



# Supporting Information of the paper: Non conventional peptide self-assemble into a highly conductive supramolecular rope

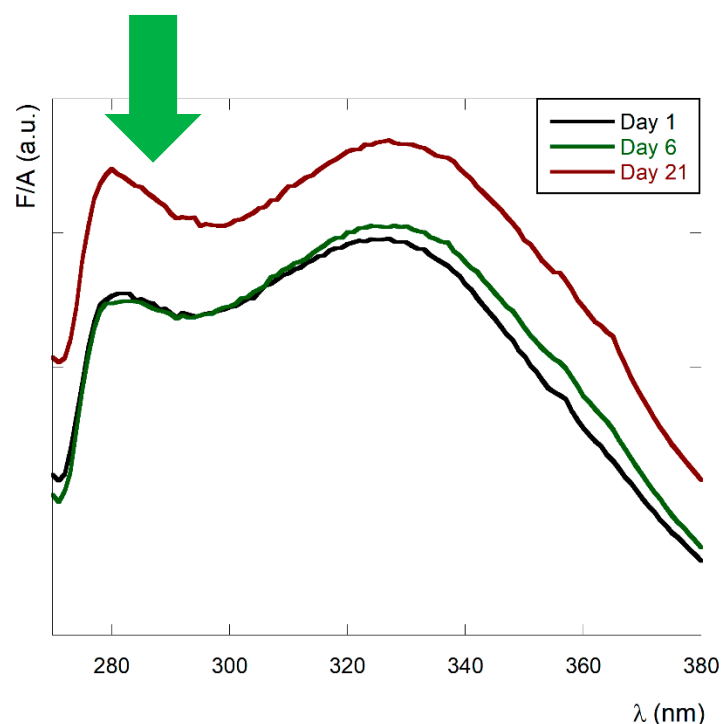
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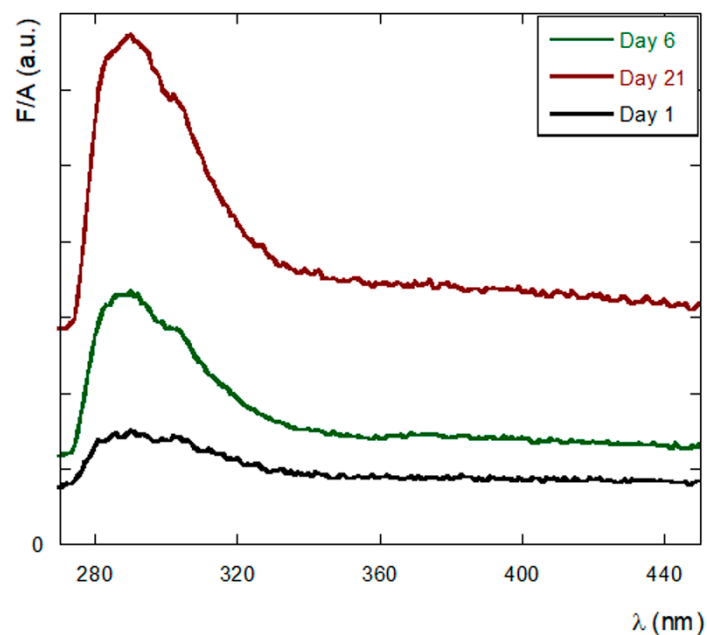
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## Fluorescence measurements.



**Figure S1.** Fluorescence excitation spectrum of peptide **1** in CH<sub>2</sub>Cl<sub>2</sub> solution at a concentration of  $2.8 \cdot 10^{-3}$  M..  $\lambda_{\text{em}}=400$  nm, followed over time.

