



Review

The Role of Ferritin in Health and Disease: Recent Advances and Understandings

Nikhil Kumar Kotla, Priyata Dutta Sanjana Parimi and Nupur K. Das *

Department of Molecular and Integrative Physiology, University of Michigan, Ann Arbor, MI 48109, USA; nkotla@med.umich.edu (N.K.K.); priyatadutta576@gmail.com (P.D.); saparimi@umich.edu (S.P.)

* Correspondence: nkdas@med.umich.edu; Tel.: +1-734-6155041

Abstract: Systemic iron homeostasis needs to be tightly controlled, as both deficiency and excess iron cause major global health concerns, such as iron deficiency anemia, hemochromatosis, etc. In mammals, sufficient dietary acquisition is critical for fulfilling the systemic iron requirement. New questions are emerging about whether and how cellular iron transport pathways integrate with the iron storage mechanism. Ferritin is the intracellular iron storage protein that stores surplus iron after all the cellular needs are fulfilled and releases it in the face of an acute demand. Currently, there is a surge in interest in ferritin research after the discovery of novel pathways like ferritinophagy and ferroptosis. This review emphasizes the most recent ferritin-related discoveries and their impact on systemic iron regulation.

Keywords: ferritin; ferritinophagy; iron disorders; ferroptosis; COVID-19



Citation: Kotla, N.K.; Dutta, P.; Parimi, S.; Das, N.K. The Role of Ferritin in Health and Disease: Recent Advances and Understandings. *Metabolites* **2022**, *12*, 609. <https://doi.org/10.3390/metabo12070609>

Academic Editors: Immacolata Andolfo and Roberta Russo

Received: 31 May 2022

Accepted: 27 June 2022

Published: 30 June 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Iron is essential for almost all living organisms on earth. Its unique ability to donate and accept electrons makes it favorable for fundamental biological processes such as DNA synthesis and repair, oxygen transport, cellular respiration, heme synthesis, and immune function [1]. Although cellular iron deficiency is detrimental, iron excess is also potentially hazardous as its divalent form (Fe^{2+}) generates toxic reactive oxygen species (ROS) via the Fenton reaction causing oxidative damage to the cells [2]. From the human health perspective, an iron imbalance is a major global concern, and both deficiency and overload disorders are prevalent in distinct geographical locations as well as in different genetic backgrounds [3]. In addition, patients with hereditary hemoglobinopathies, such as sickle cell disease and thalassemia, exhibit a complex form of iron disorders that represent both anemia and progressive tissue iron overload consequent to chronic hemolysis [4–6]. Mammalian iron metabolism is tightly regulated both at the cellular and systemic levels [7]. Although our body can extensively recycle its own iron, a regular supply of iron through diet (daily requirement is 10–20 mg, of which about 10% is absorbed) is critical for efficient homeostasis [1]. Dietary iron is absorbed by the duodenal enterocytes with the help of three main transporters, namely, the apical transporters divalent metal transporter 1 (DMT1), duodenal cytochrome b reductase (Dcytb), and the basolateral transporter ferroportin 1 (Fpn1) [8–10].

One of the most important components of cellular or organismal iron homeostasis is iron storage. Intracellular iron storage function is carried out by ferritin, which is structurally composed of 24 subunits of light (FTL) and heavy chains (FTH) that form a nano-cage complex to hold up to 4500 iron atoms [11]. Ferritin sequesters excess intracellular iron and stores it in a redox-inactive form for future use in conditions of deficiency or high demand. Cellular and systemic ferritin levels are not only crucial indicators of iron status but are also important markers of inflammatory [12], immunological [13], and malignant disorders [14,15]. Genetic alterations of ferritin are often associated with severe

pathologies, and homozygous FTH deletion in mice is incompatible with life [16]. Conditional deletion of FTH in the intestine leads to iron overload in mice characterized by intestinal hyperabsorption and dysregulated systemic control by hepatic hepcidin highlighting its requirement for accurate intestinal iron absorption [17]. On the other hand, macrophage-specific FTH deletion in mice reduces intracellular iron levels and protects from high fat diet-induced metabolic disorders by lowering inflammatory and prooxidant damages [18]. In humans, a rare FTH mutation was identified in a Japanese family suffering from iron overload [19], and mutations in the FTL gene cause various ferritin disorders, e.g., hereditary hyperferritinemia with cataract syndrome, neuroferritinopathy, benign hyperferritinemia, etc. [20].

Intracellular iron homeostasis is mediated at both transcriptional and translational levels by iron regulatory proteins (IRPs) and iron-responsive elements (IREs) [21]. IRP has two subtypes, IRP1 and IRP2, both regulated by cellular iron and oxygen levels [21–23]. During low iron conditions, increased IRP levels lead to its enhanced interaction with the IREs present on either 5'-UTR or 3'-UTR of the mRNAs of iron-related genes (e.g., transferrin receptor 1(TfR1), DMT1, Fpn1, etc.). IRP-IRE interaction at 5'-UTR of both ferritin L and H mRNAs inhibits ferritin translation [24]. In addition to being a marker of iron status, ferritin is also one of the most important biological markers of inflammation. During inflammation, there is cytokine-mediated induction of ferritin H and L synthesis at both transcriptional and translational levels [25].

In recent years, we have gained more insight regarding ferritin function in the contexts of intracellular iron dynamics among different compartments, novel iron-dependent mechanisms related to cell growth and viability, and new diagnostic or prognostic indices related to infectious and/or inflammatory disorders, especially COVID-19. This review will discuss a few seminal discoveries that reshaped our understanding of ferritin's role in biology.

2. Cytosolic Chaperones for Delivering Iron into Ferritin

The fate of intracellular iron is determined by the delicate balance between cellular iron usage and surplus. To ensure this, metabolically accessible iron, also known as cytosolic labile iron pool (LIP), must be efficiently distributed to its sites of utilization, transport, or storage. Although the mechanisms of ferritin regulation (e.g., translational regulation via iron response element (IRE)-iron regulatory protein (IRP) interaction at the 5'UTR) in response to cytosolic LIP status are well defined [24,26,27], the recent discovery of a cytosolic trafficking system that delivers iron to ferritin has significantly advanced our concepts of LIP dynamics. A member of the poly(rC)-binding protein (PCBP) family, initially known to be involved in the stabilization and translational regulation of RNAs containing C-rich motifs, PCBP1 was the first to be identified as a cytosolic iron chaperone that directly binds iron and loads it onto ferritin [28] (Figure 1). The ferritin-binding activity of PCBP1 is negatively regulated by cytosolic iron levels [29]. Subsequently, other PCBP members, namely PCPB2, PCPB3, and PCPB4, were also found to have iron chaperone activity with variable efficiency in iron delivery to ferritin [30]. Of note, PCBPs also play roles in the metallation of several iron-dependent enzymes: prolyl hydroxylases (PHDs), asparaginyl hydroxylase (FIH1), and deoxyhypusine hydroxylase (DOHH), highlighting their critical role in LIP distribution for cellular processes [30,31].

PCBP1 plays a critical role in systemic iron homeostasis, as hepatocyte-specific PCBP1 knockout (KO) mice show low hepatic ferritin levels, mitochondrial dysfunction, and ROS-mediated lipid peroxidation [32,33]. The direct involvement of PCBP in intestinal ferritin regulation is not clear. However, PCBPs might indirectly regulate enterocyte iron levels by (a) delivering iron to PHDs that in turn modulate intestinal HIF-2 α levels and (b) PCBP2 binding to both DMT1 and Fpn1 to promote iron cellular uptake [34]. More recently, it was shown that PCBP1 and 2 are essential genes for mouse development [35] and erythroid differentiation [36]. In addition, PCBP1 and nuclear receptor co-activator 4 (NCOA4) co-operatively regulate erythroid iron storage and erythropoiesis (discussed below).

