



# **Emerging Roles of Microrobots for Enhancing the Sensitivity of Biosensors**

Xiaolong Lu<sup>1,2,\*</sup>, Jinhui Bao<sup>1,2</sup>, Ying Wei<sup>1,2</sup>, Shuting Zhang<sup>1</sup>, Wenjuan Liu<sup>3,\*</sup> and Jie Wu<sup>4</sup>

- State Key Laboratory of Mechanics and Control for Aerospace Structures, Nanjing University of Aeronautics and Astronautics, Nanjing 210016, China; bao@nuaa.edu.cn (J.B.); weiying07@nuaa.edu.cn (Y.W.); zst000@nuaa.edu.cn (S.Z.)
- <sup>2</sup> Biomedical Engineering Fusion Laboratory, The Affiliated Jiangning Hospital of Nanjing Medical University, Nanjing 211100, China
- <sup>3</sup> College of Materials Science and Engineering, Nanjing Tech University, Nanjing 211816, China
- <sup>4</sup> State Key Laboratory of Analytical Chemistry for Life Science, School of Chemistry and
- Chemical Engineering, Nanjing University, Nanjing 210023, China; wujie@nju.edu.cn
- \* Correspondence: long\_8446110@nuaa.edu.cn (X.L.); liuwenjuan@njtech.edu.cn (W.L.)

Abstract: To meet the increasing needs of point-of-care testing in clinical diagnosis and daily health monitoring, numerous cutting-edge techniques have emerged to upgrade current portable biosensors with higher sensitivity, smaller size, and better intelligence. In particular, due to the controlled locomotion characteristics in the micro/nano scale, microrobots can effectively enhance the sensitivity of biosensors by disrupting conventional passive diffusion into an active enrichment during the test. In addition, microrobots are ideal to create biosensors with functions of on-demand delivery, transportation, and multi-objective detections with the capability of actively controlled motion. In this review, five types of portable biosensors and their integration with microrobots are critically introduced. Microrobots can enhance the detection signal in fluorescence intensity and surfaceenhanced Raman scattering detection via the active enrichment. The existence and quantity of detection substances also affect the motion state of microrobots for the locomotion-based detection. In addition, microrobots realize the indirect detection of the bio-molecules by functionalizing their surfaces in the electrochemical current and electrochemical impedance spectroscopy detections. We pay a special focus on the roles of microrobots with active locomotion to enhance the detection performance of portable sensors. At last, perspectives and future trends of microrobots in biosensing are also discussed.

Keywords: microrobot; biosensor; active control; locomotion; sensitivity

# 1. Introduction

Biosensors featured with high selectivity and sensitivity are ubiquitous for prevailing biological analyses, showing great potential in diverse applications for detecting trace-level chemicals and biomolecules in biomedical engineering, environmental protection, and industrial fabrication, etc. [1–3]. For the last decades, scientists devoted to this research field have developed a number of innovative biosensors according to the diverse characteristics of tested objects [4–6]. Recently, it has become possible for biosensors to measure analysts in a real-time manner, which is significant for the practical monitoring of rapid changes in biological liquids [7–9].

In modern societies, people pay more attention to effective and affordable measurements of food safety [10], disease prevention, health status, and chronic treatment in their daily life [11]. However, for a long time, accurate detections of biological substances can only be performed in laboratories to achieve high sensitivity and low detection limits [1], which requires bulky and expensive systems that can only be operated by professional



Citation: Lu, X.; Bao, J.; Wei, Y.; Zhang, S.; Liu, W.; Wu, J. Emerging Roles of Microrobots for Enhancing the Sensitivity of Biosensors. *Nanomaterials* **2023**, *13*, 2902. https://doi.org/10.3390/ nano13212902

Academic Editor: Daniela Iannazzo

Received: 29 September 2023 Revised: 29 October 2023 Accepted: 30 October 2023 Published: 4 November 2023



**Copyright:** © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). technicians [12]. Consequently, biosensors that can achieve high-quality sensing with costeffective implementation have attracted more and more attention. Recently, the combination with personal intelligent terminals such as mobile phones and iPads makes biosensors more powerful in data processing and online diagnosis [13–16]. To meet the requirements of portable devices for easy-to-go testing, miniaturization is another crucial research focus in biosensor technology [17]. However, the miniaturization of biosensors often suppresses their detection accuracy because of the unexpectedly low efficiency of passive transport in micro volume samples. Currently, the compromise between the detection performance and miniaturization is still an unmet challenge for developing state-of-the-art biosensors.

With the fast development of nanotechnologies, the upgrade of biosensors is remarkably accelerated with rapid improvement in both sensitivity and limit of detection (LOD) [18]. Generally, biosensing nanoprobes have yielded biosensors with trace-level detection capability [17,19]. The achievements and challenges of such active machines compared with passive nanoprobes have been summarized [19–23]. Moreover, an attractive revolution for biosensors is on the way by using micro/nanorobots to achieve active mass transfer in the nanoscale. The micro/nanorobot which is composed of diverse functional nanomaterial can move autonomously as it is powered by chemical fuels or external physical energy sources (light, magnetic, electrical, ultrasound, etc.) [24,25]. Pioneering researches reveal the micro/nanorobot individual can work together in a collaborative manner and accomplish complex tasks including targeted delivery and nano-surgery in intricate environments [26,27]. Such minimized machines exhibit impressive motion agility and controllability, and thus they can trap the target molecules actively to achieve ultrasensitive detections [28]. In addition, the ultrasound power energy, which has been demonstrated to be harmless for biological tissues, can also be applied to actuate micro/nanorobots in a labelfree and contactless way [24,29–31]. These driving modes determine micro/nanorobots are favorable to develop next-generation biosensors.

Focusing on the frontiers of portable biosensors combined with advanced microrobots, this review discusses their recent progress in bio-detection applications according to different assay principles, including fluorescence, surface-enhanced Raman scattering (SERS), locomotion, electrochemical current (EC), and electrochemical impedance spectroscopy (EIS). In view of the ultrasensitive detection potentials with microrobots, different manipulation methods for microrobot individuals and swarms are highlighted. Moreover, the advancement and future challenges of portable biosensors from laboratory research to practical detections are introduced to envision the prospects of the smart biosensors facilitated by micro/nanorobots. Although biosensing platforms with micro/nanorobots are still in their infancy, fantastic advantages based on active locomotion enable such smart micromachines to be an ideal partner for portable biosensors in many scenarios including accurate diagnosis, chronic disease monitoring, and explosive detections, etc., [27,32,33].

## 2. Artificial Microrobots for Biosensing

As innovative artificial machines in micro/nano scale, microrobots can be driven to operate propulsion on demand under the low Reynolds number constraints and carry payloads to move precisely under a navigation strategy for overcoming the Brownian motion [34]. With the rise and rapid development of nanotechnology, researchers have been striving to shrink the functionalized robots to cellular and molecular levels in order to perform delicate tasks in micro/nanoscale, through precise monomer or cluster control. Since then, microrobots consisting of inorganic oxides or smart materials have come into the spotlight in the field of diagnosis, sensing, microsurgery, targeted drug/cell delivery, thrombus ablation, and wound healing [35–37]. In 2016, the Nobel Prize in chemistry highly recognized the great potential of molecular motors, promoting the advancement and innovation of miniaturization technology. Since then, microrobots (also called micromotors, microengines, microrockets, etc.) have come into the spotlight as a powerful tool in various areas including drug delivery [38], nanosurgery, biosensing, and detoxification, which can convert diverse energies into efficient autonomous movement [39,40].

The detection methods, based on fluorescence, surface-enhanced Raman scattering (SERS), locomotion, electrochemical current (EC), and electrochemical impedance spectroscopy (EIS), have now been successfully combined with micro/nanorobots and are ready to be used in portable devices. Among these methods, fluorescence detection is widely accepted with high sensitivity and selectivity and has great advantages in detecting the presence of biomolecules. The enhanced Raman technique shows great potential in the detection of trace-level objects. The electrochemical method has the advantage of high sensitivity and fast response. In addition to the detection of biomolecules, electrical impedance also has great potential in cell detection. However, all these classic biosensing strategies are performed based on passive diffusion, which prevents their performance from being increased any more. In this context, microrobots featured with active locomotion and enrichment are demonstrated to be effective for overcoming such issues to develop next-generation biosensors with better detection sensitivities.

Beyond these five principles, it should also be noted that there are additional detection methods such as acoustic sensing and thermal sensing for biological detections. However, these types of biosensors are always used for low sensitive and harsh environments where microrobots cannot be implemented to perform accurate manipulations.

As listed in Scheme 1 and Table 1, the applications of different types of microrobots in biosensing involve five detection principles: fluorescence detection, SERS, locomotion, EC, and EIS. The design and utilization of these microrobots offer innovative approaches to the field of biosensing. In general, the apparent geometries of microrobots are 50 nm $\sim$ 10  $\mu$ m and their moving speed is 500 nm/s~1 mm/s. A variety of microrobots have been introduced with different materials and principles for distinct target detections. Taking fluorescence detection, for example, the fluorescence intensity needs to reach the lowest intensity for detection, which can be enhanced with the controlled motion of Janus micromotors or graphdiyne tubular catalytic microrobots [41]. The miniaturization of the fluorescence detection device has been achieved, enabling the entire detection process to be completed on site within 5 min. Microrobots made of precious metals, such as gold and silver, are also excellent probes for SERS detection to keep a close contact with analytes and on-demand enrichment, which enhanced the Raman signal approximately three times [42]. In addition, monitoring the locomotion of chemically powered microrobots is an effective mean for directly detecting chemical concentration changes [43]. Functional surface modification enables microrobots to indirectly detect substances by electrochemistry, in which the microrobot movement speeds up the electrochemical mass transfer process to improve the test sensitivity [44–46]. Microrobots can also detect 50 nm nanoplastics via the EIS change after their capture of target objects [47]. Most recently, acoustic microrobots have been applied into the biosensing field due to its high biocompatibility and superfast motility [48].

Since the implementation of microrobots has been conceptually demonstrated to improve the detection sensitivity of biosensors, researchers have therefore vigorously devoted themselves to developing microrobots to assist biosensing in recent years [49–55]. In the following part, we basically introduce the application of microrobots from five detection principles in the field of biosensing and the development of portable sensing.



**Scheme 1.** Schematic diagram for biosensors with microrobots. (SERS = surface-enhanced Raman scattering, EIS = electrochemical impedance spectroscopy).

