



Review

An Overview of Abiotic Stress in Cereal Crops: Negative Impacts, Regulation, Biotechnology and Integrated Omics

Rajendran Jeyasri ^{1,†}, Pandiyan Muthuramalingam ^{1,2,†}, Lakkakula Satish ^{1,3}, Shunmugiah Karutha Pandian ¹, Jen-Tsung Chen ⁴, Sunny Ahmar ⁵, Xiukang Wang ⁶, Freddy Mora-Poblete ^{5,*} and Manikandan Ramesh ^{1,*}

- Department of Biotechnology, Science Campus, Alagappa University, Karaikudi 630003, India; jeyasri8220@gmail.com (R.J.); pandianmuthuramalingam@gmail.com (P.M.); lsatish@post.bgu.ac.il (L.S.); pandiansk@gmail.com (S.K.P.)
- Department of Biotechnology, Sri Shakthi Institute of Engineering and Technology, Coimbatore 641062, India
- Department of Biotechnology Engineering, Ben-Gurion University of the Negev, Beer Sheva 84105, Israel
- Department of Life Sciences, National University of Kaohsiung, Kaohsiung 81148, Taiwan; jentsung@nuk.edu.tw
- Institute of Biological Sciences, University of Talca, 2 Norte 685, Talca 3460000, Chile; sunnyahmar13@gmail.com
- ⁶ College of Life Sciences, Yan'an University, Yan'an 716000, China; wangxiukang@yau.edu.cn
- * Correspondence: morapoblete@gmail.com (F.M.-P.); mrbiotech.alu@gmail.com (M.R.)
- † These authors contributed equally to this review.

Abstract: Abiotic stresses (AbS), such as drought, salinity, and thermal stresses, could highly affect the growth and development of plants. For decades, researchers have attempted to unravel the mechanisms of AbS for enhancing the corresponding tolerance of plants, especially for crop production in agriculture. In the present communication, we summarized the significant factors (atmosphere, soil and water) of AbS, their regulations, and integrated omics in the most important cereal crops in the world, especially rice, wheat, sorghum, and maize. It has been suggested that using systems biology and advanced sequencing approaches in genomics could help solve the AbS response in cereals. An emphasis was given to holistic approaches such as, bioinformatics and functional omics, gene mining and agronomic traits, genome-wide association studies (GWAS), and transcription factors (TFs) family with respect to AbS. In addition, the development of omics studies has improved to address the identification of AbS responsive genes and it enables the interaction between signaling pathways, molecular insights, novel traits and their significance in cereal crops. This review compares AbS mechanisms to omics and bioinformatics resources to provide a comprehensive view of the mechanisms. Moreover, further studies are needed to obtain the information from the integrated omics databases to understand the AbS mechanisms for the development of large spectrum AbS-tolerant crop production.

Keywords: abiotic stress; GWAS; *Oryza sativa* L.; plant omics; *Triticum aestivum* L.; *Sorghum bicolor* L.; transcription factors; *Zea mays* L.



Citation: Jeyasri, R.; Muthuramalingam, P.; Satish, L.; Pandian, S.K.; Chen, J.-T.; Ahmar, S.; Wang, X.; Mora-Poblete, F.; Ramesh, M. An Overview of Abiotic Stress in Cereal Crops: Negative Impacts, Regulation, Biotechnology and Integrated Omics. *Plants* **2021**, *10*, 1472. https://doi.org/10.3390/ plants10071472

Academic Editors: Małgorzata Nykiel, Mateusz Labudda, Beata Prabucka, Marta Gietler and Iustyna Fidler

Received: 1 June 2021 Accepted: 16 July 2021 Published: 19 July 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/licenses/by/4.0/).

1. Introduction

Cereals are grasses (a monocot family Poaceae, also known as Gramineae) cultivated for the edible components of the grain. The most important staple cereal crops are wheat, rice, maize, sorghum, barley, oats, and millet. These cereals are cultivated for the edible components of their caryopsis, composed of the endosperm, germ, and bran. These plants have evolved to live in environments where they are often exposed to various stressors such as high temperature (HT), drought, salinity and mineral toxicity, and the water deficiency [1,2]. Cereals are widely utilized crops in world agriculture, with an overall production of 2500 million tonnes being harvested globally in 2011. On a worldwide basis, rice, wheat and maize are the three most important cereal crops, which together comprise at least 75% of the world's grain production. Also, in 2011 723, 704, and 883 million tonnes

Plants 2021, 10, 1472 2 of 19

of rice, wheat, and maize were harvested, respectively. Cereals contain major nutritional and energy sources such as proteins, carbohydrates, minerals, amino acids, fiber, and micronutrients such as vitamins, magnesium, and zinc for the global population [3,4]. Asia, America, and Europe produce 80% of the world's cereal grains. Rice, sorghum, millet and wheat are widely produced in Asia; likewise, corn and sorghum in America and barley, rye, and oats in Europe. Cereals are a pivotal nutrient source in both developed and developing countries, however, the utilization pattern of these cereal grains differs. In developed countries, more than 70% of total cereal production is fed to the animals, whereas in underdeveloped countries, 68 to 98% of the cereal production is used for human consumption [5].

Abiotic stresses (AbS) (predominantly drought, cold, salinity, and heat), adversely affect diverse plant developmental stages. They are highly complex and affect the various plant dynamisms at the transcriptome, cellular, and physiological processes such as flowering, grain filling, and maturation [6,7]. The ability of cereal crops to tolerate dominant AbS comprises water deficit (drought), flood (anoxia), salinity, high/low temperature, and other osmotic stresses is an essential aspect of yield resilience, and its improvement has long been a target for plant breeders and researchers [8]. The rapidly changing global climate is affecting crop productivity and food availability due to the ever-increasing population, resulting in a demand for stress-tolerant crop varieties that have never been greater [9,10]. Understanding the molecular cross-talk of plant responses to various stresses is crucial in providing opportunities for the development of broad-spectrum stress-tolerant crops. As a result, it is crucial to understand the AbS tolerance dynamic while also devising a new and improved approach for dealing with their detrimental influence on the agricultural sector. The recent game-changing advances in bioinformatics and integrating omics technologies could serve as the most immediate and prospective strategies for improving AbS tolerance in cereal crops. Omics approaches lead to understanding the stress tolerance mechanisms at the molecular level including genomics, functional genomics, genetic engineering, gene expression, protein or metabolite profile(s), and their overall phenotypic effects. In crop breeding techniques, the identification and characterization of the genes and the specific genetic regions associated with both the quantitative and qualitative agronomic traits have been a major challenge. In recent breeding programs, a high-throughput marker-assisted system is extensively being used to enhance selection accuracy and efficiency.

2. Cereal Crops

2.1. Comparative Nutritive Values of Cereal Crops

Cereal grains have low protein content when compared to food legumes and oilseeds, with rice being the lowest. Among the essential amino acids for humans, lysine is the most limiting in all cereal grains. Most cereal proteins are rich in cysteine, methionine and sulfur-containing amino acids. The biological value (BV) ranges from 55 to 77.7%, protein digestibility (TD) 77 to 99.7%, and net protein utilization (NPU) 50 to 73.8% in different cereal diets fed to growing rats. Barley contains relatively more amounts of lysine compared with other cereal crops. The utilization of legumes is known to be affected by the presence of several antinutrients, such as metal chelates, antivitamins, goitrogens, cyanogens, inhibitors of proteases and amylases, toxic phenolic glycosides, and amino acid derivatives [11]. Hence, proper processing of cereal-legume mixture is required to minimize these antinutrients before consumption. Compared with animal foods cereal grains products are inferior in both nutritional and sensory qualities. Physical, chemical, biological, and/or physiological modifications can improve both the nutritional and evident qualities of the grains [12]. Moreover, natural processes like fermentation and controlled germination with natural microflora are highly beneficial in improving the quality of cerealbased food. Important cereal crops and their comparative nutritive values are given in Table 1.

Plants **2021**, 10, 1472 3 of 19

Factor	Wheat	Maize	Rice	Barley	Sorghum	Oat	Millet	Rye
Available CHO (%)	69.7	63.6	64.3	55.8	62.9	62.9	63.4	71.8
Energy (kJ/100 g)	1570	1660	1610	1630	1610	1640	1650	1570
Digestible energy (%)	86.4	87.2	96.3	81.0	79.9	70.6	87.2	85.0
Vitamins (mg/100 g)								
Thiamin	0.45	0.32	0.29	0.10	0.33	0.60	0.63	0.66
Riboflavin	0.10	0.10	0.04	0.04	0.13	0.14	0.33	0.25
Niacin	3.7	1.9	4.0	2.7	3.4	1.3	2.0	1.3
Amino acids (g/16 g N)								
Lysine	2.3	2.5	3.8	3.2	2.7	4.0	2.7	3.7
Threonine	2.8	3.2	3.6	2.9	3.3	3.6	3.2	3.3
Met. & Cys.	3.6	3.9	3.9	3.9	2.8	4.8	3.6	3.7
Tryptophan	1.0	0.6	1.1	1.7	1.0	0.9	1.3	1.0
Protein quality (%)								
True digestibility	96.0	95.0	99.7	88.0	84.8	84.1	93.0	77.0
Biological value	55.0	61.0	74.0	70.0	59.2	70.4	60.0	77.7
Net protein utilization	53.0	58.0	73.8	62.0	50.0	59.1	56.0	59.0
Utilization protein	5.6	5.7	5.4	6.8	4.2	5.5	6.4	5.1

Table 1. Comparative nutritive value of cereal grains.

2.2. Rice (Oryza sativa L.)

Rice is the second most widely consumed cereal, serving as a staple food for more than half of the world population and 90% of Asians. Rice, known as the grain of life, contains 80% carbohydrates, 7–8% protein, 3% fat, and 3% fiber [13]. In most countries, rice being the most dominant cereal crop can improve the health condition of millions of people who consume it. It plays an important role in health benefits and Lifestyle-related disease prevention such as high blood pressure, cancer prevention, Alzheimer's diseases, skincare, diabetes, heart disease, and dysentery [14–16]. However, rice plants are severely affected by various AbS, major stressors such as drought, cold, salinity, and high temperature [17].

Excess salinity in the soil is one of the major abiotic stress factors that affect the growth and productivity of a wide variety of crops including rice. Generally, rice can tolerate a modest amount of saltwater without affecting its growth and yield. However, it highly depends on the types and species of rice used as well as their growth stage [18]. According to Lee et al. [19], indica has a higher tolerance level than japonica at the seedlings stage. Rice is grouped as a salinity-sensitive cereal at an early stage of growth, which limits its production efficiency at the mature stage [20–22].

2.3. Maize (Zea mays L.)

Maize or corn is an important cereal crop grown in diverse agro-ecological zones and farming systems and socio-economic backgrounds in sub-Saharan Africa (SSA) [23]. After rice and wheat, maize is the world's third most important crop, and it is known as the "Queen of Cereals" because it has the highest production potential of all the cereals [24]. It is a predominant source of nutrition as well as phytochemical compounds such as carotenoids, phenolic compounds, and phytosterols [25]. Phytochemicals are naturally occurring bioactive compounds in plants that provide human health benefits while also preventing the risk of major chronic diseases [25,26]. Maize is believed to have potential anti-HIV activity due to the presence of *Galanthus nivalis* agglutinin (GNA) lectin or GNA-maize [25]. The studies revealed that phytochemicals in grains due to their potent antioxidant activities demonstrate significantly reducing the risk of many diseases such as cardiovascular disease, type 2 diabetes, Diet-related disorders, and cancers [27].

The growth and productivity of maize are severely affected by several abiotic stresses such as salinity drought, waterlogging, cold, and nitrogen stress. Among them, salinity stress causes several biochemical and physiological changes in maize, such as disruption of cellular homeostasis, ionic imbalance, nitrogen fixation, respiration, inhibition of several metabolic enzymes such as photosynthetic enzymes etc (enzyme toxicity) [28]. Drought is

Plants 2021, 10, 1472 4 of 19

one of the most detrimental AbS which are seriously affecting the productivity of cereal crops. However, while maize is known as the "Queen of Cereals" across the world, it is susceptible to drought. Drought can affect kernel weight following silking stage up to maturity. Severe drought can reduce the yields of maize during this period by 20 to 30%. During the grain filling stage because of reduced photosynthesis accelerated leaf senescence takes place [29]. More recently, new biotechnological tools have emerged to further accelerate the grain selection and improvement for enhancing the tolerance to drought conditions.

2.4. Wheat (Triticum aestivum L.)

Wheat is a major staple crop for several countries, grown on around 10 million ha in Africa and it comes from a type of grass (*T. aestivum* L.) that is grown in countless varieties worldwide. As a result of a growing population, changing food preferences, and socioeconomic change associated with urbanization and industrialization, wheat consumption steadily increased during the past two decades in all African countries [30,31]. Wheat consumption provides up to 50% of daily calories and proteins. Wheat is often considered to be a source of energy (carbohydrate) and also contains a significant amount of other important nutrients including proteins, fiber, and minor components such as lipids, vitamins, minerals, and phytochemicals that may significantly contribute to the individual diet [32,33]. It reduces the risk of cardiovascular disease, type 2 diabetes, and forms of cancer (notably colorectal cancer) [34,35]. Dietary fiber components also have high heritability and are amenable to manipulation by breeding. Therefore, plant breeders should be able to select plants with enhanced health benefits in addition to increased crop productivity [36,37].

However, wheat plants are severely affected by different AbS, such as salinity, drought, cold, and heat. Heat stress affects wheat growth and yield, particularly at grain developmental stages. The effect of a 3-day heat shock on biomass production was less than the pre- and post-treatment growing temperature [38]. Salinity stress also affects the growth and productivity of wheat. The increasing salinity of irrigation water had a significant adverse effect on the yield of wheat. Domestic wheat lines could be grown at salinity concentrations ranging from 4 and 8 g/L with minimal reduction in biological and grain yield. Moreover, many physiological and biochemical approaches have been developed in plants to survive at high salt concentration. The most effective approach to solve the salt problem is to improve wheat adaptation under salinity stress conditions and enhance its grain yield. Various biotechnological approaches are required to understand the genetic and physiological mechanisms of natural differences in salinity tolerance of wheat and to obtain methods to explore the inherent genetic differences, to get new candidate genes for improving salt tolerance in wheat [39,40].

2.5. Sorghum (Sorghum bicolor L.)

Sorghum is the fifth most important food and feed crop in the world. It is the main cereal food for semi-arid tropical regions of Africa, Asia, and Latin America. Sorghum species (*S. vulgare* and *S. bicolor*) are members of the grass family. Sorghum is resistant to drought and water-logging and is grown in different soil conditions [41]. It is mainly composed of starch, protein and unsaturated fatty acids and is an essential source of some vitamins and minerals [42]. Sorghum is the most abundant and omnivorous cereal secondary metabolites of plants, including up to 6% of 3-deoxyanthocyanidine, phenolic acid, flavonoids, and tannins [43,44]. Phenolic and soluble compounds play an important role in balancing or stabilizing the intestinal microbiota and the parameters associated with obesity, oxidative stress, inflammation, diabetes, dyslipidemia, hypertension, and cancer [45].

Among the various AbS, drought and temperature stress are of foremost important that limits sorghum production. Severe drought causes considerable yield loss in sorghum, and it has a greater impact on grain filling and flowering compared to vegetative stage.

Plants 2021, 10, 1472 5 of 19

Drought adversely affects various physiological functions, inflorescence development and leaf growth of the sorghum [46,47]. Interestingly, genome and transcriptome sequencing, annotation projects and recent literatures paved the way to identify the candidate genes through omics databases, which are predicted to be involved in individual and combined abiotic stress (CAbS) responses. This claim supports to understanding the stress tolerance mechanism and environmental adaptation, as well as promoting the green revolution of all other food crops [47,48].

3. Abiotic Stress (AbS) Dynamism on Cereal Crops

The atmosphere, soil, water and their associated factors are the major abiotic stressors affecting modern agricultural systems [49].

3.1. Atmospheric Factors

3.1.1. Rainfall

In semiarid regions, rainfall is one of the primary AbS factors affecting soil erosion and crop production in rain-fed agriculture. It controls soil salinity and acidic properties. Sulfur dioxide (SO_2) and nitrogen oxides (NOx) from fossil fuel burning merge with water and oxygen in the atmosphere resulting in acid rain. It also reduces the soil pH and removes nutrients and minerals from the soil that can be harmful to plants [50].

3.1.2. Temperature

The plant underwent a high temperature that caused certain mechanical damages including expansion-induced-lysis, phase transitions and fracture lesions in membranes, and physical damage [51]. Depending on the temperature plant species have been classified into three groups: chilling-sensitive, freezing-sensitive, and freezing-resistant plants [52]. Freezing may alter the growth and can cause frost-hardening/cold hardening and also induces the production of reactive oxygen species (ROS), which damages membrane components, and results in protein denaturation [53,54]. Chilling stress including reduced leaf expansion and wilting, affects the reproductive development of plants, chlorosis, and may lead to necrosis.

3.1.3. Gases

An increase in the level of greenhouse gases in the atmosphere such as carbon dioxide (CO_2), methane (CH_4), nitrous oxide (N_2O), ozone (O_3), water vapor (H_2O), and some artificial chemicals such as chlorofluorocarbons (CFCs) increases the atmospheric temperature. As CO_2 levels increase in the atmosphere, nitrogen concentration is decreased in plants. Thus, decreasing the protein levels and affects the growth ability of plants [55].

3.1.4. Radiation

Ultraviolet (UV) and ionizing radiations affect plant growth and development in many ways. Radiations may disrupt stomatal resistance, damage plant cells, increases cell mutations, prevents seed growth, and reduce plant fertility [56,57]. Photon irradiation (such as H_2O , CO_2 , CH_4 , N_2O , and O_3) generates cellular damages in root and leaf tissue.

3.1.5. Wind

Plant tissues can be damaged during hot, humid, hazy weather with little wind. Wind decreases the phytohormonal content of roots and shoots in cereal crops. The direction and velocity of the wind will affect plant growth and development [49].

3.2. Soil Factors

3.2.1. Soil Properties

Salt stress (salinity) causes multifarious effects in plants such as ionic effect, osmotic effect, nutrient and hormonal imbalance, and production of ROS [58]. Plant growth and productivity are severely affected by the accumulation of sodium (Na⁺) and chloride (Cl⁻)

Plants 2021, 10, 1472 6 of 19

ions that leads to creating ionic, osmotic, and oxidative stresses [59–63]. Sodium-ion (Na⁺) has been detected to intervene in multiple metabolic processes such as protein translation, transcription, and enzyme activity that ultimately led to osmotic stress. Functional genomic approaches provide new opportunities to unravel the functional roles of AbS responsive genes, enabling the identification of genes and generation of stress-tolerant plants especially on important cereal crops [64]. Plants can generally tolerate salinity stress mainly by three mechanisms, which include ion exclusion, osmotic tolerance and tissue tolerance. Osmotic tolerance is regulated by long-distance signaling waves that reduce cell expansion in root tips, leaves, and stomatal conductance [65,66]. Ion exclusion mainly involves the transport of Na⁺ and Cl⁻ into roots which prevent the reduction of Na⁺ accumulation in shoots. Tissue tolerance involves the exposure of tissues to the accumulated Na⁺ and Cl⁻ at cellular and intracellular levels, synthesis of compatible solutes, and production of the enzyme catalyzing detoxification of ROS [67].

3.2.2. Pollution

Soil contamination was widespread as a result of the rapid application of some harmful pesticide compounds which can infiltrate the natural environment in two ways depending on their solubility. The pesticide-contaminated soil may affect the availability of nutrients in plants. Rapid industrialization near agricultural lands may affect crop growth and production. Heavy metals such as Fe, Mn, Zn, Cu, Mg, Mo, B, and Ni in the soil of a particular area significantly affect the morphological, metabolic, and physiological anomalies in plants. It ranges from chlorosis of shoot to lipid peroxidation and protein degradation [68]. Among them, 'B' is an essential element for plant growth and simultaneously affects the growth and yield at higher levels.

3.2.3. Degradation

Nutrient deficiency has been considered as the main cause of poor crop productivity. Of the global land area of 13.5 billion ha, among that only 3.03 billion ha (22%) is cultivable and about 2 billion ha is degraded. Disposal of oil shales, heavy metal contamination of soil, and spillage of crude oils adversely affect the severe root damage that governs nutrient irregularity [69].

3.3. Water Factor

3.3.1. Suboptimal

The most important constraints for agriculture are the water limitation, declining rainfall, and increasing temperature which significantly affects the growth of crop plant areas. A wide range of strategies such as genetical, physiological, biochemical, and molecular levels are well defined in plants. However, recent advancements are also available to obtain important drought-tolerant crops using conventional, marker-assisted breeding, and genetic engineering [70,71].

3.3.2. Supraoptimal Salinity

High salinity can be toxic for many crops, but halophytes have adapted to the worse salt conditions, which are found in several coastal areas, such as salt marshes, and inland arid areas. These zones, having a high rate of evaporation tends to concentrate salts in the mineral content of the soil. Those halophytes are morpho-physiologically and reproductively adapted to saline, waterlogged and anaerobic conditions. Nutrient deficiency or the presence of toxic substances such as heavy metals in the soil can also result in the AbS [49].

3.3.3. Waterlogging

Flood water usually causes a water-logged situation in the field. In waterlogged soils, within 24 h the oxygen concentration drops down to zero because water replaces most of the air in the soil pore space. Roots need oxygen for their respiration and cell viability. Water-logging limits oxygen supply to the roots, if any remaining oxygen is used up by

Plants 2021, 10, 1472 7 of 19

the roots from flooded or waterlogged soils, then the normal function of the roots will be arrested. Therefore, the leaves and stems are unable to obtain enough minerals and nutrients hence the roots start to die off in water-logging conditions [72].

4. Bioinformatics and Functional Omics Approach to Explore the AbS Tolerance Mechanism

The process of gene regulatory dynamism in various cellular, physiological and biochemical mechanisms in plants was altered by environmental factors. Therefore, to analyze the processes involved in this regulatory mechanism, several functional omics projects, have been initiated across the world in recent years [49,73,74]. Moreover, multiple omics and bioinformatics approaches have been used for developing crop plants that are tolerant to AbS through molecular breeding and genetic engineering and also through advancements and increasing knowledge in genetics, genomics, and molecular physiology [75,76]. Hence, the development of new functional omics and computational biology software and tools paved the way to identify the AbS responsible candidate genes from gene pools. In addition, the use of high-throughput techniques has been employed such as expression reads by RNA-Seq, random and targeted mutagenesis, gene shifting, complementation, and synthetic promoter trapping approaches make many avenues for functional analyses of AbS responsive genes and tolerance mechanisms [77]. Transcription factors (TFs) are crucially important in knowing the appropriate molecular processes and pathways that are involved in plant growth and survival under AbS conditions [78-80]. AbS are the quantitative and multiple genes associated in nature, these stress molecular cross-talks and their pathway interconnections are found under AbS conditions. Again, understanding the post-translational modifications (PTM) degradation of proteins and non-coding miRNA interactions allow the modulation of the target proteins. Similarly, some of the siRNAs play an important role as stress-inducers and affect protein synthesis including alternative splicing. The genome-wide association studies (GWAS) have become popular to provide novel strategies to identify and characterize the unique stress-responsive genes, which are introduced into crop plants and used in building up the tolerance against various AbS conditions [80]. In this record, the identification, and characterization of specific stressresponsive genes along with their promoters are analyzed for specificity. Integrated omics and bioinformatics approaches have been used to study the AbS responsible genes, their corresponding growth regulations and their encoding global metabolic network. With the advent of newly developed functional omics can be broadly categorized into two potential approaches used in manipulating gene-pool for enhancing the AbS tolerance, which contains: (i) identification of stress-responsive genes followed by genetic engineering to improve the stress tolerant cereals development and (ii) mining of marker associated with agronomically important genes and their use in marker-assisted breeding programs.

Importance of Omics in Enhancing Nutritive Values

In the era of post-genomics, revealing the interconnected functional attributes between genes transcripts, metabolites, proteins and nutritional biology remains a major challenge. New advances in omics such as genomics, transcriptomics, proteomics and metabolomics will have an essential advantage in the bio-fortification processes in plants.

5. Gene Mining

Functionally integrated omics and computational biology tools for AbS tolerance include reconnaissance of novel AbS responsive genes and the expression levels, that may induce the AbS response. These omics approaches are used to understand the functional role of AbS -responsive genes and the generation of stress-tolerant transgenic lines. In addition, they also pave the way for guiding genetic and metabolic engineering studies. In cereals, large numbers of expressed sequence tags (ESTs) have been generated from different cDNA libraries, collected from AbS treated tissues at different developmental stages, and are considered as a significant functional genomic approach to impute the AbS responsive genes. The expressed sequence tag database (dbEST) provides information about the number and type of different AbS tolerant species. Genomics-based approaches

Plants 2021, 10, 1472 8 of 19

were started in all the crop plants such as barley, rice, maize, wheat, and sorghum (stressgenomics.org). A total of 13,022 AbS related ESTs were reported from Hordeum vulgare, 13,058 genes from O. sativa, 17,189 from S. bicolor, 2641 from Secale cereale, 20,846 from T. aestivum and 5695 regulators from Z. mays using gene index of TIGR database (http: //www.tigr.org/tdb/tgi/ accessed on 16 April 2021) [81]. However, publicly available ESTs libraries with AbS data are few. For this reason, the sequence-specific approach that is based on cDNA libraries from cereal stress resistant genotypes may be improved in different developmental periods to encompass a larger range of tissue types and organs of many species. Functional characterizations of the different AbS treated genes are studied using BlastX [82] by comparing the Swissprot dataset (http://www.uniprot.org/accessed on 24 April 2021) [83]. Moreover, the EST clustering results provides consensus sequences which are more informative than typical EST data. Drought-responsive genes have a lot of features that include metallothionein-like proteins, late embryonic abundant (LEA) proteins, heat-shock proteins, cytochrome P450 enzymes, catalases, peroxidases, kinases, phosphatases, and TFs (DREB, MYB, MYC, AP2-EREBP, ZF-HD, NAC, WRKY, and bHLH protein) that were abundantly expressed during drought stress [78,84]. Recent research was carried out to identify ESTs in the NaCl-treated cDNA library of Thellungiella halophila and also from monocots like barley, wheat, maize, and rice [85]. Furthermore, novel studies are needed to be conducted on all food crops for future food security and production.

6. Transcript Signature Usage in Stress Responsive Gene Mining

Several computational approaches are employed to quantify the expression intensities of EST on its programs in cereal crops such as rice [86], barley [87], and maize [88]. The microarray technology is used for transcript expression profiling, Mission Planning and Scheduling System (MPSS), Serial Analysis of Gene Expression (SAGE), and quantitative real-time PCR (qRT-PCR) these are further modern approaches for the quantification under the controlled and stressed tissues of large number of gene expression studies [89]. Microarray-based transcript profiling was carried out in *Arabidopsis* [90] as well as in important cereal species [91,92] have comprehensively analyzed the gene expression signature in response to unique and multiple AbS. An EST-cDNA array technology provides a pivotal tool to compare the relative expression levels between normal and treated plants to unveil the functions of AbS responsive candidates. In addition to that, it is used to identify and understand the genes involved in transcriptional reprogramming and signal transduction pathways.

TFs, are the protein-containing domains that imply the DNA binding and transcript regulatory activities. The TF families were classified into more than 50, based on the presence of highly conserved motifs (Table 2). Computational omics studies were used to identify and understand the molecular cross-talks of plant developmental stages, expression profiling, and physicochemical properties [84,93]. The TFs are mainly involved in diverse stress tolerance and plant growth metabolisms such as cold, salinity, drought, metal, submergence, heat, low/high temperature, light, UV, O₃, osmotic, oxidative stress, and signaling, tissues development, regulation of plant physiological metabolisms respectively. It is the essential key factor to unravel the candidate gene functions and their dynamism.

Table 2. C3 and C4 crop species transcription factor families and their number of family members. (Source: https://grassius.org/grasstfdb.php/ accessed on 19 May 2021) [94] (http://planttfdb.cbi.pku.edu.cn/ accessed on 19 May 2021) [95]).

S. No	Transcription Factors (TFs)	Rice	Sorghum	Maize	Wheat
1	ABI3-VP1	55	60	51	-
2	BBR/BPC	04	05	04	05
3	C2C2-GATA	23	27	36	-
4	CCAAT-HAP2	11	09	16	-
5	G2-like	44	41	59	100
6	HSF	25	24	29	53

Plants **2021**, 10, 1472 9 of 19

 Table 2. Cont.

S. No 7	Transcription Factors (TFs) Orphans	Rice 185	Sorghum 177	Maize 339	Wheat
8	WHIRLY	02	03	02	07
9	Alfin-like	09	13	19	-
10	bHLH	135	143	175	-
11	C2C2-YABBY	08	08	13	-
12	CCAAT-HAP3	12	-	04	-
13	GeBP	13	15	21	12
14	MADS	70	76	77	-
15	SBP	19	18	32	37
16 17	WRKY AP2-EREBP	103	94 156	125 212	171 -
18	bZIP	164 92	91	128	186
19	C2H2	09	07	10	224
20	CCAAT-HAP5	22	-	18	-
21	GRAS	60	76	86	121
22	MYB	114	113	167	263
23	TCP	22	28	44	28
24	ZF-HD	15	15	21	20
25	ARF	27	27	38	45
26	BZR	06	08	10	-
27	СЗН	46	44	54	100
28	CPP	11	08	13	24
29	GRF	12	10	15	16
30	MYB-related	71	80	116	227
31	Trihelix	17	19	43	46
32	ZIM	21	19	36	-
33	ARID	06	07	10	-
34	C2C2-CO-like	08	09	14	-
35	CAMTA	06	06	05	20
36	E2F-DP	09	10	19	24
37 38	Homeobox	95 144	83	133	-
36 39	NAC TUB	144 15	123 13	134 15	263
39 40	ARR-B	06	10	08	22
41	C2C2-Dof	30	29	46	- -
42	CCAAT-DR1	01	-	17	- -
43	EIL	09	07	09	16
44	HRT	01	01	-	-
45	NLP	13	13	17	-
46	VOZ	02	02	05	06
47	CCAAT	-	32	-	-
48	mTERF	-	-	30	-
49	OVATE	-	-	43	-
50	Sigma70-like	-	-	09	-
51	PLATZ	-	-	15	-
52	FAR1-like	-	-	15	-
53	Rcd1-like	-	-	10	-
54	FLO/ LFY	-	-	02	-
55	S1Fa-like	-	-	02	03
56 57	CSD	-	-	04	-
57 59	LBD	-	-	44	61
58 59	DBP SHI/STY (SRS)	-	_	04 09	-
60	AP2	_	-	U9 -	43
60 61	BES1	-	-	-	43 10
62	ERF	<u>-</u>	-	-	181
63	HRT-like	-	-	-	03
64	M-type-MADS	- -	- -	-	77
65	NF-X1	_	_	-	02
66	RAV	_	_	_	08
	1411				

Plants **2021**, 10, 1472 10 of 19

Table 2. Cont.

S. No	Transcription Factors (TFs)	Rice	Sorghum	Maize	Wheat
67	TALE	-	-	-	52
68	DBB	-	-	-	17
69	FAR1	-	-	-	201
70	MIKC_MADS	_	-	-	51
71	NF-YA	_	-	-	22
72	Dof	_	-	-	52
73	HB-PHD	_	-	-	06
74	NF-YB	_	-	-	34
75	YABBY	_	-	-	25
76	В3	_	-	-	140
77	GATA	_	-	-	48
78	HB-other	_	-	-	44
79	LFY	_	-	-	02
80	NF-YC	_	-	-	20
81	SRS	_	-	-	05
82	CO-like	_	-	-	07
83	HD-ZIP	_	-	-	62
84	LSD	_	-	-	13
85	Nin-like	_	-	-	29
86	STAT	_	-	-	02
87	WOX	_	_	-	26

^{&#}x27;-' indicates that TF family was absent in particular crop.

Diverse literary information revealed the importance of plant stress mechanisms analyzed by gene expression signature. Two kinds of pathways involved in this type of gene expression studies are (i) desiccation tolerance in an ABA-dependent manner by ABA-responsive element binding factors (ABF), MYC and MYB TFs, and (ii) ABA-independent and associated with drought-responsive element binding factors (DREB) [96]. Recent studies are providing evidence, that other than the ABA-associated pathways, there exists an interlinked relationship between cold and salinity stress signaling pathways.

In cereal crop species, microarray and RNA-Seq based stress-regulated transcripts were used for large spectrum gene expression signature analysis in rice, up-regulation of few candidates that encodes cell division, 40S ribosomal proteins, glycine-rich proteins, elongation factor, and induce the phytohormone regulating genes. The genes which are generally downregulated in the sensitive rice cultivators are glycine-rich proteins, ABA and stress-induced proteins, metallothionein-like proteins, glutathione S-transferase, ascorbate peroxidase, water channel protein isoforms, subtilisin inhibitor, tyrosine inhibitor, and so on [97,98].

During long-term AbS treatments, protease inhibitors, stress proteins, aquaporins, antioxidant components, and some unknown genes were induced and are expected to impart tolerance. A few transcriptional expression signature analyses were conducted in important cereal crops. In *Arabidopsis*, the multiple stress interactions were studied using functional genomics approaches [10,85]. Multiple stress interactions of AbS treatments were investigated to identify the key players having a pivotal role in multiple individual stresses such as drought, cold, flood, salt, UV, and temperature responses [99]. Using 1300 full-length clones, only 44 genes were identified, which are directly induced either by drought or cold stress dynamisms were reported [100]. By using 7000 whole clone inserts, 213 salinity responsible genes, 299 drought responsive genes, 245 ABA-regulating key genes, and 54 cold-regulating genes were identified [101]. Functional omics and bioinformatics tools to identify gene expression patterns related to multiple stress interactions have been considered as a significant aspect of modern plant stress biology research.

To study gene interaction and downstream elements, the analysis and characterization of the transcriptional responses in knock-out mutants or transgenic plants under abiotic stress tolerance are considered a differential method. Three members of the CBF/DREB1 family, CBF1, CBF2, and CBF3 (DREB1b, DREB1c, and DREB1a respectively) quantification

Plants **2021**, 10, 1472

of 41 downstream genes as CBF targets were identified by Fowler and Thomashow, [102]. Comparative genome analysis of the AbS responses among diverse tolerant species is extensively considered as an important approach to reconnaissance of evolutionarily conserved and unique stress defense mechanisms [103]. Various promoters of a group of the abiotic stress-responsive genes from different species of maize, rice, and *Arabidopsis* harboring DRE, GCC-box, ABF, and w-box *cis*-elements were reported [104]. Computational omics gene mining and profiling led to the novel way to understand the huge number of genes involved in AbS responses given in Table 3.

Table 3. Genes encoding enzymes/proteins associated with abiotic stress response in cereals.

Gene Category	Gene	Cellular Response	Species	Reference
Osmolyte compounds				
Glycine betaine	BADH	Heave metal stress	Rice	[105]
,	CodA	Salt, Cold and drought stress	Rice	[106]
	bet A	Cold, Drought stress	Maize	[107]
Proline	P5CS	Drought	Wheat	[108]
Regulatory genes		O .		
bZIP	bZIP4	Salinity stress	Maize	[109]
	HBP1b	Drought, Salt, cold	Rice	[110]
	bZIP16	Dehydration, salt and ABA	Rice	[111]
Transporters				
Na+-H+-dependent K+	ZmHKT1	Salt stress	Maize	[112]
transporter				
Na+-K+-symporter	HKT1	Salt stress	Wheat	[113]
	HKT1	Salt stress	Rice	[114]
Stress-responsive genes				
Transcription factors	SAP7	Abiotic stress	Rice	[115]
	DREB	Abiotic stress	Maize	[116]
	MYB6	Drought and Salt	Rice	[117]
WCS genes	WCS19	Cold stress	Wheat	[118]
	WCS120	Cold stress	Wheat	[118]
Thaumatin-like protein	TLP14	Cold stress	Barley	[119]
Heat shock protein	HSFA7	Salt and Droght	Rice	[120]
	HSP20	Heat stress	Wheat	[121]
RAB genes	RAB7	Drought and Heat stress	Rice	[122]
	RAB11	Salt stress	Rice	[123]
LEA proteins	HVA1	Drought stress	Barley	[124]
	HVA1	Salt, Cold and dehydration stress	Rice	[125]
	HVA1	Salt and Drought tolerance	Barley	[126]
Antioxidants				
Ascorbate peroxidase	APX	Drought, Salt and Cold	Rice	[127]
Catalase	CAT	Drought stress	Wheat	[128]
Superoxide dismutase	MnSOD	Abiotic stress	Rice	[129]
	ZnSOD	Salt stress	Rice	[130]
	FeSOD	Drought stress	Rice	[131]

7. Identification of Genes and Their Agronomic Traits

A case study has been carried out on the evaluation of seven well-known candidate genes for their effects on improving drought resistance of transgenic rice under field conditions [132]. Several AbS affects the growth and yield of cereal crops such as drought, extremely high temperature, low water availability, mineral deficiencies or toxicities, and salinity [133,134]. In recent years, the initiative of developing drought-resistant cereal crops has been well recognized as the most promising and effective strategy for food security against drought and water deficit. In addition, over-expression of certain stress-responsive genes or TFs regulating the multiple stress proteins were shown to confer the increased tolerance to drought as well as in salinity and freezing stresses [117]. Poor management of agricultural water resources, soil degradation, and community pressures are all the

Plants **2021**, 10, 1472

prominent stressors that play a pivotal role in the agriculture perspectives across the world. Extensive genetic studies have indicated a huge variation for AbS tolerance but it has failed to attain its maximum goal due to the relatively poor knowledge from the molecular basis for stress-tolerant cereal crop plants. In this case, functional omics and bioinformatics play a crucial role by providing several tools for dissecting AbS responses in cereal crops especially in rice, barley, and wheat through interrogation of genes that may be useful for improving resistance to AbS.

Xiao et al. [132] have selected seven genes (*CBF3*, *SOS2*, *NCED2*, *NPK1*, *LOS5*, *ZAT10*, and *NHX1*) in drought resistance breeding and transformed them into rice cultivar Zhonghua 11 (*O. sativa* L. ssp. *Japonica*) under the control of a constitutive promoter (from rice *Actin1* gene) and an inducible promoter (from a rice homologous gene of *HVA22*), respectively. The developed transgenic rice was responsible for drought resistance under natural field conditions. These traits (Table 4) are an example that can be a useful reference for drought resistance engineering in cereal crops and also pave the way to address the other stress-related issues.

Table 4	. Transgenio	crop plants	developed	for AbS	tolerance	details.

Gene	Cellular Response	Species	Reference
Osmolyte compounds			
Pyrroline carboxylate synthase (p5cs)	Drought tolerance	Wheat	[108]
Choline dehydrogenase	Drought, Salt tolerance	Rice	[135]
Arginine decarboxylase	Drought, Heat, Freezing, Salinity tolerance	Rice	[136]
Glutamine synthetase	Oxidative stress tolerance	Rice	[137]
Trehalose-6-P-synthase	Salt, Drought, Cold tolerance	Rice	[138]
Mannitol dehydrogenase	Drought, Salt tolerance	Wheat	[139]
Regulatory genes			
Calcium dependent protein kinase	Drought tolerance	Rice	[140]
DREB1A	Drought tolerance	Rice	[141]
Transporters/symporter	O		
Na+/H+ antiporter	Salt tolerance	Rice	[142]
Potassium transporter (HKT1)	Salt tolerance	Rice	[143]
Stress-responsive genes			
HVA1	Drought, Salt tolerance	Rice	[125]
Alcohol dehydrogenase	Submergence tolerance	Rice	[144]
Ferritin	Heat tolerance	Wheat	[145]
HVA1	Salt and Drought tolerance	Barley	[126]
Pyruvate decarboxylase1	Submergence tolerance	Rice	[146]
Antioxidants			
Fe-superoxide dismutase	Drought tolerance	Rice	[147]
Mn-superoxide dismutase	Salt tolerance	Rice	[148]

8. Genome-Wide Association Studies (GWAS)

By the use of an ultra- high throughput genotyping technology, GWAS became available as a powerful alternative for dissecting quantitative traits in crop plants [149]. GWAS provides an understanding of transcriptional regulation, metabolic response of rice and other cereal crops to diverse environmental conditions and also in significant relation to AbS [150]. Genome-wide association mapping identifies major QTL regions without the need of constructing a mapping population.

Currently, GWAS was categorized into two types, population-based association studies and a family-based approach [151]. In population-based association studies, it was revealed that unrelated individuals are used to examine genome-wide associations between single nucleotide polymorphisms (SNPs) and their associated phenotypic traits. Family-based mapping studies can be applied to complex pedigrees derived from the crosses among different genotypes. Both approaches have complementary pros and cons.

Plants 2021, 10, 1472 13 of 19

Population-based GWAS takes advantage of more recombination events that have accumulated over time of generations in historical populations, with the disadvantage of finding false positives or false negatives results applied in *Arabidopsis* [152]. Family-based GWAS can eliminate the effects of population structure and therefore escape the false-positives and false-negatives, but recombination accumulated over a few generations during pedigree development used in *Z. mays* NAM (nested association mapping) population was developed to characterize flowering regulation [151]. The maize NAM population of 200 recombinant inbred lines from 25 parents were crossed to the fully sequenced genotype (B73). Therefore, GWAS is an alternative and complementary approach for understanding the trait and molecular level mechanisms in plants. This approach paves the un-paralleled way for the history of plant stress and molecular biology.

9. Conclusions and Future Perspectives

Ever-increasing advances in multiple analytical stages are paving the way to address the diverse omics scale outcomes, fortifying the researcher's knowledge and long-standing questions in plant biology. Plants are acclimatizing to the environment by altering their genome, metabolome, transcriptome, lipidome, proteome, secretome and miRNAome. Even in this post-genome era, the complete understanding of plant molecular responses to the stress tolerance mechanism is not yet achieved, as the list of genes involved in stress response is increasing rapidly. The identity of these proteins and their functions are close to being determined. Post-transcriptional studies including splicing and RNA silencing and posttranslational mechanisms such as SUMOylation, phosphorylation, and ubiquitination lead to a prompt response in plants against stress. Thus, employing the developing omics approaches and GWAS will contribute to better understanding the AbS response.

At the same time, we are majorly facing multiple global issues such as climate changes, global warming, water, food, and energy security. Further, voluminous research is highly needed to unravel the specific molecular function of the plants. AbS tolerance crosstalks that are speculating in particular plant species, significantly in C_3 and C_4 grass species, are also essential in identifying the candidates to improve the diverse molecular and biochemical functions. In addition, these molecular studies are also subjected and compared to advanced omics datasets from C_3 and C_4 genome species, which are helping to improve the applied or translational research, to unveil the plant molecular systems in response to stress biologist, molecular biologist, and plant physiologist, and also the ever-increasing command of mankind. This eagle's eye review can open the penstocks to budding scientists.

Author Contributions: Conceptualization, R.J., P.M. and M.R.; Methodology, R.J., P.M., and L.S.; Investigation, R.J., P.M., L.S., S.A. and X.W., Formal analysis, R.J. and P.M.; Project administration, R.J. and P.M.; Resources, R.J., P.M., L.S., F.M.-P. and M.R.; Validation, L.S., S.K.P., J.-T.C., S.A. and X.W.; Writing—Original draft, R.J. and P.M.; Writing—review & editing, S.K.P., J.-T.C., F.M.-P. and M.R. All authors have read and agreed to the published version of the manuscript.

Funding: The APC was supported by the Chilean National Fund for Scientific and Technological Development (FONDECYT) grant number 1201973.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable. **Data Availability Statement:** Not applicable.

Acknowledgments: The authors gratefully acknowledge the use of Bioinformatics Infrastructure Facility, Alagappa University funded by Department of Biotechnology, Ministry of Science and technology, Government of India grant (No.BT/BI/25/015/2012). The authors also thankfully acknowledge DST-FIST (Grant No. SR/FST/LSI-639/2015(C)), UGC-SAP (Grant No.E.5-1/2018/DRS-II (SAP-II)) and DST-PURSE (Grant No. SR/PURSE Phase 2/38 (G)) for providing lab facilities. The authors also thank RUSA 2.0 [F. 24-51/2014-U, Policy (TN Multi-Gen), Dept of Edn, GoI].

Conflicts of Interest: Authors have no conflict of interest.

Plants **2021**, 10, 1472 14 of 19

References

1. Giordano, M.; Petropoulos, S.; Rouphael, Y. Response and Defence Mechanisms of Vegetable Crops against Drought, Heat and Salinity Stress. *Agriculture* **2021**, *11*, 463. [CrossRef]

- 2. Kumari, V.V.; Roy, A.; Vijayan, R.; Banerjee, P.; Verma, V.C.; Nalia, A.; Pramanik, M.; Mukherjee, B.; Ghosh, A.; Reja, H.; et al. Drought and Heat Stress in Cool-Season Food Legumes in Sub-Tropical Regions: Consequences, Adaptation, and Mitigation Strategies. *Plants* **2021**, *10*, 1038. [CrossRef]
- 3. O'Neil, C.E.; Nicklas, T.A.; Zanovec, M.; Cho, S. Whole-Grain Consumption Is Associated with Diet Quality and Nutrient Intake in Adults: The National Health and Nutrition Examination Survey, 1999–2004. *J. Am. Diet. Assoc.* 2010, 110, 1461–1468. [CrossRef] [PubMed]
- Papanikolaou, Y.; Fulgoni, V.L. Certain grain foods can be meaningful contributors to nutrient density in the diets of US chil-dren and adolescents: Data from the National Health and Nutrition Examination Survey, 2009–2012. Nutrients 2017, 9, 160. [CrossRef]
- 5. Olugbire, O.O.; Olorunfemi, S.; Oke, D.O. Global utilisation of cereals: Sustainability and environmental issues. *Agro-Science* **2021**, 20, 9–14. [CrossRef]
- 6. Atkinson, N.J.; Urwin, P.E. The interaction of plant biotic and abiotic stresses: From genes to the field. *J. Exp. Bot.* **2012**, *63*, 3523–3543. [CrossRef] [PubMed]
- 7. Maiti, R.; Satya, P. Research advances in major cereal crops for adaptation to abiotic stresses. *GM Crop. Food* **2014**, *5*, 259–279. [CrossRef] [PubMed]
- 8. Halford, N.G.; Curtis, T.Y.; Chen, