch, J.; et al. Conditional BDNF Delivery from Astrocytes Rescues Memory Deficits, Spine Density, and Synaptic Properties in the 5xFAD Mouse Model of Alzheimer Disease. *J. Neurosci.* 2019, *39*, 2441–2458. [CrossRef] [PubMed]
- Rajasekar, N.; Nath, C.; Hanif, K.; Shukla, R. Inhibitory Effect of Memantine on Streptozotocin-Induced Insulin Receptor Dysfunction, Neuroinflammation, Amyloidogenesis, and Neurotrophic Factor Decline in Astrocytes. *Mol. Neurobiol.* 2016, 53, 6730–6744. [CrossRef]
- Bordey, A.; Lyons, S.A.; Hablitz, J.J.; Sontheimer, H. Electrophysiological characteristics of reactive astrocytes in experimental cortical dysplasia. J. Neurophysiol. 2001, 85, 1719–1731. [CrossRef]
- 27. Griffith, C.M.; Xie, M.X.; Qiu, W.Y.; Sharp, A.A.; Ma, C.; Pan, A.; Yan, X.X.; Patrylo, P.R. Aberrant expression of the pore-forming K(ATP) channel subunit Kir6.2 in hippocampal reactive astrocytes in the 3xTg-AD mouse model and human Alzheimer's disease. *Neuroscience* **2016**, *336*, 81–101. [CrossRef]
- 28. Yi, M.; Yu, P.; Lu, Q.; Geller, H.M.; Yu, Z.; Chen, H. KCa3.1 constitutes a pharmacological target for astrogliosis associated with Alzheimer's disease. *Mol. Cell Neurosci.* **2016**, *76*, 21–32. [CrossRef]
- 29. Li, K.; Li, J.; Zheng, J.; Qin, S. Reactive Astrocytes in Neurodegenerative Diseases. Aging Dis. 2019, 10, 664–675. [CrossRef]
- Bosson, A.; Paumier, A.; Boisseau, S.; Jacquier-Sarlin, M.; Buisson, A.; Albrieux, M. TRPA1 channels promote astrocytic Ca²⁺ hyperactivity and synaptic dysfunction mediated by oligomeric forms of amyloid-beta peptide. *Mol. Neurodegener.* 2017, *12*, 53. [CrossRef]
- Agulhon, C.; Sun, M.Y.; Murphy, T.; Myers, T.; Lauderdale, K.; Fiacco, T.A. Calcium Signaling and Gliotransmission in Normal vs. *Reactive Astrocytes. Front. Pharmacol.* 2012, *3*, 139. [CrossRef]
- 32. Jung, H.; Lee, S.Y.; Lim, S.; Choi, H.R.; Choi, Y.; Kim, M.; Kim, S.; Lee, Y.; Han, K.H.; Chung, W.S.; et al. Anti-inflammatory clearance of amyloid-beta by a chimeric Gas6 fusion protein. *Nat. Med.* **2022**, *28*, 1802–1812. [CrossRef] [PubMed]
- Eugenin, J.; Vecchiola, A.; Murgas, P.; Arroyo, P.; Cornejo, F.; von Bernhardi, R. Expression Pattern of Scavenger Receptors and Amyloid-beta Phagocytosis of Astrocytes and Microglia in Culture are Modified by Acidosis: Implications for Alzheimer's Disease. J. Alzheimers Dis. 2016, 53, 857–873. [CrossRef] [PubMed]
- 34. Xiao, Q.; Yan, P.; Ma, X.; Liu, H.; Perez, R.; Zhu, A.; Gonzales, E.; Burchett, J.M.; Schuler, D.R.; Cirrito, J.R.; et al. Enhancing astrocytic lysosome biogenesis facilitates Abeta clearance and attenuates amyloid plaque pathogenesis. *J. Neurosci.* **2014**, *34*, 9607–9620. [CrossRef] [PubMed]
- Kimura, N.; Okabayashi, S.; Ono, F. Dynein dysfunction disrupts beta-amyloid clearance in astrocytes through endocytic disturbances. *Neuroreport* 2014, 25, 514–520. [CrossRef]
- Zadka, L.; Sochocka, M.; Hachiya, N.; Chojdak-Lukasiewicz, J.; Dziegiel, P.; Piasecki, E.; Leszek, J. Endocytosis and Alzheimer's disease. *Geroscience* 2024, 46, 71–85. [CrossRef]
- Dominguez-Prieto, M.; Velasco, A.; Tabernero, A.; Medina, J.M. Endocytosis and Transcytosis of Amyloid-beta Peptides by Astrocytes: A Possible Mechanism for Amyloid-beta Clearance in Alzheimer's Disease. J. Alzheimers Dis. 2018, 65, 1109–1124. [CrossRef]
- 38. Lambeth, T.R.; Julian, R.R. Proteolysis of Amyloid beta by Lysosomal Enzymes as a Function of Fibril Morphology. *ACS Omega* **2021**, *6*, 31520–31527. [CrossRef]
- Preston, S.D.; Steart, P.V.; Wilkinson, A.; Nicoll, J.A.; Weller, R.O. Capillary and arterial cerebral amyloid angiopathy in Alzheimer's disease: Defining the perivascular route for the elimination of amyloid beta from the human brain. *Neuropathol. Appl. Neurobiol.* 2003, 29, 106–117. [CrossRef]
- Zhang, X.; O'Callaghan, P.; Li, H.; Tan, Y.; Zhang, G.; Barash, U.; Wang, X.; Lannfelt, L.; Vlodavsky, I.; Lindahl, U.; et al. Heparanase overexpression impedes perivascular clearance of amyloid-beta from murine brain: Relevance to Alzheimer's disease. *Acta Neuropathol. Commun.* 2021, 9, 84. [CrossRef]
- Yamamoto, N.; Tokumon, T.; Obuchi, A.; Kono, M.; Saigo, K.; Tanida, M.; Ikeda-Matsuo, Y.; Sobue, K. Poly(I:C) promotes neurotoxic amyloid beta accumulation through reduced degradation by decreasing neprilysin protein levels in astrocytes. *J. Neurochem.* 2022, 163, 517–530. [CrossRef]
- Yin, K.J.; Cirrito, J.R.; Yan, P.; Hu, X.; Xiao, Q.; Pan, X.; Bateman, R.; Song, H.; Hsu, F.F.; Turk, J.; et al. Matrix metalloproteinases expressed by astrocytes mediate extracellular amyloid-beta peptide catabolism. *J. Neurosci.* 2006, 26, 10939–10948. [CrossRef] [PubMed]