Table 1. Summarization of typica	l detection princip	les with microro	bots.
----------------------------------	---------------------	------------------	-------

Detection Principle	Type of Microrobots <sup>a</sup>	Target	Propulsion Type	Limit of Detection	Function	Refs
Fluorescence	DNA-modified Au/Pt microrobots	HIV-1 RNA	Chemical	1000 virus particles/mL	Expanding the range of target substances	[56]
	rGO/Pt microrobots	Ricin	Chemical	0.1 ng/mL	Enhance sensitivity	[57]
	rGO/Ni/PtNPs microrobots	Fumonizin B1 Ochratoxin A	Magnetic	0.70 ng/mL 4 ng/mL	Reduce detection time	[58]
	Graphdiyne tubular microrobots	Cholera toxin B	Chemical	1.6 ng/mL	Reduce detection time	[41]
	PEDOT/Pt microrobots	Hg <sup>2+</sup>	Chemical	3 mg/L	Reduce detection time	[59]
	PEDOT/SiO <sub>2</sub> microrobots	Poly(lactic-co-glycolic acid)	Acoustic	-	Enhance sensitivity	[48]
SERS	Body of AgNW@SiO <sub>2</sub> & tail of AgCl microrobots	Crystal violet & MCF-7	Light	-	Enhance sensitivity	[42]
	Au/SiO/Fe microrobots	Rhodamine 6G	Magnetic	0.5 nM	Enhance sensitivity	[60]
Locomotion	GO-wrapped/PtNPs Janus microrobots	Glutathione	Chemical	0.89 µM	Expanding the range of target substances	[43]
EC	Ag-AuNRs microrobots	SARS-CoV-2 virus	Magnetic	1.11 PFU/mL	Enhance sensitivity	[45]
	Ag/Fe <sub>3</sub> O <sub>4</sub> nanorobots	SARS-CoV-2 RNA	Magnetic	6.1 ng/mL	Expanding the range of target substances	[44]
	Mg-Au Janus microrobots	Diphenyl phthalate	Chemical	0.039 mM	Expanding the range of target substances	[46]
EIS	MXene-derived $\gamma$ -Fe <sub>2</sub> O <sub>3</sub> /Pt/TiO <sub>2</sub> microrobots	Nanoplastics	Light	10 <sup>6</sup> nanoplastics/mL	Enhance sensitivity	[47]
	Mg/Fe <sub>3</sub> O <sub>4</sub> /P/anti-E microrobots	MCF-7, MCF-10A & GL261 cells	Chemical	-	Expanding the range of target substances	[61]

<sup>a</sup>  $\mu$ -robots = microrobots; GO = graphene oxide; rGO = reduced graphene oxide; PEDOT = 3,4-ethylenedioxythio phene; NW = nano wire; NR = nano rod; NP = nano particle.

# 3. Biosensors with Microrobots

# 3.1. Microrobots in Fluorescent Biosensing

Fluorescence detection integrated with nanomaterials is a widespread approach to detect and quantify various chemicals and biomarkers [62]. The attraction of fluorescence as an analytical tool lies in the simplicity of detection, where the devices require only a closed light environment and a light detector. Variation in fluorescence intensity and color is achieved by changing the binding state of the probe to the analyte. In divergent detection scenes, various materials and detecting methods are applied in corresponding detection conditions and exhibit different levels for the limit of detection. Among these materials,

graphene oxide (GO) or reduced graphene oxide (rGO) are more attractive because they can absorb the dye-labeled aptamer through  $\pi - \pi$  stacking interactions. The exceptional surface properties of graphene have allowed the attachment of different receptors for toxin detection [63]. Consequently, these receptors enable the capture of nerve agents and heavy metals [64], and then the detection can be accomplished by their fluorescencequenching ability [65–67]. Additionally, one of the most widely applied fluorescence is based on the aptamer, which is taken as an excellent example of functional molecules selected in vitro [68–74]. More importantly, aptamers have high specificity for certain targets, ranging from small molecules to large proteins and even cells, which offers remarkable flexibility and convenience for designing biosensors with high sensitivity and selectivity [75–78]. Compared with antibodies or enzymes, aptamers have higher chemical stability and convenience in the structure design [79-83]. Recent researches revealed the development for designing new fluorescence (bio)-sensing strategies based on artificial microrobots. Alberto Escarpa and coworkers tested two-dimensional (2D) nanomaterials combined with microrobots in fluorescence sensing approaches [84]. As shown in Figure 1A, fluorescencelabeled probes separately wrapped by graphdiyne oxide (GDYO), graphene oxide (GO), or black phosphorous (BP) were connected to Janus microrobots with a 10 µm diameter, which can swim at 40~60 μm/s in human serum. The distinct surface functions of nanomaterials coupled with active navigation play a critical role in the loading/release capacity of the peptide, which is five times higher than the traditional method without microrobots. Such advantages greatly influence the final sensing performance for the Cholera toxin B in Vibrio cholerae and Vibrio parahaemolyticus culture samples. Subjakova et al. presented ultrasound-propelled graphene-oxide-coated gold nanowire motors functionalized with fluorescein-labeled DNA aptamers (FAM-AIB1-apt) for the qualitative detection of overexpressed AIB1 oncoproteins in MCF-7 breast cancer cells (Figure 1B) [70]. The rod-shaped gold nanorobots with an average length of  $\sim$ 1.7  $\mu$ m and 400 nm diameter were driven by an ultrasound field to move at 25  $\mu$ m/s, and the fluorescence intensity for the target protein was greatly enhanced to be four times higher. This proof-of-concept strategy for functional nanorobots has great potential for fluorescence-based sensing methods. Wang and co-workers reported an aptamer-based catalytic microrobot sensing strategy for "Off-On" real-time fluorescent detection of the ricin B toxin (Figure 1C) [57]. This approach relies on self-propelled reduced graphene-oxide (rGO)/platinum (Pt) microrobots with specific ricin B aptamer tagged with a fluorescein-amidine (FAM) dye, whose fluorescence is guenched due to  $\pi - \pi$  interactions with the rGO surface. The microrobot is  $\sim 10 \ \mu m$ long and 5  $\mu$ m in diameter. The influence of the microrobot's motion state on the detection results was also studied. After incubation for three minutes, the fluorescence intensity of the static motor was 1/8 of that of the dynamic motor.

With the development of smartphones, fluorescence detection has been freed from the bondage of laboratory microscopes [10] and has become increasingly convenient for the end-users (Figure 1D) [68]. Integrated with microrobots for real-time fluorescence sensing of (bio)markers, Yuan et al. described the design of a portable device composed of a smartphone coupled with a high-resolution optical lens, custom-made emission filters, and a compartment for the insertion of low-cost commercial lasers to tailor the excitation wavelength [41]. Magnetic Janus microrobots modified with fluorescent ZnS@Cd<sub>x</sub>Se<sub>1-x</sub> quantum dots and graphdiyne tubular catalytic microrobots modified with rhodaminelabeled affinity peptide were, respectively, used for the OFF-ON detection of mercury and cholera toxin B (Figure 1E). Draz et al. presented a platform that integrates cellphone-based optical sensing, loop-mediated isothermal DNA amplification, and microrobot motion for the molecular detection of HIV-1 (Figure 1F) [56]. The microrobots used are platinum nanoparticle (PtNP)-coated spherical polystyrene (PS) beads indirectly engineered with short DNA probes through a middle piece of spherical gold nanoparticle (AuNP). Combination with the smartphone has facilitated the miniaturization and portability of biosensors; nevertheless, there are still some shortcomings of capacities and precision compared with the results obtained in the conventional laboratory. To be specific, the acquired data show

a lack of precision for quantitative detection, which calls for reliable supporting software to identify the picture and reduce the error. In addition, it is necessary to keep the detection environment as consistent as possible to reduce the impact of ambient light on the detection results. Therefore, enabling microrobots to be self-adaptable when performing long-distance enrichment based on reliable control algorithms is highly useful for the reinforcement of fluorescent signals in the next step.



**Figure 1.** Microrobots in fluorescent bio-detection: (**A**) Janus microrobots as dynamic interfaces in the fluorescence sensing of Cholera Toxin B. Reproduced with permission [84]. Copyright 2020, American Chemical Society; (**B**) Ultrasound-propelled microrobots functionalized with fluorescein-labeled DNA aptamers for the qualitative detection of overexpressed AIB1. Reproduced with permission [70]. Copyright 2019, Wiley-VCH; (**C**) Microrobot modified with a specific ricin B aptamer tagged for detection of the ricin B toxin. Reproduced with permission [57]. Copyright 2016, American Chemical Society; (**D**) A flexible smartphone-based colorimetric and fluorescence detection system for PSA detection. Reproduced with permission [68]. Copyright 2015, Elsevier; (**E**) A smartphone-based device integrated with two kinds of microrobots for real-time fluorescence assays. Scale bars, 50 µm. Reproduced with permission [41]. Copyright 2022, the Royal Society of Chemistry; (**F**) A cellphone-based assay for HIV-1 molecular detection using loop-mediated isothermal amplification (LAMP) and microrobots. Reproduced with permission [56]. Copyright 2018, Springer Nature.

## 3.2. Microrobots in Surface-Enhanced Raman Scattering Biosensing

Raman scattering refers to an inelastic light scattering process that provides a vibrational spectrum representing chemical structure information [85–88]. However, the Raman scattering is a weak process, and generally the light intensity is only approximately  $10^{-10}$  of the incident light intensity [89–91]. Surface-enhanced Raman scattering (SERS) is based on the enhancement effects of the rough surface of noble metals, which are  $10^4$ – $10^7$  times stronger than traditional Raman scattering signals [92–95]. SERS is considered to be promising [96,97] as an unlabeled and rapid biosensing technology with high specificity and sensitivity through the SERS-provided vibrational spectrum information [98–101]. Liu et al. reported a surface-enhanced Raman scattering-based lateral flow immunoassay (SERS-LFIA) for the simultaneous detection of anti-SARS-CoV-2 IgM/IgG with high sensitivity [102]. Dual-layer Raman molecule-loaded Ag-coated SiO<sub>2</sub> NPs (SiO<sub>2</sub>@Ag NPs) were used as advanced SERS tags in clinical samples to analyze anti-SARS-CoV-2 IgM/IgG (Figure 2A). The limit of detection (LOD) of SERS-LFIA for anti-SARS-CoV-2 IgM/IgG was estimated to be  $1.28 \times 10^7$ -fold dilution by the International Union of Pure and Applied Chemistry (IUPAC) standard method, which was 800 times higher than that of standard Au nanoparticle-based LFIA for target IgM and IgG.