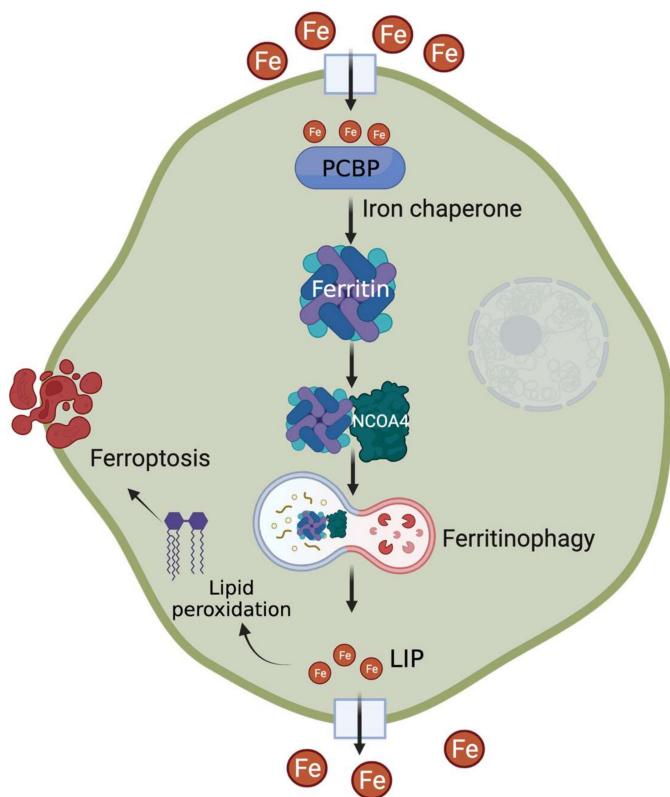


Figure 1. Recent advances in understanding the regulation of cellular iron storage mechanism. After cellular uptake, PCBPs act as iron chaperones to deliver iron to ferritin. During conditions of increased demand, NCOA4 acts as a cargo receptor for lysosomal degradation of ferritin, followed by increased labile iron pool (LIP) level. Excess iron contributes to lipid peroxidation-mediated ferroptosis.

In addition to regulating intracellular iron trafficking, PCBPs are involved in post-transcriptional gene regulation, maintenance of mitochondrial stability, and are associated with a wide variety of pathophysiologies such as rheumatoid arthritis [37], amyotrophic lateral sclerosis (ALS) [38], and Huntington's disease (HD), and certain neurodevelopmental disorders [39]. Although the iron chaperone and RNA-binding functions of PCBPs have been shown to be independently essential [40], it is not clear how these functions potentially contribute to the pathogenesis of the above-mentioned disorders.

3. Ferritinophagy—For Efficient Iron Release from the Store

Iron needs to be released from its storage form during conditions of scarcity or high demand. This involves proteolytic degradation of ferritin occurring primarily in the lysosome [41]. Earlier studies indicated that autophagic machinery are involved in ferritin degradation and subsequent iron release. More recently, NCOA4, originally known as nuclear receptor co-activator, was identified as a cargo receptor targeting ferritin for lysosomal degradation—a process coined as ferritinophagy [42]. NCOA4 colocalizes with ferritin in autophagosomes and lysosomes, and its *in vitro* or *in vivo* depletion inhibits ferritin delivery and degradation (Figure 1). The consequent reduction in ferritin turnover leads to a significant decrease in cytosolic LIP that ultimately alters cellular and systemic iron homeostasis [43,44]. NCOA4 expression and function are tightly regulated by intracellular iron status: (a) excess iron targets NCOA4 for ubiquitin-dependent turnover via its iron-dependent HERC2-mediated proteolysis [45]; (b) low iron increases NCOA4 mRNA levels via HIF-dependent transcriptional regulation [46,47].

NCOA4-mediated ferritin degradation is critical for erythroid differentiation and hemoglobinization in mice as these processes rely on efficient iron delivery and turnover in the RBCs [45,48]. In addition, both PCBP-mediated iron flux through ferritin and NCOA4-

mediated ferritinophagic iron release occurs in a sequentially coordinated manner throughout the process of erythroid differentiation to maintain a consistent iron supply [48,49]. Whole-body NCOA4 KO mice develop microcytic hypochromic anemia, which is more severe in young mice compared to adults [47,50,51]. A recent study in erythroid-specific and tamoxifen-inducible systemic NCOA4 deletion mouse models reveals that NCOA4 maintains erythropoiesis by both cell-autonomous and non-autonomous mechanisms that explain how the compensatory mechanisms operate in age- or tissue-specific NCOA4 ablation [51].

NCOA4 plays a critical role in liver iron homeostasis in a HIF-dependent manner (both HIF1 α - and HIF-2 α), as evidenced by significant impairment in iron mobilization from ferritin in phlebotomized or iron-deficient hepatocyte-specific NCOA4 KO mice [46]. Additionally, our work shows that intestinal NCOA4 is exclusively induced by HIF-2 α stabilizing conditions, such as dietary iron deficiency and chemically induced hemolytic anemia, and modulation of the intestinal NCOA4-HIF-2 α axis could be therapeutically used for attenuation of systemic iron overload [47,52]. It is important to note here that although the role of NCOA4 in cellular and systemic iron mobilization is well studied in human cell lines and various mouse models, the role of NCOA4 in human iron disorders has not been well studied. In addition, although the significance of autophagy in cancer is well established and altered NCOA4 expression is reported in various cancer types, the role of ferritinophagy in tumorigenesis is yet to be clearly defined.

4. Ferroptosis—From the Perspectives of PCBP and NCOA4

Ferroptosis is an iron-dependent non-apoptotic form of cell death characterized by the accumulation of lipid peroxides [53]. Perturbations in cellular iron homeostasis via alteration of LIP levels and/or dysregulation in the network of iron-related proteins alter the cellular sensitivity to ferroptosis (Figure 1). Ferroptosis can be inhibited by limiting intracellular iron availability [54,55] and can be induced in vitro or in vivo by high iron [53,56]. As expected, cellular FTH expression profiles can be helpful in determining their sensitivity to ferroptosis and can be used for therapeutic targeting of ferroptotic pathways [57–60]. Studies show that induction of ferritinophagy promotes ferroptosis by increasing the levels of redox-active iron, which can be suppressed by NCOA4 ablation [61–65]. In contrast to hypoxic induction of ferritinophagy in the liver and the intestine [46,47], hypoxia increases ferritin expression in primary human macrophages via miRNA-mediated ferritinophagy inhibition and protects the cells from ferroptosis [66]. Several clinical studies suggest that high ferritin levels and altered ferroptosis gene expression are associated with poor prognosis of malignant and neurodegenerative disorders [67–69]. In addition, modulation of ferritinophagy to regulate cellular ferroptosis sensitivity offers novel avenues for cancer, cardiovascular and metabolic disorder management [70–72]. For example, itaconate, a well-known immunomodulator compound, has recently been identified as a novel inducer of ferritinophagy, which suppresses tumor growth in mice [73]. As iron chaperones, PCBPs provide redox-active iron to the downstream players and thus act as a positive regulator of ferroptosis [74]. Several studies also show that PCBPs limit ferritinophagy, and in vitro or in vivo depletion of PCBP induces ferroptosis [32,75,76]. From the translational standpoint, both iron chaperone and ferritinophagy functions are attractive therapeutic targets for disorders like cancer and neurodegeneration [77].

5. Mammalian Ferritin and the Gut Microbiome

The mammalian gastrointestinal tract is a host to trillions of microorganisms, including viruses, bacteria, fungi, and protozoa, collectively known as gut microbiota [78]. The microbes co-evolved with their mammalian hosts over thousands of years to develop a mutually commensal relationship and significantly contribute to hosting nutrient homeostasis and metabolism [79]. Gut microbiota has been shown to be directly involved in a wide variety of diseases, such as obesity [80], hypertension [81], inflammatory bowel diseases (IBDs) [82], and autism [83]. Through metabolic utilization of host-derived nutrients, gut

microbiota produces a vast number of metabolites and small molecules, which interact with the host not only locally but also peripherally after their systemic absorption [84,85].

