

Review **The Role and Therapeutic Potential of Pyroptosis in Colorectal Cancer: A Review**

Qing Fang 1,2, Yunhua Xu 1,3, Xiangwen Tan 1,4, Xiaofeng Wu 1,2, Shuxiang Li ⁵ , Jinyi Yuan ⁵ , Xiguang Chen ⁵ , Qiulin Huang ⁵ , Kai Fu 6,[*](https://orcid.org/0000-0001-8571-5352) and Shuai Xiao 1,5,*

- ¹ Cancer Research Institute, The First Affiliated Hospital, Hengyang Medical School, University of South China, Hengyang 421001, China; q3145692607@163.com (Q.F.); xyh940707@163.com (Y.X.); txw1143122828@hotmail.com (X.T.); 15674919272@163.com (X.W.)
- ² Department of Clinical Laboratory Medicine, The First Affiliated Hospital, Hengyang Medical School, University of South China, Hengyang 421001, China
- 3 Institute of Clinical Medicine, The First Affiliated Hospital, Hengyang Medical School, University of South China, Hengyang 421001, China
- ⁴ Department of Hepatobiliary Surgery, The Second Affiliated Hospital, Hengyang Medical School, University of South China, Hengyang 421001, China
- ⁵ Department of Gastrointestinal Surgery, The First Affiliated Hospital, Hengyang Medical School, University of South China, Hengyang 421001, China; t17872114789@163.com (S.L.); 18075933997@163.com (J.Y.); cxgxhhncn@126.com (X.C.); 2018011993@usc.edu.cn (Q.H.)
- 6 Institute of Molecular Precision Medicine and Hunan Key Laboratory of Molecular Precision Medicine, Xiangya Hospital, Central South University, Changsha 410008, China
- ***** Correspondence: fu_kai@csu.edu.cn (K.F.); xiaoshuai1982@hotmail.com (S.X.)

Abstract: Colorectal cancer (CRC) is one of the leading causes of cancer-related mortality worldwide. The unlimited proliferation of tumor cells is one of the key features resulting in the malignant development and progression of CRC. Consequently, understanding the potential proliferation and growth molecular mechanisms and developing effective therapeutic strategies have become key in CRC treatment. Pyroptosis is an emerging type of regulated cell death (RCD) that has a significant role in cells proliferation and growth. For the last few years, numerous studies have indicated a close correlation between pyroptosis and the occurrence, progression, and treatment of many malignancies, including CRC. The development of effective therapeutic strategies to inhibit tumor growth and proliferation has become a key area in CRC treatment. Thus, this review mainly summarized the different pyroptosis pathways and mechanisms, the anti-tumor (tumor suppressor) and protective roles of pyroptosis in CRC, and the clinical and prognostic value of pyroptosis in CRC, which may contribute to exploring new therapeutic strategies for CRC.

Keywords: pyroptosis; colorectal cancer; proliferation; growth; therapy

1. Introduction

Colorectal cancer (CRC) is one of the most frequently occurring and the second leading cause of tumor mortality worldwide [\[1](#page-12-0)[,2\]](#page-12-1). CRC is considered as a multifactorial disorder [\[3\]](#page-12-2) whose development and progression are associated with age, ethnicity, lifestyle, abnormal activation of proto-oncogenes, and inactivation of tumor suppressor genes, etc. [\[4](#page-12-3)[,5\]](#page-12-4). The malignant proliferation of tumor cells is a fundamental feature of cancer occurrence, progression, and metastasis, including CRC [\[6,](#page-12-5)[7\]](#page-12-6). The proliferation and growth of tumors are closely associated with various cellular and biological processes, particularly the aberration of regulated cell death (RCD). As a result, a thorough knowledge of these molecular mechanisms and the inhibition of the proliferation of tumor cells have become a primary focus in cancer therapy, including CRC treatment.

Pyroptosis is a programmed cell death (PCD) executed by the gasdermin (GSDM) protein family [\[8\]](#page-12-7), which was first discovered in macrophages infected with *Salmonella* [\[9\]](#page-12-8)

Citation: Fang, Q.; Xu, Y.; Tan, X.; Wu, X.; Li, S.; Yuan, J.; Chen, X.; Huang, Q.; Fu, K.; Xiao, S. The Role and Therapeutic Potential of Pyroptosis in Colorectal Cancer: A Review. *Biomolecules* **2024**, *14*, 874. [https://](https://doi.org/10.3390/biom14070874) doi.org/10.3390/biom14070874

Academic Editors: Marco Corazzari and Mohsin Saleet Jafri

Received: 9 May 2024 Revised: 16 July 2024 Accepted: 18 July 2024 Published: 20 July 2024

Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license [\(https://](https://creativecommons.org/licenses/by/4.0/) [creativecommons.org/licenses/by/](https://creativecommons.org/licenses/by/4.0/) $4.0/$).

or *Shigella flexneri* [\[10\]](#page-12-9) in the 1990s. This form of cell death was initially misclassified as cell apoptosis, considering that both types of cell death are characterized by caspasedependent cell death, DNA damage induction, and nuclear condensation [\[11,](#page-12-10)[12\]](#page-12-11). In 2001, the death pattern caused by *Salmonella* infection with macrophages was formally defined as "pyroptosis" [\[13](#page-12-12)[,14\]](#page-12-13). Several studies had found that GSDM is the pyroptosis executive protein that could be cleaved by caspase proteins. The N-terminal domain of GSDM (GSDM-N) perforates (punches holes in) the cell membrane after binding to the cell membrane lipid, eventually leading to the rupture of the cell membrane. After the rupture, the contents leak out, leading to cell death and inflammation [\[15\]](#page-12-14). In recent years, pyroptosis has been reported to be associated with various diseases, including inflammatory diseases [\[16\]](#page-12-15), cardiovascular disease [\[17\]](#page-12-16), leukemia [\[18\]](#page-12-17), and especially cancer [\[19\]](#page-12-18).

The pathogenesis of cancer is a complicated biological process which involves several cellular processes including inflammation and RCD like pyroptosis, among many others. Inflammation, a physiological process of the body in response to harmful stimuli, is proven to be a critical factor in cancer development and progression. Pyroptosis is a specific form of cell death, which is also present in the context of inflammation. Numerous studies have shown that pyroptosis and relevant molecules are closely associated with the occurrence, development, and prognosis of melanoma, breast, gastric, liver, and lung cancer, as well as the CRC [\[20](#page-12-19)[–26\]](#page-12-20). Pyroptosis plays a significant tumor suppressor function in melanoma and an important role in treatment [\[19,](#page-12-18)[27\]](#page-12-21). However, cell pyroptosis in CRC appears to be a double-edged sword [\[28–](#page-13-0)[30\]](#page-13-1). While on the one hand, the excessive release of inflammatory mediators, such as interleukin-1 β (IL-1 β) and IL-18, trigger abnormal systemic inflammatory reactions, accelerating tumor progression and increasing tumor burden, on the other hand, it promotes immunogenic cell death, enhances immune activity, and selectively kills tumor cells, exhibiting potential anti-tumor activity [\[31](#page-13-2)[,32\]](#page-13-3). This review comprehensively analyzed the interplay between pyroptosis and CRC development, progression and clinical significance, which may contribute to exploring new therapeutic strategies for CRC.

2. Molecular Mechanisms of Pyroptosis and CRC

Pyroptosis is an intricate molecular biological process. Four pyroptosis-related molecular pathways have been identified, namely, the inflammasome-dependent canonical or non-canonical pathway, the caspase-3 mediated non-canonical pathway, and the granzymesbased pathway, all regulated by GSDMs (Figure [1\)](#page-2-0).

2.1. The Caspase-1-Mediated Canonical Inflammasome Pathway

The first identified pathway of pyroptosis is the canonical inflammasome pathway. The canonical inflammasome pathway is caspase-1 dependent, and the inflammasomes including NLRP1 (nucleotide-binding domain leucine-rich repeat pyrin domain containing 1), NLRP3 (nucleotide-binding domain leucine-rich repeat pyrin domain containing 3), NAIP (NOD-like receptor family apoptosis inhibitory protein)/NLRC4 (NLR-family CARDcontaining protein 4), and AIM2 (Absent in melanoma 2) are the key components of this pyroptosis pathway [\[33–](#page-13-4)[35\]](#page-13-5). In this pathway, inflammasome activation is observed. After the stimulation by bacteria, virus, or intracellular danger signal, distinct types of pattern recognition receptors (PRRs) act as intracellular molecular sensors to bind and activate caspase-1 via apoptosis-associated speck-like protein containing a CARD (ASC), which forms the inflammasome [\[36\]](#page-13-6). After the inflammasome is successfully assembled, the precursor caspase-1 is cleaved into active caspase-1. Activated caspase-1 then cleaves GSDMD to form an active N-terminal domain of GSDMD (GSDMD-N) that induces cell membrane perforation, resulting in cell rupture, death, content release, and inflammation [\[37\]](#page-13-7). Meanwhile, caspase-1 promotes the maturation and secretion of IL-1 β and IL-18 outside the cells, recruits more inflammatory cells, enhances the inflammatory response, and increases osmotic pressure and cytolysis [\[38\]](#page-13-8).

Figure 1. The molecular mechanism of pyroptosis. In the caspase-1 mediated canonical inflammasome pathway, after recognition by PRRs, PAMPs form inflammasome complexes. The inflammasome massive recall this decreases with ASC, activating case of a which induces the formation of performation ortine recruits and binds with ASC, activating caspase-1, which induces the formation of perforation-active

recruits and binds with ASC, activating caspase-1, which induces the formation of perforation-active GSDMD-N. GSDMD-N is released and forms pores in the plasma membrane, leading to secretion of IL-18/1β, influx of water, cell swelling, and rupture, ultimately resulting in pyroptosis. On the other hand, the non-canonical inflammasome pathway of pyroptosis involves recognizing LPS and activating caspase-4/5/11, which cleaves GSDMD to translocate to the cell membrane, triggering pyroptosis. Another novel non-canonical of pyroptosis mediated by caspase-3 is regulated through GSDME. Chemotherapeutic drugs activate caspase-3 to cleave GSDME, leading to widespread pyroptosis. Additionally, in the granzyme-dependent pathway of pyroptosis, GZMB and GZMA can respectively act on GSDME and GSDMB to induce pyroptosis. PAMPs: pathogen-associated molecular patterns; DAMPs: damage-associated molecular patterns; NLRP1: nucleotide-binding leucine-rich repeat particles repeat to the particle receptor family apoptosis in the certain particle receptor family apoptosis in the certain of the seconddomain leucine-rich repeat pyrin domain containing 1; NLRP3: nucleotide-binding domain leucinerich repeat pyrin domain containing 3; NAIP: NOD-like receptor family apoptosis inhibitory protein; associated speck like protein containing a CARD; LPS: Lipopolysaccharide. NLRC4: NLR-family CARD-containing protein 4; AIM2: Absent in melanoma 2; ASC: Apoptosis-

has been confirmed in many studies [39,40]. Secoisolariciresinol (SECO) diglucoside The advantageous role of the canonical inflammasome pathway of pyroptosis in CRC has been confirmed in many studies [\[39](#page-13-9)[,40\]](#page-13-10). Secoisolariciresinol (SECO) diglucoside (SDG)
has been confirmed in many studies [39,40]. Secoisolariciresinol (SECO) diglucoside (SDG) enhances cleavage of the N-terminal domain of GSDMD (GSDMD-N) and caspase-1 in EDITE IS CRC cells [\[42\]](#page-13-12). A study found that in a cell model with FOXP2 depletion, low expression of FOXP2 promoted the cell growth of CRC and inhibited cell pyroptosis by inhibiting of FOXP2 from Chinese herbal medicine inhibited CRC cell proliferation and activated caspase-1 to caspase-1 expression [\[43\]](#page-13-13). Another study found that ginsenoside Rh3 (GRh3) isolated is a component of lignans with biological and anti-tumor activity [\[41\]](#page-13-11). In CRC, SDG from Chinese herbal medicine inhibited CRC cell proliferation and activated caspase-1 to induce GSDMD-dependent cell pyroptosis [\[44\]](#page-13-14). Also, subunit CLNA1 of *Lactobacillus plantarum* ZS2058 activates caspase-1, inducing pyroptosis in CRC cells, while subunit

CLNA2 inhibits CRC cell proliferation by activating caspase-4/-5 [\[45\]](#page-13-15). Above all, the canonical inflammasome pathway in CRC is characterized by the formation of inflammasomes and activation of caspases, as well as subsequent cleavage of GSDMD, resulting in cell pyroptosis and inflammatory responses which finally lead to tumor suppression [\[8\]](#page-12-7).

2.2. The Caspase-4/5/11 Mediated Non-Canonical Inflammasome Pathway

The non-canonical inflammasome pathway is relatively rare compared to the canonical one and occurs independent of caspase-1 activation [\[46\]](#page-13-16). The activation of the non-canonical inflammasome pathway is mediated by human caspase-4/5 and mice caspase-11. Studies have found that caspase-4/5 in humans or caspase-11 in mice directly binds to LPS and is activated under stress. Activated caspase-4/5/11 cleave GSDMD to form the activated N-terminal domain of GSDMD (GSDMD-N) with a perforation function, which causes cell membrane perforation and secretion of IL-1β and IL-18, resulting in pyroptosis [\[47](#page-13-17)[,48\]](#page-13-18). According to reports, caspase-11 has been found to exert preventive and protective effects against dextran sodium sulfate (DSS)-induced colitis in mice [\[49\]](#page-13-19). Mice lacking caspase-11 exhibit prominent inflammatory responses and decreased proliferation of intestinal epithelial cells [\[50](#page-13-20)[,51\]](#page-13-21). Therefore, in the non-canonical inflammasome pathway, the inhibitory effect of pyroptosis on CRC cells is also evident.