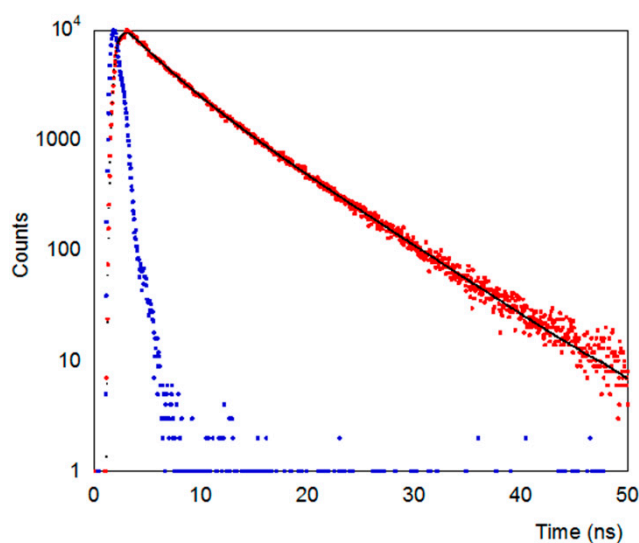


**Figure S2.** Fluorescence spectrum of peptide **1** in CH<sub>2</sub>Cl<sub>2</sub> solution at a concentration of  $2.8 \cdot 10^{-3}$  M..  $\lambda_{\text{ex}}=260$  nm, followed over time.

#### Time-resolved fluorescence measurements.

The decay profile of peptide **1** in CH<sub>2</sub>Cl<sub>2</sub> ( $\lambda_{\text{ex}}=298$  nm;  $\lambda_{\text{em}}=385$  nm) is show in Figure S3. Fluorescence time decay has been fitted through a multi-exponential analysis. In this kind of analysis, the time-decay of  $I(t)$  vs.  $t$  is fitted by the following function:

$$I(t) = \sum_i^n \alpha_i \cdot e^{-\frac{t}{\tau_i}}$$



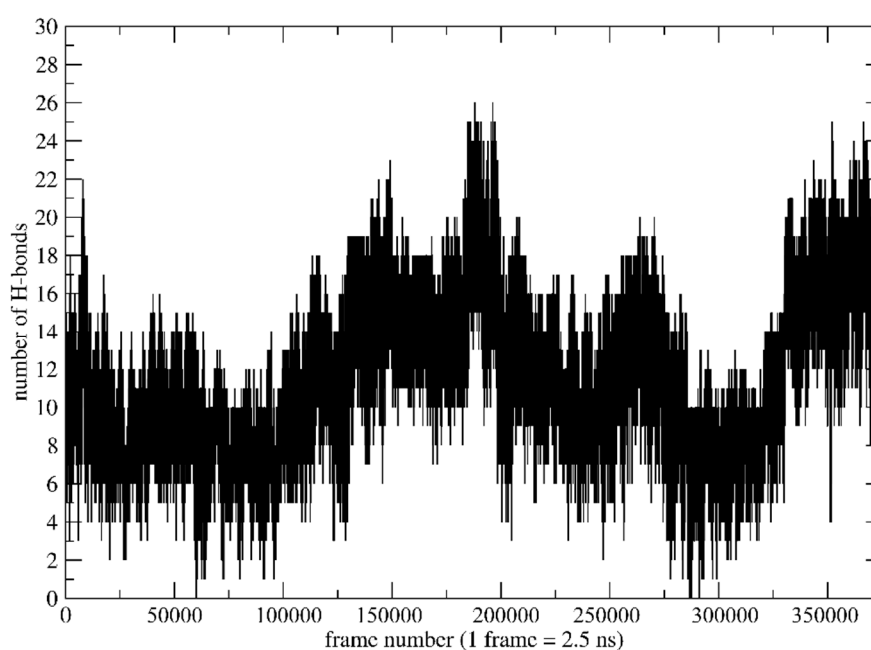
**Figure S3.** Fluorescence-decay curve of the peptide **1** in CH<sub>2</sub>Cl<sub>2</sub> ( $\lambda_{\text{ex}}=298$  nm;  $\lambda_{\text{em}}=385$  nm).