To improve the SERS performance, microrobots were attempted to be integrated in pioneer studies. The enhanced caption of SERS signal could be acquired by inducing adequate SERS probes in the detection area and maintaining close contact between the probes and analytes [85]. Wang et al. presented an active SERS probe of a light-powered micro/nanomotor (MNM) which has the matchlike  $AgNW@SiO_2$  core-shell structure (Figure 2B) [42]. The maximum speed for this type of micromotor is approximately  $9 \mu m/s$ with  $\sim$ 30 µm length and the AgCl tail. According to micromotor enrichment remotely controlled by external light, both 10<sup>-4</sup> M crystal violet and MCF-7 breast cancer cells were successfully detected with three-times-enhanced Raman signal. Using such a light-induced enrichment of the nanomotors, the Raman signals can be enhanced 6.2 times in a localized detection area in microscale as a supplement to the conventional Raman signal enhancement by SERS. Fan et al. proposed a concept of "motile hotspots" to realize ultrasensitive SERS sensing by combining hotspot engineering and active molecular enrichment. High-density plasmonic nanostructure-supporting hotspots are assembled on the tubular outer wall of microrobots via nanoimprint and rolling origami techniques (Figure 2C) [60]. The hierarchically structured microrobots (HSMs) with nanobowl- and nanocap-arrayed outer walls, respectively, show ~3.5 and ~2.2 times stronger Raman intensity when compared with the microrobot with a smooth outer wall. The size of HSMs by rolling up Au/SiO/Fe nanomembranes is affected by the thickness of nanomembranes. According to the abovementioned methods, either by aggregating the microrobots to the target area or enriching analyte into the microrobots themselves, the improved LOD could be easily achieved.

Meanwhile, portable SERS readers for a lateral flow immunoassay (LFA) were proposed to simplify the essential complicated operations in the laboratory. Li et al. proposed a LFA strip (Figure 2D) based on SERS nanotags for the simultaneous and quantitative detection of dual infection biomarkers, serum amyloid A (SAA) and C-reactive protein (CRP), respectively [103]. Such a biosensing system achieved LODs as low as 0.1 and 0.05 ng/mL, respectively, for SAA and CRP. Tran et al. presented a Raman/SERS-LFA reader that uses a custom-made fiber optic probe for rapid, quantitative, and ultrasensitive POCT [104]. In addition, an integrated portable SERS reader had been designed and built for the rapid scanning of the test strip (Figure 2E). The pregnancy hormone human chorionic gonadotropin (hCG) is detectable in clinical samples within only 2–5 s with a LOD down to approximately 1.6 mIU/mL, which is 15 times more than a commercially available lateral flow assay. Validated by these researches, the SERS-LFA strip-based POCT technology exhibits enormous potential to satisfy the needs for simplicity and convenience in varied medical situations in the near future. Nevertheless, combining microrobots into such technology is rarely explored at this moment, which may have potential implications for POCT, telemedicine, and less-developed areas lacking in medical resources. Even though LFA is a potential development trend for portable devices, we admit that traditional laboratory clinical tests cannot be replaced and still play a critical role for accurate



detection. Consequently, reliable control algorithms to realize the self-adaptable microrobot locomotion are necessary to assure the good test sensitivity for the portable SERS device.

**Figure 2.** Microrobots in SERS bio-detection: **(A)** SERS-LFIA biosensor to detect anti-SARS-CoV-2 IgM/IgG. Reproduced with permission [102]. Copyright 2021, Elsevier; **(B)** Light-powered micro/nanomotor (MNM) that can serve as an active SERS probe. Reproduced with permission [42]. Copyright 2018, Wiley-VCH; **(C)** Magnet-powered tubular microrobots with high-density plasmonic nanostructure-supporting hotspots as motile hotspots for SERS sensing. Reproduced with permission [60]. Copyright 2020, American Chemical Society; **(D)** SERS-LFA strips with a smartphone-based portable Raman spectrometer for the detection of SAA and CRP. Reproduced with permission [103]. Copyright 2021, MDPI; **(E)** Portable SERS reader for rapid scanning of the test strip for detection of hCG. Reproduced with permission [104]. Copyright 2018, Wiley-VCH.

## 3.3. Microrobots in Locomotion-Based Biosensing

The reliance of the microrobot movement on external power sources including both physical fields and chemical fuels makes it possible to design a special category of biosensors based on the relationship between the microrobot moving speed and the change in their surrounding environments [105–107]. Based on the working principle of microrobot motion-based detection, the concentration of the detected substance will affect the microrobot's speed, acceleration, or deceleration [108–110]. Once the linear or non-linear relationship was built, the quantification detection of target reagents could be easily conducted by analyzing the motion of microrobots in real time.

Moreno-Guzman et al. reported a one-millimeter-sized tubular micromotor for mobile biosensing of  $H_2O_2$  in environmental and relevant clinical samples (Figure 3A) [111]. Sodium dodecyl sulfate (SDS) surfactant and horseradish peroxidase were released from the rear of the microrobot which was propelled by the Marangoni effect. In this case, the motion of a single millimeter-sized tubular micromotor for 120 s was measured to quantify the concentration of  $H_2O_2$  in different samples. Similarly, Orozco et al. presented a novel microrobot-based strategy for water-quality testing based on changes in the propulsion behavior of artificial biocatalytic microswimmers influenced by aquatic pollutants (Figure 3B) [112]. The presence of 100  $\mu$ M Hg leads to a rapidly diminished propulsion efficiency with speed diminutions of 90–95% for enzyme-decorated PEDOT/Au millimeter-sized tubular micromotors within 6 min. Zhang et al. fabricated a chemically powered jellyfish-like microrobot by using a multimetallic shell, of which a DNA assembly with catalase decorations was modified on the concave surface to simulate the umbrella-shaped body and the muscle fibers on the inner umbrella of jellyfish (Figure 3C) [113]. The microrobot is a hemispherical shell structure with an opening of 20  $\mu$ m and can swim with speed,

exceeding 209  $\mu$ m/s in 1.5% H<sub>2</sub>O<sub>2</sub>. Once the target DNA hybridizes with the microrobot's



**Figure 3.** Microrobots in locomotion-based bio-detection: (**A**) Millimeter-sized tubular robots for biosensing of H<sub>2</sub>O<sub>2</sub>. Reproduced with permission [111]. Copyright 2015, American Chemical Society; (**B**) Enzyme-powered polymeric (PEDOT)/Au-catalase microrobots for water-quality testing. Reproduced with permission [112]. Copyright 2013, American Chemical Society; (**C**) Chemically powered jellyfish-like microrobots for detection of DNA which slow down the microrobot. Reproduced with permission [113]. Copyright 2021, American Chemical Society; (**D**) A smartphone-based platform for motion-based detection of GSH which reduces the speed of microrobots. Reproduced with permission [43]. Copyright 2018, American Chemical Society; (**E**) Immunological detection of ZIKV using Pt-nanorobots and bead motion monitor of a cellphone optical system. Reproduced with permission [114]. Copyright 2018, American Chemical Society.

Moreover, the optical equipment is essential for observing microrobot movement. Portable devices with a combination of a smartphone and an optical lens show great superiority compared with the traditional lab-used bulky microscopes. Yuan et al. built a Janus microrobot platform for the motion-based detection of glutathione by coupling an ordinary smartphone with an external magnification optical lens. (Figure 3D) [43]. The speed of 20 µm graphene-wrapped/PtNPs Janus microrobots was correlated with the concentration of glutathione, which was utilized to detect the presence of glutathione with a limit of detection (LOD) of 0.90 µM. Draz et al. reported a nanomotor-based bead-motion cellphone (NBC) system for the immunological detection of the Zika virus (ZIKV) [114]. Pt-nanomotors with a  $\sim 20$  nm diameter driven by catalyzing H<sub>2</sub>O<sub>2</sub> could be attached to the beads under the action of ZIKV (Figure 3E). Thus, the average motion velocity of the beads was quantitatively correlated to the virus concentration in the tested sample. The employed smartphone with an optical accessory is also equipped with a customized program to track the bead's movement, measure the moving velocity, and calculate the virus concentration. Obviously, mass production for high-performance microrobots with good locomotion capabilities should be given the first priority to explore for developing new synthesis strategies.

## 3.4. Microrobots in Electrochemical Current-Based Biosensing

Apart from optical methods that rely on the microscope- or smartphone-integrated magnify lenses, electro-signal-based detection offers another option to build portable biosensors independent of the delicate optical system, and thus is highly suitable for incorporating with existing MEMS devices [115–118]. The electrochemical detection reads the current signal generated by the redox reaction of the analyte on the electrode to obtain the concentration of the analyte [51,119]. As one of the most important methods in electroanalytical chemistry, cyclic voltammetry (CV) is often applied in electrochemical current detections [46,120–131]. For example, the electrochemical sensing strategy reported by Sheng et al. for H<sub>2</sub>O<sub>2</sub> concentration measurement followed this basic principle [126], of which the working electrode was modified with Ni-doped Ag@C (Ni/Ag@C) nanocomposites (Figure 4A). The performance of the sensor was studied by cyclic voltammetry measurement of H<sub>2</sub>O<sub>2</sub> to illustrate its LOD down to 0.01 mM.

The addition of microrobots in electrochemical detection accelerates the ambient liquid flow, so as to improve the detection sensitivity and LOD. Rojas et al. presented a novel Janus microrobot-based strategy for the direct determination of diphenyl phthalate (DPP) in food and biological samples. Mg/Au Janus microrobots (average diameter, 20  $\mu$ m) degraded DPP to phenol, which is directly measured by difference pulse voltammetry on disposable screen-printed electrodes (Figure 4B) [46]. Simultaneously, in the absence of microrobots, the electrochemical reaction signal has not been observed. Efficient swimming of multiple Mg/Au micromotors in complex samples introduce a 20-fold increase in the sensitivity of DPP detection compared with that in static conditions. The average speeds of microrobots are 108  $\pm$  18, 296  $\pm$  40, 223  $\pm$  38, and 40  $\pm$  8  $\mu$ m/s in milk, water, whiskey, and serum samples, respectively. Kim et al. showed another plasmonic-magnetic nanorobotbased simple but efficient COVID-19 detection assay through an electronic readout signal (Figure 4C) [44]. With a movement controlled by the magnetic field, nanorobots composed of Fe backbone and Ag surface were used to transport and release the probe nucleic acid. The microrobots with a length of  $2.49 \pm 0.59$  µm, and width of  $1.10 \pm 0.28$  µm, can move at a maximum speed of 8.9  $\mu$ m/s. This strategy has universal adaptability and can be extended to various nucleic acid detections.



