

Review

Automatic Detection and Monitoring of Insect Pests—A Review

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Abstract: Many species of insect pests can be detected and monitored automatically. Several systems have been designed in order to improve integrated pest management (IPM) in the context of precision agriculture. Automatic detection traps have been developed for many important pests. These techniques and new technologies are very promising for the early detection and monitoring of aggressive and quarantine pests. The aim of the present paper is to review the techniques and scientific state of the art of the use of sensors for automatic detection and monitoring of insect pests. The paper focuses on the methods for identification of pests based in infrared sensors, audio sensors and image-based classification, presenting the different systems available, examples of applications and recent developments, including machine learning and Internet of Things. Future trends of automatic traps and decision support systems are also discussed.

Keywords: automatic traps; sensors; integrated pest management

1. Introduction

Environmental concerns about the use of pesticides, overexploitation of natural resources, expansion of global trade, increasing human population, changes in consumption patterns, and advances in technology are thriving, leading to a new revolution in agriculture. This revolution consists of the use of digital tools to increase productivity, optimizing the management of natural resources and agricultural inputs concurrently [1]. Precision agriculture tools enable farmers to analyze the spatial-temporal variability of several key factors that affect plant health and productivity. Obtained through sensors, these data are stored and combined in digital platforms in order to guide the decision-making process [2]. Integrated pest management (IPM) systems are being developed in order to improve the management of insect pests, reducing the overall use of pesticides and focusing on more precise applications. However, the efficiency of these systems depends on the accuracy of the chosen pest population monitoring method. In addition, it is essential to gather information of population dynamics and their associated ecological factors in order to develop an appropriate pest control strategy [3]. Recently, modern technologies started to be applied in field surveys of several pests, such as radar technologies monitoring pest migration, video equipment to observe flying insects, thermal infrared imaging and chemiluminescent tags for tracking insect movement in

the darkness, Global Navigation Satellite System (GNSS) for wildlife telemetry, habitat mapping and echo-sounding detection of larvae movement [3]. Nevertheless, these techniques are usually high-cost and not affordable for farmers. More recently, advances in miniaturized sensors, microprocessors, telecommunications engineering and digital processing techniques allowed the reduction in costs in novel insect automatic detection and monitoring systems. These new devices can be easily connected to the internet, allowing real-time surveillance on the field level. Some of these devices can be connected to wireless sensors networks (Internet of Things) for monitoring field areas and/or use cloud-computing services to help in the decision-making process. This paper covers an in-depth review on recent publications in the field of insect automatic detection and monitoring. Furthermore, this review intends to highlight the methods used for different orders of insects, including Lepidoptera, Diptera (fruit flies), Coleoptera (weevil in palm trees) and sucking insects belonging to different orders of insects, considering that the majority of relevant publications were focused on them.

2. Automatic Monitoring of Lepidoptera Pest Species

A wide range of moth and butterfly species, such as gypsy moth *Lymantria dispar*, codling moth *Cydia pomonella*, diamondback moth *Plutella xylostella*, are known to cause significant yield losses in many crops worldwide [4,5]. These insects are able to oviposit a large amount of eggs, and the larval stages feed voraciously, causing direct defoliation, thus leading to huge losses when populations are well developed. Currently, the usual surveillance method is based on delta traps with pheromone lures. However, due to the multiple poses that these insects can display when attached to sticky traps, the development of automatic detection and identification models is quite challenging [6,7].

Silveira and Monteiro [8] developed a tool to automatically identify eyespot patterns of the nymphalid butterfly *Bicyclus anynana*, using a machine learning algorithm with features based on circularity and symmetry to detect eyespots on the images. The software was also able to successfully recognize patterns of other butterfly species.

For the identification of species based on images, Wen et al. [6] used a suitable combination of shape, color, texture and numerical features extracted for moth description. Later, a pyramidal stacked de-noising auto-encoder (IpSDAE) was proposed to generate a deep neural network for moth identification regardless of the pose of insects. This model reached a level of 98.13% of moth identification at genus level without classifying the species. Guarnieri et al. [9] created an automatic model for codling moth *Cydia pomonella* monitoring. Using a modified trap at field level using mobile phone cameras of different resolutions for remote visual inspection, they reached up to 100% efficacy when compared to local visual inspection.

Many models were developed using artificial neural networks (ANNs) for species identification. ANNs are computational models inspired in biological neural networks that can be trained to perform different tasks, such as identifying patterns in images. Kaya et al. [10] combined ANNs with binary patterns to identify five butterfly species of the family Papilionidae, whilst Wang et al. [11] combined ANNs with a series of morphological features and a support vector machine (SVM) to develop a high efficacy system (93%) for the identification of over 200 species from 64 families of different insect orders including Coleoptera, Hemiptera, Hymenoptera, Lepidoptera, Megaloptera, Neuroptera, Odonata and Orthoptera. Kang et al. [7] developed a model for butterfly identification based on their shapes from different angles, having efficacy ranging between 89% and 100% for 15 species of different families, and Kaya and Kayci [12] proposed a method using color and texture features.

The use of texture descriptors, especially the gray level co-occurrence matrix (GLCM) in the context of machine learning showed to be useful in the identification and monitoring of Lepidoptera species. Kayci and Kaya [12] reached 96.3% accuracy in the identification of 19 species belonging to the family Pieridae.

Kaya et al. [13] applied a texture gabor filter-based and extreme machine learning model for butterfly identification of five species with an accuracy of 97%.

Applying deep convolutional neural networks and deep learning techniques on three publicly available insect datasets, Thenmozhi and Reddy [14] were able to identify several species of Lepidoptera, as well many Coleoptera and Orthoptera with accuracy varying from 95% to 97%.

Using histograms of multi-scale curvature (HoMSC), gray-level co-occurrence matrix of image blocks (GLCMoIB) and weight-based k-nearest neighbor classifier, Li and Yin [15] reached 98% efficacy in the identification of 50 lepidopteran species.

Different approaches were conducted by two studies [16,17] that developed a mobile robot car with camera for real-time identification of Pyralidae species on field level. Applying the Gaussian Mixture Model (GMM), Aggregation Dispersion Variance (ADV) and Distance Regularization Level Set Evolution (DRLSE), Zhao et al. [17] were able to identify the target species with an accuracy of 95%. Liu and colleagues [16] designed a two-step recognition, using first a colour space (HSV) in which candidates were evaluated by applying Otsu segmentation thresholds, and secondly an object contour recognition procedure was performed based on Hu moments; the model was able to detect Pyralidae species with 94.3% accuracy, superior to the support vector machine method.

Commercial solutions are produced by EFOS (Figure 1) based on cloud computing image processing. The model integrating bucket/funnel type of trap is especially suitable for larger moth species that come in high numbers. It can remotely identify *Helicoverpa armigera*, *Autographa gamma* and *Spodoptera* spp. (Figure 1a).

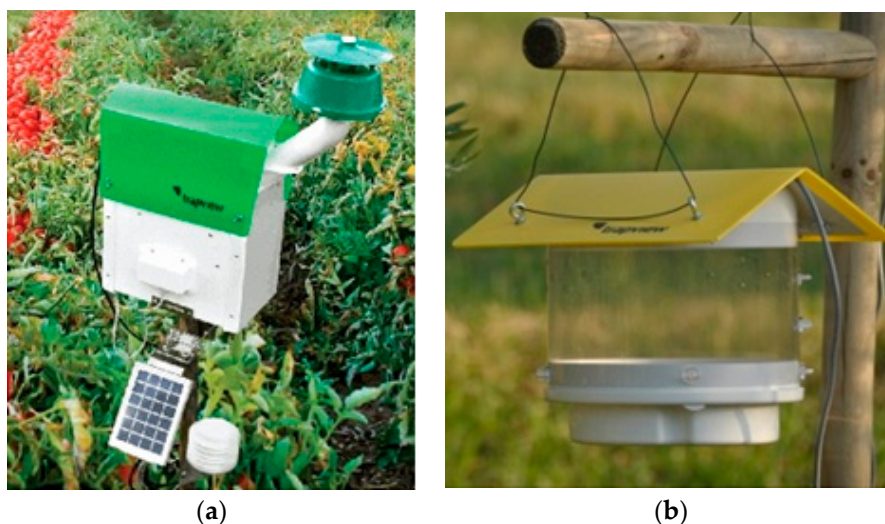


Figure 1. Automatic trap for monitoring and moth species (a) and fruit flies (b), EFOS, Trapview, Slovenia.

3. Automatic Monitoring of Sucking Insects

Sucking pests are among the critical factors causing losses in greenhouse environments. Thrips, aphids, and whiteflies are recognized as some of the most problematic pests when crops are cultivated in congested conditions in closed areas. Usually in greenhouses, sticky traps are placed in order to monitor the populations of these pests. This method can be considered a difficult task for not specialized professionals regarding their size, the complex morphology and the low efficiency score of fatigued or unskilled human observers [18].