Time-resolved fluorescence measurements confirmed that the signal was due to fluorescence, having a lifetime in the order of several ns. Table 1 reports the value of the time-decay parameters of peptide 1 in CH<sub>2</sub>Cl<sub>2</sub>.

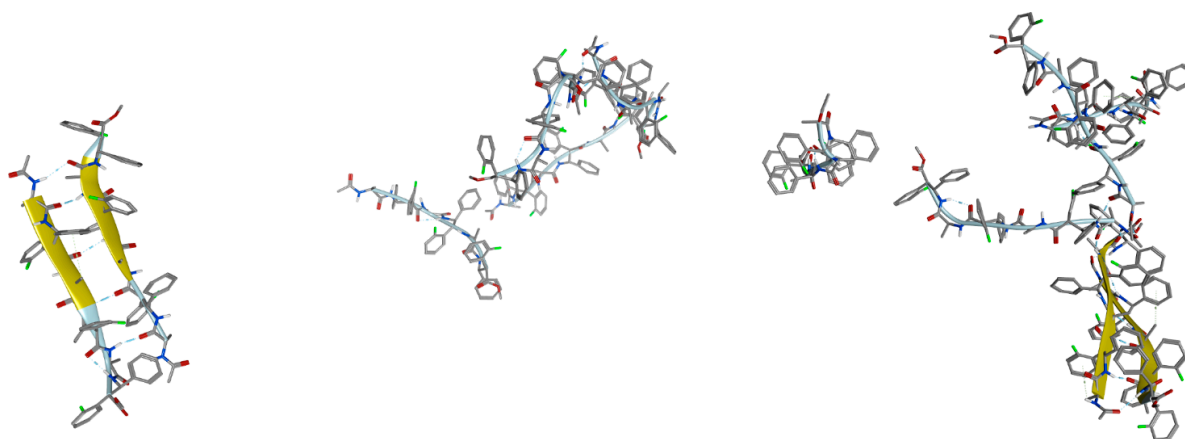
Sample	$\tau_1$	$\alpha_1$	$\tau_2$	$\alpha_2$	$\chi^2$
Peptide 1	0.72	0.37	3.63	0.63	1.3

**Table S1.** Fluorescence time-decay parameters (lifetimes ( $\tau$ ) and pre-exponents ( $\alpha$ )) of excited peptide 1 in CH<sub>2</sub>Cl<sub>2</sub>, recovered by Multiexponential (ME) analysis ( $\lambda_{\text{ex}}$ = 298 nm;  $\lambda_{\text{em}}$ =385 nm).