**Figure 4.** Microrobots in electrochemical-current bio-detection: (**A**)  $H_2O_2$  sensor modified with the Ni-doped Ag@C (Ni/Ag@C) nanocomposites on the surface of a glassy carbon electrode. Reproduced with permission [126]. Copyright 2017, the Royal Society of Chemistry; (**B**) DPP detection strategy based on Mg/Au Janus micro-robots. Reproduced with permission [46]. Copyright 2016, American Chemical Society; (**C**) Plasmonic-magnetic nanorobot-based COVID-19 detection assay. Reproduced with permission [44]. Copyright 2022, Elsevier; (**D**) Smartphone-based CV system consisting of modified electrodes, a portable electrochemical detector, and a smartphone. Reproduced with permission [123]. Copyright 2017, Elsevier;. (**E**) Mobile phone sensing platform to detect nitrate concentration. Reproduced with permission [128]. Copyright 2015, Elsevier.

Similar to the optical counterparts, electrochemical detection based on smartphones has also been reported. Smartphones serving as data processors and displayers in electrochemical detection have a great potential to combine advanced 5G technologies. Ji et al. designed a smartphone-based CV system for portable detection as shown in Figure 4D. The system consisted of screen-printing modified electrodes, a portable electrochemical detector, and a smartphone [123]. The reduced graphene oxide (rGO) and 3-amino phenylboronic acid (APBA) were modified on the screen-printed electrodes for detection. The LOD for glucose was approximately 0.026 mM with test errors less than 3.8% compared with the commercial electrochemical workstation. Wang et al. proposed a mobile phone sensing platform for nitrate in water by electrochemical measurements using the audio jack function (Figure 4E) [128]. The platform utilized a cyclic-voltammetry-based electrochemical process

with a LOD of 0.2 ppm within 1 min. In this work, the influence of external interference factors on detection stability was eliminated, such as temperature, pH, and ion interference conditions. Compared with commercial electrochemical workstations, portable electrochemical sensors hold an acceptable measuring error with a smaller equipment volume and lower cost. Considering the possibility exhibited by micro/nanorobots in electrochemical detections, diversified modification of individual micro/nanorobot and further exploration to utilize the motility of micro/nanorobots could pave a new way to improve the portability and detection performance of next-generation portable electrochemical biosensors. Under this situation, we envision that it is valuable to clarify an accurate swarm cooperation mechanism to overcome disturbance from the harsh chemical environment.

#### 3.5. Microrobots in Electrochemical Impedance Spectroscopy Biosensing

By means of measuring the intrinsic electrical properties of the target in the electric field, electrochemical impedance spectroscopy (EIS) offers a biocompatible and harmless methodology for detecting various items such as bacteria, biological cells, and tissues, etc., [132–135]. Though EIS has an unmatched advantage in cell identification by measuring the impedance signal corresponding to distinct cell lines in a biocompatible way, excluding toxic reagents, the precision of detection is highly dependent on the location of the cell during the detecting procedure or the data calibration process. Introducing microrobots and their motion control systems can convert the existing unmanageable cell passage into a controllable route and significantly improve the detection performance.

Wan et.al realized an efficient detection of circulating tumor cells (CTCs) through EIS means utilizing Mg-based microrobots (Figure 5A) [61]. Using an aldehyde–amine condensation reaction, the Mg-based microrobots can be modified with Fe<sub>3</sub>O<sub>4</sub>/P/anti-E nanoparticles to capture CTCs. With the hydrogen  $(H_2)$  propulsion by Mg reaction, microrobots maintain 16.5  $\mu$ m/s motion to increase the chance of anti-E recognition and capture of CTCs. The EIS detection platform based on Mg-based microrobots have a good linear response range and low detection limit for CTCs in untreated blood samples (~5 cells/mL). This method was also demonstrated to be effective for detecting oxidized low-density lipoprotein (Ox-LDL) in whole blood just by replacing anti-E components with the antibody of Ox-LDL for microrobots (Figure 5B) [136]. Due to the application of microrobots, the electrochemical sensor shows a good detection for Ox-LDL with range from  $1 \times 10^{-2} \,\mu\text{g/mL}$  to 10  $\mu\text{g/mL}$  and the LOD of  $9.8 \times 10^{-4} \,\mu\text{g/mL}$ . Most recently, the inducement of microrobots offers another choice to aggregate the nanoplastic pollutions in an active manner. Urso et al. developed the multifunctional MXene-derived oxide microrobots that helped to collect randomly distributed nanoplastics in the three-dimensional (3D) space onto the sensing electrodes for EIS detection (Figure 5C) [47]. Compared with the modification of electrodes, well-designed microrobots show not only the capability to trap the target within the whole volume of the liquid sample but also the controllability to locally concentrate the target effectively.

Portable electrical impedance analyzers rely heavily on smartphones for data analysis and display. Zhang et al. developed a smartphone-controlled biosensor system that consisted of a miniaturized biosensor, a hand-held EIS detector, and a smartphone to quantify different kinds of proteins by exchanging electrodes for POCT [137]. The smartphone provides control commands and receives data signals via Bluetooth, meanwhile acting as a displayer for the measurement results in form of a Nyquist plot (Figure 5D). The detection limits of bull serum albumin (BSA, also known as bovine serum albumin) and thrombin are 1.78  $\mu$ g/mL and 2.97 ng/mL, respectively. Jiang et al. presented a mobile health immunoassay platform based on audio-jack-embedded devices, such as smartphones and laptops, which uses EIS to detect a NeutrAvidin self-assembled monolayer [138]. Self-assembled monolayers have been immobilized on gold electrodes. A complementary ligand of the target analyte is modified on the sensor electrode to cause impedance changes when combined with the detecting target (Figure 5E). Regarding the fast development of a wearable and



portable biosensing device, it is essential to carry out auxiliary instrument minimization for driving microrobots for practical implementation.

**Figure 5.** Microrobots in EIS bio-detection: **(A)** A funnel-shaped device to realize the EIS detection of captured CTCs based on Mg-based microrobots. Reproduced with permission [61]. Copyright 2023, Elsevier; **(B)** An EIS sensor platform based on microrobots to actively capture and detect Ox-LDL in whole blood. Reproduced with permission [136]. Copyright 2017, Elsevier; **(C)** Nanoplastics' detection by EIS using self-propelled microrobots for preconcentration in an electrolyte solution (yellow shading). Reproduced with permission [47]. Copyright 2014, Springer Nature; **(D)** Smartphone-controlled biosensor system to detect proteins for POCT, which was constituted by the miniaturized biosensor, hand-held EIS detector, and smartphone. Reproduced with permission [137]. Copyright 2016, Elsevier; **(E)** Mobile health immunoassay platform based on audio-jack-embedded devices in which the sensing electrode is modified with a ligand complimentary to bind analyte. Reproduced with permission [138]. Copyright 2022, Elsevier.

#### 4. Conclusions and Perspective

In this review, we discussed the combination strategies of innovative microrobots with biosensors according to five classic principles, including fluorescence detection, SERS detection, microrobot motion-based detection, electrochemical-current detection, and EIS detection. Well-designed micro/nanorobots assist conventional biosensing systems to adapt to various complex scenarios at higher sensitivity and lower LOD. In addition, the

emergence of microrobots is crucial for indirectly detecting trace-level biological matters hardly detected before. Researchers often design and synthesize microrobots for specific applications based on their research goals. Considering the existing circumstances, it is still a long way to go for commercially available microrobots for users. Despite the fact that the presented merits are fantastic for actively enhancing biosensor performances, there is still plenty of room at the bottom for microrobots to be further explored from the following aspects:

- (i) Development of new synthesis strategy and surface modification to rise the loading capacity. Microrobots serving as active probes for biosensing tests are mostly fabricated with expensive materials and the surface modification is time consuming. Creating microrobots that use inorganic materials should be more efficient for mass production. Solvothermal/hydrothermal methods and template-assisted electrodeposition are adopted to synthesize microrobots just by one-step fabrication. Moreover, a large number of inorganic microrobots are featured with porous structures, which are better suited for rising the loading capacity. Based on the capability of mass production and enough space for surface modification, inorganic microrobots become excellent candidates to be applied in biosensor systems.
- (ii) Accurate swarm cooperation mechanism to overcome disturbance from harsh environments. Due to the high viscosity and large concentration of ions of tested specimens, microrobot individuals are difficult to effectively move because of the weak propulsion force. Based on swarm cooperation, one group of microrobots can aggregate together to output a large propulsion force together to overcome the disturbance. In general, the microrobot swarm powered by ultrasound maintains the highest locomotion speed and thus are primarily considered to be implemented for harsh environments in the biosensor platform. However, the method for accurately controlling the superfast movement of the ultrasonic microrobot swarm is lacking. With the development of metamaterials, unusual physical fields can be generated to modulate the spatial intensity and temporal variation, which can further improve the control accuracy of different microrobot swarms.
- (iii) Auxiliary instrument minimization for driving microrobots. It becomes easy for ordinary people to obtain commercialized biosensors, which are always designed to be portable or wearable for daily use. However, the operation of microrobots requires many auxiliary instruments closely related to the working principles. Most instruments for physically powered microrobots are bulky and expensive, and can only be implemented in the laboratory. With advanced electronic integrated circuit technology, it is possible to shrink these components to a minimized size, which can be well matched with the portable biosensing system. Regarding the costs for commercialization, detachable driving instruments are feasible if the microrobots can be effectively maneuvered to work inside biosensors.
- (iv) Reliable control algorithms to realize the self-adaptable microrobot locomotion. Currently, microrobots can be controlled to perform 2D in-plane movement, but long-distance locomotion along intricate routes is still not yet well demonstrated. For biological applications, multi-functional tests for different properties are sequentially accomplished on the same piece of biosensor with distributed testing regions. The fixed program setting is impossible to control microrobots to move effectively from one region to another in mazy microchannels. An artificial neural network control algorithm is a good option to be integrated with the manipulation system for dynamically optimizing the driving signals, enabling microrobots to be self-adaptable when performing multiple tasks in long-distance locomotion.

