

Supplementary Material

1 Supplementary Methods

Targeted metabolomics analysis

Plasma sample (20 μL) were thoroughly mixed containing 120 μL of ice-cold methanol containing partial internal standards and then centrifuged at 4 $^{\circ}\text{C}$ (4000 g, 30 min). Supernatant (30 μL) was collected, derivative reagents (20 μL) were added. The mixture was sealed, and derivatization was carried out at 30 $^{\circ}\text{C}$ for 60 min. After derivatization, the sample was diluted with 330 μL of ice-cold 50% methanol solution, stored at -20 $^{\circ}\text{C}$ for 20 min and centrifuged at 4 $^{\circ}\text{C}$ (4000 g, 30 min). The supernatant (135 μL) was pipetted and 10 μL of internal standard was added. Serial dilutions of derivatized stock standards were added to the remaining wells. An ultra-performance liquid chromatography–tandem mass spectrometry (UPLC-MS/MS) system (ACQUITY UPLC-Xevo TQ-S, Waters Corp., Milford, MA, USA) was used to quantify all targeted metabolites. The system was equipped with an ACQUITY UPLC BEH C18 1.7 μM VanGuard pre-column (2.1 \times 5 mm) and ACQUITY UPLC BEH C18 1.7 μM analytical column (2.1 \times 100 mm), with an injection volume of 5 μL and flow rate of 0.4 mL/min. The column temperature was maintained at 40 $^{\circ}\text{C}$. The mobile phase consisted of A (water with 0.1% formic acid) and B (acetonitrile/IPA; 70:30). The gradient elution procedure was as follows: 0–1 min (5% B), 1–5 min (5–30% B), 5–9 min (30–50% B), 9–11 min (50–78% B), 11–13.5 min (78–95% B), 13.5–14 min (95–100% B), 14–16 min (100% B), 16–16.1 min (100–5% B), and 16.1–18 min (5% B). The MS was alternately operated in positive-ion mode (+1.5 kV) and negative-ion mode (-2 kV). The source temperature was set to 150 $^{\circ}\text{C}$, the desolvation temperature was set to 550 $^{\circ}\text{C}$, and the desolvation gas flow was set to 1000 L/h. Raw data generated by UPLC-MS/MS were processed using MassLynx software (v4.1; Waters, Milford, MA, USA) for peak integration, calibration, and quantification of each metabolite. The iMAP platform (v1.0; MetaboProfile, Shanghai, China) was used for the statistical analyses. The concentrations of targeted metabolites were determined based on the calibration curves and corresponding regression coefficients.

2 Supplementary Tables

Supplementary Table S1 Ingredients and chemical composition of the diets for dairy cows

Item	DM basis, %	Item	DM basis, %
complete feeds	42.74	DM	49.91
fat powder	1	CP	18.48
molasses	3.76	NDF	30.5
corn silage	26.59	ADF	17.53
alfalfa hay	15.23	Ca	0.83
oat hay	2.32	P	0.45
beet pulp	3.57	NEL, Mcal/kg of DM	1.74
baking soda	0.25		
cottonseed	4.54		

Note: Guaranteed value (%) of composition of each kilogram of premix: crude protein ≥ 24.8 , crude fat ≥ 4.66 , crude fiber ≤ 5.34 , calcium ≥ 0.92 , phosphorus ≥ 0.4 , lysine ≥ 0.35 , sodium chloride ≥ 1 . The net energy of lactation is the calculated value, and others are the measured value. DM=dry matter; CP= crude protein; NDF= neutral detergent fiber; ADF= acid detergent fiber; Ca= calcium;

P=phosphorus; NEL=net energy of lactation.

Supplementary Table S2 Dry matter intake in postpartum dairy cows

Item	Testing time	Group		
		Control	L-DBT	H-DBT
DMI (kg)	0d	7.17	7.04	7.21
	+7d	17.59	17.83	17.81
	+14d	18.03	18.09	18.13

Supplementary Table S3 Effect of DBT on the relative abundance of hindgut bacteria in dairy cows(+7d)

Item	Relative abundance		SEM	P-Value
	C7 (%)	DBT7 (%)		
<i>UCG-009</i>	1.070	0.664	0.092	0.038
<i>Psychrobacillus</i>	0	0.040	0.007	0.001
<i>Bacillus</i>	0	0.033	0.006	0.0001
<i>Solibacillus</i>	0	0.021	0.004	0.0002
<i>Arthrobacter</i>	0	0.019	0.003	0.006
<i>Aerococcus</i>	0	0.013	0.003	0.015

Supplementary Table S4 Effect of DBT on the relative abundance of hindgut bacteria in dairy cows(+14d)

Item	Relative abundance		SEM	P-Value
	C14 (%)	DBT14 (%)		
<i>Paeniclostridium</i>	6.573	3.286	0.655	0.009
<i>Romboutsia</i>	4.449	2.736	0.399	0.038
<i>Alistipes</i>	2.248	3.436	0.219	0.007
<i>Clostridium_sensu_stricto_1</i>	3.748	1.395	0.477	0.017
<i>Dorea</i>	0.091	0.246	0.026	0.001
<i>Lachnospiraceae_UCG-010</i>	0.051	0.174	0.034	0.028
<i>Oscillibacter</i>	0.031	0.142	0.023	0.004
<i>Coprococcus</i>	0.032	0.112	0.013	0.004
<i>Phascolarctobacterium</i>	0.031	0.108	0.022	0.023
<i>Prevotellaceae_Ga6A1_group</i>	0.018	0.044	0.005	0.012
<i>Moryella</i>	0.013	0.036	0.004	0.002

