



Review

Pathogenesis and Management Strategies in Radioiodine-Refractory Differentiated Thyroid Cancer: From Molecular Mechanisms Toward Therapeutic Approaches: A Comprehensive Review

Iulia-Alexandra Voinea ^{1,*}, Eugenia Petrova ^{2,3}, Nicoleta Dumitru ^{2,3}, Andra Cocoloș ^{2,3}, Dumitru Ioachim ⁴, Andrei Liviu Goldstein ⁵ and Adina Mariana Ghemigian ^{2,3}

¹ PhD Doctoral School, “Carol Davila” University of Medicine and Pharmacy, 0505474 Bucharest, Romania

² Department of Endocrinology, “Carol Davila” University of Medicine and Pharmacy, 020021 Bucharest, Romania; eugenia.petrova@umfcd.ro (E.P.); nicoleta.dumitru@umfcd.ro (N.D.); andra_buruiana@yahoo.com (A.C.); adina.ghemigian@umfcd.ro (A.M.G.)

³ Department of Clinical Endocrinology V, C.I. Parhon National Institute of Endocrinology, 011863 Bucharest, Romania

⁴ Department of Pathology, C.I. Parhon National Institute of Endocrinology, 011863 Bucharest, Romania

⁵ Department of Nuclear Medicine, C.I. Parhon National Institute of Endocrinology, 011863 Bucharest, Romania; andrei_goldstein@yahoo.com

* Correspondence: iulia-alexandra.voinea@drd.umfcd.ro

Abstract: Thyroid cancer (TC) remains the most common cancer in endocrinology. Differentiated thyroid cancer (DTC), the most common type of TC, generally has a favorable outlook with conventional treatment, which typically includes surgery along with radioiodine (RAI) therapy and thyroid-stimulating hormone (TSH) suppression through thyroid hormone therapy. However, a small subset of patients (less than 5%) develop resistance to RAI. This resistance occurs due to the loss of Na/I symporter (NIS) activity, which is crucial for iodine absorption in thyroid cells. The decline in NIS activity appears to be due to gene modifications, reconfigurations with irregular stimulation of signaling pathways such as MAPK and PI3K/Akt pathways. These molecular changes lead to a diminished ability of DTC cells to concentrate iodine, which makes RAI therapy ineffective. As a consequence, patients with radioiodine-refractory DTC require alternative treatments. Therapy with tyrosine kinase inhibitors (TKIs) has emerged as the primary treatment option to inhibit proliferation and growth of RAIR-DTC, targeting the pathways responsible for tumor progression. In this article, we analyze molecular processes responsible for RAI resistance and explore both conventional and emerging therapeutic strategies for managing RAIR-DTC, aiming to improve patient outcomes.

Keywords: differentiated thyroid cancer; RAIR-DTC; NIS; signal pathways; tyrosine kinase inhibitors; surgery; iodine therapy



Citation: Voinea, I.-A.; Petrova, E.; Dumitru, N.; Cocoloș, A.; Ioachim, D.; Goldstein, A.L.; Ghemigian, A.M. Pathogenesis and Management Strategies in Radioiodine-Refractory Differentiated Thyroid Cancer: From Molecular Mechanisms Toward Therapeutic Approaches: A Comprehensive Review. *J. Clin. Med.* **2024**, *13*, 7161. <https://doi.org/10.3390/jcm13237161>

Academic Editor: Fernando Cordido

Received: 1 October 2024

Revised: 12 November 2024

Accepted: 23 November 2024

Published: 26 November 2024



Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Thyroid cancer (TC) is the most common endocrine tumor, and in most cases it is a differentiated thyroid cancer (DTC) [1,2]. Less than 5% of the subjects with poorly DTC have resistance to radioactive iodine therapy; thus, poor prediction is found, with an average life expectancy of 3–5 years. A multidisciplinary approach is needed to establish a personalized strategy [3,4].

The most prevalent forms of TC are papillary and follicular, making up over 90% of cases and collectively classified as DTCs [5]. These DTCs are usually slow-growing and have a favorable prognosis, with survival lasting 20 years or more after standard treatment [6,7].

For most patients with DTC, standard treatment—typically involving surgery followed by radioiodine (RAI) ablation, risk-adjusted monitoring, and thyroid-stimulating hormone (TSH) suppression treatment—is generally effective [8]. Nevertheless, local recurrence and distant metastases can occur in approximately 20% and 10% of patients, respectively, within the first decade after surgery [6]. Traditional treatment options for such cases include repeated radioiodine treatment, surgical treatment of metastases, and external radiation. Despite these interventions, about two-thirds of DTC cases eventually become resistant to RAI therapy, which worsens prognosis and life expectancy [1]. Once resistance develops, the survival rate at 10 years drops to about 20% [9].

In recent years, progress in genome sequencing has greatly improved our comprehension of the molecular pathways behind TC [3]. Most of the TC in this subgroup show alterations in the MAPK and PI3K/mTOR/Akt signal pathways, which are essential for regulating cell growth and division by transmitting signals from the cell membrane to the nucleus [10,11].

The excessive activation of the MAPK pathway plays a key role in the onset of papillary TC (PTC), often resulting from base substitutions in the *BRAF* oncogene. *BRAF*, part of the RAF family of serine/threonine kinases that is downstream of *RAS*, is commonly altered in PTC, associated with mutation rates reported between 29% and 83% [12–15]. This mutation triggers transcription factors that drive processes like cell expansion, maturation, cell division, and programmed cell death. While various pathogenic *BRAF* variants were identified, the *BRAF*^{V600E} mutation is the most common in classic PTC cases [16]. Research has linked this variant to more aggressive disease characteristics, such as metastasis, invasion, and recurrence [17]. *BRAF*^{V600E} mutation also promotes modulating TGF- β production, which suppresses sodium iodide symporter (NIS) expression, resulting in resistance to RAI therapy [18].

Similarly, stimulation of PI3K/mTOR/Akt signaling cascade is crucial during follicular TC (FTC) development. This pathway is activated due to mutations in the *RAS*, *PIK3CA*, and *AKT1* oncogenes or due to the loss of function of the *PTEN* oncogene, which normally plays the part of a negative regulator. *RAS* pathogenic variants, which drive both MAPK and PI3K-Akt signaling cascades, are commonly observed in FTC cases (between 28 and 68%), as well as in as many as 43% of the follicular-variant PTCs and 47% of non-invasive follicular-variant PTCs [19,20]. However, *RAS* pathogenic variants alone appear to have a relatively limited effect on clinical outcomes in TC [21].

As TC advances and loses differentiation, transforming into poorly DTC or anaplastic thyroid cancer (ATC), additional pathogenic variants—such as those affecting the p53 and Wnt/ β -catenin pathways—become involved. Recent research has also discovered alterations in *TERT* promoter across every single TC variant, more frequently found in aggressive and poorly DTC, underscoring their role in driving disease development [22–24].

As our understanding of the genetics of malignant thyroid disease advances, treatment approaches have evolved from concentrating solely on tumor type and histological features to targeting specific genetic alterations. This shift has resulted in the creation of new targeted therapies aimed at patients with more aggressive forms of the disease [25,26].

Objective

This review outlines the molecular process behind TC that contributes to refractoriness to RAI in DTC, as well as the modern diagnostic and treatment management strategies. We highlight the specific genetic alterations associated with this resistance and examine both conventional and emerging therapeutic approaches, including targeted therapies and innovative strategies to address treatment challenges. Additionally, we discuss the importance of precision medicine in optimizing patient outcomes and enhancing the effectiveness of existing treatment options.

2. Materials and Methods

An exhaustive search was made using PubMed, Scopus, and Google Scholar to identify recent studies and guidelines focused on the molecular mechanisms and the management of diagnosis and treatment for radioiodine-refractory DTC (RAIR-DTC). The search is a narrative comprehensive review and targeted English-language publications from 1998 to 2024, with a focus on both experimental and clinical trials. The search strategy incorporated key terms such as “differentiated thyroid cancer”, “RAIR-DTC”, “NIS”, “signal pathways”, “tyrosine kinase inhibitors”, “surgery”, and “iodine therapy”.

Table 1 lists the inclusion and exclusion criteria that formed the basis of this review.

Table 1. Inclusion and exclusion criteria.

Inclusion Criteria
Original studies
Topic: gene data, radiiodine-refractory
Published in PubMed
Timeframe of search: 1998–2024
Exclusion criteria
Non-human data
Case report, case series
Editorial
Non-English paper
Pediatric data
Selective inhibitor of RET
Selective inhibitor of NTRK
RAI-avid
MTC

Abbreviations: RAI-avid—radioactive iodine-avid; MTC—medullary thyroid carcinoma.

3. Pathogenesis of RAIR-DTC

Resistance to RAI occurs as a result of losing thyroid differentiation. Dedifferentiation is a consequence of damage to the NIS. Part of SLC5A5, NIS is a basolateral membrane glycoprotein in follicular epithelial cells. Iodine, as a necessary component in the follicular synthesis of thyroid hormones, enters the cell actively through NIS. Normally, NIS transcription begins when TSH binds with the TSH receptor and the cAMP pathway is immediately initiated. Then, cAMP enhances some activating pathways that contribute to NIS upstream enhances (NUE) stimulation. Thus, this stimulation of NUE is performed either in a PKA-dependent or PKA-non-dependent manner. For the case of the independent PKA pathway, Paired box gene-8 (PAX8) is activated using Ref-1, thus linking to NUE. This mechanism has a key role in the process of follicular cell differentiation. Regarding the PKA-dependent route, aAMP-response element modulator (CREM) amplifies the NUE function [27–29].

The decrease in the NIS signal, which is responsible for resistance to RAI, appears as a result of modulation of signaling pathways, chromosomal rearrangements, or aberrant gene methylation [27,29,30].

3.1. Molecular Genetic Characterisation

3.1.1. BRAF Pathogenic Variant and Rearrangement

BRAF, which is a proto-oncogene belonging to a family of serine/threonine kinases, has fundamental importance of MAPKKK in the MAPK signaling cascade [30]. T1799A point genetic alteration located in exon 15 is one of the most common mutations in the

BRAF gene [31]. This missense mutation leads to a change in B-raf protein residue 600, replacing glutamic acid with valine (V600E) and the persistent serine/threonine kinase function that damages the suppression loop. As a result, *BRAF*^{V600E} could initiate itself and also the MAPK signaling cascade [30,32].

Undoubtedly, *BRAF*^{V600E} remains the most common genetic mutation in thyroid cancers, being described in more than half of the DTCs [27]. According to clinical studies, patients with PTC with *BRAF*^{V600E} would have good prognoses. The synergistic action of *BRAF*^{V600E} with another gene mutation increases the aggressiveness. Studies suggest that patients with PTC and *BRAF*^{V600E} pathogenic variant develop aggressive pathological features, high risk of recurrence, and lack of RAI capture. The co-association of *BRAF*^{V600E} and *CYP2S1* adversely affects PTC. The presentation of *CYP2S1* is controlled by the MAPK signaling pathway mediated by *BRAF*^{V600E} with the help of the AHR-dependent cascade. The AHR/*CYP2S1* feedback mechanism increases the impact of mutations on *BRAF*^{V600E}. Moreover, the *BRAF*^{V600E} proto-oncogene can be connected with Wilm tumor gene1 (*WT1*), which has a function in transcription of a gene that is important for cell viability, differentiation, as well as proliferation [30,33,34].

BRAF fusion is an additional critical factor that determines TC progression. According to a study conducted on 65 Ukrainian-American individuals with PTC subjected to the effects of Chernobyl radiation, several alterations in MACF-*BRAF*, MBP-*BRAF*, and POR-*BRAF* were discovered through next-generation sequencing (NGS) and RNA sequencing. These may be responsible for the evolution of TC with radiation exposure [35].

3.1.2. NTRK Gene Fusion

NTRK (such as *NTRK1*, *NTRK2*, and *NTRK3*), is responsible for encoding tropomyosin receptor kinase (TRK) fusion proteins [30,36,37]. *NTRK* fusion determines carcinogenic effect in numerous tumors in both mature individuals and juveniles. Patients with DTC and *NTRK* gene fusion have a higher chance of distant metastasis as well as RAI resistance than those with DTC and *BRAF* or *RAS* pathogenic variants. Sequencing of tumor DNA and RNA, and profiling of plasma cell-free DNA are used to detect these fusions [30,36,37].

3.1.3. TERT Promoter Mutation

TERT, a ribonucleoprotein polymerase, is capable of lengthening telomeres upon activation. *TERT* reactivation that is present in many cancers is caused by the alteration of the *TERT* promoter (*TERTp*). *TERTp* is linked to RAI resistance. Several publications have shown that patients with simultaneously associated *TERT* and *BRAF*^{V600E} mutations do not respond to RAI therapy in contrast to patients with only *BRAF*^{V600E} pathogenic variant [30,38–41].

3.1.4. RAS Mutation

MAPK and PI3K cascades are activated by *RAS*. Proto-oncogenes are represented by *NRAS*, *HRAS*, and *KRAS*. Among them, the most common *RAS* genetic alteration remains *NRAS* codon 61 genetic alteration, proceeding with *HRAS* codon 61, *KRAS* codon 12/13, and *KRAS* codon 61. The association between *RAS* genetic alteration and *BRAF* mutation or *RET/PTC* rearrangement provides a negative prognosis [30,42,43].