As regular and adequate iron supply through diet is critical for both, intense competition for the metal prevails between the host and microbes. Numerous studies have shown that iron availability influences the gut microbiome composition and function [86–90]. Human studies representing various geographic and pathologic backgrounds show that modulation of dietary iron levels influences links gut microbiome with the etiopathogenesis of a variety of disorders like dysbiosis, inflammation, and colorectal cancer [91–94]. For example, a recent human study involving nonalcoholic fatty liver disease (NAFLD) patients has shown that a positive association between serum ferritin levels and high liver fat accumulation is strongly connected with the gut microbiome: (a) host serum ferritin has both negative and positive associations with distinct bacterial families; (b) ferritin is connected to host-microbiota crosstalk on amino acid sensing pathways; (c) fecal microbiota transplantation (FMT) of human-derived bacteria in mice altered their iron metabolism gene signature [95].

Although the essentiality of iron for gut microbial survival in host intestinal lumen is well established, host-microbiota crosstalk on iron acquisition was only discovered recently. Deschemin et al. provided the first evidence that host intestinal iron sensing pathways are directly regulated by the commensals [96]. Our work demonstrated that gut microbial regulation of host systemic iron homeostasis takes place via microbial metabolite-mediated inhibition of intestinal HIF-2 α signaling [97]. It is worth mentioning here that HIF-2 α , an oxygen- and iron-dependent transcription factor, is the master regulator of intestinal iron absorption [98]. Strikingly, host intestinal as well as peripheral tissue ferritin expression is undetectable or significantly downregulated in germ-free or antibiotic-treated mice, which promptly returns to basal level following FMT from healthy donor mice [96,97]. These findings suggest that microbial presence in the gut is necessary for basal expression of host ferritin. Furthermore, recovery of ferritin levels in the microbiota-depleted mice by the systemic introduction of bacterial metabolites establishes the role of microbial metabolites in the regulation of host iron storage pathways [97] (Figure 2).

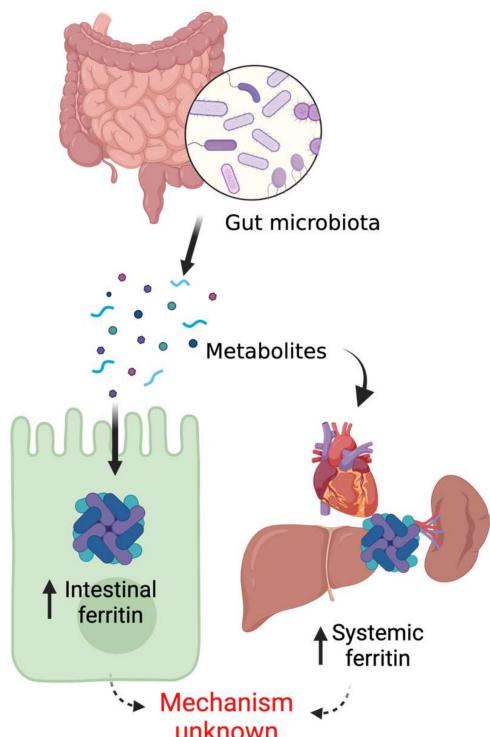


Figure 2. Gut microbial regulation of host ferritin.

Ferritin is an important indicator of cellular redox balance, which is frequently altered in a wide spectrum of diseases, including chronic inflammation, metabolic disorders, and cancer [99,100]. Identification of specific microbial communities and bacteria-derived metabolites that modulate host ferritin would be potentially beneficial for pathophysiological understanding as well as disease management. Successful development of *in vitro* co-culture with mammalian cells and *in vivo* colonization of germ-free gut models are needed to pinpoint the precise mechanisms of how gut microbiota or microbially derived metabolites regulate mammalian ferritin pathways.

6. Ferritin as a Novel Marker for COVID-19 Prognosis and Disease Severity

The ongoing global pandemic of coronavirus 2019 disease (COVID-19) caused by the acute respiratory syndrome coronavirus 2 (SARS-CoV-2) presents with heterogeneous clinical symptoms widely ranging from asymptomatic infection to fatal respiratory failure [101]. In addition to severe systemic inflammation caused by complex immunological changes, COVID-19 patients often present with dysregulated coagulation cascade system leading to further cardiovascular decompensation [102]. Massive systemic release of pro-inflammatory mediators such as interleukin (IL)-1 β , IL-2, IL-6, IL-7, IL-10, tumor necrosis factor- α (TNF α), granulocyte colony-stimulating factor (G-CSF), etc., also known as cytokine storm [103], contributes to the acute respiratory distress syndrome (ARDS) [104]. In addition, many of these patients present with hyperferritinemia, which may result from hemolysis caused by SARS-CoV-2-induced inflammation [105,106]. Considering the combined presence of excessive cytokine release and hyperferritinemia on many occasions, it was proposed that COVID-19 should be considered as a spectrum of the hyperferritinemic syndromes [107–109].

Progressively high ferritin values, along with other inflammatory biomarkers, have been consistently reported to be associated with increased disease severity and mortality [110–113]. Recently, a scoring system for predicting the disease severity, namely, the Severe COVID Prediction Estimate (SCOPE) score, has been developed, which is composed of circulating levels of ferritin, C-reactive protein, D-dimers, and IL-6. SCOPE score values of 6 or more predict the progression to severe respiratory failure or death within 2 weeks of hospital admission for pneumonia [114]. Also, serum ferritin levels dropped in patients treated with tocilizumab, an IL-6 receptor antibody [115]. Interestingly, a study reports that patients with higher ferritin levels (than the median value of 1419 μ g/L) benefitted most from tocilizumab treatment [116].

Altogether, these results establish ferritin assessment is a useful prognostic indicator that can be used as a screening test for the prediction of disease severity and mortality.

7. Conclusions and Perspectives

Ferritin plays essential roles in cellular and systemic iron homeostasis. As the iron storage protein and also as an acute phase reactant during inflammation, ferritin has been extensively studied. Over the last 15 years, several outstanding questions regarding ferritin–iron dynamics have been answered. Together, these mechanisms have allowed us to get fresh perspectives on iron-related disorders as well as infection and cancers. Sustained efforts to identify novel ferritin regulatory mechanisms will help us develop newer strategies for disease management.

