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Abstract: Background: Antimicrobial resistance (AMR) poses a global health threat, with lowermiddle-income countries bearing a disproportionate burden. Surveillance of AMR under a One Health framework is needed to elucidate the associations among clinical, animal, and environmental AMR. This review aimed to describe the state of AMR in Ghana, focusing on One Health. Method: This review utilized the PRISMA guidelines and major databases to systematically search and analyze AMR in Ghana published from 1 January 2014 to 1 May 2023. Results: Out of the 48 articles that met the inclusion criteria, 28 studies were conducted on humans, 14 studies involved animals, and 6 studies focused on the environment. A total of 48 different pathogens were identified across the human, animal, and environmental sectors, with the most common being *Escherichia coli* (67%, n = 32), *Klebsiella* spp. (52%, n = 25), *Pseudomonas* spp. (40%, n = 19), and *Salmonella* spp. (38%, n = 18). Generally, a high prevalence of antibiotic resistance was observed among various bacterial species across the sectors. These bacteria exhibited resistance to commonly used antibiotics, with resistance to ampicillin and tetracycline exceeding 80%, and multidrug resistance (MDR) ranging from 17.6% in Shigella spp. to 100% in Acinetobacter spp. Conclusion: This review reaffirms the significant challenge of AMR in Ghana, with a high prevalence observed in the human, animal, and environmental sectors. Key pathogens (e.g., Staphylococcus aureus and Escherichia coli) found across the sectors emphasize the urgent need for a One Health approach to tackle AMR in Ghana.

**Keywords:** One Health; human health; animal health; environment; antimicrobial resistance (AMR); multidrug resistance (MDR); surveillance; systematic review; Ghana

### 1. Introduction

Recently, antimicrobial resistance (AMR) has been considered by the World Health Organization (WHO) as one of the top 10 global public health threats [1]. AMR limits the effectiveness of treatment options, leading to increased morbidity and treatment failure rates, longer hospital stays, elevated mortality rates, and higher healthcare costs [2]. In 2019, bacterial AMR directly caused 1.27 million deaths and contributed to 4.95 million deaths globally [3]. The economic costs of AMR are substantial, estimated to reach USD 1 trillion additional healthcare costs by 2050, along with significant GDP losses [4]. Although AMR affects countries across all regions and income levels, lower-middle-income countries are the most impacted [5]. In 2019, Ghana faced significant challenges related to AMR, with 5900 attributable deaths and an additional 25,300 associated deaths [6]. The country also ranked 36th globally in terms of age-standardized mortality rates associated with AMR per 100,000 population [6]. A major determinant of AMR and its spread is the weak enforcement and nonadherence to practice standards, policies, and regulations that govern the access to, and use of, antimicrobial agents in both humans and animals. In many sub-Saharan African countries such as Ghana, antimicrobial agents are easily obtained over the counter, facilitating their abuse [7,8]. While most high-income countries have adopted guidelines for the rational use of antibiotics, more efforts are needed in lower-middle-income countries [9–11].



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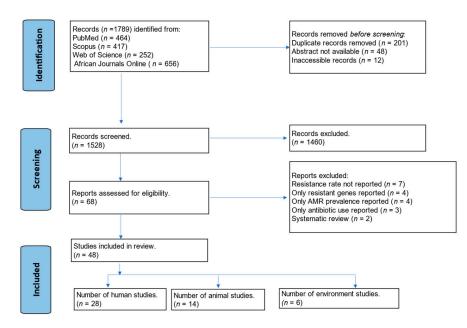
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Having recognized that the global pervasiveness of the AMR problem requires consolidated efforts for sustainable mitigation, the WHO Global Action Plan to address AMR emphasizes the application of the "One Health" approach [12]. This concept considers humans, animals, the environment, and the food chain as interconnected entities, owing to which resistance genes can be transferred from environmental bacteria to human pathogens [13]. Studying the interconnection among humans, animals, and the environment is crucial to preventing zoonotic disease outbreaks and maintaining ecosystem health, which impacts disease transmission and promotes a holistic approach to addressing health threats [14]. Surveillance of AMR under a One Health framework is needed to provide data for awareness and decision making and to enhance the understanding of the links between clinical, animal, and environmental AMR. In Ghana, a five-year National Action Plan (NAP) on AMR was instituted in 2017, focusing on enhanced surveillance through a One Health approach [15]. By monitoring the prevalence and distribution of resistant pathogens, the NAP can identify high-risk areas, track the movement of AMR, and develop targeted interventions to prevent further AMR spread. Although there has been a surge in AMR research data in Ghana, these data largely lack information on the interconnections among humans, animals, and the environment. This systematic review, therefore, examined AMR in Ghana from the One Health perspective, focusing on the antibiogram of bacterial pathogens.

#### 2. Results

# 2.1. Search Results

The initial search of the online databases identified a total of 1789 publications. After removing duplicates and inaccessible records, the titles and abstracts of the remaining 1528 records were screened. From these records, 1460 articles were excluded because they did not meet the inclusion criteria. Subsequently, 68 full-text articles were assessed for eligibility, of which 48 [16–63] met the inclusion criteria (Figure 1). These 48 articles included 28 studies on humans, 14 studies on animals, and six studies on the environment. The "human studies" included various populations, including healthy volunteers, patients, and food handlers. Studies involving animals included livestock such as fish, poultry, pigs, and cattle. The "environmental studies" included various samples: water and hospital surfaces (such as door handles and doctors' mobile phones).



**Figure 1.** PRISMA flow diagram for the identification, screening, and evaluation of articles included in the study.

### 2.2. Study Characteristics

The descriptive features of the articles used in this systematic review are grouped into three sectors: "human studies", "animal studies", and "environmental studies" (Table S1). For the "human studies", the majority (32.1%, n = 9) were conducted in the Greater Accra Region, followed by the Ashanti (18%, n = 5), Northern (11%, n = 3), Upper East (3.6%, n = 1), Western (7%, n = 2), Volta (7%, n = 2), and Central (3.6%, n = 1) Regions. Five studies were conducted at multiple centers across the Greater Accra, Eastern, Ashanti, and Volta Regions. Similarly, "animal studies" were conducted across various regions of Ghana, with the highest number of studies (35.7%, n = 5) in the Greater Accra Region, followed by the Ashanti Region (28.6%, n = 5). The Northern and Central Regions each had the least number of studies (7%, n = 1). Three studies were conducted at multiple centers across the Western, Central, Brong Ahafo, and Greater Accra Regions. Regarding "environmental studies", the region with the most studies was the Greater Accra Region (50%, n = 3), followed by the Northern Region (33%, n = 2). One study was a multicenter study across the Greater Accra and Eastern Regions. Overall, the Greater Accra Region was the most frequent sampling location (35.4%, *n* = 17), followed by the Ashanti (18.6%, *n* = 9), Northern (13%, *n* = 6), Upper East (2%, *n* = 1), Volta (4%, *n* = 2), Western (4%, *n* = 2), and Central (4%, *n* = 2) Regions. Nine studies were conducted at multiple centers, while no studies were identified in the Upper West Region.

The studies analyzed in the review varied in design and included cross-sectional, longitudinal, and retrospective studies (Table S2). Among the "human studies", the majority (39.3%, n = 11) were cross-sectional, followed by retrospective (18%, n = 5), retrospective cross-sectional (7.1%, n = 2), prospective (4%, n = 1), and prospective cross-sectional (4%, n = 1). For the "animal studies", the majority (50%, n = 7) were cross-sectional. Similarly, for the "environmental studies", the majority (33%, n = 2) were cross-sectional, followed by longitudinal (17%, n = 1). Overall, most studies used cross-sectional designs (41.7%, n = 20), followed by retrospective (10%, n = 5), retrospective cross-sectional (4.2%, n = 2), prospective (2.1%, n = 1), prospective cross-sectional (2.1%, n = 1), and longitudinal (2.1%, n = 1) designs. However, 18 studies (37.5%) did not specify the design used.