2.3. The Caspase-3 Mediated Non-Canonical Pathway

In addition to the inflammatory caspase- $1/4/5/11$, other apoptosis-associated caspases can also trigger pyroptosis. For example, under specific circumstances, such as chemotherapy or targeted therapy, pyroptosis can be induced by caspase-3 mediated gasdermin E (GSDME) [\[52\]](#page-13-22). This mode of pyroptosis is mediated by GSDME instead of GSDMD. Lobaplatin, a common chemotherapy drug, was reported to cause CRC cells to undergo pyroptosis by triggering caspase-3 to cleave GSDME [\[53\]](#page-13-23). Also, lobaplatin can activate caspase-3 in CRC cell lines HT-29 and HCT-116, leading to the cleavage of GSDME and subsequent occurrence of pyroptosis-like features, such as plasma membrane swelling and pore formation in CRC cells and inhibiting tumor growth [\[53\]](#page-13-23). The chemotherapeutic drug doxorubicin (DOX) can also induce the caspase-3 mediated pyroptosis through the ROS/JNK signaling pathway [\[54\]](#page-13-24).

In addition to chemotherapeutic drugs, natural compounds can induce pyroptosis in many cancers. Neobractatin (NBT), isolated from the edible fruit of *Garcinia bracteata*, can induce GSDME cleavage by activating caspase-3 in esophageal cancer (EC) cells, resulting in cell death and tumor growth inhibition [\[55\]](#page-14-0). Apoptin possesses the ability to cause cell death in certain human cancer cell lines [\[56\]](#page-14-1). Apoptin from the *VP3* gene of the chicken anemia virus can also trigger GSDME-mediated pyroptosis by cleaving caspase-3 [\[56\]](#page-14-1). In addition, it can increase internal reactive oxygen species (ROS) while inhibiting HCT-116 cell viability and induce pyroptosis in nude mice bearing HCT-116 xenograft, inhibiting tumor growth [\[57\]](#page-14-2).

2.4. Granzyme-A/B-Dependent Pyroptosis Pathway

According to recent research, granzymes can also target tumor cells via perforin and trigger pyroptosis [\[58,](#page-14-3)[59\]](#page-14-4). Granzyme is a serine protease secreted by cytotoxic T lymphocytes (CTL) and natural killer (NK) cells [\[58,](#page-14-3)[60\]](#page-14-5). Five kinds of granzymes have been identified in humans: GZMA, GZMB, GZMH, GZMK, and GZMM [\[58\]](#page-14-3). A previous study found that GZMA cleaves GSDMB, resulting in pyroptosis of SW837 and SKCO1 cells [\[59\]](#page-14-4). In CT26 mouse cells, the cleavage of GSDMB by GZMA significantly promotes tumor clearance [\[58\]](#page-14-3). Another serine protease, GZMB, can directly cleave GSDME and trigger caspase-independent pyroptosis in HEK293T cells. Increased expression of GSDME enhances the phagocytic activity against tumor cells, thereby inhibiting tumor growth [\[59\]](#page-14-4).

3. GSDM Family and CRC

3. GSDM Family and CRC

The GSDM protein family is characterized by pore formation, mainly expressed in the digestive tract, skin, and immune cells [\[61\]](#page-14-6). According to the conserved C-terminal and Nterminal domain sequences, the GSDM family is divided into GSDMA, GSDMB, GSDMC, GSDMD, and GSDME, except pejvakin (DFNB59/PJVK) [\[62\]](#page-14-7). GSDMs can regulate normal cell proliferation and differentiation, protect the host from pathogens [\[25\]](#page-12-22), and act as an effector protein for pyroptosis to trigger inflammation and cell death [63[,64\]](#page-14-9). GSDMs have been associated with various human diseases, including cancer and inflammationassociated diseases. Previous studies found that the GSDM family proteins are expressed in healthy normal tissues but also highly expressed in cancer tissues [65]. Recently, the GSDM family's role in cancer has become more prominent. The abnormal expression and dysfunction of GSDM family genes were linked to multiple cancer-related pathways, suggesting that GSDM genes are extensively involved in cancer occurrence and progression, including breast cancer [\[54\]](#page-13-24), lung cancer [\[66\]](#page-14-11), and gastric cancer [\[24\]](#page-12-23), as well as in CRC [\[67\]](#page-14-12). Furthermore, the GSDM genes showed significant genomic alterations, according to pancancer studies of the GSDM family $[65]$.

According to several studies, certain medications or molecules can cause GSDM-mediated pyroptosis in CRC, suggesting that GSDM family genes were associated with the develop-
GSDM have shown poor program and to the Venn analysis, GSDMB, GSDMB, GSDMB, GSDMB, GSDMB, GSDMB, GSDMB, GSDMB, ment and progression of CRC [\[68\]](#page-14-13). Notably, CRC patients with low expression of GSDMD have shown poor prognosis [\[69\]](#page-14-14). According to the Venn analysis, GSDMB, GSDMD, and have shown poor prognosis [69]. According to the Venn analysis, GSDMB, GSDMD, and GSDME are related to the invasion and metastasis of CRC $[65]$, suggesting that GSDM protein family-mediated pyroptosis has an important role in CRC (Figure [2,](#page-4-0) Table [1\)](#page-5-0). Table 1).

Figure 2. Signaling pathways of GSDMs in the CRC. GSDM proteins can regulate cell proliferation and differentiation and also serve as effector proteins in pyroptosis, triggering inflammation and cell death. Various stimuli and upstream signals such as lobaplatin, RT, 5-aza-dC, apoptin, and GA can induce GSDME cleavage and subsequent cell death. Caspase-1/GSDMD-dependent pyroptosis is induced by activating molecules, including SDG, FOXP2, GRh3, and CLNA1. CLNA2 induces cell pyroptosis through the activation of caspase-4/5. GSDMB and GSDMC participate in pyroptosis and subsequent cell death through the GZMA and TGFBR2, respectively. 5-aza-dC: 5-aza-2 ′ -deoxycytidine; RT: radiation therapy; GA: gambogic acid; DSS: dextran sulfate sodium; GRh3: ginsenoside Rh3.

Table 1. The expression and function of GSDM family members in CRC.

GSMD Family Gene	Role in CRC	Expression Pattern	Prognosis in CRC	References
GSDMA	Uncertain	Uncertain	Uncertain	None
GSDMB	Oncogene	Up-regulated	Negative	[70, 71]
GSDMC	Oncogene	Up-regulated	Negative	[23]
GSDMD	Anti-oncogene	Down-regulated	Positive	[69, 72]
GSDME	Anti-oncogene	Down-regulated	Positive	[67, 73, 74]
DFNB59/PJVK	Uncertain	Uncertain	Uncertain	None

3.1. GSDME

GSDME, also known as deafness autosomal dominant 5 (DFNA5), can induce cell swelling and death [\[75,](#page-14-20)[76\]](#page-14-21). After cleavage by the activated caspase-3, the N-terminal domain of GSDME (GSDME-N) can promote pore formation and trigger pyroptosis. GSDME expression level is different in malignant and normal tissues [\[77\]](#page-14-22). GSDME is always highly expressed in normal tissues, but the expression level is distinct among different cancers and episodically absent expression in some cancer, and the absent expression may be due to the methylation of the GSDME gene promoter [\[67](#page-14-12)[,78–](#page-14-23)[80\]](#page-15-0). The mutations of tumor-associated GSDME also could inhibit pyroptosis [\[81\]](#page-15-1).

In addition, several studies have shown that GSDME, as a molecule with known anti-tumor potential, is involved in the development and progression of many cancers [\[82\]](#page-15-2). According to a recent study, ectopic expression of GSDME in cancer cells could enhance antitumor immune responses and suppress tumor growth [\[59\]](#page-14-4). In case of CRC also, GSDME has been shown to have anti-tumor effect [\[83\]](#page-15-3). For example, an ectopic expression of GSDME significantly inhibits colony formation, proliferation, and growth of CRC cells [\[73\]](#page-14-18). The methylation inhibitor 5-aza-2′ -deoxycytidine (5-aza-dC) could promote GSDME expression and inhibit tumor cell proliferation and carcinogenesis, implying that GSDME is a potential tumor suppressor gene in CRC [\[73](#page-14-18)[,84\]](#page-15-4). Radiation therapy (RT) and chemotherapeutic agents such as lobaplatin and cisplatin can also induce caspase-3 activation and GSMDE cleavage following by pyroptosis [\[53](#page-13-23)[,85\]](#page-15-5). Activated caspase-3 and cleavage of GSDME were also observed in natural compound gambogic acid (GA)-treated CRC cells. This GA induced GSDME-dependent pyroptosis and significantly suppressed tumor proliferation, while enhancing anti-tumor activity [\[86\]](#page-15-6). Since GZMB can cleave and activate caspase-3, Zhang et al. also found that GZMB cleaved and activated GSDME in a caspase-independent manner [\[59\]](#page-14-4). These findings indicate that GSDME induces pyroptosis in CRC, acting as a cancer suppressor gene.

3.2. GSDMD

GSDMD is the extensively studied member of the GSDM family, being generally recognized as the executor of pyroptosis [\[8,](#page-12-7)[47\]](#page-13-17). GSDMD is mainly expressed in the gastrointestinal epithelium, skin, and immune cells [\[87,](#page-15-7)[88\]](#page-15-8). GSDMD is significantly expressed in esophageal cancer, gastric cancer, CRC, and other cancers [\[87,](#page-15-7)[89\]](#page-15-9). Typically, inflammatory caspase-1, which was activated by different signals/stimuli, cleaves the GSDMD to form an activated N-terminal domain of GSDMD (GSDMD-N), leading membrane pores to induce pyroptosis, which mediates the release of inflammatory cytokines IL-1 β and IL-18 [\[90](#page-15-10)[–92\]](#page-15-11). During pyroptosis, the calcium influx through the GSDMD pores is one of the signals to activate cell membrane repair, which enhances cell survival ability [\[93\]](#page-15-12). In

addition to caspase-1, caspase-11 and caspase-4/5 can also cleave GSDMD and induce pyroptosis. Caspase-11 and caspase-4/5 can be directly activated by LPS to cleave GSDMD and induce pyroptosis [\[8,](#page-12-7)[47\]](#page-13-17). Pan-cancer analysis showed that GSDMD-mediated pyroptosis might have a critical role in cancers such as adrenocortical carcinoma and CRC and is associated with the prognosis [\[94\]](#page-15-13). In a GSDMD deficient CRC mouse model induced by azoxymethane (AOM)/dextran sulfate sodium (DSS), CRC development is accompanied by the downregulation of GSDMC [\[95\]](#page-15-14). Also, a study found that a cooperative downregulated expression of GSDMD and GSDMC had a suppressive effect on gastrointestinal tumors [\[95\]](#page-15-14). Furthermore, GSDMD expression seems to be negatively correlated with lymphatic metastases and distant metastases in CRC [\[96\]](#page-15-15). According to another study, LPS-induced pyroptosis inhibits the proliferation of CRC and increases the anti-tumor activity of oxaliplatin [\[69\]](#page-14-14). Pyroptosis induced by caspase-4/GSDMD can also promote the release of inflammatory factors such as IL-1 β and IL-18, as well as recruit CD8⁺ T cell infiltration, thereby activating anti-tumor immunity [\[97\]](#page-15-16). Collectively, GSDMD-mediated pyroptosis has an important anti-tumor role in CRC.

3.3. GSDMB

GSDMB is known as gasdermin-like protein (GSDML). Interestingly, GSDMB is unique to the human genome and particularly expressed in the airway tract and gastrointestinal epithelium [\[87](#page-15-7)[,98\]](#page-15-17). Furthermore, it is abnormally expressed in many malignancies, including breast, cervical, gastric, and colon cancers [\[22,](#page-12-25)[87,](#page-15-7)[99\]](#page-15-18). GSDMB is frequently up-regulated and has a complex role in cancer. Additionally, GSDMB expression is increased in CRC and associated with inflammatory bowel disease (IBD) susceptibility. Therefore, GSDMB may have an important role in the pathogenesis of CRC under the inflammatory response in IBD [\[100](#page-15-19)[,101\]](#page-15-20).

A GSDMB-dependent cellular function study suggested that GSDMB-expressing intestinal epithelial cells (IECs) have a protective effect during gastrointestinal inflammatory infection and cancer [\[71,](#page-14-16)[102\]](#page-15-21). Recent studies have found that GSDMB could inhibit epithelial cell proliferation and motility, suggesting that epithelial-derived GSDMB has a protective function from cancer [\[70\]](#page-14-15). In addition, GSDMB has been found to specifically bind with lipid membranes, but this property appeared to be independent of pyroptosis [\[70](#page-14-15)[,102\]](#page-15-21). Since GSDMB has no orthologs in mice, identifying its role in vivo is still challenging. At present, the evidence related to the molecular mechanism of GSDMB-induced pyroptosis is limited. Therefore, the important role of GSDMB in cancer cell pyroptosis remains elusive.

3.4. GSDMC

GSDMC was originally found to be highly expressed in metastatic melanoma cells and was first known as a melanoma-derived leucine zipper-containing extranuclear factor (MLZE) [\[84\]](#page-15-4). The expression of GSDMC is low in most normal tissues but high in the gastrointestinal tract. Also, its physiological function is still unclear [\[87\]](#page-15-7). In most cancers, the methylation of the GSDMC promoter is lower in tumors than in normal tissues. GSDMC expression is tumor-specific and often expressed in melanoma [\[84\]](#page-15-4), lung cancer [\[103\]](#page-15-22), and CRC [\[104\]](#page-15-23). In addition, GSDMC exhibits cell growth inhibition activity, which indicates it may act as a potential tumor suppressor [\[87\]](#page-15-7). Like other GSDMs, GSDMC is involved in the pyroptosis biological process. Yet, because the biological function of GSDMC is rarely studied, its role in cancer is still not fully understood.

Hou et al. reported that GSDMC could be cleaved by caspase-8 and cause breast cancer cells to undergo pyroptosis [\[105\]](#page-16-0). Another study discovered that the metabolite α-ketoglutarate (α-KG) could induce pyroptosis via caspase-8-mediated GSDMC cleavage, inhibiting tumor growth and metastasis in a mouse model [\[106\]](#page-16-1). GSDMC also inhibits cell growth and decreases tumor proliferation in gastric and esophageal cancer [\[87\]](#page-15-7). On the other hand, GSDMC has also been linked to tumorigenesis in some cases. For example, GSDMC stimulates CRC proliferation under adenomatous polyposis coli (APC) mutation, which was significantly reversed by GSDMC silencing [\[23\]](#page-12-24). However, due to various

pathways of pyroptosis, the tumor suppressor or tumor-promoting effect of pyroptosis is also related to the duration time and the intensity of the inflammatory response. Therefore, the function of GSDMC in the carcinogenesis of CRC needs to be further clarified.

