



Article

Increasing Performance of Spiral-Wound Modules (SWMs) by Improving Stability against Axial Pressure Drop and Utilising Pulsed Flow

Christian Kürzl ^{1,2,*} , Martin Hartinger ¹, Patrick Ong ¹, Roland Schopf ¹ , Simon Schiffer ¹ and Ulrich Kulozik ¹

¹ Food and Bioprocess Engineering, TUM School of Life Sciences, Technical University of Munich, Weihenstephaner Berg 1, 85354 Freising, Germany

² Food Process Engineering, TUM School of Life Sciences, Technical University of Munich, Weihenstephaner Berg 1, 85354 Freising, Germany

* Correspondence: christian.kuerzl@tum.de; Tel.: +49-8-161-713-481

Abstract: Spacer-induced flow shadows and limited mechanical stability due to module construction and geometry are the main obstacles to improving the filtration performance and cleanability of microfiltration spiral-wound membranes (SWMs), applied to milk protein fractionation in this study. The goal of this study was first to improve filtration performance and cleanability by utilising pulsed flow in a modified pilot-scale filtration plant. The second goal was to enhance membrane stability against module deformation by flow-induced friction in the axial direction (“membrane telescoping”). This was accomplished by stabilising membrane layers, including spacers, at the membrane inlet by glue connections. Pulsed flow characteristics similar to those reported in previous lab-scale studies could be achieved by establishing an on/off bypass around the membrane module, thus enabling a high-frequency flow variation. Pulsed flow significantly increased filtration performance (target protein mass flow into the permeate increased by 26%) and cleaning success (protein removal increased by 28%). Furthermore, adding feed-side glue connections increased the mechanical membrane stability in terms of allowed volume throughput by $\geq 100\%$ compared to unmodified modules, thus allowing operation with higher axial pressure drops, flow velocities and pulsation amplitudes.

Keywords: pulsed flow; module stability; axial pressure loss; telescoping; membrane performance



Citation: Kürzl, C.; Hartinger, M.; Ong, P.; Schopf, R.; Schiffer, S.; Kulozik, U. Increasing Performance of Spiral-Wound Modules (SWMs) by Improving Stability against Axial Pressure Drop and Utilising Pulsed Flow. *Membranes* **2023**, *13*, 791. <https://doi.org/10.3390/membranes13090791>

Academic Editor: Sanghyun Jeong

Received: 11 August 2023

Revised: 7 September 2023

Accepted: 11 September 2023

Published: 12 September 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

The membrane-based separation of a feed solution into the permeable components (permeate) and the retained components (retentate) is widely applied across various industries. Within this process, the main challenge is controlling deposit formation, i.e., fouling, which results from the accumulation of retained feed components on and in the membrane structure [1–5]. With deposits acting as a secondary selective layer, this causes separation efficiency in terms of flux and protein permeation decreasing gradually during filtration [6–8]. Additionally, frequent cleaning cycles are required to maintain membrane performance and product quality due to the feed- and temperature-dependent occurrence and progression of biofouling [9–13]. For dairy applications, one important example is the fractionation of skim milk protein into its major protein components: whey proteins (particle diameter $d_p = 3\text{--}6\text{ nm}$ [14]) and casein micelles ($d_p = 20\text{--}300\text{ nm}$ [14]) via microfiltration (MF; nominal pore size = $0.1\text{--}0.3\text{ }\mu\text{m}$). This poses a particular challenge for the application of fouling and cleaning as casein micelles can form highly compressible and cross-linked gel layers at high concentrations and high-pressure conditions [15–19]. Deposit formation cannot be reverted by pressure release [20], and accumulated protein can only be incompletely removed by rinsing steps [1,2,21].

Besides feed composition, the membrane performance largely depends on processing conditions. Up to limiting conditions, transmembrane pressure (TMP) increases can be used

to achieve gains in flux. However, beyond limiting conditions, further increases in TMP do not cause further flux increases but solely result in additional, partly irreversible [20] deposit compaction and fouling [7]. Another option to enhance membrane performance is by increasing the flow velocity v , and thus the wall shear stress τ_w , as this reduces fouling [22–26]. Nonetheless, the maximum applicable crossflow velocities in SWMs are limited by geometry-related or constructional aspects, thus also limiting fouling control and cleanability.

Regarding membrane geometry, fouling depends on the membrane length and varies along the membrane [27]. Due to friction, an axial pressure loss (Δp_L) over the membrane length is induced. This causes a decrease in TMP and thus in the fouling intensity from the module inlet towards the module outlet [27]. The main industrially used module configurations are ceramic tubular membranes (CTMs), hollow-fibre membranes (HFMs) and SWMs, which all have their typical pros and cons. Compared to CTMs and HFMs, SWMs offer the highest packing density, i.e., active membrane area per module, and thus the highest whey protein mass flow per module [28]. On the contrary, SWMs suffer from flow shadows behind spacer filaments [29–37] and, therefore, limited cleanability [29] and mechanical stability. In SWMs, the membrane permeate pockets are formed by glueing together individual membrane sheets, which are then wrapped around a central permeate collection tube and fixed by an outer hull. Thus, the SWMs' stability mainly depends on the stability of glued bond joints and the friction between the membrane sheets, which results from the strength of the wrapping.

The stability of bond joints depends on several construction-related aspects, such as the glue composition [38], the design [39] and overlapping length of connections [40] and the glue layer thickness [38]. Apart from that, process-related aspects, such as the speed of stress application, the intensity and kind of stress [38], the process temperature [38] and the duration of stress application, have significant effects [38,41]. Due to membrane pockets formed by glueing together membrane sheets, the permeate side is susceptible to failure, especially by negative TMP, which stresses bond joints via peeling and can cause tearing of the membrane pockets. To avoid this, manufacturers usually limit the maximum negative TMP to around 0.3 bar.

A more common failure mechanism in SWMs is telescoping, which describes an axial displacement of the membrane pockets caused by frictional losses along the module (Δp_L) acting on the membrane envelope. Hence, the strength of the wrapping determines the amount of friction between membrane sheets and thus its stability against axial displacement. However, besides a lower risk of telescoping, stronger wrapping can also press spacers into the membrane surface, thus reducing the active membrane area and even disrupting the selective layer [42]. The trade-off between stability against axial deformation and membrane performance led to manufacturers limiting the friction-related axial pressure drop Δp_L to 1.3 bar m^{-1} despite the commonly added stability support against axial displacement via anti-telescoping devices (ATD). With Δp_L and thus v being limited, this significantly restricts options to control deposit formation, e.g., by conventionally established higher crossflow velocities in SWMs compared to the other module types described above.

Several process-oriented approaches trying to increase membrane performance have been investigated to cope with this limitation. One example is applying pulsed flow, i.e., a non-steady flow defined by its amplitude, in other words the difference between maximum and minimum flow and pressure conditions, and its frequency. Several studies demonstrated the positive influence of pulsed flow on filtration [31,43] and cleaning performance [29,44] for various feed solutions, including milk. For membranes containing spacers, such as SWMs, particularly strong effects of pulsed flow on filtration and cleaning performance were reported due to pulsed flow reducing flow shadows due to enhanced turbulence [29,31] at high frequencies [44–49] and amplitudes [44–47,49].

However, some aspects exacerbate the transferability of lab-scale results to industrial-scale SWM modules. Firstly, the approaches to pulsation creation used in previous lab-

scale studies either included piston or bellows units [45,50] or specialised inductively controlled pumps that could create pulsed flow by rapidly increasing and decreasing pump capacity [29,31,44]. The former approach temporarily induced distinct back-pressure and is thus incompatible with SWMs. To the authors' knowledge, the latter one is unavailable on a larger scale. Secondly, the transferability of results from studies with FSMs to SWMs has been considered to be problematic for certain spacer geometries due to the curvature of the feed channel and its influence on the radial distribution of v [37]. Thirdly, due to the limited applicable pressure drops in SWMs, the highest applicable flow velocity and pulsation amplitude are also limited. In particular, positive results for pulsed filtration in a previous study using an SWM-like flat sheet membrane system (FSM) were found for pressure losses up to 2.55 bar m^{-1} [31], which is beyond the allowed limit of SWM modules. Thus, the advantage of pulsed flow might be reduced or absent for current SWM modules.

Hence, the potential beneficial effect of pulsed flow in filtration and cleaning remains to be evaluated for industrial-scale SWMs. Therefore, a novel approach was developed to create pulsed flow without back-pressure from the permeate side or relying on rapid pump capacity ramps. Then, pulsed flow can be utilised to assess the efficacy of pulsed flow MF of skim milk and subsequent membrane cleaning in SWMs. With previous studies observing improved pulsed flow efficiency for increased amplitudes and at pressure drops above the current limits of industrial SWMs, stability-enhanced modules could support the efficiency of pulsed flow manifold by enabling higher crossflow velocities, axial pressure drops and pulsed flow amplitudes. Hence, this study also investigates an approach to improve module stability against telescoping by adding glue connections on the feed side between membrane pockets, as this should provide additional resistance against the displacement of individual sheets in the axial direction.

Accordingly, this study aims to overcome the limitations of SWMs by two means. The first one is process-oriented and functions by modifying an existing plant for utilising pulsed flow and then assessing its efficacy in filtration and cleaning. The second approach is membrane-oriented and functions by creating a more robust SWM by adding glue connections on the feed side between membrane pockets, including spacers, to enhance its mechanical stability. For comparing steady and pulsed flow, filtration performance was evaluated in terms of permeate flux, protein permeation and protein mass flow. Cleaning success was evaluated hydrodynamically by measuring the flux recovery ratio (FRR) and chemically by analysing the protein removal achieved during cleaning. To investigate the effect of additional glue connections on module stability, the axial displacement of membrane layers in an unmodified and a glued membrane system was measured at different flow rates and radial distances to the module centre.