Even though many practical troubles of applying microrobots to enhance biosensors is still unsolved and a number of challenging issues should be addressed, we believe that biocompatible, eco-friendly, and dexterous microrobots will become reliable and powerful partners for versatile biosensors to better serve diverse application scenarios in the future. **Author Contributions:** Conceptualization, X.L. and J.B.; methodology, Y.W.; software, S.Z.; validation, X.L., J.B. and W.L.; formal analysis, X.L.; investigation, J.B.; resources, J.B.; data curation, J.B.; writing—original draft preparation, J.B.; writing—review and editing, X.L., W.L. and J.W.; visualization, X.L.; supervision, Y.W.; project administration, J.B.; funding acquisition, X.L., J.B. and Y.W. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by the National Natural Science Foundation of China (Grant No. 51975278 and 52375570), Qing Lan Project, the Biomedical Engineering Fusion Laboratory of the affiliated Jiangning Hospital of Nanjing Medical University (Grant No. JNYYZXKY202217), Postgraduate Research & Practice Innovation Program of Jiangsu Province (No. KYCX23\_0353), and the Fundamental Research Funds for the Central Universities (No. xcxjh20220111 and xcxjh20220114).

**Data Availability Statement:** The authors confirm that the data supporting the findings of this study are available within the article.

Conflicts of Interest: The authors declare no conflict of interest.

#### References

- 1. Rezazadeh, M.; Seidi, S.; Lid, M.; Pedersen-Bjergaard, S.; Yamini, Y. The modern role of smartphones in analytical chemistry. *TrAC Trends Anal. Chem.* **2019**, *118*, 548–555. [CrossRef]
- Olorunyomi, J.F.; Geh, S.T.; Caruso, R.A.; Doherty, C.M. Metal-organic frameworks for chemical sensing devices. *Mater. Horiz.* 2021, *8*, 2387–2419. [CrossRef] [PubMed]
- 3. Noviana, E.; Carrao, D.B.; Pratiwi, R.; Henry, C.S. Emerging applications of paper-based analytical devices for drug analysis: A review. *Anal. Chim. Acta* 2020, 1116, 70–90. [CrossRef] [PubMed]
- Lan, J.; Chen, J.; Li, N.; Ji, X.; Yu, M.; He, Z. Microfluidic generation of magnetic-fluorescent Janus microparticles for biomolecular detection. *Talanta* 2016, 151, 126–131. [CrossRef]
- 5. Liu, Y.; Hao, M.; Chen, Z.; Liu, L.; Liu, Y.; Yang, W.; Ramakrishna, S. A review on recent advances in application of electrospun nanofiber materials as biosensors. *Curr. Opin. Biomed. Eng.* **2020**, *13*, 174–189. [CrossRef]
- 6. Mohankumar, P.; Ajayan, J.; Mohanraj, T.; Yasodharan, R. Recent developments in biosensors for healthcare and biomedical applications: A review. *Measurement* 2021, *167*, 108293. [CrossRef]
- Rocchitta, G.; Spanu, A.; Babudieri, S.; Latte, G.; Madeddu, G.; Galleri, G.; Nuvoli, S.; Bagella, P.; Demartis, M.I.; Fiore, V.; et al. Enzyme Biosensors for Biomedical Applications: Strategies for Safeguarding Analytical Performances in Biological Fluids. *Sensors* 2016, 16, 780. [CrossRef]
- Kholafazad-Kordasht, H.; Hasanzadeh, M.; Seidi, F. Smartphone based immunosensors as next generation of healthcare tools: Technical and analytical overview towards improvement of personalized medicine. *Trac-Trends Anal. Chem.* 2021, 145, 116455. [CrossRef]
- Xiao, M.; Tian, F.; Liu, X.; Zhou, Q.; Pan, J.; Luo, Z.; Yang, M.; Yi, C. Virus Detection: From State-of-the-Art Laboratories to Smartphone-Based Point-of-Care Testing. Adv. Sci. 2022, 9, 2105904. [CrossRef]
- Jafari, S.; Guercetti, J.; Geballa-Koukoula, A.; Tsagkaris, A.S.; Nelis, J.L.D.; Marco, M.P.; Salvador, J.P.; Gerssen, A.; Hajslova, J.; Elliott, C.; et al. ASSURED Point-of-Need Food Safety Screening: A Critical Assessment of Portable Food Analyzers. *Foods* 2021, 10, 1399. [CrossRef]
- 11. Zhao, W.; Tian, S.; Huang, L.; Liu, K.; Dong, L.; Guo, J. A smartphone-based biomedical sensory system. *Analyst* 2020, 145, 2873–2891. [CrossRef]
- 12. Barragan, J.T.C.; Kubota, L.T. Minipotentiostat controlled by smartphone on a micropipette: A versatile, portable, agile and accurate tool for electroanalysis. *Electrochim. Acta* 2020, 341, 136048. [CrossRef]
- Danyliuk, N.; Tatarchuk, T.; Kannan, K.; Shyichuk, A. Optimization of TiO2-P25 photocatalyst dose and H<sub>2</sub>O<sub>2</sub> concentration for advanced photooxidation using the smartphone-based colorimetry. *Water Sci. Technol.* 2021, 84, 469–483. [CrossRef]
- 14. Golicz, K.; Hallett, S.; Sakrabani, R.; Ghosh, J. Adapting smartphone app used in water testing, for soil nutrient analysis. *Comput. Electron. Agric.* **2020**, *175*, 9. [CrossRef]
- Özdemir, G.K.; Bayram, A.; Kiliç, V.; Horzum, N.; Solmaz, M.E. Smartphone-based detection of dyes in water for environmental sustainability. *Anal. Methods* 2017, 9, 579–585. [CrossRef]
- 16. Shogah, Z.A.C.; Bolshakov, D.S.; Amelin, V.G. Using Smartphones in Chemical Analysis. J. Anal. Chem. 2023, 78, 426–449. [CrossRef]
- Salinas, G.; Pavel, I.A.; Sojic, N.; Kuhn, A. Electrochemistry-Based Light-Emitting Mobile Systems. *ChemElectroChem* 2020, 7, 4853–4862. [CrossRef]
- 18. Karshalev, E.; Esteban-Fernandez de Avila, B.; Wang, J. Micromotors for "Chemistry-on-the-Fly". J. Am. Chem. Soc. 2018, 140, 3810–3820. [CrossRef]
- 19. Joshi, S.; Sharma, P.; Siddiqui, R.; Kaushal, K.; Sharma, S.; Verma, G.; Saini, A. A review on peptide functionalized graphene derivatives as nanotools for biosensing. *Microchim. Acta* **2019**, *187*, 27. [CrossRef] [PubMed]

- Campuzano, S.; Kagan, D.; Orozco, J.; Wang, J. Motion-driven sensing and biosensing using electrochemically propelled nanomotors. *Analyst* 2011, 136, 4621–4630. [CrossRef]
- 21. Jurado-Sánchez, B.; Escarpa, A. Milli, micro and nanomotors: Novel analytical tools for real-world applications. *TrAC Trends Anal. Chem.* **2016**, *84*, 48–59. [CrossRef]
- Jurado-Sánchez, B.; Escarpa, A. Janus Micromotors for Electrochemical Sensing and Biosensing Applications: A Review. *Electro*analysis 2017, 29, 14–23. [CrossRef]
- 23. Wang, J. Self-propelled affinity biosensors: Moving the receptor around the sample. *Biosens. Bioelectron.* **2016**, *76*, 234–242. [CrossRef] [PubMed]
- Lu, X.; Shen, H.; Zhao, K.; Wang, Z.; Peng, H.; Liu, W. Micro-/Nanomachines Driven by Ultrasonic Power Sources. *Chem. Asian J.* 2019, 14, 2406–2416. [CrossRef]
- 25. Zhu, C.; Yang, G.; Li, H.; Du, D.; Lin, Y. Electrochemical sensors and biosensors based on nanomaterials and nanostructures. *Anal. Chem.* **2015**, *87*, 230–249. [CrossRef]
- 26. Campuzano, S.; Esteban-Fernandez de Avila, B.; Yanez-Sedeno, P.; Pingarron, J.M.; Wang, J. Nano/microvehicles for efficient delivery and (bio)sensing at the cellular level. *Chem. Sci.* 2017, *8*, 6750–6763. [CrossRef]
- Wu, J.; Balasubramanian, S.; Kagan, D.; Manesh, K.M.; Campuzano, S.; Wang, J. Motion-based DNA detection using catalytic nanomotors. *Nat. Commun.* 2010, 1, 36. [CrossRef]
- Kong, L.; Guan, J.; Pumera, M. Micro- and nanorobots based sensing and biosensing. *Curr. Opin. Electrochem.* 2018, 10, 174–182.
  [CrossRef]
- Lu, X.; Shen, H.; Wei, Y.; Ge, H.; Wang, J.; Peng, H.; Liu, W. Ultrafast Growth and Locomotion of Dandelion-Like Microswarms with Tubular Micromotors. *Small* 2020, 16, 2003678. [CrossRef]
- Lu, X.; Wei, Y.; Ou, H.; Zhao, C.; Shi, L.; Liu, W. Universal Control for Micromotor Swarms with a Hybrid Sonoelectrode. *Small* 2021, 17, 2104516. [CrossRef] [PubMed]
- Lu, X.; Zhao, K.; Liu, W.; Yang, D.; Shen, H.; Peng, H.; Guo, X.; Li, J.; Wang, J. A Human Microrobot Interface Based on Acoustic Manipulation. ACS Nano 2019, 13, 11443–11452. [CrossRef]
- Moo, J.G.; Wang, H.; Zhao, G.; Pumera, M. Biomimetic artificial inorganic enzyme-free self-propelled microfish robot for selective detection of Pb(2+) in water. *Chemistry* 2014, 20, 4292–4296. [CrossRef] [PubMed]
- Bunea, A.I.; Pavel, I.A.; David, S.; Gaspar, S. Sensing based on the motion of enzyme-modified nanorods. *Biosens. Bioelectron*. 2015, 67, 42–48. [CrossRef] [PubMed]
- Li, J.X.; de Avila, B.E.F.; Gao, W.; Zhang, L.F.; Wang, J. Micro/nanorobots for biomedicine: Delivery, surgery, sensing, and detoxification. *Sci. Robot.* 2017, 2, eaam6431. [CrossRef] [PubMed]
- 35. Liu, W.; Chen, X.; Lu, X.; Wang, J.; Zhang, Y.; Gu, Z. From Passive Inorganic Oxides to Active Matters of Micro/Nanomotors. *Adv. Funct. Mater.* **2020**, *30*, 2003195. [CrossRef]
- 36. Soto, F.; Karshalev, E.; Zhang, F.; de Avila, B.E.F.; Nourhani, A.; Wang, J. Smart Materials for Microrobots. *Chem. Rev.* 2022, 122, 5365–5403. [CrossRef]
- Wang, B.; Kostarelos, K.; Nelson, B.J.; Zhang, L. Trends in Micro-/Nanorobotics: Materials Development, Actuation, Localization, and System Integration for Biomedical Applications. *Adv. Mater.* 2021, 33, 2002047. [CrossRef]
- Patra, D.; Sengupta, S.; Duan, W.T.; Zhang, H.; Pavlick, R.; Sen, A. Intelligent, self-powered, drug delivery systems. *Nanoscale* 2013, 5, 1273–1283. [CrossRef]
- Chatzipirpiridis, G.; Ergeneman, O.; Pokki, J.; Ullrich, F.; Fusco, S.; Ortega, J.A.; Sivaraman, K.M.; Nelson, B.J.; Pane, S. Electroforming of Implantable Tubular Magnetic Microrobots for Wireless Ophthalmologic Applications. *Adv. Healthc. Mater.* 2015, 4, 209–214. [CrossRef]
- 40. Kagan, D.; Campuzano, S.; Balasubramanian, S.; Kuralay, F.; Flechsig, G.U.; Wang, J. Functionalized Micromachines for Selective and Rapid Isolation of Nucleic Acid Targets from Complex Samples. *Nano Lett.* **2011**, *11*, 2083–2087. [CrossRef]
- 41. Yuan, K.; de la Asunción-Nadal, V.; Cuntin-Abal, C.; Jurado-Sánchez, B.; Escarpa, A. On-board smartphone micromotor-based fluorescence assays. *Lab Chip* **2022**, *22*, 928–935. [CrossRef] [PubMed]
- Wang, Y.; Zhou, C.; Wang, W.; Xu, D.; Zeng, F.; Zhan, C.; Gu, J.; Li, M.; Zhao, W.; Zhang, J.; et al. Photocatalytically Powered Matchlike Nanomotor for Light-Guided Active SERS Sensing. *Angew. Chem. Int. Ed. Engl.* 2018, 57, 13110–13113. [CrossRef] [PubMed]
- 43. Yuan, K.; Cuntín-Abal, C.; Jurado-Sánchez, B.; Escarpa, A. Smartphone-Based Janus Micromotors Strategy for Motion-Based Detection of Glutathione. *Anal. Chem.* **2021**, *93*, 16385–16392. [CrossRef] [PubMed]
- 44. Kim, J.; Mayorga-Martinez, C.C.; Vyskocil, J.; Ruzek, D.; Pumera, M. Plasmonic-magnetic nanorobots for SARS-CoV-2 RNA detection through electronic readout. *Appl. Mater. Today* 2022, 27, 101402. [CrossRef]
- Mayorga-Martinez, C.C.; Vyskočil, J.; Novotný, F.; Bednar, P.; Ruzek, D.; Alduhaishe, O.; Pumera, M. Collective behavior of magnetic microrobots through immuno-sandwich assay: On-the-fly COVID-19 sensing. *Appl. Mater. Today* 2022, 26, 101337. [CrossRef]
- 46. Rojas, D.; Jurado-Sánchez, B.; Escarpa, A. "Shoot and Sense" Janus Micromotors-Based Strategy for the Simultaneous Degradation and Detection of Persistent Organic Pollutants in Food and Biological Samples. *Anal. Chem.* **2016**, *88*, 4153–4160. [CrossRef]
- 47. Urso, M.; Ussia, M.; Novotny, F.; Pumera, M. Trapping and detecting nanoplastics by MXene-derived oxide microrobots. *Nat. Commun.* **2022**, *13*, 3573. [CrossRef]