<i>Negativibacillus</i>	0.006	0.042	0.006	0.002
<i>Lachnospira</i>	0.028	0.002	0.008	0.045
<i>Tuzzerella</i>	0.004	0.019	0.003	0.015
<i>Aeriscardovia</i>	0.013	0.002	0.002	0.006
<i>Gordonibacter</i>	0.011	0.001	0.002	0.029
<i>Hydrogenoanaerobacterium</i>	0.0004	0.009	0.002	0.016
<i>Eubacterium_xylanophilum_group</i>	0.006	0	0.001	0.035

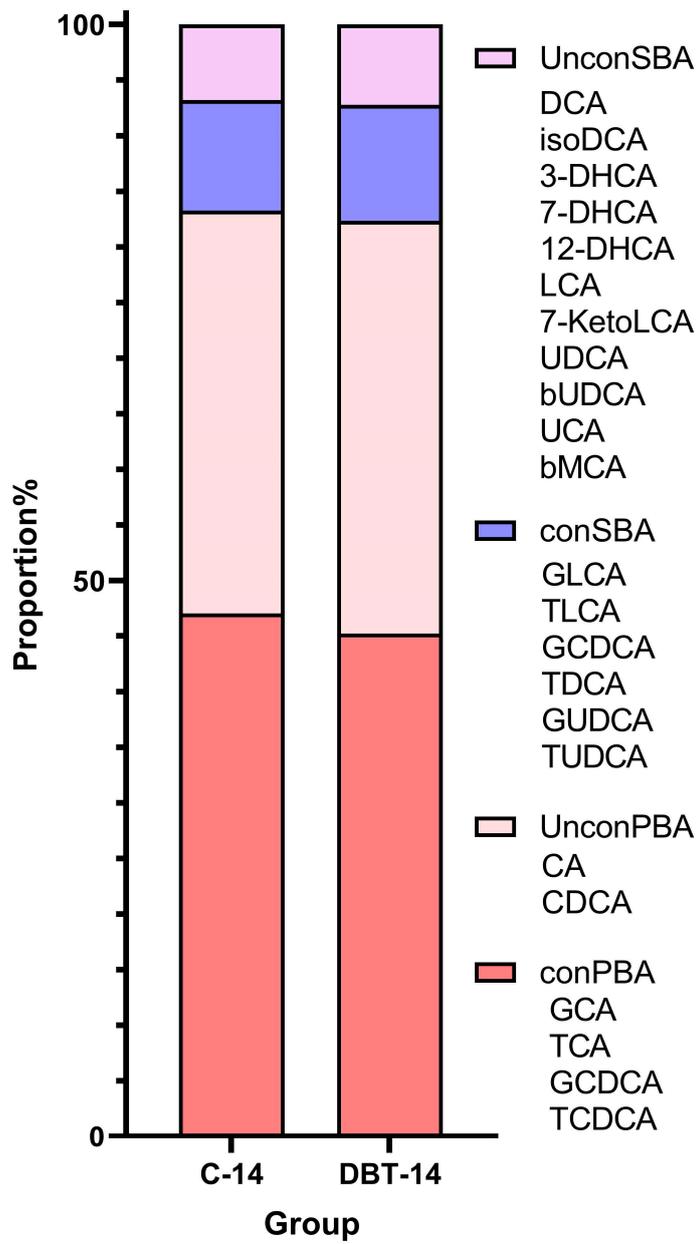
Supplementary Table S5 Differential metabolite screening of the C7 and DBT7 groups

Metabolites	C7($\mu\text{mol/L}$)	DBT7($\mu\text{mol/L}$)	Log2(FC)	P-value
Butyric acid	16.71 \pm 2.98	8.51 \pm 0.92	0.94	0.06
TCA	8.45 \pm 2.02	4.46 \pm 1.20	1.37	0.06
N-Acetylserotonin	0.48 \pm 0.05	0.62 \pm 0.04	-0.43	0.0003
2-Hydroxybutyric acid	14.64 \pm 1.37	11.04 \pm 0.99	0.41	0.05
Fumaric acid	0.62 \pm 0.10	1.30 \pm 0.21	-1.06	0.01
Malic acid	4.97 \pm 0.41	11.35 \pm 3.02	-0.89	0.001
3-Hydroxybutyric acid	582.65 \pm 47.07	449.47 \pm 22.11	0.37	0.02
Ethylmethylacetic acid	2.29 \pm 0.40	1.26 \pm 0.14	0.63	0.04
GABA	0.45 \pm 0.03	1.55 \pm 0.37	-1.10	0.002
Aspartic acid	4.93 \pm 0.33	5.77 \pm 0.22	-0.22	0.01
Ornithine	24.50 \pm 1.04	32.24 \pm 2.89	-0.39	0.02
Phenylacetylglutamine	0.59 \pm 0.03	0.55 \pm 0.04	0.18	0.06
Hydrocinnamic acid	10.62 \pm 0.92	8.25 \pm 0.53	0.37	0.04
2-Phenylpropionate	4.90 \pm 0.38	3.95 \pm 0.22	0.31	0.04
UCA	0.30 \pm 0.003	0.31 \pm 0.02	0.01	0.07
Acetylcarnitine	4.84 \pm 0.29	3.64 \pm 0.22	0.41	0.004

Note: Log2 (FC) > 0 (or < 0) indicated the metabolite concentration in C7 group were higher/lower than that in DBT7 group.

Supplementary Table S6 Differential metabolite screening of the C14 and DBT14 groups

Metabolites	C14($\mu\text{mol/L}$)	DBT14($\mu\text{mol/L}$)	Log2(FC)	P-value
GCA	25.78 \pm 3.49	17.52 \pm 2.15	0.56	0.06
GDCA	5.11 \pm 0.69	3.33 \pm 0.33	0.62	0.03



Supplementary Fig. S2 Histogram of the proportion of bile acids in plasma.