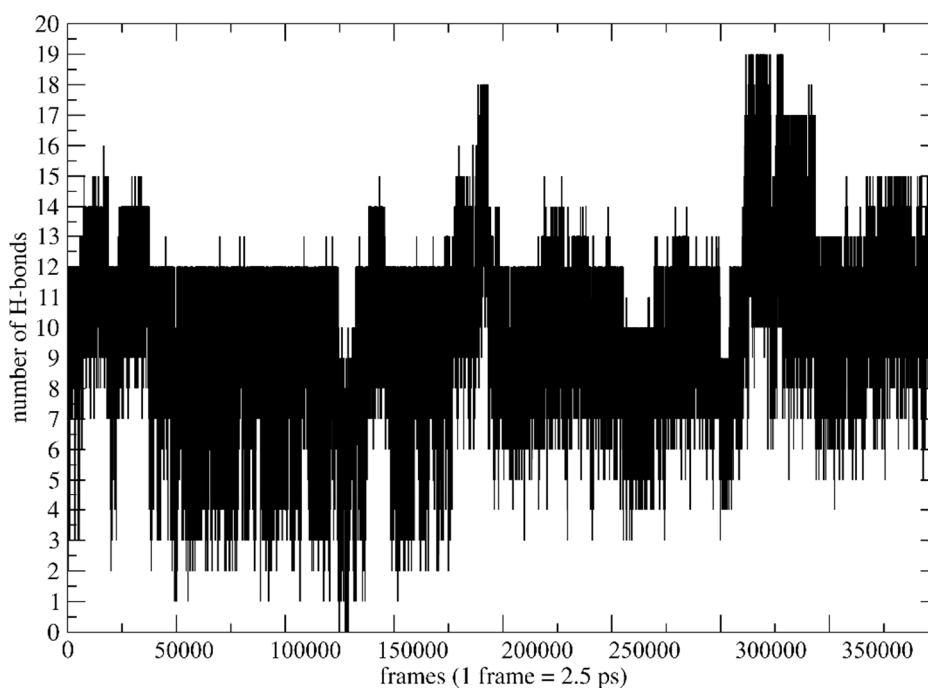
## Molecular modelling



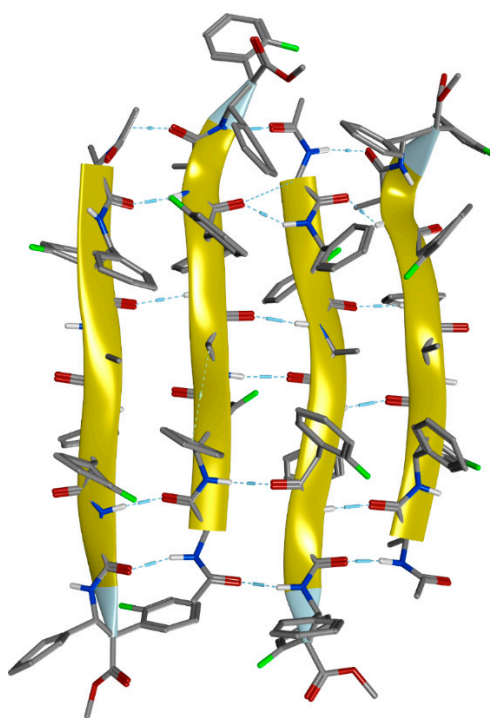
**Figure S4.** Number of intramolecular H-bonds vs simulation time obtained from the analysis of the 1  $\mu$ S trajectory of the first aMD run. The simulation was done on a system of 12 units of peptide 1 solvated by CHCl<sub>3</sub> at a concentration of about 20 mM.



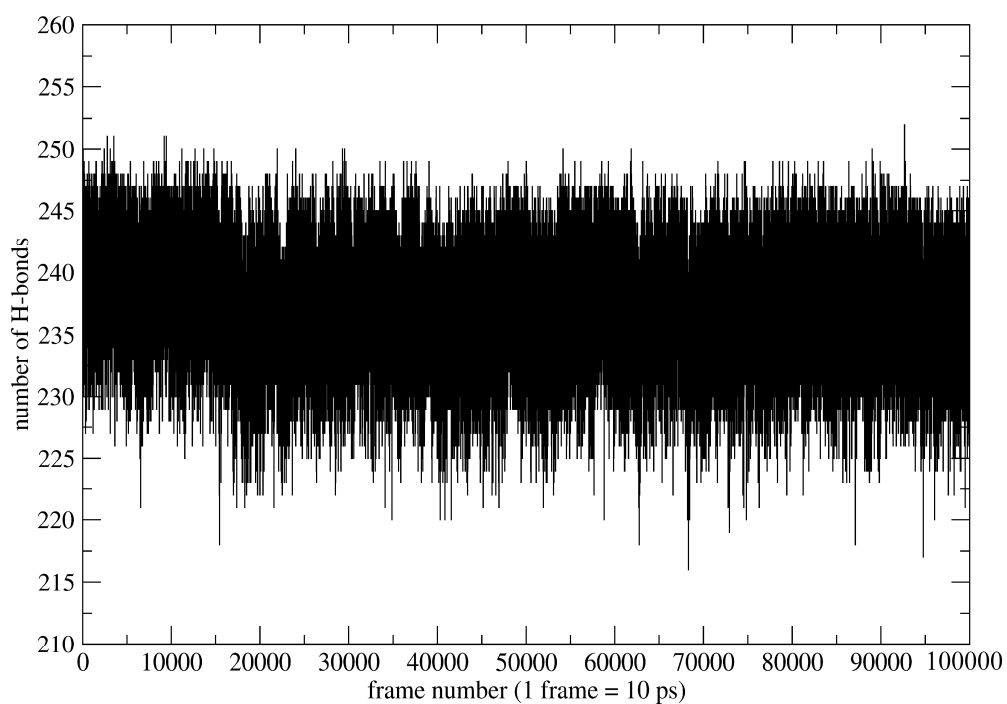
**Figure S5.** Structure of the last frame of the 1  $\mu$ S aMD trajectory of 12 units of peptide 1 randomly placed in a box of explicit  $\text{CHCl}_3$  (solvent not shown).