3.1.5. ALK Gene Mutation and Fusion

ALK gene is known to be a partner in a genetic fusion of t(2;5) chromosome translocation in anaplastic large cell lymphoma. The components of the *ALK* membrane-binding receptor are represented by extracellular receptor-binding domain, a transmembrane region and an intracellular kinase domain. The mutation or *ALK* gene fusion causes the spontaneous activation of *ALK* leading to the stimulation of MAPK, PI3K-AKT, CRK-like proto-oncogene, CRKL-C3G, MEKK2/3-MEK5-ERK5, and JAK-STAT cascades [30,44,45].

Fusion of *ALK*, which is rare in PTC, was found in thyroid carcinoma through RNA sequencing analysis. Furthermore, a correlation was observed between *ALK* fusion and

aggressive thyroid carcinoma. The most common fusion is represented by *ALK* and *STRN* gene. It was found that *STRN-ALK* dimerization leads to *ALK* kinase activation. Thus, targeted therapies on *ALK* fusion are being tried. A novel *ALK* gene fusion, *CCD149-ALK*, was reported using NGS in a woman with RAIR-DTC with disseminated metastasis [30,44,45].

3.1.6. RET Rearrangement

RET rearrangement is situated on the long arm of chromosome 10 (10q11.2) and is found in 20% of PTC. It is responsible for encoding TKR of GFL. While *RET* normally contributes to the formation of the kidney and enteric nervous systems during embryogenesis, various factors including ionizing radiation or replication-related stress in DNA fragile sites can lead to DSBs. These breaks can cause *RET* gene fusion, maintaining the kinase domain, which then activates *RET* protein aberrantly. This activation promotes cell proliferation, differentiation, and development through downstream signaling pathways. Importantly, *RET* fusion also affects the production of thyroid cell-specific genes. Consequently, *RET* fusion serves as a carcinogen in PTC, non-small cell lung cancer, and various other malignancies. *RET/PTC1* and *RET/PTC3* rearrangements are the most frequent *RET/PTC* rearrangements [30,46–52].

3.1.7. PAX8/PPAR γ

PAX8, which is a component of the transcription factors family, plays a role in promoting the activation of numerous thyroid-specific genes within mature thyroid cells through binding to their promoters. These genes include those that code for thyroglobulin, thyroid peroxidase, and NIS. On the other hand, Peroxisome Proliferator-Activated Receptor Gamma (*PPAR γ*), part of the nuclear receptor group of transcription factors, governs systemic fat metabolism and insulin responsiveness [30,53].

The combination of *PAX8* and *PPAR γ* , known as *PAX8/PPAR γ* rearrangement, arises from a relocation between chromosome regions 2q13 and 3p25. The combination results in the creation of a fusion transcript that codes for *PPFP*. In addition, *PPFP* is found in around 30–35% of FTCs and PTCs [30,53].

Functioning as a cancer-associated protein, *PPFP* can promote cell proliferation, inhibit cell death, as well as enhance DNA replication in the G0/G1 quiescent phase. Notably, the expression of *PPFP* in human thyroid cancer cell cultures modulates the regulation of thyroid-specific genes, including *SLC5A5*, *TPO*, *TG*, and *TSHR*, that are regulated through *PAX8* to different levels. Dysregulation of these *PAX8* target sequences as well as their associated pathways is believed to underlie the carcinogenic effects of *PPFP* [30,53].

3.1.8. SWI/SNF Complex Alteration

SWI/SNF chromatin remodeling complex alteration is a highly conserved molecular complex comprising 10–15 subunits. It associates with histones and transcription regulators and is categorized into BAF, PBAF, and ncBAF complexes. While these complexes contain shared subunits like *SMARCC1/2* and *SMARCD1/2/3*, they additionally possess specific subunits like *ARID1A* or *ARID1B* [30,54,55].

Gene mutations leading to *SWI/SNF* complex deletion result in decreased chromatin accessibility, thereby weakening the regulation of thyroid-specific transcription factors (TF) such as *Foxe1*, *Nkx2-1*, and *PAX8*, crucial for iodization. Deletion of specific subunits like *ARID1A*, *ARID2*, or *SMARCB1* has been linked to the progression of *BRAF^{V600E}*-driven mouse TC. Furthermore, absence of the *SWI/SNF* complex can counteract the treatment efficacy of MAPK blockers and re-differentiation treatments [30,54,55].

3.2. Regulation of Signaling Pathways

3.2.1. TSHR Pathway Activation

TSH regulates the regulation of NIS within thyroid follicular cells. TSH links to its receptor (TSHR), a glycoprotein receptor belonging to the G protein-coupled receptor (GPCR) class, located on the cell surface. Stimulation of TSHR through TSH or other

signaling factors triggers various G proteins and subsequent pathways, influencing thyroid cell proliferation and the synthesis and hormonal secretion from the thyroid [30,56–59].

The relationship between TSH and TC cells operates on two levels. From one perspective, it aids therapy by activating pathways like cAMP, promoting activation of thyroid-specific genes like NIS. On the other hand, TSH can stimulate cancer cell growth through pathways like PI3K and MAPK. Moreover, the TSH-TSHR signaling pathway can facilitate immune evasion by tumor cells by inducing expression of tumor PD-L1, suppressing T cell killing effects [30,56–59].

3.2.2. MAPK Pathway

The MAPK pathway, crucial for regulating thyroid-specific gene expression, is frequently implicated in TC development. The MAPK family comprises ERK, JNK/SAPK, and p38 MAPK, facilitating signal transmission from the extracellular environment to intracellular targets [30,59–63]. In TC, aberrant MAPK pathway activation governs cell division, expansion, and viability. Notably, MAPK activation promotes dedifferentiation of DTC, marked by reduced expression of thyroid hormone production genes including NIS, TPO, and TG, often via downregulation of histone acetylation in NIS gene promoters [30,59–63].

The predominant driver of MAPK pathway perturbation observed in RAIR-DTC is the BRAF^{V600E} pathogenic variant, complemented by a spectrum of BRAF genetic alterations, RAS genetic alterations, as well as mutations in the MEK gene [29,30,59–63] (Figure 1).

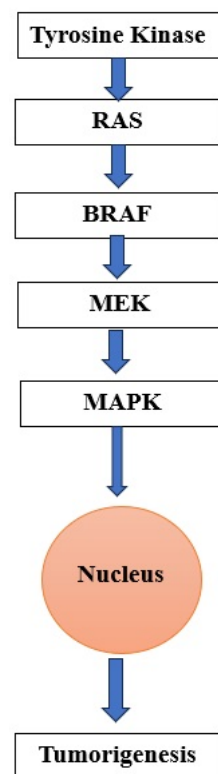


Figure 1. MAPK pathway in PTC [29,30,59–63]. Abbreviations: BRAF—V-Raf mouse sarcoma virus oncogene homologous B1; MEK—mitogen-activated protein kinase/extracellular signal-regulated kinase kinase; MAPK—mitogen-activated protein kinase.

3.2.3. PI3K Cascade

The PI3K signaling pathway contributes significantly to the development of RAIR-DTC, governing critical cellular processes including cellular growth, differentiation, and metastasis in TC. Comprising PI3K, AKT, and mTOR, activation of this pathway, along with the cAMP-independent pathway, counteracts the cAMP-dependent pathway’s promotion of thyroid-specific protein expression like NIS [30,64,65].

Besides the RAS genetic alteration and phospholipase C activation, IGF-2 has a major impact on PI3K initiation. RAI-DTC cells exhibit high expression levels of IGF-2 and its receptor IR-A. While there are no specific inhibitors for IR-A, dynamic interaction between insulin/IGF systems and discoid-domain-containing receptor (DDR) has been found. Inhibition or decrease in DDR1 expression notably reduces IR-A and IGF-2 production, causing an elevation in thyroid-related gene expression [30,66,67].

AKT activation drives mTOR signaling and simultaneously alters RUNX2 by phosphorylation. RUNX2 modulates various cellular processes including chondrocyte proliferation, maturation, as well as hypertrophy in endochondral ossification. Furthermore, it governs gene activity implicated in TC progression, infiltration, and metastasis. Excessive AKT activity leads to increased expression patterns and transcriptional processes of RUNX2, subsequently amplifying PI3K, AKT, and mTOR expression. This interplay between PI3K/AKT/mTOR cascade and RUNX2 significantly drives cancer growth. MAPK4, a distinct MAPK, can activate AKT through binding to it directly and promoting phosphorylation at threonine 308. Additionally, MAPK4 activates mTORC2, facilitating serine 473 phosphorylation of AKT. Hence, targeting MAPK4 may provide a novel treatment option for RAI-DTC [30,68–71].

The mTOR protein, situated under the influence of the PI3K/AKT signaling cascade, works as a serine-threonine protein kinase crucial for regulating various cellular activities such as metabolism, cell division, and longevity, alongside modulating gene expression of key thyroid factors including NIS, essential for RAI uptake. Research has shown that suppression of mTOR with an mTOR inhibitor enhances iodine absorption in TSH-stimulated PCCL3-derived cells from thyroid. However, the impact of rapamycin on iodine uptake appears to be less pronounced compared to inhibition of PI3K, suggesting that mTOR regulates both cell survival and the iodine absorption capacity of thyroid cells [30,72].

3.2.4. TGF- β Pathway

Aberrant TGF- β signaling is linked to multiple diseases, particularly cancer. In human thyroid malignancies, TGF- β is upregulated and serves as a strong promoter in tumor formation and metastasis. In PTC, the *BRAF*^{V600E} alteration stimulates active TGF- β 1, initiating TGF- β -induced autocrine loop. This mutation also increases levels of both total and phosphorylated Smad3. Initiation of the TGF- β /Smad signaling pathway enhances NOX4 gene expression, which, in turn, forms a heterodimeric complex with p22phox, a regulatory subunit of NOX. This complex regulates the TGF- β /Smad3 cascade by generating ROS. ROS generated by NOX4 act as second messengers, suppressing the progression of TC, especially the expression of NIS, while promoting their proliferation and metastasis. Thus, *BRAF*^{V600E}-induced RAI-DTC is significantly influenced by the TGF- β /Smad signaling pathway [30,73,74].

3.2.5. Wnt/ β -Catenin Pathway

Wnt glycoproteins release the transcription factor β -catenin from a protein complex by interacting with Frizzled and LDL receptor-related proteins. This prevents β -catenin's phosphorylation and degradation, allowing it to enter the nucleus and regulate gene expression by binding to T cell factor (TCF) [30,75,76]. The Wnt/ β -catenin pathway significantly influences TC growth and differentiation. In cancer stem cells (CSCs), β -catenin is upregulated, enhancing CSC self-renewal and proliferation, which drives TC progression. Increased lysine-specific histone demethylase 1A (LSD1) in CSCs upregulates β -catenin by downregulating adenomatous polyposis coli 2 (APC2) and Dickkopf-related protein 1 (DKK1), both of which normally promote β -catenin degradation. This pathway activation increases CSCs and contributes to TC's chemotherapy resistance [30,77,78].

The Wnt/ β -catenin signaling supports TC cell proliferation with *BRAF*^{V600E} mutations. Knocking out β -catenin slows tumor growth and reduces papillary structures. Additionally, treatment with PKF118-310, a β -catenin-specific inhibitor, enhances the responsiveness

of these cancer cells to the BRAFV600E inhibitor PLX4720, leading to substantial growth arrest, cell apoptosis in vitro, and tumor regression and differentiation in vivo [30,79,80].

The β -catenin pathway's activation can cause disrupted membrane targeting of NIS, contributing significantly to 131I resistance in thyroid cancer cells [30,81].

3.2.6. Notch-Related Pathway

The Notch receptor functions as a multifunctional transmembrane protein and is involved in regulating cell maturation, development, replication, and survival. Humans possess four Notch receptors (Notch1-4) and five ligands (δ -like 1, 3, 4, and Jagged-1, -2). When a Notch receptor interacts with its ligand, it undergoes cleavage by the γ -secretase protease complex, which releases a cytoplasmic segment that moves into the nucleus to modulate gene transcription [30,82–84].

In DTC, the levels of Notch receptors and other components of the Notch signaling pathway are markedly reduced in comparison to normal thyroid tissue. Increased expression of Notch receptors in DTC can induce them to regain differentiation by enhancing thyroid-specific genes such as NIS and TPO. Additionally, Notch can reduce cancer cell growth and proliferation rates. Therefore, Notch acts as a crucial controller of thyroid-specific genes and a tumor suppressor in DTC cells [30,82,85–87].

3.3. Modulation of microRNAs

MicroRNAs (*miRNAs*) are small, unpaired noncoding RNAs that influence gene expression by attaching to the 3'-untranslated region (3'-UTR) of target mRNAs, disrupting their integrity and inhibiting molecular translation [30,88].

Several *miRNAs*, including *miRNA-146b-3p* as well as *miRNA-339*, regulate NIS expression in PTCs by binding to *NIS mRNA 3'-UTR*. *MiRNAs* such as *miRNA-339-5p* and *miRNA-195* also impact RAI uptake in PTCs, with *miRNA-339-5p* being moderately increased and *miRNA-195* significantly decreased in these cancers. Additionally, *miRNA-146b-3p* disrupts RAI uptake by binding to *PAX8* and *NIS mRNA*, contributing to cancer cell proliferation and migration while inhibiting apoptosis. Further *miRNAs*, such as *miRNA-106a*, *miRNA-let-7*, and *miRNA-875*, reduce NIS expression or affect its membrane localization, promoting dedifferentiation in TC. Targeting these *miRNAs* to improve RAI uptake and NIS expression offers a potential therapeutic strategy for TC [30,88–96].