3.5. Other GSDMs

3.5.1. GSDMA

GSDMA is a GSDM family member with three mouse homologs [\[62\]](#page-14-7). The molecular regulation mechanism of GSDMA is still not fully understood. GSDMA is expressed in the upper gastrointestinal tract and is strongly linked with asthma susceptibility [\[107\]](#page-16-2), but its expression is frequently silenced in gastric cancer, suggesting the potential tumor suppressor role in gastrointestinal cancer [\[87\]](#page-15-7). However, since GSDMA is not expressed in the small and large intestine, the correlation of GSDMA with CRC development or progression has not been identified.

3.5.2. DFNB59

DFNB59, also known as Pejvakin (PJVK), has received little attention as a member of the GSDM family so far. Its expression has been found in the neurons of the afferent auditory pathway, inner ear cells, and testis, and it is mainly associated with hearing impairment [\[108](#page-16-3)[,109\]](#page-16-4). DFNB59 lacks the linker region between the N-terminal and Cterminal domains and has no pore-forming activity [\[110\]](#page-16-5); thus, its role in pyroptosis is still unknown. The role of DFNB59 in CRC has not yet been reported.

4. The Role of Pyroptosis in CRC

4.1. Pyroptosis Function as a Tumor Suppressor in CRC

Emerging studies demonstrate that various cellular processes and RCD may lead to cancer cell death, and the most common are apoptosis, autophagy, ferroptosis, necroptosis, and pyroptosis. RCD abnormalities are associated with cancer development and progression [\[68](#page-14-13)[,111](#page-16-6)[,112\]](#page-16-7). Pyroptosis, an inflammatory cell death associated with the inflammasomes and GSDM, affects the transformation and development of cancer. The activation of pyroptosis can lead to selective cell death in tumor cells. Cancer cells exhibit a higher sensitivity to pyroptosis compared to normal cells [\[113,](#page-16-8)[114\]](#page-16-9). What is more, studies have proven that pyroptosis can promote immunogenic cell death, improve immune activity, and kill tumor cells, thus having a strong anti-tumor activity against cancer cell invasion [\[14,](#page-12-13)[115,](#page-16-10)[116\]](#page-16-11). A study found that FOXP2 could inhibit CRC cell proliferation by promoting caspase-1 expression in a colitis-associated CRC model [\[43\]](#page-13-13). In addition, using a bioorthogonal system, it was found that in tumor cells undergoing pyroptosis, cells and genes associated with immunity and anti-tumor, such as $CD4^+$ T, $CD8^+$ T, and NK cells, are up-regulated, while various molecules that promote tumor growth and proliferation are down-regulated [\[117\]](#page-16-12). Pyroptosis-related participants, pathways, and regulatory mechanisms have been implicated in cancer development, progression, and metastasis, and pyroptosis has a powerful ant-tumor potential during cancer progression, including CRC [\[58](#page-14-3)[,59\]](#page-14-4). Therefore, the activation of pyroptosis may provide a potential tumor suppressor and treatment strategy for CRC.

4.2. Pyroptosis and the Tumorigenesis and Development of CRC

Various factors regulate the pathophysiological process of tumorigenesis. The two primary processes underlying CRC occurrence are the classical normal mucosal–adenoma–carcinoma evolution pattern and the colitis-associated pattern of CRC [\[5\]](#page-12-4). Pyroptosis takes part in most aspects of the tumorigenesis and development of CRC [\[118](#page-16-13)[–120\]](#page-16-14). According to a classical pattern, CRC originates from cancer stem cells (CSC). Previous studies indicated that pyroptosis could decrease CSC activation to inhibit cancer cell proliferation. After evaluating the relationship between GSDM genes and stemness, it was found that the GSDM family was not only related to stemness but also to patients' survival [\[65\]](#page-14-10). The relationship between the normal mucosal–adenoma–carcinoma evolution pattern and pyroptosis remains unclear and

requires further exploration and investigation. However, the relationship between pyroptosis and colitis-associated CRC is gradually becoming clearer.

For the inflammation-related pattern, IBD is highly associated with CRC and can be divided into Crohn's disease (CD) and ulcerative colitis (UC) [\[121\]](#page-16-15). During the inflammatory process, PAMPs from pathogens and DAMPs released by host damaged cells are recognized by PRRs in intestinal epithelial cells, leading to inflammasome assembly and subsequent cell pyroptosis. Accordingly, pyroptosis promotes cell death via the inflammatory response and protects from CRC. The anti-tumor role of inflammasomes in colitis-associated CRC such as maintaining epithelial cell integrity by NLRP3 inflammasomes in IBD has been extensively studied. Several studies have identified the anti-tumor effects of pyroptosis in CRC [\[18](#page-12-17)[,122,](#page-16-16)[123\]](#page-16-17). It has been found that NLRP1^{-/-} mice had significantly increased gastrointestinal inflammation and tumorigenesis compared with wild-type mice, indicating that NLRP1 has a protective role in reducing colitis and colitis-related CRC [\[40\]](#page-13-10). Another study demonstrated that NLRP3 inflammasome components act as a protective role in an animal model of acute colitis, in which NLRP3 deficiency enhanced chemically induced colitis-associated CRC occurrence [\[122\]](#page-16-16). Meanwhile, in the acute intestinal inflammation model induced by dextran sodium sulfate (DSS), caspase-11 was found to suppress and inhibit intestinal inflammation, further supporting the protective role of pyroptosis in IBD [\[50\]](#page-13-20). In mice lacking caspase-11, the damage caused by inflammation was more pronounced in the colon [\[124\]](#page-16-18). Moreover, in the colitis-associated CRC model induced by 5-aza-dC, the activation of NALP1 inflammasome increased, suppressing the growth of colon cancer and increasing lifespan [\[114\]](#page-16-9). A study found that NLRP3 could promote hepatic NK cell maturation, and the liver metastasis was increased when IL-18 signaling was impaired in a NLRP3 knockdown CRC mouse model [\[125\]](#page-16-19). All the above findings suggest that pyroptosis plays an important role in CRC development and progression.

5. The Potential Clinical Value of Pyroptosis in CRC

According to the important role of pyroptosis in the development and progression of CRC, the significance of pyroptosis in CRC treatment has also been studied (Figure [3\)](#page-9-0). Chemotherapy is currently the most widely utilized treatment for CRC. Chemotherapeutic drugs can induce pyroptosis in tumor cells, affecting cell vitality, invasion, and migration, thereby promoting tumor cell death [\[69](#page-14-14)[,114\]](#page-16-9). Additionally, the synergy between pyroptosis and chemotherapy drugs can increase chemotherapy sensitivity. Radiotherapy-induced pyroptosis enhances the radiosensitivity of CRC and boosts tumor immune infiltration, greatly improving treatment efficacy [\[126](#page-16-20)[,127\]](#page-16-21). In CRC immunotherapy, induction of cell pyroptosis promotes immune cell activity, enhances cancer cell sensitivity to immune checkpoint inhibitors (ICIs), and improves anti-tumor immune responses [\[58,](#page-14-3)[117,](#page-16-12)[128\]](#page-16-22). Furthermore, targeted pyroptosis may become an important therapeutic approach in future CRC treatments. Recently, novel delivery molecules such as biomimetic nanoparticles (BNP) and drug–polymer hybrid supramolecular nanoprodrugs (PDNP) have been developed, effectively inducing pyroptosis in tumor cells and enhancing treatment efficacy [\[129,](#page-16-23)[130\]](#page-17-0).

5.1. Pyroptosis and Chemotherapy in CRC

Chemotherapy is the most important systemic treatment option for advanced CRC. Yet, due to chemotherapy resistance, many CRC patients develop recurrence and metastasis after therapy [\[131\]](#page-17-1). Therefore, developing treatment strategies to reverse or compete against chemotherapy resistance is crucial.

Some chemotherapeutic drugs, including cisplatin, paclitaxel, 5-fluorouracil (5-FU), lobaplatin, etc., can trigger pyroptosis in tumor cells [\[132\]](#page-17-2). Compared to conventional chemotherapy, drugs inducing pyroptosis can effectively overcome resistance to apoptosis, reduce tumor immune tolerance, and enhance treatment efficacy. In CRC with high expression of GSDME, drugs such as $TNF\alpha+CHX$ and navitoclax can induce pyroptosis through the BAK/BAX caspase-3-GSDME pathway [\[81\]](#page-15-1). However, due to hypermethylation of the GSDME gene, most cancer cells lack caspase-3-derived GSDME. Four inhibitors of cell

proliferation, including obatoclax mesylate (OM), BI 2536 (BI), (S)-(+)-camptothecin (CPT), and bortezomib (BTZ), were screened for their drug efficacy in CRC using a cancer-tissueoriginated spheroid (CTOS)-based screening method and successfully induced GSDMEmediated pyroptosis. The results showed that CRC growth was inhibited and accompanied by increased CD8⁺ T cells during pyroptosis [\[133\]](#page-17-3). More recently, Alisol A, a marine herb with anti-tumor effects, was found to induce pyroptosis by increasing the levels of caspase-1, GSDMD, and GSDME in CRC cells, while reducing cancer cell migration and increasing the chemosensitivity of cisplatin [\[134\]](#page-17-4). FL118 is a novel camptothecin-based anti-tumor drug that exhibits anti-tumor effects in CRC. Recent studies have found that FL118 inhibits the growth and metastasis of CRC by activating the NLRP3 inflammasome and promoting the release of IL-18 and IL-1 β [\[135\]](#page-17-5). Additionally, it was found that administering the synthetic FXR agonist GW4064 simultaneously with oxaliplatin could have a synergistic anti-tumor impact, as GW4064 enhanced the chemosensitivity of cells to oxaliplatin by inducing BAX/caspase-3/GSDME-mediated pyroptosis [\[136\]](#page-17-6). Similarly, a study showed that GSDMD-mediated pyroptosis promoted oxaliplatin sensitivity in HT-29 cells after LPS-induced GSDMD expression [\[69\]](#page-14-14). This combination method provided a new strategy for the treatment of CRC.

In conclusion, all this evidence suggests that the induction of pyroptosis by chemotherapeutic drugs could promote drug sensitivity and improve the efficacy of chemotherapy.

Figure 3. Prospects of pyroptosis in CRC treatment. The use of chemotherapy drugs, RT, immunotherapy, and targeted pyroptosis can effectively induce CRC cells pyroptosis, thereby improving the treatment effect.

5.1. Pyroptosis and Chemotherapy in CRC 5.2. Pyroptosis and RT in CRC

RT is one of the fundamental therapies to control local recurrence and metastasis in advanced CRC. Several research investigations have shown that the induction of cancer cell pyroptosis is also an important mechanism of RT [\[126](#page-16-20)[,137\]](#page-17-7). Studies found that RT could effectively induce dose-dependent pyroptosis in HCT-116 cells [\[127\]](#page-16-21). In CRC models, RT suppressed the expression of miR-15a while inducing caspase-1 activity and GSDMD expression, decreasing tumor cell vitality, proliferation, and growth [\[137\]](#page-17-7). Additionally, caspase-1, caspase-4, and caspase-5 were up-regulated and triggered pyroptosis as a re-sult of miR-21-5p overexpression [\[138\]](#page-17-8). Another study showed that the lncRNA NEAT1

regulates the expression of GSDME, leading to pyroptosis in CRC cells during RT [\[127\]](#page-16-21). Elevated expression of GSDME enhances the RT sensitivity of CRC and increases tumor immune infiltration [\[126,](#page-16-20)[127\]](#page-16-21). Thus, the induction of pyroptosis via RT is important for enhancing the RT efficiency in controlling tumor growth and immune microenvironment in CRC.

5.3. Pyroptosis and Immunotherapy

The fully functional immune system can prevent tumor initiation and progression, but the immune function is always impaired under malignant progression [\[139,](#page-17-9)[140\]](#page-17-10). Immunotherapy is a treatment strategy to enhance the cancer cell immunogenicity and stimulate the immune response against immune escape [\[141\]](#page-17-11). Immunotherapy based on ICIs significantly affects most cancers, especially CRC. In fact, the application of ICIs is limited because of their natural anti-apoptotic ability and the individual differences in patients. Thus, studying the molecular mechanisms of cell death other than apoptosis has become a new subject of cancer therapy [\[142\]](#page-17-12).

Infiltration of CD8⁺ T cells is crucial to immunotherapy. Activated T cells can exert synergistic effects in inducing pyroptosis and immune response [\[142\]](#page-17-12). During pyroptosis, the immune cells' activity is enhanced, leading to an increased cytotoxic effect of cytotoxic lymphocytes [\[58\]](#page-14-3). Therefore, induction of tumor cell pyroptosis can improve the sensitivity to ICIs. For instance, nanomaterials enhanced the immunogenicity of cancer cells and showed favorable treatment efficacy in immunogenic cell death (ICD)-based therapy [\[143\]](#page-17-13). In addition, ICIs and pyroptosis were also found to show synergistic antitumor activity [\[117](#page-16-12)[,128\]](#page-16-22). A prodrug utilizing high paclitaxel (PTX) and a photosensitizer purine 18 (P18)-loaded ROS/glutathione (GSH) dual-responsive nano-prodrug (denoted as MCPP) was recently designed, which could efficiently induce cancer cell-specific pyroptosis, trigger adaptive immunity, enhance the effectiveness of immune checkpoint blockade (ICB), and achieve tumor regression [\[144\]](#page-17-14). These findings suggest that pyroptosis is essential for CRC immunotherapy.

