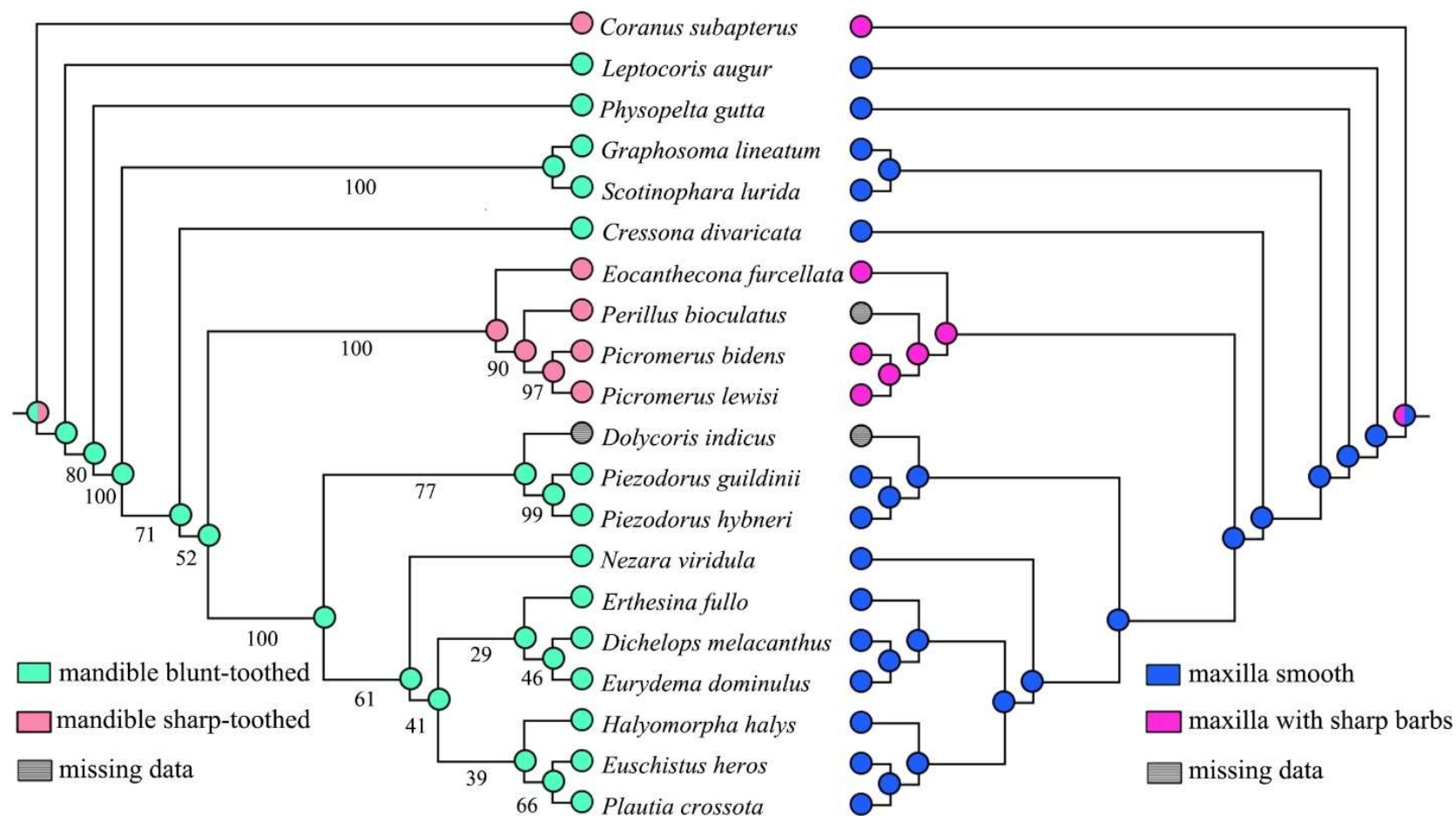


# Supplementary Materials

## Supplementary Figure S1



**Figure S1.** Ancestral state reconstruction of the shape of mandibles (left) and maxillae (right) in Pentatomidae based on parsimony. The cladogram used for reconstruction was the single tree obtained based on *cox1* and *rrnL* genes. The numbers under the branches are bootstrap values.

**Table S1–S11**

**Table S1.** Information of taxonomy, feeding habits, morphological references and genes *cox1* and *rrnL* accession numbers of all the species sampled in present study.

