

Supplementary Materials

Synthesis of Fe-Doped Peroxidase Mimetic Nanozymes from Natural Hemoglobin for Colorimetric Biosensing and In Vitro Anticancer Effects

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Materials and Methods:

Instruments

The elemental composition of BDNPs was identified by Energy-dispersive X-ray spectroscopy (EDS) analysis (MIRA3TESCAN-XMU). A FEI Tecnai G2 F20 SuperTwin (accelerating voltage: 200 kV) transmission electron microscope (TEM) was used to evaluate the morphology of the BDNPs. An ELx800™ microplate absorbance reader (BioTek Instruments) and a UV-Vis spectrophotometer (Biowave II) were used for acquiring the UV-Vis spectroscopic data. Synthesis of blood-derived BDNPs was accomplished in an AZAR furnace with controllable temperature. Hemolysis of red blood cells (RBCs) was done using a Qsonica probe sonicator (Q700). The Fourier transform infrared (FTIR) spectra were measured with a PerkinElmer Spectrum 65 spectrometer over the spectral range of 450–4000 cm^{-1} . Fluorescence measurements were carried out by a RF-6000 spectrofluorometer. The Fe content of BDNPs was measured by ICP-MS (Perkin Elmer). The H_2O_2 concentration of the stock solution was determined based on its extinction coefficient of 43.6 $\text{M}^{-1} \text{cm}^{-1}$ at 240 nm [1-3].

Extraction of Hb from human whole blood

5 mL of a blood biowaste from healthy volunteer was obtained from the Iranian Blood Transfusion Organization. The RBCs were isolated by gradient centrifugation using Ficoll-Hypaque 1077 (GE health care-Sweden). Hb extraction was performed according to a reported procedure[4]. Briefly, the isolated RBCs were washed with an equal weight of isotonic saline solution (0.9% NaCl, w/v) and centrifuged at 1000g for 30 min to remove the remaining human serum plasma. The supernatant was discarded. The RBCs were redispersed in 0.01M NaCl, considering a 1:2 (w/w) ratio of RBC/NaCl solution. Hemolysis was carried out with a probe sonicator equipped with a 13 mm diameter probe (amplitude 20%, 10 min, pulse-on time 1s, pulse-off time 2s). After breaking down the RBCs by ultrasonication, the suspension of lysed cells was heated at 60 °C for 1 h in a water bath in the dark, and centrifuged at 2000g for 1 h. The precipitate was discarded and the clear dark red supernatant was used for the synthesis experiments.

Cell culture

Human breast cancer cell line (MCF-7) and human umbilical vein endothelial cells (HUVEC) were provided by the National Cell Bank of Iran and cultivated in Dulbecco's Modified Eagle Medium (DMEM high-glucose) with 10% FBS and 1% penicillin-streptomycin. The cells were incubated at 37°C with 5% CO_2 . The cells were then re-suspended with 0.25% Trypsin-EDTA and tests were performed on cells at 80% confluence between passages 3 and 6.

3D cell spheroid formation

For the spheroid formation and 3D studies, we selected MCF-7 cells to mimic a microtumor environment. Extremely high density in a hanging drop was favored for the production of spheroids and to provide the cells with an in vivo milieu. The cells were cultured in the form of 20 μL drops on the inverted lid of a 35 mm petri dish with Ca^{+2} - and Mg^{+2} -free HBSS buffer supplemented with streptomycin (100 $\mu\text{g/mL}$), gentamycin (50 $\mu\text{g/mL}$), and penicillin 100 (U/mL).

