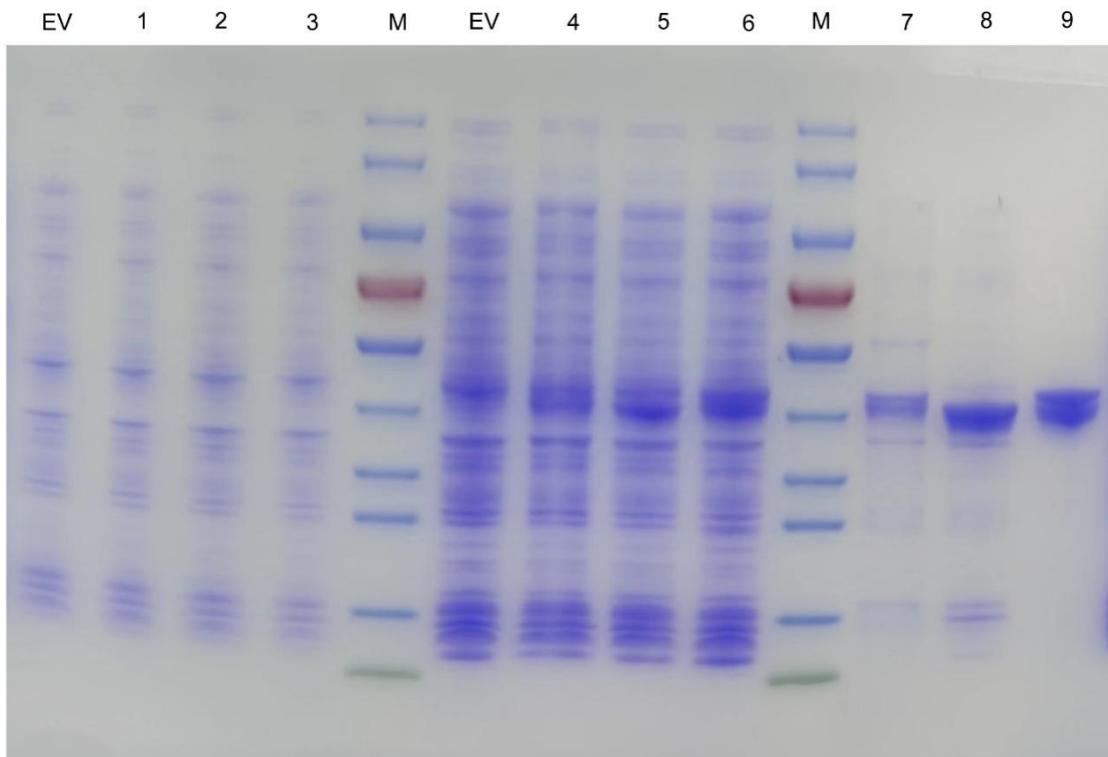
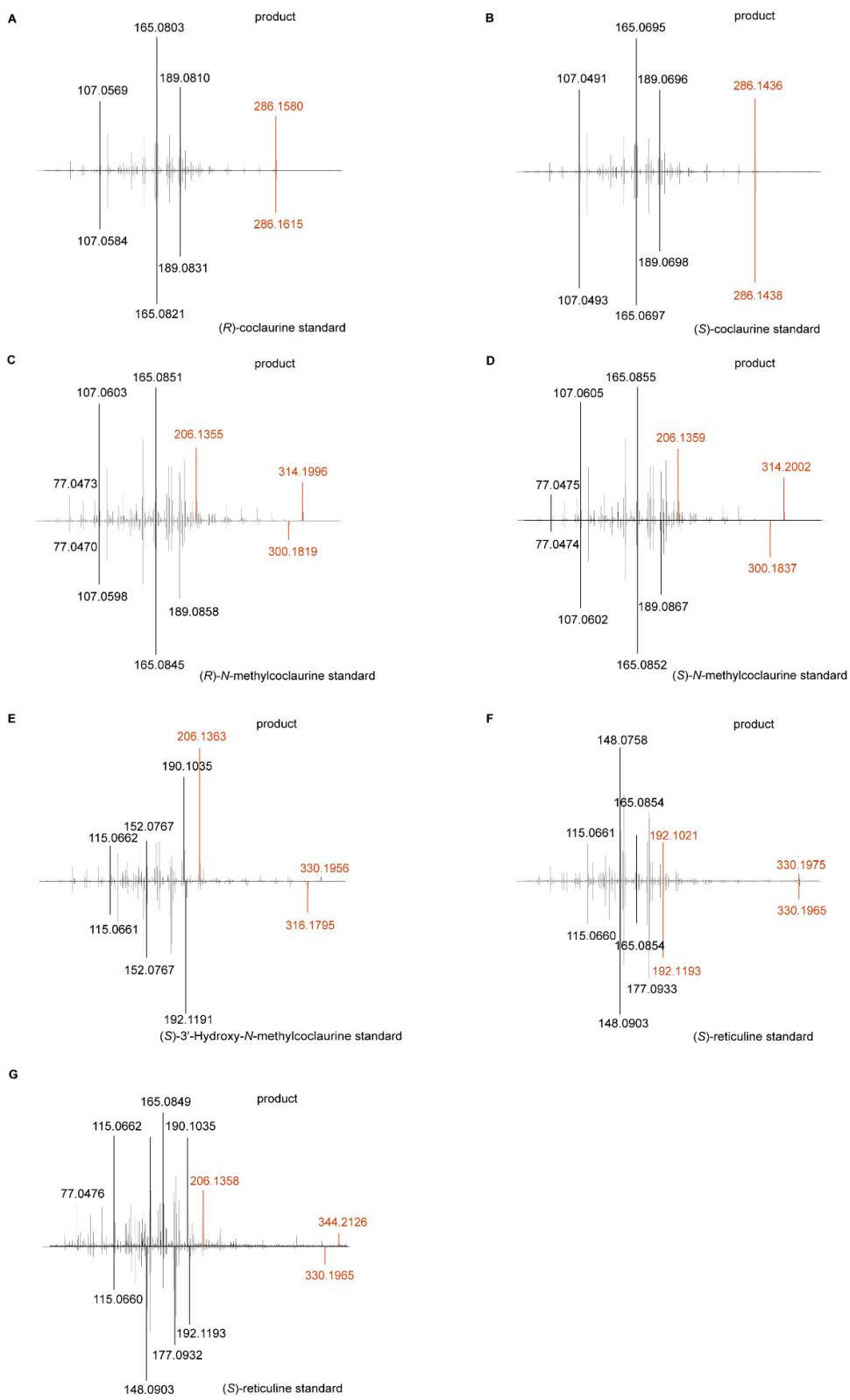