Family	Subfamily	Species	Feeding habit	Morphological reference	GenBank accession number <i>cox1</i>	<i>rrnL</i>
Largidae Amyot & Serville	-	<i>Physopelta gutta</i> (Burmeister, 1834)[1]	Phytophagous	[2]	EU427343.1	EU427343.1
Reduviidae Latreille	Harpactorinae Amyot & Serville	<i>Coranus subapterus</i> (De Geer, 1773)[3]	Predatory	[4]	KM022149.1	-
Rhopalidae Amyot & Serville	-	<i>Leptocoris augur</i> (Fabricius, 1781)[5]	Phytophagous	[6]	KX503052.1	KX523378.1
Pentatomidae Leach	Phyllocephalinae Amyot & Serville	<i>Cressona divaricata</i> Zheng & Zou, 1982*[7]	Phytophagous	present study	MK363262.1 KP142936.1 MZ673416	MZ676042
		<i>Eocanthecona furcellata</i> (Wolff, 1811)[8]	Predatory	present study	MK393432.1	-
		<i>Perillus bioculatus</i> (Fabricius, 1775)[9]	Predatory	[4,10,11]	MG398669.1	-
	Asopinae Amyot & Serville	<i>Picromerus bidens</i> (Linnaeus, 1758)[12]	Predatory	[13]	KJ541616.1	MT265462.1
		<i>Picromerus lewisi</i> Scott, 1874[14]	Predatory	[13]	KY710772.1	KC155950.1
		<i>Dichelops melacanthus</i> Dallas, 1851[15]	Phytophagous	[16]	JQ218483.1 JQ218506.1 MN257052.1 JQ218458.1	KC537022.1
		<i>Dolycoris indicus</i> Stål, 1876[17]	Phytophagous	[10,11]	KX467344.1	-
	Pentatominae Leach	<i>Erthesina fullo</i> (Thunberg, 1783)[18]	Phytophagous	[19]	MK374364.1	MK374364.1
		<i>Eurydema dominulus</i> (Scopoli, 1763)[20]	Phytophagous	present study	MG584833.1	MG584833.1
		<i>Euschistus heros</i> (Fabricius, 1798)[21]	Phytophagous	[16,22]	MN257053.1 KU892543.1	KU853769.1

Podopinae Amyot & Serville	<i>Halyomorpha halys</i> (Stål, 1855)[23]	Phytophagous	present study	NC_013272.1	NC_013272.1
	<i>Nezara viridula</i> (Linnaeus, 1758)[12]	Phytophagous	[16,24,25]	NC_011755.1	NC_011755.1
	<i>Piezodorus guildinii</i> (Westwood, 1837)[26]	Phytophagous	[16,22]	HQ985132.1	JX425407.1
				JX425420.1	
				JQ218490.1	
				JQ218510.1	
	<i>Piezodorus hybneri</i> (Gmelin, 1790)[27]	Phytophagous	[10,16]	KX467346.1	-
	<i>Plautia crossota</i> (Dallas, 1851)[15]	Phytophagous	present study	MK757497.1	MK757497.1
	<i>Graphosoma lineatum</i> (Linnaeus, 1758) [12]	Phytophagous	[4]	KM021441.1	-
				KX960066.1	
	<i>Scotinophara lurida</i> (Burmeister, 1834)[1]	Phytophagous	present study	MF497733.1	MF497733.1

\* Fragments of *coxI* and *rrnL* sequenced in present study.

**Table S2.** Taxonomic and collecting information of the stink bug species examined in present study.

Subfamily	Species	Locality	Date
Phyllocephalinae Amyot & Serville	<i>Cressona divaricata</i> Zheng & Zou, 1982	Qimaba, Honghe, Yunan, China	2012-V-20
Asopinae Amyot & Serville	<i>Eocanthecona furcellata</i> (Wolff, 1811)	Wenshan, Yunan, China	2020-VII-27
Pentatominae Leach	<i>Eurydema dominulus</i> (Scopoli, 1763)	Qifang Town, Xiangyang, Hubei, China	2018-VI-14
	<i>Halyomorpha halys</i> (Stål, 1855)	Haidian, Beijing, China	2018-X-23
	<i>Plautia crossota</i> (Dallas, 1851)	Tangfu County, HongKong, China	2019-V-12
Podopinae Amyot & Serville	<i>Scotinophara lurida</i> (Burmeister, 1834)	Fengtongzhai National Nature Reserve, Yaan, Sichuan, China	2018-VIII-29