MTT assay

MCF-7 and HUVEC cell lines were seeded into 96 well plates (9000 cells per well) and incubated overnight in the monolayer cell culture environment. In the culture media, a stock BDNP-100 stock solution (1mg/mL) was prepared, and diluted solutions were replaced with culture medium. Control wells only contained culture media. Cell proliferation and viability were evaluated using the MTT test to investigate the cytotoxic effects of BDNP-100 on MCF-7 and HUVEC cells. After 24 hours of treatment, the BDNP-containing culture media

was withdrawn, and the wells were treated with 50 μ M H₂O₂ for 6 hours. The cells were then subjected to 50 μ L MTT reagent (0.5 mg/mL) for 3 hours at 37 °C. The insoluble formazan crystals were then dissolved in 150 μ L DMSO. After 30 minutes, absorbance values at 570 and 630 nm were measured using a microplate reader (ELx800 TM, Biotech, Winooski, VT).

Apoptosis test

MCF-7 cells (in monolayer and cell spheroid conditions) and controls were seeded separately into 6cm plates at a density of 2×10^5 cells/dish. 12 hours after adhesion, cells were treated with low (12.5 μ g/mL) and high (100 μ g/mL) doses of BDNP-100 for 24 hours. In the following, cells were treated with H₂O₂ (50 μ M) for 6 hours. To identify apoptotic cells, an Annexin V-FITC/propidium iodide (PI) apoptosis test was employed. Cells were trypsinized, washed twice with PBS, and suspended in 100 μ L of 1X Annexin V binding solution. Following that, 5 μ L of Annexin V-FITC and PI staining solutions were added to the cell suspension, which was then incubated in the dark at room temperature for 15 minutes. Apoptosis was identified using flow cytometry (BD FACS Calibur (BD biosciences, San Jose, CA, USA)) and analyzed with FlowJo after the addition of 400 μ L 1X Annexin V binding solution. The apoptotic cell percentage was measured as Annexin V-positive cells/total cell number.

ROS assay

Flow cytometry was used to detect intracellular ROS inside the cancer cells. MCF-7 cells (in monolayer and cell spheroid condition) and their controls were treated with low (12.5 μ g/mL) and high (100 μ g/mL) doses of BDNP-100 for 24 hours. In the following, cells were treated with H₂O₂ (50 μ M) for 6 hours. MCF-7 cells were collected and treated with 10 μ M 2',7'-dichlorofluorescein diacetate (DCFH-DA) dye for 20 minutes at 37 °C. Flow cytometry was used to assess the fluorescence intensity of each sample and FlowJo software was used to analyze the data.

Cell Cycle

MCF-7 cells (monolayer and cell spheroids) and their controls were treated with low (12.5 μ g/mL) and high (100 μ g/mL) doses of BDNP-100 for 24 hours. In the following, cells were treated with H₂O₂ (50 μ M) for 6 hours. Then, cells were collected and fixed overnight at 4 °C in 70% ethanol. After removing the fixative solution by centrifugation at 200g for 5 minutes at room temperature, cell pellets were washed twice with ice-cold PBS and treated with RNase A (10 g/mL; cat. no. ST579; Beyotime Institute of Biotechnology) and PI (final concentration, 50 g/mL) for 30 minutes at room temperature in the dark. PI-stained cells were analyzed for cell cycle phase distribution using flow cytometry within 1 hour. FlowJo v10 was used to examine cell phase dispersion.

UV-Vis and PL of BDNP samples:

The spectral features of the extracted Hb coincide well with the spectral pattern of pure Hb in the literature (**Fig. S4**). The UV-Vis absorption spectrum of BDNPs obtained from hydrothermally-treated Hb at 100 °C (BDNP-100), shows a sharp absorption peak at around 400 nm, which is consistent with the Soret band of native Hb. Except that, none of the peaks that originally exist in the UV-Vis absorption spectrum of Hb are seen in the UV-Vis spectrum of BDNP-100.

It is possible that chemical transformation of Hb upon thermal treatment of the sample at 100 °C has proceeded partially. By increasing the synthesis temperature to 125 °C and 150 °C, the peak intensity at 400 nm decreases and disappears for BDNP-180. The spectral profile of the particles synthesized at 180 °C is close to the commonly observed absorption spectrum of carbon nanodots [5].