2. Materials and Methods

2.1. Filtration Plant and Experimental Design

An established pilot-scale filtration plant (Figure 1) was designed to resemble a typical industrial setup. It mainly consisted of a double-screw-type displacement pump (FDS 2-3, Fristam Pumpen KG, Hamburg-Bergedorf, Germany), which is insensitive to moderate pressure surges and commonly used in several dairy applications in which, e.g., highly viscous fluids such as milk concentrates need to be processed, and a membrane housing with the established module configuration 6338 (length $L = 0.96 \text{ m}$; diameter $d = 0.16 \text{ m}$). Additionally, pressure sensors and flow meters allow the monitoring and controlling of the transmembrane pressure TMP (see Equation (1)), feed flow rate and permeate flux J (see Equation (2)).

$$TMP = \frac{p_1 + p_2}{2} - p_3 \quad (1)$$

where p_1 is the feed-side pressure, p_2 is the retentate-side pressure and p_3 is the permeate-side pressure.

$$J = \frac{\dot{V}_{per}}{A_{membrane}} \tag{2}$$

where \dot{V}_{per} is the permeate flow rate and $A_{membrane}$ is the membrane area.

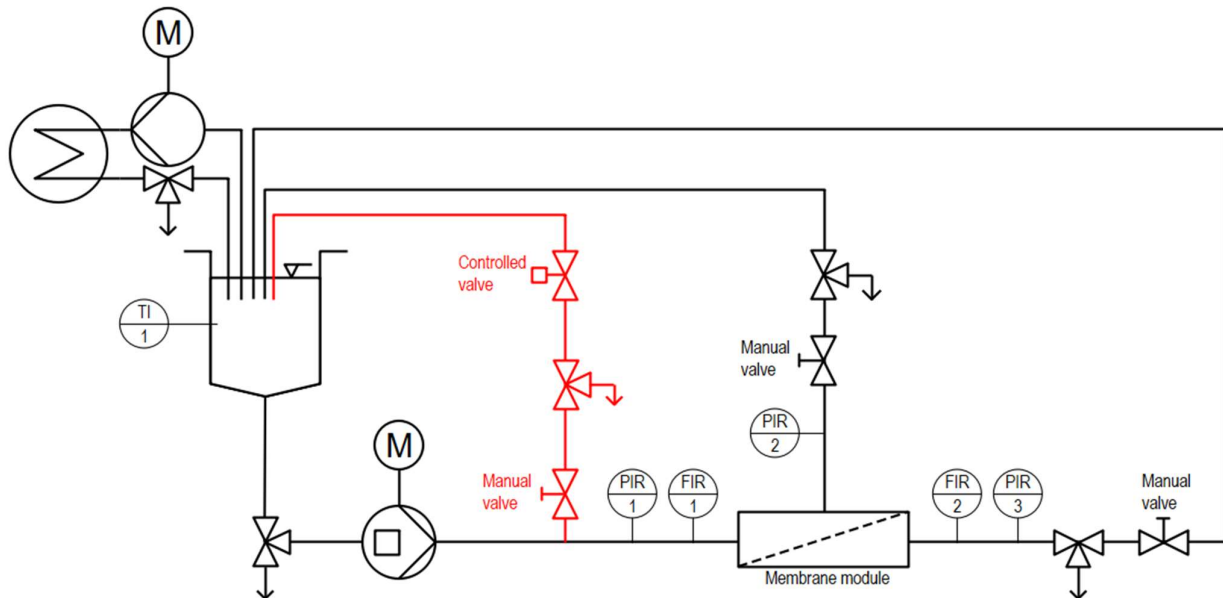


Figure 1. Piping and instrumentation (P and I) diagram of an established membrane filtration plant setup (black parts) consisting of feed pump, feed tank, sampling valves, manual throttling valves, a heating cycle consisting of another feed pump and heat exchanger, as well as various flow, pressure and temperature sensors. The red parts show the complimentary addition of a controlled bypass for utilising pulsed flow, including a manual throttling valve to control the pulsation amplitude and a controlled valve to control the pulsation frequency.

A separate heat exchanger loop combined with a temperature sensor enables precise temperature control of the filtration fluids before entering the membrane loop. Thus, the system can process various filtration feeds at defined temperatures, withstanding pressure surges and varying the installed membrane module’s geometry and pore size.

2.1.1. Plant Modification and Experimental Design to Utilise Pulsed Flow

To enable applying a pulsed flow to a standard membrane filtration plant setup without a pump capable of rapidly transitioning between high and low flow rates, a controlled bypass was added upstream of the membrane inlet flow meter and pressure sensor (see Figure 1). Accordingly, the bypass-related flow rate or pressure reductions could be monitored with installed sensors. The bypass comprised a relay-controlled pneumatic valve capable of fully closing or opening the bypass within 0.5 s at defined intervals. Hence, by determining the phase durations where the bypass was open (Δt_{min}) or closed (Δt_{max}), flow rate and pressure reach their minimum (\dot{V}_{min} , v_{min} and TMP_{min}) or maximum (\dot{V}_{max} , v_{max} and TMP_{max}), respectively, and thus control the pulsation frequency f (Equation (3)).

$$f = \frac{1}{\Delta t_{max} + \Delta t_{min}} \tag{3}$$

The additional manual valve allows control over the extent of flow rate reduction when the bypass is opened and thus the amplitude of pulsed flow in terms of flow rate (Equation (4)), flow velocity (Equation (5)) and TMP (Equation (6)).

$$\Delta\dot{V} = \dot{V}_{max} - \dot{V}_{min} \quad (4)$$

$$\Delta v = v_{max} - v_{min} \quad (5)$$

$$\Delta TMP_{cycle} = TMP_{max} - TMP_{min} \quad (6)$$

where $\Delta\dot{V}$ is the amplitude of flow rate, Δv is the amplitude of flow velocity and ΔTMP_{cycle} is the amplitude of TMP. It is to be noted that the flow velocity was calculated for a theoretical channel height without a spacer and a channel width of the SWM's spiral length. Hence, calculating v for spacer-filled channels can only provide a rough estimation, with local values highly depending on the position relative to the spacer grid.

All pulsed flow experiments were conducted at 50 °C, resembling a typical industrial filtration temperature [9]. The membrane was an MF SWM (GE JX6338C50) with a nominal pore size of 0.3 µm, the material polysulfone, a spacer height of 1.27 mm (50 mils), an active membrane area of 15.6 m², a diameter of 6.3 inches (16 cm) and a length of 38 inches (96 cm). Pasteurised skim milk (74 °C, 28 s) from a local dairy (Molkerei Weißenstephan, Freising, Germany) was used for deposit formation in all steady and pulsed flow filtration and cleaning experiments. Apart from filtration, deionised (DI) water was used in all other steps, either pure for rinsing or combined with chemicals for cleaning. As high frequencies [45–49] and amplitudes [45–47,51] were found to be beneficial for pulsed flow efficiency, the respective maximum values that were possible with the current setup were used in the pulsed flow filtration and cleaning experiments. Regarding TMP, the average values were chosen to resemble typical process conditions. TMP_{avg} during cleaning resembles the lowest possible value without reaching negative values for TMP_{min} and still enabling the identical flow velocity amplitude as during filtration (details see below).

Filtration Experiments

Before filtration, the membrane was conditioned with Ultrasil 69 (0.4% v/v , Ecolab Deutschland, Monheim am Rhein, Germany) at 50 °C for 20 min. After an intermediate rinsing step to avoid chemical residues, milk was heated to the process temperature by the heat exchanger loop, and filtration was initiated. Pulsed flow filtration was conducted with $f = 0.5$ Hz, the highest technically possible $\Delta\dot{V} = 10$ m³ h⁻¹, due to pump capacity limitations ($\dot{V}_{max} = 14$ m³ h⁻¹ with $\Delta p_{L,max} = 0.83$ bar m⁻¹ and $v_{max} = 0.37$ m s⁻¹, $\dot{V}_{min} = 4$ m³ h⁻¹ with $\Delta p_{L,min} = 0.14$ bar m⁻¹ and $v_{min} = 0.11$ m s⁻¹, $\dot{V}_{avg} = 9$ m³ h⁻¹ with $\Delta p_{L,avg} = 0.35$ bar m⁻¹ and $v_{avg} = 0.24$ m s⁻¹) and $\Delta TMP_{cycle} = 1.50$ bar ($TMP_{max} = 1.75$ bar, $TMP_{min} = 0.25$ bar, $TMP_{avg} = 1.00$ bar). The average TMP_{avg} and \dot{V}_{avg} were used for conducting comparative steady flow filtration runs. During the filtration duration of 60 min, samples were taken from permeate and retentate sample ports after 5, 10, 15, 30, 45 and 60 min. Protein permeation for a specific milk protein P_i was calculated by Equation (7) by its respective concentrations in the permeate $c_{i,p}$ and retentate $c_{i,r}$. Similarly, $c_{i,p}$ and Flux J were used to calculate an individual protein's permeating mass flow \dot{m}_i into the filtrate (Equation (8)).

$$P_i = \frac{c_{i,p}}{c_{i,r}} \quad (7)$$

$$\dot{m}_i = J \cdot c_{i,p} \quad (8)$$

After each filtration experiment, the membrane was rinsed and then cleaned with combined caustic (0.8% v/v , Ecolab Germany) and enzymatic (0.3% v/v Ultrasil 67, Ecolab

GmbH, Monheim am Rhein, Germany) cleaning agents for 40 min, followed by another rinsing step and an acidic cleaning step (0.4% *v/v* Ultrasil 75, Ecolab GmbH, Monheim am Rhein, Germany) for 20 min at 50 °C. To verify sufficient cleaning success and thus ensure long-term membrane functionality, the membrane's pure water flux was measured before each filtration run.

