



From Cells to Plaques: The Molecular Pathways of Coronary Artery Calcification and Disease

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Abstract: Coronary artery calcification (CAC) is a hallmark of atherosclerosis and a critical factor in the development and progression of coronary artery disease (CAD). This review aims to address the complex pathophysiological mechanisms underlying CAC and its relationship with CAD. We examine the cellular and molecular processes that drive the formation of calcified plaques, highlighting the roles of inflammation, lipid accumulation, and smooth muscle cell proliferation. Additionally, we explore the genetic and environmental factors that contribute to the heterogeneity in CAC and CAD presentation among individuals. Understanding these intricate mechanisms is essential for developing targeted therapeutic strategies and improving diagnostic accuracy. By integrating current research findings, this review provides a comprehensive overview of the pathways linking CAC to CAD, offering insights into potential interventions to mitigate the burden of these interrelated conditions.

Keywords: coronary artery calcification (CAC); coronary artery disease (CAD); atherosclerosis; vascular calcification; inflammation; vascular smooth muscle cells (VSMCs); plaque stability; osteogenic differentiation; molecular pathways

1. Introduction

Coronary artery calcification (CAC) is increasingly recognized as a critical component in the development and progression of coronary artery disease (CAD), the leading cause of morbidity and mortality worldwide [1]. CAC is a complex, multifaceted process characterized by the deposition of calcium in the coronary arteries, leading to obstructive CAD [2]. This calcification not only serves as a hallmark of advanced atherosclerosis but also plays a direct role in the pathophysiology of CAD, contributing to plaque stability and the risk of acute coronary events [3].

Understanding the pathophysiological mechanisms linking CAC and CAD is essential for several reasons. First, CAC is a strong and independent predictor of cardiovascular events, making it a valuable marker for assessing CAD risk. Second, the molecular and cellular processes driving CAC are closely linked with those that promote atherosclerosis, including inflammation, lipid accumulation, and smooth muscle cell proliferation. These processes contribute to the formation and progression of calcified plaques, which can significantly impact the clinical course of CAD [4]. Despite advancements in imaging modalities and treatments, the cellular and molecular pathways contributing to CAC



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Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). remain incompletely understood. This review aims to fill this gap by exploring the complex interplay of molecular signals that drive CAC, thereby providing a foundation for novel therapeutic interventions.

This review aims to provide a comprehensive overview of the current understanding of the molecular pathways involved in CAC and its relationship with CAD. By exploring the roles of vascular smooth muscle cells (VSMCs), inflammatory cytokines, lipid metabolism, and genetic factors, we seek to elucidate the complex interactions that underlie plaque calcification. Additionally, we will discuss the clinical implications of CAC, including its role in risk stratification, diagnostic imaging, and potential therapeutic interventions.

2. Methods

This narrative review is based on a comprehensive literature search conducted in databases such as PubMed, Google Scholar, and Scopus. Keywords such as 'coronary artery calcification', 'atherosclerosis', 'molecular pathways', and 'vascular smooth muscle cells' were used to identify relevant peer-reviewed articles. Studies were selected based on relevance to the topic, publication date, and citation count. The search strategy aimed to include the most recent and impactful studies to ensure the review's thoroughness. Cross-referencing of key articles was conducted to minimize the possibility of missing critical data.

3. Cellular and Molecular Mechanisms of Coronary Artery Calcification

CAC is characterized by the calcium phosphate deposition process in various conditions, such as atherosclerosis, hypertension [5], aortic valve stenosis, CAD, diabetes mellitus (DM) [6], chronic kidney disease (CKD) [7], hyperlipidemia [8], and chronic inflammatory disorders. CAC, intimal or medial, stiffens arteries and weakens vulnerable atherosclerotic plaques, increasing the risk of rupture [9]. Intimal calcification is frequently associated with damage and dysfunction of endothelial cells (EC), as well as atherosclerosis [10]. It is caused by the imbalance of the vascular microenvironment, where the pro-calcific factors (inflammation, endoplasmic reticulum (ER) stress, mitochondrial dysfunction, iron homeostasis, programmed cell death (PCD), and other cellular metabolic dynamics) are enhanced [11–13].

VSMCs play a key role in the development of CAC, and the loss of VSMCs through various forms of PCD, such as apoptosis, necrosis, necroptosis, pyroptosis, autophagy, and ferroptosis, contributes to the thinning of fibrous caps and the formation of necrotic cores, leading to calcification [14,15]. Recent studies have highlighted nucleotide-binding and oligomerization domain-like receptor family pyrin domain-containing 3 (NLRP3)-mediated pyroptosis as a protective mechanism, while autophagy has been identified as a key process in diabetes-related calcification [16]. Autophagy can inhibit CAC by suppressing the osteogenic differentiation of VSMCs through mechanisms involving pathways such as anti-differentiation non-coding RNA, b-catenin, and AMP-activated protein kinase (AMPK), while it may also promote calcification via cAMP response element-binding protein signaling, elastin degradation, and long non-coding RNA H19-mediated extracellular signal-regulated kinase (ERK) signaling [17–20].

Phosphoglycerate dehydrogenase (PHGDH) inhibits CAC in VSMCs by preventing ferroptosis via the P53/solute carrier family 7a member 11 (SLC7A11) pathways, making it a therapeutic target [21]. Indoleamine 2,3-dioxygenase 1 (IDO1) deficiency promotes calcification through enhanced Runt-related transcription factor 2 (RUNX2) activity, while kynurenine, an IDO1 catabolite, counteracts this by promoting RUNX2 degradation via aryl hydrocarbon receptor (AhR) [22]. Low C1 tumor necrosis factor-related protein 3 (CTRP3) levels are linked to CAC in diabetic patients, as CTRP3 inhibits osteogenic differentiation in VSMCs by blocking β -catenin nuclear translocation [23]. Ne-carboxymethyl-lysine (CML), an advanced glycation product, promotes calcification in diabetic atherosclerosis by enhancing foam cell formation [24]. Inhibitors like fetuin-A, matrix Gla protein (MGP), and osteoprotegerin (OPG) prevent calcium deposition [25–27]. Epidermal growth factor

receptor (EGFR) inhibition reduces calcification by blocking calcifying vesicle release from VSMCs, while sortilin regulates calcification via Rab11 trafficking [28]. Osteopontin (OPN) and matrix metalloproteinase-9 (MMP-9) contribute to CAC and atherosclerosis progression, with MMP-9 enhancing macrophage infiltration and matrix degradation [29]. CML promotes calcification in diabetic VSMCs through the CML/RAGE-ROS-p38MAPKcbf α 1-ALP pathway [30].

The Wingless-type family member (Wnt)/ β -catenin signaling pathway promotes CAC, particularly in response to high phosphorus levels [31]. Wnt3a, Wnt5a, and Wnt5b are upregulated in human coronary plaques, influencing VSMC differentiation, calcification, and cholesterol handling [32]. This pathway stimulates RUNX2, a key transcription factor in the osteogenic transformation of VSMCs [33,34]. In high-phosphate environments, β -catenin translocases to the nucleus, inducing RUNX2 expression and driving calcification, particularly in end-stage renal disease (ESRD) patients [34]. High phosphate also regulates Pit-1 transcription via this pathway, accelerating CAC [35]. Deleting peroxisome proliferator-activated receptor gamma (PPAR γ) promotes vascular calcification through a Wnt5a-driven chondrogenic pathway, while PPAR γ protects against calcification by inducing the Wnt antagonist secreted frizzled-related protein 2 (sFRP2) [36]. Osteocalcin (OCN) and low-density lipoprotein receptor-related protein 8 (LRP8) further enhance this process, while Moscatilin inhibits vascular calcification by suppressing the Wnt/ β -catenin pathway [37].

Lipoprotein a (Lp(a)) promotes CAC by increasing calcific deposits, alkaline phosphatase (ALP) activity, and pro-calcific proteins like bone morphogenetic protein 2 (BMP2) and OPN [38]. Lp(a) also stimulates VSMC mineralization and endothelial-to-mesenchymal transition (EndMT), which contributes to calcification via the Notch1-BMP2-Smad1/5/9 signaling pathway [39]. Inhibition of Notch1 reduces BMP2 expression, ALP activity, and calcification [40]. Lp(a) also activates the nuclear factor- κ B (NF- κ B) pathway, enhancing inflammation by upregulating OPN and cytokines like interleukin (IL) IL-1 β and IL-6 [38]. Clinical studies link higher Notch1 and OPN levels to CAC and show Lp(a) associated with increased CAC volume in individuals with inflammation [41,42]. Finally, adiponectin inhibits osteoblastic differentiation in VSMCs via the AMPK pathway, while indoxyl sulfate promotes CAC by suppressing Notch signaling [43], and notch1-Msx2 interactions play a key role in early-stage vascular calcification, independent of BMP-2 and RUNX2 pathways [44,45].

4. Inflammation and Coronary Artery Calcification

Chronic inflammation, driven by factors like oxidative stress, cytokines (e.g., C-reactive protein [CRP], Tumor necrosis factor- α (TNF- α), and interleukins (IL-1 β , IL-6, IL-18), plays a key role in CAC and age-related diseases [46]. Pro-inflammatory cytokines (e.g., IL-6, IL-8, TNF- α , monocyte chemoattractant protein-1 [MCP-1]) activate pathways that promote VSMC osteogenic transformation [47–49]. IL-18 enhances CAC via the ERK1/2 pathway, while TNF- α and IL-6 upregulate VSMC calcification through the AP-1/c-FOS pathway [50]. IL-29 also promotes CAC via JAK2/STAT3 signaling, increasing BMP2 expression [51]. Although some studies link inflammatory markers to CAC, their association is weak after adjusting for traditional risk factors [52]. Additionally, ER stress contributes to CAC by promoting extracellular vesicle release and interacting with iron homeostasis and mitochondrial dysfunction [53]. Lower serum levels of netrin-1 and gremlin-1, alongside higher TNF- α , may serve as early diagnostic markers for subclinical CAC [54].

Atherosclerotic plaque calcification is regulated by the receptor activator of the nuclear factor-kappa B ligand (RANKL)/OPG system, affecting both calcium deposition and plaque stability [55]. Key pathways like Notch, Wnt, and TGF-β/BMP control osteogenic differentiation and inflammation, involving macrophage activation and VSMC proliferation [56]. Ferroptosis and calcification are interconnected via Toll-like receptor 4 (TLR4) and the NF-κB axis [57]. Tetramethylpyrazine (TMP) reduces CAC by inhibiting caspase-3/GSDME-mediated pyroptosis, lowering inflammation, calcification, and oxida-

tive stress in VSMCs [58]. Clinical studies reveal that macrophage-rich plaques have a higher calcification burden, making them more vulnerable, highlighting that inflammation in conditions like rheumatoid arthritis and non-alcoholic fatty liver disease increases CAC risk [59,60]. Of note, elevated interleukin-2 receptor (IL-2R) levels in CAD patients and IL-6 in CKD are linked to CAC severity, underscoring chronic inflammation's role in CAC development and the need for early diagnosis [61,62] (Figure 1).

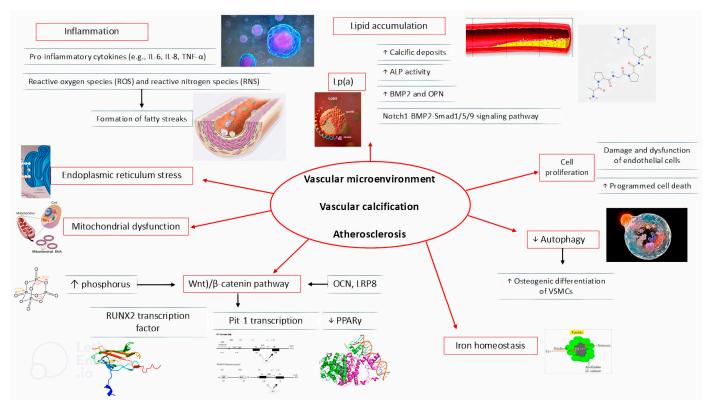


Figure 1. The mechanistic pathways leading to coronary artery calcification.

This figure illustrates the complex interplay between cellular mechanisms, inflammatory pathways, and molecular signaling contributing to coronary artery calcification (CAC). Key processes include lipid accumulation, endothelial cell dysfunction, and vascular smooth muscle cell (VSMC) osteogenic differentiation. These pathways are driven by the upregulation of pro-inflammatory cytokines, oxidative stress, and the activation of transcription factors such as RUNX2 and Wnt/ β -catenin. Collectively, these processes result in the deposition of calcific material within the vascular wall, contributing to the development and progression of atherosclerotic plaques.

ALP: Alkaline Phosphatase; BMP2: Bone Morphogenetic Protein 2; CAC: Coronary Artery Calcification; IL: Interleukin; Lp(a): Lipoprotein(a); LRP8: LDL Receptor-Related Protein 8; OCN: Osteocalcin; OPN: Osteopontin; PPAR γ : Peroxisome Proliferator-Activated Receptor Gamma; ROS: Reactive Oxygen Species; RNS: Reactive Nitrogen Species; RUNX2: Runt-Related Transcription Factor 2; TNF- α : Tumor Necrosis Factor Alpha; VSMC: Vascular Smooth Muscle Cells; Wnt: Wingless/Integrated signaling pathway.

5. Lipid Accumulation and Smooth Muscle Cell Proliferation

Atherogenesis is a chronic inflammatory process triggered by the interaction between circulating lipoproteins and subendothelial extracellular matrix molecules, specifically proteoglycans, leading to the retention of lipoproteins. Low-density lipoprotein (LDL) plays a central role, along with other apoB-containing lipoproteins smaller than 70 nm in diameter, in contributing to the formation of atherosclerotic plaques [63]. These plaques contain cholesterol, fatty substances, cellular waste products, calcium, and fibrin [64].

LDL particle concentration (LDL-P) subtypes, along with large high-density lipoprotein (HDL)-P subtypes, have been associated with CAC progression. However, after adjusting for standard risk factors and traditional lipid measurements—such as LDL cholesterol (LDL-C), HDL cholesterol (HDL-C), and triglycerides—only the medium and very small LDL-P subtypes remained significantly related to CAC progression. CAC was more prevalent in individuals with either low HDL-C or the highest HDL-P levels [65]. Additionally, elevated lipoprotein(a) [Lp(a)] levels are widely known to increase cardiovascular risk and contribute to CAC development [66,67].