- 48. Wei, Y.; Lu, X.L.; Ou, H.; Li, Z.Y.; Liu, Y.L.; Bao, J.H.; Yin, J.; Liu, W.J. Acoustically powered micro-sonobots for enhanced fluorescence biodetection. *Int. J. Mech. Sci.* **2023**, *248*, 11. [CrossRef]
- Hu, Y.; Liu, W.; Sun, Y. Self-Propelled Micro-/Nanomotors as "On-the-Move" Platforms: Cleaners, Sensors, and Reactors. *Adv. Funct. Mater.* 2021, 32, 2109181. [CrossRef]
- Jurado-Sanchez, B.; Campuzano, S.; Pingarron, J.M.; Escarpa, A. Janus particles and motors: Unrivaled devices for mastering (bio)sensing. *Microchim. Acta* 2021, 188, 416. [CrossRef] [PubMed]
- Moo, J.G.S.; Mayorga-Martinez, C.C.; Wang, H.; Khezri, B.; Teo, W.Z.; Pumera, M. Nano/Microrobots Meet Electrochemistry. Adv. Funct. Mater. 2017, 27, 1604759. [CrossRef]
- 52. Oja, S.M.; Fan, Y.; Armstrong, C.M.; Defnet, P.; Zhang, B. Nanoscale Electrochemistry Revisited. *Anal. Chem.* **2016**, *88*, 414–430. [CrossRef] [PubMed]
- 53. Pacheco, M.; Lopez, M.A.; Jurado-Sanchez, B.; Escarpa, A. Self-propelled micromachines for analytical sensing: A critical review. *Anal. Bioanal. Chem.* **2019**, *411*, 6561–6573. [CrossRef]
- 54. Yánez-Sedeño, P.; Campuzano, S.; Pingarrón, J.M. Janus particles for (bio)sensing. Appl. Mater. Today 2017, 9, 276–288. [CrossRef]
- 55. Yi, Y.; Sanchez, L.; Gao, Y.; Yu, Y. Janus particles for biological imaging and sensing. *Analyst* **2016**, 141, 3526–3539. [CrossRef]
- Draz, M.S.; Kochehbyoki, K.M.; Vasan, A.; Battalapalli, D.; Sreeram, A.; Kanakasabapathy, M.K.; Kallakuri, S.; Tsibris, A.; Kuritzkes, D.R.; Shafiee, H. DNA engineered micromotors powered by metal nanoparticles for motion based cellphone diagnostics. *Nat. Commun.* 2018, *9*, 4282. [CrossRef] [PubMed]
- 57. Esteban-Fernández de Ávila, B.; Lopez-Ramirez, M.A.; Báez, D.F.; Jodra, A.; Singh, V.V.; Kaufmann, K.; Wang, J. Aptamer-Modified Graphene-Based Catalytic Micromotors: Off–On Fluorescent Detection of Ricin. *ACS Sens.* **2016**, *1*, 217–221. [CrossRef]
- 58. Molinero-Fernández, Á.; Jodra, A.; Moreno-Guzmán, M.; Lopez, M.Á.; Escarpa, A. Magnetic Reduced Graphene Oxide/Nickel/Platinum Nanoparticles Micromotors for Mycotoxin Analysis. *Chemistry* **2018**, 24, 7172–7176. [CrossRef]
- 59. Jurado-Sánchez, B.; Escarpa, A.; Wang, J. Lighting up micromotors with quantum dots for smart chemical sensing. *Chem. Commun.* **2015**, *51*, 14088–14091. [CrossRef]
- Fan, X.; Hao, Q.; Li, M.; Zhang, X.; Yang, X.; Mei, Y.; Qiu, T. Hotspots on the Move: Active Molecular Enrichment by Hierarchically Structured Micromotors for Ultrasensitive SERS Sensing. ACS Appl. Mater. Interfaces 2020, 12, 28783–28791. [CrossRef]
- 61. Chen, Q.; Guo, W.Y.; Fang, D.; Li, T.; Chen, L.; Mao, C.; Wan, M.M.; Shen, J. Mg-based micromotors for efficient electrochemical detection of circulating tumor cells. *Sens. Actuator B-Chem.* **2023**, *390*, 10. [CrossRef]
- Chen, B.; Huang, Q.L.; Qu, Z.B.; Li, C.; Li, Q.; Shi, J.Y.; Fan, C.H.; Wang, L.H.; Zuo, X.L.; Shen, J.L.; et al. Probing Transient DNA Conformation Changes with an Intercalative Fluorescent Excimer. *Angew. Chem.-Int. Edit.* 2021, 60, 6624–6630. [CrossRef] [PubMed]
- 63. Cheng, H.H.; Hu, C.G.; Zhao, Y.; Qu, L.T. Graphene fiber: A new material platform for unique applications. *Npg Asia Mater.* **2014**, *6*, 13. [CrossRef]
- 64. Shao, Y.Y.; Wang, J.; Engelhard, M.; Wang, C.M.; Lin, Y.H. Facile and controllable electrochemical reduction of graphene oxide and its applications. *J. Mater. Chem.* **2010**, *20*, 743–748. [CrossRef]
- Lu, C.H.; Yang, H.H.; Zhu, C.L.; Chen, X.; Chen, G.N. A Graphene Platform for Sensing Biomolecules. *Angew. Chem. Int. Edit.* 2009, 48, 4785–4787. [CrossRef] [PubMed]
- 66. Morales-Narváez, E.; Merkoçi, A. Graphene Oxide as an Optical Biosensing Platform. Adv. Mater. 2012, 24, 3298–3308. [CrossRef]
- 67. Pérez-López, B.; Merkoçi, A. Carbon nanotubes and graphene in analytical sciences. *Microchim. Acta* 2012, 179, 1–16. [CrossRef]
- Barbosa, A.I.; Gehlot, P.; Sidapra, K.; Edwards, A.D.; Reis, N.M. Portable smartphone quantitation of prostate specific antigen (PSA) in a fluoropolymer microfluidic device. *Biosens. Bioelectron.* 2015, 70, 5–14. [CrossRef]
- 69. Kim, Y.S.; Raston, N.H.; Gu, M.B. Aptamer-based nanobiosensors. *Biosens. Bioelectron.* 2016, 76, 2–19.
- Beltrán-Gasttélum, M.; Esteban-Fernández de Ávila, B.; Gong, H.; Venugopalan, P.L.; Hianik, T.; Wang, J.; Subjakova, V. Rapid Detection of AIB1 in Breast Cancer Cells Based on Aptamer-Functionalized Nanomotors. *ChemPhysChem* 2019, 20, 3177–3180. [CrossRef]
- de Ávila, B.E.F.; Martin, A.; Soto, F.; Lopez-Ramirez, M.A.; Campuzano, S.; Vasquez-Machado, G.M.; Gao, W.W.; Zhang, L.F.; Wang, J. Single Cell Real-Time miRNAs Sensing Based on Nanomotors. ACS Nano 2015, 9, 6756–6764. [CrossRef]
- Khoshbin, Z.; Zamanian, J.; Davoodian, N.; Mohammad Danesh, N.; Ramezani, M.; Alibolandi, M.; Abnous, K.; Taghdisi, S.M. A simple and ultrasensitive metal-organic framework-based aptasensor for fluorescence detection of ethanolamine. *Spectrochim. Acta Part A* 2022, 267, 120488. [CrossRef] [PubMed]
- 73. Pacheco, M.; Jurado-Sánchez, B.; Escarpa, A. Sensitive Monitoring of Enterobacterial Contamination of Food Using Self-Propelled Janus Microsensors. *Anal. Chem.* 2018, *90*, 2912–2917. [CrossRef]
- Molinero-Fernández, Á.; Moreno-Guzmán, M.; López, M.Á.; Escarpa, A. Biosensing Strategy for Simultaneous and Accurate Quantitative Analysis of Mycotoxins in Food Samples Using Unmodified Graphene Micromotors. *Anal. Chem.* 2017, *89*, 10850–10857. [CrossRef]
- 75. Song, S.; Wang, L.; Li, J.; Fan, C.; Zhao, J. Aptamer-based biosensors. TrAC Trends Anal. Chem. 2008, 27, 108–117. [CrossRef]
- Zhu, C.; Zeng, Z.; Li, H.; Li, F.; Fan, C.; Zhang, H. Single-Layer MoS2-Based Nanoprobes for Homogeneous Detection of Biomolecules. J. Am. Chem. Soc. 2013, 135, 5998–6001. [CrossRef]
- Kou, H.-S.; Lo, S.-T.; Wang, C.-C. One Single Tube Reaction of Aptasensor-Based Magnetic Sensing System for Selective Fluorescent Detection of VEGF in Plasma. *Biosensors* 2023, 13, 574. [CrossRef]