- Iram, T.; Trudler, D.; Kain, D.; Kanner, S.; Galron, R.; Vassar, R.; Barzilai, A.; Blinder, P.; Fishelson, Z.; Frenkel, D. Astrocytes from old Alzheimer's disease mice are impaired in Abeta uptake and in neuroprotection. *Neurobiol. Dis.* 2016, 96, 84–94. [CrossRef] [PubMed]
- 44. Yan, L.J.; Xiao, M.; Chen, R.; Cai, Z. Metabolic Dysfunction of Astrocyte: An Initiating Factor in Beta-amyloid Pathology? *Aging Neurodegener.* **2013**, *1*, 7–14. [PubMed]
- 45. Tcw, J.; Goate, A.M. Genetics of beta-Amyloid Precursor Protein in Alzheimer's Disease. *Cold Spring Harb. Perspect. Med.* 2016, 7, a024539. [CrossRef]

- 46. Lee, H.J.; Hoe, H.S. Inhibition of CDK4/6 regulates AD pathology, neuroinflammation and cognitive function through DYRK1A/STAT3 signaling. *Pharmacol. Res.* **2023**, *190*, 106725. [CrossRef]
- 47. Pihlaja, R.; Koistinaho, J.; Malm, T.; Sikkila, H.; Vainio, S.; Koistinaho, M. Transplanted astrocytes internalize deposited betaamyloid peptides in a transgenic mouse model of Alzheimer's disease. *Glia* **2008**, *56*, 154–163. [CrossRef]
- Oksanen, M.; Hyotylainen, I.; Trontti, K.; Rolova, T.; Wojciechowski, S.; Koskuvi, M.; Viitanen, M.; Levonen, A.L.; Hovatta, I.; Roybon, L.; et al. NF-E2-related factor 2 activation boosts antioxidant defenses and ameliorates inflammatory and amyloid properties in human Presenilin-1 mutated Alzheimer's disease astrocytes. *Glia* 2020, *68*, 589–599. [CrossRef]
- 49. Etienne-Manneville, S.; Hall, A. Rho GTPases in cell biology. *Nature* 2002, 420, 629–635. [CrossRef]
- 50. Jaffe, A.B.; Hall, A. Rho GTPases: Biochemistry and biology. Annu. Rev. Cell Dev. Biol. 2005, 21, 247–269. [CrossRef]
- 51. Hannemann, S.; Madrid, R.; Stastna, J.; Kitzing, T.; Gasteier, J.; Schonichen, A.; Bouchet, J.; Jimenez, A.; Geyer, M.; Grosse, R.; et al. The Diaphanous-related Formin FHOD1 associates with ROCK1 and promotes Src-dependent plasma membrane blebbing. *J. Biol. Chem.* 2008, 283, 27891–27903. [CrossRef]
- 52. Bogucka-Janczi, K.; Harms, G.; Coissieux, M.M.; Bentires-Alj, M.; Thiede, B.; Rajalingam, K. ERK3/MAPK6 dictates CDC42/RAC1 activity and ARP2/3-dependent actin polymerization. *Elife* **2023**, *12*, e85167. [CrossRef] [PubMed]
- 53. Bonazzi, D.; Haupt, A.; Tanimoto, H.; Delacour, D.; Salort, D.; Minc, N. Actin-Based Transport Adapts Polarity Domain Size to Local Cellular Curvature. *Curr. Biol.* 2015, 25, 2677–2683. [CrossRef] [PubMed]
- 54. Ridley, A.J.; Hall, A. Signal transduction pathways regulating Rho-mediated stress fibre formation: Requirement for a tyrosine kinase. *EMBO J.* **1994**, *13*, 2600–2610. [CrossRef] [PubMed]
- 55. Watanabe, N.; Kato, T.; Fujita, A.; Ishizaki, T.; Narumiya, S. Cooperation between mDia1 and ROCK in Rho-induced actin reorganization. *Nat. Cell Biol.* **1999**, *1*, 136–143. [CrossRef]
- 56. Marlaire, S.; Dehio, C. Bartonella effector protein C mediates actin stress fiber formation via recruitment of GEF-H1 to the plasma membrane. *PLoS Pathog* **2021**, *17*, e1008548. [CrossRef]
- 57. Kozma, R.; Ahmed, S.; Best, A.; Lim, L. The Ras-related protein Cdc42Hs and bradykinin promote formation of peripheral actin microspikes and filopodia in Swiss 3T3 fibroblasts. *Mol. Cell Biol.* **1995**, *15*, 1942–1952. [CrossRef]
- 58. Grobe, H.; Wustenhagen, A.; Baarlink, C.; Grosse, R.; Grikscheit, K. A Rac1-FMNL2 signaling module affects cell-cell contact formation independent of Cdc42 and membrane protrusions. *PLoS One* **2018**, *13*, e0194716. [CrossRef]
- Kurokawa, K.; Itoh, R.E.; Yoshizaki, H.; Nakamura, Y.O.; Matsuda, M. Coactivation of Rac1 and Cdc42 at lamellipodia and membrane ruffles induced by epidermal growth factor. *Mol. Biol. Cell* 2004, *15*, 1003–1010. [CrossRef]
- 60. Nobes, C.D.; Hall, A. Rho GTPases control polarity, protrusion, and adhesion during cell movement. *J. Cell Biol.* **1999**, 144, 1235–1244. [CrossRef]
- 61. Krause, M.; Gautreau, A. Steering cell migration: Lamellipodium dynamics and the regulation of directional persistence. *Nat. Rev. Mol. Cell Biol.* **2014**, *15*, 577–590. [CrossRef]
- 62. Lamaze, C.; Chuang, T.H.; Terlecky, L.J.; Bokoch, G.M.; Schmid, S.L. Regulation of receptor-mediated endocytosis by Rho and Rac. *Nature* **1996**, *382*, 177–179. [CrossRef] [PubMed]
- Bu, W.; Lim, K.B.; Yu, Y.H.; Chou, A.M.; Sudhaharan, T.; Ahmed, S. Cdc42 interaction with N-WASP and Toca-1 regulates membrane tubulation, vesicle formation and vesicle motility: Implications for endocytosis. *PLoS ONE* 2010, 5, e12153. [CrossRef] [PubMed]
- Safavian, D.; Kim, M.S.; Xie, H.; El-Zeiry, M.; Palander, O.; Dai, L.; Collins, R.F.; Froese, C.; Shannon, R.; Nagata, K.I.; et al. Septinmediated RhoA activation engages the exocyst complex to recruit the cilium transition zone. *J. Cell Biol.* 2023, 222, e201911062. [CrossRef] [PubMed]
