Table S1. 3D-printed scaffolds for periodontal tissue engineering.

	Authors	3D printing technology	Printed material(s), bioactive factor(s) and / or cells used	Scaffold geometry	Mesh thickness	Porosity
MONOPHASIC	Kim [128]	Extrusion	PCL (80%) + HA (20%) with SDF1 and BMP-7 (100 ng/mL each) Shape of tooth (rat mandibular central incisor and human mandibular first molar)		200 μm	200 μm diameter interconnecting channels
	Mangano [129]	Extrusion	30% HA, 60% β-TCP, 10% α-TCP	Blocks with a volume of about 0.14 cm ³	300 ± 30 μm	$370 \pm 25 \ \mu m$ Open interconnecting microporosity with pores sizes of about 1 μm Total porosity: 60%
	Baba [130, 131]	Melt spinning	75/25 PLGA+BMMSCs + PRP	1.5 diameter -		-
	Carrel [132]	Extrusion	α -TCP + HA (calcium-to-phosphate ratio : 1.43)		400 μm	250 µm Total porosity: 50–659 Macroporosity: 40– 50%
	Cho [133]	Extrusion	PCL scaffolds with microspheres of 75/25 PLGA with BMP-2, BMP-7 + CTGF	Rectangular scaffolds (5×3×0.5 mm³)	200 μm	500 μm transverse pores
	Puppi [134]	Extrusion (wet spinning)	PCL fiber construct and a chitosan/poly(γ-glutamic acid) polyelectrolyte complex hydrogel	Rectangular scaffold: 10 × 10 mm, 50 layers	200–300 μm	200–1800 μm
BIPHASIC	Park [135]	3D wax printing system -	Bone compartment : 25% PCL with recombinantadenovirus- encodingmurine BMP-7	Rectangular scaffolds: $1.75 \times 5.0 \times 4.0 \text{ mm}$	-	Windows dimensions 0.75 × 0.50 × 0.05 mm ³
			PDL compartment: 25% PGA	Rectangular scaffolds: $1.5 \times 5.0 \times 4.0 \text{ mm}$	Diameter: 0.8 mm Height: 0.3 mm	-
	Park [136]	3D wax printing system	Bone compartment: PCL + BMP-7	3 × 2 × 2 mm	-	0.60 × 0.50 mm ² window pores to contact residual bone tissue

			PDL compartment: PCL+PDL cells	fiber-g architectur	DL guiding es per layer diameter,	-	0.175 mm thick interconnective space between PDL and	
	Vaquette [84, 137] Dan [138] Mathew [139] Vaquette [89]]			0.250 mm inte	n ligament rface		bone regions
			electr	hate coated melt ospun (CaP-PCL) scaffold	(approxim × 1.5 mm >	pieces ately 3 mm < 0.5 mm in ze)	500 μm	over 150 µm diameter interconnecting pores
		Electrospinning		associated) (can be azithromycin)	Small pieces (approximately 8 mm × 5 mm × 0.3 mm in size)		300–400 μm with 3 μm diameter fibers	5–10 μm
		Extrusion (FDM)		: PCL + b-tricalcium TCP, 20% wt.)	$100 \times 100 \times 2$ mm ³ sectioned into 5×2 mm ³		-	interconnectivity, 70% porosity
		Electrospinning	PDL compa	rtment: PCL	cell membrane: $7 \times 9 \times 0.4 \text{ mm}^3$		-	-
		FDM	•	:: PCL + β-TCP (20% rt.)	$100 \times 100 \times 2$ mm ³ sectioned into 5 × 5×2 mm ³		-	interconnectivity, 70% porosity
	Costa [90]	Electrospinning Extrusion (layer-	PDL compartment: PCL		membrai	cular nes with 8 iameter	Fiber diameter: 10–15 μm	Interconnected pore Pore size: 100–400 μm
			PCL/HA scaffolds and poly(lactic-	Bone compartment: +BMP2	:	2.25 mm (width)	-	300 mm microchannels
TRIPHASIC			co-glycolic acid) microspheres encapsulating factors	PDL compartment: + CTGF	5 × 5× 3 mm ³	0.5 mm (width)	-	600 mm transverse microchannels
				Cementum/denti n interface: +amelogenin		2.25 mm (width)	-	100 mm transverse microchannels
CUSTOM	Rasperini [141]	SLS	PCL and	d 4% HA	Customized scaffold: patient's defect		Mean strut length: 600 μm for support of extensions in the PDL region	Channel width for PDGF delivery was ≈ 500 μm

	Jiang [142]	Electrospinning	Biodegradable poly PCE copolymer electrospun nanofibrous mats into porous CHI	30 layers of nanofibers embedded within the porous CHI, with thickness ≈ 4 mm	Diameter (nm): - Random fibers: 574.3±218.2 - Aligned fibers: 616.1±213.1 nm	103.38 ± 49.54 μm between layers
ORIENTE	Kim [143]	Electrospinning	PCL/gelatin (Gel) nanofiber with periodontal ligament cells	30 mm × 4 mm × 200 μm	-	-
	Pilipchuk [144]	SLS	Bone compartment: PCL and 4% HA	Idem [130]: 5.1 × 4.1 × 3.2 mm	≈ 0.7 × 0.7 mm	0.7 × 0.7 mm
		Molding	Periodontal ligament: PCL	Idem [130]: 3.6 × 2.8 × 0.4 mm	400 × 250 μm, with/ without grooves	400 μm wide pores

BMMSCs: mesenchymal cells derived from bone marrow, BMP: Bone morphogenetic protein, CHI: chitosan, CTGF: Connective tissue growth factor, FDM: Fused Deposition Modeling, HA: hydroxyapatite, SLS: selective laser sintering, PCE: poly (e-caprolactone)-poly(ethylene glycol), PCL: polycaprolactone, PDGF: Platelet-derived growth factor, PDL: periodontal ligament, PGA: Polyglycolic acid, PLGA: poly-L-lactic acid, PRP: Platelet Rich Plasma, SDF: Stromal-derived factor, SLS: Selective Laser Sintering, TCP: tricalcium phosphate.