- 78. Azzouz, A.; Kumar, V.; Hejji, L.; Kim, K.-H. Advancements in nanomaterial-based aptasensors for the detection of emerging organic pollutants in environmental and biological samples. *Biotechnol. Adv.* **2023**, *66*, 108156. [CrossRef] [PubMed]
- 79. Li, Y.; Liu, L.; Qiao, L.; Deng, F. Universal CRISPR/Cas12a-associated aptasensor suitable for rapid detection of small proteins with a plate reader. *Front. Bioeng. Biotechnol.* **2023**, *11*, 1201175. [CrossRef]
- 80. Song, P.; Fan, C. Selecting aptamers with programmed affinities. Nat. Chem. 2023, 15, 747–748. [CrossRef] [PubMed]
- Kara, N.; Ayoub, N.; Ilgu, H.; Fotiadis, D.; Ilgu, M. Aptamers Targeting Membrane Proteins for Sensor and Diagnostic Applications. Molecules 2023, 28, 3728. [CrossRef]
- Mahmud, A.; Chang, D.; Das, J.; Gomis, S.; Foroutan, F.; Chen, J.B.; Pandey, L.; Flynn, C.D.; Yousefi, H.; Geraili, A.; et al. Monitoring Cardiac Biomarkers with Aptamer-Based Molecular Pendulum Sensors. *Angew. Chem. Int. Edit.* 2023, 62, e202213567. [CrossRef] [PubMed]
- Kong, F.; Luo, J.; Jing, L.; Wang, Y.; Shen, H.; Yu, R.; Sun, S.; Xing, Y.; Ming, T.; Liu, M.; et al. Reduced Graphene Oxide and Gold Nanoparticles-Modified Electrochemical Aptasensor for Highly Sensitive Detection of Doxorubicin. *Nanomaterials* 2023, 13, 1223. [CrossRef] [PubMed]
- Yuan, K.; López, M.A.; Jurado-Sánchez, B.; Escarpa, A. Janus Micromotors Coated with 2D Nanomaterials as Dynamic Interfaces for (Bio)-Sensing. ACS Appl. Mater. Interfaces 2020, 12, 46588–46597. [CrossRef]
- Cardinal, M.F.; Vander Ende, E.; Hackler, R.A.; McAnally, M.O.; Stair, P.C.; Schatz, G.C.; Van Duyne, R.P. Expanding applications of SERS through versatile nanomaterials engineering. *Chem. Soc. Rev.* 2017, 46, 3886–3903. [CrossRef] [PubMed]
- 86. Duan, B.; Hou, S.; Wang, P.; Chen, Y.; Xiong, Q.; Das, P.; Duan, H. Chemical processing of interfacially assembled metal nanowires for surface-enhanced Raman scattering detection of food contaminants. *J. Raman Spectrosc.* **2020**, *52*, 532–540. [CrossRef]
- 87. Gahlaut, S.K.; Savargaonkar, D.; Sharan, C.; Yadav, S.; Mishra, P.; Singh, J.P. SERS Platform for Dengue Diagnosis from Clinical Samples Employing a Hand Held Raman Spectrometer. *Anal. Chem.* **2020**, *92*, 2527–2534. [CrossRef]
- Li, Z.; Zhang, L.; He, X.; Bensong, C. Urchin-like ZnO-nanorod arrays templated growth of ordered hierarchical Ag/ZnO hybrid arrays for surface-enhanced Raman scattering. *Nanotechnology* 2020, 31, 165301. [CrossRef]
- 89. Feng, E.; Tian, Y. Surface-enhanced Raman Scattering of Self-assembled Superstructures. *Chem. Res. Chin. Univ.* 2021, 37, 989–1007. [CrossRef]
- García-Lojo, D.; Gómez-Graña, S.; Martin, V.F.; Solís, D.M.; Taboada, J.M.; Pérez-Juste, J.; Pastoriza-Santos, I. Integrating Plasmonic Supercrystals in Microfluidics for Ultrasensitive, Label-Free, and Selective Surface-Enhanced Raman Spectroscopy Detection. ACS Appl. Mater. Interfaces 2020, 12, 46557–46564. [CrossRef]
- 91. Wang, Y.; Liu, Y.; Li, Y.; Xu, D.; Pan, X.; Chen, Y.; Zhou, D.; Wang, B.; Feng, H.; Ma, X. Magnetic Nanomotor-Based Maneuverable SERS Probe. *Research* 2020, 2020, 7962024. [CrossRef]
- Jia, X.; Liu, Z.; Peng, Y.; Hou, G.; Chen, W.; Xiao, R. Automatic and sensitive detection of West Nile virus non-structural protein 1 with a portable SERS-LFIA detector. *Microchim. Acta* 2021, 188, 206. [CrossRef] [PubMed]
- Liu, J.; Liu, Z.; Wang, W.; Tian, Y. Real-time Tracking and Sensing of Cu(+) and Cu(2+) with a Single SERS Probe in the Live Brain: Toward Understanding Why Copper Ions Were Increased upon Ischemia. *Angew. Chem. Int. Ed. Engl.* 2021, 60, 21351–21359. [CrossRef] [PubMed]
- Liu, K.; Bai, Y.C.; Zhang, L.; Yang, Z.B.; Fan, Q.K.; Zheng, H.Q.; Yin, Y.D.; Gao, C.B. Porous Au-Ag Nanospheres with High-Density and Highly Accessible Hotspots for SERS Analysis. *Nano Lett.* 2016, 16, 3675–3681. [CrossRef]
- Liu, X.; Yang, S.; Lyu, X.; Liu, S.; Wang, Y.; Li, Y.; Wang, B.; Chen, W.; Wang, W.; Guo, J.; et al. Instant Preparation of Ultraclean Gold Nanothorns under Ambient Conditions for SERS Kit-Enabled Mobile Diagnosis. *Anal. Chem.* 2021, 93, 16628–16637. [CrossRef]
- Qian, X.M.; Nie, S.M. Single-molecule and single-nanoparticle SERS: From fundamental mechanisms to biomedical applications. *Chem. Soc. Rev.* 2008, 37, 912–920. [CrossRef]
- 97. Wang, X.; Li, J.; Shen, Y.; Xie, A. An assembled ordered W18O49 nanowire film with high SERS sensitivity and stability for the detection of RB. *Appl. Surf. Sci.* 2020, 504, 144073. [CrossRef]
- Lu, Y.; Lin, L.; Ye, J. Human metabolite detection by surface-enhanced Raman spectroscopy. *Mater. Today Bio* 2022, 13, 100205. [CrossRef]
- Shan, F.; Zhang, X.Y.; Fu, X.C.; Zhang, L.J.; Su, D.; Wang, S.J.; Wu, J.Y.; Zhang, T. Investigation of simultaneously existed Raman scattering enhancement and inhibiting fluorescence using surface modified gold nanostars as SERS probes. *Sci. Rep.* 2017, 7, 6813. [CrossRef]
- Srivastav, S.; Dankov, A.; Adanalic, M.; Grzeschik, R.; Tran, V.; Pagel-Wieder, S.; Gessler, F.; Spreitzer, I.; Scholz, T.; Schnierle, B.; et al. Rapid and Sensitive SERS-Based Lateral Flow Test for SARS-CoV2-Specific IgM/IgG Antibodies. *Anal. Chem.* 2021, 93, 12391–12399. [CrossRef]
- Wang, W.; Li, Y.; Nie, A.; Fan, G.C.; Han, H. A portable SERS reader coupled with catalytic hairpin assembly for sensitive microRNA-21 lateral flow sensing. *Analyst* 2021, 146, 848–854. [CrossRef] [PubMed]
- 102. Liu, H.; Dai, E.; Xiao, R.; Zhou, Z.; Zhang, M.; Bai, Z.; Shao, Y.; Qi, K.; Tu, J.; Wang, C.; et al. Development of a SERS-based lateral flow immunoassay for rapid and ultra-sensitive detection of anti-SARS-CoV-2 IgM/IgG in clinical samples. *Sens. Actuators B* 2021, 329, 129196. [CrossRef]
- Li, Y.; Liu, X.; Guo, J.; Zhang, Y.; Guo, J.; Wu, X.; Wang, B.; Ma, X. Simultaneous Detection of Inflammatory Biomarkers by SERS Nanotag-Based Lateral Flow Assay with Portable Cloud Raman Spectrometer. *Nanomaterials* 2021, 11, 1496. [CrossRef] [PubMed]