- Pathak, R.; Delorme-Walker, V.D.; Howell, M.C.; Anselmo, A.N.; White, M.A.; Bokoch, G.M.; Dermardirossian, C. The microtubuleassociated Rho activating factor GEF-H1 interacts with exocyst complex to regulate vesicle traffic. *Dev. Cell* 2012, 23, 397–411. [CrossRef]
- 66. Porter, A.P.; Papaioannou, A.; Malliri, A. Deregulation of Rho GTPases in cancer. Small GTPases 2016, 7, 123–138. [CrossRef]
- 67. Kalpana, G.; Figy, C.; Yeung, M.; Yeung, K.C. Reduced RhoA expression enhances breast cancer metastasis with a concomitant increase in CCR5 and CXCR4 chemokines signaling. *Sci. Rep.* **2019**, *9*, 16351. [CrossRef]
- 68. Wang, Z.; Jin, H.; Xu, R.; Mei, Q.; Fan, D. Triptolide downregulates Rac1 and the JAK/STAT3 pathway and inhibits colitis-related colon cancer progression. *Exp. Mol. Med.* **2009**, *41*, 717–727. [CrossRef]
- 69. Zhu, X.; Raina, A.K.; Boux, H.; Simmons, Z.L.; Takeda, A.; Smith, M.A. Activation of oncogenic pathways in degenerating neurons in Alzheimer disease. *Int. J. Dev. Neurosci.* 2000, *18*, 433–437. [CrossRef]
- Chan, D.; Citro, A.; Cordy, J.M.; Shen, G.C.; Wolozin, B. Rac1 protein rescues neurite retraction caused by G2019S leucine-rich repeat kinase 2 (LRRK2). J. Biol. Chem. 2011, 286, 16140–16149. [CrossRef]
- Shang, X.; Marchioni, F.; Evelyn, C.R.; Sipes, N.; Zhou, X.; Seibel, W.; Wortman, M.; Zheng, Y. Small-molecule inhibitors targeting G-protein-coupled Rho guanine nucleotide exchange factors. *Proc. Natl. Acad. Sci. USA* 2013, *110*, 3155–3160. [CrossRef]
- 72. Morstein, J.; Bowcut, V.; Fernando, M.; Yang, Y.; Zhu, L.; Jenkins, M.L.; Evans, J.T.; Guiley, K.Z.; Peacock, D.M.; Krahnke, S.; et al. Targeting Ras-, Rho-, and Rab-family GTPases via a conserved cryptic pocket. *Cell* **2024**, *187*, 6379–6392.e17. [CrossRef] [PubMed]
- 73. Endo, Y.; Even-Ram, S.; Pankov, R.; Matsumoto, K.; Yamada, K.M. Inhibition of rho GTPases by RNA interference. *Methods Enzymol.* **2006**, 406, 345–361. [CrossRef] [PubMed]

- 74. Wen, J.; Qian, C.; Pan, M.; Wang, X.; Li, Y.; Lu, Y.; Zhou, Z.; Yan, Q.; Li, L.; Liu, Z.; et al. Lentivirus-Mediated RNA Interference Targeting RhoA Slacks the Migration, Proliferation, and Myelin Formation of Schwann Cells. *Mol. Neurobiol.* 2017, 54, 1229–1239. [CrossRef] [PubMed]
- 75. Gutekunst, C.A.; Tung, J.K.; McDougal, M.E.; Gross, R.E. C3 transferase gene therapy for continuous conditional RhoA inhibition. *Neuroscience* **2016**, 339, 308–318. [CrossRef]
- 76. Threadgill, R.; Bobb, K.; Ghosh, A. Regulation of dendritic growth and remodeling by Rho, Rac, and Cdc42. *Neuron* **1997**, *19*, 625–634. [CrossRef]
- Thies, E.; Davenport, R.W. Independent roles of Rho-GTPases in growth cone and axonal behavior. J. Neurobiol. 2003, 54, 358–369.
 [CrossRef]
- Li, X.; Saint-Cyr-Proulx, E.; Aktories, K.; Lamarche-Vane, N. Rac1 and Cdc42 but not RhoA or Rho kinase activities are required for neurite outgrowth induced by the Netrin-1 receptor DCC (deleted in colorectal cancer) in N1E-115 neuroblastoma cells. J. Biol. Chem. 2002, 277, 15207–15214. [CrossRef]
- 79. Hall, A.; Lalli, G. Rho and Ras GTPases in axon growth, guidance, and branching. *Cold Spring Harb. Perspect. Biol.* **2010**, *2*, a001818. [CrossRef]
- 80. Newsome, T.P.; Schmidt, S.; Dietzl, G.; Keleman, K.; Asling, B.; Debant, A.; Dickson, B.J. Trio combines with dock to regulate Pak activity during photoreceptor axon pathfinding in Drosophila. *Cell* **2000**, *101*, 283–294. [CrossRef]
- 81. Brouns, M.R.; Matheson, S.F.; Settleman, J. p190 RhoGAP is the principal Src substrate in brain and regulates axon outgrowth, guidance and fasciculation. *Nat. Cell Biol.* **2001**, *3*, 361–367. [CrossRef]
- 82. Estrach, S.; Schmidt, S.; Diriong, S.; Penna, A.; Blangy, A.; Fort, P.; Debant, A. The Human Rho-GEF trio and its target GTPase RhoG are involved in the NGF pathway, leading to neurite outgrowth. *Curr. Biol.* **2002**, *12*, 307–312. [CrossRef] [PubMed]
- 83. Lundquist, E.A. Rac proteins and the control of axon development. Curr. Opin. Neurobiol. 2003, 13, 384–390. [CrossRef] [PubMed]
- 84. Garvalov, B.K.; Flynn, K.C.; Neukirchen, D.; Meyn, L.; Teusch, N.; Wu, X.; Brakebusch, C.; Bamburg, J.R.; Bradke, F. Cdc42 regulates cofilin during the establishment of neuronal polarity. *J. Neurosci.* 2007, 27, 13117–13129. [CrossRef] [PubMed]
- 85. Auer, M.; Hausott, B.; Klimaschewski, L. Rho GTPases as regulators of morphological neuroplasticity. *Ann. Anat.* **2011**, *193*, 259–266. [CrossRef]
- 86. Schneider, F.; Metz, I.; Rust, M.B. Regulation of actin filament assembly and disassembly in growth cone motility and axon guidance. *Brain Res. Bull.* **2023**, *192*, 21–35. [CrossRef]
- 87. Gallo, G.; Letourneau, P.C. Regulation of growth cone actin filaments by guidance cues. J. Neurobiol. 2004, 58, 92–102. [CrossRef]