*Supplementary data*

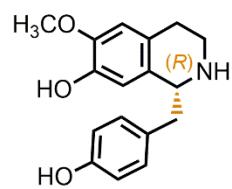
## **Identification and Characterization of Two Se6OMTs from *Stephania epigaea* Offer Novel Insights into the Biosynthetic Pathway of Cepharanthine**



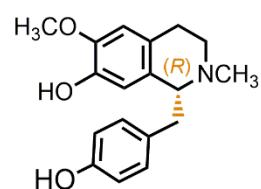
**Figure S1.** SDS-PAGE analyses was conducted on the purified proteins of St6OMT1-3 from the induced expression of the recombinant plasmid. Lane M: protein ladder, with molecular weights from top to bottom at 180, 130, 100, 70, 55, 40, 35, 25, 15, 10 kDa. Lanes 1-3: cell lysate before induction of St6OMT1-3, respectively. Lanes 2: the supernatant of induced cell lysate of St6OMT1-3. Lanes 3: the purified enzymes of St6OMT1-3. EV denotes the empty vector.



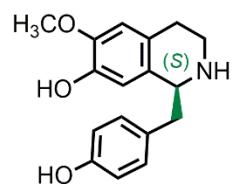
**Figure S2.** High-Resolution MS/MS spectrum fragmentation of products catalyzed by Se6OMTs. **A**, **B**, Mass spectrum of fragment ions of standard (*R*)- and (*S*)-cooclaurine (downside) and reaction products of Se6OMT1 and Se6OMT3 with substrate (*R*)- and (*S*)-cooclaurine(upside), respectively. **C**, **D**, Mass spectrum of fragment ions for standard (*R*)- and (*S*)-NMC are depicted in lower side, while the reaction products of Se6OMT3 with substrates (*R*)- and (*S*)-NMC are shown in upper side, separately. **E**, the comparison between (*S*)-HNMC standard and the products of Se6OMT3 with (*S*)-HNMC as the substrate. **F**, **G**, the comparison between (*S*)- reticuline standard and two BIAs produced by Se6OMT3 catalyzing (*S*)-HNMC and (*S*)-reticuline, respectively.