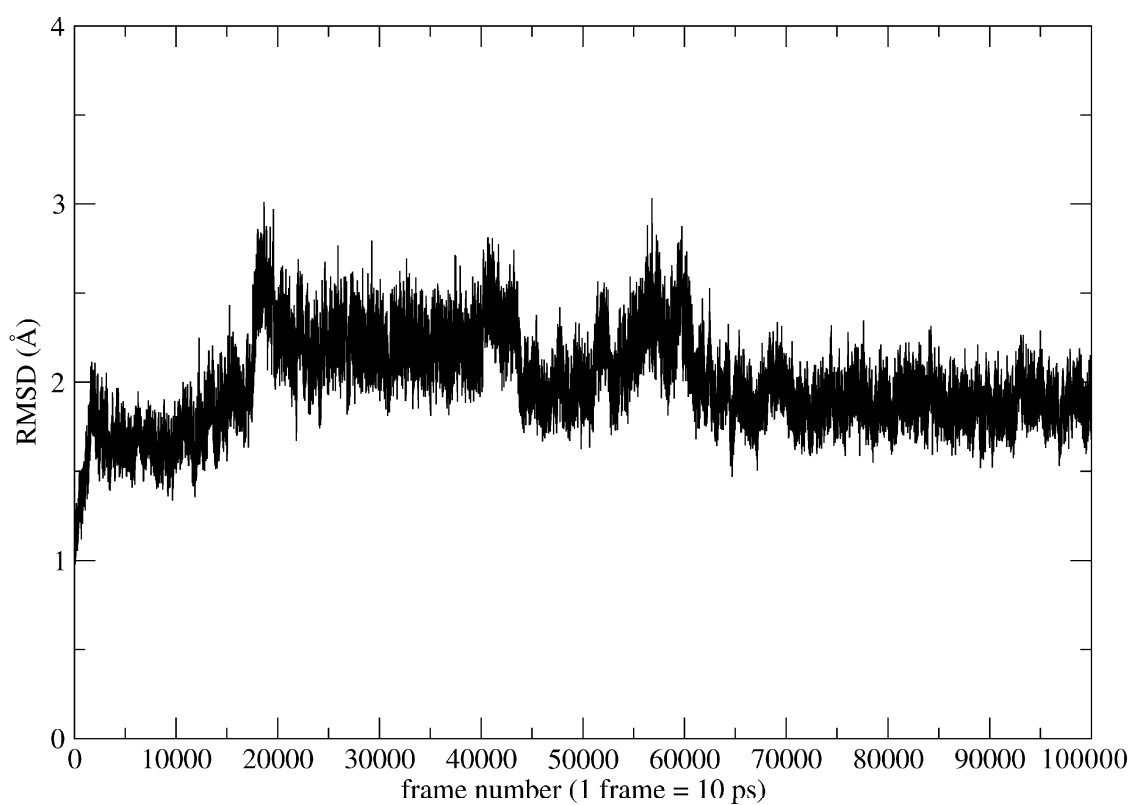
**Figure S6.** Number of intramolecular H-bonds vs simulation time obtained from the analysis of the 1  $\mu$ S trajectory of the second aMD run. The simulation was done on 2 antiparallel  $\beta$ -sheets dimers of 1 peptide solvated by  $\text{CHCl}_3$  at a concentration of about 60 mM.