- 88. Govek, E.E.; Hatten, M.E.; Van Aelst, L. The role of Rho GTPase proteins in CNS neuronal migration. *Dev. Neurobiol.* **2011**, *71*, 528–553. [CrossRef]
- Zhang, H.; Ben Zablah, Y.; Zhang, H.; Jia, Z. Rho Signaling in Synaptic Plasticity, Memory, and Brain Disorders. *Front. Cell Dev. Biol.* 2021, *9*, 729076. [CrossRef]
- Tolias, K.F.; Duman, J.G.; Um, K. Control of synapse development and plasticity by Rho GTPase regulatory proteins. *Prog. Neurobiol.* 2011, 94, 133–148. [CrossRef]
- 91. Mulherkar, S.; Tolias, K.F. RhoA-ROCK Signaling as a Therapeutic Target in Traumatic Brain Injury. Cells 2020, 9, 245. [CrossRef]
- 92. Tjalkens, R.B.; Popichak, K.A.; Kirkley, K.A. Inflammatory Activation of Microglia and Astrocytes in Manganese Neurotoxicity. *Adv. Neurobiol.* **2017**, *18*, 159–181. [CrossRef] [PubMed]
- Koch, J.C.; Tatenhorst, L.; Roser, A.E.; Saal, K.A.; Tonges, L.; Lingor, P. ROCK inhibition in models of neurodegeneration and its potential for clinical translation. *Pharmacol. Ther.* 2018, 189, 1–21. [CrossRef] [PubMed]
- 94. Sami, A.; Selzer, M.E.; Li, S. Advances in the Signaling Pathways Downstream of Glial-Scar Axon Growth Inhibitors. *Front. Cell. Neurosci.* **2020**, *14*, 174. [CrossRef] [PubMed]
- 95. Fujita, Y.; Yamashita, T. Axon growth inhibition by RhoA/ROCK in the central nervous system. *Front. Neurosci.* **2014**, *8*, 338. [CrossRef]
- Madura, T.; Yamashita, T.; Kubo, T.; Fujitani, M.; Hosokawa, K.; Tohyama, M. Activation of Rho in the injured axons following spinal cord injury. *EMBO Rep.* 2004, 5, 412–417. [CrossRef]
- Zhang, Y.; Li, K.; Wang, X.; Ding, Y.; Ren, Z.; Fang, J.; Sun, T.; Guo, Y.; Chen, Z.; Wen, J. CSE-Derived H₂S Inhibits Reactive Astrocytes Proliferation and Promotes Neural Functional Recovery after Cerebral Ischemia/Reperfusion Injury in Mice Via Inhibition of RhoA/ROCK₂ Pathway. ACS Chem. Neurosci. 2021, 12, 2580–2590. [CrossRef]
- Holtje, M.; Hoffmann, A.; Hofmann, F.; Mucke, C.; Grosse, G.; Van Rooijen, N.; Kettenmann, H.; Just, I.; Ahnert-Hilger, G. Role of Rho GTPase in astrocyte morphology and migratory response during in vitro wound healing. *J. Neurochem.* 2005, 95, 1237–1248. [CrossRef]
- 99. Etienne-Manneville, S.; Hall, A. Integrin-mediated activation of Cdc42 controls cell polarity in migrating astrocytes through PKCzeta. *Cell* **2001**, *106*, 489–498. [CrossRef]
- 100. Daub, H.; Gevaert, K.; Vandekerckhove, J.; Sobel, A.; Hall, A. Rac/Cdc42 and p65PAK regulate the microtubule-destabilizing protein stathmin through phosphorylation at serine 16. *J. Biol. Chem.* **2001**, *276*, 1677–1680. [CrossRef]
- 101. Ory, S.; Gasman, S. Rho GTPases and exocytosis: What are the molecular links? Semin. Cell Dev. Biol. 2011, 22, 27-32. [CrossRef]
- Wu, H.; Rossi, G.; Brennwald, P. The ghost in the machine: Small GTPases as spatial regulators of exocytosis. *Trends Cell Biol.* 2008, 18, 397–404. [CrossRef] [PubMed]

- 103. Zhou, Z.; Zhan, J.; Cai, Q.; Xu, F.; Chai, R.; Lam, K.; Luan, Z.; Zhou, G.; Tsang, S.; Kipp, M.; et al. The Water Transport System in Astrocytes-Aquaporins. *Cells* 2022, 11, 2564. [CrossRef] [PubMed]
- 104. van Putten, M.; Fahlke, C.; Kafitz, K.W.; Hofmeijer, J.; Rose, C.R. Dysregulation of Astrocyte Ion Homeostasis and Its Relevance for Stroke-Induced Brain Damage. *Int. J. Mol. Sci.* 2021, 22, 5679. [CrossRef] [PubMed]
- 105. Jin, L.M. Rock 'n' Rho: Regulation of ion channels. Am. J. Physiol. Heart Circ. Physiol. 2009, 296, H908–H909. [CrossRef]
- 106. Pochynyuk, O.; Stockand, J.D.; Staruschenko, A. Ion channel regulation by Ras, Rho, and Rab small GTPases. *Exp. Biol. Med.* **2007**, 232, 1258–1265. [CrossRef]
- 107. Tura, A.; Schuettauf, F.; Monnier, P.P.; Bartz-Schmidt, K.U.; Henke-Fahle, S. Efficacy of Rho-kinase inhibition in promoting cell survival and reducing reactive gliosis in the rodent retina. *Investig. Ophthalmol. Vis. Sci.* 2009, *50*, 452–461. [CrossRef]
- 108. Zhang, H.; Li, Y.; Yu, J.; Guo, M.; Meng, J.; Liu, C.; Xie, Y.; Feng, L.; Xiao, B.; Ma, C. Rho kinase inhibitor fasudil regulates microglia polarization and function. *Neuroimmunomodulation* **2013**, *20*, 313–322. [CrossRef]
- Barcia, C.; Ros, C.M.; Annese, V.; Carrillo-de Sauvage, M.A.; Ros-Bernal, F.; Gomez, A.; Yuste, J.E.; Campuzano, C.M.; de Pablos, V.; Fernandez-Villalba, E.; et al. ROCK/Cdc42-mediated microglial motility and gliapse formation lead to phagocytosis of degenerating dopaminergic neurons in vivo. *Sci. Rep.* 2012, *2*, 809. [CrossRef]
- Ding, J.; Yu, J.Z.; Li, Q.Y.; Wang, X.; Lu, C.Z.; Xiao, B.G. Rho kinase inhibitor Fasudil induces neuroprotection and neurogenesis partially through astrocyte-derived G-CSF. *Brain Behav. Immun.* 2009, 23, 1083–1088. [CrossRef]