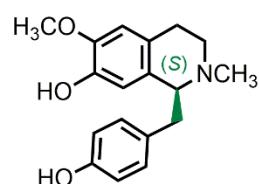
(*R*)-Coclaurine



(*R*)-*N*-methylcoclaurine



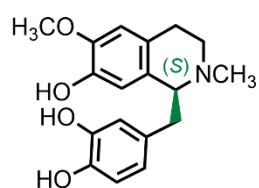
(*S*)-Coclaurine



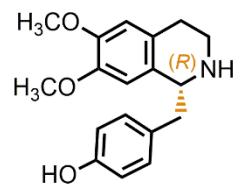
(*S*)-*N*-methylcoclaurine



(*S*)-Reticuline



(*S*)-3'-Hydroxy-*N*-methylcoclaurine



(*R*)-norarmepavine

**Figure S3.** Seven compound structures of 1-BIAs were tested as potential substrates of Se6OMTs.

**Table S1.** The full-length nucleic acid sequences of *Stephania epigaea* OMTs in this study.

gene	sequences
<i>Se6OMT1</i>	ATGGCATCAGAAAATCTAAAGCAAGACCAAGCAGCCAAAGCCAAGTTATGGAGCTTCATA- TATGGCTTGCAGAGTCACTAGTCGCAAGTCTGTATTCACTGACATACCTGATATCATTCA- AACCATGGCAAGCCCCTCCCTTCCAAACCTGCTTCTCGAATTCCCTCTCCGAAC- GGCGCCGTGGATCACGACCGCTTCCGGATCATGCGGTATTGGTGACATGAAGCTCTCA- CGAAAGAGTCTATCGACGGCGAAGATCATTGGTCTGCGCCGGCCGCGAAGTACCTAG- TGAAGGGTGGGAGAAGAGCATGGTGCCTCAATTCTATGTATAACTGATAAGGAGTTATGTC- ACCATGGTACCATCTCAAGGAGGGTTGGGAACGAGAGCAGCACAGCGTTCGA- GAAGGCCTTGGGATGAACATATGGGATTACATGTCGAAGAACCGAAGAAGAACACATCTT- CAATGAGGCCATGGCTGTGACACTAGGCTACTCATGTCATTGGTGAAGTAC- TGAATGCAAGGACATGTTCAAGGGATTAAGTCGCTAGTCATGTCGATGTCGGGGGGACTGGAAC- GCAGTTAGGCACATTGCAAGGCCATTCCACATAAATTGCACTGTC- TATGATCTCCCTCATGTTGCTGCTGATTCTCCTACTTACCCCTGAAGTGACTCGCATCGAAGGCGA- CATGTTCAAGTTCAATTCCCAAAGCAGATGCCATCTTAATGAAGT- CATCCTCATGACTGGGCCACGACAATGCAATTAAAGATTCTAAAGGAAGTGCAGGAAGCGTTG- CCGAGTAATGGGGGGAAAGTAATCCTCGAGGTCGTGTTGAATGCGCTTCCGGAG- CATCCATACACAAAGATAAGGATGGTGAATGACTGGACATGATGCTGAACGTGAGGTAAG- AAAGAACAGAGGGGGAGTGGAGAACCTCATCTATGCACTGGTTTAGCAGGTACAA- TATCAGGCAACTGGATTACACAGTATGTTATTGAGGTCTACCCCTGA
<i>Se6OMT2</i>	