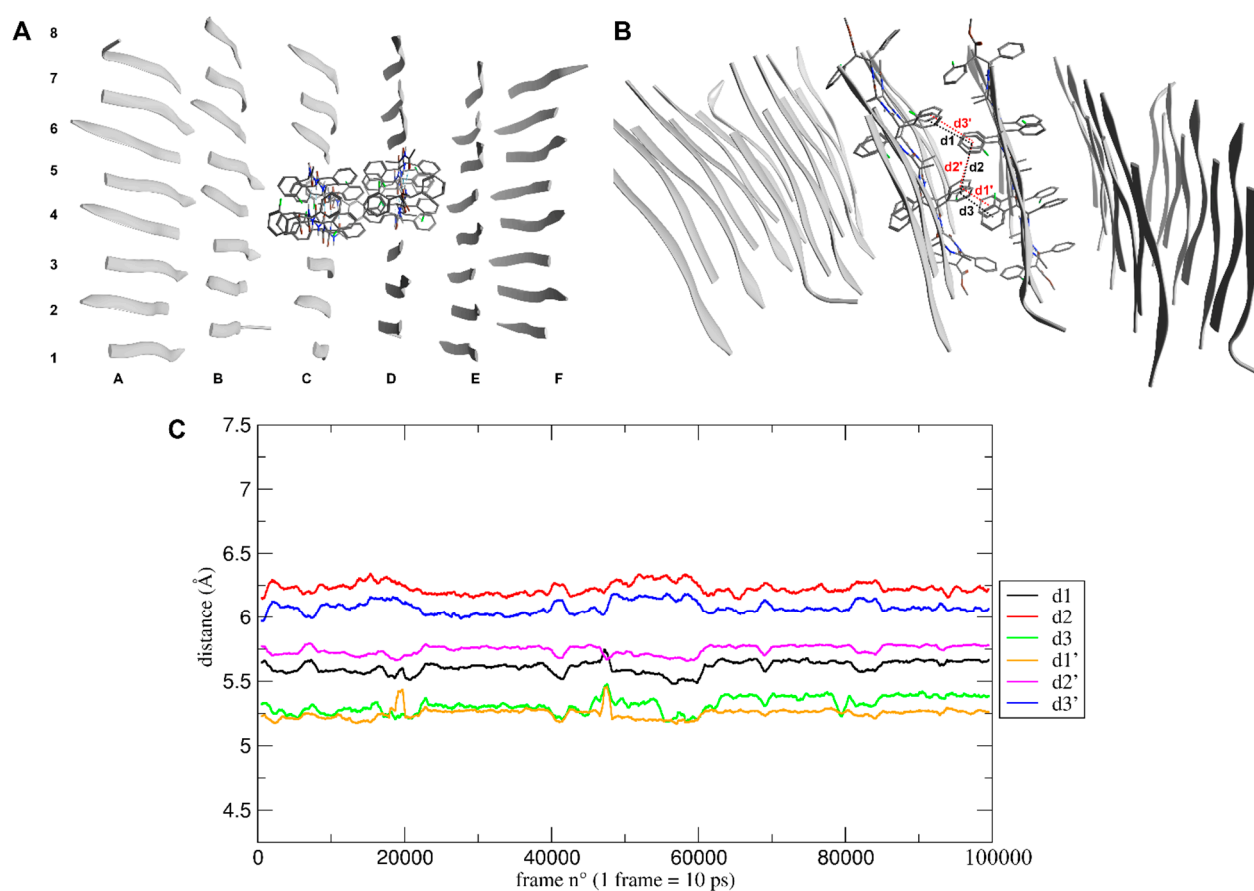
**Figure S7.** Structure of the antiparallel  $\beta$ -sheet tetramer. The geometry was retrieved from the frame number 320120 (approximately 800 ns of simulation time) of the 1  $\mu$ S aMD trajectory of two antiparallel  $\beta$ -sheets dimers of peptide 1 randomly placed in a box of explicit  $\text{CHCl}_3$  (solvent not shown).