- 104. Tran, V.; Walkenfort, B.; König, M.; Salehi, M.; Schlücker, S. Rapid, Quantitative, and Ultrasensitive Point-of-Care Testing: A Portable SERS Reader for Lateral Flow Assays in Clinical Chemistry. *Angew. Chem. Int. Ed. Engl.* 2019, 58, 442–446. [CrossRef]
- 105. Zheng, Y.; Zhao, H.; Cai, Y.; Jurado-Sanchez, B.; Dong, R. Recent Advances in One-Dimensional Micro/Nanomotors: Fabrication, Propulsion and Application. *Nano-Micro Lett.* **2023**, *15*, 20. [CrossRef]
- Thome, C.P.; Hoertdoerfer, W.S.; Bendorf, J.R.; Lee, J.G.; Shields, C.W. Electrokinetic Active Particles for Motion-Based Biomolecule Detection. *Nano Lett.* 2023, 23, 2379–2387. [CrossRef] [PubMed]
- 107. Cai, L.; Xu, D.; Zhang, Z.; Li, N.; Zhao, Y. Tailoring Functional Micromotors for Sensing. Research 2023, 6, 44. [CrossRef] [PubMed]
- 108. Popescu, M.N.; Gaspar, S. Analyte Sensing with Catalytic Micromotors. Biosensors 2023, 13, 45. [CrossRef] [PubMed]
- Wang, Y.; Zhao, P.; Zhang, S.; Zhu, K.; Shangguan, X.; Liu, L.; Zhang, S. Application of Janus Particles in Point-of-Care Testing. Biosensors 2022, 12, 689. [CrossRef]
- 110. Chang, S.H. Micro/nanomotors for metal ion detection and removal from water: A review. *Mater. Today Sustain.* **2022**, *19*, 100196. [CrossRef]
- Moreno-Guzman, M.; Jodra, A.; López, M.A.; Escarpa, A. Self-Propelled Enzyme-Based Motors for Smart Mobile Electrochemical and Optical Biosensing. *Anal. Chem.* 2015, 87, 12380–12386. [CrossRef]
- Orozco, J.; Garcia-Gradilla, V.; D'Agostino, M.; Gao, W.; Cortés, A.; Wang, J. Artificial Enzyme-Powered Microfish for Water-Quality Testing. ACS Nano 2013, 7, 818–824. [CrossRef] [PubMed]
- Zhang, X.; Chen, C.; Wu, J.; Ju, H. Bubble-Propelled Jellyfish-like Micromotors for DNA Sensing. ACS Appl. Mater. Interfaces 2019, 11, 13581–13588. [CrossRef] [PubMed]
- 114. Draz, M.S.; Lakshminaraasimulu, N.K.; Krishnakumar, S.; Battalapalli, D.; Vasan, A.; Kanakasabapathy, M.K.; Sreeram, A.; Kallakuri, S.; Thirumalaraju, P.; Li, Y.; et al. Motion-Based Immunological Detection of Zika Virus Using Pt-Nanomotors and a Cellphone. ACS Nano 2018, 12, 5709–5718. [CrossRef] [PubMed]
- 115. Ferreira, P.C.; Ataide, V.N.; Silva Chagas, C.L.; Angnes, L.; Tomazelli Coltro, W.K.; Longo Cesar Paixao, T.R.; de Araujo, W.R. Wearable electrochemical sensors for forensic and clinical applications. *Trac-Trends Anal. Chem.* **2019**, *119*, 115622. [CrossRef]
- Naghdi, T.; Ardalan, S.; Adib, Z.A.; Sharifi, A.R.; Golmohammadi, H. Moving toward smart biomedical sensing. *Biosens. Bioelectron.* 2023, 223, 115009. [CrossRef]
- 117. Madrid, R.E.; Ramallo, F.A.; Barraza, D.E.; Chaile, R.E. Smartphone-Based Biosensor Devices for Healthcare: Technologies, Trends, and Adoption by End-Users. *Bioengineering* **2022**, *9*, 101. [CrossRef]
- 118. Lyu, Q.; Gong, S.; Dyson, J.M.; Cheng, W. Soft, Disruptive and Wearable Electrochemical Biosensors. *Curr. Anal. Chem.* **2022**, *18*, 689–704. [CrossRef]
- 119. Sinha, G.N.; Subramanyam, P.; Sivaramakrishna, V.; Subrahmanyam, C. Electrodeposited copper bismuth oxide as a low-cost, non-enzymatic electrochemical sensor for sensitive detection of uric acid and hydrogen peroxide. *Inorg. Chem. Commun.* 2021, 129, 108627. [CrossRef]
- 120. Abellán-Llobregat, A.; Jeerapan, I.; Bandodkar, A.; Vidal, L.; Canals, A.; Wang, J.; Morallón, E. A stretchable and screen-printed electrochemical sensor for glucose determination in human perspiration. *Biosens. Bioelectron.* **2017**, *91*, 885–891. [CrossRef]
- Chia, H.L.; Mayorga-Martinez, C.C.; Gusmao, R.; Novotny, F.; Webster, R.D.; Pumera, M. A highly sensitive enzyme-less glucose sensor based on pnictogens and silver shell-gold core nanorod composites. *Chem. Commun.* 2020, 56, 7909–7912. [CrossRef]
- 122. Jain, U.; Chauhan, N. Glycated hemoglobin detection with electrochemical sensing amplified by gold nanoparticles embedded N-doped graphene nanosheet. *Biosens. Bioelectron.* **2017**, *89*, 578–584. [CrossRef]
- 123. Ji, D.; Liu, L.; Li, S.; Chen, C.; Lu, Y.; Wu, J.; Liu, Q. Smartphone-based cyclic voltammetry system with graphene modified screen printed electrodes for glucose detection. *Biosens. Bioelectron.* **2017**, *98*, 449–456. [CrossRef]
- Redondo, E.; Pumera, M. Fully metallic copper 3D-printed electrodes via sintering for electrocatalytic biosensing. *Appl. Mater. Today* 2021, 25, 101253. [CrossRef]
- 125. Shen, X.; Ju, F.; Li, G.; Ma, L. Smartphone-Based Electrochemical Potentiostat Detection System Using PEDOT: PSS/Chitosan/ Graphene Modified Screen-Printed Electrodes for Dopamine Detection. Sensors 2020, 20, 2781. [CrossRef] [PubMed]
- Sheng, Q.; Shen, Y.; Zhang, J.; Zheng, J. Ni doped Ag@C core–shell nanomaterials and their application in electrochemical H<sub>2</sub>O<sub>2</sub> sensing. *Anal. Methods* 2017, 9, 163–169. [CrossRef]
- 127. Shi, L.; Layani, M.; Cai, X.; Zhao, H.; Magdassi, S.; Lan, M. An inkjet printed Ag electrode fabricated on plastic substrate with a chemical sintering approach for the electrochemical sensing of hydrogen peroxide. *Sens. Actuators B* 2018, 256, 938–945. [CrossRef]
- 128. Wang, X.; Gartia, M.R.; Jiang, J.; Chang, T.-W.; Qian, J.; Liu, Y.; Liu, X.; Liu, G.L. Audio jack based miniaturized mobile phone electrochemical sensing platform. *Sens. Actuators B* 2015, 209, 677–685. [CrossRef]
- 129. Xu, Y.; Dai, Y.; Li, C.; Zhang, H.; Guo, M.; Yang, Y. PC software-based portable cyclic voltammetry system with PB-MCNT-GNPsmodified electrodes for E. coli detection. *Rev. Sci. Instrum.* **2020**, *91*, 014103. [CrossRef]
- Yusoff, N.; Rameshkumar, P.; Mehmood, M.S.; Pandikumar, A.; Lee, H.W.; Huang, N.M. Ternary nanohybrid of reduced graphene oxide-nafion@silver nanoparticles for boosting the sensor performance in non-enzymatic amperometric detection of hydrogen peroxide. *Biosens. Bioelectron.* 2017, *87*, 1020–1028. [CrossRef] [PubMed]
- Zhang, W.; Tian, Z.; Yang, S.; Rich, J.; Zhao, S.; Klingeborn, M.; Huang, P.H.; Li, Z.; Stout, A.; Murphy, Q.; et al. Electrochemical micro-aptasensors for exosome detection based on hybridization chain reaction amplification. *Microsyst. Nanoeng.* 2021, 7, 63. [CrossRef] [PubMed]

- 132. Amouzadeh Tabrizi, M.; Shamsipur, M.; Saber, R.; Sarkar, S. Isolation of HL-60 cancer cells from the human serum sample using MnO2-PEI/Ni/Au/aptamer as a novel nanomotor and electrochemical determination of thereof by aptamer/gold nanoparticlespoly(3,4-ethylene dioxythiophene) modified GC electrode. *Biosens. Bioelectron.* 2018, 110, 141–146. [CrossRef] [PubMed]
- 133. Grossi, M.; Parolin, C.; Vitali, B.; Riccò, B. Electrical Impedance Spectroscopy (EIS) characterization of saline solutions with a low-cost portable measurement system. *Eng. Sci. Technol.* **2019**, *22*, 102–108. [CrossRef]
- Jiang, J.; Wang, X.; Chao, R.; Ren, Y.; Hu, C.; Xu, Z.; Liu, G.L. Smartphone based portable bacteria pre-concentrating microfluidic sensor and impedance sensing system. *Sens. Actuators B* 2014, 193, 653–659. [CrossRef]
- Zhu, S.; Zhang, X.; Zhou, Z.; Han, Y.; Xiang, N.; Ni, Z. Microfluidic impedance cytometry for single-cell sensing: Review on electrode configurations. *Talanta* 2021, 233, 122571. [CrossRef]
- 136. Fang, D.; Tang, S.W.; Wu, Z.Y.; Chen, C.L.; Wan, M.M.; Mao, C.; Zhou, M. Electrochemical sensor based on micromotor technology for detection of Ox-LDL in whole blood. *Biosens. Bioelectron.* 2022, 217, 6. [CrossRef]
- 137. Zhang, D.; Lu, Y.; Zhang, Q.; Liu, L.; Li, S.; Yao, Y.; Jiang, J.; Liu, G.L.; Liu, Q. Protein detecting with smartphone-controlled electrochemical impedance spectroscopy for point-of-care applications. *Sens. Actuators B* 2016, 222, 994–1002. [CrossRef]
- Jiang, H.; Sun, A.; Venkatesh, A.G.; Hall, D.A. An Audio Jack-Based Electrochemical Impedance Spectroscopy Sensor for Point-of-Care Diagnostics. *IEEE Sens. J.* 2017, 17, 589–597. [CrossRef]

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.