5.4. Pyroptosis and Targeted Therapy

Distinct signaling pathways and molecules of novel cell death forms other than apoptosis have been discovered recently, providing new targets and strategies for cancer therapy [\[145\]](#page-17-15). As an emerging target for cancer therapy, pyroptosis has shown great potential in treating CRC [\[127](#page-16-21)[,146\]](#page-17-16). A series of new drugs targeting pyroptosis are being developed and tested in clinical trials in CRC. For example, a biomimetic nanoparticle (BNP) could efficiently accumulate at tumor sites, induce GSDME-mediated pyroptosis, and activate systemic anti-tumor immunity, effectively reducing the severe toxicity to normal cells and tissues [\[129\]](#page-16-23). A nanoparticle-mediated cytotoxic drug can selectively be delivered to cancer cells via exotoxin A to activate NLRP3, cleave GSDMD, and mediate pyroptosis at the primary tumor site [\[72\]](#page-14-17). Moreover, nanoparticles with high selectivity for inducing pyroptosis can effectively activate caspase-11 and NLRP3, causing CRC cell proliferation suppression. This treatment's maximum effect appears 48 h post-treatment [\[147\]](#page-17-17). Moreover, Liang and colleagues designed a drug–polymer hybrid supramolecular nanoprodrug (PDNP) as a pyroptosis inducer. PDNP solves the weakness of pyroptotic efficacy during drug delivery and promotes precise drug release, thereby effectively triggering GSDME-mediated pyroptosis and enhancing the anti-tumor immune response [\[130\]](#page-17-0). These findings provide a lot of preliminary targeting pyroptosis therapy evidence for CRC.

5.5. Diagnostic and Prognostic Value of Pyroptosis in CRC

As the important role of pyroptosis in CRC has been identified, the diagnostic and prognostic value of pyroptosis in CRC has also gradually been studied in recent years. Several pyroptosis-related biomarkers have been identified to predict patient prognosis [\[148](#page-17-18)[,149\]](#page-17-19). For example, a pyroptosis-related gene (PRG) prognosis prediction model showed that pyroptosis is associated with a higher proportion of immune infiltration and indicated a

better prognosis [\[150,](#page-17-20)[151\]](#page-17-21). A study demonstrated that the expression of NLRC3 and NLRP4 is lowered in CRC and correlated with immune infiltration, which was positively correlated with favorable prognosis [\[120\]](#page-16-14). In a recent study, functional analysis revealed a negative correlation between three genes (SLC2A3, TMPRSS11E, and UPK3B) can predict the overall survival of patients with colon cancer, finding that it is negatively correlated with the proliferation and migration of colon cancer cells [\[152\]](#page-17-22). Moreover, the lncRNA risk model related to cell pyroptosis revealed remarkable predictive capacity in CRC, indicating that the high-risk group was worse than the low-risk group in terms of survival outcomes [\[153\]](#page-17-23). The role of the predictively modeled lncRNA KCNQ1OT in CRC progression has been reported [\[153\]](#page-17-23). In addition, a CRC risk model based on nine high-risk groups of lncRNA associated with pyroptosis has been validated, demonstrating high expression levels in cancer tissues [\[154\]](#page-17-24). It was found that pyroptosis levels in CRC (such as CSP1, CASP6, GZMB, and NLRP1) and the tumor microenvironment (TME) were significantly correlated with the prognosis of individual CRC patients [\[155\]](#page-17-25). The CRC prediction model based on 13 PRGs (AIM2, CASP1, CASP5, CASP6, CASP8, CASP9, ELANE, GPX4, GSDMD, NLRP7, NOD2, PJVK, and PRKACA) revealed a higher proportion of immune infiltration and better survival outcomes in the low-risk group [\[156\]](#page-17-26). In addition, IL-17A-mediated pyroptosis in CRC cells could release immune antigens and promote the infiltration of CD8⁺ T cells, thereby improving the CRC patients' prognosis [\[97\]](#page-15-16). AIM2 inflammasome, which mediates caspase-1 activation and pyroptosis, had low expression in CRC and was negatively correlated with survival [\[157](#page-18-0)[,158\]](#page-18-1). HMGB1 is an inflammatory nuclear protein released during GSDME-mediated pyroptosis, and in the absence of GSDME, the expression of HMGB1 is significantly reduced and associated with the prognosis of CRC [\[159\]](#page-18-2).

The above studies indicate that pyroptosis plays an important function in CRC, and that pyroptosis-related molecules can be used as biomarkers for clinical diagnosis, treatment response, and prognosis prediction.

6. Conclusions

Pyroptosis has emerged as a novel form of pro-inflammatory RCD, characterized by the GSDM-activated cellular lysis and subsequent release of cellular contents, triggering a potent inflammatory response, as well as an eventual cell death. Pyroptosis is closely associated with the development and progression of cancer, including CRC. A small number of studies suggest that pyroptosis may increase tumor burden [\[160,](#page-18-3)[161\]](#page-18-4). Yet, most research has shown that pyroptosis has a significant anti-tumor effect in CRC. Consequently, pyroptosis is considered as a promising therapeutic strategy for CRC patients. Whether pyroptosis exerts its anti-tumor effects solely through known key activation components requires further investigation. The powerful therapeutic potential of inducing pyroptosis in CRC cells is apparent. It could overcome resistance to conventional chemotherapy drugs and synergize with immunotherapy to enhance anti-tumor immune responses, thereby improving treatment outcomes. The ongoing development of novel inducers targeting pyroptosis, such as biomimetic nanoparticles and supramolecular nanoprodrugs, broadens the therapeutic landscape of pyroptosis in CRC. In addition, numerous pyroptosisrelated genes have emerged as valuable diagnostic prognostic and biomarker candidates in CRC. Overall, studies on pyroptosis in CRC will undoubtedly continue to expand, and a comprehensive understanding of the relationship between pyroptosis and CRC will furthermore bring new hope for preventing and treating CRC.

Author Contributions: S.X., K.F. and Q.F. wrote the main manuscript text; S.X. and K.F. proofread and supervise during writing of the original manuscript. Y.X. made major revisions to this manuscript; X.T., X.W., S.L., J.Y., X.C. and Q.H. collected articles related to the study. All authors have read and agreed to the published version of the manuscript.

Funding: This study was supported by Natural Science Foundation of Hunan Province (2022JJ30538, 2023JJ60368), the National Natural Science Foundation of China (32170726), and the Scientific Research Fund Project of Hunan Provincial Health Commission (20201919, 202104010105).

Conflicts of Interest: There are no conflicts of interest and disclosures associated with the manuscript.