**Figure S8.** Number of intramolecular H-bonds vs simulation time obtained from the analysis of the 1  $\mu$ S trajectory of classical MD. The simulation was done on an assembly of 6  $\beta$ -sheets layers made by 8 units of peptide 1 each, in explicit  $\text{CHCl}_3$ .

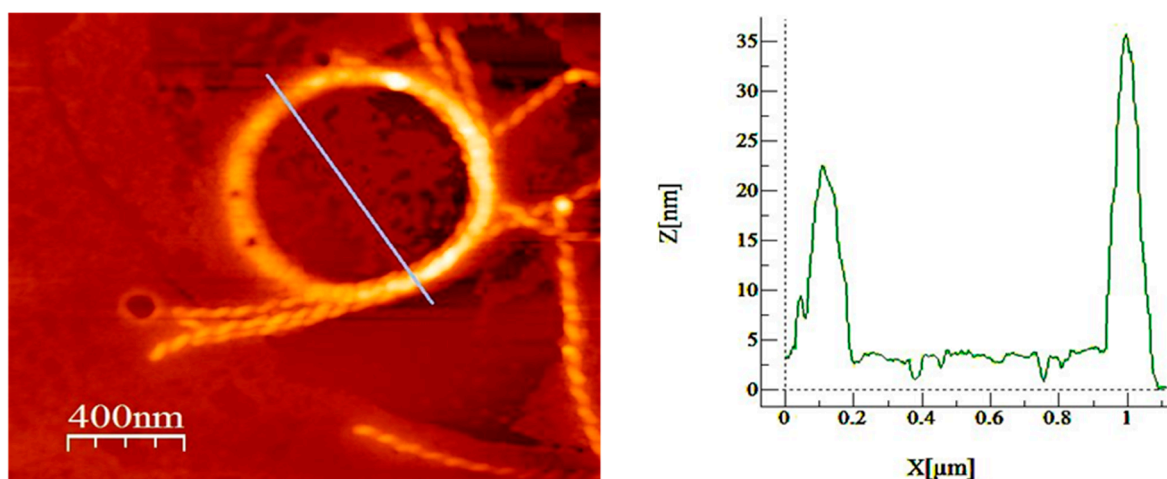


**Figure S9.** RMSD vs simulation time obtained from the analysis of the 1  $\mu$ S trajectory of classical MD simulation of the 6 x 8 peptide **1** assembly.



**Figure S10.** Time evolution (1  $\mu$ s of MD simulation) of representative distances between the centroids of the aryl groups of peptides belonging to adjacent  $\beta$ -sheet layers. Panel **A** shows the top view of the supramolecular assembly with the coordinate system used to label peptides selected for the measurements herein reported. Distances were computed for the aryl-aryl interactions between the peptide pairs C4-D4 (d1-d3) and C5-D5 (d1'-d3'), chosen because of their central position in the 48mer assembly. Panel **B** shows the side view of the assembly and selected distances are highlighted; d1 represents the interaction between the  $\beta$ -2S,3S-Fpg residues in position 4 of peptides C4 and D4; d2 describes the interaction between  $\beta$ -2S,3S-Fpg in position 2 of C4 and  $\beta$ -2S,3S-Fpg in position 4 of D4; d3 represents the interaction between  $\beta$ -2S,3S-Fpg in position 2 of C4 and D4. Distances d1'-d3' describe the same interactions but involving peptides C5 and D5.