Author Contributions: Conceptualization, N.K.D.; writing—original draft preparation, N.K.K., P.D., S.P., N.K.D.; writing—review and editing, N.K.K., N.K.D.; supervision, N.K.D. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Dev, S.; Babitt, J.L. Overview of iron metabolism in health and disease. *Hemodial. Int.* **2017**, *21* (Suppl S1), S6–S20. [[CrossRef](#)] [[PubMed](#)]
- Kappus, H. Oxidative stress in chemical toxicity. *Arch. Toxicol.* **1987**, *60*, 144–149. [[CrossRef](#)] [[PubMed](#)]
- Clasen, S.J.; Ley, R.E. Iron Deficiency Causes Wrinkles in Host-Commensal Relationships. *Cell Metab.* **2020**, *31*, 8–10. [[CrossRef](#)] [[PubMed](#)]
- Das, N.; Xie, L.; Ramakrishnan, S.K.; Campbell, A.; Rivella, S.; Shah, Y.M. Intestine-specific Disruption of Hypoxia-inducible Factor (HIF)-2alpha Improves Anemia in Sickle Cell Disease. *J. Biol. Chem.* **2015**, *290*, 23523–23527. [[CrossRef](#)] [[PubMed](#)]
- Anderson, E.R.; Taylor, M.; Xue, X.; Ramakrishnan, S.K.; Martin, A.; Xie, L.; Bredell, B.X.; Gardenghi, S.; Rivella, S.; Shah, Y.M. Intestinal HIF2alpha promotes tissue-iron accumulation in disorders of iron overload with anemia. *Proc. Natl. Acad. Sci. USA* **2013**, *110*, E4922–E4930. [[CrossRef](#)]
- Kato, G.J.; Piel, F.B.; Reid, C.D.; Gaston, M.H.; Ohene-Frempong, K.; Krishnamurti, L.; Smith, W.R.; Panepinto, J.A.; Weatherall, D.J.; Costa, F.F.; et al. Sickle cell disease. *Nat. Rev. Dis. Primers* **2018**, *4*, 18010. [[CrossRef](#)]
- Andrews, N.C. Molecular control of iron metabolism. *Best Pract. Res. Clin. Haematol.* **2005**, *18*, 159–169. [[CrossRef](#)]
- Fleming, R.E.; Migas, M.C.; Zhou, X.; Jiang, J.; Britton, R.S.; Brunt, E.M.; Tomatsu, S.; Waheed, A.; Bacon, B.R.; Sly, W.S. Mechanism of increased iron absorption in murine model of hereditary hemochromatosis: Increased duodenal expression of the iron transporter DMT1. *Proc. Natl. Acad. Sci. USA* **1999**, *96*, 3143–3148. [[CrossRef](#)]
- Frazer, D.M.; Anderson, G.J. Intestinal Iron Transport and its Regulation. *Hematology* **2001**, *6*, 193–203. [[CrossRef](#)]
- McKie, A.T.; Marciani, P.; Rolfs, A.; Brennan, K.; Wehr, K.; Barrow, D.; Miret, S.; Bomford, A.; Peters, T.J.; Farzaneh, F.; et al. A novel duodenal iron-regulated transporter, IREG1, implicated in the basolateral transfer of iron to the circulation. *Mol. Cell* **2000**, *5*, 299–309. [[CrossRef](#)]
- Melino, G.; Stefanini, S.; Chiancone, E.; Antonini, E. Stoichiometry of iron oxidation by apoferritin. *FEBS Lett.* **1978**, *86*, 136–138. [[CrossRef](#)]
- Cacoub, P.; Choukroun, G.; Cohen-Solal, A.; Luporsi, E.; Peyrin-Biroulet, L.; Peoc'h, K.; Andrieu, V.; Lasocki, S.; Puy, H.; Trochu, J.N. Iron deficiency screening is a key issue in chronic inflammatory diseases: A call to action. *J. Intern. Med.* **2022**. online ahead of print. [[CrossRef](#)] [[PubMed](#)]
- Tripathy, R.; Panda, A.K.; Das, B.K. Serum ferritin level correlates with SLEDAI scores and renal involvement in SLE. *LUPUS* **2015**, *24*, 82–89. [[CrossRef](#)] [[PubMed](#)]
- Kambara, T.; Amatya, V.J.; Kushitani, K.; Fujii, Y.; Endo, I.; Takeshima, Y. Downregulation of FTL decreases proliferation of malignant mesothelioma cells by inducing G1 cell cycle arrest. *Oncol. Lett.* **2022**, *23*, 174. [[CrossRef](#)]
- Schonberg, D.L.; Miller, T.E.; Wu, Q.; Flavahan, W.A.; Das, N.K.; Hale, J.S.; Hubert, C.G.; Mack, S.C.; Jarrar, A.M.; Karl, R.T.; et al. Preferential Iron Trafficking Characterizes Glioblastoma Stem-like Cells. *Cancer Cell* **2015**, *28*, 441–455. [[CrossRef](#)]
- Ferreira, C.; Buccini, D.; Martin, M.E.; Levi, S.; Arosio, P.; Grandchamp, B.; Beaumont, C. Early embryonic lethality of H ferritin gene deletion in mice. *J. Biol. Chem.* **2000**, *275*, 3021–3024. [[CrossRef](#)]
- Vanoaica, L.; Darshan, D.; Richman, L.; Schumann, K.; Kuhn, L.C. Intestinal ferritin H is required for an accurate control of iron absorption. *Cell Metab.* **2010**, *12*, 273–282. [[CrossRef](#)]
- Ikeda, Y.; Watanabe, H.; Shiuchi, T.; Hamano, H.; Horinouchi, Y.; Imanishi, M.; Goda, M.; Zamami, Y.; Takechi, K.; Izawa-Ishizawa, Y.; et al. Deletion of H-ferritin in macrophages alleviates obesity and diabetes induced by high-fat diet in mice. *Diabetologia* **2020**, *63*, 1588–1602. [[CrossRef](#)]
- Kato, J.; Fujikawa, K.; Kanda, M.; Fukuda, N.; Sasaki, K.; Takayama, T.; Kobune, M.; Takada, K.; Takimoto, R.; Hamada, H.; et al. A mutation, in the iron-responsive element of H ferritin mRNA, causing autosomal dominant iron overload. *Am. J. Hum. Genet.* **2001**, *69*, 191–197. [[CrossRef](#)]
- Cadenas, B.; Fita-Torro, J.; Bermudez-Cortes, M.; Hernandez-Rodriguez, I.; Fuster, J.L.; Llinares, M.E.; Galera, A.M.; Romero, J.L.; Perez-Montero, S.; Tornador, C.; et al. L-Ferritin: One Gene, Five Diseases; from Hereditary Hyperferritinemia to Hypoferritinemia—Report of New Cases. *Pharmaceutics* **2019**, *12*, 17. [[CrossRef](#)]
- Zhang, D.L.; Ghosh, M.C.; Rouault, T.A. The physiological functions of iron regulatory proteins in iron homeostasis—An update. *Front. Pharmacol.* **2014**, *5*, 124. [[CrossRef](#)] [[PubMed](#)]
- Koskenkorva-Frank, T.S.; Weiss, G.; Koppenol, W.H.; Burckhardt, S. The complex interplay of iron metabolism, reactive oxygen species, and reactive nitrogen species: Insights into the potential of various iron therapies to induce oxidative and nitrosative stress. *Free Radic. Biol. Med.* **2013**, *65*, 1174–1194. [[CrossRef](#)] [[PubMed](#)]
- Ito, H.; Kurokawa, H.; Matsui, H. Mitochondrial reactive oxygen species and heme, non-heme iron metabolism. *Arch Biochem Biophys* **2021**, *700*, 108695. [[CrossRef](#)]
- Zhou, Z.D.; Tan, E.K. Iron regulatory protein (IRP)-iron responsive element (IRE) signaling pathway in human neurodegenerative diseases. *Mol. Neurodegener.* **2017**, *12*, 75. [[CrossRef](#)] [[PubMed](#)]
- Kernan, K.F.; Carcillo, J.A. Hyperferritinemia and inflammation. *Int. Immunopharmacol.* **2017**, *29*, 401–409. [[CrossRef](#)]
- Huang, B.; Zhang, X.; Liu, Q.; Bai, C.; Li, C.; Wang, C.; Xin, L. Iron Regulatory Protein 1 Inhibits Ferritin Translation Responding to OsHV-1 Infection in Ark Clams, *Scapharca Broughtonii*. *Cells* **2022**, *11*, 982. [[CrossRef](#)]
- Miyazawa, M.; Bogdan, A.R.; Hashimoto, K.; Tsuji, Y. Regulation of transferrin receptor-1 mRNA by the interplay between IRE-binding proteins and miR-7/miR-141 in the 3'-IRE stem-loops. *RNA* **2018**, *24*, 468–479. [[CrossRef](#)]