References

- 1. Siegel, R.L.; Miller, K.D.; Wagle, N.S.; Jemal, A. Cancer statistics, 2023. *CA Cancer J. Clin.* **2023**, *73*, 17–48. [\[CrossRef\]](https://doi.org/10.3322/caac.21763) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36633525)
- 2. Zheng, R.; Zhang, S.; Zeng, H.; Wang, S.; Sun, K.; Chen, R.; Li, L.; Wei, W.; He, J. Cancer incidence and mortality in China, 2016. *J. Natl. Cancer Cent.* **2022**, *2*, 1–9. [\[CrossRef\]](https://doi.org/10.1016/j.jncc.2022.02.002)
- 3. Wan, M.L.; Wang, Y.; Zeng, Z.; Deng, B.; Zhu, B.S.; Cao, T.; Li, Y.K.; Xiao, J.; Han, Q.; Wu, Q. Colorectal cancer (CRC) as a multifactorial disease and its causal correlations with multiple signaling pathways. *Biosci. Rep.* **2020**, *40*, BSR20200265. [\[CrossRef\]](https://doi.org/10.1042/BSR20200265) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32149326)
- 4. Brody, H. Colorectal cancer. *Nature* **2015**, *521*, S1. [\[CrossRef\]](https://doi.org/10.1038/521S1a)
- 5. Dekker, E.; Tanis, P.J.; Vleugels, J.L.A.; Kasi, P.M.; Wallace, M.B. Colorectal cancer. *Lancet* **2019**, *394*, 1467–1480. [\[CrossRef\]](https://doi.org/10.1016/S0140-6736(19)32319-0)
- 6. Trosko, J.E. On the potential origin and characteristics of cancer stem cells. *Carcinogenesis* **2021**, *42*, 905–912. [\[CrossRef\]](https://doi.org/10.1093/carcin/bgab042) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34014276)
- 7. Deshpande, A.; Sicinski, P.; Hinds, P.W. Cyclins and cdks in development and cancer: A perspective. *Oncogene* **2005**, *24*, 2909–2915. [\[CrossRef\]](https://doi.org/10.1038/sj.onc.1208618)
- 8. Shi, J.; Zhao, Y.; Wang, K.; Shi, X.; Wang, Y.; Huang, H.; Zhuang, Y.; Cai, T.; Wang, F.; Shao, F. Cleavage of GSDMD by inflammatory caspases determines pyroptotic cell death. *Nature* **2015**, *526*, 660–665. [\[CrossRef\]](https://doi.org/10.1038/nature15514)
- 9. Monack, D.M.; Raupach, B.; Hromockyj, A.E.; Falkow, S. *Salmonella* typhimurium invasion induces apoptosis in infected macrophages. *Proc. Natl. Acad. Sci. USA* **1996**, *93*, 9833–9838. [\[CrossRef\]](https://doi.org/10.1073/pnas.93.18.9833)
- 10. Zychlinsky, A.; Prevost, M.C.; Sansonetti, P.J. *Shigella flexneri* induces apoptosis in infected macrophages. *Nature* **1992**, *358*, 167–169. [\[CrossRef\]](https://doi.org/10.1038/358167a0) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/1614548)
- 11. Galluzzi, L.; Vitale, I.; Aaronson, S.A.; Abrams, J.M.; Adam, D.; Agostinis, P.; Alnemri, E.S.; Altucci, L.; Amelio, I.; Andrews, D.W.; et al. Molecular mechanisms of cell death: Recommendations of the Nomenclature Committee on Cell Death 2018. *Cell Death Differ.* **2018**, *25*, 486–541. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29362479)
- 12. Hilbi, H.; Moss, J.E.; Hersh, D.; Chen, Y.; Arondel, J.; Banerjee, S.; Flavell, R.A.; Yuan, J.; Sansonetti, P.J.; Zychlinsky, A. Shigella-induced apoptosis is dependent on caspase-1 which binds to IpaB. *J. Biol. Chem.* **1998**, *273*, 32895–32900. [\[CrossRef\]](https://doi.org/10.1074/jbc.273.49.32895)
- 13. Boise, L.H.; Collins, C.M. Salmonella-induced cell death: Apoptosis, necrosis or programmed cell death? *Trends Microbiol.* **2001**, *9*, 64–67. [\[CrossRef\]](https://doi.org/10.1016/S0966-842X(00)01937-5) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11173244)
- 14. Cookson, B.T.; Brennan, M.A. Pro-inflammatory programmed cell death. *Trends Microbiol.* **2001**, *9*, 113–114. [\[CrossRef\]](https://doi.org/10.1016/S0966-842X(00)01936-3) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11303500)
- 15. Lu, X.; Guo, T.; Zhang, X. Pyroptosis in Cancer: Friend or Foe? *Cancers* **2021**, *13*, 3620. [\[CrossRef\]](https://doi.org/10.3390/cancers13143620)
- 16. Rao, Z.; Zhu, Y.; Yang, P.; Chen, Z.; Xia, Y.; Qiao, C.; Liu, W.; Deng, H.; Li, J.; Ning, P.; et al. Pyroptosis in inflammatory diseases and cancer. *Theranostics* **2022**, *12*, 4310–4329. [\[CrossRef\]](https://doi.org/10.7150/thno.71086)
- 17. Zhaolin, Z.; Guohua, L.; Shiyuan, W.; Zuo, W. Role of pyroptosis in cardiovascular disease. *Cell Prolif.* **2019**, *52*, e12563. [\[CrossRef\]](https://doi.org/10.1111/cpr.12563)
- 18. Johnson, D.C.; Taabazuing, C.Y.; Okondo, M.C.; Chui, A.J.; Rao, S.D.; Brown, F.C.; Reed, C.; Peguero, E.; de Stanchina, E.; Kentsis, A.; et al. DPP8/DPP9 inhibitor-induced pyroptosis for treatment of acute myeloid leukemia. *Nat. Med.* **2018**, *24*, 1151–1156. [\[CrossRef\]](https://doi.org/10.1038/s41591-018-0082-y)
- 19. Yu, P.; Zhang, X.; Liu, N.; Tang, L.; Peng, C.; Chen, X. Pyroptosis: Mechanisms and diseases. *Signal Transduct. Target. Ther.* **2021**, *6*, 128. [\[CrossRef\]](https://doi.org/10.1038/s41392-021-00507-5)
- 20. Rogers, C.; Erkes, D.A.; Nardone, A.; Aplin, A.E.; Fernandes-Alnemri, T.; Alnemri, E.S. Gasdermin pores permeabilize mitochondria to augment caspase-3 activation during apoptosis and inflammasome activation. *Nat. Commun.* **2019**, *10*, 1689. [\[CrossRef\]](https://doi.org/10.1038/s41467-019-09397-2)
- 21. Verma, D.; Bivik, C.; Farahani, E.; Synnerstad, I.; Fredrikson, M.; Enerbäck, C.; Rosdahl, I.; Söderkvist, P. Inflammasome polymorphisms confer susceptibility to sporadic malignant melanoma. *Pigment Cell Melanoma Res.* **2012**, *25*, 506–513. [\[CrossRef\]](https://doi.org/10.1111/j.1755-148X.2012.01008.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22524199)
- 22. Hergueta-Redondo, M.; Sarrió, D.; Molina-Crespo, Á.; Megias, D.; Mota, A.; Rojo-Sebastian, A.; García-Sanz, P.; Morales, S.; Abril, S.; Cano, A.; et al. Gasdermin-B promotes invasion and metastasis in breast cancer cells. *PLoS ONE* **2014**, *9*, e90099. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0090099) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24675552)
- 23. Miguchi, M.; Hinoi, T.; Shimomura, M.; Adachi, T.; Saito, Y.; Niitsu, H.; Kochi, M.; Sada, H.; Sotomaru, Y.; Ikenoue, T.; et al. Gasdermin C Is Upregulated by Inactivation of Transforming Growth Factor β Receptor Type II in the Presence of Mutated Apc, Promoting Colorectal Cancer Proliferation. *PLoS ONE* **2016**, *11*, e0166422. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0166422) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27835699)
- 24. Wang, W.J.; Chen, D.; Jiang, M.Z.; Xu, B.; Li, X.W.; Chu, Y.; Zhang, Y.J.; Mao, R.; Liang, J.; Fan, D.M. Downregulation of gasdermin D promotes gastric cancer proliferation by regulating cell cycle-related proteins. *J. Dig. Dis.* **2018**, *19*, 74–83. [\[CrossRef\]](https://doi.org/10.1111/1751-2980.12576)
- 25. Wang, C.J.; Tang, L.; Shen, D.W.; Wang, C.; Yuan, Q.Y.; Gao, W.; Wang, Y.K.; Xu, R.H.; Zhang, H. The expression and regulation of DFNA5 in human hepatocellular carcinoma DFNA5 in hepatocellular carcinoma. *Mol. Biol. Rep.* **2013**, *40*, 6525–6531. [\[CrossRef\]](https://doi.org/10.1007/s11033-013-2581-8)
- 26. Gao, J.; Qiu, X.; Xi, G.; Liu, H.; Zhang, F.; Lv, T.; Song, Y. Downregulation of GSDMD attenuates tumor proliferation via the intrinsic mitochondrial apoptotic pathway and inhibition of EGFR/Akt signaling and predicts a good prognosis in non-small cell lung cancer. *Oncol. Rep.* **2018**, *40*, 1971–1984. [\[CrossRef\]](https://doi.org/10.3892/or.2018.6634)
- 27. Zaffaroni, N.; Beretta, G.L. The Therapeutic Potential of Pyroptosis in Melanoma. *Int. J. Mol. Sci.* **2023**, *24*, 1285. [\[CrossRef\]](https://doi.org/10.3390/ijms24021285) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36674798)
- 28. Xia, X.; Wang, X.; Cheng, Z.; Qin, W.; Lei, L.; Jiang, J.; Hu, J. The role of pyroptosis in cancer: Pro-cancer or pro-"host"? *Cell Death Dis.* **2019**, *10*, 650. [\[CrossRef\]](https://doi.org/10.1038/s41419-019-1883-8)
- 29. Hu, B.; Elinav, E.; Huber, S.; Booth, C.J.; Strowig, T.; Jin, C.; Eisenbarth, S.C.; Flavell, R.A. Inflammation-induced tumorigenesis in the colon is regulated by caspase-1 and NLRC4. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 21635–21640. [\[CrossRef\]](https://doi.org/10.1073/pnas.1016814108)
- 30. Janowski, A.M.; Kolb, R.; Zhang, W.; Sutterwala, F.S. Beneficial and Detrimental Roles of NLRs in Carcinogenesis. *Front. Immunol.* **2013**, *4*, 370. [\[CrossRef\]](https://doi.org/10.3389/fimmu.2013.00370)
- 31. Hanahan, D.; Weinberg, R.A. Hallmarks of cancer: The next generation. *Cell* **2011**, *144*, 646–674. [\[CrossRef\]](https://doi.org/10.1016/j.cell.2011.02.013) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21376230)
- 32. Thi, H.T.H.; Hong, S. Inflammasome as a Therapeutic Target for Cancer Prevention and Treatment. *J. Cancer Prev.* **2017**, *22*, 62–73. [\[CrossRef\]](https://doi.org/10.15430/JCP.2017.22.2.62) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28698859)
- 33. Rathinam, V.A.; Vanaja, S.K.; Fitzgerald, K.A. Regulation of inflammasome signaling. *Nat. Immunol.* **2012**, *13*, 333–342. [\[CrossRef\]](https://doi.org/10.1038/ni.2237) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22430786)
- 34. Martinon, F.; Burns, K.; Tschopp, J. The inflammasome: A molecular platform triggering activation of inflammatory caspases and processing of proIL-beta. *Mol. Cell* **2002**, *10*, 417–426. [\[CrossRef\]](https://doi.org/10.1016/S1097-2765(02)00599-3) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12191486)
- 35. Wei, X.; Xie, F.; Zhou, X.; Wu, Y.; Yan, H.; Liu, T.; Huang, J.; Wang, F.; Zhou, F.; Zhang, L. Role of pyroptosis in inflammation and cancer. *Cell. Mol. Immunol.* **2022**, *19*, 971–992. [\[CrossRef\]](https://doi.org/10.1038/s41423-022-00905-x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35970871)
- 36. Broz, P.; Dixit, V.M. Inflammasomes: Mechanism of assembly, regulation and signalling. *Nat. Rev. Immunol.* **2016**, *16*, 407–420. [\[CrossRef\]](https://doi.org/10.1038/nri.2016.58) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27291964)
- 37. Broz, P.; Pelegrín, P.; Shao, F. The gasdermins, a protein family executing cell death and inflammation. *Nat. Rev. Immunol.* **2020**, *20*, 143–157. [\[CrossRef\]](https://doi.org/10.1038/s41577-019-0228-2) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31690840)
- 38. Liu, X.; Zhang, Z.; Ruan, J.; Pan, Y.; Magupalli, V.G.; Wu, H.; Lieberman, J. Inflammasome-activated gasdermin D causes pyroptosis by forming membrane pores. *Nature* **2016**, *535*, 153–158. [\[CrossRef\]](https://doi.org/10.1038/nature18629)
- 39. Williams, T.M.; Leeth, R.A.; Rothschild, D.E.; Coutermarsh-Ott, S.L.; McDaniel, D.K.; Simmons, A.E.; Heid, B.; Cecere, T.E.; Allen, I.C. The NLRP1 inflammasome attenuates colitis and colitis-associated tumorigenesis. *J. Immunol.* **2015**, *194*, 3369–3380. [\[CrossRef\]](https://doi.org/10.4049/jimmunol.1402098)
- 40. Sharma, D.; Malik, A.; Guy, C.S.; Karki, R.; Vogel, P.; Kanneganti, T.D. Pyrin Inflammasome Regulates Tight Junction Integrity to Restrict Colitis and Tumorigenesis. *Gastroenterology* **2018**, *154*, 948–964.e8. [\[CrossRef\]](https://doi.org/10.1053/j.gastro.2017.11.276)
- 41. Jenab, M.; Thompson, L.U. The influence of flaxseed and lignans on colon carcinogenesis and beta-glucuronidase activity. *Carcinogenesis* **1996**, *17*, 1343–1348. [\[CrossRef\]](https://doi.org/10.1093/carcin/17.6.1343) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/8681453)
- 42. Chen, T.; Wang, Z.; Zhong, J.; Zhang, L.; Zhang, H.; Zhang, D.; Xu, X.; Zhong, X.; Wang, J.; Li, H. Secoisolariciresinol diglucoside induces pyroptosis by activating caspase-1 to cleave GSDMD in colorectal cancer cells. *Drug Dev. Res.* **2022**, *83*, 1152–1166. [\[CrossRef\]](https://doi.org/10.1002/ddr.21939) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35472101)
- 43. Liao, P.; Huang, W.H.; Cao, L.; Wang, T.; Chen, L.M. Low expression of FOXP2 predicts poor survival and targets caspase-1 to inhibit cell pyroptosis in colorectal cancer. *J. Cancer* **2022**, *13*, 1181–1192. [\[CrossRef\]](https://doi.org/10.7150/jca.62433)
- 44. Wu, Y.; Pi, D.; Zhou, S.; Yi, Z.; Dong, Y.; Wang, W.; Ye, H.; Chen, Y.; Zuo, Q.; Ouyang, M. Ginsenoside Rh3 induces pyroptosis and ferroptosis through the Stat3/p53/NRF2 axis in colorectal cancer cells. *Acta Biochim. Biophys. Sin.* **2023**, *55*, 587–600. [\[CrossRef\]](https://doi.org/10.3724/abbs.2023068)
- 45. Ren, Q.; Yang, B.; Zhu, G.; Wang, S.; Fu, C.; Zhang, H.; Ross, R.P.; Stanton, C.; Chen, H.; Chen, W. Antiproliferation Activity and Mechanism of c9, t11, c15-CLNA and t9, t11, c15-CLNA from Lactobacillus plantarum ZS2058 on Colon Cancer Cells. *Molecules* **2020**, *25*, 1225. [\[CrossRef\]](https://doi.org/10.3390/molecules25051225) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32182796)