According to the response-to-retention atherosclerosis model, atheroma development in the vessel wall is driven by a complex inflammatory process [68]. Reactive oxygen species (ROS) and reactive nitrogen species (RNS) oxidize lipids and LDL [69]. Oxidized LDL, concentrated in the intimal layer of the vessel, plays a crucial role in initiating and advancing atherosclerosis by damaging endothelial cells, enhancing leukocyte adhesion, and influencing the production of leukocyte and monocyte adhesion molecules (e.g., VCAM, ICAM, E-selectin, and P-selectin) in the endothelium [70,71]. This damage promotes the infiltration of the intimal vessel layer by monocytes, T cells, and mast cells [72].

Through the action of MCP-1, M-CSF, and IL-8, monocytes differentiate into macrophages, which phagocytose oxidized LDL using scavenger receptors (SR-A family) and transform into lipid-rich foam cells [73]. While foam cells were traditionally thought to originate from bone marrow-derived macrophages, recent studies suggest that vascular smooth muscle cells (VSMCs) can also convert into foam cells when exposed to aggregated or oxidized LDL. In fact, the majority of foam cells in atherosclerotic plaques may derive from VSMCs [74,75].

The accumulation of foam cells in the arterial wall leads to the formation of fatty streaks [76]. VSMCs migrate from the media to the intima, proliferating to form fibrous atherosclerotic plaques from these fatty streaks [77]. As atherosclerosis progresses, proteolytic enzymes such as metalloproteinases—produced by macrophages and T-lymphocytes—destabilize the fibrous plaque. Damage to the fibrous cap and collagen results in coagulation, thrombus formation, and ultimately, occlusion of the arteries [78]. Table 1 provides a summary of all the molecular pathways involved in CAC (Table 1).

Pathway	Key Molecular Players	Role in Calcification	
Inflammatory Pathways	IL-1β [46], TNF-α, NF-kB [47–49]	Promote VSMCs differentiation and osteogenic transformation, contributing to calcified plaques	
Oxidative Stress Pathway	ROS, NADPH oxidase [69]	Enhances inflammation and calcification through oxidative damage and VSMCs differentiation	
Osteogenic Pathways	BMPs [79,80], RANKL [55,81], RUNX2 [33,34]	Induce VSMCs transformation into osteoblast-like cells, driving calcification	
Autophagy	AMPK, mTOR, Beclin-1, b-catenin [17–20]	Dysregulated autophagy promotes calcification by increasing osteogenic differentiation	

Table 1. Molecular Pathways Involved in Coronary Artery Calcification.

AMPK: AMP-activated protein kinase; BMP: Bone Morphogenetic Protein; IL-1 β : Interleukin 1 Beta; mTOR: Mechanistic Target of Rapamycin; NF-kB: Nuclear Factor Kappa-light-chain-enhancer of activated B cells; RANKL: Receptor Activator of Nuclear Factor Kappa-B Ligand; ROS: Reactive Oxygen Species; RUNX2: Runt-related transcription factor 2; TNF- α : Tumor Necrosis Factor Alpha; VSMCs: Vascular Smooth Muscle Cells.

6. Genetic and Environmental Factors in CAC and CAD

As in many complex diseases, the risk of developing CAC and CAD is modulated by a set of genetic and environmental factors. Accessibility to human genome sequencing enables the performance of genome-wide association studies and meta-analyses studies supporting the strong genetic component that drives CAC and CAD events [82]. More than 160 loci have been associated with the risk of developing CAD. Interestingly, a great percentage of these variants are not associated with the traditional pathogenesis pathways of CAD such as lipid metabolism, blood pressure, inflammation, extracellular matrix function and structure, and vascular remodeling [83–85], supporting the existence of undiscovered pathological mechanisms. From these studies, only four loci (9p21, PHACTR1/EDN1, APOE, and APOB) have been associated with CAC [86]. Recent studies confirmed the association of these four loci with CAC and identified new loci that are implicated in VSMC calcification, bone mineralization, phosphate, and vitamin metabolism [87,88].

Additionally, rare variants association studies enable the identification of rare mutations in different genes which influence the risk of CAD. Inactivating mutations in LDLR, LPL, and APOA5 are associated with increased risk while mutations in PCSK9, NPC1L1, ASGR1, APOC3, ANGPTL4, and LPA are associated with decreased risk of CAD [89–91]. These genes are implicated in pathways related to LDL, triglyceride-rich lipoproteins, cholesterol, or lipoprotein metabolisms [92]. Arterial calcification is also connected with rare mutations in different genes. The related genes can be divided into three groups: (1) genes associated with extracellular purine/phosphate/phosphate metabolism such as ENPP1, ABCC6, PDGFRB, SLC20A2, XPR1, MYORG, and LMNA; (2) genes associated with interferonopathies such as IFIH1 and DDX58; and (3) the GBA gene which is associated with Gaucher disease, a known lysosomal storage disorder characterized by increased levels of glucosylsphingosine and glucosylceramide in various organs [93–97]. Further clinical studies deciphering the molecular mechanisms of these mutations would lead to the development of novel therapeutic options for CAC and CAD.

Not only genetic predisposition but also various environmental factors play a role in the development of CAC and CAD. The European Society of Cardiology and the American College of Cardiology guidelines state cholesterol, blood pressure, cigarette smoking, diabetes, and adiposity are the major risk factors for coronary disease [98]. Age, gender, and ethnicity have been shown to play a role in the onset of CAC [99,100]. The Multi-Ethnic Study of Atherosclerosis (MESA) study has shown, by measuring coronary calcification, that whites have the greatest CAC, followed by Chinese, Hispanics, and African Americans [101]. Other studies, such as the Heinz Nixdorf Recall [102] and Coronary Artery Risk Development in Young Adults (CARDIA) studies [103] have shown a positive correlation between smoking and CAC onset. Cigarette smoking through the production of toxic chemicals causes vascular calcification [104]. Additionally, the relationship between physical activity and CAC remains controversial. While the CARDIA study had shown that physical activity reduces the risk of having CAC [105], the MESA study failed to show this correlation [106]. Studies in athletes have shown that participants with exercise volume > 2000 MET-min/wk have a significantly higher CAC score [107]. Recently, the MARC-2 study suggests that exercise intensity but not volume is associated with coronary artery calcification progression [108]. Studies on favorable cardiovascular health highlight the importance of a healthy lifestyle and early prevention to preserve a healthy cardiovascular state and reduce the risk of developing CAD later in life [109].

7. Clinical Implications and Diagnostic Approaches

The management of CAC and its implications for CAD requires a comprehensive approach that includes early detection, advanced diagnostic methods, and the identification of potential biomarkers [110,111]. Early detection of CAC is crucial for effective risk stratification and patient management [112]. By identifying CAC at an early stage, clinicians can better assess the risk of CAD-related events and implement preventive measures [113]. Monitoring the progression of CAC over time allows for timely interventions that could slow the advancement of atherosclerosis, potentially reducing the incidence of major cardiovascular events [114]. Furthermore, regular monitoring aids in adjusting the intensity of preventive strategies, such as lifestyle modifications or pharmacological treatments, to better manage the patient's condition [115,116].

7.1. Imaging Modalities for the Detection of CAC

A variety of imaging modalities are available for detecting and quantifying CAC, each with its own advantages and limitations. Cardiac multi-slice computed tomography (MSCT) imaging remains the most commonly used imaging modality [117]. Cardiac CT is promoted by both the European Society of Cardiology [118] and the American Heart Association [119] for the non-invasive evaluation of patients with low-to-intermediate risk for CAD. Cardiac CT can be used either for the detection of CAC or for the precise visualization of coronary arteries. Coronary computed tomography angiography (CTA) is often favored over plain coronary arteries but also provides a qualitative assessment of plaque morphology by visualizing both calcified and non-calcified plaques, assessing their characteristics, and directly measuring the severity of arterial narrowing [120].

CAC with MSCT can be assessed by various scoring systems. The three primary methods for assessing coronary artery calcification by CT are the CT Calcium Scoring (CACS), Calcium Volume Score, and Calcium Mass Score. The CACS, or Agatston score, is widely used and provides a simple, standardized measure of calcified plaque burden but focuses only on calcified plaques and can be influenced by image noise [121]. The CACS offers a valuable measure of calcified plaque burden and helps predict cardiovascular risk [122]. This method is highly predictive of future cardiovascular events, making it a valuable tool in clinical practice [123]. The Calcium Volume Score assesses the total volume of calcified plaques, offering a more comprehensive evaluation and greater stability over time, though it is less commonly used and requires more detailed processing [124,125]. Finally, the determination of the calcium mass score is another option for the quantification of CAC [126]. The Calcium Mass Score combines plaque volume and density, providing a physiologically relevant measure of plaque burden that may offer better risk stratification; however, it is more complex and less widely adopted in clinical practice. Each method has unique advantages and disadvantages, making them suitable for different clinical scenarios depending on the goals of the assessment [127].

The Coronary Artery Calcium Data and Reporting System (CAC-DRS) provides a standardized framework for reporting CAC scores derived from CT scans, which are crucial in assessing the risk of cardiovascular events, particularly CAD. CAC-DRS categorizes CAC scores into four distinct groups: 0 (indicating no detectable coronary calcium), 1–99 (reflecting mild plaque buildup), 100–299 (indicating moderate calcium levels and more significant plaque), and \geq 300 (suggesting extensive calcium deposits and a high risk of cardiovascular complications) [128,129]. This stratification allows healthcare providers to accurately gauge a patient's risk and tailor prevention and treatment strategies accordingly. A higher CAC-DRS category is associated with a greater likelihood of cardiovascular events, prompting more intensive management [130]. By ensuring consistent and clear reporting of CAC scores, CAC-DRS enhances clinical decision-making and supports more effective risk assessment and intervention for patients at varying levels of cardiovascular risk [131].

CAC detection using positron emission tomography (PET)/CT scans offers a powerful combination of anatomical and functional imaging, enhancing the assessment of CAD [132]. The CT component provides high-resolution images for detecting and quantifying calcified plaques, while the PET component adds functional insight by identifying metabolically active or inflamed plaques that may not yet be calcified but pose significant risk [133]. This hybrid approach improves diagnostic accuracy and offers enhanced risk assessment by capturing both calcified and non-calcified plaques, making it particularly valuable in high-risk patients or those with complex coronary anatomy [134]. However, PET/CT is more costly, less widely available, and involves higher radiation exposure compared to standard CT, which may limit its use to specialized centers or select patient populations [135]. Despite these limitations, PET/CT's ability to integrate anatomical and metabolic information provides a comprehensive tool for guiding treatment decisions and monitoring disease progression in CAD.

Cardiac magnetic resonance imaging (CMR) is not typically utilized for the detection of CAC due to its limited sensitivity to calcium deposits [136]. CMR excels in providing detailed assessments of cardiac structure and function, offering valuable information on myocardial tissue characterization, including the detection of fibrosis, edema, and infarction. Additionally, CMR is highly effective in evaluating ventricular function, wall motion abnormalities, and myocardial perfusion, particularly under stress conditions, making it an important tool for diagnosing and managing various aspects of CAD [137]. CMR offers the advantage of avoiding ionizing radiation, making it a safer option for some patients. However, CMR does not play a direct role in detecting calcified plaques within the coronary arteries, it serves as a complementary modality alongside CT imaging by offering comprehensive insights into the functional and structural implications of CAD on the heart [138].

Intravascular imaging modalities, such as intravascular ultrasound (IVUS) and optical coherence tomography (OCT), can provide additional insights into plaque activity and coronary artery wall structure [139], though they are less frequently used for routine CAC assessment due to their invasiveness and cost. IVUS offers deeper penetration, making it effective for visualizing the entire vessel wall and assessing the overall plaque burden, including the quantification of calcification [140]. However, it has a lower spatial resolution, which may miss finer details like micro-calcifications. On the other hand, OCT provides much higher resolution, allowing for detailed visualization of plaque morphology and precise delineation of calcified plaques, though it has limited penetration depth and requires blood clearance during imaging [141]. The choice between IVUS and OCT depends on the specific clinical scenario, with IVUS being better for assessing deeper or more extensive calcifications, while OCT is superior for high-resolution imaging of plaque details. Often, these modalities are used complementarily to provide a comprehensive assessment of coronary artery disease.

In developing a comprehensive diagnostic approach for CAC, it is essential to integrate the various imaging modalities by assessing their strengths according to the specific clinical scenario (Table 2). For instance, CTA and CACS are often first-line choices for non-invasive evaluation due to their accessibility and ability to provide both anatomical and, in the case of CTA, functional insights. PET/CT can be reserved for high-risk patients or those with complex coronary anatomy, where the combined anatomical and metabolic information can guide more nuanced therapeutic decisions. Intravascular modalities like IVUS and OCT, while more invasive, are particularly valuable during interventional procedures where detailed plaque characterization is critical for optimal stent placement. Patient characteristics play a crucial role in modality selection; for example, CMR might be preferred in patients requiring frequent imaging due to its lack of ionizing radiation, while renal function considerations may influence the use of contrast-enhanced studies like CTA. Ultimately, the choice of imaging modality should be tailored to the patient's risk profile, clinical needs, and specific diagnostic or therapeutic goals, ensuring a personalized and effective approach to cardiovascular care.

Imaging Modality	Pros	Cons	Key Characteristic
Cardiac Multi-Slice CT (MSCT) [118,119]	Most common modality for CAC detection	Focuses mainly on calcified plaques	Broad availability and relatively low cost of this modality.
	Supported by the European Society of Cardiology and the American Heart Association	Can be influenced by image noise	
	Non-invasive evaluation of coronary arteries	Higher radiation exposure	
	Quantifies degree of stenosis	Requires the use of contrast agents	

Table 2. Comparison of Imaging Modalities for Coronary Artery Calcification (CAC) Detection.

Imaging Modality	Pros	Cons	Key Characteristic	
CT Calcium Scoring (CACS) [122,123]	Simple, standardized measure of calcified plaque burden	Only assesses calcified plaques	Important role in routine clinical practice for risk stratification.	
	Highly predictive of future cardiovascular events	Can be affected by image noise		
Calcium Volume Score [124,125]	Assesses total volume of calcified plaques	Less commonly used	Useful for tracking changes in	
	Offers greater stability over time	Requires more detailed processing	plaque burden over time.	
Calcium Mass Score [126]	Combines plaque volume and density	More complex to calculate	More valuable in research settings or for detailed patient evaluations.	
	Provides a physiologically relevant measure	Less widely adopted in clinical practice		
	Potentially better for risk stratification			
PET/CT scans [132–135]	Combines anatomical and functional imaging	More costly	Important role in guiding therapeutic decisions, especially in complex cases.	
	Enhances diagnostic accuracy by identifying metabolically active plaques	Less widely available		
	Particularly valuable in high-risk patients	Higher radiation exposure		
Cardiac Magnetic	Provides detailed assessments of cardiac structure and function	Limited sensitivity to calcium	Important role as a complementary modality,	
Resonance Imaging	No ionizing radiation	Not typically used for	especially for patients requiring	
(CMR) [136]	Effective for evaluating myocardial tissue	detecting calcification	frequent imaging without radiation exposure.	
Intravascular Ultrasound (IVUS) [139,140]	Deeper penetration into vessel wall	Lower spatial resolution	Invasive method typically reserved for specific clinical scenarios or interventional procedures.	
	Effective for assessing overall plaque burden	May miss fine details like micro-calcifications		
	Useful for quantifying calcification	Invasive procedure		
Optical Coherence Tomography (OCT) [141]	High spatial resolution	Limited penetration depth		
	Detailed visualization of plaque morphology	Requires blood clearance	Important role in pre-intervention planning, particularly when fine detail is crucial.	
	Precise delineation of calcified plaques	Invasive and costly		

Table 2. Cont.

7.2. Biomarkers for the Detection of CAC

The search for reliable biomarkers to enhance risk assessment for CAC and CAD is an ongoing area of research. Inflammatory markers such as CRP, soluble intercellular adhesion molecule- 1 (sICAM-1), and fibrinogen have been associated with increased CAC and may serve as indicators of active disease [142]. CRP is significantly associated with CAC progression among clinical parameters [143] and can be used to predict the risk of an elevated coronary artery calcium score [144]. However, a recent large meta-analysis did not show the significant role of CRP in risk stratification for CAC scores [145]. IL-6 has been found to be associated with the progression of CAC in patients with chronic renal dysfunction on dialysis [146]. IL-6 is positively associated with vascular calcification, promoting inflammation and contributing to the progression of calcification in vascular tissues [147].

Lipid-related biomarkers, including LDL-C and ApoB, are central to the process of atherosclerosis and could help predict the likelihood of calcification [148,149]. The atherogenic index of plasma (AIP), an indicator calculated based on TG levels and HDL-C levels, has been proposed to assess the degree of CAC, showing greater predictive power for atherosclerosis and cardiovascular events [150]. AIP has been found to be an independent predictor of CAD and has been used as a risk factor for CAC and CVD [151,152].

Genetic markers, particularly specific polymorphisms, are also being investigated for their potential to identify individuals at higher risk for CAC [153]. Recently, it has been described that genetic polymorphisms on the NPC1L1 gene were associated with high-degree CAC in male patients with premature CAD [154]. Additionally, emerging biomarkers like MGP, which inhibits vascular calcification, adiponectin which is inversely associated with vascular calcification, suggesting a protective role against the progression of calcification in vascular tissues, and OPG, which is involved in bone metabolism, are being studied for their roles in vascular health and may offer new opportunities for assessing CAC risk [155]. Low plasma adiponectin levels [156] and elevated plasma OPG [157] are associated with the progression of CAC and aortic plaque, while MGP has been proposed as a major factor in the development of vascular calcification [158]. Fetuin-A is a glycoprotein that plays a crucial role in inhibiting calcium deposition in blood vessels [159]. Lower levels of Fetuin-A are associated with increased vascular calcification, making it a potential biomarker for assessing cardiovascular risk [160]. Finally, BMPs, and especially BMP-2, are key regulators of bone formation and are also involved in the pathological process of vascular calcification [79]. BMP-2 promotes the osteogenic differentiation of vascular smooth muscle cells, leading them to adopt bone-like properties and contribute to the deposition of calcium in the arterial walls. Elevated levels of BMP-2 have been linked to increased calcification, making it an important biomarker and potential therapeutic target in cardiovascular disease [80].

The integration of early detection with advanced imaging techniques and biomarker research represents a comprehensive approach to managing CAC and CAD. By combining these strategies, clinicians can better identify high-risk patients, monitor disease progression, and tailor interventions to prevent adverse cardiovascular outcomes. The continued exploration of these diagnostic approaches and biomarkers is essential for enhancing the overall management of CAD associated with CAC, ultimately improving patient care and outcomes.

8. Therapeutic Strategies Targeting CAC

Pharmacological strategies for managing CAC primarily focus on reducing the progression of calcification through lipid-lowering agents, anti-inflammatory drugs, and agents that modulate mineral metabolism [161]. These interventions aim to stabilize plaques, reduce cardiovascular risk, and potentially slow the calcification process (Table 3). Calcium channel blockers (CCBs) and renin-angiotensin system (RAS) inhibitors play significant roles in modulating vascular calcification. CCBs, by preventing calcium influx into VSMCs, help reduce the cellular processes that contribute to calcification, potentially slowing the progression of arterial stiffness [162,163]. RAS inhibitors, such as ACE inhibitors and angiotensin II receptor blockers (ARBs), reduce the effects of angiotensin II, a key driver of vascular calcification through its promotion of inflammation, oxidative stress, and osteogenic differentiation of vascular smooth muscle cells [164,165]. By inhibiting these pathways, both CCBs [166,167] and RAS inhibitors [168,169] offer therapeutic potential in reducing vascular calcification and the associated cardiovascular risks, particularly in patients with hypertension or chronic kidney disease.

Statins are the cornerstone of pharmacological therapy for CAC due to their dual action of significantly lowering LDL-C levels and exerting potent anti-inflammatory effects, both of which are crucial in reducing the progression of calcification [170]. By decreasing LDL-C, statins help reduce the lipid core of atherosclerotic plaques, thereby stabilizing them and preventing further calcification. Moreover, their anti-inflammatory properties play a

key role in mitigating the inflammatory processes that contribute to vascular calcification resulting in plaque stabilization [171]. Furthermore, statins can modulate the calcification of VSMCs, providing further evidence of their beneficial effects beyond cholesterol reduction [172]. Clinical studies have demonstrated that statin therapy is associated with a slower progression of calcified plaque burden, underscoring their importance in managing CAD [173–175]. These findings collectively reinforce the critical role of statins in not only managing lipid levels but also in directly influencing the progression of CAC, making them a fundamental component of CVD management.

Anti-inflammatory drugs, such as colchicine, are gaining attention for their potential role in reducing CAC by targeting the underlying inflammatory processes that contribute to plaque instability and calcification [176]. Colchicine, traditionally used for gout, has been shown in recent studies to reduce cardiovascular events [177,178], potentially by mitigating inflammation within the arterial walls [179]. In addition to anti-inflammatory agents, novel therapies targeting specific pathways involved in calcification, such as PCSK9 inhibitors and RANKL inhibitors, are also being explored. PCSK9 inhibitors, initially developed to lower LDL cholesterol levels, have shown potential in reducing coronary plaque volume and may also play a role in limiting calcification within the arterial walls. Inhibitors like evolocumab and alirocumab, by reducing LDL and lipoprotein(a), can mitigate calcific progression [180,181]. Additionally, calcification inhibitor therapies are promising areas of exploration. These interventions target molecular pathways involved in osteogenic differentiation of VSMCs, such as the Wnt/ β -catenin and BMP signaling pathways [182]. RANKL inhibitors, which interfere with the osteogenic pathways that drive vascular calcification, offer a promising approach to directly targeting the calcification process [81]. In addition to traditional therapeutic strategies, natural compounds such as resveratrol have shown promise in preventing atherosclerosis progression. For instance, Sirasanagandla et al. demonstrated that maternal resveratrol supplementation ameliorates bisphenol A-induced atherosclerotic lesion formation in offspring, suggesting that dietary intake of foods rich in resveratrol could reduce future cardiovascular risk [183]. These emerging therapies represent a significant advancement in the potential treatment options for patients at risk of CAC, offering hope for more effective management strategies in the future.

Therapeutic Approach	Target Pathway	Mechanism of Action	Potential Clinical Impact
Statins [170–175]	Lipid metabolism	Reduces cholesterol and inflammation, indirectly inhibiting calcification	Lower cardiovascular event risk
PCSK9 Inhibitors [180,181]	Lipid metabolism	Lowers LDL cholesterol, potentially influencing calcification	Reduces plaque progression and improves outcomes
Vitamin K [184,185]	Calcium metabolism	Activates MGP to inhibit vascular calcification	Prevents progression of calcified plaques
RANKL Inhibitors [81]	Osteogenic pathway	Blocks osteoclast differentiation, limiting calcification	May reduce the progression of coronary artery calcification
Omega-3 Fatty Acids (EPA) [186,187]	Inflammation	Suppresses inflammatory pathways and inhibits calcification	Potential role in reducing coronary artery calcification

Table 3. Therapeutic Approaches Targeting Calcification Pathways.

EPA: Eicosapentaenoic Acid; LDL: Low-Density Lipoprotein; MGP: Matrix Gla Protein; PCSK9: Proprotein Convertase Subtilisin/Kexin Type 9; RANKL: Receptor Activator of Nuclear Factor Kappa-B Ligand.

Lifestyle interventions, including regular physical activity, smoking cessation, and a heart-healthy diet rich in fruits, vegetables, and whole grains, are essential components of CAC management [188]. Dietary modifications, particularly those reducing saturated fats and increasing omega-3 fatty acids, can lower inflammation and lipid levels, indirectly influencing calcification progression [186]. Specifically, eicosapentaenoic acid (EPA), an omega-3 polyunsaturated fatty acid found in fatty fish and fish oils, has been reported to directly inhibit arterial calcification [187]. Vitamin K supplementation plays a crucial role in

the modulation of vascular calcification as it is essential for the activation of MGP and the overall stimulation of osteoblastogenesis [189]. Supplementing with Vitamin K, particularly in individuals with low dietary intake or those at risk for vascular calcification, may help reduce the progression of calcification and improve cardiovascular outcomes [184]. Finally, magnesium supplements may be associated with less vascular calcification [185]. Of note, increased dietary calcium intake has not been associated with an increased risk of CAC, suggesting that calcium consumed through diet does not contribute to the development of calcification therapies could be valuable not only as preventive measures for the general population but also particularly beneficial for patients with low bone turnover conditions, such as those with osteoporosis or a significant number of individuals with CKD.

Future therapeutic strategies for CAC are likely to focus on more targeted interventions that specifically inhibit the molecular pathways responsible for calcification. One promising area of research involves the development of therapies that target osteogenic differentiation in VSMCs [192–194]. This process, where VSMCs undergo a transformation into osteoblast-like cells, is a key driver of vascular calcification [195]. Inhibitors of this differentiation process could potentially prevent or reduce calcification, offering a more direct approach to managing CAC. For instance, research into inhibitors of the BMP pathway, which is heavily involved in osteogenic differentiation, is showing potential in preclinical studies [196].

Additionally, advancements in precision medicine are paving the way for more personalized therapeutic approaches. By leveraging genetic and biochemical profiling, future therapies could be tailored to an individual's specific risk factors and underlying biological mechanisms driving calcification [197]. For example, individuals with specific genetic polymorphisms that influence calcium metabolism, or inflammatory pathways could benefit from customized treatments that target these specific processes. Precision medicine also holds promise in identifying patients who may respond particularly well to novel therapies, such as PCSK9 inhibitors or RANKL inhibitors, further optimizing the management of CAD and CAC [198,199].

These approaches, supported by ongoing research and clinical trials (NCT05720156, NCT04889053, NCT05482399, and NCT05259046) suggest a future where the treatment of CAC is not only more effective but also highly individualized, addressing the unique needs of each patient based on their genetic and molecular profiles [200]. As our understanding of the molecular mechanisms underlying CAC deepens, these targeted and personalized therapies are expected to play a crucial role in the prevention and treatment of coronary artery disease.

9. Conclusions

CAC is a complex process driven by multiple molecular and cellular pathways, including inflammation, oxidative stress, and osteogenic differentiation of vascular smooth muscle cells. As a significant contributor to CAD, CAC not only serves as a marker of advanced atherosclerosis but also plays a critical role in plaque stability and progression. Current research efforts are aimed at understanding the intricate mechanisms that regulate calcification and exploring therapeutic interventions to slow or reverse this process. The integration of advanced imaging techniques and personalized medicine offers new hope for risk stratification and tailored treatment strategies. As the molecular underpinnings of CAC continue to be uncovered, innovative therapeutic approaches targeting specific calcification pathways could lead to better clinical outcomes in CAD patients.

While this review aims to provide a comprehensive overview of the molecular mechanisms underlying CAC, there are inherent limitations. The narrative nature of the review may result in selection bias, as studies with inconclusive or negative results may be underrepresented. Additionally, emerging molecular pathways not yet extensively studied could present new therapeutic targets. Future research should focus on large-scale, multi-center studies that validate the clinical applicability of the pathways discussed here. Investigating the role of novel compounds, such as anti-inflammatory agents and calcification inhibitors, in clinical trials would be a valuable next step in targeting CAC.

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References

- GBD 2017 Causes of Death Collaborators. Global, Regional, and National Age-Sex-Specific Mortality for 282 Causes of Death in 195 Countries and Territories, 1980–2017: A Systematic Analysis for the Global Burden of Disease Study 2017. *Lancet* 2018, 392, 1736–1788. [CrossRef] [PubMed]
- Liu, W.; Zhang, Y.; Yu, C.-M.; Ji, Q.-W.; Cai, M.; Zhao, Y.-X.; Zhou, Y.-J. Current Understanding of Coronary Artery Calcification. J. Geriatr. Cardiol. 2015, 12, 668–675. [CrossRef] [PubMed]
- 3. Mitsis, A.; Gragnano, F. Myocardial Infarction with and without ST-Segment Elevation: A Contemporary Reappraisal of Similarities and Differences. *Curr. Cardiol. Rev.* 2021, *17*, e230421189013. [CrossRef] [PubMed]
- 4. Onnis, C.; Virmani, R.; Kawai, K.; Nardi, V.; Lerman, A.; Cademartiri, F.; Scicolone, R.; Boi, A.; Congiu, T.; Faa, G.; et al. Coronary Artery Calcification: Current Concepts and Clinical Implications. *Circulation* **2024**, *149*, 251–266. [CrossRef] [PubMed]
- Rattazzi, M.; Bertacco, E.; Puato, M.; Faggin, E.; Pauletto, P. Hypertension and Vascular Calcification: A Vicious Cycle? J. Hypertens. 2012, 30, 1885–1893. [CrossRef]
- Nicoll, R.; Zhao, Y.; Ibrahimi, P.; Olivecrona, G.; Henein, M. Diabetes and Hypertension Consistently Predict the Presence and Extent of Coronary Artery Calcification in Symptomatic Patients: A Systematic Review and Meta-Analysis. *Int. J. Mol. Sci.* 2016, 17, 1481. [CrossRef]
- 7. Dai, Z.; Zhang, X. Pathophysiology and Clinical Impacts of Chronic Kidney Disease on Coronary Artery Calcification. J. Cardiovasc. Dev. Dis. 2023, 10, 207. [CrossRef]
- 8. Fantus, D.; Awan, Z.; Seidah, N.G.; Genest, J. Aortic Calcification: Novel Insights from Familial Hypercholesterolemia and Potential Role for the Low-Density Lipoprotein Receptor. *Atherosclerosis* **2013**, *226*, 9–15. [CrossRef]
- 9. Nakahara, T.; Dweck, M.R.; Narula, N.; Pisapia, D.; Narula, J.; Strauss, H.W. Coronary Artery Calcification. *JACC Cardiovasc. Imaging* **2017**, *10*, 582–593. [CrossRef]
- 10. Proudfoot, D.; Shanahan, C.M. Biology of Calcification in Vascular Cells: Intima versus Media. Herz 2001, 26, 245–251. [CrossRef]
- 11. Zhou, X.; Xu, S.-N.; Yuan, S.-T.; Lei, X.; Sun, X.; Xing, L.; Li, H.-J.; He, C.-X.; Qin, W.; Zhao, D.; et al. Multiple Functions of Autophagy in Vascular Calcification. *Cell Biosci.* 2021, *11*, 159. [CrossRef] [PubMed]
- 12. Li, M.; Zhu, Y.; Jaiswal, S.K.; Liu, N.-F. Mitochondria Homeostasis and Vascular Medial Calcification. *Calcif. Tissue Int.* **2021**, 109, 113–120. [CrossRef] [PubMed]
- 13. Ribeiro-Silva, J.C.; Nolasco, P.; Krieger, J.E.; Miyakawa, A.A. Dynamic Crosstalk between Vascular Smooth Muscle Cells and the Aged Extracellular Matrix. *Int. J. Mol. Sci.* 2021, 22, 10175. [CrossRef] [PubMed]
- Lee, S.J.; Lee, I.-K.; Jeon, J.-H. Vascular Calcification—New Insights into Its Mechanism. Int. J. Mol. Sci. 2020, 21, 2685. [CrossRef] [PubMed]
- 15. Peng, Y.-Q.; Xiong, D.; Lin, X.; Cui, R.-R.; Xu, F.; Zhong, J.-Y.; Zhu, T.; Wu, F.; Mao, M.-Z.; Liao, X.-B.; et al. Oestrogen Inhibits Arterial Calcification by Promoting Autophagy. *Sci. Rep.* **2017**, *7*, 3549. [CrossRef]
- 16. Durham, A.L.; Speer, M.Y.; Scatena, M.; Giachelli, C.M.; Shanahan, C.M. Role of Smooth Muscle Cells in Vascular Calcification: Implications in Atherosclerosis and Arterial Stiffness. *Cardiovasc. Res.* **2018**, *114*, 590–600. [CrossRef]
- Yang, R.; Zhu, Y.; Wang, Y.; Ma, W.; Han, X.; Wang, X.; Liu, N. HIF-1α/PDK4/Autophagy Pathway Protects against Advanced Glycation End-Products Induced Vascular Smooth Muscle Cell Calcification. *Biochem. Biophys. Res. Commun.* 2019, 517, 470–476. [CrossRef]
- 18. Chen, K.; Zhou, X.; Sun, Z. Haplodeficiency of *Klotho* Gene Causes Arterial Stiffening via Upregulation of Scleraxis Expression and Induction of Autophagy. *Hypertension* **2015**, *66*, 1006–1013. [CrossRef]
- 19. Lanzer, P.; Boehm, M.; Sorribas, V.; Thiriet, M.; Janzen, J.; Zeller, T.; St Hilaire, C.; Shanahan, C. Medial Vascular Calcification Revisited: Review and Perspectives. *Eur. Heart J.* **2014**, *35*, 1515–1525. [CrossRef]
- 20. Lee, K.; Kim, H.; Jeong, D. Microtubule Stabilization Attenuates Vascular Calcification through the Inhibition of Osteogenic Signaling and Matrix Vesicle Release. *Biochem. Biophys. Res. Commun.* **2014**, 451, 436–441. [CrossRef]

- Zou, Y.; Li, D.; Guan, G.; Liu, W. Phosphoglycerate Dehydrogenase Overexpression Inhibits Ferroptosis to Repress Calcification of Human Coronary Artery Vascular Smooth Muscle Cells via the P53/SLC7A11 Pathway. Int. J. Gen. Med. 2024, 17, 3673–3687. [CrossRef] [PubMed]
- Ouyang, L.; Yu, C.; Xie, Z.; Su, X.; Xu, Z.; Song, P.; Li, J.; Huang, H.; Ding, Y.; Zou, M.-H. Indoleamine 2,3-Dioxygenase 1 Deletion-Mediated Kynurenine Insufficiency in Vascular Smooth Muscle Cells Exacerbates Arterial Calcification. *Circulation* 2022, 145, 1784–1798. [CrossRef] [PubMed]
- Liu, D.; Cui, X.; Lu, R.; Hu, H.; Gu, G. CTRP3 Is a Coronary Artery Calcification Biomarker and Protects against Vascular Calcification by Inhibiting β-Catenin Nuclear Translocation to Prevent Vascular Smooth Muscle Cell Osteogenic Differentiation. *J. Cardiol.* 2022, 79, 551–558. [CrossRef] [PubMed]
- Xu, S.-N.; Zhou, X.; Zhu, C.-J.; Qin, W.; Zhu, J.; Zhang, K.-L.; Li, H.-J.; Xing, L.; Lian, K.; Li, C.-X.; et al. Nε-Carboxymethyl-Lysine Deteriorates Vascular Calcification in Diabetic Atherosclerosis Induced by Vascular Smooth Muscle Cell-Derived Foam Cells. *Front. Pharmacol.* 2020, *11*, 626. [CrossRef] [PubMed]
- 25. Guzman, R.J. Clinical, Cellular, and Molecular Aspects of Arterial Calcification. J. Vasc. Surg. 2007, 45 (Suppl. SA), A57–A63. [CrossRef]
- Kadoglou, N.P.; Stasinopoulou, M.; Velidakis, N.; Khattab, E.; Christodoulou, E.; Gkougkoudi, E.; Valsami, G. The Complex Mechanisms and the Potential Effects of Statins on Vascular Calcification: A Narrative Review. *Rev. Cardiovasc. Med.* 2024, 25, 51. [CrossRef]
- Dekker, M.; Waissi, F.; Silvis, M.J.M.; Bennekom, J.V.; Schoneveld, A.H.; de Winter, R.J.; Isgum, I.; Lessmann, N.; Velthuis, B.K.; Pasterkamp, G.; et al. High Levels of Osteoprotegerin Are Associated with Coronary Artery Calcification in Patients Suspected of a Chronic Coronary Syndrome. *Sci. Rep.* 2021, *11*, 18946. [CrossRef]
- Bakhshian Nik, A.; Ng, H.H.; Ashbrook, S.K.; Sun, P.; Iacoviello, F.; Shearing, P.R.; Bertazzo, S.; Mero, D.; Khomtchouk, B.B.; Hutcheson, J.D. Epidermal Growth Factor Receptor Inhibition Prevents Vascular Calcifying Extracellular Vesicle Biogenesis. *Am. J. Physiol. Heart Circ. Physiol.* 2023, 324, H553–H570. [CrossRef]
- 29. Kadoglou, N.P.E.; Khattab, E.; Velidakis, N.; Gkougkoudi, E. The Role of Osteopontin in Atherosclerosis and Its Clinical Manifestations (Atherosclerotic Cardiovascular Diseases)-A Narrative Review. *Biomedicines* **2023**, *11*, 3178. [CrossRef]
- 30. Wang, Z.; Li, L.; Du, R.; Yan, J.; Liu, N.; Yuan, W.; Jiang, Y.; Xu, S.; Ye, F.; Yuan, G.; et al. CML/RAGE Signal Induces Calcification Cascade in Diabetes. *Diabetol. Metab. Syndr.* 2016, *8*, 83. [CrossRef]
- Qin, Z.; Li, Y.; Li, J.; Jiang, L.; Zhang, Z.; Chang, K.; Yang, Q.; Chen, S.; Liao, R.; Su, B. Exosomal STAT1 Derived from High Phosphorus-stimulated Vascular Endothelial Cells Induces Vascular Smooth Muscle Cell Calcification via the Wnt/B-catenin Signaling Pathway. *Int. J. Mol. Med.* 2022, 50, 139. [CrossRef] [PubMed]
- 32. Khan, K.; Yu, B.; Tardif, J.-C.; Rhéaume, E.; Al-Kindi, H.; Filimon, S.; Pop, C.; Genest, J.; Cecere, R.; Schwertani, A. Significance of the Wnt Signaling Pathway in Coronary Artery Atherosclerosis. *Front. Cardiovasc. Med.* **2024**, *11*, 1360380. [CrossRef] [PubMed]
- Voelkl, J.; Tuffaha, R.; Luong, T.T.D.; Zickler, D.; Masyout, J.; Feger, M.; Verheyen, N.; Blaschke, F.; Kuro-O, M.; Tomaschitz, A.; et al. Zinc Inhibits Phosphate-Induced Vascular Calcification through TNFAIP3-Mediated Suppression of NF-κB. *J. Am. Soc. Nephrol.* 2018, 29, 1636–1648. [CrossRef] [PubMed]
- Leopold, J.A. Vascular Calcification: Mechanisms of Vascular Smooth Muscle Cell Calcification. *Trends Cardiovasc. Med.* 2015, 25, 267–274. [CrossRef] [PubMed]
- Zhang, D.; Bi, X.; Liu, Y.; Huang, Y.; Xiong, J.; Xu, X.; Xiao, T.; Yu, Y.; Jiang, W.; Huang, Y.; et al. High Phosphate-Induced Calcification of Vascular Smooth Muscle Cells Is Associated with the TLR4/NF-Kb Signaling Pathway. *Kidney Blood Press. Res.* 2017, 42, 1205–1215. [CrossRef]
- Woldt, E.; Terrand, J.; Mlih, M.; Matz, R.L.; Bruban, V.; Coudane, F.; Foppolo, S.; El Asmar, Z.; Chollet, M.E.; Ninio, E.; et al. The Nuclear Hormone Receptor PPARγ Counteracts Vascular Calcification by Inhibiting Wnt5a Signalling in Vascular Smooth Muscle Cells. *Nat. Commun.* 2012, *3*, 1077. [CrossRef]
- Rashdan, N.A.; Sim, A.M.; Cui, L.; Phadwal, K.; Roberts, F.L.; Carter, R.; Ozdemir, D.D.; Hohenstein, P.; Hung, J.; Kaczynski, J.; et al. Osteocalcin Regulates Arterial Calcification Via Altered Wnt Signaling and Glucose Metabolism. *J. Bone Miner. Res.* 2020, 35, 357–367. [CrossRef]
- 38. Peng, J.; Liu, M.-M.; Liu, H.-H.; Xu, R.-X.; Zhu, C.-G.; Guo, Y.-L.; Wu, N.-Q.; Dong, Q.; Cui, C.-J.; Li, J.-J. Lipoprotein (a)-Mediated Vascular Calcification: Population-Based and in Vitro Studies. *Metabolism* **2022**, *127*, 154960. [CrossRef]
- Hsu, J.J.; Tintut, Y.; Demer, L.L. Regulation of Cardiovascular Calcification by Lipids and Lipoproteins. *Curr. Opin. Lipidol.* 2022, 33, 289–294. [CrossRef]
- Shimizu, T.; Tanaka, T.; Iso, T.; Matsui, H.; Ooyama, Y.; Kawai-Kowase, K.; Arai, M.; Kurabayashi, M. Notch Signaling Pathway Enhances Bone Morphogenetic Protein 2 (BMP2) Responsiveness of Msx2 Gene to Induce Osteogenic Differentiation and Mineralization of Vascular Smooth Muscle Cells. J. Biol. Chem. 2011, 286, 19138–19148. [CrossRef]
- Ong, K.L.; McClelland, R.L.; Allison, M.A.; Cushman, M.; Garg, P.K.; Tsai, M.Y.; Rye, K.-A.; Tabet, F. Lipoprotein (a) and Coronary Artery Calcification: Prospective Study Assessing Interactions with Other Risk Factors. *Metabolism* 2021, 116, 154706. [CrossRef] [PubMed]
- Greif, M.; Arnoldt, T.; von Ziegler, F.; Ruemmler, J.; Becker, C.; Wakili, R.; D'Anastasi, M.; Schenzle, J.; Leber, A.W.; Becker, A. Lipoprotein (a) Is Independently Correlated with Coronary Artery Calcification. *Eur. J. Intern. Med.* 2013, 24, 75–79. [CrossRef] [PubMed]
- 43. Zhan, J.-K.; Wang, Y.-J.; Wang, Y.; Tang, Z.-Y.; Tan, P.; Huang, W.; Liu, Y.-S. Adiponectin Attenuates the Osteoblastic Differentiation of Vascular Smooth Muscle Cells through the AMPK/mTOR Pathway. *Exp. Cell Res.* **2014**, *323*, 352–358. [CrossRef] [PubMed]
- 44. Mahmoud, A.M.; Jones, A.M.; Sidgwick, G.P.; Arafat, A.M.; Alexander, Y.M.; Wilkinson, F.L. Small Molecule Glycomimetics Inhibit Vascular Calcification via C-Met/Notch3/HES1 Signalling. *Cell Physiol. Biochem.* **2019**, *53*, 323–336. [CrossRef] [PubMed]

- Shimizu, T.; Tanaka, T.; Iso, T.; Doi, H.; Sato, H.; Kawai-Kowase, K.; Arai, M.; Kurabayashi, M. Notch Signaling Induces Osteogenic Differentiation and Mineralization of Vascular Smooth Muscle Cells: Role of Msx2 Gene Induction via Notch-RBP-Jk Signaling. *Arterioscler. Thromb. Vasc. Biol.* 2009, 29, 1104–1111. [CrossRef]
- Choi, S.R.; Lee, Y.-K.; Cho, A.J.; Park, H.C.; Han, C.H.; Choi, M.-J.; Koo, J.-R.; Yoon, J.-W.; Noh, J.W. Malnutrition, Inflammation, Progression of Vascular Calcification and Survival: Inter-Relationships in Hemodialysis Patients. *PLoS ONE* 2019, 14, e0216415. [CrossRef]
- Reilly, M.P.; Wolfe, M.L.; Localio, A.R.; Rader, D.J. Study of Inherited Risk of Coronary Atherosclerosis C-Reactive Protein and Coronary Artery Calcification: The Study of Inherited Risk of Coronary Atherosclerosis (SIRCA). *Arterioscler. Thromb. Vasc. Biol.* 2003, 23, 1851–1856. [CrossRef]
- 48. Yu, K.; Min, X.; Lin, Y.; Huang, Y.; Huang, S.; Liu, L.; Peng, Y.; Meng, K.; Li, D.; Ji, Q.; et al. Increased IL-37 Concentrations in Patients with Arterial Calcification. *Clin. Chim. Acta* **2016**, *461*, 19–24. [CrossRef]
- 49. Raaz-Schrauder, D.; Klinghammer, L.; Baum, C.; Frank, T.; Lewczuk, P.; Achenbach, S.; Cicha, I.; Stumpf, C.; Wiltfang, J.; Kornhuber, J.; et al. Association of Systemic Inflammation Markers with the Presence and Extent of Coronary Artery Calcification. *Cytokine* **2012**, *57*, 251–257. [CrossRef]
- Zhang, K.; Zhang, Y.; Feng, W.; Chen, R.; Chen, J.; Touyz, R.M.; Wang, J.; Huang, H. Interleukin-18 Enhances Vascular Calcification and Osteogenic Differentiation of Vascular Smooth Muscle Cells Through TRPM7 Activation. *Arterioscler. Thromb. Vasc. Biol.* 2017, 37, 1933–1943. [CrossRef]
- Hao, N.; Zhou, Z.; Zhang, F.; Li, Y.; Hu, R.; Zou, J.; Zheng, R.; Wang, L.; Xu, L.; Tan, W.; et al. Interleukin-29 Accelerates Vascular Calcification via JAK2/STAT3/BMP2 Signaling. J. Am. Heart Assoc. 2023, 12, e027222. [CrossRef] [PubMed]
- 52. Hamirani, Y.S.; Pandey, S.; Rivera, J.J.; Ndumele, C.; Budoff, M.J.; Blumenthal, R.S.; Nasir, K. Markers of Inflammation and Coronary Artery Calcification: A Systematic Review. *Atherosclerosis* **2008**, *201*, 1–7. [CrossRef] [PubMed]
- 53. Zhu, Y.; Qu, J.; He, L.; Zhang, F.; Zhou, Z.; Yang, S.; Zhou, Y. Calcium in Vascular Smooth Muscle Cell Elasticity and Adhesion: Novel Insights Into the Mechanism of Action. *Front. Physiol.* **2019**, *10*, 852. [CrossRef] [PubMed]
- Muñoz, J.C.; Martín, R.; Alonso, C.; Gutiérrez, B.; Nieto, M.L. Relation between Serum Levels of Chemotaxis-Related Factors and the Presence of Coronary Artery Calcification as Expression of Subclinical Atherosclerosis. *Clin. Biochem.* 2017, *50*, 1048–1055. [CrossRef] [PubMed]
- Quercioli, A.; Luciano Viviani, G.; Dallegri, F.; Mach, F.; Montecucco, F. Receptor Activator of Nuclear Factor Kappa B Ligand/Osteoprotegerin Pathway Is a Promising Target to Reduce Atherosclerotic Plaque Calcification. *Crit. Pathw. Cardiol.* 2010, 9, 227–230. [CrossRef]
- 56. Rusanescu, G.; Weissleder, R.; Aikawa, E. Notch Signaling in Cardiovascular Disease and Calcification. *Curr. Cardiol. Rev.* 2008, *4*, 148–156. [CrossRef]
- Guerrero-Hue, M.; García-Caballero, C.; Palomino-Antolín, A.; Rubio-Navarro, A.; Vázquez-Carballo, C.; Herencia, C.; Martín-Sanchez, D.; Farré-Alins, V.; Egea, J.; Cannata, P.; et al. Curcumin Reduces Renal Damage Associated with Rhabdomyolysis by Decreasing Ferroptosis-Mediated Cell Death. *FASEB J.* 2019, 33, 8961–8975. [CrossRef]
- 58. Yang, H.; Xu, G.; Li, Q.; Zhu, L. Ligustrazine Alleviates the Progression of Coronary Artery Calcification by Inhibiting Caspase-3/GSDME Mediated Pyroptosis. *Biosci. Trends* 2024. [CrossRef]
- Kim, J.; Lee, D.Y.; Park, S.E.; Park, C.-Y.; Lee, W.-Y.; Oh, K.-W.; Park, S.-W.; Rhee, E.-J. Increased Risk for Development of Coronary Artery Calcification in Subjects with Non-Alcoholic Fatty Liver Disease and Systemic Inflammation. *PLoS ONE* 2017, *12*, e0180118. [CrossRef]
- 60. Wahlin, B.; Meedt, T.; Jonsson, F.; Henein, M.Y.; Wållberg-Jonsson, S. Coronary Artery Calcification Is Related to Inflammation in Rheumatoid Arthritis: A Long-Term Follow-Up Study. *Biomed. Res. Int.* **2016**, 2016, 1261582. [CrossRef]
- 61. Wang, C.; Liu, S.; Kamronbek, R.; Ni, S.; Yang, K.; Yang, Y.; Zhou, D.; Zhou, C.; Yin, C.; Zhang, M. Association between IL-2 Receptor and Severe Coronary Artery Calcification in Patients with Coronary Artery Disease. *Rev. Cardiovasc. Med.* **2024**, 25, 186. [CrossRef] [PubMed]
- Kamińska, J.; Stopiński, M.; Mucha, K.; Jędrzejczak, A.; Gołębiowski, M.; Niewczas, M.A.; Pączek, L.; Foroncewicz, B. IL 6 but Not TNF Is Linked to Coronary Artery Calcification in Patients with Chronic Kidney Disease. *Cytokine* 2019, 120, 9–14. [CrossRef] [PubMed]
- Ference, B.A.; Ginsberg, H.N.; Graham, I.; Ray, K.K.; Packard, C.J.; Bruckert, E.; Hegele, R.A.; Krauss, R.M.; Raal, F.J.; Schunkert, H.; et al. Low-Density Lipoproteins Cause Atherosclerotic Cardiovascular Disease. 1. Evidence from Genetic, Epidemiologic, and Clinical Studies. A Consensus Statement from the European Atherosclerosis Society Consensus Panel. *Eur. Heart J.* 2017, 38, 2459–2472. [CrossRef] [PubMed]
- 64. Villa-Bellosta, R. Vascular Calcification: A Passive Process That Requires Active Inhibition. *Biology* **2024**, *13*, 111. [CrossRef] [PubMed]
- 65. Sandesara, P.B.; Mehta, A.; O'Neal, W.T.; Mohamed Kelli, H.; Sathiyakumar, V.; Martin, S.S.; Blaha, M.J.; Blumenthal, R.S.; Sperling, L.S. Association of Elevated High-Density Lipoprotein Cholesterol and Particle Concentration with Coronary Artery Calcium: The Multi-Ethnic Study of Atherosclerosis. *Circ. Cardiovasc. Imaging* **2020**, *13*, e010473. [CrossRef]
- 66. Chung, Y.H.; Lee, B.-K.; Kwon, H.M.; Min, P.-K.; Choi, E.-Y.; Yoon, Y.W.; Hong, B.-K.; Rim, S.-J.; Kim, J.-Y. Coronary Calcification Is Associated with Elevated Serum Lipoprotein (a) Levels in Asymptomatic Men over the Age of 45 Years: A Cross-Sectional Study of the Korean National Health Checkup Data. *Medicine* 2021, 100, e24962. [CrossRef]

- 67. Kronenberg, F.; Mora, S.; Stroes, E.S.G.; Ference, B.A.; Arsenault, B.J.; Berglund, L.; Dweck, M.R.; Koschinsky, M.; Lambert, G.; Mach, F.; et al. Lipoprotein(a) in Atherosclerotic Cardiovascular Disease and Aortic Stenosis: A European Atherosclerosis Society Consensus Statement. *Eur. Heart J.* **2022**, *43*, 3925–3946. [CrossRef]
- Goldstein, J.L.; Brown, M.S. A Century of Cholesterol and Coronaries: From Plaques to Genes to Statins. *Cell* 2015, 161, 161–172. [CrossRef]
- 69. Sies, H.; Stahl, W.; Sevanian, A. Nutritional, Dietary and Postprandial Oxidative Stress. J. Nutr. 2005, 135, 969–972. [CrossRef]
- Malekmohammad, K.; Sewell, R.D.E.; Rafieian-Kopaei, M. Antioxidants and Atherosclerosis: Mechanistic Aspects. *Biomolecules* 2019, 9, 301. [CrossRef]
- Di Pietro, N.; Formoso, G.; Pandolfi, A. Physiology and Pathophysiology of oxLDL Uptake by Vascular Wall Cells in Atherosclerosis. *Vascul. Pharmacol.* 2016, 84, 1–7. [CrossRef]
- 72. Malekmohammad, K.; Bezsonov, E.E.; Rafieian-Kopaei, M. Role of Lipid Accumulation and Inflammation in Atherosclerosis: Focus on Molecular and Cellular Mechanisms. *Front. Cardiovasc. Med.* **2021**, *8*, 707529. [CrossRef]
- 73. Lind, L. Circulating Markers of Inflammation and Atherosclerosis. *Atherosclerosis* 2003, 169, 203–214. [CrossRef]
- 74. Allahverdian, S.; Chehroudi, A.C.; McManus, B.M.; Abraham, T.; Francis, G.A. Contribution of Intimal Smooth Muscle Cells to Cholesterol Accumulation and Macrophage-like Cells in Human Atherosclerosis. *Circulation* **2014**, *129*, 1551–1559. [CrossRef]
- Wang, Y.; Dubland, J.A.; Allahverdian, S.; Asonye, E.; Sahin, B.; Jaw, J.E.; Sin, D.D.; Seidman, M.A.; Leeper, N.J.; Francis, G.A. Smooth Muscle Cells Contribute the Majority of Foam Cells in ApoE (Apolipoprotein E)-Deficient Mouse Atherosclerosis. *Arterioscler. Thromb. Vasc. Biol.* 2019, 39, 876–887. [CrossRef]
- 76. Libby, P. Inflammation in Atherosclerosis. Nature 2002, 420, 868–874. [CrossRef]
- Patel, S.; Celermajer, D.S.; Bao, S. Atherosclerosis-Underlying Inflammatory Mechanisms and Clinical Implications. *Int. J. Biochem. Cell. Biol.* 2008, 40, 576–580. [CrossRef]
- 78. Ouweneel, A.B.; Van Eck, M. Lipoproteins as Modulators of Atherothrombosis: From Endothelial Function to Primary and Secondary Coagulation. *Vascul. Pharmacol.* **2016**, *82*, 1–10. [CrossRef]
- 79. Yang, P.; Troncone, L.; Augur, Z.M.; Kim, S.S.J.; McNeil, M.E.; Yu, P.B. The Role of Bone Morphogenetic Protein Signaling in Vascular Calcification. *Bone* 2020, *141*, 115542. [CrossRef]
- Luna-Luna, M.; Criales-Vera, S.; Medina-Leyte, D.; Díaz-Zamudio, M.; Flores-Zapata, A.; Cruz-Robles, D.; López-Meneses, M.; Olvera-Cruz, S.; Ramírez-Marroquín, S.; Flores-Castillo, C.; et al. Bone Morphogenetic Protein-2 and Osteopontin Gene Expression in Epicardial Adipose Tissue from Patients with Coronary Artery Disease Is Associated with the Presence of Calcified Atherosclerotic Plaques. *Diabetes Metab. Syndr. Obes.* 2020, *13*, 1943–1951. [CrossRef]
- Vossen, L.M.; Kroon, A.A.; Schurgers, L.J.; de Leeuw, P.W. Pharmacological and Nutritional Modulation of Vascular Calcification. *Nutrients* 2019, 12, 100. [CrossRef] [PubMed]
- Klarin, D.; Zhu, Q.M.; Emdin, C.A.; Chaffin, M.; Horner, S.; McMillan, B.J.; Leed, A.; Weale, M.E.; Spencer, C.C.A.; Aguet, F.; et al. Genetic Analysis in UK Biobank Links Insulin Resistance and Transendothelial Migration Pathways to Coronary Artery Disease. *Nat. Genet.* 2017, 49, 1392–1397. [CrossRef] [PubMed]
- Howson, J.M.M.; Zhao, W.; Barnes, D.R.; Ho, W.-K.; Young, R.; Paul, D.S.; Waite, L.L.; Freitag, D.F.; Fauman, E.B.; Salfati, E.L.; et al. Fifteen New Risk Loci for Coronary Artery Disease Highlight Arterial-Wall-Specific Mechanisms. *Nat. Genet.* 2017, 49, 1113–1119. [CrossRef] [PubMed]
- Matsunaga, H.; Ito, K.; Akiyama, M.; Takahashi, A.; Koyama, S.; Nomura, S.; Ieki, H.; Ozaki, K.; Onouchi, Y.; Sakaue, S.; et al. Transethnic Meta-Analysis of Genome-Wide Association Studies Identifies Three New Loci and Characterizes Population-Specific Differences for Coronary Artery Disease. *Circ. Genom. Precis. Med.* 2020, 13, e002670. [CrossRef] [PubMed]
- Webb, T.R.; Erdmann, J.; Stirrups, K.E.; Stitziel, N.O.; Masca, N.G.D.; Jansen, H.; Kanoni, S.; Nelson, C.P.; Ferrario, P.G.; König, I.R.; et al. Systematic Evaluation of Pleiotropy Identifies 6 Further Loci Associated with Coronary Artery Disease. J. Am. Coll. Cardiol. 2017, 69, 823–836. [CrossRef]
- O'Donnell, C.J.; Kavousi, M.; Smith, A.V.; Kardia, S.L.R.; Feitosa, M.F.; Hwang, S.-J.; Sun, Y.V.; Province, M.A.; Aspelund, T.; Dehghan, A.; et al. Genome-Wide Association Study for Coronary Artery Calcification with Follow-up in Myocardial Infarction. *Circulation* 2011, 124, 2855–2864. [CrossRef]
- de Vries, P.S.; Conomos, M.P.; Singh, K.; Nicholson, C.J.; Jain, D.; Hasbani, N.R.; Jiang, W.; Lee, S.; Lino Cardenas, C.L.; Lutz, S.M.; et al. Whole-Genome Sequencing Uncovers Two Loci for Coronary Artery Calcification and Identifies ARSE as a Regulator of Vascular Calcification. *Nat. Cardiovasc. Res.* 2023, *2*, 1159–1172. [CrossRef]
- Kavousi, M.; Bos, M.M.; Barnes, H.J.; Lino Cardenas, C.L.; Wong, D.; Lu, H.; Hodonsky, C.J.; Landsmeer, L.P.L.; Turner, A.W.; Kho, M.; et al. Multi-Ancestry Genome-Wide Study Identifies Effector Genes and Druggable Pathways for Coronary Artery Calcification. *Nat. Genet.* 2023, 55, 1651–1664. [CrossRef]
- Do, R.; Stitziel, N.O.; Won, H.-H.; Jørgensen, A.B.; Duga, S.; Angelica Merlini, P.; Kiezun, A.; Farrall, M.; Goel, A.; Zuk, O.; et al. Exome Sequencing Identifies Rare LDLR and APOA5 Alleles Conferring Risk for Myocardial Infarction. *Nature* 2015, *518*, 102–106. [CrossRef]
- Khera, A.V.; Won, H.-H.; Peloso, G.M.; O'Dushlaine, C.; Liu, D.; Stitziel, N.O.; Natarajan, P.; Nomura, A.; Emdin, C.A.; Gupta, N.; et al. Association of Rare and Common Variation in the Lipoprotein Lipase Gene with Coronary Artery Disease. *JAMA* 2017, 317, 937–946. [CrossRef]

- 91. Myocardial Infarction Genetics and CARDIoGRAM Exome Consortia Investigators; Stitziel, N.O.; Stirrups, K.E.; Masca, N.G.D.; Erdmann, J.; Ferrario, P.G.; König, I.R.; Weeke, P.E.; Webb, T.R.; Auer, P.L.; et al. Coding Variation in ANGPTL4, LPL, and SVEP1 and the Risk of Coronary Disease. *N. Engl. J. Med.* **2016**, *374*, 1134–1144. [CrossRef] [PubMed]
- 92. Khera, A.V.; Kathiresan, S. Genetics of Coronary Artery Disease: Discovery, Biology and Clinical Translation. *Nat. Rev. Genet.* 2017, *18*, 331–344. [CrossRef] [PubMed]
- Rutsch, F.; Ruf, N.; Vaingankar, S.; Toliat, M.R.; Suk, A.; Höhne, W.; Schauer, G.; Lehmann, M.; Roscioli, T.; Schnabel, D.; et al. Mutations in ENPP1 Are Associated with "idiopathic" Infantile Arterial Calcification. *Nat. Genet.* 2003, 34, 379–381. [CrossRef] [PubMed]
- 94. Bergen, A.A.; Plomp, A.S.; Schuurman, E.J.; Terry, S.; Breuning, M.; Dauwerse, H.; Swart, J.; Kool, M.; van Soest, S.; Baas, F.; et al. Mutations in ABCC6 Cause Pseudoxanthoma Elasticum. *Nat. Genet.* **2000**, *25*, 228–231. [CrossRef]
- 95. Rice, G.I.; Park, S.; Gavazzi, F.; Adang, L.A.; Ayuk, L.A.; Van Eyck, L.; Seabra, L.; Barrea, C.; Battini, R.; Belot, A.; et al. Genetic and Phenotypic Spectrum Associated with IFIH1 Gain-of-Function. *Hum. Mutat.* 2020, *41*, 837–849. [CrossRef]
- Jang, M.-A.; Kim, E.K.; Now, H.; Nguyen, N.T.H.; Kim, W.-J.; Yoo, J.-Y.; Lee, J.; Jeong, Y.-M.; Kim, C.-H.; Kim, O.-H.; et al. Mutations in DDX58, Which Encodes RIG-I, Cause Atypical Singleton-Merten Syndrome. Am. J. Hum. Genet. 2015, 96, 266–274. [CrossRef]
- Feigenbaum, A.; Müller, C.; Yale, C.; Kleinheinz, J.; Jezewski, P.; Kehl, H.G.; MacDougall, M.; Rutsch, F.; Hennekam, R.C.M. Singleton-Merten Syndrome: An Autosomal Dominant Disorder with Variable Expression. *Am. J. Med. Genet. A* 2013, 161A, 360–370. [CrossRef]
- Visseren, F.L.J.; Mach, F.; Smulders, Y.M.; Carballo, D.; Koskinas, K.C.; Bäck, M.; Benetos, A.; Biffi, A.; Boavida, J.-M.; Capodanno, D.; et al. 2021 ESC Guidelines on Cardiovascular Disease Prevention in Clinical Practice. *Eur. Heart J.* 2021, 42, 3227–3337. [CrossRef]
- 99. Kim, B.S.; Chan, N.; Hsu, G.; Makaryus, A.N.; Chopra, M.; Cohen, S.L.; Makaryus, J.N. Sex Differences in Coronary Arterial Calcification in Symptomatic Patients. *Am. J. Cardiol.* **2021**, *149*, 16–20. [CrossRef]
- 100. Obisesan, O.H.; Boakye, E.; Wang, F.M.; Dardari, Z.; Dzaye, O.; Cainzos-Achirica, M.; Meyer, M.L.; Gottesman, R.; Palta, P.; Coresh, J.; et al. Coronary Artery Calcium as a Marker of Healthy and Unhealthy Aging in Adults Aged 75 and Older: The Atherosclerosis Risk in Communities (ARIC) Study. *Atherosclerosis* 2024, 392, 117475. [CrossRef]
- 101. Bild, D.E.; Detrano, R.; Peterson, D.; Guerci, A.; Liu, K.; Shahar, E.; Ouyang, P.; Jackson, S.; Saad, M.F. Ethnic Differences in Coronary Calcification: The Multi-Ethnic Study of Atherosclerosis (MESA). *Circulation* 2005, 111, 1313–1320. [CrossRef] [PubMed]
- 102. Lehmann, N.; Möhlenkamp, S.; Mahabadi, A.A.; Schmermund, A.; Roggenbuck, U.; Seibel, R.; Grönemeyer, D.; Budde, T.; Dragano, N.; Stang, A.; et al. Effect of Smoking and Other Traditional Risk Factors on the Onset of Coronary Artery Calcification: Results of the Heinz Nixdorf Recall Study. *Atherosclerosis* 2014, 232, 339–345. [CrossRef] [PubMed]
- 103. Loria, C.M.; Liu, K.; Lewis, C.E.; Hulley, S.B.; Sidney, S.; Schreiner, P.J.; Williams, O.D.; Bild, D.E.; Detrano, R. Early Adult Risk Factor Levels and Subsequent Coronary Artery Calcification: The CARDIA Study. J. Am. Coll. Cardiol. 2007, 49, 2013–2020. [CrossRef] [PubMed]
- 104. Petsophonsakul, P.; Burgmaier, M.; Willems, B.; Heeneman, S.; Stadler, N.; Gremse, F.; Reith, S.; Burgmaier, K.; Kahles, F.; Marx, N.; et al. Nicotine Promotes Vascular Calcification via Intracellular Ca²⁺-Mediated, Nox5-Induced Oxidative Stress, and Extracellular Vesicle Release in Vascular Smooth Muscle Cells. *Cardiovasc. Res.* 2021, 118, 2196–2210. [CrossRef] [PubMed]
- 105. Lee, C.-D.; Jacobs, D.R.; Hankinson, A.; Iribarren, C.; Sidney, S. Cardiorespiratory Fitness and Coronary Artery Calcification in Young Adults: The CARDIA Study. *Atherosclerosis* **2009**, *203*, 263–268. [CrossRef]
- 106. Bertoni, A.G.; Whitt-Glover, M.C.; Chung, H.; Le, K.Y.; Barr, R.G.; Mahesh, M.; Jenny, N.S.; Burke, G.L.; Jacobs, D.R. The Association between Physical Activity and Subclinical Atherosclerosis: The Multi-Ethnic Study of Atherosclerosis. Am. J. Epidemiol. 2009, 169, 444–454. [CrossRef]
- 107. Aengevaeren, V.L.; Mosterd, A.; Braber, T.L.; Prakken, N.H.J.; Doevendans, P.A.; Grobbee, D.E.; Thompson, P.D.; Eijsvogels, T.M.H.; Velthuis, B.K. Relationship Between Lifelong Exercise Volume and Coronary Atherosclerosis in Athletes. *Circulation* 2017, 136, 138–148. [CrossRef]
- 108. Aengevaeren, V.L.; Mosterd, A.; Bakker, E.A.; Braber, T.L.; Nathoe, H.M.; Sharma, S.; Thompson, P.D.; Velthuis, B.K.; Eijsvogels, T.M.H. Exercise Volume Versus Intensity and the Progression of Coronary Atherosclerosis in Middle-Aged and Older Athletes: Findings from the MARC-2 Study. *Circulation* 2023, 147, 993–1003. [CrossRef]
- 109. Ogunmoroti, O.; Osibogun, O.; Mathews, L.; Esuruoso, O.A.; Ndumele, C.E.; Okunrintemi, V.; Burke, G.L.; Blumenthal, R.S.; Budoff, M.J.; Michos, E.D. Favorable Cardiovascular Health Is Associated With Lower Prevalence, Incidence, Extent, and Progression of Extracoronary Calcification: MESA. *Circ. Cardiovasc. Imaging* 2022, 15, e013762. [CrossRef]
- Hoffmann, U.; Massaro, J.M.; D'Agostino, R.B.; Kathiresan, S.; Fox, C.S.; O'Donnell, C.J. Cardiovascular Event Prediction and Risk Reclassification by Coronary, Aortic, and Valvular Calcification in the Framingham Heart Study. J. Am. Heart Assoc. 2016, 5, e003144. [CrossRef]
- 111. Gore, M.O.; Ayers, C.R.; Khera, A.; deFilippi, C.R.; Wang, T.J.; Seliger, S.L.; Nambi, V.; Selvin, E.; Berry, J.D.; Hundley, W.G.; et al. Combining Biomarkers and Imaging for Short-Term Assessment of Cardiovascular Disease Risk in Apparently Healthy Adults. *J. Am. Heart Assoc.* 2020, 9, e015410. [CrossRef] [PubMed]
- 112. Greenland, P.; LaBree, L.; Azen, S.P.; Doherty, T.M.; Detrano, R.C. Coronary Artery Calcium Score Combined with Framingham Score for Risk Prediction in Asymptomatic Individuals. *JAMA* 2004, 291, 210–215. [CrossRef] [PubMed]