Under the UV light excitation at 365 nm, we observed that samples prepared at 150 °C (BDNP-150) and 180 °C (BDNP-180) glow while the other two (BDNP-100 and BDNP-125) did not exhibit any fluorescence signal that could be detected by the naked eye. To evaluate and compare the emission profiles, all samples were excited by two wavelengths (250 nm and 300 nm), and the corresponding emission spectra were recorded by a spectrofluorometer. As seen in **Fig. S4b, c**, native Hb exhibits one emission peak at 300-350 nm and another weaker peak at around 450 nm. Regardless of the excitation wavelength, the emission profiles of BDNP-100 and BDNP-125 are similar to Hb. In contrast, BDNP-150 and BDNP-180 emit a blue light upon UV excitation (**Fig. S4a**, inset). Accordingly, a single emission peak at 400-420 nm is observed for BDNP-150, BDNP-180, similar to the emission profile of carbon nanodots in the previous reports. Based on the UV-Vis and fluorescence emission data, we inspect that at low temperatures (100 and 125 °C), chemical transformation of Hb is incomplete, while samples treated at 150 and 180 °C carbonize into fluorescent particles.

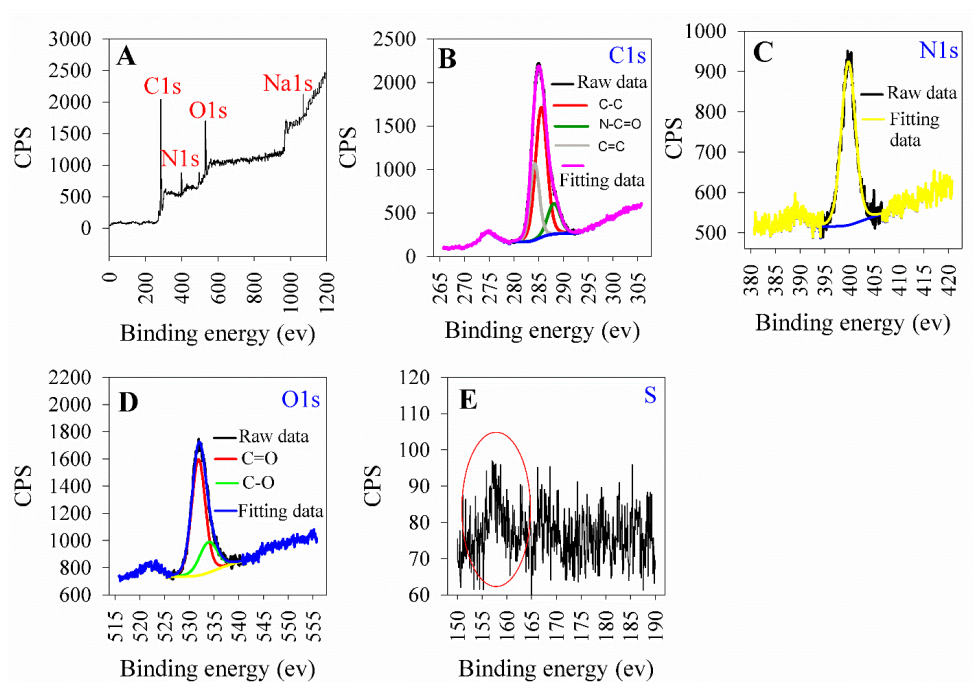


Figure S1. (A) XPS survey spectrum and high resolution (B) C1s, (C) N1s, and (D) O1s, spectra of BDNP-100. (E) sulfur region.