Several studies were conducted for automatic identification of greenhouse sucking pests. For the identification of the greenhouse whitefly *Trialeurodes vaporariorum* at the mature stage, models applying computer vision techniques were proposed by a few authors [19,20]. Using threshold algorithms, Bodhe and Mukherji [21] developed a system to detect and count whiteflies using image analysis. Applying texture and shape analysis, Ghods and Shojaeddini [22] created an algorithm that could identify whiteflies in plant leaves with 85% accuracy. Blasco et al. [23] developed a prototype to monitor insect traps placed in the field by capturing and sending images of the trapped insects to a remote server. The device was created on the basis of a Raspberry Pi platform and incorporates a

camera to capture the images, a control board to program the image capture intervals and a modem to send the images and additional information to a remote media server.

Applying noise removal, contrast enhancement techniques and k-means, Dey et al. [24] used statistical feature extraction methods such as GLCM and gray level run length matrix (GLRLM) before the use of classifiers like support vector machine, Bayesian classifier, artificial neural network, binary decision tree classifier and k-nearest neighbor to distinguish white fly pest infested from healthy leaf images. All these classifiers present a high accuracy (90–98%) and high sensitivity (93.9–98.8%).

Using a support vector machine, Ebrahimi et al. [25] developed a model to detect and classify multiple stages of whitefly. Automatic models based on color transformations were proposed for counting and measuring whiteflies in soybean leaves [26], in different leaves [27], and in yellow sticky traps [28,29].

With a more practical IPM-based approach, multiple techniques were developed. Using samples of yellow sticky traps from greenhouses, Qiao et al. [30] proposed a model for density estimation of the silverleaf whitefly *Bemisia tabaci* based on image processing system. In this work, the system proved to be more efficient with medium and high densities of the pest.

For score and identification of *Bemisia tabaci* in yellow sticky traps and the western flower thrips *Franklinella occidentalis* on blue sticky traps, Sun et al. [31] applied a novel smart vision algorithm using two-dimensional Fourier transformed spectra, finding high correlation with human vision accuracy. Similar results were obtained by Solis-Sanchez [32].

Combining image processing (segmentation, morphological and color property estimation) and ANN, Espinoza et al. [33] proposed a model for the identification and monitoring of *Franklinella occidentalis* and *Bemisia tabaci* with high precision (96%). Bauch and Rath [34] used shape and colour properties for the identification of *Bemisia tabaci* and *Trialeurodes vaporariorum*, achieving 85% efficacy.

Furthermore, Lu et al. [35] proposed a convolutional network (CNN) classifier model in combination with a generative adversarial network (GAN) image augmentation. For this method, a Raspberry Pi v2 camera was used, both whiteflies and thrips were analyzed, and synthetic images were created through the GAN-based data augmentation method, in order to enhance CNN classifier with limited image data. This method resulted in a precision of a range between 85% and 95%.

Seeking out a way to process images with a low computational cost, a model fitting embedded system for the detection of aphids, whiteflies and thrips was proposed by Xia et al. [18], whilst Xuesong et al. [36] developed a model for the counting and identification of aphids based on machine learning and an adapted smartphone. In yellow sticky traps placed in greenhouse conditions, this method showed 95% accuracy and 92.5% when placed outside. Liu et al. [37] developed a model that uses a maximally stable extremal region descriptor, and then used histograms of oriented gradient features and a support vector machine for identification of wheat aphids in field conditions. This new method provides an 86.81% identification rate. In addition, Li et al. [38] developed a convolutional neural network (CNN) of Zeiler and Fergus model and a region proposal network (RPN) with non-maximum suppression (NMS), achieving a precision of over 88.5%.

Using leaf samples, Maharlooei et al. [39] designed a model for identification of soybean aphid (*Aphis glycines*) based on image processing techniques with different types of cameras and two illumination conditions and compared results with human counting. The best results were obtained with low illumination and Sony camera with 96% accuracy. Also using leaf samples (of pakchoi), Chen et al. [40] proposed a model for the segmentation and counting of aphid nymphs. This system showed a high accuracy (99%) when compared to human counting, but it is not selective and can be used for other pests. As a limitation, this and some of the systems described appear just to count and not distinguish between the whiteflies or aphid species.

Using sticky traps, a camera, a temperature sensor and an ambient light sensor, Rustia et al. [41] developed a RGB to LUV color model conversion in order to extract the V-channel color component, achieving an accuracy of between 90% and 96%.

Furthermore, Gutierrez et al. [42] conducted a study for monitoring and identifying whiteflies. For this purpose, two cameras, a dataset generator and two microcontrollers were used in combination with a K-nearest neighbor (KNN) and multilayer perceptron (MLP), resulting in an accuracy of between 66% and 81%.

4. Automatic Identification and Monitoring of Fruit Flies

One of the biggest challenges of horticultural and fruit production in the Mediterranean, tropical, and subtropical areas of the world is the frugivorous fruit flies (Diptera: Tephritidae) [43]. This group of pest causes crop losses amounting to billions of dollars each year worldwide, totaling USD 242 million/year in Brazil alone [44]. Depending on the crop and on the lack of control methods, fruit fly damages can lead to from 80% to 100% of crop losses [45]. The fruit fly belongs to the tribe Dacini and have 932 recognized species, in which about 10% are currently recognized as pests of commercial fruit and vegetable production, causing quarantine issues and trade embargos [46]. Females of *Ceratitis capitata* (Wiedemann), *Bactrocera Dorsalis* (Hendel) and *Bactrocera oleae* (Gmelin), among other species, exhibit high reproductive rates. These insects lay eggs below the fruit surface and when they hatch, the larvae feed inside the fruit. For this reason, the fruit drops or loses quality [45].

The global fruit fly issue is intensified by a small group of highly polyphagous and highly invasive pest species that competitively dominate the local fauna if they enter and establish in a region [47–49]. These pests can be controlled with pesticide sprays, mass trapping containing pheromone lures [50], and by the release of sterile males [51]. The efficiency of the control method depends on the time of application, being more effective when the pest is detected early. Due to a low threshold of control of these pests, an automatic detection system will be useful to prevent and monitoring the infestation of these quarantine pests in a faster way.

In order to improve the operation of a low-cost McPhail trap, Potamitis et al. [47] inserted optoelectronics sensors to monitor the entrance of the pests and identify the species of incoming insects from the optoacoustic spectrum analysis of their wingbeat. With this system, it was possible to distinguish fruit flies from other insects with 91% accuracy, but not between fruit fly species. Improving this system, Potamitis et al. [52] presented a novel bimodal optoelectronic sensor based on a stereo-recording device that records the wingbeat of an insect in flight and Fresnel lens. This system was able to distinguish between *Ceratitis capitata* and *Bactrocera oleae* with 98.99% accuracy (Figures 2 and 3).

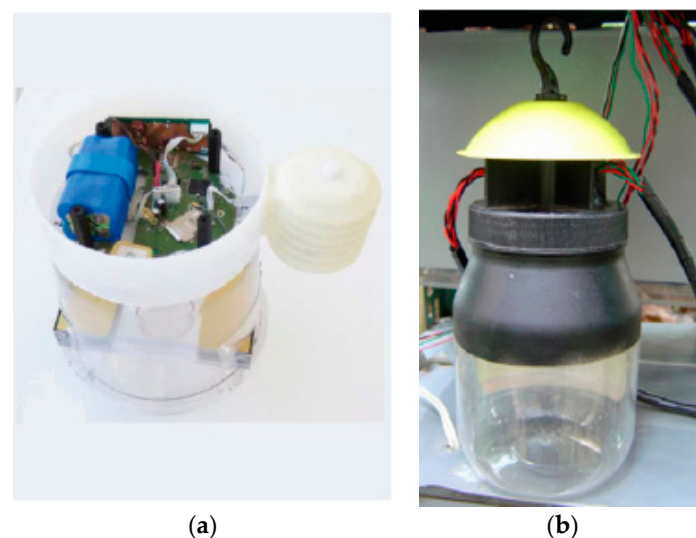


Figure 2. Electronic traps: conventional traps converted to automatic fruit fly monitoring units. McPhail trap equipped with fresnel lens (a) [50]. Automatic trap with infrared sensor for *Bactrocera dorsalis* (b) [3].

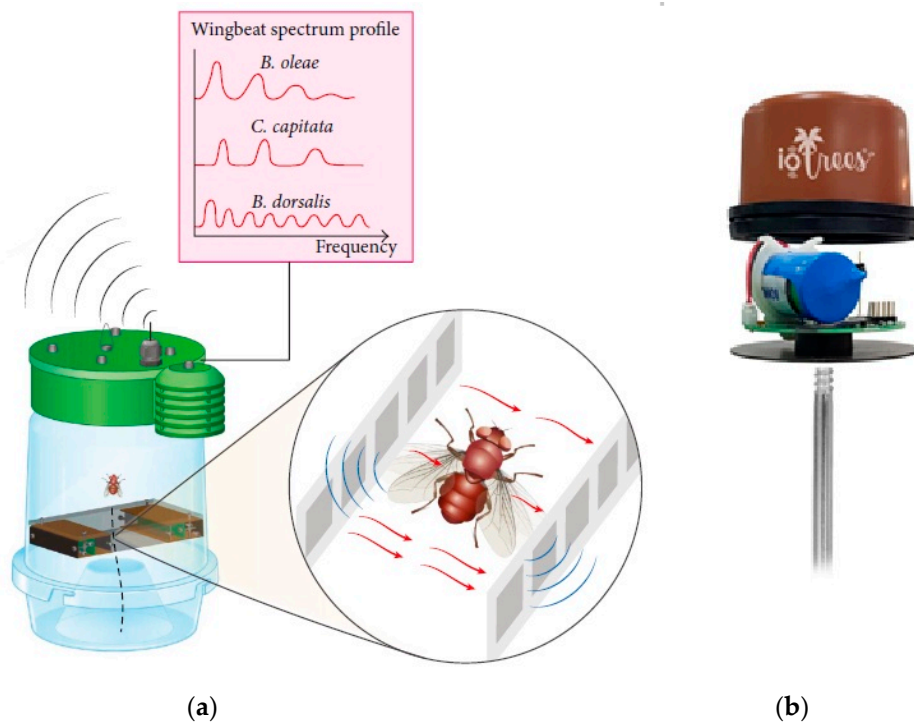


Figure 3. Echo-acoustic trap and sensor: scheme detailing the functioning of an automatic acoustic insect trap (a) [52]. Automatic detection device with seismic sensor for detection of weevils in palm trees (IOSTrees, Spain) (b).