4. Management of RAI-DTC

4.1. Monitoring

RAI-DTC is asymptomatic for years. Thus, a careful clinical and laboratory assessment should be performed. Every patient with metastatic TC depends on thyroid function regulation to keep the TSH value suppressed. Therefore, laboratory evaluations ought to involve TSH, fT4, and calcium level post-surgery hypoparathyroidism every 6–12 months. Tumor burden can be evaluated using Tg levels, knowing that Tg doubling time under one year indicates negative predicted outcome and suggests rapid progression of the disease [1,27,97–100].

Regular imaging every 6 to 12 months using CT scanning and implementing RECIST criteria helps to evaluate the growth of neoplastic mass. Additionally, 18-FDG-PET/CT scanning may provide prognostic indicators in advanced TC. According to studies, patients with lesions with increased glucose uptake have negative prognoses and shorter survival than patients with FDG-PET-negative tumor lesions. The extension of local tumor as well as complications can also be appreciated by other imaging techniques such as bronchoscopy or esophagoduodenoscopy [1,27,97–100].

4.2. Local Treatments

In order to sustain the patients' standard of living, before starting targeted therapies with tyrosine kinase inhibitors (TKIs), a complete anamnesis regarding age, medical history, size, position, and rate of lesion progression should be completed. Surgery, including the

dissection of the central and lateral regions, remains the standard in therapeutic management of locoregional relapse every time the surgical procedure can be safely performed amid re-intervention or distant spreading of the originating malignancy [101–103]. Studies have demonstrated that surgery and external-beam radiation treatment (EBRT) in doses of 40–50 Gy for patients older than 45 years, offers a locoregional control and an overall good prognosis in most cases. Local therapies are recommended before targeted therapies for patients with lung nodules or bone metastasis [1,27,97–100].

In case of infiltration of the trachea, ablative laser therapy should be performed in order to reduce the obstruction. This treatment can be repeated every 6 months. In case of a compression of the trachea as a result of the local tumor mass, an endotracheal stent should be used. Surgery is essential for the resection of bone and lung metastases [1,27,97,99,101].

Depending on the evolution of the TC and the behavior of metastases, percutaneous interventional techniques may be vascular, ablative, or consolidative treatments. Trans-arterial chemoembolization (TACE) is a vascular technique, and it is part of the category of palliative therapies for both advanced hepatocellular cancers and aggressive TCs. This procedure is used in the case of metastases that do not exceed 3 cm and with liver damage of less than 30% [1,104,105].

Radiofrequency thermoablation uses electromagnetic waves that cause movement and heating of the tumor cells. In other words, the technique is used to reduce the volume of the metastatic lesion in the case of metastases involving the lymph nodes, bones, liver, and lung [1,27,106–108].

Lymph nodes with metastases smaller than 1 cm may be monitored periodically every 6 months and if they increase in size, ultrasound-guided percutaneous ethanol ablation can be performed. For bone metastases with osteolytic lesions, combinations of local and palliative treatments such as cementoplasty can enhance the patient's quality of life by alleviating pain and ensuring bone stability [1,27,109].

4.3. TKIs as Targeted Therapies

Since RAIR-DTC does not respond to RAI due to the previously presented mechanisms, clinical trials and preclinical studies are being conducted with new drugs that would be successful in treating these patients. Currently, TKIs are now considered the first therapeutic line to inhibit the expansion and progression of RAIR-DTC [1,28,30] (Figure 2).

Sorafenib, lenvatinib, and cabozantinib have been approved by the US Food and Drug Administration (FDA) for treating RAIR-DTC [1,30,100] (Figures 3 and 4).

Sorafenib targets RAF and blocks VEGFR1/2/3, c-KIT, RET, PDGFR, and FLT receptors. In the phase 3 DECISION trial, 417 subjects with advanced or metastatic DTC who had progressive RAIR disease were administered 400 mg of sorafenib, taken twice a day. The trial showed that 12.2% of patients receiving sorafenib achieved a partial response (PR), compared to just 0.5% in the placebo group. Progression-free survival (PFS) improved from 5.8 months to 10.8 months, while overall survival (OS) remained stable. Notably, 78% of subjects needed dose modifications due to side effects [1,4,28,30,110].

Lenvatinib (E7080), a multi-kinase oral inhibitor that targets VEGFR, FGFR, PDGFR α , RET, and KIT, was approved by the FDA in 2015 for treating RAIR-DTC. In the phase 3 SELECT trial, lenvatinib significantly improved PFS and response rates compared to placebo in RAIR-DTC patients. A sub-analysis revealed that while lenvatinib improved PFS in both younger and older patients, older patients experienced more toxicity. Despite allowing crossover after disease progression, an OS benefit was noted in older subjects. However, lenvatinib used alone was found to be less effective for treating ATC, warranting further investigation [1,4,28,30,108–114].

A phase 3 study (NCT02966093) was performed across 24 sites in China to investigate the safety and efficacy of lenvatinib in treating RAIR-DTC in this population. The results showed that a starting dose of 24 mg/day led to a significant improvement in PFS and objective response rates compared to placebo, with no new or unexpected side effects reported. These findings are consistent with the SELECT trial results [1,4,28,30,111–117].

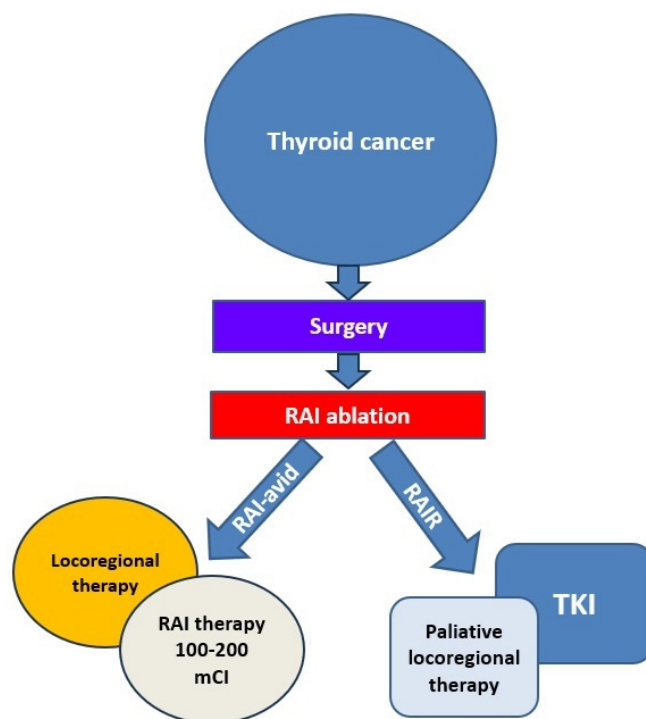


Figure 2. Treatment approach for advanced/metastatic DTC [1,28,30]. Abbreviations: RAI—radioactive iodine; RAI-avid—radioactive iodine-avid; TKI—tyrosine kinase inhibitor.

Additionally, a correlation was found between lung metastases in RAIR-DTC patients and reduced survival rates. A post hoc analysis by Tahara et al. (2021) of the SELECT data indicated that lenvatinib led to improved OS in patients with lung metastases greater than or equal to 1.0 cm, despite a crossover rate of 89%. Prompt initiation of treatment can enhance outcomes in these patients [1,4,28,30,111–117].

While lenvatinib’s toxicity is usually manageable through dose adjustments, Tahara et al. (2019) noted that shorter treatment interruptions were associated with better outcomes. This emphasizes the significance of early management of lenvatinib-related side effects to optimize its effectiveness in RAIR-DTC subjects [1,4,28,30,111–117].

Cabozantinib is an inhibitor targeting c-MET, RET, and VEGFR that has received FDA approval for MTC after the phase 3 trial, demonstrating a 7.2-month rise in median PFS. Initial phase 1 studies revealed a 62% objective response rate in eight subjects with DTC who had undergone prior VEGFR-targeted treatment [1,4,28,30,118–122].

Building on these encouraging findings, a phase 2 study highlighted cabozantinib’s efficacy in subjects with RAIR-DTC that had disease progression after previous treatments. A later phase 3 trial further confirmed that cabozantinib significantly improved PFS among RAIR-DTC subjects that lacked conventional treatment options [1,4,28,30,118–122].

On 17 September 2021, cabozantinib received FDA approval for use in adults and pediatric patients 12 years and older with locally advanced or metastatic DTC who showed progression following previous VEGFR-targeted therapy. This approval marked a significant advancement in treatment options, providing hope for patients facing limited alternatives and demonstrating the viability of cabozantinib in treating RAIR-DTC. Additionally, ongoing research is expected to further elucidate the long-term effects and potential combination therapies involving cabozantinib to enhance outcomes for patients with this challenging disease [1,4,28,30,118–122].

Vandetanib is an inhibitor that targets multiple pathways, including VEGFR2/3, EGFR, c-KIT, and RET. Although its use in treating RAIR-DTC has not yet received approval, an earlier phase 2 randomized trial indicated a favorable response in this subject population, with a median PFS of 11.1 months in the vandetanib group, versus 5.9 months in the placebo group. A phase 3 trial (VERIFY) was completed in 2020 with 119 patients suffering

from progressive RAIR-DTCs. The preliminary data indicated no significant difference in PFS between the vandetanib and placebo groups (10 months vs. 5.7 months, $p = 0.08$) (NCT01876784, ClinicalTrials.gov accessed on 20 August 2024) [1,4,28,30,123].

Long-term use of TKIs in clinical practice can lead to moderate to severe adverse effects, particularly in patients aged 65 and older, necessitating careful monitoring for dosage adjustments [113]. Additionally, research showed that RAIR-TC patients often develop TKI resistance, resulting in “tumor escape” due to alterations in alternative signaling pathways, such as HER2/3 hyper-expression. Nonetheless, a retrospective analysis indicates that RAIR-DTC refractory to initial TKI treatment may continue to show effectiveness with salvage therapies, including candetanib, cabozantinib, sunitinib, pazopanib, and vemurafenib [1,4,28,30,124,125].

Table 2 provides an overview of the targeted kinase inhibitors evaluated in randomized-controlled trials for advanced, metastatic RAIR-DTC.

Table 2. FDA-approved TKIs for RAIR-DTC (PubMed search based on key terms “RAIR-DTC”, “tyrosine kinase inhibitors”) [112,121,123].

Randomized Control Trial	Drug	Molecular Targets	Phase	Results: PFS	ORR
DECISION [123]	Sorafenib	VEGFR, PDGFR, c-KIT, RET, RAF	III	from 10.8 months to 5.8 months (placebo)	12.2% (vs. 0.5%)
SELECT [112]	Lenvatinib	VEGFR, PDGFR, c-KIT, RET, FGFR	III	18.3 months vs. 3.6 months (placebo)	64.8% (vs. 1.5)
COSMIC-311 [121]	Cabozantinib	VEGFR, RET, c-MET, FLT3, TEK	III	11.0 months vs. 1.9 months placebo	15%

Abbreviations: PFS—progression-free survival; ORR—overall response rate, c-MET—hepatocyte growth factor receptor or HGFR; c-KIT—stem cell factor receptor or SCFR; EGFR—epidermal growth factor receptor; FGFR—fibroblast growth factor receptor; FLT3—FMS-like tyrosine kinase 3 (or CD135); PDGFR—platelet-derived growth factor receptor; RET—ret proto-oncogene; RAF—rapidly accelerated fibrosarcoma; VEGFR—vascular endothelial growth factor receptor.

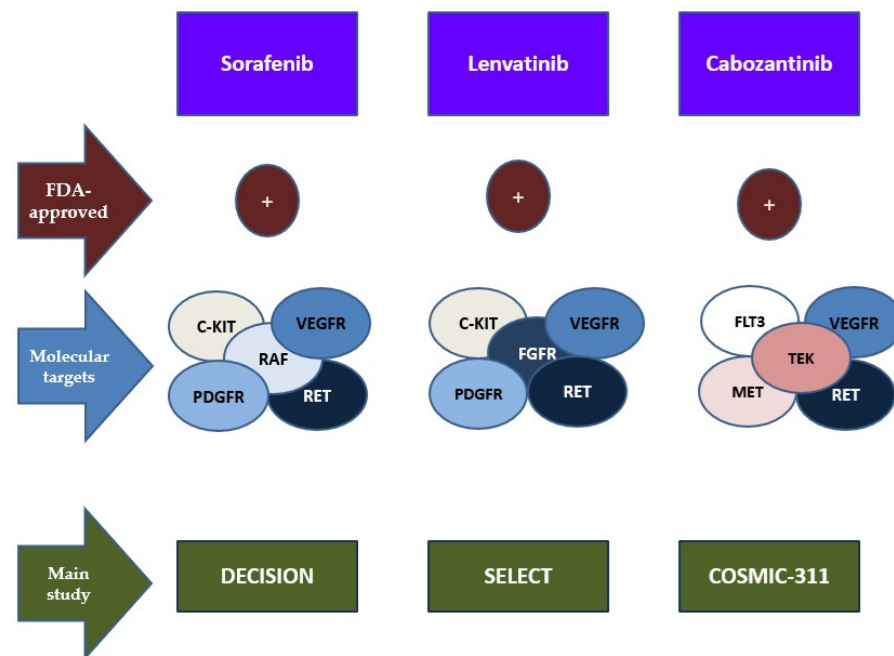


Figure 3. FDA-approved TKIs for RAIR-DTC [112,121,123]. Abbreviations: FDA—Food and Drug Administration; c-MET—hepatocyte growth factor receptor or HGFR; c-KIT—stem cell factor receptor or SCFR; EGFR—epidermal growth factor receptor; FGFR—fibroblast growth factor receptor; FLT3—FMS-like tyrosine kinase 3 (or CD135); PDGFR—platelet-derived growth factor receptor; RET—ret proto-oncogene; RAF—rapidly accelerated fibrosarcoma; VEGFR—vascular endothelial growth factor receptor; DECISION ClinicalTrials.gov number, NCT00984282; SELECT ClinicalTrials.gov number, NCT01321554; COSMIC-311 ClinicalTrials.gov number NCT03690388.

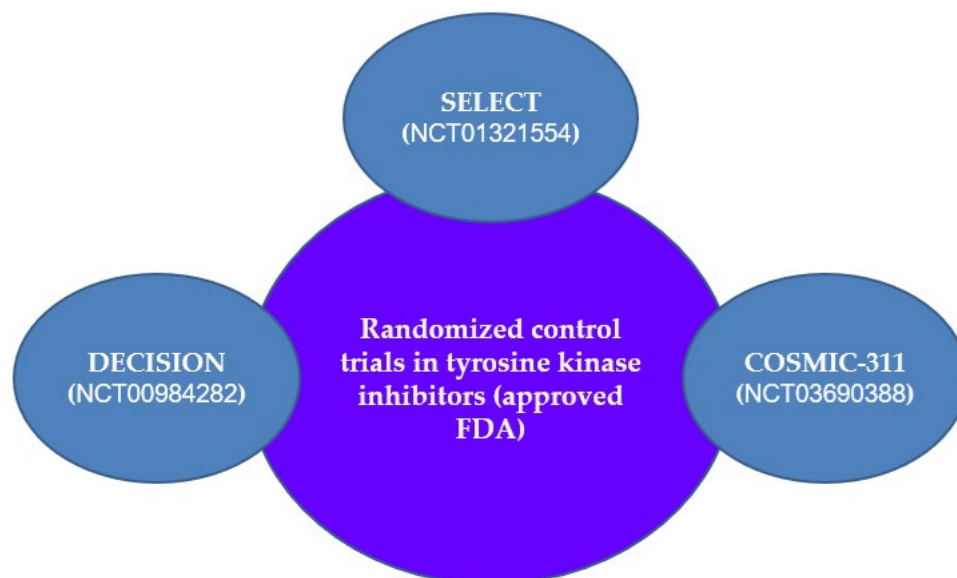


Figure 4. Randomized control trial of FDA-approved TKIs for RAI-R DTC [112,121,123]. Abbreviations: FDA—Food and Drug Administration; DECISION ClinicalTrials.gov number, NCT00984282; SELECT ClinicalTrials.gov number, NCT01321554; COSMIC-311 ClinicalTrials.gov number NCT03690388.

4.4. Redifferentiation Therapy

In light of the pathophysiology associated with RAI refractoriness, research has focused on re-inducing NIS expression to restore RAI avidity. Inhibitors targeting the MAPK pathway have demonstrated potential in facilitating redifferentiation process in RAI-R DTC. For instance, the MEK inhibitor selumetinib increased RAI avidity in 12 out of 20 RAI-R DTC patients, enabling 8 of these subjects to receive RAI treatment, in which 7 exhibited a partial response; however, information on the duration of these responses is not available [4,126,127].

Additionally, the application of *BRAF* inhibitors, including vemurafenib and dabrafenib, has been investigated in patients with *BRAF*-mutated RAI-R TC, producing similar outcomes. Nevertheless, it has been noted that thyroid cancers with *BRAF* pathogenic variants tend to respond less favorably to redifferentiation therapies, suggesting that a stronger inhibition of the MAPK pathway may be necessary, potentially through a dual therapy of *BRAF* and MEK inhibitors. A short duration of these treatment regimens (typically lasting 4–8 weeks in most studies) may lead to significantly lower toxicity compared to long-term use of multi-kinase inhibitors (MKIs), thus alleviating some of the economic burdens associated with treatment. While redifferentiation therapies appear promising, the current evidence regarding their clinical efficacy remains preliminary, necessitating larger clinical trials to confirm these results [4,126–134].

4.5. Immunotherapy

The introduction of checkpoint blockade therapies, such as anti-PD-1, PD-L1, and PD-L1-4, marks a significant advancement in treating various tumors. Current research indicates that PD-L1 could be used as a prognostic biomarker for PTC as well as indicate recurrence in MTC. A retrospective analysis identified high PD-L1 level in ATC, correlating with worse overall life expectancy plus PFS, positioning PD-L1 as a possible predictive marker of ATC outcomes. Furthermore, immunotherapy approaches have been studied in subjects with advanced RAI-R DTC [28,135–137].

The non-randomized phase Ib KEYNOTE-028 trial (NCT02054806) investigated the effectiveness of pembrolizumab in 22 patients with advanced RAI-R DTC expressing PD-L1. Pembrolizumab, a PD-1 antibody, was administered biweekly at a dose of 10 mg/kg for

a maximum of 24 months. Among the participants, two patients (9%) showed a partial response, with durations of response ranging from 8 to 20 months. Median PFS was 7 months, with the median OS yet to be reached. Adverse events occurred in 18 subjects (82%), with gastrointestinal distress and tiredness being the most common [28,138].

A phase I/II trial (NCT02404441) involving 30 ATC patients treated with spartalizumab (400 mg every 4 weeks) demonstrated an overall response rate (ORR) of 17% and disease control in 27%. Common adverse events included gastrointestinal discomfort, such as diarrhea, pruritus, fatigue, and hematologic and oncologic complications. In another phase 2 trial led by Capdevila, 42 patients received spartalizumab (400 mg/month), with an ORR of 19%. PD-L1-positive patients responded better (29% vs. 0%) and those with the *BRAF* pathogenic variant had long-lasting responses, with a 1-year survival rate of 52.1% [28,139,140].

A phase 2 trial assessing pembrolizumab in combination with chemoradiotherapy in three subjects with ATC initially showed favorable tumor reactions, but every patient died within 6 months from metastases or pulmonary disorders, highlighting concerns over the high toxicity of chemoradiotherapy in ATC [28,141]. According to very recent data, PD-L1 expression is not correlated with the response to combined treatment [142,143].

The novelty of the topic is as follows: immunotherapy in radioiodine-refractory thyroid cancer is an emerging and innovative area of research, offering new potential for patient recruitment. Despite its limited accessibility, it provides an additional prognostic approach and underscores the importance of a multidisciplinary team in optimizing patient outcomes. [27,30,144,145].

5. Conclusions and Future Perspectives

Despite the generally good prognosis of thyroid tumors, a small portion of subjects with advanced or progressive TC will not respond to radioiodine treatment, which is responsible for the majority of TC-related deaths. Significant efforts have been devoted to understanding the molecular mechanisms behind this, leading to notable advancements in identifying the genetic and epigenetic changes associated with iodine resistance. This progress has facilitated the development of several possible treatments for RAIR-DTC.

Three TKIs are approved for RAIR-DTC treatment, and several more are in clinical trials. However, the considerable toxicity related to these drugs presents serious concerns. Given this risk, the use of TKIs should be restricted to carefully selected patient populations, with thorough evaluations and interdisciplinary input from experienced clinicians required before personalizing treatment or considering clinical trial enrollment.

Redifferentiation therapies, particularly those involving *BRAF* and *MEK* antagonists, have proven notable progress in enhancing responsiveness in RAIR-DTC patients, offering comparable response to TKIs with reduced adverse effects. PD-1/PD-L1 blockade, a key immunotherapeutic approach in oncology, shows promise, but its application in RAIR-DTC is still not well established, requiring larger studies to evaluate its potential.

Looking ahead, the advancement of targeted therapies, such as TKIs, MAPK inhibitors, and checkpoint inhibitors, holds significant promise for RAIR-DTC. Combination therapies targeting different pathways may offer new treatment options, with dual targeting of key molecules like *BRAF* and *MEK* potentially overcoming compensatory mechanisms that lead to drug resistance.

Author Contributions: Conceptualization, I.-A.V., E.P., D.I., A.L.G., and A.M.G.; methodology, I.-A.V., D.I., A.L.G., and A.M.G.; software, I.-A.V., E.P., D.I., and A.M.G.; validation, I.-A.V., E.P., A.L.G., and A.M.G.; formal analysis, I.-A.V., D.I., A.L.G., and A.M.G.; investigation, I.-A.V., E.P., N.D., A.C., D.I., and A.M.G.; resources, I.-A.V., E.P., N.D., A.C., and A.M.G.; data curation, I.-A.V., E.P., D.I., A.L.G., and A.M.G.; writing—original draft preparation, I.-A.V., and A.M.G.; writing—review and editing, I.-A.V., and A.M.G.; visualization, I.-A.V., and A.M.G.; supervision, A.M.G.; project administration, I.-A.V., E.P., D.I., A.L.G., and A.M.G.; funding acquisition, I.-A.V., E.P., D.I., A.L.G., and A.M.G. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by “Carol Davila” University of Medicine and Pharmacy, Bucharest, Romania, granted via institutional program “Publish not Perish”.

Acknowledgments: This work is part of PhD research of PhD Doctoral School of “Carol Davila” University of Medicine and Pharmacy, entitled “Molecular and immunohistochemical profile of Radioiodine-Refractory Thyroid Cancers”. Publication of this paper was supported by “Carol Davila” University of Medicine and Pharmacy, Bucharest, Romania through the institutional program “Publish not Perish”.

Conflicts of Interest: The authors declare no conflicts of interest.

Abbreviations

TC	Thyroid cancer
DTC	Differentiated thyroid cancer
RAI	Radioactive iodine
TSH	Thyroid-stimulating hormone
MAPK	Mitogen-activated protein kinase
PI3K/mTOR/Akt	Phosphatidylinositol-3-kinase/mammalian target of rapamycin/protein kinase B
BRAF	V-Raf mouse sarcoma virus oncogene homologous B1
TGF- β	Transforming growth factor- β
NIS	Sodium iodine symporter
PTEN	Phosphatase and tensin homolog
ATC	Anaplastic thyroid cancer
TERT	Telomerase reverse transcriptase
RAIR-DTC	Radioiodine-refractory differentiated thyroid cancer
SLC5A5	Solute carrier family 5A
cAMP	Cyclic adenosine monophosphate
NUE	NIS upstream enhances
PKA	Protein kinase A
PAX8	Paired box gene-8
Ref-1	Redox effector factor-1
CREM	cAMP-response element modulator
MAPKKK	Mitogen activated protein kinase kinase kinase
V600E	B-raf protein residue 600 from glutamic acid to valine
AHR	Aromatic hydrocarbon receptor
WT1	Wilm tumor gene 1
NTRK	Neurotrophic receptor tyrosine kinase
TRK	Tropomyosin receptor kinase
TERT _p	TERT promoter
ALK	Anaplastic lymphoma kinase
CRKL-C3G	Adaptor protein-Rap guanine nucleotide exchange factor 1
MEKK2/3-MEK5-ERK5	Mitogen-activated protein kinase kinase kinase 2/3-Mitogen-activated protein kinase kinase 5-extracellular signal regulated kinase 5
JAK-STAT	Janus kinase-signal transducer and activator of transcription
STRN	Recurrent striatal protein
NGS	Next-generation sequencing
TKR	Tyrosine kinase membrane receptor
GDNK	Glial cell line-derived neurotrophic factor
GFL	GDNF family ligand
DSBs	DNA double-strand breaks
PPAR- γ	Peroxisome proliferator-activated receptor gamma
PPFP	PAX8-PPAR- γ fusion protein
TPO	Thyroid peroxidase
TG	Thyroglobulin
TSHR	Thyroid-stimulating hormone receptor
SWI/SNF	SWIItch/sucrose nonfermentable

BAF	BRG1/BRM related factor
PBAF	Polybromine-related factor
ncBAF	Atypical BAF
TF	Transcription factors
GPCR	G protein-coupled receptor
PD-L1	Tumor programmed death-ligand 1
ERK	Extracellular-signal-regulated kinase
JNK/SAPK	Jun kinase
MEK	Mitogen-activated protein kinase/extracellular signal-regulated kinase kinase
IGF-2	Insulin-like growth factor 2
IR-A	Insulin receptor subtype A
IIGFs	Insulin/insulin-like growth factor systems
DDRs	Discoid domain receptors
RUNX2	Runt-related transcription factor 2
mTORC2	mTOR complex 2
NADPH	Nicotinamide adenine dinucleotide phosphate
NOX4	NADPH oxidase 4
ROS	Reactive oxygen species
TCF	T cell factor
CSCs	Cancer stem cells
LSD1	Lysine-specific histone demethylase 1 A
APC2	Adenomatous polyposis coli 2
DKK1	Dickkopf-related protein 1
Notch1-4	Notch receptors
miRNAs	MicroRNAs
3'-UTR	3'-untranslated region
ft4	Free thyroxine
TKIs	Tyrosine kinase inhibitors
EBRT	External-beam radiation therapy
TACE	Trans-arterial chemoembolization
VEGFR1/2/3	Vascular endothelial growth factor receptor 1, 2, 3
c-KIT	Cellular kit
PDGFR	Platelet-derived growth factor receptor
FLT	Fms-like tyrosine kinase
FDA	Food and Drug Administration
PR	Partial response
PFS	Progression-free survival
OS	Overall survival
FGFR	Fibroblast growth factor receptor
c-MET	Mesenchymal-epithelial transition factor
MTC	Medullary thyroid carcinoma
EGFR	Epidermal growth factor receptor
ORR	Overall response rate
RAF	Rapidly accelerated fibrosarcoma
MKIs	Multi-kinase inhibitors
CTLA-4	Cytotoxic T-lymphocyte antigen 4