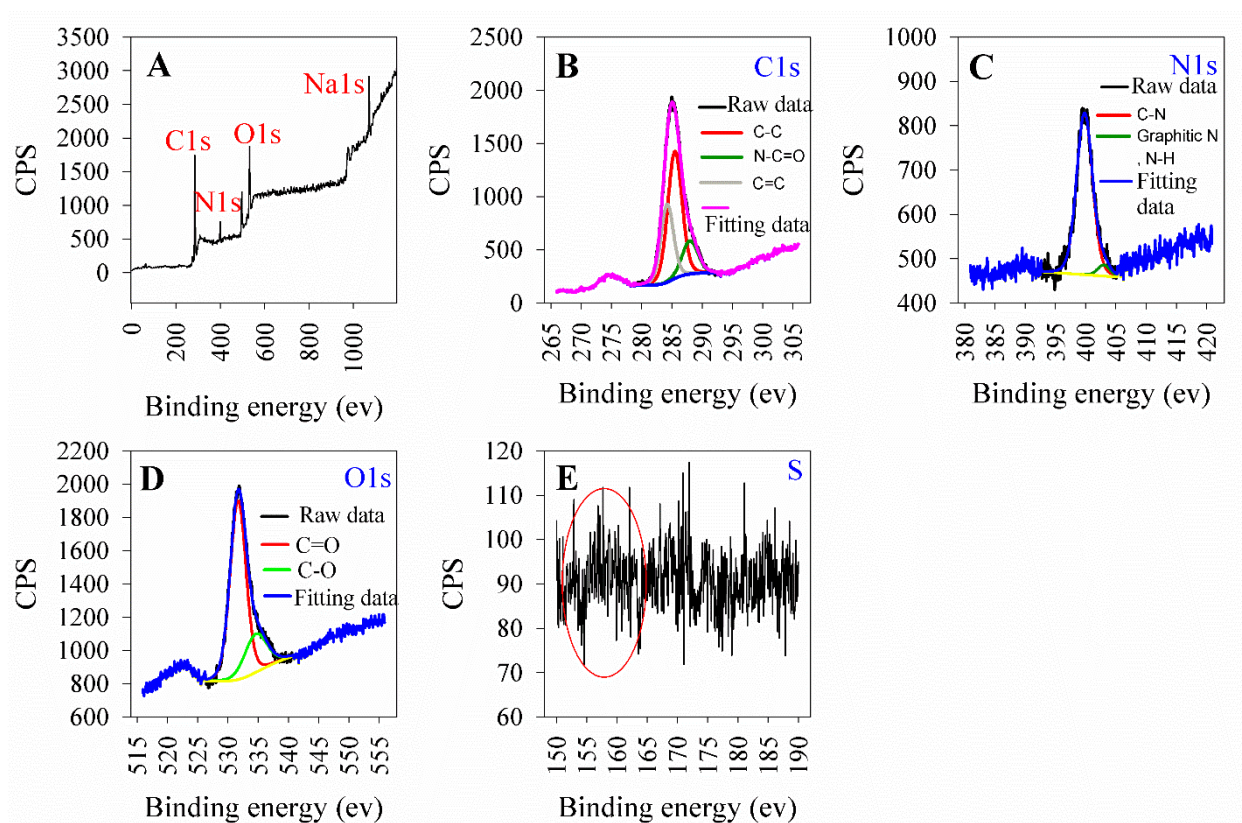


Figure S2. (A) XPS survey spectrum and high resolution (B) C1s, (C) N1s, and (D) O1s, spectra of BDNP-150. (E) sulfur region.