28. Shi, H.; Bencze, K.Z.; Stemmler, T.L.; Philpott, C.C. A cytosolic iron chaperone that delivers iron to ferritin. *Science* **2008**, *320*, 1207–1210. [[CrossRef](#)]
29. Ryu, M.S.; Duck, K.A.; Philpott, C.C. Ferritin iron regulators, PCBP1 and NCOA4, respond to cellular iron status in developing red cells. *Blood Cells Mol. Dis.* **2018**, *69*, 75–81. [[CrossRef](#)]
30. Leidgens, S.; Bulloch, K.Z.; Shi, H.; Li, F.; Shakoury-Elizeh, M.; Yabe, T.; Subramanian, P.; Hsu, E.; Natarajan, N.; Nandal, A.; et al. Each member of the poly-r(C)-binding protein 1 (PCBP) family exhibits iron chaperone activity toward ferritin. *J. Biol. Chem.* **2013**, *288*, 17791–17802. [[CrossRef](#)]
31. Frey, A.G.; Nandal, A.; Park, J.H.; Smith, P.M.; Yabe, T.; Ryu, M.S.; Ghosh, M.C.; Lee, J.; Rouault, T.A.; Park, M.H.; et al. Iron chaperones PCBP1 and PCBP2 mediate the metallation of the dinuclear iron enzyme deoxyhypusine hydroxylase. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, 8031–8036. [[CrossRef](#)] [[PubMed](#)]
32. Protchenko, O.; Baratz, E.; Jadhav, S.; Li, F.; Shakoury-Elizeh, M.; Gavrilova, O.; Ghosh, M.C.; Cox, J.E.; Maschek, J.A.; Tyurin, V.A.; et al. Iron Chaperone Poly rC Binding Protein 1 Protects Mouse Liver From Lipid Peroxidation and Steatosis. *Hepatology* **2021**, *73*, 1176–1193. [[CrossRef](#)] [[PubMed](#)]
33. Jadhav, S.; Protchenko, O.; Li, F.; Baratz, E.; Shakoury-Elizeh, M.; Maschek, A.; Cox, J.; Philpott, C.C. Mitochondrial dysfunction in mouse livers depleted of iron chaperone PCBP1. *Free Radic. Biol. Med.* **2021**, *175*, 18–27. [[CrossRef](#)]
34. Yanatori, I.; Richardson, D.R.; Imada, K.; Kishi, F. Iron Export through the Transporter Ferroportin 1 Is Modulated by the Iron Chaperone PCBP2. *J. Biol. Chem.* **2016**, *291*, 17303–17318. [[CrossRef](#)] [[PubMed](#)]
35. Ghanem, L.R.; Kromer, A.; Silverman, I.M.; Chatterji, P.; Traxler, E.; Penzo-Mendez, A.; Weiss, M.J.; Stanger, B.Z.; Liebhaber, S.A. The Poly(C) Binding Protein Pcbp2 and Its Retrotransposed Derivative Pcbp1 Are Independently Essential to Mouse Development. *Mol. Cell Biol.* **2016**, *36*, 304–319. [[CrossRef](#)]
36. Ji, X.; Jha, A.; Humenik, J.; Ghanem, L.R.; Kromer, A.; Duncan-Lewis, C.; Traxler, E.; Weiss, M.J.; Barash, Y.; Liebhaber, S.A. RNA-Binding Proteins PCBP1 and PCBP2 Are Critical Determinants of Murine Erythropoiesis. *Mol. Cell Biol.* **2021**, *41*, e0066820. [[CrossRef](#)] [[PubMed](#)]
37. Cao, X.; Li, P.; Song, X.; Shi, L.; Qin, L.; Chen, D.; Chu, T.; Cheng, Y. PCBP1 is associated with rheumatoid arthritis by affecting RNA products of genes involved in immune response in Th1 cells. *Sci. Rep.* **2022**, *12*, 8398. [[CrossRef](#)] [[PubMed](#)]
38. Yoshimura, M.; Honda, H.; Sasagasko, N.; Mori, S.; Hamasaki, H.; Suzuki, S.O.; Ishii, T.; Ninomiya, T.; Kira, J.I.; Iwaki, T. PCBP2 Is Downregulated in Degenerating Neurons and Rarely Observed in TDP-43-Positive Inclusions in Sporadic Amyotrophic Lateral Sclerosis. *J. Neuropathol. Exp. Neurol.* **2021**, *80*, 220–228. [[CrossRef](#)]
39. Gillentine, M.A.; Wang, T.; Hoekzema, K.; Rosenfeld, J.; Liu, P.; Guo, H.; Kim, C.N.; De Vries, B.B.A.; Vissers, L.; Nordenskjold, M.; et al. Rare deleterious mutations of HNRNP genes result in shared neurodevelopmental disorders. *Genome Med.* **2021**, *13*, 63. [[CrossRef](#)]
40. Patel, S.J.; Protchenko, O.; Shakoury-Elizeh, M.; Baratz, E.; Jadhav, S.; Philpott, C.C. The iron chaperone and nucleic acid-binding activities of poly(rC)-binding protein 1 are separable and independently essential. *Proc. Natl. Acad. Sci. USA* **2021**, *118*. [[CrossRef](#)]
41. Radisky, D.C.; Kaplan, J. Iron in cytosolic ferritin can be recycled through lysosomal degradation in human fibroblasts. *Biochem. J.* **1998**, *336 Pt 1*, 201–205. [[CrossRef](#)]
42. Mancias, J.D.; Wang, X.; Gygi, S.P.; Harper, J.W.; Kimmelman, A.C. Quantitative proteomics identifies NCOA4 as the cargo receptor mediating ferritinophagy. *Nature* **2014**, *509*, 105–109. [[CrossRef](#)] [[PubMed](#)]
43. Santana-Codina, N.; Mancias, J.D. The Role of NCOA4-Mediated Ferritinophagy in Health and Disease. *Pharmaceuticals* **2018**, *11*, 114. [[CrossRef](#)]
44. Xie, Y.; Hou, W.; Song, X.; Yu, Y.; Huang, J.; Sun, X.; Kang, R.; Tang, D. Ferroptosis: Process and function. *Cell Death Differ.* **2016**, *23*, 369–379. [[CrossRef](#)] [[PubMed](#)]
45. Mancias, J.D.; Pontano Vaites, L.; Nissim, S.; Biancur, D.E.; Kim, A.J.; Wang, X.; Liu, Y.; Goessling, W.; Kimmelman, A.C.; Harper, J.W. Ferritinophagy via NCOA4 is required for erythropoiesis and is regulated by iron dependent HERC2-mediated proteolysis. *eLife* **2015**, *4*, e10308. [[CrossRef](#)] [[PubMed](#)]
46. Li, X.; Lozovatsky, L.; Sukumaran, A.; Gonzalez, L.; Jain, A.; Liu, D.; Ayala-Lopez, N.; Finberg, K.E. NCOA4 is regulated by HIF and mediates mobilization of murine hepatic iron stores after blood loss. *Blood* **2020**, *136*, 2691–2702. [[CrossRef](#)]
47. Das, N.K.; Jain, C.; Sankar, A.; Schwartz, A.J.; Santana-Codina, N.; Solanki, S.; Zhang, Z.; Ma, X.; Parimi, S.; Rui, L.; et al. Modulation of the HIF2alpha-NCOA4 axis in enterocytes attenuates iron loading in a mouse model of hemochromatosis. *Blood* **2022**, *139*, 2547–2552. [[CrossRef](#)]
48. Ryu, M.S.; Zhang, D.; Protchenko, O.; Shakoury-Elizeh, M.; Philpott, C.C. PCBP1 and NCOA4 regulate erythroid iron storage and heme biosynthesis. *J. Clin. Investg.* **2017**, *127*, 1786–1797. [[CrossRef](#)] [[PubMed](#)]
49. Philpott, C.C. The flux of iron through ferritin in erythrocyte development. *Curr. Opin. Hematol.* **2018**, *25*, 183–188. [[CrossRef](#)]
50. Bellelli, R.; Federico, G.; Matte, A.; Colecchia, D.; Iolascon, A.; Chiariello, M.; Santoro, M.; De Franceschi, L.; Carlomagno, F. NCOA4 Deficiency Impairs Systemic Iron Homeostasis. *Cell Rep.* **2016**, *14*, 411–421. [[CrossRef](#)]
51. Santana-Codina, N.; Gableske, S.; Quiles del Rey, M.; Malachowska, B.; Jedrychowski, M.P.; Biancur, D.E.; Schmidt, P.J.; Fleming, M.D.; Fendler, W.; Harper, J.W.; et al. NCOA4 maintains murine erythropoiesis via cell autonomous and non-autonomous mechanisms. *Haematologica* **2019**, *104*, 1342–1354. [[CrossRef](#)]
52. Finberg, K.E. Blocking (iron) traffic in the gut. *Blood* **2022**, *139*, 2424–2425. [[CrossRef](#)] [[PubMed](#)]