References

1. Fugazzola, L.; Elisei, R.; Fuhrer, D.; Jarzab, B.; Leboulleux, S.; Newbold, K.; Smit, J. 2019 European Thyroid Association Guidelines for the Treatment and Follow-Up of Advanced Radioiodine-Refractory Thyroid Cancer. *Eur. Thyroid J.* **2019**, *8*, 227–245. [[CrossRef](#)] [[PubMed](#)]
2. Shobab, L.; Gomes-Lima, C.; Zeymo, A.; Feldman, R.; Jonklaas, J.; Wartofsky, L.; Burman, K.D. Clinical, Pathological, and Molecular Profiling of Radioactive Iodine Refractory Differentiated Thyroid Cancer. *Thyroid* **2019**, *29*, 1262–1268. [[CrossRef](#)] [[PubMed](#)]
3. Durante, C.; Haddy, N.; Baudin, E.; Leboulleux, S.; Hartl, D.; Travagli, J.P.; Caillou, B.; Ricard, M.; Lombroso, J.D.; De Vathaire, F.; et al. Long-Term Outcome of 444 Patients with Distant Metastases from Papillary and Follicular Thyroid Carcinoma: Benefits and Limits of Radioiodine Therapy. *J. Clin. Endocrinol. Metab.* **2006**, *91*, 2892–2899. [[CrossRef](#)] [[PubMed](#)]

4. Nervo, A.; Retta, F.; Ragni, A.; Piovesan, A.; Gallo, M.; Arvat, E. Management of Progressive Radioiodine-Refractory Thyroid Carcinoma: Current Perspective. *Cancer Manag. Res.* **2022**, *14*, 3047–3062. [[CrossRef](#)] [[PubMed](#)]
5. Saïe, C.; Wassermann, J.; Mathy, E.; Chereau, N.; Leenhardt, L.; Tezenas du Montcel, S.; Buffet, C. Impact of age on survival in radioiodine refractory differentiated thyroid cancer patients. *Eur. J. Endocrinol.* **2021**, *184*, 667–676. [[CrossRef](#)]
6. Brenner, H. Long-term survival rates of cancer patients achieved by the end of the 20th century: A period analysis. *Lancet* **2002**, *360*, 1131–1135. [[CrossRef](#)]
7. Cooper, D.S.; Doherty, G.M.; Haugen, B.R.; Kloos, R.T.; Lee, S.L.; Mandel, S.J.; Mazzaferri, E.L.; McIver, B.; Pacini, F.; Schlumberger, M.; et al. Revised American Thyroid Association Management Guidelines for Patients with Thyroid Nodules and Differentiated Thyroid Cancer. *Thyroid* **2009**, *19*, 1167–1214. [[CrossRef](#)]
8. Saftencu, M.; Braicu, C.; Cojocneanu, R.; Buse, M.; Irimie, A.; Piciu, D.; Berindan-Neagoe, I. Gene Expression Patterns Unveil New Insights in Papillary Thyroid Cancer. *Medicina* **2019**, *55*, 500. [[CrossRef](#)]
9. Filetti, S.; Durante, C.; Hartl, D.; Leboulleux, S.; Locati, L.D.; Newbold, K.; Papotti, M.G.; Berruti, A. Thyroid cancer: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann. Oncol.* **2019**, *30*, 1856–1883. [[CrossRef](#)]
10. Mazzaferri, E.L. An Overview of the Management of Papillary and Follicular Thyroid Carcinoma. *Thyroid* **1999**, *9*, 421–427. [[CrossRef](#)]
11. Hanahan, D.; Weinberg, R.A. Hallmarks of Cancer: The Next Generation. *Cell* **2011**, *144*, 646–674. [[CrossRef](#)] [[PubMed](#)]
12. Agrawal, N.; Akbani, R.; Aksoy, B.A.; Ally, A.; Arachchi, H.; Asa, S.L.; Auman, J.T.; Balasundaram, M.; Balu, S.; Baylin, S.B.; et al. Integrated Genomic Characterization of Papillary Thyroid Carcinoma. *Cell* **2014**, *159*, 676–690. [[CrossRef](#)]
13. Kim, K.H.; Kang, D.W.; Kim, S.H.; Seong, I.O.; Kang, D.Y. Mutations of the BRAF Gene in Papillary Thyroid Carcinoma in a Korean Population. *Yonsei Med. J.* **2004**, *45*, 818–821. [[CrossRef](#)]
14. Soares, P.; Trovisco, V.; Rocha, A.S.; Lima, J.; Castro, P.; Preto, A.; Máximo, V.; Botelho, T.; Seruca, R.; Sobrinho-Simões, M. BRAF mutations and RET/PTC rearrangements are alternative events in the etiopathogenesis of PTC. *Oncogene* **2003**, *22*, 4578–4580. [[CrossRef](#)] [[PubMed](#)]
15. Nikiforova, M.N.; Kimura, E.T.; Gandhi, M.; Biddinger, P.W.; Knauf, J.A.; Basolo, F.; Zhu, Z.; Giannini, R.; Salvatore, G.; Fusco, A.; et al. BRAF Mutations in Thyroid Tumors Are Restricted to Papillary Carcinomas and Anaplastic or Poorly Differentiated Carcinomas Arising from Papillary Carcinomas. *J. Clin. Endocrinol. Metab.* **2003**, *88*, 5399–5404. [[CrossRef](#)] [[PubMed](#)]
16. Fugazzola, L.; Mannavola, D.; Cirello, V.; Vannucchi, G.; Muzza, M.; Vicentini, L.; Beck-Peccoz, P. BRAF mutations in an Italian cohort of thyroid cancers. *Clin. Endocrinol.* **2004**, *61*, 239–243. [[CrossRef](#)] [[PubMed](#)]
17. Trovisco, V.; Vieira de Castro, I.; Soares, P.; Máximo, V.; Silva, P.; Magalhães, J.; Abrosimov, A.; Guiu, X.M.; Sobrinho-Simões, M. BRAF mutations are associated with some histological types of papillary thyroid carcinoma. *J. Pathol.* **2004**, *202*, 247–251. [[CrossRef](#)]
18. Cappola, A.R.; Mandel, S.J. Molecular Testing in Thyroid Cancer. *JAMA* **2013**, *309*, 1529–1530. [[CrossRef](#)]
19. Biondi, B.; Filetti, S.; Schlumberger, M. Thyroid-hormone therapy and thyroid cancer: A reassessment. *Nat. Clin. Pract. Endocrinol. Metab.* **2005**, *1*, 32–40. [[CrossRef](#)]
20. Kimura, E.T.; Nikiforova, M.N.; Zhu, Z.; Knauf, J.A.; Nikiforov, Y.E.; Fagin, J.A. High prevalence of BRAF mutations in thyroid cancer: Genetic evidence for constitutive activation of the RET/PTC-RAS-BRAF signaling pathway in papillary thyroid carcinoma. *Cancer Res.* **2003**, *63*, 1454–1457.
21. Kim, M.; Jeon, M.J.; Oh, H.-S.; Park, S.; Kim, T.Y.; Shong, Y.K.; Kim, W.B.; Kim, K.; Kim, W.G.; Song, D.E. BRAF and RAS Mutational Status in Noninvasive Follicular Thyroid Neoplasm with Papillary-Like Nuclear Features and Invasive Subtype of Encapsulated Follicular Variant of Papillary Thyroid Carcinoma in Korea. *Thyroid* **2018**, *28*, 504–510. [[CrossRef](#)] [[PubMed](#)]
22. Nikiforov, Y.E.; Seethala, R.R.; Tallini, G.; Baloch, Z.W.; Basolo, F.; Thompson, L.D.R.; Barletta, J.A.; Wenig, B.M.; Al Ghuzlan, A.; Kakudo, K.; et al. Nomenclature Revision for Encapsulated Follicular Variant of Papillary Thyroid Carcinoma. *JAMA Oncol.* **2016**, *2*, 1023–1029. [[CrossRef](#)] [[PubMed](#)]
23. Prete, A.; Borges de Souza, P.; Censi, S.; Muzza, M.; Nucci, N.; Sponziello, M. Update on Fundamental Mechanisms of Thyroid Cancer. *Front. Endocrinol.* **2020**, *11*, 102. [[CrossRef](#)] [[PubMed](#)]
24. Al Rasheed, M.R.H.; Xu, B. Molecular Alterations in Thyroid Carcinoma. *Surg. Pathol. Clin.* **2019**, *12*, 921–930. [[CrossRef](#)] [[PubMed](#)]
25. Oikonomou, E.; Koustas, E.; Goulielmaki, M.; Pintzas, A. BRAF vs. RAS oncogenes: Are mutations of the same pathway equal? Differential signalling and therapeutic implications. *Oncotarget* **2014**, *5*, 11752–11777. [[CrossRef](#)]
26. Ratajczak, M.; Gawel, D.; Godlewska, M. Novel Inhibitor-Based Therapies for Thyroid Cancer—An Update. *Int. J. Mol. Sci.* **2021**, *22*, 11829. [[CrossRef](#)]
27. Aashiq, M.; Silverman, D.A.; Na'ara, S.; Takahashi, H.; Amit, M. Radioiodine-Refractory Thyroid Cancer: Molecular Basis of Redifferentiation Therapies, Management, and Novel Therapies. *Cancers* **2019**, *11*, 1382. [[CrossRef](#)]
28. Yu, Q.; Zhang, X.; Li, L.; Zhang, C.; Huang, J.; Huang, W. Molecular basis and targeted therapies for radioiodine refractory thyroid cancer. *Asia Pac. J. Clin. Oncol.* **2023**, *19*, 279–289. [[CrossRef](#)]
29. Liu, J.; Liu, Y.; Lin, Y.; Liang, J. Radioactive Iodine-Refractory Differentiated Thyroid Cancer and Redifferentiation Therapy. *Endocrinol. Metab.* **2019**, *34*, 215–225. [[CrossRef](#)]