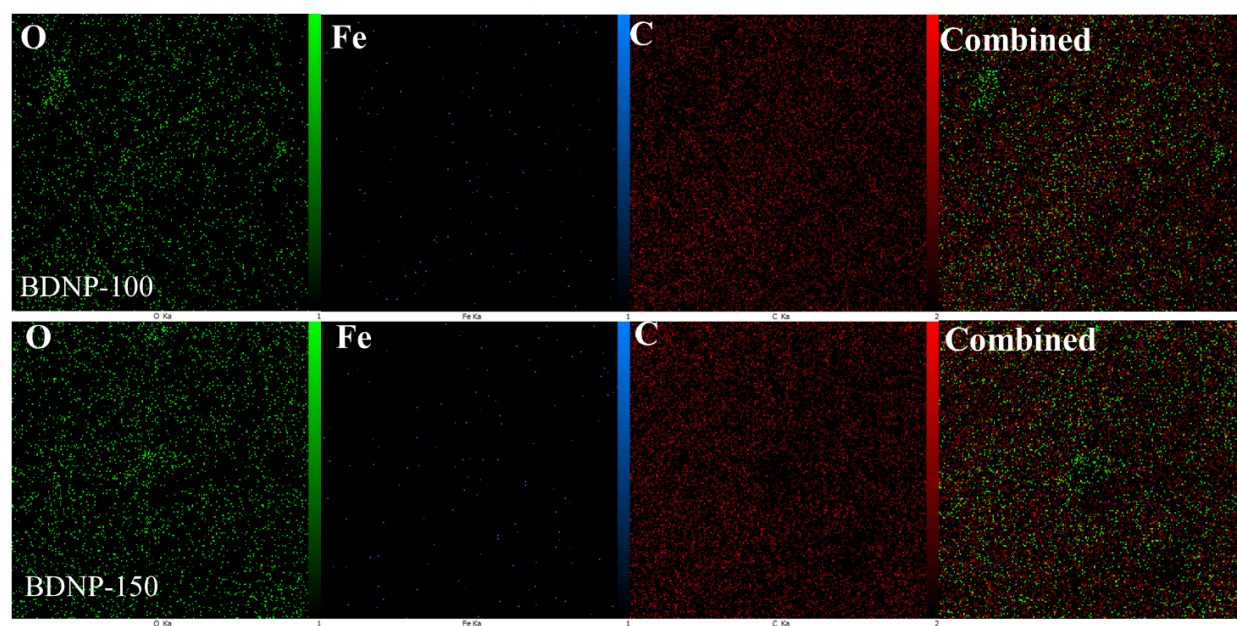


Figure S3. EDS elemental mapping of BDNP-100 and BDNP-150.