Cleaning Experiments

Before filtration, the membrane was conditioned, and the initial water flux J_0 was measured. Filtration was then conducted for 40 min at 50 °C, $\dot{V} = 5 \text{ m}^3 \text{ h}^{-1}$ and $\text{TMP} = 1.7 \text{ bar}$ with skim milk. Afterwards, the milk was drained, and the membrane system was carefully rinsed to remove bulk milk and loosely bound material. The following cleaning experiments were conducted with NaOH at pH 11.3 ($c_{\text{NaOH}} = 0.03\%$) for 20 min at 50 °C in circulation under either steady or pulsed flow mode. Due to the NaOH solution volume being high compared to the membrane area to be cleaned (yielding a specific cleaning volume of 6.4 L per square meter of active membrane area), an excess of cleaning agent compared to the amount of protein to be removed was present. Thus, the experiments should not be affected by the excessive consumption of cleaning agents causing distorted protein removal or changes in the pH. Pulsed flow cleaning was conducted with $f = 0.5 \text{ Hz}$, the maximum technically viable $\Delta\dot{V} = 10 \text{ m}^3 \text{ h}^{-1}$ ($\dot{V}_{\text{max}} = 14 \text{ m}^3 \text{ h}^{-1}$ with $\Delta p_{\text{L, max}} = 0.83 \text{ bar m}^{-1}$ and $v_{\text{max}} = 0.37 \text{ m s}^{-1}$, $\dot{V}_{\text{min}} = 4 \text{ m}^3 \text{ h}^{-1}$ with $\Delta p_{\text{L, min}} = 0.14 \text{ bar m}^{-1}$ and $v_{\text{min}} = 0.11 \text{ m s}^{-1}$, $\dot{V}_{\text{avg}} = 9 \text{ m}^3 \text{ h}^{-1}$ with $\Delta p_{\text{L, avg}} = 0.35 \text{ bar m}^{-1}$ and $v_{\text{avg}} = 0.24 \text{ m s}^{-1}$) and $\Delta\text{TMP}_{\text{cycle}} = 1.00 \text{ bar}$ ($\text{TMP}_{\text{max}} = 1.15 \text{ bar}$, $\text{TMP}_{\text{min}} = 0.15 \text{ bar}$, $\text{TMP}_{\text{avg}} = 0.60 \text{ bar}$). The average TMP_{avg} and \dot{V}_{avg} were used for comparative steady flow cleaning runs. For evaluating chemical cleaning success in terms of protein removal, samples were taken from the feed vessel after 20 min cleaning. Subsequently, the cleaning solution was drained, the system rinsed, and the water flux after cleaning J_1 was measured to evaluate the hydraulic cleanliness in terms of flux recovery ratio (FRR) (see Equation (9)) reached by the applied cleaning protocol. If the cleaning evaluation indicated incomplete cleaning ($\text{FRR} < 90\%$), an additional cleaning procedure with industrial cleaning agents, analogous to filtration experiments, was conducted to evaluate long-term membrane functionality.

$$\text{FRR} = \frac{J_1}{J_0} \quad (9)$$

It is to be noted that while identical pulsation frequencies as in previous FSM studies could be achieved with this approach and setup, the maximum applicable amplitudes and flow velocities were significantly lower in the current study due to limitations in pump capacity.

2.1.2. Membrane Modification and Experimental Design to Investigate Increased Axial Pressure Drops

To investigate increased axial pressure drops, the plant's double-screw-type displacement pump (see Figure 1) was replaced by a larger centrifugal pump capable of creating a feed pressure of 4.8 bar and a maximum feed flow rate of $45 \text{ m}^3 \text{ h}^{-1}$. In this scenario, experiments were conducted with used membranes put out of operation at an industrial plant to be free for establishing potentially destructive conditions. The membranes were provided by a local dairy, where they had been used for the filtration of dairy fluids for several months. The membranes (Koch Industries, Wichita, KS, USA) had a separation range of 10 kDa, a 31 mil (0.79 mm) diamond-shaped spacer and an active membrane area of 19.1 m^2 , a diameter of 6.3 inch (16 cm) and a length of 38 inch (96 cm). It is to be noted that the used membranes showed no apparent membrane failures despite a few areas with dislocated spacers between non-displaced membrane sheets.

To assess the effect of feed-side glue connections on membrane stability, modified membranes were obtained by inserting a two-component adhesive (Araldite 2014-1,

Huntsman Corporation, Salt Lake City, UT, USA) into the dry spacer channels with a syringe and thus glueing together the membrane sheets. This procedure resulted in semi-circular glue connections ($d = 2$ cm) placed in a radial direction along the membrane diameter (Figure 2). After hardening for several days, the modified membranes were comparatively assessed with unmodified membranes for their axial pressure drop stability.



Figure 2. The membrane module was modified by adding glue dots (see black areas) radially along the membrane diameter.

For axial stability experiments, the membranes were initially rinsed with deionised (DI) water with open permeate valves to allow the permeate pockets to be filled. To simulate the filtration of fouling-intensive feeds such as skim milk, where permeate production is substantially low and thus the influence of flux on the length dependency of crossflow velocity is negligibly small, axial stability experiments with water were conducted with the permeate valve closed ($TMP = 0.0$ bar). The membrane was then subjected to an initial axial pressure drop of 0.3 bar m^{-1} for 5 min. After assessing the axial displacement relative to the permeate collection tube at four equidistant points (radial distances 3.3 cm, 4.5 cm, 5.8 cm, 7.0 cm) in the radial direction of the SWM with a Vernier calliper, this procedure was repeated, increasing the axial pressure drop by 0.2 bar m^{-1} up to 1.5 bar m^{-1} . This approach allowed the evaluation of the displacement depending both on the applied axial pressure drop and the radial distance of displaced membrane sheets to the permeate collection tube.

Preliminary experiments with unmodified membranes and an ATD showed no significant displacement at either radial position for pressure drops $< 4.0 \text{ bar m}^{-1}$ (see Figure 3), contrary to industrial reports and restrictions stated by membrane manufacturers. This contradicting observation is presumably due to displacements with ATD only caused by long-term stress, as bond joints and polymers are known to migrate under constant stress [41]. As these long-term scenarios are hard to reproduce at lab scale, the following experiments were conducted without an ATD to exclusively assess the axial stability of the membrane module without the support of an ATD.

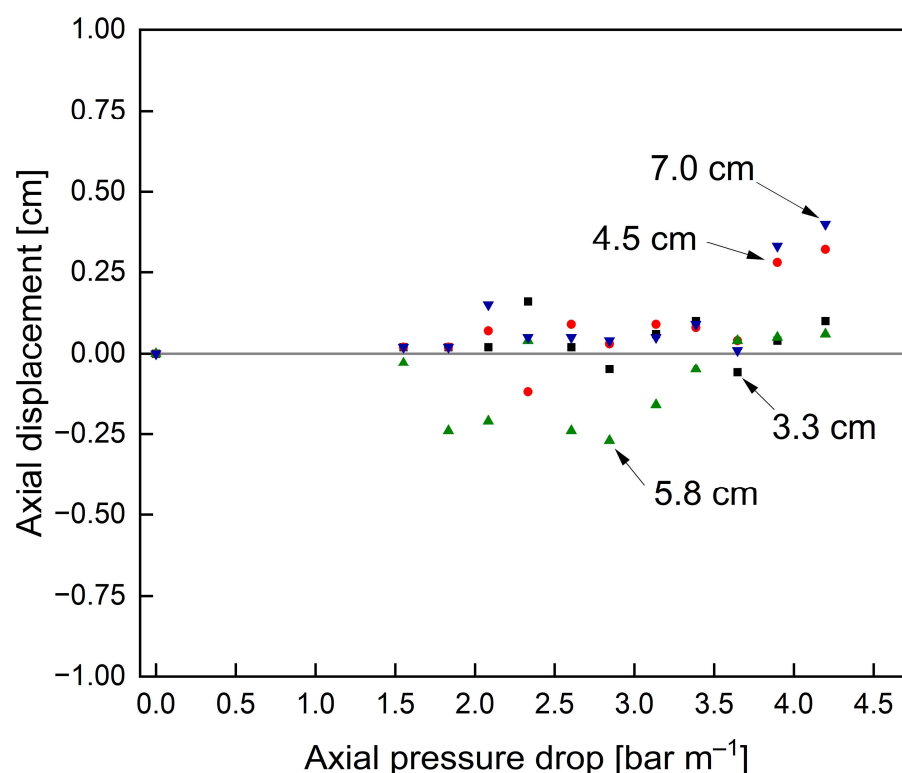


Figure 3. Influence of the axial pressure drop and radial position on the axial displacement of membrane sheets with an ATD. The grey reference line indicates no axial displacement.

2.2. Chemical and Statistical Analyses

The contents of caseins and whey proteins in filtration and cleaning samples were analysed by reversed-phase high-performance liquid chromatography (RP-HPLC) as described by Dümpler et al. [52]. Agilent ChemStation software (Rev. B.04.03) was used to analyse RP-HPLC chromatograms.

Data were plotted, fitted and statistically evaluated using OriginPro 2021 (OriginLab Corporation, Northampton, MA, USA). Statistical significance between data sets was assessed using a one-way analysis of variance (ANOVA) at the 5% level ($p < 0.05$). Depicted error bars represent the standard deviation of replicates, whereas all cleaning and filtration experiments were conducted at least in triplicates. Due to membrane failure/rupture accompanying axial displacements, stability experiments could only be conducted as single runs.

3. Results and Discussion

3.1. Optimisation of SWM's Process Efficiency via the Utilisation of Pulsed Flow

3.1.1. Validation of Plant Modifications

As larger pumps are normally incapable of rapidly producing quickly transitioning conditions between high and low flow rates, pulsed flow was created by installing a bypass, temporarily reducing the pressure and flow rates reaching the membrane module. An overview of the resulting pulsed flow characteristics is given in Figure 4.

Figure 4 depicts the time-resolved progression of flow rate \dot{V} and TMP. With this approach to pulsation creation and the specific setup used in this study, a maximum frequency of 0.5 Hz with an amplitude $>10 \text{ m}^3 \text{ h}^{-1}$ could be realised. Hence, compared to previous lab-scale studies using steep transitioning ramps of inductively controlled pumps to generate pulsed flow [29,31,44,53], the same maximum frequencies can be achieved at tenfold higher flow rates. Also, the profiles of flow rate and TMP progression correspond to those of lab-scale experiments with rapid ramps creating pulsed flow [44]. It is to be noted that while other valves with shorter opening and closing times might enable higher

pulsation frequencies, they might also induce intensified pressure surges on the plant equipment, which could cause enhanced wear. However, the current setup with $f = 0.5$ Hz did not cause any damage or wear on sensitive plant components, such as sensors or valves, within a pulsed flow operation of four months. Contrary to other approaches of creating pulsed flow, such as via bellows or piston units [45,50], it is also of advantage that the occurrence of negative TMP values can be avoided. This is particularly important for SWMs, where negative TMP must not exceed 0.3 bar, according to membrane manufacturers' specifications, as this would stress the bond joints of membrane pockets and could result in membrane failure. Overall, the bypass as a technical option to produce pulsed flow conditions resembles a low-effort and cost-efficient approach to creating similar pulsed flow profiles on a pilot scale with industrially sized membranes as in lab-scale studies. Nonetheless, it is to be noted that with this novel approach to pulsation creation, the energy efficiency is decreased compared to that of the previous system using rapid pump capacity ramps [31,44]. In this case, pumps will not alternately increase and decrease in pump capacity, but are instead continuously run at maximum capacity, despite a large share of flow temporarily not reaching the membrane during the low-flow pulsation phases.

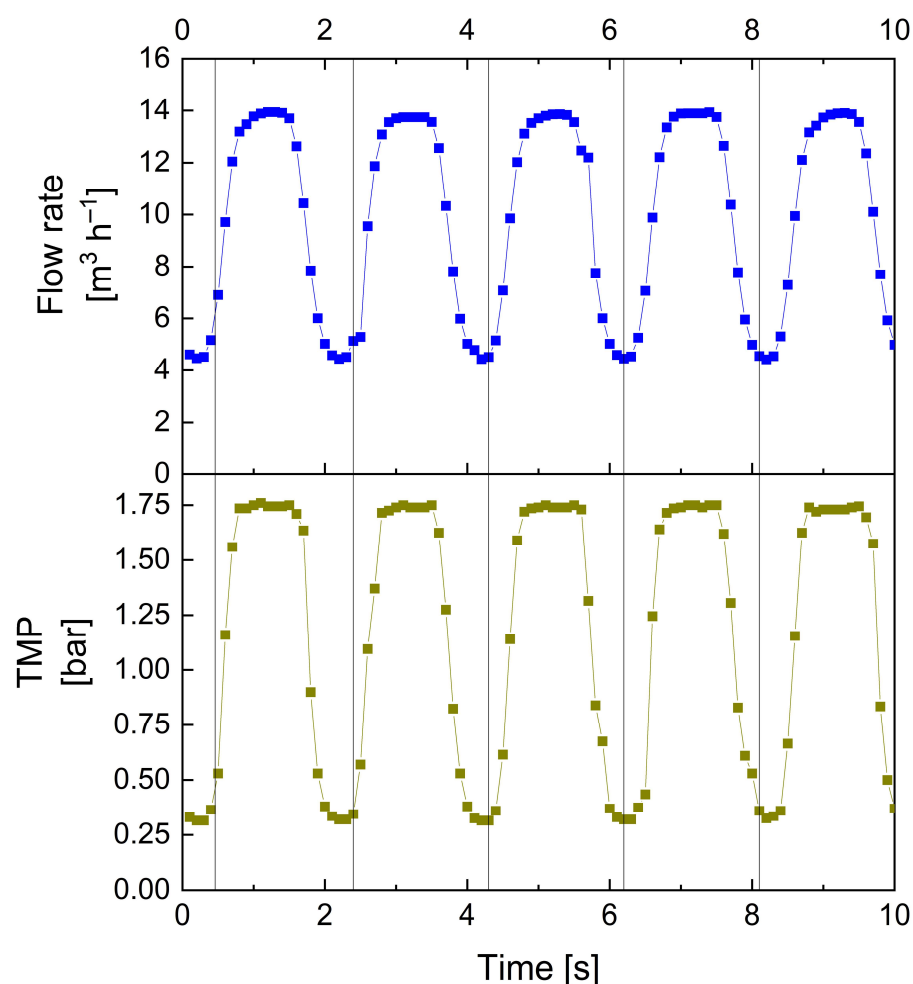


Figure 4. Time-resolved progression of feed flow rate and TMP over several flow cycles during pulsed filtration. Pulsed flow conditions: $\Delta\dot{V} = 10 \text{ m}^3 \text{ h}^{-1}$ ($\dot{V}_{\text{max}} = 14 \text{ m}^3 \text{ h}^{-1}$, $\dot{V}_{\text{min}} = 4 \text{ m}^3 \text{ h}^{-1}$, $\dot{V}_{\text{avg}} = 9 \text{ m}^3 \text{ h}^{-1}$) and $\Delta\text{TMP}_{\text{cycle}} = 1.50 \text{ bar}$ ($\text{TMP}_{\text{max}} = 1.75 \text{ bar}$, $\text{TMP}_{\text{min}} = 0.25 \text{ bar}$, $\text{TMP}_{\text{avg}} = 1.00 \text{ bar}$).

3.1.2. Influence of Pulsed Flow on Filtration Performance in Industrial-Scale SWMs

The mass flow resulting from flux and permeation was analysed to assess the effect of pulsed flow on the time-resolved filtration performance in industrial SWMs during 60 min filtration. The strongest impacts of pulsed flow were reported for high frequencies [44–49]

and amplitudes [44–47,49]. Accordingly, pulsed flow experiments were run with the best combination of frequency and amplitude applicable to the current setup. The mass flow of whey protein, i.e., the targeted permeating component (Figure 5), for pulsed flow was permanently increased over that of steady flow throughout filtration. While the whey protein mass flow with steady flow decreased from $38.9 \text{ g m}^{-2} \text{ h}^{-1}$ by 39% to $23.8 \text{ g m}^{-2} \text{ h}^{-1}$, pulsed flow decreased from $43.7 \text{ g m}^{-2} \text{ h}^{-1}$ by 31% to $30.0 \text{ g m}^{-2} \text{ h}^{-1}$. Hence, the initial mass flow (+12%), steady-state mass flow (+26%) and its decrease during filtration (−21%) were all improved with pulsed flow. These results demonstrate an improved initial and continuous deposit control with pulsed flow resulting in a 26% increased whey protein mass flow compared to steady flow at steady-state.

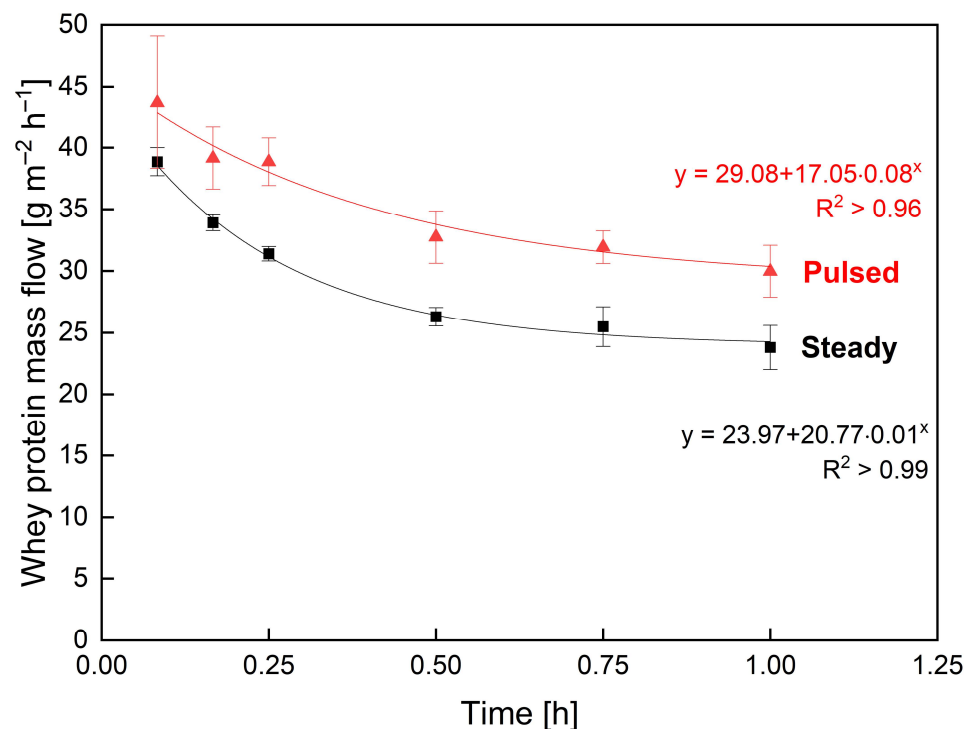


Figure 5. Influence of steady (black squares) and pulsed (red triangles) flow on whey protein mass flow during skim milk MF. Filtration conditions $\Delta\dot{V} = 10 \text{ m}^3 \text{ h}^{-1}$ ($\dot{V}_{\text{max}} = 14 \text{ m}^3 \text{ h}^{-1}$, $\dot{V}_{\text{min}} = 4 \text{ m}^3 \text{ h}^{-1}$, $\dot{V}_{\text{avg}} = 9 \text{ m}^3 \text{ h}^{-1}$) and $\Delta\text{TMP}_{\text{cycle}} = 1.50 \text{ bar}$ ($\text{TMP}_{\text{max}} = 1.75 \text{ bar}$, $\text{TMP}_{\text{min}} = 0.25 \text{ bar}$, $\text{TMP}_{\text{avg}} = 1.00 \text{ bar}$).

The observed improvements in filtration performance with pulsed flow are generally in accordance with our previous lab-scale study [31]. The increased mass flow, induced by enhanced flux and permeation, is the result of improved access to flow shadows causing improved deposit control with less fouling [31]. The small differences in performance improvement with pulsed flow between lab-scale and industrial-scale membranes, particularly regarding whey protein permeation, could arise from the fact that the highest applied axial pressure drop, and thus the flow velocity, was much lower ($\Delta p_{L, \text{max}} = 0.83 \text{ bar m}^{-1}$) compared to that in the previous study ($\Delta p_{L, \text{max}} = 2.55 \text{ bar m}^{-1}$) with FSM. The same applies to the amplitude ($\Delta v = 0.26 \text{ m s}^{-1}$ versus $\Delta v = 0.60 \text{ m s}^{-1}$) [31], as explained above. Frequency and amplitude have both been previously identified to be critical aspects for pulsed flow efficiency [44–47,54].

3.1.3. Influence of Pulsed Flow on Cleaning Efficiency in Industrial-Scale SWMs

To also examine the effect of pulsed flow on cleaning efficiency with the modified filtration plant for industrial-scale SWMs, cleaning experiments with NaOH at pH 11.3 ($c_{\text{NaOH}} = 0.03\%$) were conducted after steady flow filtration. Again, pulsed flow experiments were conducted at the maximum frequency and amplitude settings possible with

the current setup. The results of comparing steady and pulsed flow cleaning experiments were evaluated using FRR (Figure 6 left) and total protein removal (Figure 6 right) as assessment criteria.

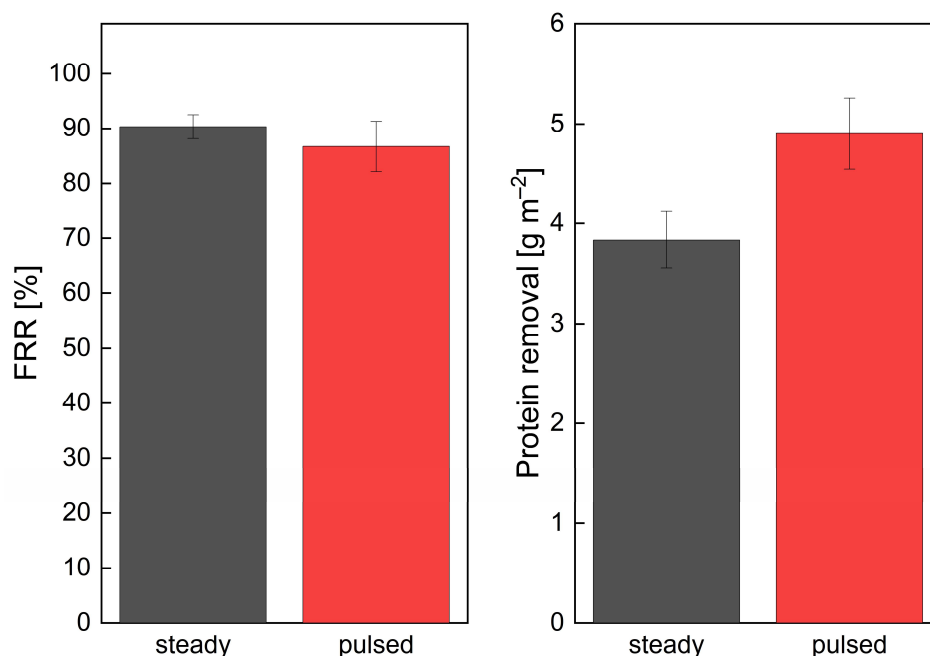


Figure 6. Influence of steady (black) and pulsed (red) flow on FRR (left) and protein removal (right) during membrane cleaning after skim milk MF. Cleaning conditions: $\Delta\dot{V} = 10 \text{ m}^3 \text{ h}^{-1}$ ($\dot{V}_{\text{max}} = 14 \text{ m}^3 \text{ h}^{-1}$, $\dot{V}_{\text{min}} = 4 \text{ m}^3 \text{ h}^{-1}$, $\dot{V}_{\text{avg}} = 9 \text{ m}^3 \text{ h}^{-1}$) and $\Delta\text{TMP}_{\text{cycle}} = 1.00 \text{ bar}$ ($\text{TMP}_{\text{max}} = 1.15 \text{ bar}$, $\text{TMP}_{\text{min}} = 0.15 \text{ bar}$, $\text{TMP}_{\text{avg}} = 0.60 \text{ bar}$).

Regarding FRR, there were no significant differences between flow modes, with $90 \pm 2\%$ for steady and $87 \pm 5\%$ for pulsed flow. Nonetheless, the protein removal achieved with pulsed flow ($4.90 \pm 0.36 \text{ g m}^{-2}$) was significantly increased by 28% over that achieved with steady flow cleaning ($3.83 \pm 0.29 \text{ g m}^{-2}$). With pulsed flow improving access to flow shadows [29,31,44,51,55–59] and thus improving removal particularly in these areas, the reason for the absence of improvements in FRR could be due to the steady water flux measurements being prone to the same flow shadows behind spacer filaments as steady flow cleaning. As shown in a previous study for FSMs, fouling residues in areas subject to flow shadows could only partially be removed by steady flow cleaning, whereas no distinct residues in those areas could be observed for pulsed flow cleaning [29]. As these observations were only reflected by an increased protein removal but not an increase in FRR, it can be assumed that the additional protein removal near spacer filaments could not be detected by steady flux measurements. This is presumably due to these areas hardly contributing to flux under steady flow, regardless of fouling being present or absent, and translates to an overestimation of hydraulic cleanliness for steady flow and an underestimation thereof for pulsed flow cleaning. This explanation also highlights FRR being insufficient as a singular tool for cleaning evaluation, particularly for membranes subject to flow shadows, such as FSMs or SWMs. Overall, similar to the filtration experiments (Section 3.1.2), the positive results from lab-scale trials could be confirmed, but the benefits were less pronounced, due to reasons explained above.

Another factor when comparing FSM and SWM results is the membrane length, which could also affect the results of pulsation efficiency. Due to the length dependency of Δp_L , TMP and fouling, the membrane length was previously identified in HFM to affect the cleaning efficiency for flow modes inducing flow reversal [53] but not for conventional steady or pulsed flow [29]. Nonetheless, due to significant geometrical differences between HFMs and SWMs, a declining efficiency of pulsation effects with increasing membrane

length in SWMs, e.g., due to propagating flow and pressure waves being partially absorbed by the friction with spacer filaments, cannot be excluded. Nonetheless, the lower flow velocity and amplitude, limited by the maximum pressure drop applicable and thus the stability of SWMs, remain the most probable causes for the observed differences between FSMs and SWMs. Hence, the following sections will investigate an approach to improve module stability in SWMs under operating conditions currently out of reach.

3.2. Optimisation of SWMs' Mechanical Stability by Feed-Side Glue Connections

First, the effect of glue connections on the filtration behaviour was to be assessed since the glued areas reduce the inlet cross-section of the module. Therefore, the relationship between axial pressure drop and volume flow rates was compared for a glued and an unmodified membrane (Figure 7).

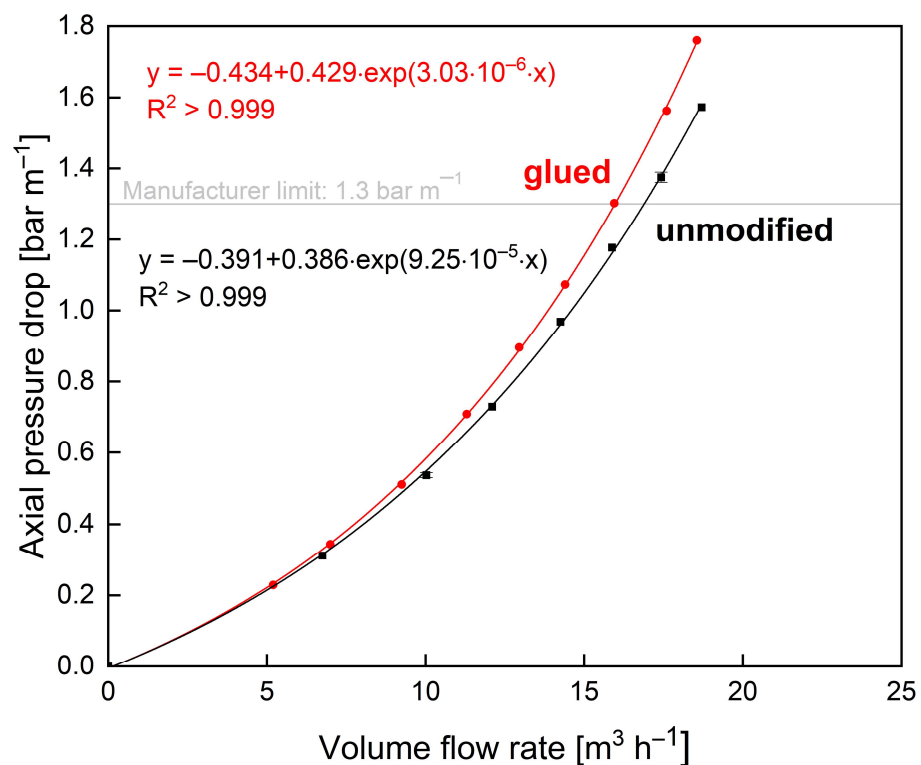


Figure 7. Axial pressure drop at different feed flow rates of the glued (red) and unmodified (black) membrane without an ATD.