53. Dixon, S.J.; Lemberg, K.M.; Lamprecht, M.R.; Skouta, R.; Zaitsev, E.M.; Gleason, C.E.; Patel, D.N.; Bauer, A.J.; Cantley, A.M.; Yang, W.S.; et al. Ferroptosis: An iron-dependent form of nonapoptotic cell death. *Cell* **2012**, *149*, 1060–1072. [CrossRef] [PubMed]
54. Friedmann Angeli, J.P.; Schneider, M.; Proneth, B.; Tyurina, Y.Y.; Tyurin, V.A.; Hammond, V.J.; Herbach, N.; Aichler, M.; Walch, A.; Eggenhofer, E.; et al. Inactivation of the ferroptosis regulator Gpx4 triggers acute renal failure in mice. *Nat. Cell Biol.* **2014**, *16*, 1180–1191. [CrossRef]
55. Yang, W.S.; SriRamaratnam, R.; Welsch, M.E.; Shimada, K.; Skouta, R.; Viswanathan, V.S.; Cheah, J.H.; Clemons, P.A.; Shamji, A.F.; Clish, C.B.; et al. Regulation of ferroptotic cancer cell death by GPX4. *Cell* **2014**, *156*, 317–331. [CrossRef]
56. Wang, H.; An, P.; Xie, E.; Wu, Q.; Fang, X.; Gao, H.; Zhang, Z.; Li, Y.; Wang, X.; Zhang, J.; et al. Characterization of ferroptosis in murine models of hemochromatosis. *Hepatology* **2017**, *66*, 449–465. [CrossRef] [PubMed]
57. Hu, W.; Zhou, C.; Jing, Q.; Li, Y.; Yang, J.; Yang, C.; Wang, L.; Hu, J.; Li, H.; Wang, H.; et al. FTH promotes the proliferation and renders the HCC cells specifically resist to ferroptosis by maintaining iron homeostasis. *Cancer Cell Int.* **2021**, *21*, 709. [CrossRef] [PubMed]
58. Rui, T.; Wang, H.; Li, Q.; Cheng, Y.; Gao, Y.; Fang, X.; Ma, X.; Chen, G.; Gao, C.; Gu, Z.; et al. Deletion of ferritin H in neurons counteracts the protective effect of melatonin against traumatic brain injury-induced ferroptosis. *J. Pineal Res.* **2021**, *70*, e12704. [CrossRef] [PubMed]
59. Fang, X.; Cai, Z.; Wang, H.; Han, D.; Cheng, Q.; Zhang, P.; Gao, F.; Yu, Y.; Song, Z.; Wu, Q.; et al. Loss of Cardiac Ferritin H Facilitates Cardiomyopathy via Slc7a11-Mediated Ferroptosis. *Circ. Res.* **2020**, *127*, 486–501. [CrossRef]
60. Lu, R.; Jiang, Y.; Lai, X.; Liu, S.; Sun, L.; Zhou, Z.W. A Shortage of FTH Induces ROS and Sensitizes RAS-Proficient Neuroblastoma N2A Cells to Ferroptosis. *Int. J. Mol. Sci.* **2021**, *22*, 8898. [CrossRef]
61. Hou, W.; Xie, Y.; Song, X.; Sun, X.; Lotze, M.T.; Zeh, H.J., 3rd; Kang, R.; Tang, D. Autophagy promotes ferroptosis by degradation of ferritin. *Autophagy* **2016**, *12*, 1425–1428. [CrossRef] [PubMed]
62. Gao, M.; Monian, P.; Pan, Q.; Zhang, W.; Xiang, J.; Jiang, X. Ferroptosis is an autophagic cell death process. *Cell Res.* **2016**, *26*, 1021–1032. [CrossRef] [PubMed]
63. Gryzik, M.; Asperti, M.; Denardo, A.; Arosio, P.; Poli, M. NCOA4-mediated ferritinophagy promotes ferroptosis induced by erastin, but not by RSL3 in HeLa cells. *Biochim. Biophys. Acta. Mol. Cell Res.* **2021**, *1868*, 118913. [CrossRef]
64. Yoshida, M.; Minagawa, S.; Araya, J.; Sakamoto, T.; Hara, H.; Tsubouchi, K.; Hosaka, Y.; Ichikawa, A.; Saito, N.; Kadota, T.; et al. Involvement of cigarette smoke-induced epithelial cell ferroptosis in COPD pathogenesis. *Nat. Commun.* **2019**, *10*, 3145. [CrossRef]
65. Li, N.; Wang, W.; Zhou, H.; Wu, Q.; Duan, M.; Liu, C.; Wu, H.; Deng, W.; Shen, D.; Tang, Q. Ferritinophagy-mediated ferroptosis is involved in sepsis-induced cardiac injury. *Free Radic. Biol. Med.* **2020**, *160*, 303–318. [CrossRef] [PubMed]
66. Fuhrmann, D.C.; Mondorf, A.; Beifuss, J.; Jung, M.; Brune, B. Hypoxia inhibits ferritinophagy, increases mitochondrial ferritin, and protects from ferroptosis. *Redox Biol.* **2020**, *36*, 101670. [CrossRef] [PubMed]
67. Park, J.M.; Mau, C.Z.; Chen, Y.C.; Su, Y.H.; Chen, H.A.; Huang, S.Y.; Chang, J.S.; Chiu, C.F. A case-control study in Taiwanese cohort and meta-analysis of serum ferritin in pancreatic cancer. *Sci. Rep.* **2021**, *11*, 21242. [CrossRef]
68. Devos, D.; Moreau, C.; Kyheng, M.; Garcon, G.; Rolland, A.S.; Blasco, H.; Gele, P.; Timothee Lenglet, T.; Veyrat-Durebex, C.; Corcia, P.; et al. A ferroptosis-based panel of prognostic biomarkers for Amyotrophic Lateral Sclerosis. *Sci. Rep.* **2019**, *9*, 2918. [CrossRef]
69. Patin, F.; Corcia, P.; Madji Hounoum, B.; Veyrat-Durebex, C.; Respaud, E.; Piver, E.; Benz-de Bretagne, I.; Vourc'h, P.; Andres, C.R.; Blasco, H. Biological follow-up in amyotrophic lateral sclerosis: Decrease in creatinine levels and increase in ferritin levels predict poor prognosis. *Eur. J. Neurol.* **2015**, *22*, 1385–1390. [CrossRef]
70. Chen, H.C.; Tang, H.H.; Hsu, W.H.; Wu, S.Y.; Cheng, W.H.; Wang, B.Y.; Su, C.L. Vulnerability of Triple-Negative Breast Cancer to Saponin Formosanin C-Induced Ferroptosis. *Antioxidants* **2022**, *11*, 298. [CrossRef]
71. He, J.; Li, Z.; Xia, P.; Shi, A.; FuChen, X.; Zhang, J.; Yu, P. Ferroptosis and ferritinophagy in diabetes complications. *Mol. Metab.* **2022**, *60*, 101470. [CrossRef] [PubMed]
72. Qin, Y.; Qiao, Y.; Wang, D.; Tang, C.; Yan, G. Ferritinophagy and ferroptosis in cardiovascular disease: Mechanisms and potential applications. *Biomed. Pharmacother.* **2021**, *141*, 111872. [CrossRef] [PubMed]
73. Qu, C.; Dai, E.; Lai, T.; Cao, G.; Liu, J.; Kang, R.; Han, L.; Tang, D.; Zhou, D. Itaconic acid induces ferroptosis by activating ferritinophagy. *Biochem. Biophys. Res. Commun.* **2021**, *583*, 56–62. [CrossRef] [PubMed]
74. Philpott, C.C.; Patel, S.J.; Protchenko, O. Management versus miscues in the cytosolic labile iron pool: The varied functions of iron chaperones. *Biochim. Biophys. Acta. Mol. Cell Res.* **2020**, *1867*, 118830. [CrossRef]
75. Lee, J.; You, J.H.; Roh, J.L. Poly(rC)-binding protein 1 represses ferritinophagy-mediated ferroptosis in head and neck cancer. *Redox Biol.* **2022**, *51*, 102276. [CrossRef] [PubMed]
76. Yue, L.; Luo, Y.; Jiang, L.; Sekido, Y.; Toyokuni, S. PCBP2 knockdown promotes ferroptosis in malignant mesothelioma. *Pathol. Int.* **2022**, *72*, 242–251. [CrossRef]
77. Santana-Codina, N.; Gikandi, A.; Mancias, J.D. The Role of NCOA4-Mediated Ferritinophagy in Ferroptosis. *Adv. Exp. Med. Biol.* **2021**, *1301*, 41–57. [CrossRef]
78. Valdes, A.M.; Walter, J.; Segal, E.; Spector, T.D. Role of the gut microbiota in nutrition and health. *BMJ* **2018**, *361*, k2179. [CrossRef]