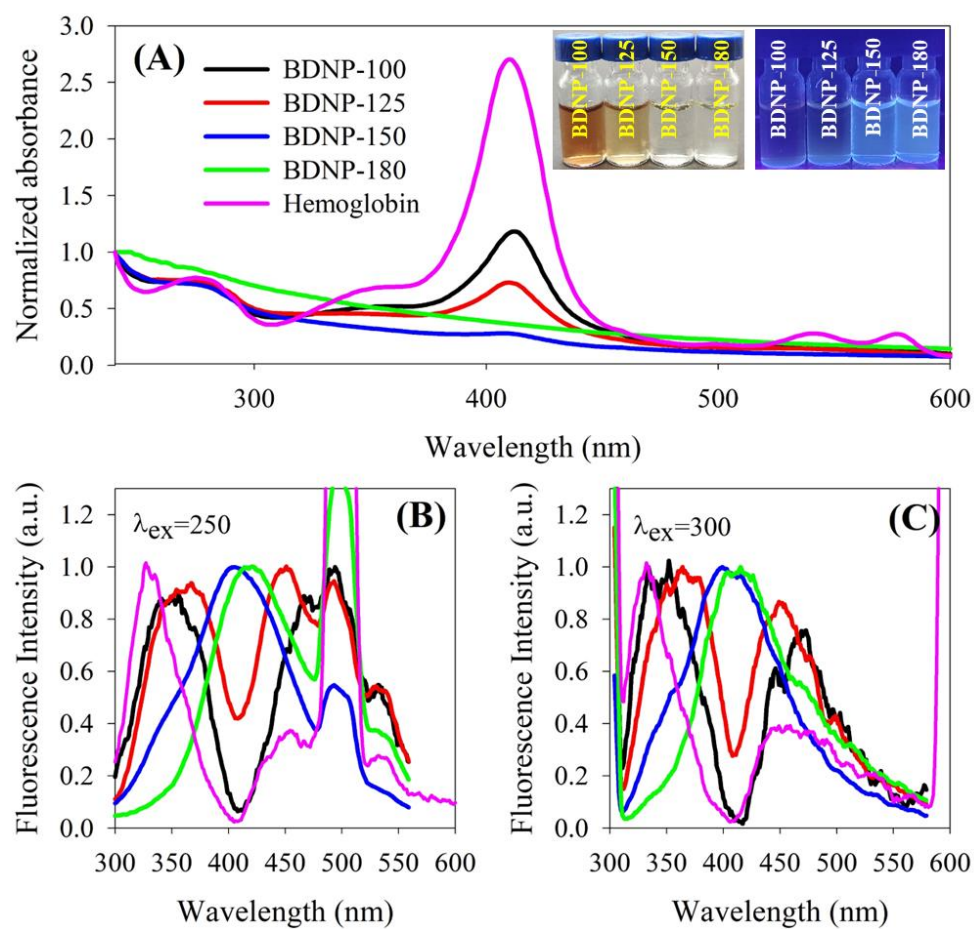


Figure S4. (A) the UV-Vis and (B) fluorescence spectra of BDNP samples under the excitation wavelengths of 250 nm and 300 nm.

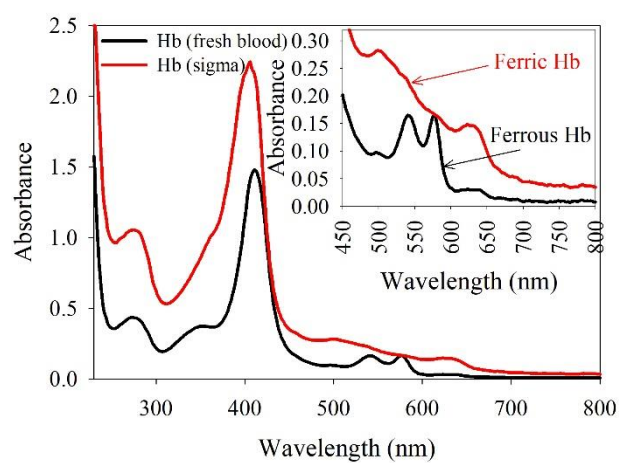


Figure S5. UV-Vis absorption spectra of Hb samples of two origins dispersed in deionized water.