The unmodified membrane reaches the maximum axial pressure drop of 1.3 bar m⁻¹ at 16.9 m³ h⁻¹, and the glued membrane already at 15.9 m³ h⁻¹. This means that for an identical maximum pressure drop of 1.30 bar m⁻¹, the modified membrane could only be operated at a six percent lower volume flow rate than the unmodified membrane. Hence, potential improvements in module stability need to be more pronounced than the enhanced axial pressure drop induced by the glued sections. Otherwise, if glued connections could not provide sufficient stability improvements, the enhanced axial pressure drop would further limit the highest applicable flow velocities. Also, it is to be noted that the glued connections were added to the SWM after its manufacture. Therefore, the areas covered with glue were larger and less well-shaped than they could be when created during the SWM manufacturing process.

Furthermore, the effect of additional glue connections on the short-term stability of the membranes without an ATD was assessed in terms of the axial displacement at different radial positions caused by different axial pressure drops (Figure 8).

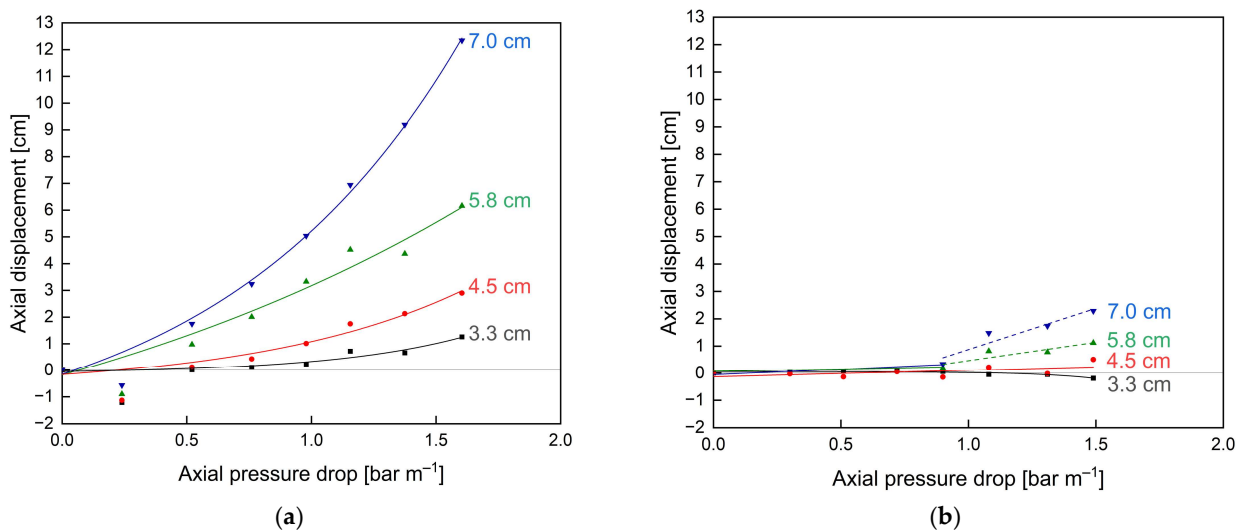


Figure 8. Axial displacement of the unmodified (a) and glued (b) membrane caused by axial pressure drops at different radial distances to the module centre without an ATD. The grey reference lines indicate no axial displacement. Lines are a guide for the eye.

Due to the absence of an ATD, the critical axial pressure drop, where axial displacement starts to occur, was reached at 0.5 bar m^{-1} for the unmodified membrane (Figure 8a). Beyond this point, the axial displacement increased exponentially as a function of axial pressure drop. Also, the displacement was most pronounced for the membrane parts in the radially outer positions, i.e., farthest away from the central collection tube (7.0 cm), as the pockets are only fixed to the central collection tube and the outer part is only held in place by the friction induced by the module wrapping. Hence, the outer part cannot take up high forces in the axial direction and thus is pushed towards the rear part of the module 879% further than the inner part (12.34 cm displacement at 7.0 cm radial distance versus 1.26 cm displacement at 3.3 cm radial distance) where most of the axial forces can be taken up by the connection to the central collection tube. Even at pressure drops of 1.5 bar m^{-1} , above the manufacturer limit of 1.3 bar m^{-1} , only a slight axial displacement $<1.0 \text{ cm}$ of the inner membrane envelope could be observed. Overall, these results emphasise both the instability, particularly of the outer SWM parts, against telescoping as well as the necessity and potential advantages of additional stabilisers, such as glued connections, for module stability.

The glued membrane (Figure 8b) showed vastly different results with no displacement up to an axial pressure drop of 1.0 bar m^{-1} . This translates to an overall 100% stability increase compared to the unmodified membrane with significant displacements already observed at 0.5 bar m^{-1} . Considering the displacements at different radial positions, they are all significantly reduced. For the inner part, i.e., 3.3 cm and 4.5 cm, no significant displacement can be observed for axial pressure drops up to 1.5 bar m^{-1} . At 4.5 cm radial distance and 1.5 bar m^{-1} , the displacement in the glued membrane was 82% reduced compared to that of the unmodified membrane (0.5 cm versus 2.9 cm). In the outer part (7.0 cm radial distance) at 1.5 bar m^{-1} , where displacement was most pronounced for both membranes, the displacement could be reduced by 81% (2.3 cm versus 12.3 cm). An overview of the achieved reductions in axial displacements with the modified membrane compared to the unmodified membrane shows an exponential increase for increasing pressure drops (see Figure 9a) and radial distances to the module centre (see Figure 9b). Hence, improvements of the modified membrane are most pronounced for outer membrane parts and at increased axial pressure drops or flow velocities.

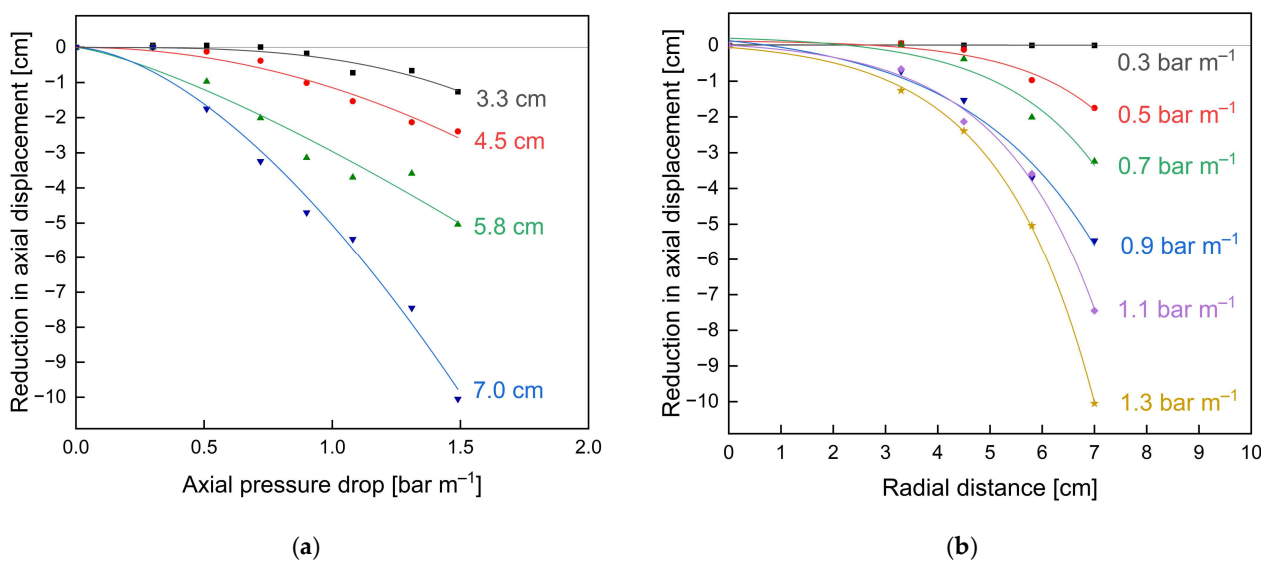


Figure 9. Reductions in axial displacement achieved by the glued membrane compared to the unmodified membrane at different feed flow rates (a), i.e., axial pressure drops and different radial distances to the module centre (b) without an ATD. The values were obtained by subtracting the displacement of the unmodified membrane from the glued membrane (see Figure 8). Lines are a guide for the eye.

Besides the observed stability increases, axial displacement still occurred due to the increasingly stressed bond joints eventually rupturing at their weakest point. As a consequence, the supportive effect of this bond joint vanished, and axial displacement occurred. Nevertheless, due to the supportive effect of the remaining membrane sheet connections, the resulting axial displacement could be significantly reduced compared to an envelope without glued bond joints. The related failure mechanism was a rupture of the glue connections. An additional failure mechanism, e.g., the axial displacement of the feed spacer, as observed in the used membranes, could not be observed for the glued membranes, which also underlines their enhanced stability.

It is to be noted that due to glue connections being added after SWM manufacturing and for used membranes, the adhesion between the selective and support layer, as well as the glue connections and their geometry, might not be ideal. Hence, it can be assumed that if prepared under ideal conditions, the stability improvements gained by glue connections would be more pronounced.