Another system was developed to identify different species of the *Anastrepha* group based on image datasets of wings and aculeus, a specialized ovipositor structure. Faria et al. [53] used a multimodal fusion classifier approach to distinguish images of three species: *Anastrepha fraterculus*, *Anastrepha obliqua*, *Anastrepha sororcula*. In these experiments, the fuzzy support vector machines (FSVM) multimodal approach could account for 98.8% of classification accuracy in the laboratory conditions. In a more applied study, Doitsidis et al. [54] modified a McPhail trap with a camera connected to a Web system that automatically counts the *Bactrocera oleae* and provides the images to expert entomologists that can remotely assess the potential threat at any time and rate, reducing the need for visiting and collect data on site. Okuyama et al. [55] applied an automatic count system to monitor the dynamics of *Bactrocera dorsalis* at the field level using an automatic fly census developed by Jiang et al. [3]. This automatic fly census system reports the environmental conditions and the pests in real time. It is composed of an infrared device counting the insects that enter the trap and recording environmental data (remote monitoring platform—RMP) (Figure 2). The device also has an external host control platform that receives data from the RMP through a short cell phone message (GSM).

In order to improve this system and develop a more accurate and precise device, Liao et al. [56] designed a monitoring system built on two different wireless protocols: GSM and ZigBee, with three major components: remote sensing information gateway (RSIG), a host control platform (HCP) and wireless monitoring nodes (WMNs). The WMNs transmit the collected data (relative humidity, illumination, temperature and the number of Oriental fruit flies captured) to the RSIG, and the RSIG delivers the data to the database server (HCP) for storage and analysis. The server can process the data and classify the information after analysis in three event types: a normal status event, a pest outbreak event and a sensor fault event, which can be accessed through an online platform (Figure 4).

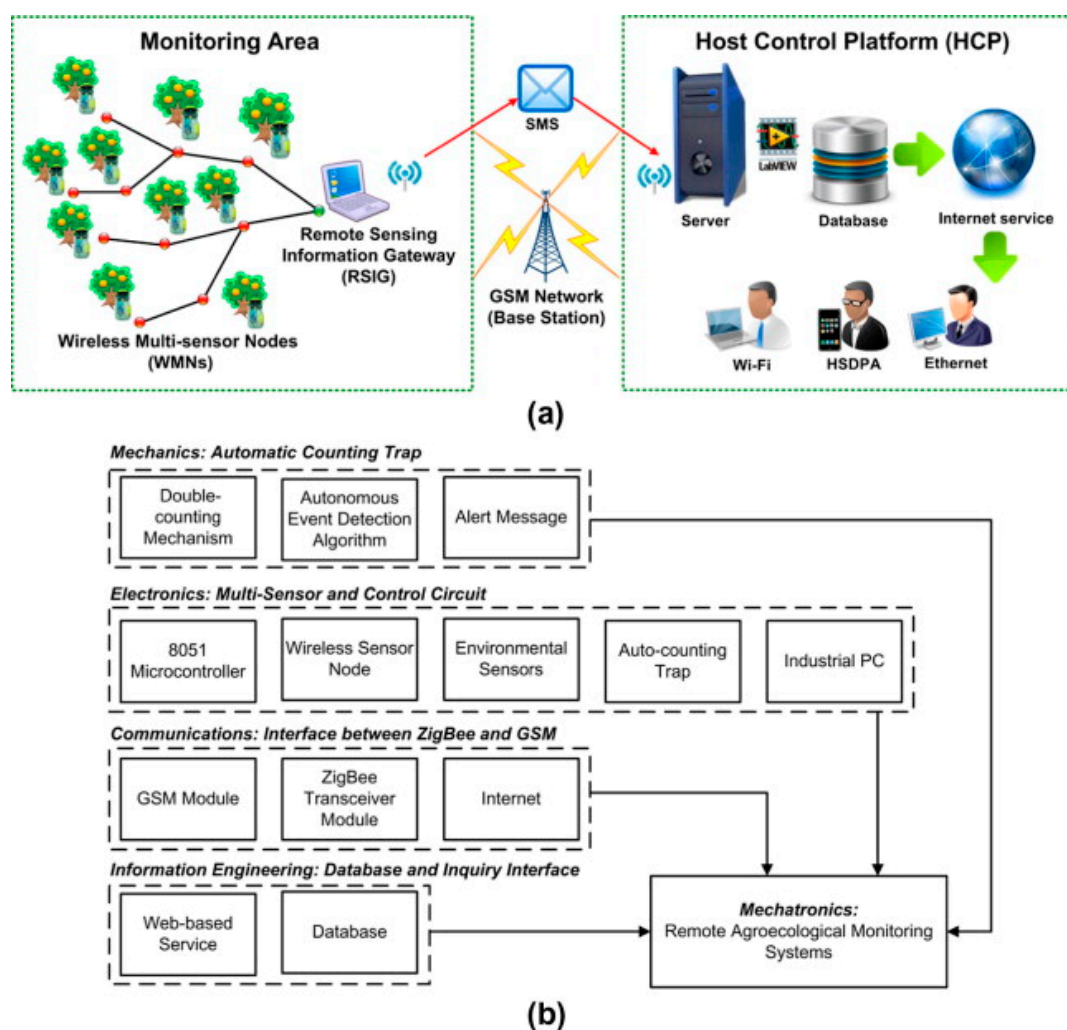


Figure 4. Monitoring system proposed by Liao et al. [54]. Overall conceptual model of the system (a). Block diagram integrating technologies in the remote agroecological monitoring system (b). HSDPA: High Speed Downlink Packet Access. PC: GSM.

Another electronic trap for the monitoring of adult fruit flies was developed by Shaked et al. [57] and extensively tested at the field level. One of the traps was based on specific volatiles for male and female adult *Ceratitis capitata*, and another on the attraction of the adults to the color yellow in order to capture *Bactrocera oleae*, *Dacus ciliates* and *Rhagoletis cerasi*. These traps were image based. Real-time images of the surface of the traps were taken automatically and sent to a server. From the office, the entomologists could classify fruit flies from images with an accuracy superior to 88%. These traps showed good specificity among the different fruit fly species, and do not differ from the results of conventional traps that were operating simultaneously.

A different approach was proposed by Haff et al. [58] using hyperspectral images. It was possible to identify spots caused by fruit fly larvae in mangoes fruits in post-harvest on line with 87.7% accuracy. This approach used image processing parameters such as gaussian blur radius, ball radius, threshold for binary conversion, and minimum particle size.

A commercial automatic trap with a high-resolution camera is manufactured by TrapView, EFOS, Slovenia, for the monitoring of Mediterranean fruit fly (*Ceratitis capitata*) in citrus and peaches and spotted wing drosophila (*Drosophila suzukii*) in fruits and grapes (Figure 1b).

5. Automatic Monitoring of Weevil in Palm Trees and Other Borer Insects

The *Rhynchophorus ferrugineus* or red palm weevil (RPW) is a key pest of palm species in the Mediterranean region, Middle East, Asia and North Africa. It is one of the most important invasive pests worldwide, being considered the single most destructive pest in 40 species of palm trees. RPW has an important socio-economic impact; present in more than 60 countries, this pest is reducing date production and destroying ornamental palms, causing economic losses of millions of dollars each year [59]. The RPW larvae are located deep inside of the palm crowns, offshoots and trunks, causing considerable damage without being visually detected [60]. The intensive trade of palms without visual symptoms was one of the reasons for spreading the pest worldwide. The early detection of the RPW is the key of success to control and eradicate this pest. The difficulty in controlling with insecticides requires an early detection method for minimizing the losses.

Currently, the use of pheromone traps and human inspection are the most widely and effectively used techniques. Recently, the use of entomopathogenic nematodes in green areas inside Spain have presented successful results in controlling the pest. In addition, the overall efficiency and speed of detection can be increased by using promising technologies in automatic detection such as thermal cameras and acoustic detectors, in order to develop an easy-to-handle, cost-effective, quick and reliable device for early detection of RPW [59].