- 111. Alkam, D.; Feldman, E.Z.; Singh, A.; Kiaei, M. Profilin1 biology and its mutation, actin(g) in disease. *Cell Mol. Life Sci.* 2017, 74, 967–981. [CrossRef]
- 112. Lu, E.; Wang, Q.; Li, S.; Chen, C.; Wu, W.; Xu, Y.X.Z.; Zhou, P.; Tu, W.; Lou, X.; Rao, G.; et al. Profilin 1 knockdown prevents ischemic brain damage by promoting M2 microglial polarization associated with the RhoA/ROCK pathway. *J. Neurosci. Res.* 2020, *98*, 1198–1212. [CrossRef] [PubMed]
- Zhang, Y.; Miao, L.; Peng, Q.; Fan, X.; Song, W.; Yang, B.; Zhang, P.; Liu, G.; Liu, J. Parthenolide modulates cerebral ischemiainduced microglial polarization and alleviates neuroinflammatory injury via the RhoA/ROCK pathway. *Phytomedicine* 2022, 105, 154373. [CrossRef] [PubMed]
- 114. John, G.R.; Chen, L.; Rivieccio, M.A.; Melendez-Vasquez, C.V.; Hartley, A.; Brosnan, C.F. Interleukin-1beta induces a reactive astroglial phenotype via deactivation of the Rho GTPase-Rock axis. *J. Neurosci.* **2004**, *24*, 2837–2845. [CrossRef] [PubMed]
- 115. Renault-Mihara, F.; Mukaino, M.; Shinozaki, M.; Kumamaru, H.; Kawase, S.; Baudoux, M.; Ishibashi, T.; Kawabata, S.; Nishiyama, Y.; Sugai, K.; et al. Regulation of RhoA by STAT3 coordinates glial scar formation. *J. Cell Biol.* **2017**, *216*, 2533–2550. [CrossRef]
- 116. Haditsch, U.; Anderson, M.P.; Freewoman, J.; Cord, B.; Babu, H.; Brakebusch, C.; Palmer, T.D. Neuronal Rac1 is required for learning-evoked neurogenesis. *J. Neurosci.* 2013, *33*, 12229–12241. [CrossRef]
- Wang, P.L.; Niidome, T.; Akaike, A.; Kihara, T.; Sugimoto, H. Rac1 inhibition negatively regulates transcriptional activity of the amyloid precursor protein gene. J. Neurosci. Res. 2009, 87, 2105–2114. [CrossRef]
- Boo, J.H.; Sohn, J.H.; Kim, J.E.; Song, H.; Mook-Jung, I. Rac1 changes the substrate specificity of gamma-secretase between amyloid precursor protein and Notch1. *Biochem. Biophys. Res. Commun.* 2008, 372, 913–917. [CrossRef]
- 119. Guiler, W.; Koehler, A.; Boykin, C.; Lu, Q. Pharmacological Modulators of Small GTPases of Rho Family in Neurodegenerative Diseases. *Front. Cell Neurosci.* 2021, 15, 661612. [CrossRef]
- 120. Aguilar, B.J.; Zhu, Y.; Lu, Q. Rho GTPases as therapeutic targets in Alzheimer's disease. *Alzheimers Res. Ther.* 2017, 9, 97. [CrossRef]
- 121. Kitamura, Y.; Shibagaki, K.; Takata, K.; Tsuchiya, D.; Taniguchi, T.; Gebicke-Haerter, P.J.; Miki, H.; Takenawa, T.; Shimohama, S. Involvement of Wiskott-Aldrich syndrome protein family verprolin-homologous protein (WAVE) and Rac1 in the phagocytosis of amyloid-beta(1–42) in rat microglia. *J. Pharmacol. Sci.* 2003, *92*, 115–123. [CrossRef]
- 122. Thal, D.R. The role of astrocytes in amyloid beta-protein toxicity and clearance. *Exp. Neurol.* 2012, 236, 1–5. [CrossRef] [PubMed]
- 123. Bell, R.D.; Sagare, A.P.; Friedman, A.E.; Bedi, G.S.; Holtzman, D.M.; Deane, R.; Zlokovic, B.V. Transport pathways for clearance of human Alzheimer's amyloid beta-peptide and apolipoproteins E and J in the mouse central nervous system. *J. Cereb. Blood Flow. Metab.* 2007, 27, 909–918. [CrossRef] [PubMed]
- 124. Shibata, M.; Yamada, S.; Kumar, S.R.; Calero, M.; Bading, J.; Frangione, B.; Holtzman, D.M.; Miller, C.A.; Strickland, D.K.; Ghiso, J.; et al. Clearance of Alzheimer's amyloid-ss(1–40) peptide from brain by LDL receptor-related protein-1 at the blood-brain barrier. *J. Clin. Investig.* 2000, 106, 1489–1499. [CrossRef] [PubMed]
- 125. Abe, K.; Misawa, M. The extracellular signal-regulated kinase cascade suppresses amyloid beta protein-induced promotion of glutamate clearance in cultured rat cortical astrocytes. *Brain Res.* 2003, 979, 179–187. [CrossRef]
- Nalivaeva, N.N.; Beckett, C.; Belyaev, N.D.; Turner, A.J. Are amyloid-degrading enzymes viable therapeutic targets in Alzheimer's disease? J. Neurochem. 2012, 120 (Suppl. S1), 167–185. [CrossRef]
- 127. Turner, A.J.; Nalivaeva, N.N. New insights into the roles of metalloproteinases in neurodegeneration and neuroprotection. *Int. Rev. Neurobiol.* 2007, *82*, 113–135. [CrossRef]
- 128. Fukami, S.; Watanabe, K.; Iwata, N.; Haraoka, J.; Lu, B.; Gerard, N.P.; Gerard, C.; Fraser, P.; Westaway, D.; St George-Hyslop, P.; et al. Abeta-degrading endopeptidase, neprilysin, in mouse brain: Synaptic and axonal localization inversely correlating with Abeta pathology. *Neurosci. Res.* **2002**, *43*, 39–56. [CrossRef]
- 129. Yasojima, K.; Akiyama, H.; McGeer, E.G.; McGeer, P.L. Reduced neprilysin in high plaque areas of Alzheimer brain: A possible relationship to deficient degradation of beta-amyloid peptide. *Neurosci. Lett.* **2001**, 297, 97–100. [CrossRef]

- Dorfman, V.B.; Pasquini, L.; Riudavets, M.; Lopez-Costa, J.J.; Villegas, A.; Troncoso, J.C.; Lopera, F.; Castano, E.M.; Morelli, L. Differential cerebral deposition of IDE and NEP in sporadic and familial Alzheimer's disease. *Neurobiol. Aging* 2010, *31*, 1743–1757. [CrossRef]
- Yamamoto, N.; Tanida, M.; Ono, Y.; Kasahara, R.; Fujii, Y.; Ohora, K.; Suzuki, K.; Sobue, K. Leptin inhibits amyloid beta-protein degradation through decrease of neprilysin expression in primary cultured astrocytes. *Biochem. Biophys. Res. Commun.* 2014, 445, 214–217. [CrossRef]
- 132. Rofo, F.; Ugur Yilmaz, C.; Metzendorf, N.; Gustavsson, T.; Beretta, C.; Erlandsson, A.; Sehlin, D.; Syvanen, S.; Nilsson, P.; Hultqvist, G. Enhanced neprilysin-mediated degradation of hippocampal Abeta42 with a somatostatin peptide that enters the brain. *Theranostics* **2021**, *11*, 789–804. [CrossRef] [PubMed]
- Yamamoto, N.; Arima, H.; Naruse, K.; Kasahara, R.; Taniura, H.; Hirate, H.; Sugiura, T.; Suzuki, K.; Sobue, K. Ketamine reduces amyloid beta-protein degradation by suppressing neprilysin expression in primary cultured astrocytes. *Neurosci. Lett.* 2013, 545, 54–58. [CrossRef] [PubMed]
- 134. Eckman, E.A.; Adams, S.K.; Troendle, F.J.; Stodola, B.A.; Kahn, M.A.; Fauq, A.H.; Xiao, H.D.; Bernstein, K.E.; Eckman, C.B. Regulation of steady-state beta-amyloid levels in the brain by neprilysin and endothelin-converting enzyme but not angiotensinconverting enzyme. J. Biol. Chem. 2006, 281, 30471–30478. [CrossRef] [PubMed]
- 135. Eckman, E.A.; Reed, D.K.; Eckman, C.B. Degradation of the Alzheimer's amyloid beta peptide by endothelin-converting enzyme. *J. Biol. Chem.* **2001**, *276*, 24540–24548. [CrossRef]
- 136. Iwata, N.; Tsubuki, S.; Takaki, Y.; Shirotani, K.; Lu, B.; Gerard, N.P.; Gerard, C.; Hama, E.; Lee, H.J.; Saido, T.C. Metabolic regulation of brain Abeta by neprilysin. *Science* **2001**, *292*, 1550–1552. [CrossRef]
- 137. Padilla, B.E.; Cottrell, G.S.; Roosterman, D.; Pikios, S.; Muller, L.; Steinhoff, M.; Bunnett, N.W. Endothelin-converting enzyme-1 regulates endosomal sorting of calcitonin receptor-like receptor and beta-arrestins. *J. Cell Biol.* 2007, *179*, 981–997. [CrossRef]
- 138. Palmer, J.C.; Kehoe, P.G.; Love, S. Endothelin-converting enzyme-1 in Alzheimer's disease and vascular dementia. *Neuropathol. Appl. Neurobiol.* **2010**, *36*, 487–497. [CrossRef]
- 139. Eckman, E.A.; Watson, M.; Marlow, L.; Sambamurti, K.; Eckman, C.B. Alzheimer's disease beta-amyloid peptide is increased in mice deficient in endothelin-converting enzyme. *J. Biol. Chem.* 2003, 278, 2081–2084. [CrossRef]
- Nortley, R.; Korte, N.; Izquierdo, P.; Hirunpattarasilp, C.; Mishra, A.; Jaunmuktane, Z.; Kyrargyri, V.; Pfeiffer, T.; Khennouf, L.; Madry, C.; et al. Amyloid beta oligomers constrict human capillaries in Alzheimer's disease via signaling to pericytes. *Science* 2019, 365, eaav9518. [CrossRef]
- 141. Guo, Y.; Chung, S.K.; Siu, C.W.; Kwan, S.C.; Ho, P.W.; Yeung, P.K.; Chan, K.H. Endothelin-1 overexpression exacerbate experimental allergic encephalomyelitis. *J. Neuroimmunol.* **2014**, 276, 64–70. [CrossRef]
- 142. D'Haeseleer, M.; Beelen, R.; Fierens, Y.; Cambron, M.; Vanbinst, A.M.; Verborgh, C.; Demey, J.; De Keyser, J. Cerebral hypoperfusion in multiple sclerosis is reversible and mediated by endothelin-1. *Proc. Natl. Acad. Sci. USA* 2013, *110*, 5654–5658. [CrossRef] [PubMed]
- 143. Hostenbach, S.; D'Haeseleer, M.; Kooijman, R.; De Keyser, J. Modulation of Cytokine-Induced Astrocytic Endothelin-1 Production as a Possible New Approach to the Treatment of Multiple Sclerosis. *Front. Pharmacol.* **2019**, *10*, 1491. [CrossRef] [PubMed]
- 144. Pacheco-Quinto, J.; Eckman, C.B.; Eckman, E.A. Major amyloid-beta-degrading enzymes, endothelin-converting enzyme-2 and neprilysin, are expressed by distinct populations of GABAergic interneurons in hippocampus and neocortex. *Neurobiol. Aging* 2016, 48, 83–92. [CrossRef] [PubMed]
- 145. Palmer, J.C.; Baig, S.; Kehoe, P.G.; Love, S. Endothelin-converting enzyme-2 is increased in Alzheimer's disease and up-regulated by Abeta. *Am. J. Pathol.* **2009**, *175*, 262–270. [CrossRef]
- 146. Rodriguiz, R.M.; Gadnidze, K.; Ragnauth, A.; Dorr, N.; Yanagisawa, M.; Wetsel, W.C.; Devi, L.A. Animals lacking endothelinconverting enzyme-2 are deficient in learning and memory. *Genes. Brain Behav.* 2008, 7, 418–426. [CrossRef]
- 147. Chen, S.; Guo, D.; Zhu, Y.; Xiao, S.; Xie, J.; Zhang, Z.; Hu, Y.; Huang, J.; Ma, X.; Ning, Z.; et al. Amyloid beta oligomer induces cerebral vasculopathy via pericyte-mediated endothelial dysfunction. *Alzheimers Res. Ther.* **2024**, *16*, 56. [CrossRef]
- 148. Ganz, T.; Fainstein, N.; Ben-Hur, T. When the infectious environment meets the AD brain. *Mol. Neurodegener.* **2022**, *17*, 53. [CrossRef]
- 149. Botella Lucena, P.; Heneka, M.T. Inflammatory aspects of Alzheimer's disease. Acta Neuropathol. 2024, 148, 31. [CrossRef]
- 150. Shi, M.; Chu, F.; Zhu, F.; Zhu, J. Peripheral blood amyloid-beta involved in the pathogenesis of Alzheimer's disease via impacting on peripheral innate immune cells. *J. Neuroinflammation* **2024**, *21*, 5. [CrossRef]
- 151. Freeman, B.D.; Machado, F.S.; Tanowitz, H.B.; Desruisseaux, M.S. Endothelin-1 and its role in the pathogenesis of infectious diseases. *Life Sci.* 2014, *118*, 110–119. [CrossRef]
- 152. Takano, T.; Oberheim, N.; Cotrina, M.L.; Nedergaard, M. Astrocytes and ischemic injury. *Stroke* 2009, 40, S8–S12. [CrossRef] [PubMed]
- 153. Hostenbach, S.; D'Haeseleer, M.; Kooijman, R.; De Keyser, J. The pathophysiological role of astrocytic endothelin-1. *Prog. Neurobiol.* **2016**, 144, 88–102. [CrossRef]
- 154. Eto, M.; Barandier, C.; Rathgeb, L.; Kozai, T.; Joch, H.; Yang, Z.; Luscher, T.F. Thrombin suppresses endothelial nitric oxide synthase and upregulates endothelin-converting enzyme-1 expression by distinct pathways: Role of Rho/ROCK and mitogen-activated protein kinase. *Circ. Res.* 2001, *89*, 583–590. [CrossRef] [PubMed]

- 155. Minamino, T.; Kurihara, H.; Takahashi, M.; Shimada, K.; Maemura, K.; Oda, H.; Ishikawa, T.; Uchiyama, T.; Tanzawa, K.; Yazaki, Y. Endothelin-converting enzyme expression in the rat vascular injury model and human coronary atherosclerosis. *Circulation* 1997, 95, 221–230. [CrossRef] [PubMed]
- 156. Lau, C.L.; Perreau, V.M.; Chen, M.J.; Cate, H.S.; Merlo, D.; Cheung, N.S.; O'Shea, R.D.; Beart, P.M. Transcriptomic profiling of astrocytes treated with the Rho kinase inhibitor fasudil reveals cytoskeletal and pro-survival responses. *J. Cell Physiol.* 2012, 227, 1199–1211. [CrossRef] [PubMed]
- 157. Tonges, L.; Gunther, R.; Suhr, M.; Jansen, J.; Balck, A.; Saal, K.A.; Barski, E.; Nientied, T.; Gotz, A.A.; Koch, J.C.; et al. Rho kinase inhibition modulates microglia activation and improves survival in a model of amyotrophic lateral sclerosis. *Glia* **2014**, *62*, 217–232. [CrossRef]
- 158. Kimura, T.; Horikoshi, Y.; Kuriyagawa, C.; Niiyama, Y. Rho/ROCK Pathway and Noncoding RNAs: Implications in Ischemic Stroke and Spinal Cord Injury. *Int. J. Mol. Sci.* 2021, 22, 11573. [CrossRef]
- 159. Collu, R.; Yin, Z.; Giunti, E.; Daley, S.; Chen, M.; Morin, P.; Killick, R.; Wong, S.T.C.; Xia, W. Effect of the ROCK inhibitor fasudil on the brain proteomic profile in the tau transgenic mouse model of Alzheimer's disease. *Front. Aging Neurosci.* **2024**, *16*, 1323563. [CrossRef]
- 160. Hamano, T.; Shirafuji, N.; Yen, S.H.; Yoshida, H.; Kanaan, N.M.; Hayashi, K.; Ikawa, M.; Yamamura, O.; Fujita, Y.; Kuriyama, M.; et al. Rho-kinase ROCK inhibitors reduce oligomeric tau protein. *Neurobiol. Aging* **2020**, *89*, 41–54. [CrossRef]
- 161. Saray, H.; Suer, C.; Kosar, B.; Tan, B.; Dursun, N. Rho-associated kinases contribute to the regulation of tau phosphorylation and amyloid metabolism during neuronal plasticity. *Pharmacol. Rep.* **2021**, *73*, 1303–1314. [CrossRef]
- 162. Wen, J.Y.; Gao, S.S.; Chen, F.L.; Chen, S.; Wang, M.; Chen, Z.W. Role of CSE-Produced H₂S on Cerebrovascular Relaxation via RhoA-ROCK Inhibition and Cerebral Ischemia-Reperfusion Injury in Mice. ACS Chem. Neurosci. 2019, 10, 1565–1574. [CrossRef] [PubMed]
- Barker, R.; Ashby, E.L.; Wellington, D.; Barrow, V.M.; Palmer, J.C.; Kehoe, P.G.; Esiri, M.M.; Love, S. Pathophysiology of white matter perfusion in Alzheimer's disease and vascular dementia. *Brain* 2014, 137, 1524–1532. [CrossRef] [PubMed]
- 164. Zimmer, S.; Goody, P.R.; Oelze, M.; Ghanem, A.; Mueller, C.F.; Laufs, U.; Daiber, A.; Jansen, F.; Nickenig, G.; Wassmann, S. Inhibition of Rac1 GTPase Decreases Vascular Oxidative Stress, Improves Endothelial Function, and Attenuates Atherosclerosis Development in Mice. *Front. Cardiovasc. Med.* 2021, *8*, 680775. [CrossRef] [PubMed]
- 165. Laufs, U.; Adam, O.; Strehlow, K.; Wassmann, S.; Konkol, C.; Laufs, K.; Schmidt, W.; Bohm, M.; Nickenig, G. Down-regulation of Rac-1 GTPase by Estrogen. J. Biol. Chem. 2003, 278, 5956–5962. [CrossRef] [PubMed]
- 166. Friedrich, E.B.; Clever, Y.P.; Wassmann, S.; Hess, C.; Nickenig, G. 17Beta-estradiol inhibits monocyte adhesion via down-regulation of Rac1 GTPase. J. Mol. Cell Cardiol. 2006, 40, 87–95. [CrossRef]
- 167. Hu, Q.; Zheng, G.; Zweier, J.L.; Deshpande, S.; Irani, K.; Ziegelstein, R.C. NADPH oxidase activation increases the sensitivity of intracellular Ca²⁺ stores to inositol 1,4,5-trisphosphate in human endothelial cells. *J. Biol. Chem.* 2000, 275, 15749–15757. [CrossRef]
- 168. Schiattarella, G.G.; Carrizzo, A.; Ilardi, F.; Damato, A.; Ambrosio, M.; Madonna, M.; Trimarco, V.; Marino, M.; De Angelis, E.; Settembrini, S.; et al. Rac1 Modulates Endothelial Function and Platelet Aggregation in Diabetes Mellitus. *J. Am. Heart Assoc.* 2018, 7, e007322. [CrossRef]