30. Shen, H.; Zhu, R.; Liu, Y.; Hong, Y.; Ge, J.; Xuan, J.; Niu, W.; Yu, X.; Qin, J.J.; Li, Q. Radioiodine-refractory differentiated thyroid cancer: Molecular mechanisms and therapeutic strategies for radioiodine resistance. *Drug Resist. Updates* **2024**, *72*, 101013. [[CrossRef](#)]
31. Davies, H.; Bignell, G.R.; Cox, C.; Stephens, P.; Edkins, S.; Clegg, S.; Teague, J.; Woffendin, H.; Garnett, M.J.; Bottomley, W.; et al. Mutations of the BRAF gene in human cancer. *Nature* **2002**, *417*, 949–954. [[CrossRef](#)] [[PubMed](#)]
32. Valvo, V.; Nucera, C. Coding Molecular Determinants of Thyroid Cancer Development and Progression. *Endocrinol. Metab. Clin. N. Am.* **2019**, *48*, 37–59. [[CrossRef](#)] [[PubMed](#)]
33. Spitzweg, C.; Bible, K.C.; Hofbauer, L.C.; Morris, J.C. Advanced radioiodine-refractory differentiated thyroid cancer: The sodium iodide symporter and other emerging therapeutic targets. *Lancet Diabetes Endocrinol.* **2014**, *2*, 830–842. [[CrossRef](#)]
34. Song, E.; Jin, M.; Jang, A.; Jeon, M.J.; Song, D.E.; Yoo, H.J.; Kim, W.B.; Shong, Y.K.; Kim, W.G. Mutation in Genes Encoding Key Functional Groups Additively Increase Mortality in Patients with BRAFV600E-Mutant Advanced Papillary Thyroid Carcinoma. *Cancers* **2021**, *13*, 5846. [[CrossRef](#)]
35. Efanov, A.A.; Brenner, A.V.; Bogdanova, T.I.; Kelly, L.M.; Liu, P.; Little, M.P.; Wald, A.I.; Hatch, M.; Zurnadzy, L.Y.; Nikiforova, M.N.; et al. Investigation of the Relationship between Radiation Dose and Gene Mutations and Fusions in Post-Chernobyl Thyroid Cancer. *J. Natl. Cancer Inst.* **2018**, *110*, 371–378, Erratum in *J. Natl. Cancer Inst.* **2018**, *110*, 685. [[CrossRef](#)]
36. Cocco, E.; Scaltriti, M.; Drilon, A. NTRK fusion-positive cancers and TRK inhibitor therapy. *Nat. Rev. Clin. Oncol.* **2018**, *15*, 731–747. [[CrossRef](#)]
37. Prasad, M.L.; Vyas, M.; Horne, M.J.; Virk, R.K.; Morotti, R.; Liu, Z.; Tallini, G.; Nikiforova, M.N.; Christison-Lagay, E.R.; Udelsman, R.; et al. NTRK fusion oncogenes in pediatric papillary thyroid carcinoma in northeast United States. *Cancer* **2016**, *122*, 1097–1107. [[CrossRef](#)]
38. Bodnar, A.G.; Ouellette, M.; Frolkis, M.; Holt, S.E.; Chiu, C.P.; Morin, G.B.; Harley, C.B.; Shay, J.W.; Lichtsteiner, S.; Wright, W.E. Extension of life-span by introduction of telomerase into normal human cells. *Science* **1998**, *279*, 349–352. [[CrossRef](#)] [[PubMed](#)]
39. Hafezi, F.; Perez Bercoff, D. The Solo Play of TERT Promoter Mutations. *Cells* **2020**, *9*, 749. [[CrossRef](#)]
40. Liu, Y.; Li, Z.; Tang, X.; Li, M.; Shi, F. Association between hTERT Polymorphisms and Female Papillary Thyroid Carcinoma. *Recent Pat. Anticancer Drug Discov.* **2019**, *14*, 268–279. [[CrossRef](#)]
41. Yang, X.; Li, J.; Li, X.; Liang, Z.; Gao, W.; Liang, J.; Cheng, S.; Lin, Y. TERT Promoter Mutation Predicts Radioiodine-Refractory Character in Distant Metastatic Differentiated Thyroid Cancer. *J. Nucl. Med.* **2017**, *58*, 258–265. [[CrossRef](#)] [[PubMed](#)]
42. Jang, E.K.; Song, D.E.; Sim, S.Y.; Kwon, H.; Choi, Y.M.; Jeon, M.J.; Han, J.M.; Kim, W.G.; Kim, T.Y.; Shong, Y.K.; et al. NRAS codon 61 mutation is associated with distant metastasis in patients with follicular thyroid carcinoma. *Thyroid* **2014**, *24*, 1275–1281. [[CrossRef](#)]
43. Zou, M.; Baitei, E.Y.; Alzahrani, A.S.; BinHumaid, F.S.; Alkhafaji, D.; Al-Rijjal, R.A.; Meyer, B.F.; Shi, Y. Concomitant RAS, RET/PTC, or BRAF mutations in advanced stage of papillary thyroid carcinoma. *Thyroid* **2014**, *24*, 1256–1266. [[CrossRef](#)] [[PubMed](#)]
44. Kelly, L.M.; Barila, G.; Liu, P.; Evdokimova, V.N.; Trivedi, S.; Panebianco, F.; Gandhi, M.; Carty, S.E.; Hodak, S.P.; Luo, J.; et al. Identification of the transforming STRN-ALK fusion as a potential therapeutic target in the aggressive forms of thyroid cancer. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, 4233–4238. [[CrossRef](#)]
45. Lee, H.; Krishnan, V.; Wirth, L.J.; Nucera, C.; Venturina, M.; Sadow, P.M.; Mita, A.; Sacks, W. Case Report of CCDC149-ALK Fusion: A Novel Genetic Alteration and a Clinically Relevant Target in Metastatic Papillary Thyroid Carcinoma. *Thyroid* **2022**, *32*, 1580–1585. [[CrossRef](#)]
46. Subbiah, V.; Yang, D.; Velcheti, V.; Drilon, A.; Meric-Bernstam, F. State-of-the-Art Strategies for Targeting RET-Dependent Cancers. *J. Clin. Oncol.* **2020**, *38*, 1209–1221. [[CrossRef](#)]
47. Ishizaka, Y.; Itoh, F.; Tahira, T.; Ikeda, I.; Sugimura, T.; Tucker, J.; Fertitta, A.; Carrano, A.V.; Nagao, M. Human ret proto-oncogene mapped to chromosome 10q11.2. *Oncogene* **1989**, *4*, 1519–1521. [[PubMed](#)]
48. Takahashi, M.; Buma, Y.; Iwamoto, T.; Inaguma, Y.; Ikeda, H.; Hiai, H. Cloning and expression of the ret proto-oncogene encoding a tyrosine kinase with two potential transmembrane domains. *Oncogene* **1988**, *3*, 571–578.
49. Chi, X.; Michos, O.; Shakya, R.; Riccio, P.; Enomoto, H.; Licht, J.D.; Asai, N.; Takahashi, M.; Ohgami, N.; Kato, M.; et al. Ret-dependent cell rearrangements in the Wolffian duct epithelium initiate ureteric bud morphogenesis. *Dev. Cell* **2009**, *17*, 199–209. [[CrossRef](#)]
50. Dacic, S.; Luvison, A.; Evdokimova, V.; Kelly, L.; Siegfried, J.M.; Villaruz, L.C.; Socinski, M.A.; Nikiforov, Y.E. RET rearrangements in lung adenocarcinoma and radiation. *J. Thorac. Oncol.* **2014**, *9*, 118–120. [[CrossRef](#)]
51. Romei, C.; Ciampi, R.; Elisei, R. A comprehensive overview of the role of the RET proto-oncogene in thyroid carcinoma. *Nat. Rev. Endocrinol.* **2016**, *12*, 192–202. [[CrossRef](#)] [[PubMed](#)]
52. Raman, P.; Koenig, R.J. Pax-8-PPAR- γ fusion protein in thyroid carcinoma. *Nat. Rev. Endocrinol.* **2014**, *10*, 616–623. [[CrossRef](#)] [[PubMed](#)]
53. Wang, Y.; Hoang, L.; Ji, J.X.; Huntsman, D.G. SWI/SNF Complex Mutations in Gynecologic Cancers: Molecular Mechanisms and Models. *Annu. Rev. Pathol.* **2020**, *15*, 467–492. [[CrossRef](#)] [[PubMed](#)]
54. Saqcena, M.; Leandro-Garcia, L.J.; Maag, J.L.; Tchekmedyan, V.; Krishnamoorthy, G.P.; Tamarapu, P.P.; Tiedje, V.; Reuter, V.; Knauf, J.A.; de Stanchina, E.; et al. SWI/SNF Complex Mutations Promote Thyroid Tumor Progression and Insensitivity to Redifferentiation Therapies. *Cancer Discov.* **2021**, *11*, 1158–1175. [[CrossRef](#)]

55. Szkudlinski, M.W.; Fremont, V.; Ronin, C.; Weintraub, B.D. Thyroid-stimulating hormone and thyroid-stimulating hormone receptor structure-function relationships. *Physiol. Rev.* **2002**, *82*, 473–502. [[CrossRef](#)]
56. Morgan, S.J.; Neumann, S.; Marcus-Samuels, B.; Gershengorn, M.C. Thyrotropin and Insulin-Like Growth Factor 1 Receptor Crosstalk Upregulates Sodium-Iodide Symporter Expression in Primary Cultures of Human Thyrocytes. *Thyroid* **2016**, *26*, 1794–1803. [[CrossRef](#)]
57. Chu, Y.D.; Yeh, C.T. The Molecular Function and Clinical Role of Thyroid Stimulating Hormone Receptor in Cancer Cells. *Cells* **2020**, *9*, 1730. [[CrossRef](#)]
58. Wu, Z.; Xi, Z.; Xiao, Y.; Zhao, X.; Li, J.; Feng, N.; Hu, L.; Zheng, R.; Zhang, N.; Wang, S.; et al. TSH-TSHR axis promotes tumor immune evasion. *J. Immunother. Cancer* **2022**, *10*, e004049. [[CrossRef](#)]
59. Hou, P.; Bojdani, E.; Xing, M. Induction of thyroid gene expression and radioiodine uptake in thyroid cancer cells by targeting major signaling pathways. *J. Clin. Endocrinol. Metab.* **2010**, *95*, 820–828. [[CrossRef](#)]
60. Zhang, W.; Liu, H.T. MAPK signal pathways in the regulation of cell proliferation in mammalian cells. *Cell Res.* **2002**, *12*, 9–18. [[CrossRef](#)]
61. Zhang, Z.; Liu, D.; Murugan, A.K.; Liu, Z.; Xing, M. Histone deacetylation of NIS promoter underlies BRAF V600E-promoted NIS silencing in thyroid cancer. *Endocr. Relat. Cancer* **2014**, *21*, 161–173. [[CrossRef](#)] [[PubMed](#)]
62. Nagarajah, J.; Le, M.; Knauf, J.A.; Ferrandino, G.; Montero-Conde, C.; Pillarsetty, N.; Bolaender, A.; Irwin, C.; Krishnamoorthy, G.P.; Saqccena, M.; et al. Sustained ERK inhibition maximizes responses of BrafV600E thyroid cancers to radioiodine. *J. Clin. Investig.* **2016**, *126*, 4119–4124. [[CrossRef](#)]
63. Borrelli, N.; Panebianco, F.; Condello, V.; Barletta, J.A.; Kaya, C.; Yip, L.; Nikiforova, M.N.; Nikiforov, Y.E. Characterization of Activating Mutations of the MEK1 Gene in Papillary Thyroid Carcinomas. *Thyroid* **2019**, *29*, 1279–1285. [[CrossRef](#)]
64. Petrulea, M.S.; Plantinga, T.S.; Smit, J.W.; Georgescu, C.E.; Netea-Maier, R.T. PI3K/Akt/mTOR: A promising therapeutic target for non-medullary thyroid carcinoma. *Cancer Treat. Rev.* **2015**, *41*, 707–713. [[CrossRef](#)] [[PubMed](#)]
65. Liu, J.; Dong, H.; Yang, Y.; Qian, Y.; Liu, J.; Li, Z.; Guan, H.; Chen, Z.; Li, C.; Zhang, K.; et al. Upregulation of long noncoding RNA MALAT1 in papillary thyroid cancer and its diagnostic value. *Future Oncol.* **2018**, *14*, 3015–3022. [[CrossRef](#)] [[PubMed](#)]
66. Vella, V.; Nicolosi, M.L.; Cantafio, P.; Massimino, M.; Lappano, R.; Vigneri, P.; Ciuni, R.; Gangemi, P.; Morrione, A.; Malaguarnera, R.; et al. DDR1 regulates thyroid cancer cell differentiation via IGF-2/IR-A autocrine signaling loop. *Endocr. Relat. Cancer* **2019**, *26*, 197–214. [[CrossRef](#)]
67. Chen, H.; Ghori-Javed, F.Y.; Rashid, H.; Adhami, M.D.; Serra, R.; Gutierrez, S.E.; Javed, A. Runx2 regulates endochondral ossification through control of chondrocyte proliferation and differentiation. *J. Bone Miner. Res.* **2014**, *29*, 2653–2665. [[CrossRef](#)]
68. Sancisi, V.; Boretini, G.; Maramotti, S.; Ragazzi, M.; Tamagnini, I.; Nicoli, D.; Piana, S.; Ciarrocchi, A. Runx2 isoform I controls a panel of proinvasive genes driving aggressiveness of papillary thyroid carcinomas. *J. Clin. Endocrinol. Metab.* **2012**, *97*, E2006–E2015. [[CrossRef](#)]
69. Cohen-Solal, K.A.; Boregowda, R.K.; Lasfar, A. RUNX2 and the PI3K/AKT axis reciprocal activation as a driving force for tumor progression. *Mol. Cancer* **2015**, *14*, 137. [[CrossRef](#)]
70. Wang, W.; Shen, T.; Dong, B.; Creighton, C.J.; Meng, Y.; Zhou, W.; Shi, Q.; Zhou, H.; Zhang, Y.; Moore, D.D.; et al. MAPK4 overexpression promotes tumor progression via noncanonical activation of AKT/mTOR signaling. *J. Clin. Investig.* **2019**, *129*, 1015–1029. [[CrossRef](#)]
71. de Souza, E.C.; Padron, A.S.; Braga, W.M.; de Andrade, B.M.; Vaisman, M.; Nasciutti, L.E.; Ferreira, A.C.; de Carvalho, D.P. MTOR downregulates iodide uptake in thyrocytes. *J. Endocrinol.* **2010**, *206*, 113–120. [[CrossRef](#)] [[PubMed](#)]
72. Azouzi, N.; Cailloux, J.; Cazarin, J.M.; Knauf, J.A.; Cracchiolo, J.; Al Ghuzlan, A.; Hartl, D.; Polak, M.; Carré, A.; El Mzibri, M.; et al. NADPH Oxidase NOX4 Is a Critical Mediator of BRAFV600E-Induced Downregulation of the Sodium/Iodide Symporter in Papillary Thyroid Carcinomas. *Antioxid. Redox Signal.* **2017**, *26*, 864–877. [[CrossRef](#)] [[PubMed](#)]
73. Weyemi, U.; Caillou, B.; Talbot, M.; Ameziane-El-Hassani, R.; Lacroix, L.; Lagent-Chevallier, O.; Al Ghuzlan, A.; Roos, D.; Bidart, J.M.; Virion, A.; et al. Intracellular expression of reactive oxygen species-generating NADPH oxidase NOX4 in normal and cancer thyroid tissues. *Endocr. Relat. Cancer* **2010**, *17*, 27–37. [[CrossRef](#)]
74. Liu, J.; Xiao, Q.; Xiao, J.; Niu, C.; Li, Y.; Zhang, X.; Zhou, Z.; Shu, G.; Yin, G. Wnt/ β -catenin signalling: Function, biological mechanisms, and therapeutic opportunities. *Signal Transduct. Target. Ther.* **2022**, *7*, 3. [[CrossRef](#)]
75. Sastre-Perona, A.; Santisteban, P. Role of the wnt pathway in thyroid cancer. *Front. Endocrinol.* **2012**, *3*, 31. [[CrossRef](#)]
76. Zhang, W.; Ruan, X.; Li, Y.; Zhi, J.; Hu, L.; Hou, X.; Shi, X.; Wang, X.; Wang, J.; Ma, W.; et al. KDM1A promotes thyroid cancer progression and maintains stemness through the Wnt/ β -catenin signaling pathway. *Theranostics* **2022**, *12*, 1500–1517. [[CrossRef](#)] [[PubMed](#)]
77. Antonelli, A.; La Motta, C. Novel therapeutic clues in thyroid carcinomas: The role of targeting cancer stem cells. *Med. Res. Rev.* **2017**, *37*, 1299–1317. [[CrossRef](#)]
78. Zou, M.; BinEssa, H.A.; Al-Malki, Y.H.; Al-Yahya, S.; Al-Alwan, M.; Al-Jammaz, I.; Khabar, K.S.; Almohanna, F.; Assiri, A.M.; Meyer, B.F.; et al. β -Catenin Attenuation Inhibits Tumor Growth and Promotes Differentiation in a BRAFV600E-Driven Thyroid Cancer Animal Model. *Mol. Cancer Ther.* **2021**, *20*, 1603–1613. [[CrossRef](#)]
79. Leow, P.C.; Tian, Q.; Ong, Z.Y.; Yang, Z.; Ee, P.L. Antitumor activity of natural compounds, curcumin and PKF118-310, as Wnt/ β -catenin antagonists against human osteosarcoma cells. *Investig. New Drugs* **2010**, *28*, 766–782. [[CrossRef](#)]

80. Lan, L.; Basourakos, S.; Cui, D.; Zuo, X.; Deng, W.; Huo, L.; Chen, L.; Zhang, G.; Deng, L.; Shi, B.; et al. Inhibiting β -catenin expression promotes efficiency of radioiodine treatment in aggressive follicular thyroid cancer cells probably through mediating NIS localization. *Oncol. Rep.* **2017**, *37*, 426–434. [[CrossRef](#)]
81. Ferretti, E.; Tosi, E.; Po, A.; Scipioni, A.; Morisi, R.; Espinola, M.S.; Russo, D.; Durante, C.; Schlumberger, M.; Screpanti, I.; et al. Notch signaling is involved in expression of thyrocyte differentiation markers and is down-regulated in thyroid tumors. *J. Clin. Endocrinol. Metab.* **2008**, *93*, 4080–4087. [[CrossRef](#)] [[PubMed](#)]
82. Bolós, V.; Grego-Bessa, J.; de la Pompa, J.L. Notch signaling in development and cancer. *Endocr. Rev.* **2007**, *28*, 339–363. [[CrossRef](#)] [[PubMed](#)]
83. Miele, L. Notch signaling. *Clin. Cancer Res.* **2006**, *12*, 1074–1079. [[CrossRef](#)] [[PubMed](#)]
84. Somnay, Y.R.; Yu, X.M.; Lloyd, R.V.; Levenson, G.; Aburjania, Z.; Jang, S.; Jaskula-Sztul, R.; Chen, H. Notch3 expression correlates with thyroid cancer differentiation, induces apoptosis, and predicts disease prognosis. *Cancer* **2017**, *123*, 769–782. [[CrossRef](#)] [[PubMed](#)]
85. Talora, C.; Sgroi, D.C.; Crum, C.P.; Dotto, G.P. Specific down-modulation of Notch1 signaling in cervical cancer cells is required for sustained HPV-E6/E7 expression and late steps of malignant transformation. *Genes Dev.* **2002**, *16*, 2252–2263. [[CrossRef](#)] [[PubMed](#)]
86. Xiao, X.; Ning, L.; Chen, H. Notch1 mediates growth suppression of papillary and follicular thyroid cancer cells by histone deacetylase inhibitors. *Mol. Cancer Ther.* **2009**, *8*, 350–356. [[CrossRef](#)]
87. Zhang, X.; Li, D.; Li, M.; Ye, M.; Ding, L.; Cai, H.; Fu, D.; Lv, Z. MicroRNA-146a targets PRKCE to modulate papillary thyroid tumor development. *Int. J. Cancer* **2014**, *134*, 257–267. [[CrossRef](#)]
88. Yang, S.J.; Wang, D.D.; Zhong, S.L.; Chen, W.Q.; Wang, F.L.; Zhang, J.; Xu, W.X.; Xu, D.; Zhang, Q.; Li, J.; et al. Tumor-derived exosomal circPSMA1 facilitates the tumorigenesis, metastasis, and migration in triple-negative breast cancer (TNBC) through miR-637/Akt1/ β -catenin (cyclin D1) axis. *Cell Death Dis.* **2021**, *12*, 420. [[CrossRef](#)]
89. Lakshmanan, A.; Wojcicka, A.; Kotlarek, M.; Zhang, X.; Jazdzewski, K.; Jhiang, S.M. microRNA-339-5p modulates Na⁺/I⁻ symporter-mediated radioiodide uptake. *Endocr. Relat. Cancer* **2015**, *22*, 11–21. [[CrossRef](#)]
90. Montero-Conde, C.; Graña-Castro, O.; Martín-Serrano, G.; Martínez-Montes, Á.M.; Zarzuela, E.; Muñoz, J.; Torres-Perez, R.; Pita, G.; Cordero-Barreal, A.; Leandro-García, L.J.; et al. Hsa-miR-139-5p is a prognostic thyroid cancer marker involved in HNRNPF-mediated alternative splicing. *Int. J. Cancer* **2020**, *146*, 521–530. [[CrossRef](#)]
91. Riesco-Eizaguirre, G.; Wert-Lamas, L.; Perales-Patón, J.; Sastre-Perona, A.; Fernández, L.P.; Santisteban, P. The miR-146b-3p/PAX8/NIS Regulatory Circuit Modulates the Differentiation Phenotype and Function of Thyroid Cells during Carcinogenesis. *Cancer Res.* **2015**, *75*, 4119–4130. [[CrossRef](#)] [[PubMed](#)]
92. Hou, S.; Xie, X.; Zhao, J.; Wu, C.; Li, N.; Meng, Z.; Cai, C.; Tan, J. Downregulation of miR-146b-3p Inhibits Proliferation and Migration and Modulates the Expression and Location of Sodium/Iodide Symporter in Dedifferentiated Thyroid Cancer by Potentially Targeting MUC20. *Front. Oncol.* **2021**, *10*, 566365. [[CrossRef](#)] [[PubMed](#)]
93. Shen, C.T.; Qiu, Z.L.; Song, H.J.; Wei, W.J.; Luo, Q.Y. miRNA-106a directly targeting RARB associates with the expression of Na(+)/I(-) symporter in thyroid cancer by regulating MAPK signaling pathway. *J. Exp. Clin. Cancer Res.* **2016**, *35*, 101. [[CrossRef](#)] [[PubMed](#)]
94. Ricarte-Filho, J.C.; Fuziwara, C.S.; Yamashita, A.S.; Rezende, E.; da-Silva, M.J.; Kimura, E.T. Effects of let-7 microRNA on Cell Growth and Differentiation of Papillary Thyroid Cancer. *Transl. Oncol.* **2009**, *2*, 236–241. [[CrossRef](#)]
95. Oh, J.M.; Ahn, B.C. Molecular mechanisms of radioactive iodine refractoriness in differentiated thyroid cancer: Impaired sodium iodide symporter (NIS) expression owing to altered signaling pathway activity and intracellular localization of NIS. *Theranostics* **2021**, *11*, 6251–6277. [[CrossRef](#)] [[PubMed](#)]
96. Mu, Z.; Zhang, X.; Liang, D.; Fang, J.; Chen, G.; Guo, W.; Sun, D.; Sun, Y.; Kai, Z.; Huang, L.; et al. Risk stratification for radioactive iodine refractoriness using molecular alterations in distant metastatic differentiated thyroid cancer. *Chin. J. Cancer Res.* **2024**, *36*, 25–35. [[CrossRef](#)]
97. Haugen, B.R.; Alexander, E.K.; Bible, K.C.; Doherty, G.M.; Mandel, S.J.; Nikiforov, Y.E.; Pacini, F.; Randolph, G.W.; Sawka, A.M.; Schlumberger, M.; et al. 2015 American Thyroid Association Management Guidelines for Adult Patients with Thyroid Nodules and Differentiated Thyroid Cancer: The American Thyroid Association Guidelines Task Force on Thyroid Nodules and Differentiated Thyroid Cancer. *Thyroid* **2016**, *26*, 1–133. [[CrossRef](#)] [[PubMed](#)]
98. Filetti, S.; Durante, C.; Hartl, D.M.; Leboulleux, S.; Locati, L.D.; Newbold, K.; Papotti, M.G.; Berruti, A.; ESMO Guidelines Committee. ESMO Clinical Practice Guideline update on the use of systemic therapy in advanced thyroid cancer. *Ann. Oncol.* **2022**, *33*, 674–684. [[CrossRef](#)]
99. Schlumberger, M.; Brose, M.; Elisei, R.; Leboulleux, S.; Luster, M.; Pitoia, F.; Pacini, F. Definition and management of radioactive iodine-refractory differentiated thyroid cancer. *Lancet Diabetes Endocrinol.* **2014**, *2*, 356–358. [[CrossRef](#)]
100. Silaghi, H.; Lozovanu, V.; Georgescu, C.E.; Pop, C.; Nasui, B.A.; Cătoi, A.F.; Silaghi, C.A. State of the Art in the Current Management and Future Directions of Targeted Therapy for Differentiated Thyroid Cancer. *Int. J. Mol. Sci.* **2022**, *23*, 3470. [[CrossRef](#)]
101. Nistor, C.; Ciuche, A.; Constantinescu, I. Emergency surgical tracheal decompression in a huge retrosternal goiter. *Acta Endocrinol.* **2017**, *13*, 370–374. [[CrossRef](#)]

102. Kiss, A.; Szili, B.; Bakos, B.; Ármós, R.; Putz, Z.; Árvai, K.; Kocsis-Deák, B.; Tobiás, B.; Balla, B.; Pikó, H.; et al. Comparison of surgical strategies in the treatment of low-risk differentiated thyroid cancer. *BMC Endocr. Disord.* **2023**, *23*, 23. [[CrossRef](#)] [[PubMed](#)]
103. Nistor, C.E.; Găvan, C.S.; Cirtel, A.A.; Nemes, A.F.; Ciuche, A. The Association of Minimally Invasive Surgical Approaches and Mortality in Patients with Malignant Pleuropericarditis—A 10 Year Retrospective Observational Study. *Medicina* **2022**, *58*, 718. [[CrossRef](#)] [[PubMed](#)]
104. Young, M.; John, S. *Hepatic Chemoembolization*. StatPearls [Internet]; StatPearls Publishing: Treasure Island, FL, USA, 2018.
105. Minocha, J.; Salem, R.; Lewandowski, R.J. Transarterial chemoembolization and yttrium-90 for liver cancer and other lesions. *Clin. Liver Dis.* **2014**, *18*, 877–890. [[CrossRef](#)] [[PubMed](#)]
106. Choy, P.Y.; Koea, J.; McCall, J.; Holden, A.; Osbourne, M. The role of radiofrequency ablation in the treatment of primary and metastatic tumours of the liver: Initial lessons learned. *N. Z. Med. J.* **2002**, *115*, 1–7.
107. Mazzeo, S.; Cervelli, R.; Elisei, R.; Tarantini, G.; Cappelli, C.; Molinaro, E.; Galleri, D.; De Napoli, L.; Comite, C.; Cioni, R.; et al. mRECIST criteria to assess recurrent thyroid carcinoma treatment response after radiofrequency ablation: A prospective study. *J. Endocrinol. Investig.* **2018**, *41*, 1389–1399. [[CrossRef](#)]
108. Hay, I.D.; Lee, R.A.; Davidge-Pitts, C.; Reading, C.C.; Charboneau, J.W. Long-term outcome of ultrasound-guided percutaneous ethanol ablation of selected “recurrent” neck nodal metastases in 25 patients with TNM stages III or IVA papillary thyroid carcinoma previously treated by surgery and 131I therapy. *Surgery* **2013**, *154*, 1448–1455. [[CrossRef](#)] [[PubMed](#)]
109. Deschamps, F.; de Baere, T. Cementoplasty of bone metastases. *Diagn. Interv. Imaging* **2012**, *93*, 685–689. [[CrossRef](#)]
110. Karapanou, O.; Simeakis, G.; Vlassopoulou, B.; Alevizaki, M.; Saltiki, K. Advanced RAI-refractory thyroid cancer: An update on treatment perspectives. *Endocr. Relat. Cancer* **2022**, *29*, R57–R66. [[CrossRef](#)]
111. Brose, M.S.; Nutting, C.M.; Jarzab, B.; Elisei, R.; Siena, S.; Bastholt, L.; De La Fouchardiere, C.; Pacini, F.; Paschke, R.; Shong, Y.K.; et al. Sorafenib in radioactive iodine-refractory, locally advanced or metastatic differentiated thyroid cancer: A randomised, double-blind, phase 3 trial. *Lancet* **2014**, *384*, 319–328. [[CrossRef](#)]
112. Schlumberger, M.; Tahara, M.; Wirth, L.J.; Robinson, B.; Brose, M.S.; Elisei, R.; Habra, M.A.; Newbold, K.; Shah, M.H.; Hoff, A.O.; et al. Lenvatinib versus placebo in radioiodine-refractory thyroid cancer. *N. Engl. J. Med.* **2015**, *372*, 621–630. [[CrossRef](#)] [[PubMed](#)]
113. Brose, M.S.; Worden, F.P.; Newbold, K.L.; Guo, M.; Hurria, A. Effect of Age on the Efficacy and Safety of Lenvatinib in Radioiodine-Refractory Differentiated Thyroid Cancer in the Phase III SELECT Trial. *J. Clin. Oncol.* **2017**, *35*, 2692–2699. [[CrossRef](#)] [[PubMed](#)]
114. Zheng, X.; Xu, Z.; Ji, Q.; Ge, M.; Shi, F.; Qin, J.; Wang, F.; Chen, G.; Zhang, Y.; Huang, R.; et al. A Randomized, Phase III Study of Lenvatinib in Chinese Patients with Radioiodine-Refractory Differentiated Thyroid Cancer. *Clin. Cancer Res.* **2021**, *27*, 5502–5509. [[CrossRef](#)] [[PubMed](#)]
115. Wirth, L.J.; Brose, M.S.; Sherman, E.J.; Licitra, L.; Schlumberger, M.; Sherman, S.I.; Bible, K.C.; Robinson, B.; Rodien, P.; Godbert, Y.; et al. Open-Label, Single-Arm, Multicenter, Phase II Trial of Lenvatinib for the Treatment of Patients with Anaplastic Thyroid Cancer. *J. Clin. Oncol.* **2021**, *39*, 2359–2366. [[CrossRef](#)]
116. Tahara, M.; Kiyota, N.; Hoff, A.O.; Badiu, C.; Owonikoko, T.K.; Dutcus, C.E.; Suzuki, T.; Ren, M.; Wirth, L.J. Impact of lung metastases on overall survival in the phase 3 SELECT study of lenvatinib in patients with radioiodine-refractory differentiated thyroid cancer. *Eur. J. Cancer* **2021**, *147*, 51–57. [[CrossRef](#)]
117. Tahara, M.; Brose, M.S.; Wirth, L.J.; Suzuki, T.; Miyagishi, H.; Fujino, K.; Dutcus, C.E.; Gianoukakis, A. Impact of dose interruption on the efficacy of lenvatinib in a phase 3 study in patients with radioiodine-refractory differentiated thyroid cancer. *Eur. J. Cancer* **2019**, *106*, 61–68. [[CrossRef](#)]
118. Gild, M.L.; Tsang, V.H.M.; Clifton-Bligh, R.J.; Robinson, B.G. Multikinase inhibitors in thyroid cancer: Timing of targeted therapy. *Nat. Rev. Endocrinol.* **2021**, *17*, 225–234. [[CrossRef](#)]
119. Cabanillas, M.E.; Brose, M.S.; Holland, J.; Ferguson, K.C.; Sherman, S.I. A phase I study of cabozantinib (XL184) in patients with differentiated thyroid cancer. *Thyroid* **2014**, *24*, 1508–1514. [[CrossRef](#)]
120. Cabanillas, M.E.; De Souza, J.A.; Geyer, S.; Wirth, L.J.; Menefee, M.E.; Liu, S.V.; Shah, K.; Wright, J.; Shah, M.H. Cabozantinib as Salvage Therapy for Patients with Tyrosine Kinase Inhibitor-Refractory Differentiated Thyroid Cancer: Results of a Multicenter Phase II International Thyroid Oncology Group Trial. *J. Clin. Oncol.* **2017**, *35*, 3315–3321. [[CrossRef](#)]
121. Brose, M.S.; Robinson, B.; Sherman, S.I.; Krajewska, J.; Lin, C.C.; Vaisman, F.; Hoff, A.O.; Hitre, E.; Bowles, D.W.; Hernando, J.; et al. Cabozantinib for radioiodine-refractory differentiated thyroid cancer (COSMIC-311): A randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet Oncol.* **2021**, *22*, 1126–1138. [[CrossRef](#)]
122. Duke, E.S.; Barone, A.K.; Chatterjee, S.; Mishra-Kalyani, P.S.; Shen, Y.L.; Isikwei, E.; Zhao, H.; Bi, Y.; Liu, J.; Rahman, N.A.; et al. FDA Approval Summary: Cabozantinib for Differentiated Thyroid Cancer. *Clin. Cancer Res.* **2022**, *28*, 4173–4177. [[CrossRef](#)] [[PubMed](#)]
123. Leboulleux, S.; Bastholt, L.; Krause, T.; de la Fouchardiere, C.; Tennvall, J.; Awada, A.; Gómez, J.M.; Bonichon, F.; Leenhardt, L.; Soufflet, C.; et al. Vandetanib in locally advanced or metastatic differentiated thyroid cancer: A randomised, double-blind, phase 2 trial. *Lancet Oncol.* **2012**, *13*, 897–905. [[CrossRef](#)] [[PubMed](#)]
124. Lorusso, L.; Pieruzzi, L.; Biagini, A.; Sabini, E.; Valerio, L.; Giani, C.; Passannanti, P.; Pontillo-Contillo, B.; Battaglia, V.; Mazzeo, S.; et al. Lenvatinib and other tyrosine kinase inhibitors for the treatment of radioiodine refractory, advanced, and progressive thyroid cancer. *Oncotargets Ther.* **2016**, *9*, 6467–6477. [[CrossRef](#)] [[PubMed](#)]