**Table S3.** Host plant information of the stink bug species sampled in present study.

Subfamily	Species	Host plant family*
Phyllocephalinae Amyot & Serville	<i>Cressona divaricata</i> Zheng & Zou, 1982	Poaceae
	<i>Dichelops melacanthus</i> Dallas, 1851	Fabaceae
Pentatominae Leach	<i>Dolycoris indicus</i> Stål, 1876	Oleaceae
		Poaceae
		Apiaceae
		Asteraceae
		Brassicaceae
		Cannabaceae
		Chenopodiaceae
		Cuscutaceae
		Fabaceae
		Liliaceae
		Linaceae
		Malvaceae
		Plantaginaceae
		Poaceae
		Polygonaceae
	<i>Erthesina fullo</i> (Thunberg, 1783)	Santalaceae
		Solanaceae
		Anacardiaceae
		Apocynaceae
		Araliaceae
		Bignoniaceae
		Burseraceae
		Cannabinaceae
		Casuarinaceae
		Cupressaceae
		Ebenaceae
		Euphorbiaceae
		Fabaceae
		Hamamelidaceae
		Juglandaceae
		Lauraceae
		Magnoliaceae
		Malvaceae
		Meliaceae
		Mimosaceae
		Moraceae
		Myrtaceae
		Oxalidaceae
		Pinaceae
		Platanaceae
		Poaceae
		Proteaceae
		Punicaceae
		Rhamnaceae
		Rosaceae
		Rubiaceae
		Salicaceae
		Santalaceae
		Scrophulariaceae
		Simaroubaceae
		Ulmaceae
		Verbenaceae

<i>Eurydema dominulus</i> (Scopoli, 1763)	Apiaceae
	Balsaminaceae
	Brassicaceae
	Lamiaceae
	Scrophulariaceae
<i>Euschistus heros</i> (Fabricius, 1798)	Brassicaceae
	Fabaceae
	Oleaceae
	Solanaceae
<i>Halyomorpha halys</i> (Stål, 1855)	Amaranthaceae
	Basellaceae
	Ebanaceae
	Fabaceae
	Malvaceae
	Moraceae
	Oleaceae
	Rosaceae
	Rutaceae
	Solanaceae
	Vitaceae
<i>Nezara viridula</i> (Linnaeus, 1758)	Amaranthaceae
	Anacardiaceae
	Apiaceae
	Aquifoliaceae
	Asteraceae
	Brassicaceae
	Cannabaceae
	Capparaceae
	Chenopodiaceae
	Cupressaceae
	Euphorbiaceae
	Fagaceae
	Lamiaceae
	Liliaceae
	Linaceae
	Malvaceae
	Meliaceae
	Mimosaceae
	Oleaceae
	Pedaliaceae
	Plantaginaceae
	Poaceae
	Polygonaceae
	Rosaceae
	Rutaceae
	Scrophulariaceae
	Solanaceae
<i>Piezodorus guildinii</i> (Westwood, 1837)	Apiaceae
	Aquifoliaceae
	Brassicaceae
	Cucurbitaceae
	Fabaceae
	Linaceae
	Malpighiaceae
	Oleaceae
	Phytolaccaceae
	Rosaceae

	<i>Piezodorus hybneri</i> (Gmelin, 1790)	Solanaceae Anacardiaceae Fabaceae Poaceae Verbenaceae
	<i>Plautia crossota</i> (Dallas, 1851)	Anacardiaceae Asteraceae Basellaceae Brassicaceae Cannaceae Caprifoliaceae Convolvulaceae Fabaceae Moraceae Poaceae Rhamnaceae Rubiaceae Santalaceae Scrophulariaceae Solanaceae Tiliaceae Verbenaceae
Podopinae Amyot & Serville	<i>Graphosoma lineatum</i> (Linnaeus, 1758)	Apiaceae Asteraceae
	<i>Scotinophara lurida</i> (Burmeister, 1834)	Cyperaceae Marsileaceae Poaceae Pontederiaceae

Information of host plant family was summarized from the online database Pentatomoidea Home Page, with the related literatures available on <https://www.ndsu.edu/pubweb/~rider/Pentatomoidea/>.