- 46. Kayagaki, N.; Stowe, I.B.; Lee, B.L.; O'Rourke, K.; Anderson, K.; Warming, S.; Cuellar, T.; Haley, B.; Roose-Girma, M.; Phung, Q.T.; et al. Caspase-11 cleaves gasdermin D for non-canonical inflammasome signalling. *Nature* **2015**, *526*, 666–671. [\[CrossRef\]](https://doi.org/10.1038/nature15541) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26375259)
- 47. Kayagaki, N.; Warming, S.; Lamkanfi, M.; Vande Walle, L.; Louie, S.; Dong, J.; Newton, K.; Qu, Y.; Liu, J.; Heldens, S.; et al. Non-canonical inflammasome activation targets caspase-11. *Nature* **2011**, *479*, 117–121. [\[CrossRef\]](https://doi.org/10.1038/nature10558) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22002608)
- 48. Kayagaki, N.; Wong, M.T.; Stowe, I.B.; Ramani, S.R.; Gonzalez, L.C.; Akashi-Takamura, S.; Miyake, K.; Zhang, J.; Lee, W.P.; Muszy´nski, A.; et al. Noncanonical inflammasome activation by intracellular LPS independent of TLR4. *Science* **2013**, *341*, 1246–1249. [\[CrossRef\]](https://doi.org/10.1126/science.1240248)
- 49. Demon, D.; Kuchmiy, A.; Fossoul, A.; Zhu, Q.; Kanneganti, T.D.; Lamkanfi, M. Caspase-11 is expressed in the colonic mucosa and protects against dextran sodium sulfate-induced colitis. *Mucosal Immunol.* **2014**, *7*, 1480–1491. [\[CrossRef\]](https://doi.org/10.1038/mi.2014.36)
- 50. Oficjalska, K.; Raverdeau, M.; Aviello, G.; Wade, S.C.; Hickey, A.; Sheehan, K.M.; Corr, S.C.; Kay, E.W.; O'Neill, L.A.; Mills, K.H.; et al. Protective role for caspase-11 during acute experimental murine colitis. *J. Immunol.* **2015**, *194*, 1252–1260. [\[CrossRef\]](https://doi.org/10.4049/jimmunol.1400501)
- 51. Williams, T.M.; Leeth, R.A.; Rothschild, D.E.; McDaniel, D.K.; Coutermarsh-Ott, S.L.; Simmons, A.E.; Kable, K.H.; Heid, B.; Allen, I.C. Caspase-11 attenuates gastrointestinal inflammation and experimental colitis pathogenesis. *Am. J. Physiol. Gastrointest. Liver. Physiol.* **2015**, *308*, G139–G150. [\[CrossRef\]](https://doi.org/10.1152/ajpgi.00234.2014)
- 52. Wang, Y.; Gao, W.; Shi, X.; Ding, J.; Liu, W.; He, H.; Wang, K.; Shao, F. Chemotherapy drugs induce pyroptosis through caspase-3 cleavage of a gasdermin. *Nature* **2017**, *547*, 99–103. [\[CrossRef\]](https://doi.org/10.1038/nature22393) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28459430)
- 53. Yu, J.; Li, S.; Qi, J.; Chen, Z.; Wu, Y.; Guo, J.; Wang, K.; Sun, X.; Zheng, J. Cleavage of GSDME by caspase-3 determines lobaplatin-induced pyroptosis in colon cancer cells. *Cell Death Dis.* **2019**, *10*, 193. [\[CrossRef\]](https://doi.org/10.1038/s41419-019-1441-4) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30804337)
- 54. Zhang, Z.; Zhang, H.; Li, D.; Zhou, X.; Qin, Q.; Zhang, Q. Caspase-3-mediated GSDME induced Pyroptosis in breast cancer cells through the ROS/JNK signalling pathway. *J. Cell. Mol. Med.* **2021**, *25*, 8159–8168. [\[CrossRef\]](https://doi.org/10.1111/jcmm.16574) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34369076)
- 55. Tan, J.Q.; Li, Z.; Chen, G.; Wu, M.; Feng, J.L.; Kong, S.Y.; Shi, X.Q.; Zhang, H.M.; Lao, Y.Z.; Zhou, H.; et al. The natural compound from *Garcinia bracteata* mainly induces GSDME-mediated pyroptosis in esophageal cancer cells. *Phytomedicine* **2022**, *102*, 154142. [\[CrossRef\]](https://doi.org/10.1016/j.phymed.2022.154142)
- 56. Liu, X.; Zeidan, Y.H.; Elojeimy, S.; Holman, D.H.; El-Zawahry, A.M.; Guo, G.W.; Bielawska, A.; Bielawski, J.; Szulc, Z.; Rubinchik, S.; et al. Involvement of sphingolipids in apoptin-induced cell killing. *Mol. Ther.* **2006**, *14*, 627–636. [\[CrossRef\]](https://doi.org/10.1016/j.ymthe.2006.07.001)
- 57. Liu, Z.; Li, Y.; Zhu, Y.; Li, N.; Li, W.; Shang, C.; Song, G.; Li, S.; Cong, J.; Li, T.; et al. Apoptin induces pyroptosis of colorectal cancer cells via the GSDME-dependent pathway. *Int. J. Biol. Sci.* **2022**, *18*, 717–730. [\[CrossRef\]](https://doi.org/10.7150/ijbs.64350) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35002520)
- 58. Zhou, Z.; He, H.; Wang, K.; Shi, X.; Wang, Y.; Su, Y.; Wang, Y.; Li, D.; Liu, W.; Zhang, Y.; et al. Granzyme A from cytotoxic lymphocytes cleaves GSDMB to trigger pyroptosis in target cells. *Science* **2020**, *368*, eaaz7548. [\[CrossRef\]](https://doi.org/10.1126/science.aaz7548)
- 59. Zhang, Z.; Zhang, Y.; Xia, S.; Kong, Q.; Li, S.; Liu, X.; Junqueira, C.; Meza-Sosa, K.F.; Mok, T.M.Y.; Ansara, J.; et al. Gasdermin E suppresses tumour growth by activating anti-tumour immunity. *Nature* **2020**, *579*, 415–420. [\[CrossRef\]](https://doi.org/10.1038/s41586-020-2071-9)
- 60. Voskoboinik, I.; Whisstock, J.C.; Trapani, J.A. Perforin and granzymes: Function, dysfunction and human pathology. *Nat. Rev. Immunol.* **2015**, *15*, 388–400. [\[CrossRef\]](https://doi.org/10.1038/nri3839)
- 61. Saeki, N.; Kuwahara, Y.; Sasaki, H.; Satoh, H.; Shiroishi, T. Gasdermin (Gsdm) localizing to mouse Chromosome 11 is predominantly expressed in upper gastrointestinal tract but significantly suppressed in human gastric cancer cells. *Mamm. Genome* **2000**, *11*, 718–724. [\[CrossRef\]](https://doi.org/10.1007/s003350010138)
- 62. Tamura, M.; Tanaka, S.; Fujii, T.; Aoki, A.; Komiyama, H.; Ezawa, K.; Sumiyama, K.; Sagai, T.; Shiroishi, T. Members of a novel gene family, Gsdm, are expressed exclusively in the epithelium of the skin and gastrointestinal tract in a highly tissue-specific manner. *Genomics* **2007**, *89*, 618–629. [\[CrossRef\]](https://doi.org/10.1016/j.ygeno.2007.01.003) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17350798)
- 63. Tan, Y.; Chen, Q.; Li, X.; Zeng, Z.; Xiong, W.; Li, G.; Li, X.; Yang, J.; Xiang, B.; Yi, M. Pyroptosis: A new paradigm of cell death for fighting against cancer. *J. Exp. Clin. Cancer Res.* **2021**, *40*, 153. [\[CrossRef\]](https://doi.org/10.1186/s13046-021-01959-x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33941231)
- 64. Jiang, M.; Qi, L.; Li, L.; Li, Y. The caspase-3/GSDME signal pathway as a switch between apoptosis and pyroptosis in cancer. *Cell Death Discov.* **2020**, *6*, 112. [\[CrossRef\]](https://doi.org/10.1038/s41420-020-00349-0) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33133646)
- 65. Mu, M.; Yu, Q.; Zhang, Q.; Guo, J.; Wang, X.; Sun, X.; Yu, J. A pan-cancer analysis of molecular characteristics and oncogenic role of gasdermins. *Cancer Cell Int.* **2022**, *22*, 80. [\[CrossRef\]](https://doi.org/10.1186/s12935-022-02483-4) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35164740)
- 66. Traughber, C.A.; Deshpande, G.M.; Neupane, K.; Bhandari, N.; Khan, M.R.; McMullen, M.R.; Swaidani, S.; Opoku, E.; Muppala, S.; Smith, J.D.; et al. Myeloid-cell-specific role of Gasdermin D in promoting lung cancer progression in mice. *iScience* **2023**, *26*, 106076. [\[CrossRef\]](https://doi.org/10.1016/j.isci.2023.106076) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36844454)
- 67. Yokomizo, K.; Harada, Y.; Kijima, K.; Shinmura, K.; Sakata, M.; Sakuraba, K.; Kitamura, Y.; Shirahata, A.; Goto, T.; Mizukami, H.; et al. Methylation of the DFNA5 gene is frequently detected in colorectal cancer. *Anticancer Res.* **2012**, *32*, 1319–1322. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22493364)
- 68. Fang, Y.; Tian, S.; Pan, Y.; Li, W.; Wang, Q.; Tang, Y.; Yu, T.; Wu, X.; Shi, Y.; Ma, P.; et al. Pyroptosis: A new frontier in cancer. *Biomed. Pharmacother.* **2020**, *121*, 109595. [\[CrossRef\]](https://doi.org/10.1016/j.biopha.2019.109595) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31710896)
- 69. Wu, L.S.; Liu, Y.; Wang, X.W.; Xu, B.; Lin, Y.L.; Song, Y.; Dong, Y.; Liu, J.L.; Wang, X.J.; Liu, S.; et al. LPS Enhances the Chemosensitivity of Oxaliplatin in HT29 Cells via GSDMD-Mediated Pyroptosis. *Cancer Manag. Res.* **2020**, *12*, 10397–10409. [\[CrossRef\]](https://doi.org/10.2147/CMAR.S244374)
- 70. Ivanov, A.I.; Rana, N.; Privitera, G.; Pizarro, T.T. The enigmatic roles of epithelial gasdermin B: Recent discoveries and controversies. *Trends Cell Biol.* **2022**, *33*, 48–59. [\[CrossRef\]](https://doi.org/10.1016/j.tcb.2022.06.006)
- 71. Rana, N.; Privitera, G.; Kondolf, H.C.; Bulek, K.; Lechuga, S.; De Salvo, C.; Corridoni, D.; Antanaviciute, A.; Maywald, R.L.; Hurtado, A.M.; et al. GSDMB is increased in IBD and regulates epithelial restitution/repair independent of pyroptosis. *Cell* **2022**, *185*, 283–298.e17. [\[CrossRef\]](https://doi.org/10.1016/j.cell.2021.12.024) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35021065)
- 72. Sala, R.; Rioja-Blanco, E.; Serna, N.; Sánchez-García, L.; Álamo, P.; Alba-Castellón, L.; Casanova, I.; López-Pousa, A.; Unzueta, U.; Céspedes, M.V.; et al. GSDMD-dependent pyroptotic induction by a multivalent CXCR4-targeted nanotoxin blocks colorectal cancer metastases. *Drug Deliv.* **2022**, *29*, 1384–1397. [\[CrossRef\]](https://doi.org/10.1080/10717544.2022.2069302) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35532120)
- 73. Kim, M.S.; Chang, X.; Yamashita, K.; Nagpal, J.K.; Baek, J.H.; Wu, G.; Trink, B.; Ratovitski, E.A.; Mori, M.; Sidransky, D. Aberrant promoter methylation and tumor suppressive activity of the DFNA5 gene in colorectal carcinoma. *Oncogene* **2008**, *27*, 3624–3634. [\[CrossRef\]](https://doi.org/10.1038/sj.onc.1211021)
- 74. Liao, X.X.; Dai, Y.Z.; Zhao, Y.Z.; Nie, K. Gasdermin E: A Prospective Target for Therapy of Diseases. *Front. Pharmacol.* **2022**, *13*, 855828. [\[CrossRef\]](https://doi.org/10.3389/fphar.2022.855828) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35462927)
- 75. Op de Beeck, K.; Van Camp, G.; Thys, S.; Cools, N.; Callebaut, I.; Vrijens, K.; Van Nassauw, L.; Van Tendeloo, V.F.; Timmermans, J.P.; Van Laer, L. The DFNA5 gene, responsible for hearing loss and involved in cancer, encodes a novel apoptosis-inducing protein. *Eur. J. Hum. Genet.* **2011**, *19*, 965–973. [\[CrossRef\]](https://doi.org/10.1038/ejhg.2011.63)
- 76. Van Rossom, S.; Op de Beeck, K.; Franssens, V.; Swinnen, E.; Schepers, A.; Ghillebert, R.; Caldara, M.; Van Camp, G.; Winderickx, J. The splicing mutant of the human tumor suppressor protein DFNA5 induces programmed cell death when expressed in the yeast Saccharomyces cerevisiae. *Front. Oncol.* **2012**, *2*, 77. [\[CrossRef\]](https://doi.org/10.3389/fonc.2012.00077)
- 77. Liu, X.; Xia, S.; Zhang, Z.; Wu, H.; Lieberman, J. Channelling inflammation: Gasdermins in physiology and disease. *Nat. Rev. Drug Discov.* **2021**, *20*, 384–405. [\[CrossRef\]](https://doi.org/10.1038/s41573-021-00154-z)
- 78. Akino, K.; Toyota, M.; Suzuki, H.; Imai, T.; Maruyama, R.; Kusano, M.; Nishikawa, N.; Watanabe, Y.; Sasaki, Y.; Abe, T.; et al. Identification of DFNA5 as a target of epigenetic inactivation in gastric cancer. *Cancer Sci.* **2007**, *98*, 88–95. [\[CrossRef\]](https://doi.org/10.1111/j.1349-7006.2006.00351.x)
- 79. Kim, M.S.; Lebron, C.; Nagpal, J.K.; Chae, Y.K.; Chang, X.; Huang, Y.; Chuang, T.; Yamashita, K.; Trink, B.; Ratovitski, E.A.; et al. Methylation of the DFNA5 increases risk of lymph node metastasis in human breast cancer. *Biochem. Biophys. Res. Commun.* **2008**, *370*, 38–43. [\[CrossRef\]](https://doi.org/10.1016/j.bbrc.2008.03.026)
- 80. Fujikane, T.; Nishikawa, N.; Toyota, M.; Suzuki, H.; Nojima, M.; Maruyama, R.; Ashida, M.; Ohe-Toyota, M.; Kai, M.; Nishidate, T.; et al. Genomic screening for genes upregulated by demethylation revealed novel targets of epigenetic silencing in breast cancer. *Breast Cancer Res. Treat* **2010**, *122*, 699–710. [\[CrossRef\]](https://doi.org/10.1007/s10549-009-0600-1)
- 81. Hu, L.; Chen, M.; Chen, X.; Zhao, C.; Fang, Z.; Wang, H.; Dai, H. Chemotherapy-induced pyroptosis is mediated by BAK/BAXcaspase-3-GSDME pathway and inhibited by 2-bromopalmitate. *Cell Death Dis.* **2020**, *11*, 281. [\[CrossRef\]](https://doi.org/10.1038/s41419-020-2476-2)
- 82. Yang, X.; Tang, Z. Role of gasdermin family proteins in cancers (Review). *Int. J. Oncol.* **2023**, *63*, 1–13.
- 83. Croes, L.; Fransen, E.; Hylebos, M.; Buys, K.; Hermans, C.; Broeckx, G.; Peeters, M.; Pauwels, P.; Op de Beeck, K.; Van Camp, G. Determination of the Potential Tumor-Suppressive Effects of Gsdme in a Chemically Induced and in a Genetically Modified Intestinal Cancer Mouse Model. *Cancers* **2019**, *11*, 1214. [\[CrossRef\]](https://doi.org/10.3390/cancers11081214) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31434357)
- 84. Watabe, K.; Ito, A.; Asada, H.; Endo, Y.; Kobayashi, T.; Nakamoto, K.; Itami, S.; Takao, S.; Shinomura, Y.; Aikou, T.; et al. Structure, expression and chromosome mapping of MLZE, a novel gene which is preferentially expressed in metastatic melanoma cells. *Jpn. J. Cancer Res.* **2001**, *92*, 140–151. [\[CrossRef\]](https://doi.org/10.1111/j.1349-7006.2001.tb01076.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11223543)
- 85. Zhang, C.C.; Li, C.G.; Wang, Y.F.; Xu, L.H.; He, X.H.; Zeng, Q.Z.; Zeng, C.Y.; Mai, F.Y.; Hu, B.; Ouyang, D.Y. Chemotherapeutic paclitaxel and cisplatin differentially induce pyroptosis in A549 lung cancer cells via caspase-3/GSDME activation. *Apoptosis* **2019**, *24*, 312–325. [\[CrossRef\]](https://doi.org/10.1007/s10495-019-01515-1) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30710195)
- 86. Xu, H.; Zhang, D.; Wei, R.; Zhou, Y.; Dai, G.; Li, J.; Sun, Y.; Li, F.; Xi, L. Gambogic Acid Induces Pyroptosis of Colorectal Cancer Cells through the GSDME-Dependent Pathway and Elicits an Antitumor Immune Response. *Cancers* **2022**, *14*, 5505. [\[CrossRef\]](https://doi.org/10.3390/cancers14225505)
- 87. Saeki, N.; Usui, T.; Aoyagi, K.; Kim, D.H.; Sato, M.; Mabuchi, T.; Yanagihara, K.; Ogawa, K.; Sakamoto, H.; Yoshida, T.; et al. Distinctive expression and function of four GSDM family genes (GSDMA-D) in normal and malignant upper gastrointestinal epithelium. *Genes Chromosomes Cancer* **2009**, *48*, 261–271. [\[CrossRef\]](https://doi.org/10.1002/gcc.20636)
- 88. Liu, X.; Lieberman, J. Knocking e'm Dead: Pore-Forming Proteins in Immune Defense. *Annu. Rev. Immunol.* **2020**, *38*, 455–485. [\[CrossRef\]](https://doi.org/10.1146/annurev-immunol-111319-023800)
- 89. Katoh, M.; Katoh, M. Identification and characterization of human DFNA5L, mouse Dfna5l, and rat Dfna5l genes in silico. *Int. J. Oncol.* **2004**, *25*, 765–770. [\[CrossRef\]](https://doi.org/10.3892/ijo.25.4.1193)
- 90. Ramos-Junior, E.S.; Morandini, A.C. Gasdermin: A new player to the inflammasome game. *Biomed. J.* **2017**, *40*, 313–316. [\[CrossRef\]](https://doi.org/10.1016/j.bj.2017.10.002)
- 91. Kuang, S.; Zheng, J.; Yang, H.; Li, S.; Duan, S.; Shen, Y.; Ji, C.; Gan, J.; Xu, X.W.; Li, J. Structure insight of GSDMD reveals the basis of GSDMD autoinhibition in cell pyroptosis. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, 10642–10647. [\[CrossRef\]](https://doi.org/10.1073/pnas.1708194114) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28928145)
- 92. Liu, Z.; Wang, C.; Rathkey, J.K.; Yang, J.; Dubyak, G.R.; Abbott, D.W.; Xiao, T.S. Structures of the Gasdermin D C-Terminal Domains Reveal Mechanisms of Autoinhibition. *Structure* **2018**, *26*, 778–784.e3. [\[CrossRef\]](https://doi.org/10.1016/j.str.2018.03.002) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29576317)
- 93. Rühl, S.; Shkarina, K.; Demarco, B.; Heilig, R.; Santos, J.C.; Broz, P. ESCRT-dependent membrane repair negatively regulates pyroptosis downstream of GSDMD activation. *Science* **2018**, *362*, 956–960. [\[CrossRef\]](https://doi.org/10.1126/science.aar7607) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30467171)
- 94. Qiu, S.; Hu, Y.; Dong, S. Pan-cancer analysis reveals the expression, genetic alteration and prognosis of pyroptosis key gene GSDMD. *Int. Immunopharmacol.* **2021**, *101 Pt A*, 108270. [\[CrossRef\]](https://doi.org/10.1016/j.intimp.2021.108270)
- 95. Tanaka, S.; Orita, H.; Kataoka, T.; Miyazaki, M.; Saeki, H.; Wada, R.; Brock, M.V.; Fukunaga, T.; Amano, T.; Shiroishi, T. Gasdermin D Represses Inflammation-Induced Colon Cancer Development by Regulating Apoptosis. *Carcinogenesis* **2023**, *44*, 341–349. [\[CrossRef\]](https://doi.org/10.1093/carcin/bgad005) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36753047)
- 96. Ma, Y.; Chen, Y.; Lin, C.; Hu, G. Biological functions and clinical significance of the newly identified long non-coding RNA RP1-85F18.6 in colorectal cancer. *Oncol. Rep.* **2018**, *40*, 2648–2658. [\[CrossRef\]](https://doi.org/10.3892/or.2018.6694) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30226619)
- 97. Feng, W.Q.; Zhang, Y.C.; Xu, Z.Q.; Yu, S.Y.; Huo, J.T.; Tuersun, A.; Zheng, M.H.; Zhao, J.K.; Zong, Y.P.; Lu, A.G. IL-17A-mediated mitochondrial dysfunction induces pyroptosis in colorectal cancer cells and promotes CD8 + T-cell tumour infiltration. *J. Transl. Med.* **2023**, *21*, 335. [\[CrossRef\]](https://doi.org/10.1186/s12967-023-04187-3) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37211606)
- 98. Carl-McGrath, S.; Schneider-Stock, R.; Ebert, M.; Röcken, C. Differential expression and localisation of gasdermin-like (GSDML), a novel member of the cancer-associated GSDMDC protein family, in neoplastic and non-neoplastic gastric, hepatic, and colon tissues. *Pathology* **2008**, *40*, 13–24. [\[CrossRef\]](https://doi.org/10.1080/00313020701716250)
- 99. Sun, Q.; Yang, J.; Xing, G.; Sun, Q.; Zhang, L.; He, F. Expression of GSDML Associates with Tumor Progression in Uterine Cervix Cancer. *Transl. Oncol.* **2008**, *1*, 73–83. [\[CrossRef\]](https://doi.org/10.1593/tlo.08112)
- 100. Söderman, J.; Berglind, L.; Almer, S. Gene Expression-Genotype Analysis Implicates GSDMA, GSDMB, and LRRC3C as Contributors to Inflammatory Bowel Disease Susceptibility. *Biomed. Res. Int.* **2015**, *2015*, 834805. [\[CrossRef\]](https://doi.org/10.1155/2015/834805)
- 101. Zhang, S.; Liang, Y.; Yao, J.; Li, D.F.; Wang, L.S. Role of Pyroptosis in Inflammatory Bowel Disease (IBD): From Gasdermins to DAMPs. *Front. Pharmacol.* **2022**, *13*, 833588. [\[CrossRef\]](https://doi.org/10.3389/fphar.2022.833588) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35677444)
- 102. Hansen, J.M.; de Jong, M.F.; Wu, Q.; Zhang, L.S.; Heisler, D.B.; Alto, L.T.; Alto, N.M. Pathogenic ubiquitination of GSDMB inhibits NK cell bactericidal functions. *Cell* **2021**, *184*, 3178–3191.e18. [\[CrossRef\]](https://doi.org/10.1016/j.cell.2021.04.036) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34022140)
- 103. Wei, J.; Xu, Z.; Chen, X.; Wang, X.; Zeng, S.; Qian, L.; Yang, X.; Ou, C.; Lin, W.; Gong, Z.; et al. Overexpression of GSDMC is a prognostic factor for predicting a poor outcome in lung adenocarcinoma. *Mol. Med. Rep.* **2020**, *21*, 360–370. [\[CrossRef\]](https://doi.org/10.3892/mmr.2019.10837) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31939622)
- 104. Sarrió, D.; Martínez-Val, J.; Molina-Crespo, Á.; Sánchez, L.; Moreno-Bueno, G. The multifaceted roles of gasdermins in cancer biology and oncologic therapies. *Biochim. Biophys. Acta Rev. Cancer* **2021**, *1876*, 188635. [\[CrossRef\]](https://doi.org/10.1016/j.bbcan.2021.188635) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34656686)
- 105. Hou, J.; Zhao, R.; Xia, W.; Chang, C.W.; You, Y.; Hsu, J.M.; Nie, L.; Chen, Y.; Wang, Y.C.; Liu, C.; et al. PD-L1-mediated gasdermin C expression switches apoptosis to pyroptosis in cancer cells and facilitates tumour necrosis. *Nat. Cell Biol.* **2020**, *22*, 1264–1275. [\[CrossRef\]](https://doi.org/10.1038/s41556-020-0575-z) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32929201)
- 106. Zhang, J.Y.; Zhou, B.; Sun, R.Y.; Ai, Y.L.; Cheng, K.; Li, F.N.; Wang, B.R.; Liu, F.J.; Jiang, Z.H.; Wang, W.J.; et al. The metabolite α-KG induces GSDMC-dependent pyroptosis through death receptor 6-activated caspase-8. *Cell Res.* **2021**, *31*, 980–997. [\[CrossRef\]](https://doi.org/10.1038/s41422-021-00506-9)
- 107. Yu, J.; Kang, M.J.; Kim, B.J.; Kwon, J.W.; Song, Y.H.; Choi, W.A.; Shin, Y.J.; Hong, S.J. Polymorphisms in GSDMA and GSDMB are associated with asthma susceptibility, atopy and BHR. *Pediatr. Pulmonol.* **2011**, *46*, 701–708. [\[CrossRef\]](https://doi.org/10.1002/ppul.21424)
- 108. Delmaghani, S.; del Castillo, F.J.; Michel, V.; Leibovici, M.; Aghaie, A.; Ron, U.; Van Laer, L.; Ben-Tal, N.; Van Camp, G.; Weil, D.; et al. Mutations in the gene encoding pejvakin, a newly identified protein of the afferent auditory pathway, cause DFNB59 auditory neuropathy. *Nat. Genet.* **2006**, *38*, 770–778. [\[CrossRef\]](https://doi.org/10.1038/ng1829)
- 109. Schwander, M.; Sczaniecka, A.; Grillet, N.; Bailey, J.S.; Avenarius, M.; Najmabadi, H.; Steffy, B.M.; Federe, G.C.; Lagler, E.A.; Banan, R.; et al. A forward genetics screen in mice identifies recessive deafness traits and reveals that pejvakin is essential for outer hair cell function. *J. Neurosci.* **2007**, *27*, 2163–2175. [\[CrossRef\]](https://doi.org/10.1523/JNEUROSCI.4975-06.2007) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17329413)
- 110. Ding, J.; Wang, K.; Liu, W.; She, Y.; Sun, Q.; Shi, J.; Sun, H.; Wang, D.C.; Shao, F. Pore-forming activity and structural autoinhibition of the gasdermin family. *Nature* **2016**, *535*, 111–116. [\[CrossRef\]](https://doi.org/10.1038/nature18590)
- 111. Zuo, Y.B.; Zhang, Y.F.; Zhang, R.; Tian, J.W.; Lv, X.B.; Li, R.; Li, S.P.; Cheng, M.D.; Shan, J.; Zhao, Z.; et al. Ferroptosis in Cancer Progression: Role of Noncoding RNAs. *Int. J. Biol. Sci.* **2022**, *18*, 1829–1843. [\[CrossRef\]](https://doi.org/10.7150/ijbs.66917) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35342359)
- 112. Koren, E.; Fuchs, Y. Modes of Regulated Cell Death in Cancer. *Cancer Discov.* **2021**, *11*, 245–265. [\[CrossRef\]](https://doi.org/10.1158/2159-8290.CD-20-0789) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33462123)
- 113. Cheng, M.Z.; Yang, B.B.; Zhan, Z.T.; Lin, S.M.; Fang, Z.P.; Gao, Y.; Zhou, W.J. MACC1 and Gasdermin-E (GSDME) regulate the resistance of colorectal cancer cells to irinotecan. *Biochem. Biophys. Res. Commun.* **2023**, *671*, 236–245. [\[CrossRef\]](https://doi.org/10.1016/j.bbrc.2023.06.002) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37307707)
- 114. Chen, C.; Wang, B.; Sun, J.; Na, H.; Chen, Z.; Zhu, Z.; Yan, L.; Ren, S.; Zuo, Y. DAC can restore expression of NALP1 to suppress tumor growth in colon cancer. *Cell Death Dis.* **2015**, *6*, e1602. [\[CrossRef\]](https://doi.org/10.1038/cddis.2014.532) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25611377)
- 115. Ruan, J.; Wang, S.; Wang, J. Mechanism and regulation of pyroptosis-mediated in cancer cell death. *Chem. Biol. Interact.* **2020**, *323*, 109052. [\[CrossRef\]](https://doi.org/10.1016/j.cbi.2020.109052) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32169591)
- 116. Wang, Y.Y.; Liu, X.L.; Zhao, R. Induction of Pyroptosis and Its Implications in Cancer Management. *Front. Oncol.* **2019**, *9*, 971. [\[CrossRef\]](https://doi.org/10.3389/fonc.2019.00971) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31616642)
- 117. Wang, Q.; Wang, Y.; Ding, J.; Wang, C.; Zhou, X.; Gao, W.; Huang, H.; Shao, F.; Liu, Z. A bioorthogonal system reveals antitumour immune function of pyroptosis. *Nature* **2020**, *579*, 421–426. [\[CrossRef\]](https://doi.org/10.1038/s41586-020-2079-1)
- 118. Zaki, M.H.; Vogel, P.; Body-Malapel, M.; Lamkanfi, M.; Kanneganti, T.D. IL-18 production downstream of the Nlrp3 inflammasome confers protection against colorectal tumor formation. *J. Immunol.* **2010**, *185*, 4912–4920. [\[CrossRef\]](https://doi.org/10.4049/jimmunol.1002046) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20855874)
- 119. Hu, B.; Elinav, E.; Huber, S.; Strowig, T.; Hao, L.; Hafemann, A.; Jin, C.; Wunderlich, C.; Wunderlich, T.; Eisenbarth, S.C.; et al. Microbiota-induced activation of epithelial IL-6 signaling links inflammasome-driven inflammation with transmissible cancer. *Proc. Natl. Acad. Sci. USA* **2013**, *110*, 9862–9867. [\[CrossRef\]](https://doi.org/10.1073/pnas.1307575110) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23696660)
- 120. Liu, R.; Truax, A.D.; Chen, L.; Hu, P.; Li, Z.; Chen, J.; Song, C.; Chen, L.; Ting, J.P. Expression profile of innate immune receptors, NLRs and AIM2, in human colorectal cancer: Correlation with cancer stages and inflammasome components. *Oncotarget* **2015**, *6*, 33456–33469. [\[CrossRef\]](https://doi.org/10.18632/oncotarget.5587) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26378020)
- 121. Maloy, K.J.; Powrie, F. Intestinal homeostasis and its breakdown in inflammatory bowel disease. *Nature* **2011**, *474*, 298–306. [\[CrossRef\]](https://doi.org/10.1038/nature10208) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21677746)
- 122. Allen, I.C.; TeKippe, E.M.; Woodford, R.M.; Uronis, J.M.; Holl, E.K.; Rogers, A.B.; Herfarth, H.H.; Jobin, C.; Ting, J.P. The NLRP3 inflammasome functions as a negative regulator of tumorigenesis during colitis-associated cancer. *J. Exp. Med.* **2010**, *207*, 1045–1056. [\[CrossRef\]](https://doi.org/10.1084/jem.20100050) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20385749)
- 123. Zaki, M.H.; Lamkanfi, M.; Kanneganti, T.D. The Nlrp3 inflammasome: Contributions to intestinal homeostasis. *Trends Immunol.* **2011**, *32*, 171–179. [\[CrossRef\]](https://doi.org/10.1016/j.it.2011.02.002) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21388882)