Most of the "animal studies" (50%, n = 7) were conducted on chicken/poultry, followed by cattle (36%, n = 5), fish (14%, n = 2), and pigs (7%, n = 1) (Table S2). Information on the common infections investigated in the "human studies" was included. The majority (14%, n = 4) were conducted on patients with urinary tract infection (UTI), followed by those with bloodstream infections (11%, n = 3), wound infection (7%, n = 2), cholera (7%, n = 2), HIV (4%, n = 1), diarrhea (4%, n = 1), and sepsis (4%, n = 1). Information on the specimen types used in the studies was also included. For samples reported from "human studies", the most abundant was urine (46%, n = 13), followed by blood (43%, n = 12), wound swabs (36%, n = 10), high vaginal swabs (21%, n = 6), ear swabs (18%, n = 5), urethral swabs (14%, n = 5)n = 4), stool (14%, n = 4), sputum (11%, n = 3), aspirates (11%, n = 3), nasopharyngeal swabs (11%, n = 3), semen (7%, n = 2), cerebrospinal fluid (7%, n = 2), fecal sludge (4%, n = 1), and palm swabs (4%, n = 1). Samples from "animal studies" were obtained from meat (36%, n = 5), feces (36%, n = 5), intestines (7%, n = 1), poultry feed (7%, n = 1), poultry drinking water (7%, n = 1), and rectal swabs (7%, n = 1). Samples from the environmental sector were obtained from water sources (67%, n = 4), mobile phones (17%, n = 1), and door handles (17%, n = 1).

The identification methods utilized in these studies included culture-based techniques (92.9%, n = 26), standard biochemical tests (78.6%, n = 22), PCR (10.7%, n = 3), and MALDI-TOF assays (25%, n = 7). In terms of susceptibility testing, the Kirby-Bauer disk diffusion method was the most commonly used method (75%, n = 21), while a few studies (14.3%, n = 4) employed the VITEK 2 system. Additionally, most studies (71.4%, n = 20) adopted the CLSI and EUCAST (17.9%, n = 5) guidelines (Table S3).

#### 2.3. Bacterial Agents and Antibiotic Resistance

Forty-eight different bacterial agents were isolated across the "human", "animal" and "environmental" sectors, with approximately one-third belonging to the Enterobacteriaceae family (Figure 2). *E. coli* was the most commonly reported organism for each of the three sectors: 19 "human studies", eight "animal studies", and five "environmental studies". *Klebsiella* spp. was the next most commonly reported organism for each of the three sectors: 18 "human studies", five "animal studies" and two "environmental studies".

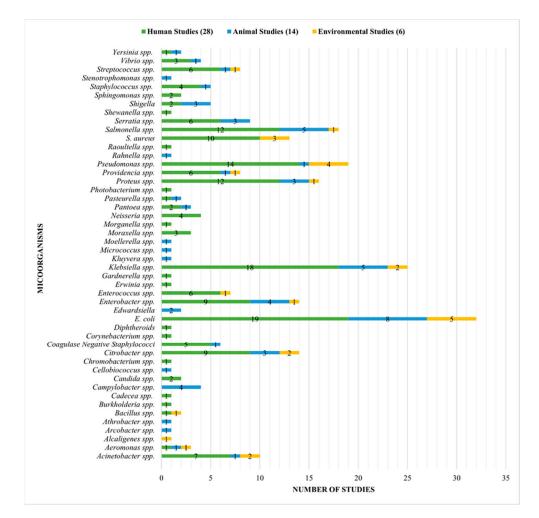


Figure 2. List of all bacteria isolated across the studies.

A high prevalence of resistance was observed among various bacterial species isolated from humans (Table 1). For instance, the prevalence of resistance in *Acinetobacter* spp. ranged from 35% to 67% for ciprofloxacin, tetracycline, gentamicin, and trimethoprim-sulfamethoxazole, while the prevalence of resistance to ampicillin and ceftriaxone was >80%. *Citrobacter* spp. exhibited resistance prevalence ranging from 22.1% to 93% for ciprofloxacin, tetracycline, gentamicin, trimethoprim-sulfamethoxazole, and ceftriaxone. *Enterococcus* showed high resistance prevalence of 100% to tetracycline [25,31], and trimethoprim-sulfamethoxazole [31]. Similarly, a high prevalence of resistance was observed among bacterial agents isolated from animals, as well as from the environment. For example, in the case of *Campylobacter* spp., which were reported from only animals, one study reported a resistance prevalence of 44% to ciprofloxacin, 81% to tetracycline [55], while another reported a resistance prevalence of 44% to ciprofloxacin, 81% to tetracycline, 81% to ampicillin, 56% to trimethoprim-sulfamethoxazole and 88% to chloramphenicol [52]. The prevalence of resistance to several antibiotics, including ciprofloxacin, ampicillin, trimethoprim-sulfamethoxazole, and ceftriaxone [60].

Bacteria Isolates	CIP			TET			GEN			AMP			SXT			СТХ			CHL			References
	Hu	An	En	Hu	An	En	Hu	An	En	Hu	An	En	Hu	An	En	Hu	An	En	Hu	An	En	
Acinetobacter spp.	35-44	-	-	55	-	-	37-63	-	-	95	-	-	58-67	-	-	88-89	-	-	-	-	-	[22,31,35]
Aeromonas spp.	-	-	13	_	-	23	-	-	17	-	-	-	-	-	-	_	-	-	-	-	-	[58]
Campylobacter spp.	-	44-75	-	-	70-100	-	-	-	-	-	81-96	-	-	56	-	-	-	-	-	88	-	[50,53,55]
Citrobacter spp.	22-80	-	100	78-93	53	-	22.2-66.7	5	97	-	100	100	50	-	100	78	45	-	-	58	-	[18,23,29,34,44,60]
E. coli	46-89	2-54	6–17	25-92	45-100	37	17-62	0-39	3.7–27	88-100	100	100	69–92	8-21	66–100	49-78	0-17	34-100	9-83	0-46	-	[17,18,22,23,25,29,31,34,35,37,41,43, 44,46,48,49,51,56,58–60]
Enterobacter spp.	11-73	-	-	87-100	82	-	25.9-47	0	-	100	100	-	37-68	-	-	52-68	45	-	-	36	-	[22,23,31,34,44]
Enterococcus spp.	-	-	-	100	-	-	-	-	-	0	-	-	58-100	-	-	-	-	-	44	-	-	[25,31]
Klebsiella spp.	14-76	-	12	71-89	55	-	29-83	14	0	100	100	100	48-95	-	46	64-91	36	-	50-92	55	-	[18,22-25,31,34,35,41,43,44,60]
Moraxella spp.	69	-	-	50-90	-	-	28	-	-	-	-	-	67	-	-	-	-	-	-	-	-	[18,22]
Neisseria gonorrhoeae	82	-	-	100	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	[19]
Proteus spp.	5-100	-	-	80-100	50	-	5-100	0	-	70-90	100	-	43-75	-	-	-	50	-	64	0	-	[18,25,29,31,34,44]
Pseudomonas spp.	15-100	-	5-100	80-100	-	-	10-40	-	11-100	96	-	100	92	-	100	-	-	-	-	-	-	[18,22,23,29,31,34,35,59,60]
Staphylococcus spp.	0-71	-	5-9	57-100	24	28-64	4-67	0	9	-	100	13	32-100	-	11	-	-	-	60-71	-	81.8	[16,18,21,29,34,44,56,61]
Salmonella spp.	0-67	63-65	-	35	60-100	-	0-67	0	-	33-57	26-80	_	17-66	69	-	0-22	20	-	34	6-20	-	[17,23,32,34,44]
Serratia spp.	-	-	-	>90	-	-	-	-	-	>90	-	-	-	-	-	>90	-	-	>90	_	-	[18]
Shigella spp.	-	-	-	90-100	67-100	-	-	0	-	-	98-100	-	39	-	-	-	100	-	-	100	-	[17,18,44,45]
Streptococcus spp.	-	-	-	-	17	-	-	17	-	100	83	-	-	-	-	-	-	-	63	-	-	[25,44]
Vibrio spp.	25-90	-	-	65	-	-	-	-	-	-	-	-	38	-	-	83.3	-	-	50	-	-	[17,26]

 Table 1. Antibiotic resistance profile of commonly isolated bacteria across sectors.