79. Milani, C.; Duranti, S.; Bottacini, F.; Casey, E.; Turroni, F.; Mahony, J.; Belzer, C.; Delgado Palacio, S.; Arboleya Montes, S.; Mancabelli, L.; et al. The First Microbial Colonizers of the Human Gut: Composition, Activities, and Health Implications of the Infant Gut Microbiota. *Microbiol. Mol. Biol. Rev.* **2017**, *81*, e00036-17. [[CrossRef](#)]
80. Turnbaugh, P.J.; Ley, R.E.; Mahowald, M.A.; Magrini, V.; Mardis, E.R.; Gordon, J.I. An obesity-associated gut microbiome with increased capacity for energy harvest. *Nature* **2006**, *444*, 1027–1031. [[CrossRef](#)]
81. Holmes, E.; Loo, R.L.; Stamler, J.; Bictash, M.; Yap, I.K.; Chan, Q.; Ebbels, T.; De Iorio, M.; Brown, I.J.; Veselkov, K.A.; et al. Human metabolic phenotype diversity and its association with diet and blood pressure. *Nature* **2008**, *453*, 396–400. [[CrossRef](#)] [[PubMed](#)]
82. Xia, Y.; Wang, J.; Fang, X.; Dou, T.; Han, L.; Yang, C. Combined analysis of metagenomic data revealed consistent changes of gut microbiome structure and function in inflammatory bowel disease. *J. Appl. Microbiol.* **2021**, *131*, 3018–3031. [[CrossRef](#)] [[PubMed](#)]
83. Li, N.; Chen, H.; Cheng, Y.; Xu, F.; Ruan, G.; Ying, S.; Tang, W.; Chen, L.; Chen, M.; Lv, L.; et al. Fecal Microbiota Transplantation Relieves Gastrointestinal and Autism Symptoms by Improving the Gut Microbiota in an Open-Label Study. *Front. Cell Infect. Microbiol.* **2021**, *11*, 759435. [[CrossRef](#)]
84. Ma, Y.; Liu, X.; Wang, J. Small molecules in the big picture of gut microbiome-host cross-talk. *EBioMedicine* **2022**, *81*, 104085. [[CrossRef](#)]
85. Sumida, K.; Han, Z.; Chiu, C.Y.; Mims, T.S.; Bajwa, A.; Demmer, R.T.; Datta, S.; Kovacs, C.P.; Pierre, J.F. Circulating Microbiota in Cardiometabolic Disease. *Front. Cell Infect. Microbiol.* **2022**, *12*, 892232. [[CrossRef](#)]
86. Parmanand, B.; Watson, M.; Boland, K.J.; Ramamurthy, N.; Wharton, V.; Morovat, A.; Lund, E.K.; Collier, J.; Le Gall, G.; Kellingray, L.; et al. Systemic iron reduction by venesection alters the gut microbiome in patients with haemochromatosis. *JHEP Rep.* **2020**, *2*, 100154. [[CrossRef](#)] [[PubMed](#)]
87. Ippolito, J.R.; Piccolo, B.D.; Robeson, M.S.; Barney, D.E., Jr.; Ali, J.; Singh, P.; Hennigar, S.R. Iron deficient diets modify the gut microbiome and reduce the severity of enteric infection in a mouse model of *S. Typhimurium*-induced enterocolitis. *J. Nutr. Biochem.* **2022**, *107*, 109065. [[CrossRef](#)] [[PubMed](#)]
88. McMillen, S.; Thomas, S.; Liang, E.; Nonnecke, E.B.; Slupsky, C.; Lonnerdal, B. Gut Microbiome Alterations following Postnatal Iron Supplementation Depend on Iron Form and Persist into Adulthood. *Nutrients* **2022**, *14*, 412. [[CrossRef](#)]
89. Popovic, A.; Bourdon, C.; Wang, P.W.; Guttman, D.S.; Soofi, S.; Bhutta, Z.A.; Bandsma, R.H.J.; Parkinson, J.; Pell, L.G. Micronutrient supplements can promote disruptive protozoan and fungal communities in the developing infant gut. *Nat. Commun.* **2021**, *12*, 6729. [[CrossRef](#)]
90. Cuisinier, T.; Calve, A.; Fragoso, G.; Oliero, M.; Hajjar, R.; Gonzalez, E.; Santos, M.M. Oral iron supplementation after antibiotic exposure induces a deleterious recovery of the gut microbiota. *BMC Microbiol.* **2021**, *21*, 259. [[CrossRef](#)]
91. Kim, S.R.; Kim, K.; Lee, S.A.; Kwon, S.O.; Lee, J.K.; Keum, N.; Park, S.M. Effect of Red, Processed, and White Meat Consumption on the Risk of Gastric Cancer: An Overall and Dose(-)Response Meta-Analysis. *Nutrients* **2019**, *11*, 826. [[CrossRef](#)]
92. Jahani-Sherafat, S.; Alebouyeh, M.; Moghim, S.; Ahmadi Amoli, H.; Ghasemian-Safaei, H. Role of gut microbiota in the pathogenesis of colorectal cancer; A review article. *Gastroenterol. Hepatol. Bed Bench.* **2018**, *11*, 101–109. [[PubMed](#)]
93. Louis, P.; Hold, G.L.; Flint, H.J. The gut microbiota, bacterial metabolites and colorectal cancer. *Nat. Rev. Microbiol.* **2014**, *12*, 661–672. [[CrossRef](#)] [[PubMed](#)]
94. Goosen, C.; Proost, S.; Tito, R.Y.; Baumgartner, J.; Barnabas, S.L.; Cotton, M.F.; Zimmermann, M.B.; Raes, J.; Blaauw, R. The effect of oral iron supplementation on the gut microbiota, gut inflammation, and iron status in iron-depleted South African school-age children with virally suppressed HIV and without HIV. *Eur. J. Nutr.* **2022**, *61*, 2067–2078. [[CrossRef](#)]
95. Mayneris-Perxachs, J.; Cardellini, M.; Hoyles, L.; Latorre, J.; Davato, F.; Moreno-Navarrete, J.M.; Arnoriaga-Rodriguez, M.; Serino, M.; Abbott, J.; Barton, R.H.; et al. Iron status influences non-alcoholic fatty liver disease in obesity through the gut microbiome. *Microbiome* **2021**, *9*, 104. [[CrossRef](#)] [[PubMed](#)]
96. Deschemin, J.C.; Noordine, M.L.; Remot, A.; Willemetz, A.; Afif, C.; Canonne-Hergaux, F.; Langella, P.; Karim, Z.; Vaulont, S.; Thomas, M.; et al. The microbiota shifts the iron sensing of intestinal cells. *FASEB J.* **2016**, *30*, 252–261. [[CrossRef](#)]
97. Das, N.K.; Schwartz, A.J.; Barthel, G.; Inohara, N.; Liu, Q.; Sankar, A.; Hill, D.R.; Ma, X.; Lamberg, O.; Schnizlein, M.K.; et al. Microbial Metabolite Signaling Is Required for Systemic Iron Homeostasis. *Cell Metab.* **2020**, *31*, 115–130.e6. [[CrossRef](#)]
98. Shah, Y.M.; Matsubara, T.; Ito, S.; Yim, S.H.; Gonzalez, F.J. Intestinal hypoxia-inducible transcription factors are essential for iron absorption following iron deficiency. *Cell Metab.* **2009**, *9*, 152–164. [[CrossRef](#)]
99. Knovich, M.A.; Storey, J.A.; Coffman, L.G.; Torti, S.V.; Torti, F.M. Ferritin for the clinician. *Blood Rev.* **2009**, *23*, 95–104. [[CrossRef](#)]
100. Kell, D.B.; Pretorius, E. Serum ferritin is an important inflammatory disease marker, as it is mainly a leakage product from damaged cells. *Metalomics* **2014**, *6*, 748–773. [[CrossRef](#)]
101. Lentz, S.; Roginski, M.A.; Montrief, T.; Ramzy, M.; Gottlieb, M.; Long, B. Initial emergency department mechanical ventilation strategies for COVID-19 hypoxic respiratory failure and ARDS. *Am. J. Emerg. Med.* **2020**, *38*, 2194–2202. [[CrossRef](#)] [[PubMed](#)]
102. Boechat, J.L.; Chora, I.; Morais, A.; Delgado, L. The immune response to SARS-CoV-2 and COVID-19 immunopathology—Current perspectives. *Pulmonology* **2021**, *27*, 423–437. [[CrossRef](#)] [[PubMed](#)]
103. Mangalmurti, N.; Hunter, C.A. Cytokine Storms: Understanding COVID-19. *Immunity* **2020**, *53*, 19–25. [[CrossRef](#)] [[PubMed](#)]
104. Giamparellos-Bourboulis, E.J.; Netea, M.G.; Rovina, N.; Akinosoglou, K.; Antoniadou, A.; Antonakos, N.; Damoraki, G.; Gkavogianni, T.; Adami, M.E.; Katsaounou, P.; et al. Complex Immune Dysregulation in COVID-19 Patients with Severe Respiratory Failure. *Cell Host Microbe* **2020**, *27*, 992–1000.e3. [[CrossRef](#)]