The acoustic activity of *Rhynchophorus ferrugineus* was studied in detail by several researchers [52,61–63] that concluded that the sound produced by the pest can be isolated and differentiated from environmental sounds and other insects. Dosunmu et al. [64] collected RPW sounds inside the palm trees in different conditions and create a model based in the pattern and frequency spectra of the emitted sounds. In their study, the RPW could be differentiated from *Chamaemyia elegans* based on the burst duration, and the detection of the larvae in the offshoots was easier than inside the trunk. Studying the activities of RPW in the larvae stage, Martin et al. [61] concluded that the sound spectrum is constant during chewing and biting activities and differs during the insect movement. Using a speech recognition technique, the presence of the RPW larvae was identified and validated by tree dissections. For the identification of hidden RPW inside coconut trunks, Martin and Juliet [65] used the MfCC (Mel Frequency Cepstral Coefficient) algorithm and vector quantization. Pinhas et al. [61] developed a mathematical model based on speech recognition (Gaussian Mixture Model and Vector Quantization) with a detection rate of 98%. Using Gaussian Mixture Models, Potamitis et al. [52] also had detection rates above 94%. A bioacoustic sensor was developed to detect the weevil in the early infestation stages with 90% success, despite the environmental noises. This prototype also was solar energy based and was able to send information to a control station in schedule defined by the user via wireless [66].

Multiple studies were conducted in order to identify the RPW using acoustic sensors [67–73]. A wireless network connected with acoustic sensors was proposed by Srinivas et al. [71] in order to precisely monitor palm fields. Hetzroni et al. [72] developed a ‘learning data set’ based on the multivariate distribution of nine pre-selected frequencies to detect RPW larvae activity, using a piezoelectric sensor. The sensitivity was low (around 30%) in early larval stages but had a significant improvement (up to 95%) as larvae developed. Herrick and Mankin [74] developed a custom-written insect signal analysis program: “Digitize, Analyze, View, Insect Sounds” (DAVIS), and were able to detect larval burst in 80% of palms inoculated with neonates in the previous day.

Makin et al. [75], using microphones and amplifiers had applied the DAVIS (digitize, analyze, view, insect sounds), achieved 90% accuracy.

Soroker et al. [76] developed a method for identifying digital signature of larvae on trunks, resulting in improvements in detection efficiency of weevils in palms offshoots.

Using a titanium drill bit inserted into the palm tree trunk and a sensor-preamplifier module, Fiaboe et al. [77] could detect the activity of RPW larvae through oscillogram and spectrogram analysis.

A portable, user-friendly acoustic sensor system enabled the identification of larvae in individual infested trees through the use of signal processing analyses that screened out bird and wind noise.

Nowadays, automatic acoustic sensor (seismic) for early detection of RPW in ornamental and date palm trees using wireless sensor networks is commercialized by Agrint, Rockville, USA and for IOTrees, Spain. The platform is connected to a network of palm tree sensor devices and gives the user treat alerts in real-time (Figure 4).

6. Conclusions

The identification and monitoring of insect pests using automatic traps brings a novel approach to the integrated pest management. Systems that use image recognition techniques and neural networks are the most studied ones, being reliable for the fully automatized identification of orders and counting of insects; however, not so many proposed models are able to identify the species level. Other promising image-based systems developed are the ones that aim to send the insect image to a specialist and then the insects could be identified and counted remotely in real time. The infrared sensor traps were shown to be useful for counting insects, but are limited because they cannot identify the species, which can result to misleading data in the survey. Audio traps are another deeply studied approach for monitoring pests. The research presented in the RPW resulted in the creation of commercial solutions. Image-based commercial solutions can be also be found nowadays in the market.

These new systems promise to facilitate the implementation of IPM systems soon. Appendix A summarizes the automatic detection techniques and sensors used for the different insect groups and Appendix B brings a list of commercial providers previously discussed in this review. The record of data will be useful for population dynamics studies and, if related with climate data, can be used in decision support systems and provide real-time information about pest infestation risk.

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Appendix A

Table A1. Automatic Detection Techniques, Sensors and Efficacy for Different Insect Groups.

Group of Insect	Species	Sensors	Automatic Detection Technique	Efficacy	Authors
Aphids	<i>Aphis glycines</i>	SONY camera, images of leaves	Hue, Intensity components and similarity algorithm	>90%	Maharlooei et al., 2017
Aphids	<i>Myzus persicae</i>	Digital camera, images of Pakchoi leaves	Convolutional neural networks	>80%	Chen et al., 2018
Aphids	<i>Not specified</i>	Yellow sticky traps and smartphone camera	GrabCut method, OTSU algorithm and boundary extraction	92.5–95%	Xuesong et al., 2017
Aphids	<i>Wheat aphids - Not specified species</i>	Digital camera, images at field level	Maximally stable extremal region descriptor to simplify the background of field images containing aphids, and then used histograms of oriented gradient features and a support vector machine.	86.81%	Liu et al., 2016
Borer Insects	<i>Anoplophora glabripennis</i>	Screw with piezoelectric sensor (Oyster 723), amplifier (model ENC1485) and stored on an audio recorder (model TCD-D10 Pro II)	Custom-written insect signal analysis program: “Digitize, Analyze, View, Insect Sounds” (DAVIS).	79–84%	Mankin et al., 2008b
Coleoptera, Hemiptera, Hymenoptera, Lepidoptera, Megaloptera, Neuroptera, Odonata and Orthoptera	64 families, 221 species		Series of morphological features, Artificial neural networks (ANNs) and a support vector machine (SVM)	93%	Wang et al., 2012
Fruit Flies	<i>Anastrepha fraterculus</i> , <i>Anastrepha obliqua</i> , <i>Anastrepha sororcula</i>	Nikon DS-Fi1 camera attached to a Nikon SMZ 1500 stereomicroscope	K-nearest neighbors classifier, Naive Bayes, Naive Bayes Tree, support vector machine and multimodal classifiers	88–96%	Faria et al., 2014

Table A1. Cont.

Group of Insect	Species	Sensors	Automatic Detection Technique	Efficacy	Authors
Fruit Flies	<i>Bactrocera dorsalis</i>	Sensors for measuring wind speed, temperature, and humidity, microcontroller (TI MSP430F449 chip), GSM module, GPS receiver, PC and Infrared counting device.	Complex system based on a remote monitoring platform (RMP) and a host control platform (HCP).	72–92%	Jiang et al., 2008
Fruit Flies	<i>Bactrocera dorsalis</i>	Sensors for measuring wind speed, temperature, and humidity, microcontroller (TI MSP430F449 chip), GSM module, GPS receiver, PC, Infrared counting device and wireless sensor network	Complex system based on a remote monitoring platform (RMP) and a host control platform (HCP).	-	Okuyama et al., 2011
Fruit Flies	<i>Bactrocera dorsalis</i>	GSM and ZigBee, with three major components: remote sensing information gateway (RSIG), a host control platform (HCP) and a wireless monitoring nodes (WMNs). The WMNs transmits the collected data (relative humidity, illumination, temperature and the number of Oriental fruit flies captured) to the RSIG, the RSIG deliver the data to the database server (HCP) for storage and analysis.	Cloud computing image processing and environmental data	98–100%	Liao et al., 2012
Fruit Flies	<i>Bactrocera oleae</i>	Several webcams associated in a modified McPhail trap connected to internet	Gaussian blur filter, OTSU algorithm threshold and counting	75%	Doitsidis et al., 2017

Table A1. Cont.

Group of Insect	Species	Sensors	Automatic Detection Technique	Efficacy	Authors
Fruit Flies	<i>Bactrocera oleae</i> , <i>Ceratitis capitata</i>	Modified traps with Fresnel lenses and associated wingbeat stereo-recording device	Linear support vector classifier, radial basis function support vector machine, random forests, adaboost metaclassifier, extra randomized trees, gradient boosting classifier, convolutional neural network.	98–99%	Potamitis et al., 2018
Fruit Flies	<i>Bactrocera oleae</i> , <i>Ceratitis capitata</i> , <i>Bactrocera Dorsalis</i>	Modified traps with optoelectronic sensor	Linear support vector classifier, radial basis function support vector machine, random forests, adaboost metaclassifier, extra randomized trees, gradient boosting classifier, convolutional neural network.	81–90%	Potamitis et al., 2017
Fruit Flies	<i>Dacus ciliatus</i> , <i>Rhagoletis cerasi</i> , <i>Bactrocera oleae</i>	Digital camera inserted in a modified trap	Remote visual inspection	>88%	Shaked et al., 2018
Lepidoptera	19 species of the family Pieridae	Image database	Texture and color filter with gray level co-occurrence matrix (GLCM)	92.85%	Kaya and Kayci 2014
Lepidoptera Coleoptera Orthoptera	40 species	Image database	Deep convolutional neural networks and transfer learning	95–97%	Thenmozhi and Reddy, 2019
Lepidoptera	5 species of the family Papilionidae	Image database	Artificial neural network with binary patterns	70–98%	Kaya et al., 2015
Lepidoptera	5 species of the family Papilionidae	Nikon Professional camera	Texture gabor filter-based, and extreme machine learning	97%	Kaya et al., 2013
Lepidoptera	50 species	Digital camera	Histograms of multi-scale curvature (HoMSC), gray-level co-occurrence matrix of image blocks (GLCMoIB) and weight-based k-nearest neighbor classifier	98%	Li and Xiong 2017
Lepidoptera	<i>Bicyclus anynana</i>	Nikon SMZ1500 dissecting microscope at 3.8× magnification and a Digital Camera	Machine learning algorithm with features based on circularity and symmetry (1D Hough Transform which corresponds to histogramming)	96%	Silveira and Monteiro 2009

Table A1. Cont.