CHL: chloramphenicol, CTX: ceftriaxone, SXT: trimethoprim-sulfamethoxazole, AMP: ampicillin, GEN: gentamicin, TET: tetracycline, CIP: ciprofloxacin. Hu: "human studies", An: "animal studies", En: "environmental studies", and -: no data available.

Multidrug resistance, generally defined as the ability of bacteria to resist three or more antibiotics or three or more classes of antimicrobial drugs, was commonly observed in "human", "animal" and "environmental" studies across a wide spectrum of bacterial pathogens. For the "human studies", two reported MDR in *S. aureus* at a prevalence of 35.7% and 84.6% [16,20]. Two others reported MDR in *Salmonella* spp. at a prevalence of 41.5% and 99.6% [39,62]. In six "human studies", *E. coli* exhibited MDR, with the prevalence ranging from 41.5% to 99.6% [18,23,27,34,36,41]. MDR prevalence of 17.6%, 78.4%, 88%, 89.5% and 100% were reported for *Shigella* spp. [17], *Vibrio cholerae* [26], *K. pneumoniae* [35], *P. aeruginosa* [28], and *A. baumannii* [30], respectively. Six "animal studies" reported MDR prevalence ranging from 14.9% to 100% in *E. coli* [45,46,48,49,51,63]. Four "animal studies" reported MDR traits ranging from 20% to 66.6% in *Campylobacter* spp. [50,52,53,55]. One animal study reported an MDR prevalence of 40.4% in *Salmonella* spp. [32], while another reported an MDR prevalence of 19.1% in coagulase-negative Staphylococci [47]. Two "environmental studies" reported MDR in *E. coli* at 28% and 58.3% prevalence [59,60].

# 2.4. Risk of Bias

The risk of bias assessment in Figure 3 provides a comprehensive evaluation of the 48 studies included in this systematic review. The Robvis tool used for evaluation categorizes the risk of bias into three levels: low risk (shown in green), some concerns (in yellow), and high risk (in red). The overall low risk of bias across all evaluated domains indicates that the studies are methodologically sound and robust.

	1	D1	D2	Risk of bia D3	s domains D4	D5	Overal
	Dekker et al. [16]	X	+	+	+	+	+
	Afum et al. [17]	+	+	•	+	+	+
	Inusah et al. [18]	+	+	+	×	+	+
	Attram ct al. [19]	?	+	×	x	+	×
Sa	mpane-Donkor et al. [20]	+	+	<b>•</b>	+	+	+
	Bekoe et al. [21]	?	<b>(</b>	X	X	(+)	×
	Gnimatin et al. [22]	+	+	+	+	+	+
_	Sah et al. [23]	+	+	+	-	+	Ŧ
_	Quansah et al. [24]	?	+	×	×	+	×
_	Krumkamp et al. [25]	×	+	×	×	+	×
	Abana et al. [26]	×	+	×	×	+	×
_	Deku et al. [27]	+	+	-	+	+	+
	Agyepong et al. [28]	+	+	X	+	+	+
	Deininger et al. [29]	+	+	-	+	+	+
	Osei et al. [30]	+	+	•	+	+	+
	Janssen et al. [31]	+	+	•	+	+	+
	Andoh et al. [32]	×	+	×	+	+	÷
	Mohammed et al. [33]	+	+	X	-	+	-
	Asafo-Adjei et al. [34]	+	+	-	+	+	Ŧ
	Donkor et al. [35]	+	<b>(+</b> )	+	+	+	+
	Karikari et al. [36]	+	+	+	+	+	+
	Asare et al. [37]	+	+	-	+	Ŧ	+
	Omenako et al. [38]	+	+	<u> </u>	+	+	+
	Labi et al. [39]	+	Ŧ	+	+	+	+
	Codjoe et al. [40]	+	+	-	+	+	+
	Dwomoh et al. [41]	•	+	Ŧ	+	+	+
	Vicar et al. [42]	+	+	-	+	+	+
	Asamoah et al. [43]	+	+	+	+	+	(+)
	Agbeko et al. [44]	X	+	-	+	+	-
	Adinortey et al. [45]	X	+	-	+	+	- + -
	Mensah et al. [46]	+	+	+	+	+	+
	Boamah et al. [47]	-	+	X	+	+	-
	Ohene Larbi et al. [48]	+	+	+	+	+	+
	Adzitey et al. [49]	-	+	X	+	+	-
	Paintsil et al. [50]	+	+	-	+	+	+
	Dsani et al. [51]	+	+	+	+	+	+
	Karikari et al. [52]	+	+	-	+	+	+
As	uming-Bediako et al. [53]	X	+	-	+	+	-
	Eibach et al. [54]	X	+	-	+	+	-
	Dekker et al. [55]	+	+	+	+	+	
	Saba et al. [56]	X	+	-	+	+	-
	Odonkor et al. [57]	×	+	-	+	+	-
	Adomako et al. [58]	-	+	X	+	+	-
	Ahmed et al. [59]	+	+	+	+	+	+
	Addae-Nuku et al. [60]	+	+	+	+	+	+
	Saba et al. [61]	-	+	X	+	+	-
	Andoh et al. [62]	+	+	+	+	+	+
	Baah et al. [63]	+	+	-	+	+	+

Figure 3. Assessment of bias of the included studies using the Cochrane risk-of-bias tool (ROB2) [16-63].

### 3. Discussion

Antimicrobial resistance is a significant global health threat that affects human health, as well as animal and environmental health [64]. The One Health approach emphasizes the importance of monitoring and surveillance systems that combine data from human, animal, and environmental sources [13,65]. Thus, to gain a comprehensive understanding of the AMR situation in Ghana, this systematic review employed a One Health approach. Greater Accra Region was the most frequently sampled location for "human", "animal", and "environmental" studies, likely due to its high population density and industrial activities. Furthermore, this could be attributed to the presence of numerous research institutions, academic centers, and tertiary hospitals in the region. Interestingly, the Ashanti Region had more animal research than clinical research, which may be due to the high prevalence of agricultural activities in the region. It is essential to note that the Central, Volta, Western, Upper East, and Eastern Regions had fewer studies conducted across all sectors, which could be due to various factors, such as inadequate research infrastructure and funding. There is an urgent need for increased investment in research and surveillance programs, particularly in regions of the country that have received less focus regarding AMR surveillance efforts.

A decade ago, neglected tropical diseases, malaria, HIV/AIDS, sexually transmitted infections, and other infectious conditions in Ghana had higher mortality rates compared to AMR-related deaths [6]. However, presently, AMR-related deaths in Ghana have significantly increased, surpassing deaths from all the aforementioned diseases [6]. This rise can be attributed to resistant pathogens, as evident from this systematic review. The results indicate that AMR is a major concern across multiple bacterial species in Ghana; there are significant levels of resistance in bacteria commonly associated with human infections, such as E. coli, Klebsiella spp., S. aureus, and Salmonella spp. Notably, the presence of MDR bacteria (that is bacteria exhibiting resistance to multiple antibiotics) is particularly alarming. MDR was commonly observed among members of the Enterobacteriaceae, including E. coli and K. pneumoniae, which can cause severe infections and have multiple resistance mechanisms, such as extended-spectrum beta-lactamase (ESBL) and carbapenemase production [28,66,67]. The high prevalence of MDR-Enterobacteriaceae in animals can pose a risk to human public health, both directly and indirectly. E. coli, a common colonizer of the gastrointestinal tract in humans and animals [68,69], was the most studied bacterial species, followed by species of Klebsiella, Pseudomonas, and Salmonella. Klebsiella pneumoniae is naturally present in the respiratory and gastrointestinal tracts of humans and has a lower occurrence in animals, possibly due to diet, competition with other bacteria, or genetic virulence factors [70–72]. Pseudomonas spp., on the other hand, typically take advantage of weakened immune systems and causes infections that can be acquired in hospitals or the community, affecting both humans and animals [73]. Salmonella spp. are foodborne pathogens that cause gastroenteritis and other serious illnesses in humans and animals [74]. Given the ability of these pathogens to infect multiple host types, it is crucial to adopt a One Health approach that recognizes the interdependence of human, animal, and environmental health in addressing antibiotic resistance.

MDR bacteria can be transmitted from animals to humans through direct contact or the consumption of contaminated food products [75]. Studies have revealed the presence of MDR bacteria, including methicillin-resistant *S. aureus* (MRSA), ESBL-producing Enterobacteriaceae, and MDR-*Salmonella*, in meat products [63,76–79]. Similarly, a recent study in Romania found that pigs, and to some extent cattle, serve as significant natural reservoirs for zoonotic MDR *Campylobacter* strains [80]. However, in Ghana, AMR surveillance has mostly overlooked animal-derived foods, which are a significant source of antibiotic-resistant zoonotic pathogens. Monitoring AMR in animal-derived foods is just as essential as monitoring it in humans. The diverse range of bacterial contaminants found reflects the various infections that consumers may be exposed to, particularly if the meat is not cooked thoroughly before consumption, leading to foodborne illnesses and the transmission of zoonotic diseases. The involvement of MDR organisms in meat contamination could exacerbate the already severe problem of AMR, which is projected to cause 10 million deaths annually, amidst other adverse impacts [81]. One potential approach to broaden AMR surveillance is to incorporate wastewater as a significant reservoir of MDR pathogens. Although there is mounting evidence of bacterial resistance to antibiotics in Ghana, the available data primarily originate from clinical samples, with limited information on environmental AMR. The environmental dimension of AMR is closely linked to the use and disposal of antibiotics in various sectors. For instance, animal husbandry practices mainly utilize tetracyclines, penicillin, streptomycin, and ciprofloxacin as prophylaxes [82,83]. The release of antibiotic residues and resistant bacteria into the environment through wastewater effluents or agricultural runoffs contributes to the dissemination of resistance genes in environmental bacteria [84]. This environmental reservoir of AMR genes can potentially spread to human and animal pathogens, further exacerbating this problem. Raising awareness about AMR across sectors (human health, animal health, and the environment) can promote responsible antibiotic use and infection prevention, ultimately contributing to collective efforts in combating AMR.

## 4. Methods

#### 4.1. Preferred Reporting Items for Systematic Reviews (PRISMA) Guidelines

We followed the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidelines [85] to ensure a systematic and transparent approach to our literature search and review. The PRISMA guidelines provide a comprehensive checklist and flow diagram for record identification, screening, and evaluation.

#### 4.2. Search Strategy

A literature search was conducted using various databases and indexing services (PubMed, Scopus, Web of Science and African Journals Online) to identify all published studies on AMR in Ghana. We searched for English-language articles published between 1 January 2014, and 1 May 2023, as most relevant studies were published during that period. Additionally, we checked the reference lists of relevant articles for additional studies to include in our review. The search terms used included "antimicrobial resistance", "antibiotic resistance", "multidrug resistant bacteria", and "Ghana". A combination of keywords and MeSH terms was used to ensure that the search was comprehensive and specific to our research question. Boolean operators such as "AND" and "OR" were used to combine search terms and increase the search sensitivity. The PubMed, Scopus, Web of Science and African Journals Online databases were chosen for the search because they are widely used and have broad coverage of scientific publications. The search was not limited to any specific study design, population group, or outcome measure to ensure the capture of all relevant studies.

# 4.3. Inclusion and Exclusion Criteria

A two-step screening process was employed to identify relevant studies for the systematic review. First, the titles and abstracts of all studies identified were screened to exclude any irrelevant or duplicate studies. Second, we assessed the full texts of the remaining studies to determine their eligibility for inclusion in the review. We employed predefined inclusion and exclusion criteria to evaluate the studies. Studies reporting antibiotic resistance in bacteria from humans, animals, and the environment in Ghana were considered eligible. We excluded studies that focused solely on resistance genes without reporting phenotypic antibiogram data. Additionally, studies that reported only antibiotic use and those that were systematic reviews were excluded. Additionally, studies published before 2014 and those not conducted in Ghana were excluded. Two independent reviewers performed the screening process with fixed inclusion criteria. Mendeley Desktop version 1.19.8 was used to manage the search results and identify any duplicate records from the databases.

#### 4.4. Data Extraction and Analysis

Microsoft Excel 365 software was used to manage the data from the studies reviewed. Two authors worked independently using a data abstraction format prepared in Microsoft Excel 365 to extract data from the studies. The extracted information included author(s), year of publication, geographical area of the study, study design, sector of study (human, animal, and the environment), specimen type, bacteria isolated, antibiotics tested, and antibiotic resistance data. Microsoft Excel 365 was used for all computations and data visualization. The frequencies and prevalence of bacterial agents, including their antibiogram, resistance and multidrug resistance, were calculated; comparisons of these data were made across the different sectors of humans, animals and the environment. A statistical significance threshold was set at p < 0.05. To measure the variability between studies, the I<sup>2</sup> statistic and Cochran's Q test were used, with cutoff values of 25%, 50%, and 75% indicating low, moderate, and high heterogeneity, respectively [86]. The results are presented in tables, text, and figures.

#### 4.5. Evaluation of Bias

The Cochrane risk-of-bias tool (ROB2) [87] was used to assess the risk of bias in each study, and the results were visualized using the Robvis tool [88]. The Robvis tool evaluates five domains of bias: randomization, deviations from intended interventions, missing outcome data, measurement of outcomes, and selection of reported results. Each domain was classified as low risk, high risk, or some concerns, and studies were categorized as low risk if all domains showed low risk, high risk if any domain showed high risk, or some concerns if there were concerns in at least one domain.

## 5. Conclusions

This comprehensive review underscores the considerable burden of AMR in Ghana, with a high prevalence observed across the human, animal, and environmental sectors. This study revealed alarming levels of resistance among key pathogens, particularly the widespread resistance to multiple antibiotics among *S. aureus* and *E. coli* across various sectors. These findings emphasize the urgent need for a One Health approach for addressing AMR in Ghana.

**Supplementary Materials:** The following supporting information can be downloaded: https:// www.mdpi.com/article/10.3390/antibiotics13070662/s1, Figure S1: Antibiotics used in the studies; Table S1: List of full-text articles included in the systematic review; Table S2: Demographic features of studies included in the systematic review; Table S3: Characteristics of studies included in the systematic review; Table S4: Medically important bacteria isolated in the studies.

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### References

- 10 Global Health Issues to Track in 2021. Available online: https://www.who.int/news-room/spotlight/10-global-health-issuesto-track-in-2021 (accessed on 22 May 2024).
- Salam, A.; Al-Amin, Y.; Salam, M.T.; Pawar, J.S.; Akhter, N.; Rabaan, A.A.; Alqumber, M.A.A. Antimicrobial Resistance: A Growing Serious Threat for Global Public Health. *Healthcare* 2023, 11, 1946. [CrossRef] [PubMed]
- Murray, C.J.L.; Ikuta, K.S.; Sharara, F.; Swetschinski, L.; Aguilar, G.R.; Gray, A.; Han, C.; Bisignano, C.; Rao, P.; Wool, E.; et al. Global burden of bacterial antimicrobial resistance in 2019: A systematic analysis. *Lancet* 2022, 399, 629–655. [CrossRef] [PubMed]
- 4. Antimicrobial Resistance. Available online: https://www.who.int/news-room/fact-sheets/detail/antimicrobial-resistance (accessed on 6 June 2024).
- Pokharel, S.; Raut, S.; Adhikari, B. Tackling antimicrobial resistance in low-income and middle-income countries. *BMJ Glob. Health* 2024, 4, e002104. Available online: https://gh.bmj.com/content/4/6/e002104 (accessed on 1 November 2019). [CrossRef]
- 6. Antimicrobial Resistance (AMR). Available online: https://www.healthdata.org/research-analysis/health-risks-issues/ antimicrobial-resistance-amr (accessed on 11 July 2024).
- Yevutsey, S.K.; Buabeng, K.O.; Aikins, M.; Anto, B.P.; Biritwum, R.B.; Frimodt-Møller, N.; Gyansa-Lutterodt, M. Situational analysis of antibiotic use and resistance in Ghana: Policy and regulation. *BMC Public Health* 2017, 17, 896. [CrossRef] [PubMed]
- Duedu, K.O.; Offei, G.; Codjoe, F.S.; Donkor, E.S. Multidrug Resistant Enteric Bacterial Pathogens in a Psychiatric Hospital in Ghana: Implications for Control of Nosocomial Infections. *Int. J. Microbiol.* 2017, 2017, 9509087. [CrossRef] [PubMed]
- Ahiabu, M.-A.; Tersbøl, B.P.; Biritwum, R.; Bygbjerg, I.C.; Magnussen, P. A retrospective audit of antibiotic prescriptions in primary health-care facilities in Eastern Region, Ghana. *Health Policy Plan.* 2016, *31*, 250–258. [CrossRef] [PubMed]
- 10. Jimah, T.; Fenny, A.P.; Ogunseitan, O.A. Antibiotics stewardship in Ghana: A cross-sectional study of public knowledge, attitudes, and practices among communities. *One Health Outlook* **2020**, *2*, 12. [CrossRef] [PubMed]
- 11. Opoku, M.M.; Bonful, H.A.; Koram, K.A. Antibiotic prescription for febrile outpatients: A health facility-based secondary data analysis for the Greater Accra region of Ghana. *BMC Health Serv. Res.* 2020, 20, 978. [CrossRef] [PubMed]
- 12. Hailat, E.; Amiri, M.; Debnath, N.; Rahman, M.; Islam, N.; Fatima, Z.; Khader, Y.; Al Nsour, M. Strengthening the One Health Approach in the Eastern Mediterranean Region. *Interact. J. Med. Res.* **2023**, *12*, e41190. [CrossRef] [PubMed]
- 13. Aslam, B.; Khurshid, M.; Arshad, M.I.; Muzammil, S.; Rasool, M.; Yasmeen, N.; Shah, T.; Chaudhry, T.H.; Rasool, M.H.; Shahid, A.; et al. Antibiotic Resistance: One Health One World Outlook. *Front. Cell. Infect. Microbiol.* **2021**, *11*, 771510. [CrossRef] [PubMed]
- 14. Sharan, M.; Vijay, D.; Yadav, J.P.; Bedi, J.S.; Dhaka, P. Surveillance and response strategies for zoonotic diseases: A comprehensive review. *Sci. One Health* **2023**, *2*, 100050. [CrossRef]
- 15. Koduah, A.; Gyansa-Lutterodt, M.; Hedidor, G.K.; Sekyi-Brown, R.; Asiedu-Danso, M.; Asare, B.A.; Ackon, A.A.; Annan, E.A. Antimicrobial resistance national level dialogue and action in Ghana: Setting and sustaining the agenda and outcomes. *One Health Outlook* **2021**, *3*, 18. [CrossRef] [PubMed]
- Dekker, D.; Wolters, M.; Mertens, E.; Boahen, K.G.; Krumkamp, R.; Eibach, D.; Schwarz, N.G.; Adu-Sarkodie, Y.; Rohde, H.; Christner, M.; et al. Antibiotic Resistance and Clonal Diversity of Invasive *Staphylococcus aureus* in the Rural Ashanti Region, Ghana. *BMC Infect. Dis.* 2016, *16*, 720. [CrossRef] [PubMed]
- Afum, T.; Asandem, D.A.; Asare, P.; Asante-Poku, A.; Mensah, G.I.; Musah, A.B.; Opare, D.; Taniguchi, K.; Guinko, N.M.; Aphour, T.; et al. Diarrhea-Causing Bacteria and Their Antibiotic Resistance Patterns among Diarrhea Patients from Ghana. *Front. Microbiol.* 2022, 13, 894319. [CrossRef] [PubMed]
- Inusah, A.; Quansah, E.; Fosu, K.; Dadzie, I. Resistance Status of Bacteria from a Health Facility in Ghana: A Retrospective Study. J. Pathog. 2021, 2021, 6648247. [CrossRef] [PubMed]
- Attram, N.; Agbodzi, B.; Dela, H.; Behene, E.; Nyarko, E.O.; Kyei, N.N.A.; Larbi, J.A.; Lawson, B.W.L.; Addo, K.K.; Newman, M.J.; et al. Antimicrobial Resistance (AMR) and Molecular Characterization of Neisseria Gonorrhoeae in Ghana, 2012–2015. *PLoS ONE* 2019, 14, e0223598. [CrossRef] [PubMed]
- Sampane-Donkor, E.; Badoe, E.V.; Annan, J.A.; Nii-Trebi, N.I. Colonisation of Antibiotic Resistant Bacteria in a Cohort of HIV Infected Children in Ghana. *Pan. Afr. Med. J.* 2017, 26, 1937–8688. [PubMed Central]
- 21. Bekoe, S.O.; Hane-Weijman, S.; Trads, S.L.; Orman, E.; Opintan, J.; Hansen, M.; Frimodt-Møller, N.; Styrishave, B. Reservoir of Antibiotic Residues and Resistant Coagulase Negative Staphylococci in a Healthy Population in the Greater Accra Region, Ghana. *Antibiotics* **2022**, *11*, 119. [CrossRef] [PubMed]
- 22. Gnimatin, J.-P.; Weyori, E.W.; Agossou, S.M.; Adokiya, M.N. Bacterial Infections Epidemiology and Factors Associated with Multidrug Resistance in the Northern Region of Ghana. *Sci. Rep.* **2022**, *12*, 22069. [CrossRef] [PubMed]
- 23. Sah, A.K.; Feglo, P.K. Plasmid-mediated quinolone resistance determinants in clinical bacterial pathogens isolated from the Western Region of Ghana: A cross-sectional study. *Pan Afr. Med. J.* **2022**, *43*, 207. [CrossRef] [PubMed]
- Quansah, E.; Amoah Barnie, P.; Omane Acheampong, D.; Obiri-Yeboah, D.; Odarkor Mills, R.; Asmah, E.; Cudjoe, O.; Dadzie, I. Geographical Distribution of β-Lactam Resistance among *Klebsiella* spp. from Selected Health Facilities in Ghana. *Trop. Med. Infect. Dis.* 2019, 4, 117. [CrossRef] [PubMed]
- Krumkamp, R.; Oppong, K.; Hogan, B.; Strauss, R.; Frickmann, H.; Wiafe-Akenten, C.; Boahen, K.G.; Rickerts, V.; Smith, I.M.; Groß, U.; et al. Spectrum of antibiotic resistant bacteria and fungi isolated from chronically infected wounds in a rural district hospital in Ghana. *PLoS ONE* 2020, *15*, e0237263. [CrossRef] [PubMed]

- Abana, D.; Gyamfi, E.; Dogbe, M.; Opoku, G.; Opare, D.; Boateng, G.; Mosi, L. Investigating the virulence genes and antibiotic susceptibility patterns of Vibrio cholerae O1 in environmental and clinical isolates in Accra, Ghana. *BMC Infect. Dis.* 2019, 19, 76. [CrossRef] [PubMed]
- 27. Deku, J.G.; Duedu, K.O.; Ativi, E.; Kpene, G.E.; Feglo, P.K. Occurrence and distribution of extended-spectrum β-lactamase in clinical *Escherichia coli* isolates at Ho Teaching Hospital in Ghana. *Ghana Med. J.* **2021**, *55*, 298–307. [CrossRef] [PubMed]
- 28. Agyepong, N.; Govinden, U.; Owusu-Ofori, A.; Essack, S.Y. Multidrug-resistant gram-negative bacterial infections in a teaching hospital in Ghana. *Antimicrob. Resist. Infect. Control.* **2018**, *7*, 37. [CrossRef] [PubMed]
- Deininger, S.; Gründler, T.; Deininger, S.H.M.; Lütcke, K.; Lütcke, H.; Agbesi, J.; Ladzaka, W.; Gyamfi, E.; Wichlas, F.; Hofmann, V.; et al. The Antimicrobial Resistance (AMR) Rates of Uropathogens in a Rural Western African Area—A Retrospective Single-Center Study from Kpando, Ghana. *Antibiotics* 2022, *11*, 1808. [CrossRef] [PubMed]
- Osei, M.-M.; Dayie, N.T.K.D.; Azaglo, G.S.K.; Tettey, E.Y.; Nartey, E.T.; Fenny, A.P.; Manzi, M.; Kumar, A.M.V.; Labi, A.-K.; Opintan, J.A.; et al. Alarming Levels of Multidrug Resistance in Aerobic Gram-Negative Bacilli Isolated from the Nasopharynx of Healthy Under-Five Children in Accra, Ghana. *Int. J. Environ. Res. Public Health* 2022, 19, 10927. [CrossRef] [PubMed]
- Janssen, H.; Janssen, I.; Cooper, P.; Kainyah, C.; Pellio, T.; Quintel, M.; Monnheimer, M.; Groß, U.; Schulze, M.H. Antimicrobial-Resistant Bacteria in Infected Wounds, Ghana, 2014. *Emerg. Infect. Dis.* 2018, 24, 916. [CrossRef] [PubMed]
- Andoh, L.A.; Dalsgaard, A.; Obiri-Danso, K.; Newman, M.J.; Barco, L.; Olsen, J.E. Prevalence and antimicrobial resistance of Salmonella serovars isolated from poultry in Ghana. *Epidemiol. Infect.* 2016, 144, 3288–3299. [CrossRef] [PubMed]
- 33. Mohammed, J.; Hounmanou, Y.M.G.; Thomsen, L.E. Antimicrobial resistance among clinically relevant bacterial isolates in Accra: A retrospective study. *BMC Res. Notes* **2018**, *11*, 254. [CrossRef] [PubMed]
- 34. Asafo-Adjei, K.; Mensah, J.E.; Labi, A.-K.; Dayie, N.T.K.D.; Donkor, E.S. Urinary Tract Infections among Bladder Outlet Obstruction Patients in Accra, Ghana: Aetiology, Antibiotic Resistance, and Risk Factors. *Diseases* **2018**, *6*, 65. [CrossRef] [PubMed]
- Donkor, E.S.; Muhsen, K.; Johnson, S.A.M.; Kotey, F.C.N.; Dayie, N.T.K.D.; Tetteh-Quarcoo, P.B.; Tette, E.M.A.; Osei, M.-M.; Egyir, B.; Nii-Trebi, N.I.; et al. Multicenter Surveillance of Antimicrobial Resistance among Gram-Negative Bacteria Isolated from Bloodstream Infections in Ghana. *Antibiotics* 2023, 12, 255. [CrossRef] [PubMed]
- Karikari, A.B.; Saba, C.K.; Yamik, D.Y. Reported Cases of Urinary Tract Infections and the Susceptibility of Uropathogens from Hospitals in Northern Ghana. *Microbiol. Insights* 2022, 15, 11786361221106108. [CrossRef] [PubMed]
- 37. Asare, K.K.; Amoah, S.; Coomson, C.A., Jr.; Banson, C.; Yaro, D.; Mbata, J.; Aaron Arthur, R.; Mayeem, P.B.; Afrifa, J.; Bentsi-Enchill, F.; et al. Antibiotic-resistant pathogenic bacterial isolates from patients attending the outpatient department of university of Cape Coast hospital, Ghana: A retrospective study between 2013–2015. PLoS Glob. Public Health 2022, 2, e0000417. [CrossRef] [PubMed]
- Omenako, K.A.; Enimil, A.; Marfo, A.F.A.; Timire, C.; Chinnakali, P.; Fenny, A.P.; Jeyashree, K.; Buabeng, K.O. Pattern of Antimicrobial Susceptibility and Antimicrobial Treatment of Neonates Admitted with Suspected Sepsis in a Teaching Hospital in Ghana, 2021. Int. J. Environ. Res. Public Health 2022, 19, 12968. [CrossRef] [PubMed]
- Labi, A.-K.; Obeng-Nkrumah, N.; Addison, N.O.; Donkor, E.S. Salmonella blood stream infections in a tertiary care setting in Ghana. BMC Infect. Dis. 2014, 14, 3857. [CrossRef] [PubMed]
- Codjoe, F.S.; Donkor, E.S.; Smith, T.J.; Miller, K. Phenotypic and Genotypic Characterization of Carbapenem-Resistant Gram-Negative Bacilli Pathogens from Hospitals in Ghana. *Microb. Drug Resist.* 2019, 25, 1449–1457. [CrossRef] [PubMed]
- Dwomoh, F.P.; Kotey, F.C.N.; Dayie, N.T.K.D.; Osei, M.-M.; Amoa-Owusu, F.; Bannah, V.; Alzahrani, F.M.; Halawani, I.F.; Alzahrani, K.J.; Egyir, B.; et al. Phenotypic and genotypic detection of carbapenemase-producing *Escherichia coli* and *Klebsiella* pneumoniae in Accra, Ghana. PLoS ONE 2022, 17, e0279715. [CrossRef] [PubMed]
- Vicar, E.K.; Alo, D.B.; Koyiri, V.C.; Opare-Asamoah, K.; Obeng-Bempong, M.; Mensah, G.I. Carriage of Antibiotic Resistant Bacteria and Associated Factors Among Food Handlers in Tamale Metropolis, Ghana: Implications for Food Safety. *Microbiol. Insights* 2023, 16, 11786361221150696. [CrossRef] [PubMed]
- Asamoah, B.; Labi, A.-K.; Gupte, H.A.; Davtyan, H.; Peprah, G.M.; Adu-Gyan, F.; Nair, D.; Muradyan, K.; Jessani, N.S.; Sekyere-Nyantakyi, P. High Resistance to Antibiotics Recommended in Standard Treatment Guidelines in Ghana: A Cross-Sectional Study of Antimicrobial Resistance Patterns in Patients with Urinary Tract Infections between 2017–2021. *Int. J. Environ. Res. Public Health* 2022, 19, 16556. [CrossRef] [PubMed]
- Agbeko, R.; Aheto, D.W.; Asante, D.K.; Asare, N.K.; Boateng, A.A.; Adinortey, C.A. Identification of molecular determinants of antibiotic resistance in some fish farms of Ghana. *Heliyon* 2022, 8, e10431. [CrossRef] [PubMed]
- 45. Adinortey, C.A.; Aheto, D.W.; Boateng, A.A.; Agbeko, R. Multiple Antibiotic Resistance-Coliform Bacteria in Some Selected Fish Farms of the Central Region of Ghana. *Scientifica* **2020**, 2020, 6641461. [CrossRef] [PubMed]
- Mensah, G.I.; Adjei, V.Y.; Vicar, E.K.; Atsu, P.S.; Blavo, D.L.; Johnson, S.A.M.; Addo, K.K. Safety of Retailed Poultry: Analysis of Antibiotic Resistance in *Escherichia coli* From Raw Chicken and Poultry Fecal Matter From Selected Farms and Retail Outlets in Accra, Ghana. *Microbiol. Insights* 2022, 15, 11786361221093278. [CrossRef] [PubMed]
- Boamah, V.E.; Agyare, C.; Odoi, H.; Adu, F.; Gbedema, S.Y.; Dalsgaard, A. Prevalence and antibiotic resistance of coagulasenegative Staphylococci isolated from poultry farms in three regions of Ghana. *Infect. Drug Resist.* 2017, 10, 175–183. [CrossRef] [PubMed]
- Ohene Larbi, R.; Adeapena, W.; Ayim-Akonor, M.; Ansa, E.D.O.; Tweya, H.; Terry, R.F.; Labi, A.-K.; Harries, A.D. Antimicrobial, Multi-Drug and Colistin Resistance in Enterobacteriaceae in Healthy Pigs in the Greater Accra Region of Ghana, 2022: A Cross-Sectional Study. Int. J. Environ. Res. Public Health 2022, 19, 10449. [CrossRef] [PubMed]

- Adzitey, F.; Assoah-Peprah, P.; Teye, G.A.; Somboro, A.M.; Kumalo, H.M.; Amoako, D.G. Prevalence and Antimicrobial Resistance of *Escherichia coli* Isolated from Various Meat Types in the Tamale Metropolis of Ghana. *Int. J. Food Sci.* 2020, 2020, 8877196. [CrossRef] [PubMed]
- Paintsil, E.K.; Ofori, L.A.; Akenten, C.W.; Zautner, A.E.; Mbwana, J.; Jaeger, A.; Lamshöft, M.; May, J.; Obiri-Danso, K.; Philipps, R.O.; et al. Antibiotic-resistant Campylobacter coli and Campylobacter jejuni in commercial and smallholder farm animals in the Asante Akim North Municipality of Ghana. *Front. Microbiol.* 2022, *13*, 983047. [CrossRef] [PubMed]
- Dsani, E.; Afari, E.A.; Danso-Appiah, A.; Kenu, E.; Kaburi, B.B.; Egyir, B. Antimicrobial resistance and molecular detection of extended spectrum β-lactamase producing *Escherichia coli* isolates from raw meat in Greater Accra region, Ghana. *BMC Microbiol.* 2020, 20, 253. [CrossRef] [PubMed]
- 52. Karikari, A.B.; Obiri-Danso, K.; Frimpong, E.H.; Krogfelt, K.A. Antibiotic Resistance of *Campylobacter* Recovered from Faeces and Carcasses of Healthy Livestock. *BioMed Res. Int.* 2017, 2017, 4091856. [CrossRef] [PubMed]
- Asuming-Bediako, N.; Kunadu, A.P.-H.; Jordan, D.; Abraham, S.; Habib, I. Prevalence and antimicrobial susceptibility pattern of Campylobacter jejuni in raw retail chicken meat in Metropolitan Accra, Ghana. Int. J. Food Microbiol. 2022, 376, 109760. [CrossRef]
- Eibach, D.; Dekker, D.; Boahen, K.G.; Akenten, C.W.; Sarpong, N.; Campos, C.B.; Berneking, L.; Aepfelbacher, M.; Krumkamp, R.; Owusu-Dabo, E.; et al. Extended-spectrum beta-lactamase-producing *Escherichia coli* and *Klebsiella pneumoniae* in local and imported poultry meat in Ghana. *Vet. Microbiol.* 2018, 217, 7–12. [CrossRef] [PubMed]
- 55. Dekker, D.; Eibach, D.; Boahen, K.G.; Akenten, C.W.; Pfeifer, Y.; Zautner, A.E.; Mertens, E.; Krumkamp, R.; Jaeger, A.; Flieger, A.; et al. Fluoroquinolone-Resistant Salmonella enterica, Campylobacter spp., and Arcobacter butzleri from Local and Imported Poultry Meat in Kumasi, Ghana. Foodborne Pathog. Dis. 2019, 16, 352–358. [CrossRef] [PubMed]
- 56. Saba, C.K.S.; Naa-Inour, F.; Kpordze, S.W. Antibiotic resistance pattern of methicillin-resistant *Staphylococcus aureus* and *Escherichia coli* from mobile phones of healthcare workers in public hospitals in Ghana. *Pan Afr. Med. J.* **2022**, *41*, 259. [CrossRef] [PubMed]
- 57. Odonkor, S.T.; Simpson, S.V.; Medina, W.R.M.; Fahrenfeld, N.L. Antibiotic-Resistant Bacteria and Resistance Genes in Isolates from Ghanaian Drinking Water Sources. J. Environ. Public Health 2022, 2022, 2850165. [CrossRef] [PubMed]
- Adomako, L.A.B.; Yirenya-Tawiah, D.; Nukpezah, D.; Abrahamya, A.; Labi, A.-K.; Grigoryan, R.; Ahmed, H.; Owusu-Danquah, J.; Annang, T.Y.; Banu, R.A.; et al. Reduced Bacterial Counts from a Sewage Treatment Plant but Increased Counts and Antibiotic Resistance in the Recipient Stream in Accra, Ghana—A Cross-Sectional Study. *Trop. Med. Infect. Dis.* 2021, *6*, 79. [CrossRef] [PubMed]
- Ahmed, H.; Zolfo, M.; Williams, A.; Ashubwe-Jalemba, J.; Tweya, H.; Adeapena, W.; Labi, A.-K.; Adomako, L.A.B.; Addico, G.N.D.; Banu, R.A.; et al. Antibiotic-Resistant Bacteria in Drinking Water from the Greater Accra Region, Ghana: A Cross-Sectional Study, December 2021–March 2022. *Int. J. Environ. Res. Public Health* 2022, *19*, 12300. [CrossRef] [PubMed]
- Addae-Nuku, D.S.; Kotey, F.C.; Dayie, N.T.; Osei, M.-M.; Tette, E.M.; Debrah, P.; Donkor, E.S. Multidrug-Resistant Bacteria in Hospital Wastewater of the Korle Bu Teaching Hospital in Accra, Ghana. *Environ. Health Insights* 2022, 16, 11786302221130612. [CrossRef] [PubMed]
- Saba, C.K.S.; Amenyona, J.K.; Kpordze, S.W. Prevalence and pattern of antibiotic resistance of Staphylococcus aureus isolated from door handles and other points of contact in public hospitals in Ghana. *Antimicrob. Resist. Infect. Control.* 2017, *6*, 44. Available online: https://aricjournal.biomedcentral.com/articles/10.1186/s13756-017-0203-2 (accessed on 10 May 2017). [CrossRef]
- 62. Andoh, L.A.; Ahmed, S.; Olsen, J.E.; Obiri-Danso, K.; Newman, M.J.; Opintan, J.A.; Barco, L.; Dalsgaard, A. Prevalence and characterization of Salmonella among humans in Ghana. *Trop. Med. Health* **2017**, *45*, 3. [CrossRef] [PubMed]
- 63. Baah, D.A.; Kotey, F.C.N.; Dayie, N.T.K.D.; Codjoe, F.S.; Tetteh-Quarcoo, P.B.; Donkor, E.S. Multidrug-Resistant Gram-Negative Bacteria Contaminating Raw Meat Sold in Accra, Ghana. *Pathogens* **2022**, *11*, 1517. [CrossRef]
- 64. Velazquez-Meza, M.E.; Galarde-López, M.; Carrillo-Quiróz, B.; Alpuche-Aranda, C.M. Antimicrobial resistance: One Health approach. *Vet. World* 2022, *15*, 743–749. [CrossRef] [PubMed]
- 65. Badau, E. A One Health perspective on the issue of the antibiotic resistance. Parasite 2021, 28, 16. [CrossRef] [PubMed]
- Shaikh, S.; Fatima, J.; Shakil, S.; Rizvi, S.M.D.; Kamal, M.A. Antibiotic resistance and extended spectrum beta-lactamases: Types, epidemiology and treatment. Saudi J. Biol. Sci. 2015, 22, 90–101. [CrossRef] [PubMed]
- Alkofide, H.; Alhammad, A.M.; Alruwaili, A.; Aldemerdash, A.; A Almangour, T.; Alsuwayegh, A.; Almoqbel, D.; Albati, A.; Alsaud, A.; Enani, M. Multidrug-Resistant and Extensively Drug-Resistant Enterobacteriaceae: Prevalence, Treatments, and Outcomes—A Retrospective Cohort Study. *Infect. Drug Resist.* 2020, *13*, 4653–4662. [CrossRef] [PubMed]
- 68. Loayza, F.; Graham, J.P.; Trueba, G. Factors Obscuring the Role of E. coli from Domestic Animals in the Global Antimicrobial Resistance Crisis: An Evidence-Based Review. *Int. J. Environ. Res. Public Health* **2020**, *17*, 3061. [CrossRef] [PubMed]
- Ramos, S.; Silva, V.; Dapkevicius, M.d.L.E.; Caniça, M.; Tejedor-Junco, M.T.; Igrejas, G.; Poeta, P. *Escherichia coli* as Commensal and Pathogenic Bacteria among Food-Producing Animals: Health Implications of Extended Spectrum β-Lactamase (ESBL) Production. *Animals* 2020, 10, 2239. [CrossRef] [PubMed]
- Khan, I.; Bai, Y.; Zha, L.; Ullah, N.; Ullah, H.; Shah, S.R.H.; Sun, H.; Zhang, C. Mechanism of the Gut Microbiota Colonization Resistance and Enteric Pathogen Infection. *Front. Cell. Infect. Microbiol.* 2021, 11, 716299. [CrossRef] [PubMed]
- 71. Wareth, G.; Neubauer, H. The Animal-foods-environment interface of *Klebsiella pneumoniae* in Germany: An observational study on pathogenicity, resistance development and the current situation. *Vet. Res.* **2021**, *52*, 16. [CrossRef] [PubMed]
- 72. Young, T.M.; Bray, A.S.; Nagpal, R.K.; Caudell, D.L.; Yadav, H.; Zafar, M.A. Animal Model To Study *Klebsiella pneumoniae* Gastrointestinal Colonization and Host-to-Host Transmission. *Infect. Immun.* **2020**, *88*, e00071-20. [CrossRef] [PubMed]

- 73. Verdial, C.; Serrano, I.; Tavares, L.; Gil, S.; Oliveira, M. Mechanisms of Antibiotic and Biocide Resistance That Contribute to *Pseudomonas aeruginosa* Persistence in the Hospital Environment. *Biomedicines* **2023**, *11*, 1221. [CrossRef] [PubMed]
- 74. Ehuwa, O.; Jaiswal, A.K.; Jaiswal, S. *Salmonella*, Food Safety and Food Handling Practices. *Foods* **2021**, *10*, 907. [CrossRef] [PubMed]
- 75. Pokharel, S.; Shrestha, P.; Adhikari, B. Antimicrobial use in food animals and human health: Time to implement 'One Health' approach. *Antimicrob. Resist. Infect. Control* 2020, *9*, 181. [CrossRef] [PubMed]
- 76. Ahmed, H.A.; Elsohaby, I.; Elamin, A.M.; El-Ghafar, A.E.A.; Elsaid, G.A.; Elbarbary, M.; Mohsen, R.A.; El Feky, T.M.; El Bayomi, R.M. Extended-spectrum β-lactamase-producing E. coli from retail meat and workers: Genetic diversity, virulotyping, pathotyping and the antimicrobial effect of silver nanoparticles. *BMC Microbiol.* 2023, 23, 212. [CrossRef] [PubMed]
- 77. Gambino, D.; Gargano, V.; Butera, G.; Sciortino, S.; Pizzo, M.; Oliveri, G.; Cardamone, C.; Piraino, C.; Cassata, G.; Vicari, D.; et al. Food Is Reservoir of MDR *Salmonella*: Prevalence of ESBLs Profiles and Resistance Genes in Strains Isolated from Food. *Microorganisms* **2022**, *10*, 780. [CrossRef]
- 78. Igbinosa, E.O.; Beshiru, A.; Igbinosa, I.H.; Cho, G.S.; Franz, C.M.A.P. Multidrug-resistant extended spectrum β-lactamase (ESBL)-producing *Escherichia coli* from farm produce and agricultural environments in Edo State, Nigeria. *PLoS ONE* 2023, 18, e0282835. [CrossRef] [PubMed]
- 79. Komodromos, D.; Kotzamanidis, C.; Giantzi, V.; Pappa, S.; Papa, A.; Zdragas, A.; Angelidis, A.; Sergelidis, D. Prevalence, Infectious Characteristics and Genetic Diversity of *Staphylococcus aureus* and Methicillin-Resistant *Staphylococcus aureus* (MRSA) in Two Raw-Meat Processing Establishments in Northern Greece. *Pathogens* 2022, 11, 1370. [CrossRef]
- Popa, S.A.; Morar, A.; Ban-Cucerzan, A.; Tîrziu, E.; Herman, V.; Imre, M.; Florea, T.; Morar, D.; Pătrînjan, R.-T.; Imre, K. First study in the frequency of isolation and phenotypic antimicrobial resistance profiles of pig and cattle origin Campylobacter strains in Romania. *Vet. Res. Commun.* 2024, 1–7. Available online: https://link.springer.com/article/10.1007/s11259-024-10360-w (accessed on 6 June 2024). [CrossRef] [PubMed]
- 81. Taneja, N.; Sharma, M. Antimicrobial resistance in the environment: The Indian scenario. *Indian J. Med. Res.* **2019**, 149, 119–128. [CrossRef] [PubMed]
- 82. Chowdhury, S.; Ghosh, S.; Aleem, M.A.; Parveen, S.; Islam, M.A.; Rashid, M.M.; Akhtar, A.; Chowdhury, F. Antibiotic Usage and Resistance in Food Animal Production: What Have We Learned from Bangladesh? *Antibiotics* **2021**, *10*, 1032. [CrossRef] [PubMed]
- 83. Silva, A.; Silva, V.; Pereira, J.E.; Maltez, L.; Igrejas, G.; Valentão, P.; Falco, V.; Poeta, P. Antimicrobial Resistance and Clonal Lineages of *Escherichia coli* from Food-Producing Animals. *Antibiotics* **2023**, *12*, 1061. [CrossRef] [PubMed]
- 84. Manyi-Loh, C.; Mamphweli, S.; Meyer, E.; Okoh, A. Antibiotic Use in Agriculture and Its Consequential Resistance in Environmental Sources: Potential Public Health Implications. *Molecules* **2018**, *23*, 795. [CrossRef] [PubMed]
- Page, M.J.; McKenzie, J.E.; Bossuyt, P.M.; Boutron, I.; Hoffmann, T.C.; Mulrow, C.D.; Shamseer, L.; Tetzlaff, J.M.; Akl, E.A.; Brennan, S.E.; et al. The PRISMA 2020 statement: An updated guideline for reporting systematic reviews. *BMJ* 2021, 372, n71. [CrossRef] [PubMed]
- Huedo-Medina, T.B.; Sánchez-Meca, J.; Marín-Martínez, F.; Botella, J. Assessing Heterogeneity in Meta-Analysis: Q Statistic or I 2 Index? *Psychol. Methods* 1998, 11, 193–206. [CrossRef] [PubMed]
- 87. Sterne, J.A.C.; Savović, J.; Page, M.J.; Elbers, R.G.; Blencowe, N.S.; Boutron, I.; Cates, C.J.; Cheng, H.Y.; Corbett, M.S.; Eldridge, S.M.; et al. RoB 2: A revised tool for assessing risk of bias in randomised trials. *BMJ* **2019**, *366*, 14898. [CrossRef]
- McGuinness, L.A.; Higgins, J.P.T. Risk-of-bias VISualization (robvis): An R package and Shiny web app for visualizing risk-of-bias assessments. *Res. Synth. Methods* 2021, 12, 55–61. Available online: https://onlinelibrary.wiley.com/doi/10.1002/jrsm.1411 (accessed on 11 June 2024). [CrossRef]

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