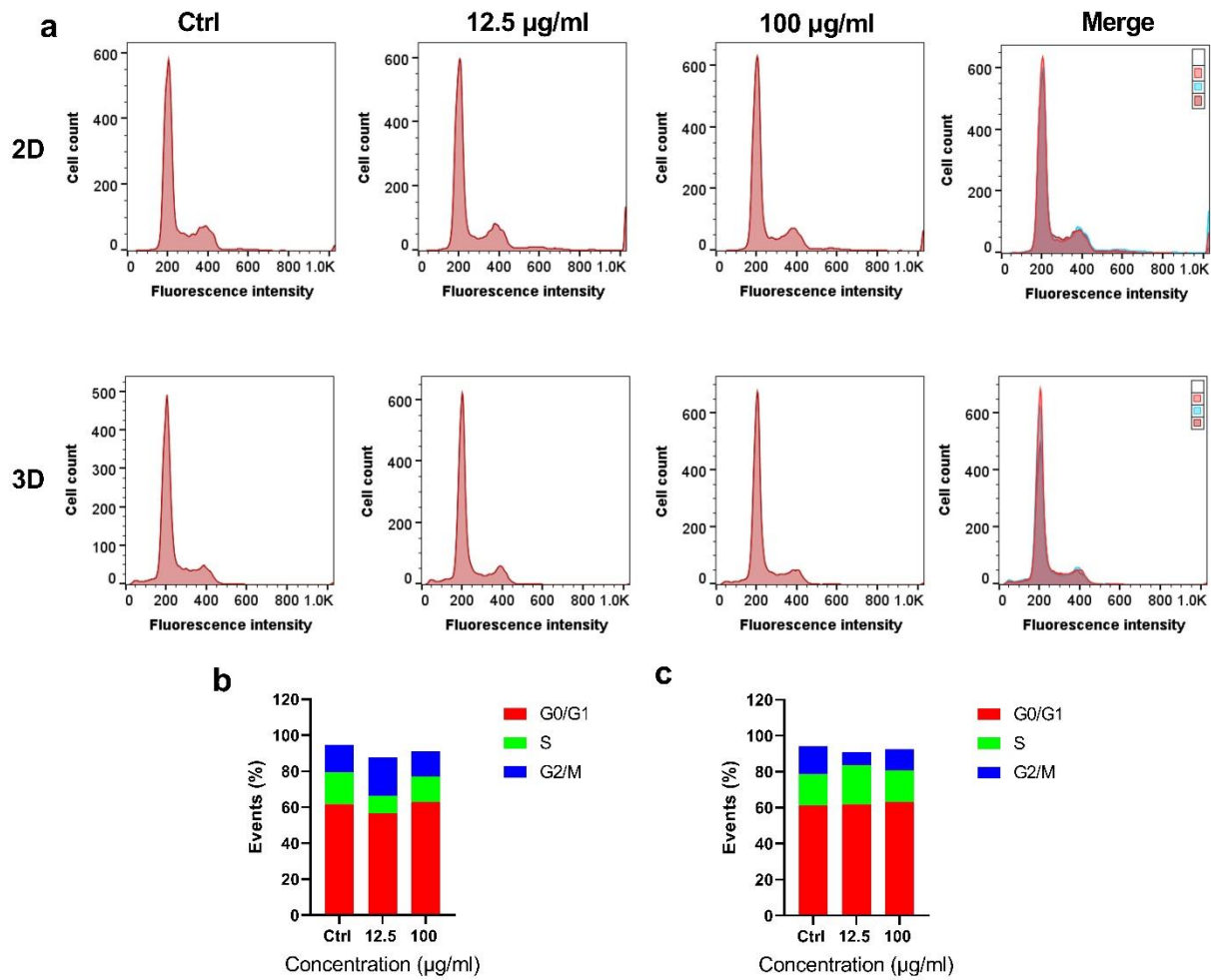


Figure S6. Flow cytometric analysis of the cell cycle of MCF-7 cell lines treated with BDNPs at concentrations of 12.5 and 100 $\mu\text{g/ml}$ in monolayer and 3D spheroid model condition.

Table S1. Performance of glucose colorimetric sensors.

	Linear range (μM)	LoD (μM)	Response time (min)	Reference
single iron site nanozyme on porous N-doped carbon	10-60	2.1	10	[6]
MoS ₂ -MIL-101(Fe)	0.01-20	0.01	20	[7]
2Fe ₂ O ₃ /30Pt/CNTs	5-20	0.92	-	[8]
GO _x @HP-MIL88B-BA	2-100	0.98	10	[9]
Fe-CNNPs	0-100	0.29	10	[10]
FNHCs	1.25-25	1.7	10	[11]
SO ₄ ²⁻ /CoFe ₂ O ₄	0-300	6.4	10	[12]
Pt nanoparticles/ hollow mesoporous carbon nanospheres	100–1330	35.4	30	[13]
Fe doped silica hollow spheres	5-600	0.24	5	[14]
3D porous graphene decorated with Fe ₃ O ₄ NPs	5-500	0.8	10 ^a	[15]
Fe@PCN-224	30-800	22	10	[16]
superparamagnetic iron oxide-doped mesoporous carbon	25-10000 (R ² =0.9415)	2	10	[17]
Fe-MoS ₂	5-2000	1.2	-	[18]
BDNP	50-700	40	4	This work
^a measurement was made at 50 °C				

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