4. Conclusions

This study presented two approaches to successfully reduce the limitations of SWMs regarding cleanability and mechanical stability. The first approach, focusing on processing, transferred and utilised the concept of pulsed flow to industrial-scale membranes by adding a controlled bypass. This led to similar flow characteristics but less distinct process improvements during filtration and cleaning compared to previous lab-scale studies using FSMs [29,31]. While the underlying causes for the observed differences between lab-scale FSMs and industrial-scale SWMs could not entirely be determined, they are presumably due to the reduced flow velocity and pulsation amplitude applicable in SWMs. Nonetheless, significant improvements for both filtration (mass flow +26%) and cleaning performance (protein removal +28%) could be confirmed for pulsed flow. It is to be noted that while this approach was associated with low effort and investment cost on the pilot scale, a transfer to industrial-scale systems, often encompassing several membrane housings, i.e., filtration units, would require an efficient implementation of the additional controlled bypass into each filtration unit. This could be performed by, e.g., combining two filtration units into one pulsation unit where the flow control is managed by a controlled three-way diverting

valve instead of a controlled bypass. This way, one filtration unit would be in the high flow rate phase while the other filtration unit would be in the low flow rate phase. One advantage of this approach would be that no pump energy would be left unused as bypass flow but instead split between two filtration units pulsing inversely. Despite the advantages of pulsed flow, the necessity of adding the respective type of pulsation creation to every filtration unit is given for each type of pulsation creation and should thus be considered upon implementation.

The second approach, focusing on membrane construction, applied glue connections between membrane sheets in the radial direction across the membrane width of a used SWM. As a result, the axial pressure drop at a given flow rate was slightly increased. However, the stability against axial displacement without an ATD was increased by $\geq 100\%$ across the whole membrane diameter. Consequently, the enhanced axial pressure drop at the inlet partly compensated the glued connections' positive effect. Nevertheless, the positive effect of enhanced stability predominates over the disadvantage of enhanced axial pressure drop. Thus, higher flow rates and increased amplitudes under pulsed flow are expected to be applicable to the glue-connected SWM. Due to glue connections being added under non-ideal conditions, i.e., after manufacturing and for used membranes, it can be assumed that stability improvements would be more pronounced under ideal glueing conditions. In the case of manufacturing the interconnections between the membrane pockets simultaneously with the SWM itself, the bonds could be formed slimmer but longer to leave more free inlet cross-sections. Additionally, the effect of those glue connections might be enhanced by optimising their location, orientation and extent within the membrane module. However, since the effects on module stability could only be assessed with short-term experiments under extreme conditions, i.e., without an ATD, long-term stability tests should be conducted to confirm the enhanced stability for conventional setups with an ATD.

Finally, the combined maximum achievable advantages of using a stabilised SWM with pulsed flow allowing for increased pulsation amplitudes at increased flow rates should be assessed to facilitate weighing the required implementation effort versus the gained advantage, particularly from an economic and ecological point of view.

Author Contributions: Conceptualization, C.K., M.H., R.S. and S.S.; data curation, C.K., M.H. and P.O.; formal analysis, C.K. and M.H.; funding acquisition, U.K.; investigation, C.K., M.H. and P.O.; methodology, C.K. and M.H.; project administration, C.K.; resources, U.K.; software, C.K. and M.H.; supervision, U.K.; validation, C.K. and M.H.; visualization, C.K. and M.H.; writing—original draft, C.K. and M.H.; writing—review and editing, C.K., M.H. and U.K. All authors have read and agreed to the published version of the manuscript.

Funding: This IGF Project of the FEI was supported via AiF to promote the Industrial Collective Research (IGF) of the German Ministry of Economic Affairs and Energy (BMWi), based on a resolution of the German Parliament. Project: AiF 57 EWN.

Institutional Review Board Statement: Not applicable.

Data Availability Statement: The datasets generated during and/or analysed during the current study are available from the corresponding author on reasonable request.

Acknowledgments: We gratefully thank Heidi Wohlschläger for her assistance with the RP-HPLC analysis of samples. Furthermore, we want to thank Christian Ederer, Erich Schneider and Franz Fraunhofer in our workshop for technical support. We also want to thank Diana Kellhammer and Franz Kuhnert for their experimental support. Finally, we want to thank Nikolaus Adams and Steffen Schmidt from the Aerodynamics and Fluid Mechanics for organisationally supporting parts of this work.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Bartlett, M.; Bird, M.R.; Howell, J.A. An experimental study for the development of a qualitative membrane cleaning model. *J. Membr. Sci.* **1995**, *105*, 147–157. [[CrossRef](#)]
2. Bird, M.R.; Bartlett, M. Measuring and modelling flux recovery during the chemical cleaning of MF membranes for the processing of whey protein concentrate. *J. Food Eng.* **2002**, *53*, 143–152. [[CrossRef](#)]
3. Cui, Z.F.; Jiang, Y.; Field, R.W. Fundamentals of pressure-driven membrane separation processes. In *Membrane Technology*; Elsevier: Oxford, UK, 2010; pp. 1–18. ISBN 9781856176323.
4. Saxena, A.; Tripathi, B.P.; Kumar, M.; Shahi, V.K. Membrane-based techniques for the separation and purification of proteins: An overview. *Adv. Colloid. Interface Sci.* **2009**, *145*, 1–22. [[CrossRef](#)]
5. Baker, R.W. *Membrane Technology and Applications*; John Wiley & Sons, Ltd.: Chichester, UK, 2012; ISBN 9781118359686.
6. Ng, K.S.; Haribabu, M.; Harvie, D.J.; Dunstan, D.E.; Martin, G.J. Mechanisms of flux decline in skim milk ultrafiltration: A review. *J. Membr. Sci.* **2017**, *523*, 144–162. [[CrossRef](#)]
7. Ripperger, S.; Altmann, J. Crossflow microfiltration—state of the art. *Sep. Purif. Technol.* **2002**, *26*, 19–31. [[CrossRef](#)]
8. Ripperger, S.; Grein, T. Filtrationsverfahren mit Membranen und ihre Modellierung. *Chem. Ing. Tech.* **2007**, *79*, 1765–1776. [[CrossRef](#)]
9. Schiffer, S.; Kulozik, U. Effect of temperature-dependent bacterial growth during milk protein fractionation by means of 0.1 μM microfiltration on the length of possible production cycle times. *Membranes* **2020**, *10*, 326. [[CrossRef](#)] [[PubMed](#)]
10. Chamberland, J.; Messier, T.; Dugat-Bony, E.; Lessard, M.-H.; Labrie, S.; Doyen, A.; Pouliot, Y. Influence of feed temperature to biofouling of ultrafiltration membrane during skim milk processing. *Int. Dairy J.* **2019**, *93*, 99–105. [[CrossRef](#)]
11. Burgess, S.A.; Lindsay, D.; Flint, S.H. Thermophilic bacilli and their importance in dairy processing. *Int. J. Food Microbiol.* **2010**, *144*, 215–225. [[CrossRef](#)]
12. Carrascosa, C.; Raheem, D.; Ramos, F.; Saraiva, A.; Raposo, A. Microbial Biofilms in the Food Industry—A Comprehensive Review. *Int. J. Environ. Res. Public Health* **2021**, *18*, 2014. [[CrossRef](#)]
13. Ng, K.S.; Dunstan, D.E.; Martin, G.J. Influence of processing temperature on flux decline during skim milk ultrafiltration. *Sep. Purif. Technol.* **2018**, *195*, 322–331. [[CrossRef](#)]
14. Brans, G.; Schroën, C.G.; van der Sman, R.G.; Boom, R.M. Membrane fractionation of milk: State of the art and challenges. *J. Membr. Sci.* **2004**, *243*, 263–272. [[CrossRef](#)]
15. Qu, P.; Gésan-Guiziou, G.; Bouchoux, A. Dead-end filtration of sponge-like colloids: The case of casein micelle. *J. Membr. Sci.* **2012**, *417–418*, 10–19. [[CrossRef](#)]
16. Qu, P.; Bouchoux, A.; Gésan-Guiziou, G. On the cohesive properties of casein micelles in dense systems. *Food Hydrocoll.* **2015**, *43*, 753–762. [[CrossRef](#)]
17. Bouchoux, A.; Gésan-Guiziou, G.; Pérez, J.; Cabane, B. How to squeeze a sponge: Casein micelles under osmotic stress, a SAXS study. *Biophys. J.* **2010**, *99*, 3754–3762. [[CrossRef](#)]
18. Bouchoux, A.; Cayemite, P.-E.; Jardin, J.; Gésan-Guiziou, G.; Cabane, B. Casein micelle dispersions under osmotic stress. *Biophys. J.* **2009**, *96*, 693–706. [[CrossRef](#)]
19. Horne, D.S. Casein micelle structure and stability. In *Milk Proteins*; Elsevier: Cambridge, UK, 2020; pp. 213–250. ISBN 9780128152515.
20. Hartinger, M.; Heidebrecht, H.-J.; Schiffer, S.; Dimpler, J.; Kulozik, U. Technical concepts for the investigation of spatial effects in spiral-wound microfiltration membranes. *Membranes* **2019**, *9*, 80. [[CrossRef](#)] [[PubMed](#)]
21. Blanpain-Avet, P.; Migdal, J.F.; Bénézech, T. Chemical cleaning of a tubular ceramic microfiltration membrane fouled with a whey protein concentrate suspension—Characterization of hydraulic and chemical cleanliness. *J. Membr. Sci.* **2009**, *337*, 153–174. [[CrossRef](#)]
22. Samuelsson, G.; Dejmek, P.; Trägårdh, G.; Paulsson, M. Minimizing whey protein retention in cross-flow microfiltration of skim milk. *Int. Dairy J.* **1997**, *7*, 237–242. [[CrossRef](#)]
23. Defrance, L.; Jaffrin, M.Y. Comparison between filtrations at fixed transmembrane pressure and fixed permeate flux: Application to a membrane bioreactor used for wastewater treatment. *J. Membr. Sci.* **1999**, *152*, 203–210. [[CrossRef](#)]
24. Qiu, T.Y.; Davies, P.A. Concentration polarization model of spiral-wound membrane modules with application to batch-mode RO desalination of brackish water. *Desalination* **2015**, *368*, 36–47. [[CrossRef](#)]
25. Farhat, N.M.; Staal, M.; Bucs, S.; van Loosdrecht, M.; Vrouwenvelder, J.S. Spatial heterogeneity of biofouling under different cross-flow velocities in reverse osmosis membrane systems. *J. Membr. Sci.* **2016**, *520*, 964–971. [[CrossRef](#)]
26. Altmann, J.; Ripperger, S. Particle deposition and layer formation at the crossflow microfiltration. *J. Membr. Sci.* **1997**, *124*, 119–128. [[CrossRef](#)]
27. Hartinger, M.; Schiffer, S.; Heidebrecht, H.-J.; Dimpler, J.; Kulozik, U. Milk protein fractionation by custom-made prototypes of spiral-wound microfiltration membranes operated at extreme crossflow velocities. *J. Membr. Sci.* **2020**, *605*, 118110. [[CrossRef](#)]
28. Schopf, R.; Schmidt, F.; Linner, J.; Kulozik, U. Comparative assessment of tubular ceramic, spiral wound, and hollow fiber membrane microfiltration Module Systems for Milk Protein Fractionation. *Foods* **2021**, *10*, 692. [[CrossRef](#)]
29. Kürzl, C.; Kulozik, U. Comparison of the efficiency of pulsed flow membrane cleaning in hollow fibre (HFM) and spiral-wound microfiltration membranes (SWM). *Food Bioprod. Process.* **2023**, *139*, 166–177. [[CrossRef](#)]