Group of Insect	Species	Sensors	Automatic Detection Technique	Efficacy	Authors
Lepidoptera	<i>Celastrina argiolus</i> , <i>Cynthia cardui</i> , <i>Dilipa fenestra</i> , <i>Favonius orientalis</i> , <i>Graphium sarpedon</i> , <i>Libythea celtis</i> , <i>Luehdorfia puziloi</i> , <i>Lycaena dispar</i> , <i>Lycaena phlaeas</i> , <i>Ochlodes subhyalina</i> , <i>Papilio maackii</i> , <i>Papilio xuthus</i> , <i>Parantica sita</i> , <i>Parnassius bremeri</i> , <i>Sasakia charonda</i>	Image database	BLS entropy profile and artificial neural network	89–100%	Kang et al., 2014
Lepidoptera	<i>Cydia pomonella</i>	Modified commercial trap with mobile camera with different resolutions	Remote visual inspection	Up to 100%	Guarnieri et al., 2011
Lepidoptera	<i>Cydia pomonella</i> , <i>Choristoneura rosaceana</i> , <i>Argyrotaenia velutinana</i> , <i>Grapholita molesta</i> , <i>Platynota idaeusalis</i> , <i>Spilonota ocellana</i> , <i>Rhagoletis pomonella</i> , <i>Rhagoletis cingulata</i> , <i>Grapholita prunivora</i>	Yellow sticky traps and webcam camera (Creative Inc., USA)	Combination of shape, color, texture and numerical features. Then, a pyramidal stacked de-noising auto-encoder (IpSDAE) was proposed to generate a deep neural network	98.13%	Wen et al., 2015
Lepidoptera	<i>Helicoverpa armigera</i> , <i>Autographa gamma</i> and <i>Spodoptera spp.</i>	Bucket/funnel trap with Camera	Cloud computing image processing	-	EFOS, Slovenia
Lepidoptera	Pyralidae family (do not specify species)	Mobile robot car with camera	Gaussian Mixture Model (GMM), Aggregation Dispersion Variance (ADV) and Distance Regularization Level Set Evolution (DRLSE)	95%	Zhao et al., 2019

Table A1. Cont.

Group of Insect	Species	Sensors	Automatic Detection Technique	Efficacy	Authors
Lepidoptera	Pyralidae family (do not specify species)	Mobile robot car with camera	Conversion into HSV space, extraction of the H spatial matrix, normalization of histogram, Otsu segmentation and object contour recognition based on Hu moments.	94.3%	Liu et al., 2019
Palm Weevil	<i>Rhynchophorus cruentatus</i>	Sensor-preamplifier module (model SP-1L Acoustic Emission Consulting AEC) and an amplifier AED-2000 connected to a digital audio recorder (model HD-P2, Tascam)	Custom-written insect signal analysis program: "Digitize, Analyze, View, Insect Sounds" (DAVIS). A bird-noise profile and a Traffic profile was created, after that a 512-point Fourier transformation was performed	48.6–93.7%	Dosunmu et al., 2014
Palm Weevil	<i>Rhynchophorus ferrugineus</i>	Piezoelectric microphone and 12v amplifier	Vector quantization, Gaussian mixture modeling and nearest neighborhood classifier	92–98.1%	Pinhas et al., 2008
Palm Weevil	<i>Rhynchophorus ferrugineus</i>	Digital voice recorder and soundproof chamber	Analysis of frequency and decibels	-	Martin et al., 2015
Palm Weevil	<i>Rhynchophorus ferrugineus</i>	Digital Voice Recorder	Mel frequency coefficient characteristics and vector quantization	-	Martin and Juliet, 2010
Palm Weevil	<i>Rhynchophorus ferrugineus</i>	An MCE-100 microphone, low-power processor, wireless communication interface and power supply unit	Hanning filter, decibel threshold, wavelet packet transform and vector quantization	>90%	Rach et al., 2013
Palm Weevil	<i>Rhynchophorus ferrugineus</i>	SP-1 probe with an AED-2000 amplifier and also a Sony model TCD-D10 Pro II recorder device	Time-frequency distribution (TFD) based on spectrogram	-	Al-Manie and Alkanhal 2007

Table A1. Cont.

Group of Insect	Species	Sensors	Automatic Detection Technique	Efficacy	Authors
Palm Weevil	<i>Rhynchophorus ferrugineus</i>	Laar ultrasound gate hard disk recording System (frequency range 50 Hz–250 kHz) and Laar WD 60 detector with amplifying system and insertion sensors of different types (contact microphone, airborne ultrasound microphone, contact acceleration sensor and a combined contact/airborne probe sensor)	High pass filter, several time domain features and several frequency domain features	>94%	Hussein et al., 2010
Palm Weevil	<i>Rhynchophorus ferrugineus</i>	POM-3542P-R acoustic microphone coupled a preamplifier stage based on OP37 operational amplifier and a 4th order pass-band, continuous-time active filter (MAX274) applied to the signal in order to select the range of frequencies of interest.	Active filter, frequency spectra analysis	70%	Gutiérrez et al., 2010
Palm Weevil	<i>Rhynchophorus ferrugineus</i>	Magnetic cartridge head	Time-frequency wavelet analysis to relate the spectral frequencies available in the unique acoustic signature of red palm weevil larvae to its time of occurrence.	92–97%	Siriwardena et al., 2010
Palm Weevil	<i>Rhynchophorus ferrugineus</i>	Microphones, radio amplifier, repeater and transmitter connected to a wireless sensor network	Low-pass anti-aliasing filter and Down sampling; Butterworth IIR filter; Threshold filter		Srinivas et al., 2013

Table A1. Cont.

Group of Insect	Species	Sensors	Automatic Detection Technique	Efficacy	Authors
Palm Weevil	<i>Rhynchophorus ferrugineus</i>	Piezo-electric microphone with 50 mm (dia.) membrane was connected to a 20 mm (dia.) magnet via a hollow metal cone. Signal was preamplifier using model MP13. Recordings were performed using Raven Pro 64	'Learning data set' based on the multivariate distribution of nine pre-selected frequencies. threshold criterion (dynamically set) and multivariate classifier	75–95%	Hetzroni et al., 2016
Palm Weevil	<i>Rhynchophorus ferrugineus</i>	AED-2000, digital recorder, and sensor-preamplifier module magnetically attached to a screw and inserted into the base of a pruned palm frond	Custom-written insect signal analysis program: "Digitize, Analyze, View, Insect Sounds" (DAVIS).	>80%	Herrick and Mankin 2012
Palm Weevil	<i>Rhynchophorus ferrugineus</i>	1.59-mm titanium drill bit was inserted into the palm tree trunk and a sensor-preamplifier module (model SP-1) was attached magnetically, AED2000 amplifier and a digital recorder (HD-P2)	Oscillogram and spectrogram analysis	-	Fiaboe et al., 2011
Palm Weevil	<i>Rhynchophorus ferrugineus</i>	Larva Lausher sensor	Sampling frequencies to create digital signature	-	Soroker et al., 2004
Palm Weevil	<i>Rhynchophorus ferrugineus</i> , <i>Oryctes rhinoceros</i>	Digital recorder device	Analysis of frequency and decibels, Los Mel Frequency Cepstral Coefficients and Euclidean distance	-	Martin and Juliet, 2013
Palm Weevil	<i>Rhynchophorus ferrugineus</i> , <i>Rhynchophorus cruentatus</i>	Microphone and amplifier	Custom-written insect signal analysis program: "Digitize, Analyze, View, Insect Sounds" (DAVIS).	90%	Mankin et al., 2008a

Table A1. Cont.

Group of Insect	Species	Sensors	Automatic Detection Technique	Efficacy	Authors
Sucking pests	<i>Bemisia tabaci</i> , <i>Aphis gossypii</i> <i>Thrips tabaci</i>	Scanned sticky traps	Convert image to YCbCr, Segmentation by watershed, colour features and Melanobis distance	>80% relating to human counting	Xia et al., 2015
Sucking pests	<i>Bemisia tabaci</i> , <i>Myzus persicae</i> subsp. <i>nicotianae</i> , <i>Frankliniella fusca</i>	Scanned sticky traps	Colour transformation YUV, fixed threshold and Prewittedge detection method	66–100%	Cho et al., 2008
Sucking pests	<i>Bemisia tabaco</i> , <i>Frankliniella occidentalis</i>	Digital camera (Nikon Coolpix S9200) and yellow and blue sticky traps	Two-dimensional Fourier transformation spectrum	96%	Sun et al., 2017
Sucking pests	<i>Bemisia tabaco</i> , <i>Frankliniella occidentalis</i>	Digitalization of sticky traps	Image-processing algorithm and artificial neural networks	92–96%	Espinoza et al., 2016
Sucking pests	Whiteflies and thrips (do not specify species)	Yellow sticky traps, Raspberry Pi v2 cameras	Convolutional neural network (CNN) classifier model through a generative adversarial network (GAN) image augmentation	85–95%	Lu et al., 2019
Sucking pests	Whiteflies, aphids and thrips (Do not Specify Species)	Scanned yellow sticky traps	Colour transformation, fixed threshold, morphological analysis	66–100%	Cho et al., 2007
Sucking pests	Whiteflies, thrips and aphids (do not specify species)	Sticky traps, camera, humidity and temperature sensor (AM2301 (Guangzhou Aosong Electronics Co., Guangzhou, China). Ambient light sensor (BH1750 (ROHM Semiconductor, Kyoto, Japan)	RGB-to-LUV color model conversion, Extraction of the V-channel color component. Static thresholding for image segmentation. Selective blob filtering.	90–96%	Rustia et al., 2020
<i>Psyllids</i>	(Not Specify)	Camera Raspberry Pi V2 (3280 × 2464pixels)	Insects trap and automatic image collection and storage in a server.	-	Blasco, et al., 2019

Table A1. Cont.