125. Dadu, R.; Devine, C.; Hernandez, M.; Waguespack, S.G.; Busaidy, N.L.; Hu, M.I.; Jimenez, C.; Habra, M.A.; Sellin, R.V.; Ying, A.K.; et al. Role of salvage targeted therapy in differentiated thyroid cancer patients who failed first-line sorafenib. *J. Clin. Endocrinol. Metab.* **2014**, *99*, 2086–2094. [[CrossRef](#)]
126. Lamartina, L.; Anizan, N.; Dupuy, C.; Leboulleux, S.; Schlumberger, M. Redifferentiation-facilitated radioiodine therapy in thyroid cancer. *Endocr. Relat. Cancer* **2021**, *28*, T179–T191. [[CrossRef](#)]
127. Ho, A.L.; Grewal, R.K.; Leboeuf, R.; Sherman, E.J.; Pfister, D.G.; Deandreis, D.; Pentlow, K.S.; Zanzonico, P.B.; Haque, S.; Gavane, S.; et al. Selumetinib-enhanced radioiodine uptake in advanced thyroid cancer. *N. Engl. J. Med.* **2013**, *368*, 623–632. [[CrossRef](#)]
128. Rothenberg, S.M.; McFadden, D.G.; Palmer, E.L.; Daniels, G.H.; Wirth, L.J. Redifferentiation of iodine-refractory BRAF V600E-mutant metastatic papillary thyroid cancer with dabrafenib. *Clin. Cancer Res.* **2015**, *21*, 1028–1035. [[CrossRef](#)]
129. Dunn, L.A.; Sherman, E.J.; Baxi, S.S.; Tchekmedyian, V.; Grewal, R.K.; Larson, S.M.; Pentlow, K.S.; Haque, S.; Tuttle, R.M.; Sabra, M.M.; et al. Vemurafenib Redifferentiation of BRAF Mutant, RAI-Refractory Thyroid Cancers. *J. Clin. Endocrinol. Metab.* **2019**, *104*, 1417–1428. [[CrossRef](#)]
130. Pešorda, M.; Kuna, S.K.; Huić, D.; Herceg, D.; Despot, M.; Samardžić, T.; Gnjidić, M.; Belev, B. Kinase Inhibitors in the Treatment of Thyroid Cancer: Institutional Experience. *Acta Clin. Croat.* **2020**, *59* (Suppl. S1), 73–80. [[CrossRef](#)]
131. Jaber, T.; Waguespack, S.G.; Cabanillas, M.E.; Elbanan, M.; Vu, T.; Dadu, R.; Sherman, S.I.; Amit, M.; Santos, E.B.; Zafereo, M.; et al. Targeted Therapy in Advanced Thyroid Cancer to Resensitize Tumors to Radioactive Iodine. *J. Clin. Endocrinol. Metab.* **2018**, *103*, 3698–3705. [[CrossRef](#)]
132. Iravani, A.; Solomon, B.; Pattison, D.A.; Jackson, P.; Ravi Kumar, A.; Kong, G.; Hofman, M.S.; Akhurst, T.; Hicks, R.J. Mitogen-Activated Protein Kinase Pathway Inhibition for Redifferentiation of Radioiodine Refractory Differentiated Thyroid Cancer: An Evolving Protocol. *Thyroid* **2019**, *29*, 1634–1645. [[CrossRef](#)] [[PubMed](#)]
133. Leboulleux, S.; Dupuy, C.; Lacroix, L.; Attard, M.; Grimaldi, S.; Corre, R.; Ricard, M.; Nasr, S.; Berdelou, A.; Hadoux, J.; et al. Redifferentiation of a BRAFK601E-Mutated Poorly Differentiated Thyroid Cancer Patient with Dabrafenib and Trametinib Treatment. *Thyroid* **2019**, *29*, 735–742. [[CrossRef](#)] [[PubMed](#)]
134. Weber, M.; Kersting, D.; Riemann, B.; Brandenburg, T.; Führer-Sakel, D.; Grünwald, F.; Kreissl, M.C.; Dralle, H.; Weber, F.; Schmid, K.W.; et al. Enhancing Radioiodine Incorporation into Radio Iodine Refractory Thyroid Cancer with MAPK Inhibition (ERRITI): A Single-Center Prospective Two-Arm Study. *Clin. Cancer Res.* **2022**, *28*, 4194–4202. [[CrossRef](#)]
135. Girolami, I.; Pantanowitz, L.; Mete, O.; Brunelli, M.; Marletta, S.; Colato, C.; Trimboli, P.; Crescenzi, A.; Bongiovanni, M.; Barbareschi, M.; et al. Programmed death-ligand 1 (PD-L1) is a potential biomarker of disease-free survival in papillary thyroid carcinoma: A systematic review and meta-analysis of PD-L1 immunoexpression in follicular epithelial derived thyroid carcinoma. *Endocr. Pathol.* **2020**, *31*, 291–300. [[CrossRef](#)]
136. Shi, X.; Li, C.W.; Tan, L.C.; Wen, S.S.; Liao, T.; Zhang, Y.; Chen, T.Z.; Ma, B.; Yu, P.C.; Lu, Z.W.; et al. Immune co-inhibitory receptors PD-1, CTLA-4, TIM-3, LAG-3 and TIGIT in medullary thyroid cancers: A large cohort study. *J. Clin. Endocrinol. Metab.* **2020**, *106*, 120–132. [[CrossRef](#)]
137. Chintakuntlawar, A.V.; Rumilla, K.M.; Smith, C.Y.; Jenkins, S.M.; Foote, R.L.; Kasperbauer, J.L.; Morris, J.C.; Ryder, M.; Alsidawi, S.; Hilger, C.; et al. Expression of PD-1 and PD-L1 in anaplastic thyroid cancer patients treated with multimodal therapy: Results from a retrospective study. *J. Clin. Endocrinol. Metab.* **2017**, *102*, 1943–1950. [[CrossRef](#)]
138. Mehnert, J.M.; Varga, A.; Brose, M.S.; Aggarwal, R.R.; Lin, C.C.; Prawira, A.; De Braud, F.; Tamura, K.; Doi, T.; Piha-Paul, S.A.; et al. Safety and antitumor activity of the anti-PD-1 antibody pembrolizumab in patients with advanced, PD-L1-positive papillary or follicular thyroid cancer. *BMC Cancer* **2019**, *19*, 196. [[CrossRef](#)] [[PubMed](#)]
139. Naing, A.; Gainor, J.F.; Gelderblom, H.; Forde, P.M.; Butler, M.O.; Lin, C.C.; Sharma, S.; De Olza, M.O.; Varga, A.; Taylor, M.; et al. A first-in-human phase 1 dose escalation study of spartalizumab (PDR001), an anti-PD-1 antibody, in patients with advanced solid tumors. *J. Immunother. Cancer* **2020**, *8*, e000530. [[CrossRef](#)] [[PubMed](#)]
140. Capdevila, J.; Wirth, L.J.; Ernst, T.; Ponce Aix, S.; Lin, C.C.; Ramlau, R.; Butler, M.O.; Delord, J.P.; Gelderblom, H.; Ascierto, P.A.; et al. PD-1 blockade in anaplastic thyroid carcinoma. *J. Clin. Oncol. Off. J. Am. Soc. Clin. Oncol.* **2020**, *38*, 2620–2627. [[CrossRef](#)]
141. Chintakuntlawar, A.V.; Yin, J.; Foote, R.L.; Kasperbauer, J.L.; Rivera, M.; Asmus, E.; Garces, N.I.; Janus, J.R.; Liu, M.; Ma, D.J.; et al. A phase 2 study of pembrolizumab combined with chemoradiotherapy as initial treatment for anaplastic thyroid cancer. *Thyroid Off. J. Am. Thyroid Assoc.* **2019**, *29*, 1615–1622. [[CrossRef](#)]
142. Dierks, C.; Ruf, J.; Seufert, J.; Kreissl, M.; Klein, C.; Spitzweg, C.; Kroiss, M.; Thomusch, O.; Lorenz, K.; Zielke, A.; et al. 1646MO Phase II ATLEP trial: Final results for lenvatinib/pembrolizumab in metastasized anaplastic and poorly differentiated thyroid carcinoma. *Ann. Oncol.* **2022**, *33*, S1295. [[CrossRef](#)]
143. French, J.D.; Haugen, B.R.; Worden, F.P.; Bowles, D.W.; Gianoukakis, A.G.; Konda, B.; Dadu, R.; Sherman, E.J.; McCue, S.; Foster, N.R.; et al. Combination Targeted Therapy with Pembrolizumab and Lenvatinib in Progressive, Radioiodine-Refractory Differentiated Thyroid Cancers. *Clin. Cancer Res.* **2024**, *30*, 3757–3767. [[CrossRef](#)] [[PubMed](#)]

144. Volpe, D.F.; Nappi, C.; Zampella, E.; Di Donna, E.; Maurea, S.; Cuocolo, A.; Klain, M. Current Advances in Radioactive Iodine-Refractory Differentiated Thyroid Cancer. *Curr. Oncol.* **2024**, *31*, 3870–3884. [[CrossRef](#)]
145. Jin, Y.; Van Nostrand, D.; Cheng, L.; Liu, M.; Chen, L. Radioiodine refractory differentiated thyroid cancer. *Crit. Rev. Oncol. Hematol.* **2018**, *125*, 111–120. [[CrossRef](#)] [[PubMed](#)]

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.