ATGGCAGCAGAAAATCTAGAGCAACACCAAGCAGCCAAAGCCAAGCTATGGAGC- TACATCTATGGCTATAACAGAGTCACTAGTACTCAAGTCTGCTGCTCAGCTGACATCCCTGATATC- ATCCACAACCATGGCAAGCCCATTCCCTCTCCGACCTGCTCGCCGGATTCCCTCACCA- GCGCGCCGTGGATCACACCGCTTGTATCGGATCATGCGCTACTTGGTGACATGAAGTCTTC- ACGAAGGAGTCCGACGTCATGGTGAAGATCGATATGGGCTTGCACCCCGCAAAGTAC- CTAGTGAAGGGTGGGAGAAGAGCATGGTACCTCAATTCTAAGCCTACTGATGAGGAATTCA- TGTACCATGGTACCATCTCAAGGATGGTTGTCGAACAATGGCACACCGCGTTAGA- GAAGGCCTAGGGATGGCATATGGGAATACATGTCGAAGAACCCCAAGACGAACAGCATCTT- CAATGAGGCCATGGCTGTGACACTAGGCTGGTACATGTCATTGGTGAAGTAC- TGAATGCAAGGACATGTTCAAGGAATTAAAGTCACTAGTCATGTCGATGTCGGGGGGACTGGACT- GCTGTTAGGTACATTGCAAGGCCCTCCCTACATAAATTGCACTGTC- TATGATCTCCCTCATGTTGCTGCTGATTCTCTACTTACCCGAAGTGACTCGCATCGAAGGCGA- CATGTTCAAATACATTCTAAAGCTGATGCCATCTTAATGAAGTACATT- TACATGACTGGGCCACGAGGAAGCATTCAAGATACTGAAGAAGTGTAAAGGAAGCATTGCCGA- GTGATGGGGGGAAAGTAATCCTAGTCGATATCGTGTGAATCCGTTCCGGAG- CATCCATACCGAAGGTGAGGATGGTGAACGACGTGGACATGATGCAAGAGCTGTGGAGGGAAA- GAGAGGACAGAGGAGGAGTGGAGAACAGCTCATCTATGCTGAGGTTTAG- TAGGTACAACATTAGGCACATAAATACCACATCTGTTATTGAGGTTTCCCTGA
<i>Se6OMT3</i>	ATGGCAACACAAAATCTAAAGCAAGACCAAGCAGCATCCAAAGCAAAGCTATGGAGCTTCATA- TATGGCTTGCAGAGTCACTAGTCCTCAAGTCTGCCATTCAACTGACATACCTGATATCATTCA- AACCATGGCAAGCCCATTCCACATCCAAACCTGCTTCTCGAATTCCCTCTCCGAAC- GGCGCCCGGATCACGACCGCTTCCGGATCATGCGCTATTGGTGACATGAATCTCTTCAC- GAAAGAGTCTATCGACGGCGAAGATCATTGGTCTGCGCCGGCCGCGAAGTACCTAG- TGAAGGGTGGGAGAAGAGCATGGTGCCTCAATTCTATGTATAACTGATAAGGAGTTATGTC- ACCATGGTACCATCTCAAGGAGGGTTGGGAACGAGAGCAGCACAGCGTTCGA- GAAGGCCTTGGGATGAACATATGGGATTACATGTCGAAGAACCGAAGAACACATCTT- CAATGAGGCCATGGCTGTGACACTAGGCTACTCATGTCATTGGTGAAGTAC- TGAATGCAAGGACATGTTCAAGGGATTAAGTCGCTAGTCATGTCGATGTCGGGGGGACTGGAAC- GCAGTTAGGCACATTGCAAGGCCATTCCACATAAATTGCACTGTC- TATGATCTCCCTCATGTTGCTGCTGATTCTCTACTTACCCGAAGTGACTCGCATCGAAGGCGA- CATGTTCAAGTTCAATTCCCAAAGCAGATGCCATCTTAATGAAGT- CATCCTCATGACTGGGCCACGACGAATGCAATTAGATTCTAAAGAAGTGCAGGAAGCGTTG- CCGAGTAACGGGGGGAAAGTAATCCTAGTCGATATCGTGTGAATGCGTTCCGGAG- CATCCATACACGAAGATGAGGATGGTGAATGGACTTGGACATGATGCTGAACGTGAGGGAAA- GAGAGGACAGAGGAGGAGTGGAGAACAGCTCATCTATGCTGAGGTTCACTGAGGTACAA- GATTAGGCAAGTGAATGCCATACAGTCTGTTATTGAGGTTACCCCTACTGA

**Table S2.** The primer sequences for cloning Se6OMTs-pET-28a.

gene	PET-28a primer sequences
<i>Se6OMT1</i> -F	CAGCAAATGGGT CGCGGATCCATGGCATCAGAAAATCTAAAGCAA
<i>Se6OMT1</i> -R	ACGGAGCTCGAATT CGGATCCTCAAGGGTAGACCTCAATAACAT
<i>Se6OMT2</i> -F	CAGCAAATGGGT CGCGGATCCATGGCAGCAGAAAATCTAGAG
<i>Se6OMT2</i> -R	ACGGAGCTCGAATT CGGATCCTCAAGGAAAAACCTCAATAACAGATG
<i>Se6OMT3</i> -F	CAGCAAATGGGT CGCGGATCCATGGCACACACAAAATCTAAAGCA
<i>Se6OMT3</i> -R	ACGGAGCTCGAATT CGGATCCTCAGTAAGGGTAAACCTCAATAACA

**Table S3.** The Physical and chemical properties of Se6OMTs in this study.

<b>protein</b>	<b>Se6OMT1</b>	<b>Se6OMT2</b>	<b>Se6OMT3</b>
Molecular weight	39406.59	39605.66	39645.02
Theoretical pI	6.27	6.02	6.60
Total number of negatively charged residues	43	44	43
Total number of positively charged residues	39	39	41
Formula	C <sub>1768</sub> H <sub>2767</sub> N <sub>463</sub> O <sub>510</sub> S <sub>23</sub>	C <sub>1783</sub> H <sub>2780</sub> N <sub>460</sub> O <sub>519</sub> S <sub>20</sub>	C <sub>1776</sub> H <sub>2785</sub> N <sub>465</sub> O <sub>510</sub> S <sub>26</sub>
Total number of atoms	5531	5562	5562
Instability index	41.64	40.76	42.82
Aliphatic index	89.74	88.63	86.47
Grand average of hydropathicity	-0.156	-0.183	-0.184

**Table S4.** Abbreviations and GenBank accessions numbers for functionally characterized plants OMTs used for phylogenetic analysis.

Gene	Source	Accession
Ps6OMT	<i>Papaver somniferum</i>	AAP45315
Ps4'OMT	<i>Papaver somniferum</i>	AAP45313
Ps7OMT	<i>Papaver somniferum</i>	AAQ01668
PsNMT4	<i>Papaver somniferum</i>	AOR51553
PsTNMT	<i>Papaver somniferum</i>	Q108P1
Cj6OMT	<i>Coptis japonica</i>	BAB08004
St6OMT	<i>Stephania tetrandra</i>	Reference[20]
Cj4'OMT	<i>Coptis japonica</i>	BAB08005
CjSOMT	<i>Coptis japonica</i>	BAA06192
CjCoOMT	<i>Coptis japonica</i>	BAC22084
CjCNMT	<i>Coptis japonica</i>	BAB71802
GflOMT7	<i>Glaucium flavum</i>	AKO60158
NnOMT5	<i>Nelumbo nucifera</i>	XP_010276063
SiCNMT1	<i>Stephania intermedia</i>	MK749412
SiCNMT2	<i>Stephania intermedia</i>	MK749413
SiCNMT3	<i>Stephania intermedia</i>	MK749414
MsChOMT	<i>Medicago sativa</i>	AAB48059
Cc6OMT1	<i>Coptis chinensis</i>	AXC09385