- 113. Ilgar, M.; Dağ, N.; Türkoğlu, C. Importance of Incidental Coronary Artery Calcification in Early Diagnosis of Obstructive Coronary Artery Disease. *Pol. J. Radiol.* 2023, *88*, e338–e342. [CrossRef] [PubMed]
- 114. Arad, Y.; Goodman, K.J.; Roth, M.; Newstein, D.; Guerci, A.D. Coronary Calcification, Coronary Disease Risk Factors, C-Reactive Protein, and Atherosclerotic Cardiovascular Disease Events: The St. Francis Heart Study. J. Am. Coll. Cardiol. 2005, 46, 158–165. [CrossRef] [PubMed]
- 115. Nasir, K.; Michos, E.D.; Blumenthal, R.S.; Raggi, P. Detection of High-Risk Young Adults and Women by Coronary Calcium and National Cholesterol Education Program Panel III Guidelines. J. Am. Coll. Cardiol. 2005, 46, 1931–1936. [CrossRef]
- 116. Michos, E.D.; Nasir, K.; Braunstein, J.B.; Rumberger, J.A.; Budoff, M.J.; Post, W.S.; Blumenthal, R.S. Framingham Risk Equation Underestimates Subclinical Atherosclerosis Risk in Asymptomatic Women. *Atherosclerosis* **2006**, *184*, 201–206. [CrossRef]
- 117. Feuchtner, G.M.; Plank, F.; Beyer, C.; Barbieri, F.; Widmann, G.; Spitaler, P.; Dichtl, W. Cardiac Computed Tomography: State of the Art and Future Horizons. *J. Clin. Med.* **2022**, *11*, 4429. [CrossRef]
- 118. Authors/Task Force members; Windecker, S.; Kolh, P.; Alfonso, F.; Collet, J.-P.; Cremer, J.; Falk, V.; Filippatos, G.; Hamm, C.; Head, S.J.; et al. 2014 ESC/EACTS Guidelines on Myocardial Revascularization: The Task Force on Myocardial Revascularization of the European Society of Cardiology (ESC) and the European Association for Cardio-Thoracic Surgery (EACTS)Developed with the Special Contribution of the European Association of Percutaneous Cardiovascular Interventions (EAPCI). *Eur. Heart J.* 2014, 35, 2541–2619. [CrossRef]
- 119. Gulati, M.; Levy, P.D.; Mukherjee, D.; Amsterdam, E.; Bhatt, D.L.; Birtcher, K.K.; Blankstein, R.; Boyd, J.; Bullock-Palmer, R.P.; Conejo, T.; et al. 2021 AHA/ACC/ASE/CHEST/SAEM/SCCT/SCMR Guideline for the Evaluation and Diagnosis of Chest Pain: A Report of the American College of Cardiology/American Heart Association Joint Committee on Clinical Practice Guidelines. *Circulation* 2021, 144, e368–e454. [CrossRef]
- 120. Serruys, P.W.; Hara, H.; Garg, S.; Kawashima, H.; Nørgaard, B.L.; Dweck, M.R.; Bax, J.J.; Knuuti, J.; Nieman, K.; Leipsic, J.A.; et al. Coronary Computed Tomographic Angiography for Complete Assessment of Coronary Artery Disease: JACC State-of-the-Art Review. J. Am. Coll. Cardiol. 2021, 78, 713–736. [CrossRef]
- 121. Agatston, A.S.; Janowitz, W.R.; Hildner, F.J.; Zusmer, N.R.; Viamonte, M.; Detrano, R. Quantification of Coronary Artery Calcium Using Ultrafast Computed Tomography. J. Am. Coll. Cardiol. 1990, 15, 827–832. [CrossRef] [PubMed]
- 122. Hisamatsu, T.; Kinuta, M. Coronary Artery Calcium in Assessment of Atherosclerotic Cardiovascular Disease Risk and Its Role in Primary Prevention. *J. Atheroscler. Thromb.* **2023**, *30*, 1289–1302. [CrossRef] [PubMed]
- 123. Razavi, A.C.; Agatston, A.S.; Shaw, L.J.; De Cecco, C.N.; van Assen, M.; Sperling, L.S.; Bittencourt, M.S.; Daubert, M.A.; Nasir, K.; Blumenthal, R.S.; et al. Evolving Role of Calcium Density in Coronary Artery Calcium Scoring and Atherosclerotic Cardiovascular Disease Risk. JACC Cardiovasc. Imaging 2022, 15, 1648–1662. [CrossRef] [PubMed]
- 124. McCollough, C.H.; Ulzheimer, S.; Halliburton, S.S.; Shanneik, K.; White, R.D.; Kalender, W.A. Coronary Artery Calcium: A Multi-Institutional, Multimanufacturer International Standard for Quantification at Cardiac CT. *Radiology* 2007, 243, 527–538. [CrossRef] [PubMed]
- Callister, T.Q.; Cooil, B.; Raya, S.P.; Lippolis, N.J.; Russo, D.J.; Raggi, P. Coronary Artery Disease: Improved Reproducibility of Calcium Scoring with an Electron-Beam CT Volumetric Method. *Radiology* 1998, 208, 807–814. [CrossRef]
- 126. Yoon, H.C.; Greaser, L.E.; Mather, R.; Sinha, S.; McNitt-Gray, M.F.; Goldin, J.G. Coronary Artery Calcium: Alternate Methods for Accurate and Reproducible Quantitation. *Acad. Radiol.* **1997**, *4*, 666–673. [CrossRef]
- 127. Hecht, H.S. Coronary Artery Calcium Scanning: Past, Present, and Future. JACC Cardiovasc. Imaging 2015, 8, 579–596. [CrossRef]
- 128. Hecht, H.S.; Blaha, M.J.; Kazerooni, E.A.; Cury, R.C.; Budoff, M.; Leipsic, J.; Shaw, L. CAC-DRS: Coronary Artery Calcium Data and Reporting System. An Expert Consensus Document of the Society of Cardiovascular Computed Tomography (SCCT). *J. Cardiovasc. Comput. Tomogr.* **2018**, *12*, 185–191. [CrossRef]
- 129. Cury, R.C.; Abbara, S.; Achenbach, S.; Agatston, A.; Berman, D.S.; Budoff, M.J.; Dill, K.E.; Jacobs, J.E.; Maroules, C.D.; Rubin, G.D.; et al. Coronary Artery Disease—Reporting and Data System (CAD-RADS): An Expert Consensus Document of SCCT, ACR and NASCI: Endorsed by the ACC. *JACC Cardiovasc. Imaging* 2016, *9*, 1099–1113. [CrossRef]
- Williams, M.C.; Moss, A.; Dweck, M.; Hunter, A.; Pawade, T.; Adamson, P.D.; Shah, A.S.V.; Alam, S.; Maroules, C.D.; van Beek, E.J.; et al. Standardized Reporting Systems for Computed Tomography Coronary Angiography and Calcium Scoring: A Real-World Validation of CAD-RADS and CAC-DRS in Patients with Stable Chest Pain. J. Cardiovasc. Comput. Tomogr. 2020, 14, 3–11. [CrossRef]
- 131. Cury, R.C.; Abbara, S.; Achenbach, S.; Agatston, A.; Berman, D.S.; Budoff, M.J.; Dill, K.E.; Jacobs, J.E.; Maroules, C.D.; Rubin, G.D.; et al. CAD-RADSTM: Coronary Artery Disease—Reporting and Data System: An Expert Consensus Document of the Society of Cardiovascular Computed Tomography (SCCT), the American College of Radiology (ACR) and the North American Society for Cardiovascular Imaging (NASCI). Endorsed by the American College of Cardiology. *J. Am. Coll. Radiol.* 2016, 13, 1458–1466.e9. [CrossRef] [PubMed]
- 132. Mannarino, T.; D'Antonio, A.; Assante, R.; Zampella, E.; Gaudieri, V.; Petretta, M.; Cuocolo, A.; Acampa, W. Combined Evaluation of CAC Score and Myocardial Perfusion Imaging in Patients at Risk of Cardiovascular Disease: Where Are We and What Do the Data Say. J. Nucl. Cardiol. 2023, 30, 2349–2360. [CrossRef] [PubMed]
- 133. Guaricci, A.I.; Neglia, D.; Acampa, W.; Andreini, D.; Baggiano, A.; Bianco, F.; Carrabba, N.; Conte, E.; Gaudieri, V.; Mushtaq, S.; et al. Computed Tomography and Nuclear Medicine for the Assessment of Coronary Inflammation: Clinical Applications and Perspectives. *J. Cardiovasc. Med.* **2023**, *24*, e67–e76. [CrossRef] [PubMed]

- 134. Kwiecinski, J.; Wolny, R.; Chwala, A.; Slomka, P. Advances in the Assessment of Coronary Artery Disease Activity with PET/CT and CTA. *Tomography* **2023**, *9*, 328–341. [CrossRef] [PubMed]
- 135. Alavi, A.; Werner, T.J.; Raynor, W.; Høilund-Carlsen, P.F.; Revheim, M.-E. Critical Review of PET Imaging for Detection and Characterization of the Atherosclerotic Plaques with Emphasis on Limitations of FDG-PET Compared to NaF-PET in This Setting. *Am. J. Nucl. Med. Mol. Imaging.* 2021, 11, 337–351.
- 136. Kim, W.Y.; Danias, P.G.; Stuber, M.; Flamm, S.D.; Plein, S.; Nagel, E.; Langerak, S.E.; Weber, O.M.; Pedersen, E.M.; Schmidt, M.; et al. Coronary Magnetic Resonance Angiography for the Detection of Coronary Stenoses. N. Engl. J. Med. 2001, 345, 1863–1869. [CrossRef]
- 137. Foley, J.R.J.; Plein, S.; Greenwood, J.P. Assessment of Stable Coronary Artery Disease by Cardiovascular Magnetic Resonance Imaging: Current and Emerging Techniques. *World J. Cardiol.* **2017**, *9*, 92–108. [CrossRef]
- Hamdan, A.; Asbach, P.; Wellnhofer, E.; Klein, C.; Gebker, R.; Kelle, S.; Kilian, H.; Huppertz, A.; Fleck, E. A Prospective Study for Comparison of MR and CT Imaging for Detection of Coronary Artery Stenosis. *JACC Cardiovasc. Imaging* 2011, 4, 50–61. [CrossRef]
- Mitsis, A.; Eftychiou, C.; Kadoglou, N.P.E.; Theodoropoulos, K.C.; Karagiannidis, E.; Nasoufidou, A.; Ziakas, A.; Tzikas, S.; Kassimis, G. Innovations in Intracoronary Imaging: Present Clinical Practices and Future Outlooks. J. Clin. Med. 2024, 13, 4086.
 [CrossRef]
- 140. Peng, C.; Wu, H.; Kim, S.; Dai, X.; Jiang, X. Recent Advances in Transducers for Intravascular Ultrasound (IVUS) Imaging. *Sensors* **2021**, *21*, 3540. [CrossRef]
- 141. Tearney, G.J.; Regar, E.; Akasaka, T.; Adriaenssens, T.; Barlis, P.; Bezerra, H.G.; Bouma, B.; Bruining, N.; Cho, J.; Chowdhary, S.; et al. Consensus Standards for Acquisition, Measurement, and Reporting of Intravascular Optical Coherence Tomography Studies: A Report from the International Working Group for Intravascular Optical Coherence Tomography Standardization and Validation. J. Am. Coll. Cardiol. 2012, 59, 1058–1072. [CrossRef] [PubMed]
- Jenny, N.S.; Brown, E.R.; Detrano, R.; Folsom, A.R.; Saad, M.F.; Shea, S.; Szklo, M.; Herrington, D.M.; Jacobs, D.R. Associations of Inflammatory Markers with Coronary Artery Calcification: Results from the Multi-Ethnic Study of Atherosclerosis. *Atherosclerosis* 2010, 209, 226–229. [CrossRef] [PubMed]
- 143. Lee, H.; Park, H.E.; Yoon, J.W.; Choi, S.-Y. Clinical Significance of Body Fat Distribution in Coronary Artery Calcification Progression in Korean Population. *Diabetes Metab. J.* 2021, 45, 219–230. [CrossRef] [PubMed]
- 144. Oh, J.; Park, S.; Yu, H.T.; Chang, H.J.; Lee, S.H.; Kang, S.M.; Choi, D. Lack of Superiority for Soluble ST2 over High Sensitive C-Reactive Protein in Predicting High Risk Coronary Artery Calcium Score in a Community Cohort. *Yonsei Med. J.* 2016, 57, 1347–1353. [CrossRef] [PubMed]
- 145. Tajani, A.; Sadeghi, M.; Omidkhoda, N.; Mohammadpour, A.H.; Samadi, S.; Jomehzadeh, V. The Association between C-Reactive Protein and Coronary Artery Calcification: A Systematic Review and Meta-Analysis. BMC Cardiovasc. Disord 2024, 24, 204. [CrossRef]
- 146. Roy, N.; Rosas, S.E. IL-6 Associated with Progression of Coronary Artery Calcification and Mortality in Incident Dialysis Patients. *Am. J. Nephrol.* **2021**, *52*, 745–752. [CrossRef]
- Kurozumi, A.; Nakano, K.; Yamagata, K.; Okada, Y.; Nakayamada, S.; Tanaka, Y. IL-6 and sIL-6R Induces STAT3-Dependent Differentiation of Human VSMCs into Osteoblast-like Cells through JMJD2B-Mediated Histone Demethylation of RUNX2. *Bone* 2019, 124, 53–61. [CrossRef]
- 148. Wong, N.D.; Kawakubo, M.; LaBree, L.; Azen, S.P.; Xiang, M.; Detrano, R. Relation of Coronary Calcium Progression and Control of Lipids According to National Cholesterol Education Program Guidelines. *Am. J. Cardiol.* **2004**, *94*, 431–436. [CrossRef]
- 149. Wang, J.-S.; Chiang, H.-Y.; Wang, Y.-C.; Yeh, H.-C.; Ting, I.-W.; Liang, C.-C.; Wang, M.-C.; Lin, C.-C.; Hsiao, C.-T.; Shen, M.-Y.; et al. Dyslipidemia and Coronary Artery Calcium: From Association to Development of a Risk-Prediction Nomogram. *Nutr. Metab. Cardiovasc. Dis.* 2022, 32, 1944–1954. [CrossRef]
- 150. Fernández-Macías, J.C.; Ochoa-Martínez, A.C.; Varela-Silva, J.A.; Pérez-Maldonado, I.N. Atherogenic Index of Plasma: Novel Predictive Biomarker for Cardiovascular Illnesses. *Arch. Med. Res.* **2019**, *50*, 285–294. [CrossRef]
- Yao, H.; Feng, G.; Liu, Y.; Chen, Y.; Shao, C.; Wang, Z. Coronary Artery Calcification Burden, Atherogenic Index of Plasma, and Risk of Adverse Cardiovascular Events in the General Population: Evidence from a Mediation Analysis. *Lipids Health Dis.* 2024, 23, 258. [CrossRef] [PubMed]
- 152. Guo, Q.; Zhou, S.; Feng, X.; Yang, J.; Qiao, J.; Zhao, Y.; Shi, D.; Zhou, Y. The Sensibility of the New Blood Lipid Indicator-Atherogenic Index of Plasma (AIP) in Menopausal Women with Coronary Artery Disease. *Lipids Health Dis.* 2020, 19, 27. [CrossRef] [PubMed]
- 153. Sitinjak, B.D.P.; Murdaya, N.; Rachman, T.A.; Zakiyah, N.; Barliana, M.I. The Potential of Single Nucleotide Polymorphisms (SNPs) as Biomarkers and Their Association with the Increased Risk of Coronary Heart Disease: A Systematic Review. Vasc. Health Risk Manag. 2023, 19, 289–301. [CrossRef] [PubMed]
- 154. Li, Y.; Li, J.; Tang, X.; Xu, J.; Liu, R.; Jiang, L.; Tian, J.; Zhang, Y.; Wang, D.; Sun, K.; et al. Association of NPC1L1 and HMGCR Gene Polymorphisms with Coronary Artery Calcification in Patients with Premature Triple-Vessel Coronary Disease. *BMC Med. Genomics* 2024, *17*, 22. [CrossRef]
- 155. Golüke, N.M.S.; Schoffelmeer, M.A.; De Jonghe, A.; Emmelot-Vonk, M.H.; De Jong, P.A.; Koek, H.L. Serum Biomarkers for Arterial Calcification in Humans: A Systematic Review. *Bone Rep.* 2022, 17, 101599. [CrossRef]