Group of Insect	Species	Sensors	Automatic Detection Technique	Efficacy	Authors
Thrips	Do not Specify	Digital camera (Canon EOS M, 18 MP, CMOS, Japan) mounted in robot arm to capture the flower images	Support Vector Machine classification method with region index and intensifyas color index	>97%	Ebrahimi et al., 2017
Wheat mites	<i>Not specified</i>	Image database	Convolutional neural network (CNN) of ZF (Zeiler and Fergus model) and a region proposal network (RPN) with Non-Maximum Suppression (NMS).	88.5%	Li et al., 2019
Wheat mites Rice planthopper	<i>Not specified</i>	Sony CX-10 GCD camera	Deep convolutional neural networks		Wang et al., 2020
Whiteflies	<i>Bemisia tabaci</i>	Camera coupled with a tube of 10 cm in diameter	Binary masks using colour transformation and fixed threshold	83–95%	Barbedo (2013)
Whiteflies	<i>Bemisia tabaci</i>	Scanned yellow sticky traps	Image transformation 8-bit grayscale, binarization, boundary tracking and counting	94.1–98.1%	Qiao et al., 2008
Whiteflies	<i>Bemisia tabaci</i>	Digital camera, sticky traps and plants	Features corresponding to the eccentricity, area and machine learning	97%	Soliz-Sánchez et al., 2009
Whiteflies	Do not Specify	Image database	Shape, Gray Scale Intensity and Texture analysis	74–85%	Ghods and Shojaeddini 2015
Whiteflies	Do not Specify	Image database	Gray level run length matrix (GLRLM) and gray level co-occurrence matrix (GLCM). Various classifiers like support vector machine, artificial neural network, Bayesian classifier, binary decision tree classifier and k-nearest neighbor classifier	90–98%	Dey et al., 2016

Table A1. Cont.

Group of Insect	Species	Sensors	Automatic Detection Technique	Efficacy	Authors
Whiteflies	Do not specify	Low resolution digital camera	Algorithm based on relative difference in pixel intensities (RDI) using image processing	96%	Huddar et al., 2012
Whiteflies	<i>Trialeurodes vaporariorum</i>	Scanned leaves	Object extraction (Gaussian blur and Laplacian filter) and feature extraction	-	Bhadane et al., 2013
Whiteflies	<i>Trialeurodes vaporariorum</i>	Scanned leaves	Cognitive vision with knowledge-based systems	>85%	Boissard et al., 2008
Whiteflies	<i>Trialeurodes vaporariorum</i> and <i>Bemisia Tabaci</i>	Mobile aspiration mechanism and CCD Camera	Shape and colour properties	85%	Bauch and Rath (2005)
Whitefly	<i>Trialeurodes vaporariorum</i> and <i>Bemisia tabaci</i>	Automatic dataset generator, which is composed by two microcontrollers, two cameras, two tripods, two USB flash drives, two artificial illumination systems, one IP65 box and a portable Wi-Fi 4G router	K-nearest neighbor (KNN) and multilayer perceptron (MLP)	66–81%	Gutierrez et al., 2019

Appendix B

Table A2. List of Commercial Suppliers of Automatic Detection of Pests Devices.

Supplier	Trap Type	Pest	Website
Agrint, USA	Seismic Sensor	<i>Rhynchophorus ferrugineus</i> in palm trees	https://www.agrint.net/
IOTrees, Spain	Seismic Sensor	<i>Rhynchophorus ferrugineus</i> in palm trees	https://www.iotrees.es/
TrapView, Slovenia	Delta trap (Standard)	<i>Cydia pomonella</i> in apples and pears <i>Grapholita funebrana</i> in plums <i>Tuta absoluta</i> in tomatoes <i>Lobesia botrana</i> and <i>Eupoecilia ambiguella</i> in wine grapes <i>Cydia molesta</i> in peaches <i>Plutella Xylostella</i> in plants from Cruciferae family/Brassica	https://www.trapview.com
TrapView, Slovenia	Bucket/funnel (Self Cleaning)	<i>Helicoverpa armigera</i> <i>Autographa gamma</i> <i>Spodoptera spp.</i>	https://www.trapview.com
TrapView, Slovenia	Polarized UV light (Aura)	<i>Ostrinia nubilalis</i> in corn	https://www.trapview.com
TrapView, Slovenia	McPhail (Fly)	<i>Ceratitis capitata</i> in citrus and peaches <i>Drosophila suzukii</i> in grapes and other fruits	https://www.trapview.com
TrapView, Slovenia	Support for Camera and Sticky Trap (Vertical)	Flying and Sucking Pests (not selective for species)	https://www.trapview.com

References

1. Fraser, A. Land grab/data grab: Precision agriculture and its new horizons. *J. Peasant Stud.* **2019**, *46*, 893–912. [[CrossRef](#)]
2. Pedersen, S.M.; Lind, K.M. Precision Agriculture—From Mapping to Site-Specific Application. In *Precision Agriculture: Technology and Economic Perspectives*, 1st ed.; Springer Nature: Basel, Switzerland, 2017; pp. 1–20.
3. Jiang, J.A.; Tseng, C.L.; Lu, F.M.; Yang, E.C.; Wu, Z.S.; Chen, C.P.; Lin, S.H.; Lin, K.C.; Liao, C.S. A GSM-based remote wireless automatic monitoring system for field information: A case study for ecological monitoring of the oriental fruit fly, *Bactrocera dorsalis* (Hendel). *Comput. Electron. Agric.* **2008**, *62*, 243–259. [[CrossRef](#)]
4. Bradshaw, C.J.; Leroy, B.; Bellard, C.; Roiz, D.; Albert, C.; Fournier, A.; Barbet-Massin, M.; Salles, J.M.; Simard, F.; Courchamp, F.; et al. Massive yet grossly underestimated global costs of invasive insects. *Nat. Commun.* **2016**, *7*, 12986. [[CrossRef](#)]
5. Gautam, M.P.; Singh, H.; Kumar, S.; Kumar, V.; Singh, G.; Singh, S.N. Diamondback moth, *Plutella xylostella* (Linnaeus) (Insecta: Lepidoptera: Plutellidae) a major insect of cabbage in India: A review. *J. Entomol. Zool. Stud.* **2018**, *6*, 1394–1399.
6. Wen, C.; Wu, D.; Hu, H.; Pan, W. Pose estimation-dependent identification method for field moth images using deep learning architecture. *Biosyst. Eng.* **2015**, *136*, 117–128. [[CrossRef](#)]
7. Kang, S.H.; Cho, J.H.; Lee, S.H. Identification of butterfly based on their shapes when viewed from different angles using an artificial neural network. *J. Asia-Pac. Entomol.* **2014**, *17*, 143–149. [[CrossRef](#)]
8. Silveira, M.; Monteiro, A. Automatic recognition and measurement of butterfly eyespot patterns. *Biosystems* **2009**, *95*, 130–136. [[CrossRef](#)] [[PubMed](#)]
9. Guarnieri, A.; Maini, S.; Molari, G.; Rondelli, V. Automatic trap for moth detection in integrated pest management. *Bull. Insectol.* **2011**, *64*, 247–251.
10. Kaya, Y.; Kayci, L.; Uyar, M. Automatic identification of butterfly species based on local binary patterns and artificial neural network. *Appl. Soft Comput.* **2015**, *28*, 132–137. [[CrossRef](#)]
11. Wang, J.; Lin, C.; Ji, L.; Liang, A. A new automatic identification system of insect images at the order level. *Knowl. Based Syst.* **2012**, *33*, 102–110. [[CrossRef](#)]
12. Kaya, Y.; Kayci, L. Application of artificial neural network for automatic detection of butterfly species using color and texture features. *Vis. Comput.* **2014**, *30*, 71–79. [[CrossRef](#)]
13. Kaya, Y.; Kayci, L.; Tekin, R. A computer vision system for the automatic identification of butterfly species via gabor-filter-based texture features and extreme learning machine: GF+ELM. *TEM J.* **2013**, *2*, 13–20.
14. Thenmozhi, K.; Reddy, U.S. Crop pest classification based on deep convolutional neural network and transfer learning. *Comput. Electron. Agric.* **2019**, *164*, 104906. [[CrossRef](#)]
15. Li, F.; Yin, X. Automatic identification of butterfly species based on HoMSC and GLCMoIB. *Vis. Comput.* **2018**, *34*, 1525–1533. [[CrossRef](#)]
16. Liu, B.; Hu, Z.; Zhao, Y.; Bai, Y.; Wang, Y. Recognition of Pyralidae Insects Using Intelligent Monitoring Autonomous Robot Vehicle in Natural Farm Scene. *arXiv* **2019**, arXiv:1903.10827.
17. Zhao, Y.; Wang, Y.; Wang, J.; Hu, Z.; Lin, F.; Xu, M. GMM and DRLSE Based Detection and Segmentation of Pests: A Case Study. In Proceedings of the 2019 4th International Conference on Multimedia Systems and Signal Processing, Guangzhou China, 10–12 May 2019; pp. 62–66.
18. Xia, C.; Chon, T.S.; Ren, Z.; Lee, J.M. Automatic identification and counting of small size pests in greenhouse conditions with low computational cost. *Ecol. Inform.* **2015**, *29*, 139–146. [[CrossRef](#)]
19. Bhadane, G.; Sharma, S.; Nerkar, V.B. Early pest identification in agricultural crops using image processing techniques. *Int. J. Electr. Electron. Comput. Eng.* **2013**, *2*, 77–82.
20. Boissard, P.; Martin, V.; Moisan, S. A cognitive vision approach to early pest detection in greenhouse crops. *Comput. Electron. Agric.* **2008**, *62*, 81–93. [[CrossRef](#)]
21. Bodhe, T.S.; Mukherji, P. Selection of color space for image segmentation in pest detection. In Proceedings of the International Conference on Advances in Technology and Engineering, Mumbai, India, 23–25 January 2015.
22. Ghods, S.; Shojaeddini, V. A novel automated image analysis method for counting the population of whiteflies on leaves of crops. *J. Crop Prot.* **2015**, *5*, 59–73. [[CrossRef](#)]