- 169. Arfian, N.; Suzuki, Y.; Hartopo, A.B.; Anggorowati, N.; Nugrahaningsih, D.A.A.; Emoto, N. Endothelin converting enzyme-1 (ECE-1) deletion in association with Endothelin-1 downregulation ameliorates kidney fibrosis in mice. *Life Sci.* 2020, 258, 118223. [CrossRef]
- 170. Cattaruzza, F.; Cottrell, G.S.; Vaksman, N.; Bunnett, N.W. Endothelin-converting enzyme 1 promotes re-sensitization of neurokinin 1 receptor-dependent neurogenic inflammation. *Br. J. Pharmacol.* **2009**, *156*, 730–739. [CrossRef]
- 171. Roosterman, D.; Cottrell, G.S.; Padilla, B.E.; Muller, L.; Eckman, C.B.; Bunnett, N.W.; Steinhoff, M. Endothelin-converting enzyme 1 degrades neuropeptides in endosomes to control receptor recycling. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 11838–11843. [CrossRef]
- 172. Zeug, A.; Muller, F.E.; Anders, S.; Herde, M.K.; Minge, D.; Ponimaskin, E.; Henneberger, C. Control of astrocyte morphology by Rho GTPases. *Brain Res. Bull.* 2018, 136, 44–53. [CrossRef] [PubMed]
- 173. Silva, A.I.; Socodato, R.; Pinto, C.; Terceiro, A.F.; Canedo, T.; Relvas, J.B.; Saraiva, M.; Summavielle, T. IL-10 and Cdc42 modulate astrocyte-mediated microglia activation in methamphetamine-induced neuroinflammation. *Glia* 2024, 72, 1501–1517. [CrossRef] [PubMed]
- 174. Barone, F.C.; Globus, M.Y.; Price, W.J.; White, R.F.; Storer, B.L.; Feuerstein, G.Z.; Busto, R.; Ohlstein, E.H. Endothelin levels increase in rat focal and global ischemia. *J. Cereb. Blood Flow. Metab.* **1994**, *14*, 337–342. [CrossRef] [PubMed]
- 175. Michinaga, S.; Inoue, A.; Yamamoto, H.; Ryu, R.; Inoue, A.; Mizuguchi, H.; Koyama, Y. Endothelin receptor antagonists alleviate blood-brain barrier disruption and cerebral edema in a mouse model of traumatic brain injury: A comparison between bosentan and ambrisentan. *Neuropharmacology* **2020**, *175*, 108182. [CrossRef]
- 176. Lampl, Y.; Fleminger, G.; Gilad, R.; Galron, R.; Sarova-Pinhas, I.; Sokolovsky, M. Endothelin in cerebrospinal fluid and plasma of patients in the early stage of ischemic stroke. *Stroke* **1997**, *28*, 1951–1955. [CrossRef]
- 177. Kobayashi, H.; Ide, H.; Ishii, H.; Kabuto, M.; Handa, Y.; Kubota, T. Endothelin-1 levels in plasma and cerebrospinal fluidfollowing subarachnoid haemorrhage. *J. Clin. Neurosci.* **1995**, *2*, 252–256. [CrossRef]

- 178. Maier, B.; Lehnert, M.; Laurer, H.L.; Marzi, I. Biphasic elevation in cerebrospinal fluid and plasma concentrations of endothelin 1 after traumatic brain injury in human patients. *Shock* **2007**, *27*, 610–614. [CrossRef]
- 179. Koyama, Y. Endothelin ET_B Receptor-Mediated Astrocytic Activation: Pathological Roles in Brain Disorders. *Int. J. Mol. Sci.* 2021, 22, 4333. [CrossRef]
- 180. Koyama, Y. Endothelin systems in the brain: Involvement in pathophysiological responses of damaged nerve tissues. *Biomol. Concepts* **2013**, *4*, 335–347. [CrossRef]
- 181. Skopal, J.; Turbucz, P.; Vastag, M.; Bori, Z.; Pek, M.; deChatel, R.; Nagy, Z.; Toth, M.; Karadi, I. Regulation of endothelin release from human brain microvessel endothelial cells. *J. Cardiovasc. Pharmacol.* **1998**, *31* (Suppl. S1), S370–S372. [CrossRef]
- Michinaga, S.; Hishinuma, S.; Koyama, Y. Roles of Astrocytic Endothelin ET_B Receptor in Traumatic Brain Injury. *Cells* 2023, 12, 719. [CrossRef] [PubMed]
- Prasanna, G.; Krishnamoorthy, R.; Yorio, T. Endothelin, astrocytes and glaucoma. *Exp. Eye Res.* 2011, 93, 170–177. [CrossRef]
 [PubMed]
- Didier, N.; Romero, I.A.; Creminon, C.; Wijkhuisen, A.; Grassi, J.; Mabondzo, A. Secretion of interleukin-1beta by astrocytes mediates endothelin-1 and tumour necrosis factor-alpha effects on human brain microvascular endothelial cell permeability. *J. Neurochem.* 2003, *86*, 246–254. [CrossRef] [PubMed]
- 185. Robel, S.; Bardehle, S.; Lepier, A.; Brakebusch, C.; Gotz, M. Genetic deletion of cdc42 reveals a crucial role for astrocyte recruitment to the injury site in vitro and in vivo. *J. Neurosci.* 2011, *31*, 12471–12482. [CrossRef]
- Yuan, Y.; Liu, H.; Dai, Z.; He, C.; Qin, S.; Su, Z. From Physiology to Pathology of Astrocytes: Highlighting Their Potential as Therapeutic Targets for CNS Injury. *Neurosci. Bull.* 2024. [CrossRef] [PubMed]
- 187. Chiareli, R.A.; Carvalho, G.A.; Marques, B.L.; Mota, L.S.; Oliveira-Lima, O.C.; Gomes, R.M.; Birbrair, A.; Gomez, R.S.; Simao, F.; Klempin, F.; et al. The Role of Astrocytes in the Neurorepair Process. *Front. Cell Dev. Biol.* **2021**, *9*, 665795. [CrossRef]
- Liu, S.; Li, Q.; Na, Q.; Liu, C. Endothelin-1 stimulates human trophoblast cell migration through Cdc42 activation. *Placenta* 2012, 33, 712–716. [CrossRef]

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