105. Sahu, K.K.; Borogovac, A.; Cerny, J. COVID-19 related immune hemolysis and thrombocytopenia. *J. Med. Virol.* **2021**, *93*, 1164–1170. [[CrossRef](#)]
106. Jawed, M.; Hart, E.; Saeed, M. Haemolytic anaemia: A consequence of COVID-19. *BMJ Case Rep.* **2020**, *13*, e238118. [[CrossRef](#)]
107. Mehta, P.; McAuley, D.F.; Brown, M.; Sanchez, E.; Tattersall, R.S.; Manson, J.J.; Hlh Across Speciality Collaboration, U.K. COVID-19: Consider cytokine storm syndromes and immunosuppression. *Lancet* **2020**, *395*, 1033–1034. [[CrossRef](#)]
108. Perricone, C.; Bartoloni, E.; Bursi, R.; Cafaro, G.; Guidelli, G.M.; Shoenfeld, Y.; Gerli, R. COVID-19 as part of the hyperferritinemic syndromes: The role of iron depletion therapy. *Immunol. Res.* **2020**, *68*, 213–224. [[CrossRef](#)]
109. Colafrancesco, S.; Alessandri, C.; Conti, F.; Priori, R. COVID-19 gone bad: A new character in the spectrum of the hyperferritinemic syndrome? *Autoimmun. Rev.* **2020**, *19*, 102573. [[CrossRef](#)]
110. Mueller, A.A.; Tamura, T.; Crowley, C.P.; DeGrado, J.R.; Haider, H.; Jezmir, J.L.; Keras, G.; Penn, E.H.; Massaro, A.F.; Kim, E.Y. Inflammatory Biomarker Trends Predict Respiratory Decline in COVID-19 Patients. *Cell Rep. Med.* **2020**, *1*, 100144. [[CrossRef](#)]
111. Mahat, R.K.; Panda, S.; Rathore, V.; Swain, S.; Yadav, L.; Sah, S.P. The dynamics of inflammatory markers in coronavirus disease-2019 (COVID-19) patients: A systematic review and meta-analysis. *Clin. Epidemiol. Glob. Health* **2021**, *11*, 100727. [[CrossRef](#)] [[PubMed](#)]
112. Terpos, E.; Ntanasis-Stathopoulos, I.; Elalamy, I.; Kastritis, E.; Sergentanis, T.N.; Politou, M.; Psaltopoulou, T.; Gerotziafas, G.; Dimopoulos, M.A. Hematological findings and complications of COVID-19. *Am. J. Hematol.* **2020**, *95*, 834–847. [[CrossRef](#)] [[PubMed](#)]
113. Zhou, F.; Yu, T.; Du, R.; Fan, G.; Liu, Y.; Liu, Z.; Xiang, J.; Wang, Y.; Song, B.; Gu, X.; et al. Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: A retrospective cohort study. *Lancet* **2020**, *395*, 1054–1062. [[CrossRef](#)]
114. Giamarellos-Bourboulis, E.J.; Poulakou, G.; de Nooijer, A.; Milionis, H.; Metallidis, S.; Ploumidis, M.; Grigoropoulou, P.; Rapti, A.; Segala, F.V.; Balis, E.; et al. Development and validation of SCOPE score: A clinical score to predict COVID-19 pneumonia progression to severe respiratory failure. *Cell Rep. Med.* **2022**, *3*, 100560. [[CrossRef](#)]
115. Toniati, P.; Piva, S.; Cattalini, M.; Garrafa, E.; Regola, F.; Castelli, F.; Franceschini, F.; Airo, P.; Bazzani, C.; Beindorf, E.A.; et al. Tocilizumab for the treatment of severe COVID-19 pneumonia with hyperinflammatory syndrome and acute respiratory failure: A single center study of 100 patients in Brescia, Italy. *Autoimmun. Rev.* **2020**, *19*, 102568. [[CrossRef](#)]
116. Ramiro, S.; Mostard, R.L.M.; Magro-Checa, C.; van Dongen, C.M.P.; Dormans, T.; Buijs, J.; Gronenschild, M.; de Kruif, M.D.; van Haren, E.H.J.; van Kraaij, T.; et al. Historically controlled comparison of glucocorticoids with or without tocilizumab versus supportive care only in patients with COVID-19-associated cytokine storm syndrome: Results of the CHIC study. *Ann. Rheum. Dis.* **2020**, *79*, 1143–1151. [[CrossRef](#)]