23. Blasco, J.; Sanjuan, S.; Chueca, P.; Fereres, A.; Cubero, S.; Lopez, S.; Alegre, V. *Dispositivo de Captura y Envío de Imágenes a un Servidor Remoto para Monitorizar Trampas para Insectos en el Campo*; No. COMON-2019-agri-3469; X Congreso Ibérico de Agroingeniería: Huesca, Spain, 2019. [\[CrossRef\]](#)
24. Dey, A.; Bhoumik, D.; Dey, K.N. Automatic Detection of Whitefly Pest using Statistical Feature Extraction and Image Classification Methods. *Int. Res. J. Eng. Technol.* **2016**, *3*, 950–959.
25. Ebrahimi, M.A.; Khoshtaghaza, M.H.; Minaei, S.; Jamshidi, B. Vision-based pest detection based on SVM classification method. *Comput. Electron. Agric.* **2017**, *137*, 52–58. [\[CrossRef\]](#)
26. Barbedo, J.G.A. Automatic method for counting and measuring whiteflies in soybean leaves using digital image processing. In Proceedings of the IX Brazilian Congress of Agro-Informatics, Cuiaba, Brazil, 21–25 October 2013.
27. Huddar, S.R.; Gowri, S.; Keerthana, K.; Vasanthi, S.; Rupanagudi, S.R. Novel algorithm for segmentation and automatic identification of pests on plants using image processing. In Proceedings of the Third International Conference on Computing Communication and Networking Technologies, Karur, India, 26–28 July 2012.
28. Cho, J.; Choi, J.; Qiao, M.; Ji, C.W.; Kim, H.Y.; Uhm, K.B.; Chon, T.S. Automatic identification of whiteflies, aphids and thrips in greenhouse based on image analysis. *Int. J. Math. Comput. Simul.* **2007**, *1*, 46–53.
29. Cho, J.; Choi, J.; Qiao, M.; Ji, C.; Kim, H.; Uhm, K.; Chon, T. Automatic identification of tobacco whiteflies, aphids and thrips in greenhouse using image processing techniques. In Proceedings of the 4th WSEAS International Conference on Mathematical Biology and Ecology, Acapulco, Mexico, 25–27 January 2008.
30. Qiao, M.; Lim, J.; Ji, C.W.; Chung, B.K.; Kim, H.Y.; Uhm, K.B.; Myung, C.S.; Cho, J.; Chon, T.S. Density estimation of *Bemisia tabaci* (Hemiptera: Aleyrodidae) in a greenhouse using sticky traps in conjunction with an image processing system. *J. Asia-Pac. Entomol.* **2008**, *11*, 25–29. [\[CrossRef\]](#)
31. Sun, Y.; Cheng, H.; Cheng, Q.; Zhou, H.; Li, M.; Fan, Y.; Shan, G.; Damerow, L.; Lammers, P.S.; Jones, S.B. A smart-vision algorithm for counting whiteflies and thrips on sticky traps using two-dimensional Fourier transform spectrum. *Biosyst. Eng.* **2017**, *153*, 82–88. [\[CrossRef\]](#)
32. Solis-Sánchez, J.J.; García-Escalante, R.; Castañeda-Miranda, I.; Torres-Pacheco, R.G. González. Machine vision algorithm for whiteflies (*Bemisia tabaci* Genn.) scouting under greenhouse environment. *J. Appl. Entomol.* **2009**, *133*, 546–552. [\[CrossRef\]](#)
33. Espinoza, K.; Valera, D.L.; Torres, J.A.; López, A.; Molina-Aiz, F.D. Combination of image processing and artificial neural networks as a novel approach for the identification of *Bemisia tabaci* and *Frankliniella occidentalis* on sticky traps in greenhouse agriculture. *Comput. Electron. Agric.* **2016**, *127*, 495–505. [\[CrossRef\]](#)
34. Bauch, C.; Rath, T. Prototype of a vision based system for measurements of white fly infestation. In Proceedings of the International Conference on Sustainable Greenhouse Systems (Greensys 2004), Leuven, Belgium, 12–16 September 2004.
35. Lu, C.Y.; Rustia, D.J.A.; Lin, T.T. Generative Adversarial Network Based Image Augmentation for Insect Pest Classification Enhancement. *IFAC-PapersOnLine* **2019**, *52*, 1–5. [\[CrossRef\]](#)
36. Suo, X.; Liu, Z.; Sun, L.; Wang, J.; Zhao, Y. Aphid Identification and Counting Based on Smartphone and Machine Vision. *J. Sens.* **2017**, 1–7. [\[CrossRef\]](#)
37. Liu, T.; Chen, W.; Wu, W. Detection of aphids in wheat fields using a computer vision technique. *Biosyst. Eng.* **2016**, *141*, 82–93. [\[CrossRef\]](#)
38. Li, W.; Chen, P.; Wang, B.; Xie, C. Automatic localization and count of agricultural crop pests based on an improved deep learning pipeline. *Sci. Rep.* **2019**, *9*, 1–11.
39. Maharlooei, M.; Sivarajan, S.; Bajwa, S.G.; Harmon, J.P.; Nowatzki, J. Detection of soybean aphids in a greenhouse using an image processing technique. *Comput. Electron. Agric.* **2017**, *132*, 63–70. [\[CrossRef\]](#)
40. Chen, J.; Fan, Y.; Wang, T.; Zhang, C.; Qiu, Z.; He, Y. Automatic Segmentation and Counting of Aphid Nymphs on Leaves Using Convolutional Neural Networks. *Agronomy* **2018**, *8*, 129. [\[CrossRef\]](#)
41. Rustia, D.J.A.; Lin, C.E.; Chung, J.Y.; Zhuang, Y.J.; Hsu, J.C.; Lin, T.T. Application of an image and environmental sensor network for automated greenhouse insect pest monitoring. *J. Asia-Pac. Entomol.* **2020**, *23*, 17–28. [\[CrossRef\]](#)
42. Gutierrez, A.; Ansuategi, A.; Susperregi, L.; Tubío, C.; Rankić, I.; Lenža, L. A Benchmarking of Learning Strategies for Pest Detection and Identification on Tomato Plants for Autonomous Scouting Robots Using Internal Databases. *J. Sens.* **2019**, *2019*, 5219471. [\[CrossRef\]](#)
43. Hendrichs, J.; Vera, M.T.; De Meyer, M.; Clarke, A.R. Resolving cryptic species complexes of major tephritid pests. *Zookeys* **2015**, *540*, 5–39. [\[CrossRef\]](#)