- 156. Maahs, D.M.; Ogden, L.G.; Kinney, G.L.; Wadwa, P.; Snell-Bergeon, J.K.; Dabelea, D.; Hokanson, J.E.; Ehrlich, J.; Eckel, R.H.; Rewers, M. Low Plasma Adiponectin Levels Predict Progression of Coronary Artery Calcification. *Circulation* 2005, 111, 747–753. [CrossRef]
- Abedin, M.; Omland, T.; Ueland, T.; Khera, A.; Aukrust, P.; Murphy, S.A.; Jain, T.; Gruntmanis, U.; McGuire, D.K.; de Lemos, J.A. Relation of Osteoprotegerin to Coronary Calcium and Aortic Plaque (from the Dallas Heart Study). *Am. J. Cardiol.* 2007, 99, 513–518. [CrossRef]
- 158. Jono, S.; Ikari, Y.; Vermeer, C.; Dissel, P.; Hasegawa, K.; Shioi, A.; Taniwaki, H.; Kizu, A.; Nishizawa, Y.; Saito, S. Matrix Gla Protein Is Associated with Coronary Artery Calcification as Assessed by Electron-Beam Computed Tomography. *Thromb. Haemost.* 2004, 91, 790–794. [CrossRef]
- Schafer, C.; Heiss, A.; Schwarz, A.; Westenfeld, R.; Ketteler, M.; Floege, J.; Muller-Esterl, W.; Schinke, T.; Jahnen-Dechent, W. The Serum Protein Alpha 2-Heremans-Schmid Glycoprotein/Fetuin-A Is a Systemically Acting Inhibitor of Ectopic Calcification. *J. Clin. Investig.* 2003, 112, 357–366. [CrossRef]
- 160. Zheng, J.; Huang, M.; Huang, Q.; Chen, Q.; Chen, Z. The Relationship between Fetuin-A and Coronary Atherosclerotic Heart Disease (CHD) and CHD-Related Risk Factors. *Medicine* **2021**, *100*, e27481. [CrossRef]
- Pan, W.; Jie, W.; Huang, H. Vascular Calcification: Molecular Mechanisms and Therapeutic Interventions. *MedComm* 2023, 4, e200. [CrossRef] [PubMed]
- 162. Chen, N.X.; Kircelli, F.; O'Neill, K.D.; Chen, X.; Moe, S.M. Verapamil Inhibits Calcification and Matrix Vesicle Activity of Bovine Vascular Smooth Muscle Cells. *Kidney Int.* **2010**, *77*, 436–442. [CrossRef] [PubMed]
- 163. Shimizu, T.; Tanaka, T.; Iso, T.; Kawai-Kowase, K.; Kurabayashi, M. Azelnidipine Inhibits Msx2-Dependent Osteogenic Differentiation and Matrix Mineralization of Vascular Smooth Muscle Cells. *Int. Heart J.* **2012**, *53*, 331–335. [CrossRef] [PubMed]
- 164. Armstrong, Z.B.; Boughner, D.R.; Drangova, M.; Rogers, K.A. Angiotensin II Type 1 Receptor Blocker Inhibits Arterial Calcification in a Pre-Clinical Model. *Cardiovasc. Res.* 2011, *90*, 165–170. [CrossRef] [PubMed]
- 165. Montezano, A.C.; Nguyen Dinh Cat, A.; Rios, F.J.; Touyz, R.M. Angiotensin II and Vascular Injury. *Curr. Hypertens. Rep.* 2014, 16, 431. [CrossRef]
- 166. Lichtlen, P.R.; Hugenholtz, P.G.; Rafflenbeul, W.; Hecker, H.; Jost, S.; Deckers, J.W. Retardation of Angiographic Progression of Coronary Artery Disease by Nifedipine. Results of the International Nifedipine Trial on Antiatherosclerotic Therapy (INTACT). INTACT Group Investigators. *Lancet* 1990, 335, 1109–1113. [CrossRef]
- 167. Motro, M.; Shemesh, J. Calcium Channel Blocker Nifedipine Slows down Progression of Coronary Calcification in Hypertensive Patients Compared with Diuretics. *Hypertension* **2001**, *37*, 1410–1413. [CrossRef]
- 168. Nissen, S.E.; Tuzcu, E.M.; Libby, P.; Thompson, P.D.; Ghali, M.; Garza, D.; Berman, L.; Shi, H.; Buebendorf, E.; Topol, E.J.; et al. Effect of Antihypertensive Agents on Cardiovascular Events in Patients with Coronary Disease and Normal Blood Pressure: The CAMELOT Study: A Randomized Controlled Trial. *JAMA* 2004, 292, 2217–2225. [CrossRef]
- 169. Bruining, N.; de Winter, S.; Roelandt, J.R.T.C.; Rodriguez-Granillo, G.A.; Heller, I.; van Domburg, R.T.; Hamers, R.; de Feijter, P.J.; EUROPA/PERSPECTIVE Investigators. Coronary Calcium Significantly Affects Quantitative Analysis of Coronary Ultrasound: Importance for Atherosclerosis Progression/Regression Studies. *Coron. Artery Dis.* 2009, 20, 409–414. [CrossRef]
- 170. Kizu, A.; Shioi, A.; Jono, S.; Koyama, H.; Okuno, Y.; Nishizawa, Y. Statins Inhibit in Vitro Calcification of Human Vascular Smooth Muscle Cells Induced by Inflammatory Mediators. J. Cell Biochem. 2004, 93, 1011–1019. [CrossRef]
- 171. Bryniarski, K.L.; den Dekker, W.; Legutko, J.; Gasior, P.; Tahon, J.; Diletti, R.; Wilschut, J.M.; Nuis, R.-J.; Daemen, J.; Kleczynski, P.; et al. Role of Lipid-Lowering and Anti-Inflammatory Therapies on Plaque Stabilization. J. Clin. Med. 2024, 13, 3096. [CrossRef] [PubMed]
- 172. Trion, A.; Schutte-Bart, C.; Bax, W.H.; Jukema, J.W.; van der Laarse, A. Modulation of Calcification of Vascular Smooth Muscle Cells in Culture by Calcium Antagonists, Statins, and Their Combination. *Mol. Cell Biochem.* 2008, 308, 25–33. [CrossRef] [PubMed]
- 173. Budoff, M.J.; Lane, K.L.; Bakhsheshi, H.; Mao, S.; Grassmann, B.O.; Friedman, B.C.; Brundage, B.H. Rates of Progression of Coronary Calcium by Electron Beam Tomography. *Am. J. Cardiol.* **2000**, *86*, 8–11. [CrossRef] [PubMed]
- 174. Achenbach, S.; Ropers, D.; Pohle, K.; Leber, A.; Thilo, C.; Knez, A.; Menendez, T.; Maeffert, R.; Kusus, M.; Regenfus, M.; et al. Influence of Lipid-Lowering Therapy on the Progression of Coronary Artery Calcification: A Prospective Evaluation. *Circulation* 2002, 106, 1077–1082. [CrossRef] [PubMed]
- 175. Lee, S.-E.; Sung, J.M.; Andreini, D.; Budoff, M.J.; Cademartiri, F.; Chinnaiyan, K.; Choi, J.H.; Chun, E.J.; Conte, E.; Gottlieb, I.; et al. Differential Association between the Progression of Coronary Artery Calcium Score and Coronary Plaque Volume Progression According to Statins: The Progression of AtheRosclerotic PlAque DetermIned by Computed TomoGraphic Angiography Imaging (PARADIGM) Study. *Eur. Heart J. Cardiovasc. Imaging* **2019**, *20*, 1307–1314. [CrossRef]
- 176. Mitsis, A.; Kyriakou, M.; Sokratous, S.; Karmioti, G.; Drakomathioulakis, M.; Myrianthefs, M.; Ziakas, A.; Tzikas, S.; Kassimis, G. Exploring the Landscape of Anti-Inflammatory Trials: A Comprehensive Review of Strategies for Targeting Inflammation in Acute Myocardial Infraction. *Biomedicines* 2024, 12, 701. [CrossRef]
- 177. Tardif, J.-C.; Kouz, S.; Waters, D.D.; Bertrand, O.F.; Diaz, R.; Maggioni, A.P.; Pinto, F.J.; Ibrahim, R.; Gamra, H.; Kiwan, G.S.; et al. Efficacy and Safety of Low-Dose Colchicine after Myocardial Infarction. *N. Engl. J. Med.* **2019**, *381*, 2497–2505. [CrossRef]

- 178. Nidorf, S.M.; Fiolet, A.T.L.; Eikelboom, J.W.; Schut, A.; Opstal, T.S.J.; Bax, W.A.; Budgeon, C.A.; Tijssen, J.G.P.; Mosterd, A.; Cornel, J.H.; et al. The Effect of Low-Dose Colchicine in Patients with Stable Coronary Artery Disease: The LoDoCo2 Trial Rationale, Design, and Baseline Characteristics. Am. Heart J. 2019, 218, 46–56. [CrossRef]
- 179. González, L.; Bulnes, J.F.; Orellana, M.P.; Muñoz Venturelli, P.; Martínez Rodriguez, G. The Role of Colchicine in Atherosclerosis: From Bench to Bedside. *Pharmaceutics* **2022**, *14*, 1395. [CrossRef]
- 180. Gao, F.; Li, Y.P.; Ma, X.T.; Wang, Z.J.; Shi, D.M.; Zhou, Y.J. Effect of Alirocumab on Coronary Calcification in Patients With Coronary Artery Disease. *Front. Cardiovasc. Med.* **2022**, *9*, 907662. [CrossRef]
- 181. Ikegami, Y.; Inoue, I.; Inoue, K.; Shinoda, Y.; Iida, S.; Goto, S.; Nakano, T.; Shimada, A.; Noda, M. The Annual Rate of Coronary Artery Calcification with Combination Therapy with a PCSK9 Inhibitor and a Statin Is Lower than That with Statin Monotherapy. NPJ Aging Mech. Dis. 2018, 4, 7. [CrossRef] [PubMed]
- 182. Kawakami, R.; Nakagami, H.; Noma, T.; Ohmori, K.; Kohno, M.; Morishita, R. RANKL System in Vascular and Valve Calcification with Aging. *Inflamm. Regener.* 2016, 36, 10. [CrossRef] [PubMed]
- Sharma, T.; Mandal, C.C. Omega-3 Fatty Acids in Pathological Calcification and Bone Health. J. Food Biochem. 2020, 44, e13333.
 [CrossRef] [PubMed]
- 184. Samelson, E.J.; Booth, S.L.; Fox, C.S.; Tucker, K.L.; Wang, T.J.; Hoffmann, U.; Cupples, L.A.; O'Donnell, C.J.; Kiel, D.P. Calcium Intake Is Not Associated with Increased Coronary Artery Calcification: The Framingham Study. Am. J. Clin. Nutr. 2012, 96, 1274–1280. [CrossRef] [PubMed]
- 185. Kim, J.H.; Yoon, J.W.; Kim, K.W.; Lee, E.J.; Lee, W.; Cho, S.-H.; Shin, C.S. Increased Dietary Calcium Intake Is Not Associated with Coronary Artery Calcification. *Int. J. Cardiol.* **2012**, *157*, 429–431. [CrossRef]
- 186. Koshihara, Y.; Hoshi, K.; Okawara, R.; Ishibashi, H.; Yamamoto, S. Vitamin K Stimulates Osteoblastogenesis and Inhibits Osteoclastogenesis in Human Bone Marrow Cell Culture. *J. Endocrinol.* **2003**, 176, 339–348. [CrossRef]
- Shioi, A.; Morioka, T.; Shoji, T.; Emoto, M. The Inhibitory Roles of Vitamin K in Progression of Vascular Calcification. *Nutrients* 2020, 12, 583. [CrossRef]
- 188. Saito, Y.; Nakamura, K.; Miura, D.; Yunoki, K.; Miyoshi, T.; Yoshida, M.; Kawakita, N.; Kimura, T.; Kondo, M.; Sarashina, T.; et al. Suppression of Wnt Signaling and Osteogenic Changes in Vascular Smooth Muscle Cells by Eicosapentaenoic Acid. *Nutrients* 2017, 9, 858. [CrossRef]
- 189. Hénaut, L.; Massy, Z.A. Magnesium as a Calcification Inhibitor. Adv. Chronic. Kidney Dis. 2018, 25, 281–290. [CrossRef]
- Schelski, N.; Luong, T.T.D.; Lang, F.; Pieske, B.; Voelkl, J.; Alesutan, I. SGK1-Dependent Stimulation of Vascular Smooth Muscle Cell Osteo-/Chondrogenic Transdifferentiation by Interleukin-18. *Pflugers Arch.* 2019, 471, 889–899. [CrossRef]
- He, L.; Xu, J.; Bai, Y.; Zhang, H.; Zhou, W.; Cheng, M.; Zhang, D.; Zhang, L.; Zhang, S. MicroRNA-103a Regulates the Calcification of Vascular Smooth Muscle Cells by Targeting Runt-Related Transcription Factor 2 in High Phosphorus Conditions. *Exp. Ther. Med.* 2021, 22, 1036. [CrossRef] [PubMed]
- 192. Li, W.; Feng, W.; Su, X.; Luo, D.; Li, Z.; Zhou, Y.; Zhu, Y.; Zhang, M.; Chen, J.; Liu, B.; et al. SIRT6 Protects Vascular Smooth Muscle Cells from Osteogenic Transdifferentiation via Runx2 in Chronic Kidney Disease. J. Clin. Investig. 2022, 132, e150051. [CrossRef] [PubMed]
- 193. Li, T.; Yu, H.; Zhang, D.; Feng, T.; Miao, M.; Li, J.; Liu, X. Matrix Vesicles as a Therapeutic Target for Vascular Calcification. *Front. Cell Dev. Biol.* **2022**, *10*, 825622. [CrossRef] [PubMed]
- 194. Schurgers, L.J.; Uitto, J.; Reutelingsperger, C.P. Vitamin K-Dependent Carboxylation of Matrix Gla-Protein: A Crucial Switch to Control Ectopic Mineralization. *Trends Mol. Med.* 2013, *19*, 217–226. [CrossRef] [PubMed]
- 195. Wang, R.C.; Wang, Z. Precision Medicine: Disease Subtyping and Tailored Treatment. Cancers 2023, 15, 3837. [CrossRef]
- 196. Shchekochikhin, D.; Kopylov, P. Personalized Medicine in Coronary Artery Disease: Where Are We in 2022? *J. Pers. Med.* 2022, 12, 1446. [CrossRef]
- 197. Sethi, Y.; Patel, N.; Kaka, N.; Kaiwan, O.; Kar, J.; Moinuddin, A.; Goel, A.; Chopra, H.; Cavalu, S. Precision Medicine and the Future of Cardiovascular Diseases: A Clinically Oriented Comprehensive Review. J. Clin. Med. **2023**, 12, 1799. [CrossRef]
- 198. Mitsis, A.; Myrianthefs, M.; Sokratous, S.; Karmioti, G.; Kyriakou, M.; Drakomathioulakis, M.; Tzikas, S.; Kadoglou, N.P.E.; Karagiannidis, E.; Nasoufidou, A.; et al. Emerging Therapeutic Targets for Acute Coronary Syndromes: Novel Advancements and Future Directions. *Biomedicines* 2024, 12, 1670. [CrossRef]
- 199. Bundy, K.; Boone, J.; Simpson, C.L. Wnt Signaling in Vascular Calcification. Front. Cardiovasc. Med. 2021, 8, 708470. [CrossRef]
- 200. Sirasanagandla, S.R.; Al-Huseini, I.; Al Mushaiqri, M.; Al-Abri, N.; Al-Ghafri, F. Maternal resveratrol supplementation ameliorates bisphenol A-induced atherosclerotic lesions formation in adult offspring ApoE-/- mice. *3 Biotech* **2022**, *12*, 36. [CrossRef]

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