**Table S4.** Morphometric data of labial and antennal sensilla in *Cressona divaricata* Zheng & Zou, 1982.

Sensilla type	Length (μm)	Basal Diameter (μm)	Distribution
AnCh I	71.62 ± 11.88	6.37 ± 1.84	Sc, BPd, DPd, Bf, Df.
AnTr I	58.67 ± 6.70	4.25 ± 0.66	Bf, Df.
AnTr II	44.80 ± 5.23	2.73 ± 0.45	Bf, Df.
AnBa I	15.28 ± 0.88	2.40 ± 0.26	Bf, Df.

Data are mean ± SD values obtained from scanning electron microscopy based on about 20 antennal sensilla of each type selected from antennal scape to distiflagellomere of males and females. AnBa I, antennal sensilla basiconica I; AnCh I, antennal sensilla chaetica I; AnTr I–II, antennal sensilla trichodea I–II; Bf, basiflagellomere; BPd, basal pedicel; Df, distiflagellomere; DPd, distal pedicel; Sc, scape.

**Table S5.** Morphometric data of labial and antennal sensilla in *Eocanthecona furcellata* (Wolff, 1811).

Sensilla type	Length (μm)	Basal Diameter (μm)	Distribution
AnCh I	44.62 ± 2.50	4.16 ± 0.67	Sc, BPd, DPd, Bf, Df.
AnCh II	43.18 ± 8.01	3.38 ± 0.69	Sc, BPd.
LaTr I	44.59 ± 8.21	2.14 ± 0.63	La.
LaTr II	21.94 ± 2.79	3.16 ± 0.99	La.
AnTr I	47.77 ± 2.93	3.43 ± 0.87	DPd, Bf, Df.

AnTr II	37.67 ± 1.08	1.41 ± 0.15	DPd, Bf, Df.
LaBa I	19.86 ± 1.22	3.14 ± 0.63	La.
LaBa II	7.07 ± 2.02	2.19 ± 0.24	La.
LaBa III	22.15 ± 1.68	8.36 ± 1.91	DPd, Bf, Df.
AnBa I	13.15 ± 2.75	1.93 ± 0.10	DPd, Bf, Df.
AnCo I	2.37 ± 0.08	1.63 ± 0.05	Df.
AnCo II	-	0.35 ± 0.06	Df.

Data are mean ± SD values obtained from scanning electron microscopy based on about 20 antennal sensilla of each type selected from antennal scape to distiflagellomere of males and females. Antennal sensilla coeloconica I and II are tiny in size, few in number and sparsely scattered on distiflagellomere; therefore, their measurements were got from all the observed ones (around 3 to 5). AnBa I, antennal sensilla basiconica I; AnCh I–II, antennal sensilla chaetica I–II; AnCo I–II, antennal sensilla coeloconica I–II; AnTr I–II, antennal sensilla trichodea I–II; LaBa I–III, labial sensilla basiconica I–III; Bf, basiflagellomere; BPd, basal pedicel; Df, distiflagellomere; DPd, distal pedicel; LaBa I–III, labial sensilla basiconica I–III; LaTr I–II, labial sensilla trichodea I–II; Sc, scape.

**Table S6.** Morphometric data of labial and antennal sensilla in *Eurydema dominulus* (Scopoli, 1763).

Sensilla type	Length (µm)	Basal Diameter (µm)	Distribution
AnCh I	46.80 ± 5.50	4.22 ± 0.24	Sc, BPd, DPd, Bf, Df.
LaTr I	44.01 ± 4.52	3.54 ± 0.12	La.
LaTr II	23.01 ± 1.65	3.42 ± 0.30	La.
AnTr I	34.40 ± 2.71	3.23 ± 0.43	Bf, Df.
AnTr II	32.36 ± 2.83	1.51 ± 0.29	Bf, Df.
LaBa I	11.97 ± 1.61	3.36 ± 0.14	La.
LaBa II	6.93 ± 0.82	2.52 ± 0.30	La.
AnBa I	10.87 ± 0.92	2.46 ± 0.30	Bf, Df.