44. Oliveira, C.M.; Auad, A.M.; Mendes, S.M.; Frizzas, M.R. Economic impact of insect pests in Brazilian agriculture. *J. Appl. Entomol.* **2012**, *137*, 1–15. [[CrossRef](#)]
45. White, I.M.; Elson-Harris, M.M. *Fruit Flies of Economic Significance: Their Identification and Bionomics*, 1st ed.; CABI: Wallingford, UK, 1992; p. 70.
46. Downweerd, C.; Leblanc, L.; Norrbom, A.L.; Jose, M.S.; Rubinoff, D. A global checklist of the 932 fruit fly species in the tribe Dacini (Diptera, Tephritidae). *Zookeys* **2018**, *730*, 19. [[CrossRef](#)]
47. Potamitis, I.; Rigakis, I.; Tatlas, N.A. Automated surveillance of fruit flies. *Sensors* **2017**, *17*, 110. [[CrossRef](#)]
48. Duyck, P.; David, P.; Quilici, S. A review of relationships between interspecific competition and invasions of fruit flies (Diptera: Tephritidae). *Ecol. Entomol.* **2004**, *29*, 511–520. [[CrossRef](#)]
49. Duyck, P.; David, P.; Junod, G.; Brunel, C.; Dupont, R.; Quilici, S. Importance of competition mechanisms in successive invasions by polyphagous tephritis in La Reunion. *Ecology* **2006**, *87*, 1770–1780. [[CrossRef](#)]
50. Villalobos, J.; Flores, S.; Liedo, P.; Malo, E. Mass trapping is as effective as ground bait sprays for the control of *Anastrepha* (Diptera: Tephritidae) fruit flies in mango orchards. *Pest Manag. Sci.* **2017**, *73*, 2105–2110. [[CrossRef](#)] [[PubMed](#)]
51. Hendrichs, J.; Franz, G.; Rendon, P. Increased effectiveness and applicability of the sterile insect technique through male-only releases for control of Mediterranean fruit flies during fruiting seasons. *J. Appl. Entomol.* **1995**, *119*, 371–377. [[CrossRef](#)]
52. Potamitis, I.; Rigakis, I.; Vidakis, N.; Petousis, M.; Weber, M. Affordable Bimodal Optical Sensors to Spread the Use of Automated Insect Monitoring. *J. Sens.* **2018**, 1–25. [[CrossRef](#)]
53. Faria, F.A.; Perre, P.; Zucchi, R.A.; Jorge, L.R.; Lewinsohn, T.M.; Rocha, A.; Torres, R.D.S. Automatic identification of fruit flies (Diptera: Tephritidae). *J. Vis. Commun. Image Represent.* **2014**, *25*, 1516–1527. [[CrossRef](#)]
54. Doitsidis, L.; Fouskitakis, G.N.; Varikou, K.N.; Rigakis, I.I.; Chatzichristofis, S.A.; Papafilippaki, A.K.; Birouraki, A.E. Remote monitoring of the *Bactrocera oleae* (Gmelin)(Diptera: Tephritidae) population using an automated McPhail trap. *Comput. Electron. Agric.* **2017**, *137*, 69–78. [[CrossRef](#)]
55. Okuyama, T.; Yang, E.C.; Chen, C.P.; Lin, T.S.; Chuang, C.L.; Jiang, J.A. Using automated monitoring systems to uncover pest population dynamics in agricultural fields. *Agric. Syst.* **2011**, *104*, 666–670. [[CrossRef](#)]
56. Liao, M.S.; Chuang, C.L.; Lin, T.S.; Chen, C.P.; Zheng, X.Y.; Chen, P.T.; Liao, K.C.; Jiang, J.A. Development of an autonomous early warning system for *Bactrocera dorsalis* (Hendel) outbreaks in remote fruit orchards. *Comput. Electron. Agric.* **2012**, *88*, 1–12. [[CrossRef](#)]
57. Shaked, B.; Amore, A.; Ioannou, C.; Valdés, F.; Alorda, B.; Papanastasiou, S.; Goldshtein, E.; Shenderey, C.; Leza, M.; Pontikakos, C.; et al. Electronic traps for detection and population monitoring of adult fruit flies (Diptera: Tephritidae). *J. Appl. Entomol.* **2018**, *142*, 43–51. [[CrossRef](#)]
58. Haff, R.P.; Saranwong, S.; Thanapase, W.; Janhiran, A.; Kasemsumran, S.; Kawano, S. Automatic image analysis and spot classification for detection of fruit fly infestation in hyperspectral images of mangoes. *Postharvest Biol. Technol.* **2013**, *86*, 23–28. [[CrossRef](#)]
59. FAO. Current Situation of Red Palm Weevil in the NENA Region. In *Current Situation of Management Practices, Challenges/Weaknesses and Available Research and Technologies for Its Improvement, Proceedings of Scientific Consultation and High-Level Meeting on Red Palm Weevil Management, Rome, Italy, 29–31 March 2017*; FAO: Rome, Italy, 2017.
60. Pinhas, J.; Soroker, V.; Hetzroni, A.; Mizrach, A.; Teicher, M.; Goldberger, J. Automatic acoustic detection of the red palm weevil. *Comput. Electron. Agric.* **2008**, *63*, 131–139. [[CrossRef](#)]
61. Martin, B.; Shaby, S.M.; Premi, M.G. Studies on acoustic activity of red palm weevil the deadly pest on coconut crops. *Procedia Mater. Sci.* **2015**, *10*, 455–466. [[CrossRef](#)]
62. Nangai, V.L. Interpreting the Acoustic Characteristics of Rpw Towards Its Detection—A Review. *IOP Conf. Ser. Mater. Sci. Eng.* **2017**, *225*, 1–9. [[CrossRef](#)]
63. Martin, B.; Juliet, V. A novel approach to identify red palm weevil on palms. *Adv. Mater. Res.* **2013**, *634*, 3853–3857. [[CrossRef](#)]
64. Dosunmu, O.G.; Herrick, N.J.; Haseeb, M.; Hix, R.L.; Mankin, R.W. Acoustic detectability of *Rhynchophorus cruentatus* (Coleoptera: Dryophthoridae). *Fla. Entomol.* **2014**, *97*, 431–438. [[CrossRef](#)]
65. Martin, B.; Juliet, V. Discriminating human whispers from pest sound during recordings in coconut palm grooves using MFCC and vector quantization. *Int. J. Appl. Bioeng.* **2010**, *4*, 29–33. [[CrossRef](#)]

66. Rach, M.M.; Gomis, H.M.; Granado, O.L.; Malumbres, M.P.; Campoy, A.M.; Martín, J.J.S. On the design of a bioacoustic sensor for the early detection of the red palm weevil. *Sensors* **2013**, *13*, 1706–1729. [[CrossRef](#)]
67. Al-Manie, M.A.; Alkanhalm, M.I. Acoustic detection of the red date palm weevil. *Int. J. Electr. Comput. Energ. Electron. Commun. Eng.* **2007**, *1*, 345–348.
68. Hussein, W.; Hussein, M.; Becker, T. Detection of the red palm weevil *Rhynchophorus Ferrugineus* using its bioacoustics features. *Bioacoustics* **2010**, *19*, 177–194. [[CrossRef](#)]
69. Gutiérrez, A.; Ruiz, V.; Moltó, E.; Tapia, G.; Téllez, M. Development of a bioacoustic sensor for the early detection of Red Palm Weevil (*Rhynchophorus ferrugineus* Olivier). *Crop Prot.* **2010**, *29*, 671–676. [[CrossRef](#)]
70. Siriwardena, K.A.P.; Fernando, L.C.P.; Nanayakkara, N.; Perera, K.F.G.; Kumara, A.D.N.T.; Nanayakkara, T. Portable acoustic device for detection of coconut palms infested by *Rhynchophorus ferrugineus* (Coleoptera: Curculionidae). *Crop Prot.* **2010**, *29*, 25–29. [[CrossRef](#)]
71. Srinivas, S.; Harsha, K.S.; Sujatha, A.; Kumar, N.G. Efficient protection of palms from RPW larvae using wireless sensor networks. *Int. J. Comput. Sci. Issues* **2013**, *10*, 192–200.
72. Hetzroni, A.; Soroker, V.; Cohen, Y. Toward practical acoustic red palm weevil detection. *Comput. Electron. Agric.* **2016**, *124*, 100–106. [[CrossRef](#)]
73. Mankin, R.W.; Mizrach, A.; Hetzroni, A.; Levsky, S.; Nakache, Y.; Soroker, V. Temporal and spectral features of sounds of wood-boring beetle larvae: Identifiable patterns of activity enable improved discrimination from background noise. *Fla. Entomol.* **2008**, *91*, 241–248. [[CrossRef](#)]
74. Herrick, N.J.; Mankin, R.W. Acoustical detection of early instar *Rhynchophorus ferrugineus* (Coleoptera: Curculionidae) in Canary Island date palm, *Phoenix canariensis* (Arecaceae: Arecaceae). *Fla. Entomol.* **2012**, *95*, 983–990. [[CrossRef](#)]
75. Mankin, R.W.; Smith, M.T.; Tropp, J.M.; Atkinson, E.B.; Jong, D.Y. Detection of *Anoplophora glabripennis* (Coleoptera: Cerambycidae) larvae in different host trees and tissues by automated analyses of sound-impulse frequency and temporal patterns. *J. Econ. Entomol.* **2008**, *101*, 838–849. [[CrossRef](#)]
76. Soroker, V.; Nakache, Y.; Landau, U.; Mizrach, A.; Hetzroni, A.; Gerling, D. Note: Utilization of sounding methodology to detect infestation by *Rhynchophorus ferrugineus* on palm offshoots. *Phytoparasitica* **2004**, *32*, 6–8.
77. Fiaboe, K.K.M.; Mankin, R.W.; Roda, A.L.; Kairo, M.T.K.; Johanns, C. Pheromone-Food-Bait Trap and Acoustic Surveys of *Rhynchophorus ferrugineus* (Coleoptera: Curculionidae) in Curacao. *Fla. Entomol.* **2011**, *94*, 766–773. [[CrossRef](#)]



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