- 124. Flood, B.; Manils, J.; Nulty, C.; Flis, E.; Kenealy, S.; Barber, G.; Fay, J.; Mills, K.H.G.; Kay, E.W.; Creagh, E.M. Caspase-11 regulates the tumour suppressor function of STAT1 in a murine model of colitis-associated carcinogenesis. *Oncogene* **2019**, *38*, 2658–2674. [\[CrossRef\]](https://doi.org/10.1038/s41388-018-0613-5) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30538296)
- 125. Dupaul-Chicoine, J.; Arabzadeh, A.; Dagenais, M.; Douglas, T.; Champagne, C.; Morizot, A.; Rodrigue-Gervais, I.G.; Breton, V.; Colpitts, S.L.; Beauchemin, N.; et al. The Nlrp3 Inflammasome Suppresses Colorectal Cancer Metastatic Growth in the Liver by Promoting Natural Killer Cell Tumoricidal Activity. *Immunity* **2015**, *43*, 751–763. [\[CrossRef\]](https://doi.org/10.1016/j.immuni.2015.08.013) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26384545)
- 126. Tan, G.; Lin, C.; Huang, C.; Chen, B.; Chen, J.; Shi, Y.; Zhi, F. Radiosensitivity of colorectal cancer and radiation-induced gut damages are regulated by gasdermin E. *Cancer Lett.* **2022**, *529*, 1–10. [\[CrossRef\]](https://doi.org/10.1016/j.canlet.2021.12.034) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34979164)
- 127. Su, F.; Duan, J.; Zhu, J.; Fu, H.; Zheng, X.; Ge, C. Long non-coding RNA nuclear paraspeckle assembly transcript 1 regulates ionizing radiation-induced pyroptosis via microRNA-448/gasdermin E in colorectal cancer cells. *Int. J. Oncol.* **2021**, *59*, 79. [\[CrossRef\]](https://doi.org/10.3892/ijo.2021.5259) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34476497)
- 128. Erkes, D.A.; Cai, W.; Sanchez, I.M.; Purwin, T.J.; Rogers, C.; Field, C.O.; Berger, A.C.; Hartsough, E.J.; Rodeck, U.; Alnemri, E.S.; et al. Mutant BRAF and MEK Inhibitors Regulate the Tumor Immune Microenvironment via Pyroptosis. *Cancer Discov.* **2020**, *10*, 254–269. [\[CrossRef\]](https://doi.org/10.1158/2159-8290.CD-19-0672) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31796433)
- 129. Zhao, P.; Wang, M.; Chen, M.; Chen, Z.; Peng, X.; Zhou, F.; Song, J.; Qu, J. Programming cell pyroptosis with biomimetic nanoparticles for solid tumor immunotherapy. *Biomaterials* **2020**, *254*, 120142. [\[CrossRef\]](https://doi.org/10.1016/j.biomaterials.2020.120142)
- 130. Liang, M.Y.; Zhang, M.J.; Qiu, W.; Xiao, Y.; Ye, M.J.; Xue, P.; Kang, Y.J.; Sun, Z.J.; Xu, Z. Stepwise Size Shrinkage Cascade-Activated Supramolecular Prodrug Boosts Antitumor Immunity by Eliciting Pyroptosis. *Adv. Sci.* **2022**, *9*, e2203353. [\[CrossRef\]](https://doi.org/10.1002/advs.202203353)
- 131. Liu, Z.; Xie, Y.; Xiong, Y.; Liu, S.; Qiu, C.; Zhu, Z.; Mao, H.; Yu, M.; Wang, X. TLR 7/8 agonist reverses oxaliplatin resistance in colorectal cancer via directing the myeloid-derived suppressor cells to tumoricidal M1-macrophages. *Cancer Lett.* **2020**, *469*, 173–185. [\[CrossRef\]](https://doi.org/10.1016/j.canlet.2019.10.020) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31629935)
- 132. Yang, F.; Bettadapura, S.N.; Smeltzer, M.S.; Zhu, H.; Wang, S. Pyroptosis and pyroptosis-inducing cancer drugs. *Acta Pharmacol. Sin.* 2022. [\[CrossRef\]](https://doi.org/10.1038/s41401-022-00887-6) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35288674)
- 133. Zhang, Y.; Xu, Z.; Feng, W.; Gao, H.; Xu, Z.; Miao, Y.; Li, W.; Chen, F.; Lv, Z.; Huo, J.; et al. Small molecule inhibitors from organoid-based drug screen induce concurrent apoptosis and gasdermin E-dependent pyroptosis in colorectal cancer. *Clin. Transl. Med.* **2022**, *12*, e812. [\[CrossRef\]](https://doi.org/10.1002/ctm2.812) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35415883)
- 134. Han, W.; Xing, W.; Wang, K.; Wang, B.; Bai, K. Alisol A attenuates malignant phenotypes of colorectal cancer cells by inactivating PI3K/Akt signaling. *Oncol. Lett.* **2022**, *24*, 249. [\[CrossRef\]](https://doi.org/10.3892/ol.2022.13369) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35761944)
- 135. Tang, Z.; Ji, L.; Han, M.; Xie, J.; Zhong, F.; Zhang, X.; Su, Q.; Yang, Z.; Liu, Z.; Gao, H.; et al. Pyroptosis is involved in the inhibitory effect of FL118 on growth and metastasis in colorectal cancer. *Life Sci.* **2020**, *257*, 118065. [\[CrossRef\]](https://doi.org/10.1016/j.lfs.2020.118065) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32659366)
- 136. Guo, J.; Zheng, J.; Mu, M.; Chen, Z.; Xu, Z.; Zhao, C.; Yang, K.; Qin, X.; Sun, X.; Yu, J. GW4064 enhances the chemosensitivity of colorectal cancer to oxaliplatin by inducing pyroptosis. *Biochem. Biophys. Res. Commun.* **2021**, *548*, 60–66. [\[CrossRef\]](https://doi.org/10.1016/j.bbrc.2021.02.043) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33631675)
- 137. Rana, S.; Espinosa-Diez, C.; Ruhl, R.; Chatterjee, N.; Hudson, C.; Fraile-Bethencourt, E.; Agarwal, A.; Khou, S.; Thomas, C.R., Jr.; Anand, S. Differential regulation of microRNA-15a by radiation affects angiogenesis and tumor growth via modulation of acid sphingomyelinase. *Sci. Rep.* **2020**, *10*, 5581. [\[CrossRef\]](https://doi.org/10.1038/s41598-020-62621-8) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32221387)
- 138. Jiang, R.; Chen, X.; Ge, S.; Wang, Q.; Liu, Y.; Chen, H.; Xu, J.; Wu, J. MiR-21-5p Induces Pyroptosis in Colorectal Cancer via TGFBI. *Front. Oncol.* **2020**, *10*, 610545. [\[CrossRef\]](https://doi.org/10.3389/fonc.2020.610545) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33614494)
- 139. Zou, W. Immunosuppressive networks in the tumour environment and their therapeutic relevance. *Nat. Rev. Cancer* **2005**, *5*, 263–274. [\[CrossRef\]](https://doi.org/10.1038/nrc1586)
- 140. Mohme, M.; Riethdorf, S.; Pantel, K. Circulating and disseminated tumour cells—Mechanisms of immune surveillance and escape. *Nat. Rev. Clin. Oncol.* **2017**, *14*, 155–167. [\[CrossRef\]](https://doi.org/10.1038/nrclinonc.2016.144)
- 141. Wei, S.C.; Duffy, C.R.; Allison, J.P. Fundamental Mechanisms of Immune Checkpoint Blockade Therapy. *Cancer Discov.* **2018**, *8*, 1069–1086. [\[CrossRef\]](https://doi.org/10.1158/2159-8290.CD-18-0367) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30115704)
- 142. Tang, R.; Xu, J.; Zhang, B.; Liu, J.; Liang, C.; Hua, J.; Meng, Q.; Yu, X.; Shi, S. Ferroptosis, necroptosis, and pyroptosis in anticancer immunity. *J. Hematol. Oncol.* **2020**, *13*, 110. [\[CrossRef\]](https://doi.org/10.1186/s13045-020-00946-7) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32778143)
- 143. Catanzaro, E.; Feron, O.; Skirtach, A.G.; Krysko, D.V. Immunogenic Cell Death and Role of Nanomaterials Serving as Therapeutic Vaccine for Personalized Cancer Immunotherapy. *Front. Immunol.* **2022**, *13*, 925290. [\[CrossRef\]](https://doi.org/10.3389/fimmu.2022.925290) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35844506)
- 144. Xiao, Y.; Zhang, T.; Ma, X.; Yang, Q.C.; Yang, L.L.; Yang, S.C.; Liang, M.; Xu, Z.; Sun, Z.J. Microenvironment-Responsive Prodrug-Induced Pyro44ptosis Boosts Cancer Immunotherapy. *Adv. Sci. (Weinh)* **2021**, *8*, e2101840. [\[CrossRef\]](https://doi.org/10.1002/advs.202101840) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34705343)
- 145. Yu, Y.Q.; Gamez-Belmonte, R.; Patankar, J.V.; Liebing, E.; Becker, C. The Role of Programmed Necrosis in Colorectal Cancer. *Cancers* **2022**, *14*, 4295. [\[CrossRef\]](https://doi.org/10.3390/cancers14174295) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36077828)
- 146. Ji, F.; Shi, C.; Shu, Z.; Li, Z. Nanomaterials Enhance Pyroptosis-Based Tumor Immunotherapy. *Int. J. Nanomed.* **2024**, *19*, 5545–5579. [\[CrossRef\]](https://doi.org/10.2147/IJN.S457309) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38882539)
- 147. Serna, N.; Álamo, P.; Ramesh, P.; Vinokurova, D.; Sánchez-García, L.; Unzueta, U.; Gallardo, A.; Céspedes, M.V.; Vázquez, E.; Villaverde, A.; et al. Nanostructured toxins for the selective destruction of drug-resistant human CXCR4(+) colorectal cancer stem cells. *J. Control. Release* **2020**, *320*, 96–104. [\[CrossRef\]](https://doi.org/10.1016/j.jconrel.2020.01.019) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31931052)
- 148. Jiang, X.; Zhu, Z.; Ding, L.; Du, W.; Pei, D. ALKBH4 impedes 5-FU Sensitivity through suppressing GSDME induced pyroptosis in gastric cancer. *Cell Death Dis.* **2024**, *15*, 435. [\[CrossRef\]](https://doi.org/10.1038/s41419-024-06832-1) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38902235)
- 149. Tan, L.; Zhang, H.; Ding, Y.; Huang, Y.; Sun, D. CRTAC1 identified as a promising diagnosis and prognostic biomarker in lung adenocarcinoma. *Sci. Rep.* **2024**, *14*, 11223. [\[CrossRef\]](https://doi.org/10.1038/s41598-024-61804-x)
- 150. Lu, H.; Sun, Y.; Zhu, Z.; Yao, J.; Xu, H.; Huang, R.; Huang, B. Pyroptosis is related to immune infiltration and predictive for survival of colon adenocarcinoma patients. *Sci. Rep.* **2022**, *12*, 9233. [\[CrossRef\]](https://doi.org/10.1038/s41598-022-13212-2)
- 151. Dai, J.; Chen, S.; Bai, Y.; Fu, Y.; Pan, Y.; Ye, L. A Novel Pyroptosis-Associated Gene Signature to Predict Prognosis in Patients with Colorectal Cancer. *Evid. Based Complement. Alternat. Med.* **2022**, *2022*, 6965308. [\[CrossRef\]](https://doi.org/10.1155/2022/6965308) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35620407)
- 152. Wu, Z.; Wang, B.; Ye, Y.; Wang, S.; Jiang, K. Development and verification of a prognostic model for colon cancer on pyroptosisrelated genes. *Front. Genet.* **2022**, *13*, 922055. [\[CrossRef\]](https://doi.org/10.3389/fgene.2022.922055) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36246625)
- 153. Chen, Y.; Tian, Z.; Hou, H.; Gai, W. The noncoding RNAs regulating pyroptosis in colon adenocarcinoma were derived from the construction of a ceRNA network and used to develop a prognostic model. *BMC Med. Genom.* **2022**, *15*, 201. [\[CrossRef\]](https://doi.org/10.1186/s12920-022-01359-w) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36127676)
- 154. Liu, Q.; Chen, N.; Liu, L.; Zheng, Q.; Liao, W.; Zhao, M.; Zeng, J.; Tang, J. Construction and Validation of Pyroptosis-Related lncRNA Prediction Model for Colon Adenocarcinoma and Immune Infiltration Analysis. *Dis. Markers* **2022**, *2022*, 4492608. [\[CrossRef\]](https://doi.org/10.1155/2022/4492608) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36168326)
- 155. Li, Z.; Liu, Y.; Lin, B.; Yan, W.; Yi, H.; Wang, H.; Wei, Y. Pyroptosis-Related Signature as Potential Biomarkers for Predicting Prognosis and Therapy Response in Colorectal Cancer Patients. *Front. Genet.* **2022**, *13*, 925338. [\[CrossRef\]](https://doi.org/10.3389/fgene.2022.925338) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35937993)
- 156. Hu, J.; Tian, C.; Zhao, Y.; Guo, Y.; Chen, S. Prognostic prediction of systemic immune-inflammation status for patients with colorectal cancer: A novel pyroptosis-related model. *World J. Surg. Oncol.* **2022**, *20*, 234. [\[CrossRef\]](https://doi.org/10.1186/s12957-022-02697-w) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35836259)
- 157. Qin, Y.; Pan, L.; Qin, T.; Ruan, H.; Zhang, Y.; Zhang, Y.; Li, J.; Yang, J.; Li, W. Pan-cancer analysis of AIM2 inflammasomes with potential implications for immunotherapy in human cancer: A bulk omics research and single cell sequencing validation. *Front. Immunol.* **2022**, *13*, 998266. [\[CrossRef\]](https://doi.org/10.3389/fimmu.2022.998266)
- 158. Dihlmann, S.; Tao, S.; Echterdiek, F.; Herpel, E.; Jansen, L.; Chang-Claude, J.; Brenner, H.; Hoffmeister, M.; Kloor, M. Lack of Absent in Melanoma 2 (AIM2) expression in tumor cells is closely associated with poor survival in colorectal cancer patients. *Int. J. Cancer* **2014**, *135*, 2387–2396. [\[CrossRef\]](https://doi.org/10.1002/ijc.28891) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24729378)
- 159. Tan, G.; Huang, C.; Chen, J.; Zhi, F. HMGB1 released from GSDME-mediated pyroptotic epithelial cells participates in the tumorigenesis of colitis-associated colorectal cancer through the ERK1/2 pathway. *J. Hematol. Oncol.* **2020**, *13*, 149. [\[CrossRef\]](https://doi.org/10.1186/s13045-020-00985-0)
- 160. Yu, X.; He, S. GSDME as an executioner of chemotherapy-induced cell death. *Sci. China Life Sci.* **2017**, *60*, 1291–1294. [\[CrossRef\]](https://doi.org/10.1007/s11427-017-9142-2)
- 161. Hu, B.; Jin, C.; Li, H.B.; Tong, J.; Ouyang, X.; Cetinbas, N.M.; Zhu, S.; Strowig, T.; Lam, F.C.; Zhao, C.; et al. The DNA-sensing AIM2 inflammasome controls radiation-induced cell death and tissue injury. *Science* **2016**, *354*, 765–768. [\[CrossRef\]](https://doi.org/10.1126/science.aaf7532) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27846608)

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.