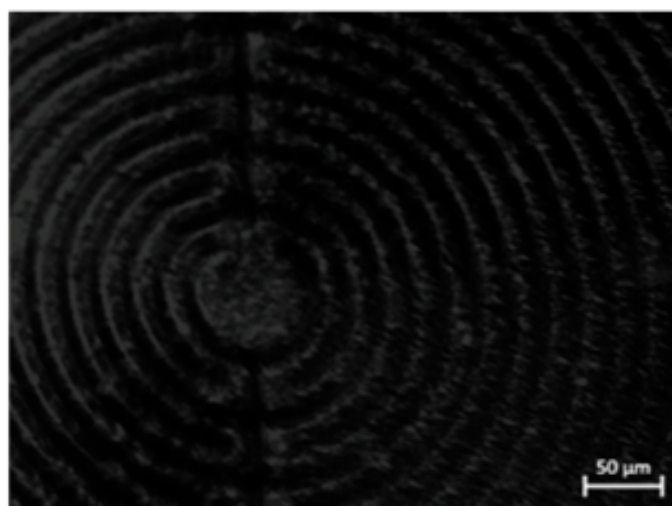
### AFM measurements.



**Figure S11.** AFM characterization of peptide 1 supramolecular structure. Right: line profile reported in the AFM image on the left.

### Fluorescence microscopy measurements.

Figure S12 shows the circular shape of the concentric interdigitated electrode, with have gaps of 10 μm. The image has been obtained with an optical microscope, using a 20x magnification.

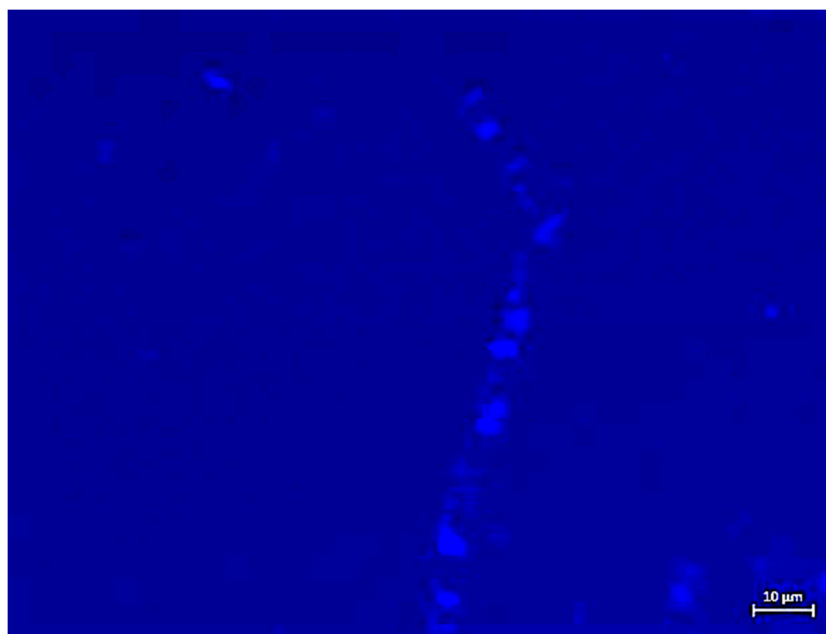


**Figure S12.** Image of the interdigitated electrode, obtained with a fluorescence optical microscope, applying a 20x magnification.

Figure S13 shows the fluorescent filaments of peptide 1 in the blue region. The fibrils were deposited onto the mica surface by cast deposition of a 10 μl drop of a  $2.8 \cdot 10^{-3}$  M fibril suspension in  $\text{CH}_2\text{Cl}_2$ .



After solvent evaporation, several fluorescent fibrils were seen, confirming that the emission is due to aggregation.



**Figure S13.** Image of the filaments of peptide **1**, obtained with a fluorescence optical microscope, using a 20x magnification in the DAPI region (blue channel).