

**Table S1. A comparison between different biosensor techniques for tau detection.**

| <i>Signal transduction</i> | <i>Analytical method<sup>(A)</sup></i> | <i>Sensor platform</i>                                     | <i>Target</i> | <i>Linear detection range</i>   | <i>Limit of detection<sup>(C)</sup></i>              | <i>Sample tested</i>           | <i>Label</i>      | <i>Ref.</i>      |
|----------------------------|--|--|---------------|---|--|--------------------------------|-------------------|------------------|
| <i>Optical</i>             | BLI                                    | Streptavidin interface probe sensor                        | Tau441        | 2-55 nM   | 6.7 nM (8.02 pM)                                     | Buffer and FBS                 | Label-free        | [37]             |
|                            | Spectrophotometry/ Interferometry      | Nanopore sensor with integrated microfluidic network       | T-Tau         | 15.6-2000 pg/mL   | 15.6 pg/mL in buffer (0.284pM)                       | Buffer and CSF                 | Label-free        | [38]             |
|                            | LSPR                                   | Resonance-based immunochip                                 | T-Tau         | 10-100000 pg/mL   | 10 pg/mL (0.15 pM)                                   | Buffer                         | Label-free        | [40]             |
|                            | SPR                                    | Surface plasmon resonance coupled to carbon nanostructures | T-Tau         | 125-1000 pM   | 125 pM   | Buffer and aCSF <sup>(D)</sup> | Label-free        | [38]             |
|                            | <b>Interferometry</b>                  | <b>Two Fabry-Perot interferometers</b>                     | <b>T-Tau</b>  | <b>12.5 µg/mL to 10 pg/mL in buffer</b><br><b>12.5 µg/mL to 10 pg/mL in serum</b> | <b>0.18 pM in buffer</b><br><b>0.18 pM in serum</b>  | <b>Buffer and Human serum</b>  | <b>Label-free</b> | <b>This work</b> |
| <i>Electrochemical</i>     | CV and EIS                             | Anti-Tau antibodies on polycrystalline Au surface          | Tau441        | 10-100 µg/mL  | 10 µg/mL   | Buffer                         | Label-free        | [41]             |
|                            | DPV                                    | Aptamer-antibody-sandwich                                  | Tau381        | 0.5-100 pM  | 0.42 pM in PBS (0.50 pM)                             | Human serum diluted 1/100      | Label-free        | [42]             |
|                            | DPV                                    | Neutral charged immunosensor                               | T-Tau         | 0.968-454 pM  | 0.968 pM   | Human serum                    | Label-free        | [43]             |
|                            | CV and EIS                             | Aptasensor   | Tau381        | 1-100 pM  | 0.7 pM   | Human serum diluted 1/100      | Label-free        | [44]             |
|                            | DVP and EIS                            | GO/pPG/anti-Tau nanoimmunosensor <sup>(B)</sup>            | T-Tau         | 0.25-250 nM   | 150 pM   | CSF and human serum            | Label-free        | [45]             |
| <i>Potentiometric</i>      | FET                                    | Transistor-based biosensor                                 | T-Tau         | 1-10 pM   | 1 pM in buffer and cell culture media, 10 pM in aCSF | Buffer, cell culture and aCSF  | Label-free        | [19]             |

(A) Abbreviations: BLI = Biolayer interferometry; LSPR = Localized surface plasmon resonance; SPR = Surface plasmon resonance; CV = Cyclic voltammetry; EIS = Electrochemical impedance spectroscopy; DPV = Differential pulse voltammetry; FET = Field effect transistor. (B) Abbreviations: GO = Graphene oxide; pPG = poly (propyleneglycol). (C) Conversion to molarity was carried out by assuming a mean tau molecular weight of 55KDa. (D) Abbreviation: aCSF = artificial cerebral spinal fluid.