30. Fischer, N.; Masoudian, M.; Germann, N. Impact of non-Newtonian fluid behavior on hydrodynamics and mass transfer in spacer-filled channels. *Fluid. Dyn. Res.* **2020**, *52*, 65502. [[CrossRef](#)]
31. Kürzl, C.; Kulozik, U. Influence of Pulsed and Alternating Flow on the Filtration Performance during Skim Milk Microfiltration with Flat-Sheet Membranes. *Sep. Purif. Technol.* **2023**, *321*, 124234. [[CrossRef](#)]
32. Geraldes, V. Flow management in nanofiltration spiral wound modules with ladder-type spacers. *J. Membr. Sci.* **2002**, *203*, 87–102. [[CrossRef](#)]
33. Han, Z.; Terashima, M.; Liu, B.; Yasui, H. CFD investigation of the effect of the feed spacer on hydrodynamics in spiral wound membrane modules. *MCA* **2018**, *23*, 80. [[CrossRef](#)]
34. Kaviani-pour, O.; Ingram, G.D.; Vuthaluru, H.B. Investigation into the effectiveness of feed spacer configurations for reverse osmosis membrane modules using Computational Fluid Dynamics. *J. Membr. Sci.* **2017**, *526*, 156–171. [[CrossRef](#)]
35. Schwinge, J.; Wiley, D.E.; Fletcher, D.F. A CFD study of unsteady flow in narrow spacer-filled channels for spiral-wound membrane modules. *Desalination* **2002**, *146*, 195–201. [[CrossRef](#)]
36. Koutsou, C.P.; Yiantsios, S.G.; Karabelas, A.J. Direct numerical simulation of flow in spacer-filled channels: Effect of spacer geometrical characteristics. *J. Membr. Sci.* **2007**, *291*, 53–69. [[CrossRef](#)]
37. Hartinger, M.; Napiwotzki, J.; Schmid, E.-M.; Hoffmann, D.; Kurz, F.; Kulozik, U. Influence of spacer design and module geometry on the filtration performance during skim milk microfiltration with flat sheet and spiral-wound membranes. *Membrane* **2020**, *10*, 57. [[CrossRef](#)]
38. Habenicht, G. *Kleben: Grundlagen, Technologien, Anwendungen*, 6. Aufl. 2009; Springer: Berlin/Heidelberg, Germany, 2008; ISBN 978-3-540-85266-7.
39. Dilger, K. Selecting the right joint design and fabrication techniques. In *Advances in Structural Adhesive Bonding*; Elsevier: Cambridge, UK, 2010; pp. 295–315. ISBN 9781845694357.
40. Grote, K.-H.; Bender, B.; Göhlich, D. (Eds.) *Dubbel-Taschenbuch für den Maschinenbau*; 25. Auflage; Springer Vieweg: Berlin/Heidelberg, Germany, 2018; ISBN 9783662548042.
41. Althof, W. Verformungs-Und Festigkeitseigenschaften von Klebstoffen bei Kurz-Und Langzeitbeanspruchung. *N39* **1984**, 141–162.
42. Karabelas, A.J.; Koutsou, C.P.; Sioutopoulos, D.C. Comprehensive performance assessment of spacers in spiral-wound membrane modules accounting for compressibility effects. *J. Membr. Sci.* **2018**, *549*, 602–615. [[CrossRef](#)]
43. Weinberger, M.E.; Kulozik, U. Pulsatile crossflow improves microfiltration fractionation of cells and proteins. *J. Membr. Sci.* **2021**, *629*, 119295. [[CrossRef](#)]
44. Kürzl, C.; Tran, T.; Kulozik, U. Application of a pulsed crossflow to improve chemical cleaning efficiency in hollow fibre membranes following skim milk microfiltration. *Sep. Purif. Technol.* **2022**, *302*, 122123. [[CrossRef](#)]
45. Gillham, C.R.; Fryer, P.J.; Hasting, A.P.; Wilson, D.I. Enhanced cleaning of whey protein soils using pulsed flows. *J. Food Eng.* **2000**, *46*, 199–209. [[CrossRef](#)]
46. Blé, W.; Le Gentil-Lelièvre, C.; Bénézech, T.; Legrand, J.; Legentilhomme, P. Application of turbulent pulsating flows to the bacterial removal during a cleaning in place procedure. Part 1: Experimental analysis of wall shear stress in a cylindrical pipe. *J. Food Eng.* **2009**, *90*, 422–432. [[CrossRef](#)]
47. Blé, W.; Legentilhomme, P.; Bénézech, T.; Legrand, J.; Le Gentil-Lelièvre, C. Application of turbulent pulsating flows to the bacterial removal during a cleaning in place procedure. Part 2: Effects on cleaning efficiency. *J. Food Eng.* **2009**, *90*, 433–440. [[CrossRef](#)]
48. Yang, J.; Kjellberg, K.; Jensen, B.B.B.; Nordkvist, M.; Gernaey, K.V.; Krühne, U. Investigation of the cleaning of egg yolk deposits from tank surfaces using continuous and pulsed flows. *Food Bioprod. Process.* **2019**, *113*, 154–167. [[CrossRef](#)]
49. Weidemann, C.; Vogt, S.; Nirschl, H. Cleaning of filter media by pulsed flow—Establishment of dimensionless operation numbers describing the cleaning result. *J. Food Eng.* **2014**, *132*, 29–38. [[CrossRef](#)]
50. Bode, K.; Hooper, R.J.; Paterson, W.R.; Wilson, D.I.; Augustin, W.; Scholl, S. Pulsed flow cleaning of whey protein fouling layers. *Heat Transf. Eng.* **2007**, *28*, 202–209. [[CrossRef](#)]
51. Augustin, W.; Fuchs, T.; Föste, H.; Schöler, M.; Majschak, J.-P.; Scholl, S. Pulsed flow for enhanced cleaning in food processing. *Food Bioprod. Process.* **2010**, *88*, 384–391. [[CrossRef](#)]
52. Dümpler, J.; Wohlschläger, H.; Kulozik, U. Dissociation and coagulation of caseins and whey proteins in concentrated skim milk heated by direct steam injection. *Dairy Sci. Technol.* **2017**, *96*, 807–826. [[CrossRef](#)]
53. Kürzl, C.; Kulozik, U. Alternating flow direction improves chemical cleaning efficiency in hollow fibre membranes following skim milk microfiltration. *J. Food Eng.* **2023**, *356*, 111587. [[CrossRef](#)]
54. Augustin, W.; Böhnet, M. Influence of pulsating flow on fouling behaviour. In *Proceedings of the International Conference on Mitigation of Heat Exchanger Fouling and Its Economic and Environmental Implications*, Banff, Canada, 18–23 July 1999; ISBN 1-56700-172-6.
55. Föste, H.; Schöler, M.; Majschak, J.-P.; Augustin, W.; Scholl, S. Modeling and validation of the mechanism of pulsed flow cleaning. *Heat Transf. Eng.* **2013**, *34*, 753–760. [[CrossRef](#)]
56. Gu, B.; Adjiman, C.S.; Xu, X.Y. The effect of feed spacer geometry on membrane performance and concentration polarisation based on 3D CFD simulations. *J. Membr. Sci.* **2017**, *527*, 78–91. [[CrossRef](#)]
57. Schwinge, J.; Wiley, D.E.; Fletcher, D.F. Simulation of unsteady flow and vortex shedding for narrow spacer-filled channels. *Ind. Eng. Chem. Res.* **2003**, *42*, 4962–4977. [[CrossRef](#)]

58. Schwinge, J.; Neal, P.R.; Wiley, D.E.; Fletcher, D.F.; Fane, A.G. Spiral wound modules and spacers. *J. Membr. Sci.* **2004**, *242*, 129–153. [[CrossRef](#)]
59. Koutsou, C.P.; Yiantsios, S.G.; Karabelas, A.J. A numerical and experimental study of mass transfer in spacer-filled channels: Effects of spacer geometrical characteristics and Schmidt number. *J. Membr. Sci.* **2009**, *326*, 234–251. [[CrossRef](#)]

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.