Data are mean ± SD values obtained from scanning electron microscopy based on about 20 antennal sensilla of each type selected from antennal scape to distiflagellomere of males and females. AnBa I, antennal sensilla basiconica I; AnCh I, antennal sensilla chaetica I; AnTr I–II, antennal sensilla trichodea I–II; Bf, basiflagellomere; BPd, basal pedicel; Df, distiflagellomere; DPd, distal pedicel; LaBa I–II, labial sensilla basiconica I–II; LaTr I–II, labial sensilla trichodea I–II; Sc, scape.

**Table S7.** Morphometric data of labial and antennal sensilla in *Halyomorpha halys* (Stål, 1855).

Sensilla type	Length (µm)	Basal Diameter (µm)	Distribution
AnCh I	52.12 ± 7.49	4.38 ± 0.62	Sc, BPd, DPd, Bf, Df.
LaTr I	38.60 ± 9.17	2.85 ± 0.26	La.
LaTr II	24.11 ± 9.36	2.49 ± 0.07	La.
AnTr I	49.03 ± 5.40	3.51 ± 0.45	BPd, DPd, Bf, Df.
AnTr II	38.68 ± 2.28	1.74 ± 0.41	BPd, DPd, Bf, Df.
LaBa I	12.95 ± 2.80	3.59 ± 0.22	La.
LaBa II	6.44 ± 1.70	2.95 ± 0.33	La.
AnBa I	19.50 ± 1.09	2.30 ± 0.20	DPd, Bf, Df.
AnBa II	11.74 ± 1.08	2.48 ± 0.10	DPd, Bf, Df.

AnCo I	3.71 ± 0.03	3.35 ± 0.20	Df.
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Data are mean ± SD values obtained from scanning electron microscopy based on about 20 antennal sensilla of each type selected from antennal scape to distiflagellomere of males and females. Antennal sensilla coeloconica I are tiny in size, few in number and sparsely scattered on distiflagellomere; therefore, their measurements were got from all the observed ones (around 3 to 5). AnBa I–II, antennal sensilla basiconica I–II; AnCh I, antennal sensilla chaetica I; AnCo I, antennal sensilla coeloconica I; AnTr I–II, antennal sensilla trichodea I–II; Bf, basiflagellomere; BPd, basal pedicel; Df, distiflagellomere; DPd, distal pedicel; LaBa I–II, labial sensilla basiconica I–II; LaTr I–II, labial sensilla trichodea I–II; Sc, scape.

**Table S8.** Morphometric data of labial and antennal sensilla in *Plautia crossota* (Dallas, 1851).

Sensilla type	Length (μm)	Basal Diameter (μm)	Distribution
AnCh I	61.95 ± 6.17	4.16 ± 0.76	Sc, BPd, DPd, Bf, Df.
LaTr I	56.28 ± 5.49	2.76 ± 0.23	La.
LaTr II	20.47 ± 1.29	3.10 ± 0.60	La.
AnTr I	41.75 ± 6.08	3.11 ± 0.12	DPd, Bf, Df.
AnTr II	35.98 ± 3.09	1.66 ± 0.26	DPd, Bf, Df.
LaBa I	14.37 ± 0.75	3.89 ± 0.22	La.
LaBa II	10.96 ± 1.12	3.14 ± 0.23	La.
AnBa I	17.51 ± 1.21	1.48 ± 0.15	DPd, Bf, Df.
AnBa II	11.65 ± 0.83	1.93 ± 0.30	DPd, Bf, Df.
AnCo I	2.07 ± 0.08	1.43 ± 0.09	Df.
AnCo II	-	0.54 ± 0.02	Df.

Data are mean ± SD values obtained from scanning electron microscopy based on about 20 antennal sensilla of each type selected from antennal scape to distiflagellomere of males and females. Antennal sensilla coeloconica I and II are tiny in size, few in number and sparsely scattered on distiflagellomere; therefore, their measurements were got from all the observed ones (around 3 to 5). AnBa I–II, antennal sensilla basiconica I–II; AnCh I, antennal sensilla chaetica I; AnCo I–II, antennal sensilla coeloconica I–II; AnTr I–II, antennal sensilla trichodea I–II; Bf, basiflagellomere; BPd, basal pedicel; Df, distiflagellomere; DPd, distal pedicel; LaBa I–II, labial sensilla basiconica I–II; LaTr I–II, labial sensilla trichodea I–II; Sc, scape.

**Table S9.** Morphometric data of labial and antennal sensilla in *Scotinophara lurida* (Burmeister, 1834).

Sensilla type	Length (μm)	Basal Diameter (μm)	Distribution
AnCh I	63.48 ± 4.63	3.17 ± 0.42	Sc, BPd, DPd, Bf, Df.
AnTr I	67.85 ± 9.17	3.69 ± 0.63	Bf, Df.
AnTr II	43.81 ± 3.41	2.09 ± 0.54	Bf, Df.
AnBa I	17.59 ± 2.43	2.30 ± 0.40	Bf, Df.
AnCo II	-	0.90 ± 0.09	Df.

Data are mean ± SD values obtained from scanning electron microscopy based on about 20 antennal sensilla of each type selected from antennal scape to distiflagellomere of males and females. Antennal sensilla coeloconica II are tiny in size, few in number and sparsely scattered on distiflagellomere; therefore, their measurements were got from all the observed ones (around 3 to 5). AnBa I, antennal sensilla basiconica I; AnCh I, antennal sensilla chaetica I; AnCo II, antennal sensilla coeloconica II; AnTr I–II, antennal sensilla trichodea I–II; Bf, basiflagellomere; BPd, basal pedicel; Df, distiflagellomere; DPd, distal pedicel; Sc, scape.



**Table S10.** Average density of basiconica sensilla on antennal distiflagellomere of the stink bug species examined in present study.

Species	Density (/10 <sup>4</sup> μm <sup>2</sup> )
<i>Cressona divaricata</i> Zheng & Zou, 1982	1.25 ± 0.32
<i>Eocanthecona furcellata</i> (Wolff, 1811)	22.63 ± 7.48
<i>Eurydema dominulus</i> (Scopoli, 1763)	3.10 ± 1.67
<i>Halyomorpha halys</i> (Stål, 1855)	5.10 ± 0.76
<i>Plautia crossota</i> (Dallas, 1851)	7.74 ± 1.88
<i>Scotinophara lurida</i> (Burmeister, 1834)	5.59 ± 1.66

Data are mean ± SD values obtained from scanning electron microscopy. The densities were calculated by averaging the number of basiconica sensilla in two randomly selected 1000 μm<sup>3</sup> quadrats on the electron microscopic images from males and females.

**Table S11.** Morphological characters used for ancestral state reconstruction.

Family	Subfamily	Species	Mandible (0: sharp-toothed; 1: blunt toothed)	Maxilla (0: smooth; 1: with sharp barbs)	Reference
Largidae Amyot & Serville	-	<i>Physopelta gutta</i> (Burmeister, 1834)	0	0	[2]
Reduviidae Latreille	Harpactorinae Amyot & Serville	<i>Coranus subapterus</i> (De Geer, 1773)	1	1	[4]
Rhopalidae Amyot & Serville	-	<i>Leptocoris augur</i> (Fabricius, 1781)	0	0	[6]
Pentatomidae Leach	Phyllocephalinae Amyot & Serville	<i>Cressona divaricata</i> Zheng & Zou, 1982	0	0	present study
	Asopinae Amyot & Serville	<i>Eocanthecona furcellata</i> (Wolff, 1811)	1	1	present study
		<i>Perillus bioculatus</i> (Fabricius, 1775)	1	?	[4]
		<i>Picromerus bidens</i> (Linnaeus, 1758)	1	1	[13]
		<i>Picromerus lewisi</i> Scott, 1874	1	1	[13]
	Pentatominae Leach	<i>Dichelops melacanthus</i> Dallas, 1851	0	0	[16]
		<i>Dolycoris indicus</i> Stål, 1876	?	?	N.A. (with antennal characters available for this species)

<i>Erthesina fullo</i> (Thunberg, 1783)	0	0	[19]
<i>Eurydema dominulus</i> (Scopoli, 1763)	0	0	present study
<i>Euschistus heros</i> (Fabricius, 1798)	0	0	[16]
<i>Halyomorpha halys</i> (Stål, 1855)	0	0	present study
<i>Nezara viridula</i> (Linnaeus, 1758)	0	0	[16]
<i>Piezodorus guildinii</i> (Westwood, 1837)	0	0	[16]
<i>Piezodorus hybneri</i> (Gmelin, 1790)	0	0	[16]
<i>Plautia crossota</i> (Dallas, 1851)	0	0	present study
<i>Graphosoma lineatum</i> (Linnaeus, 1758)	0	0	[4]
<i>Scotinophara lurida</i> (Burmeister, 1834)	0	0	present study

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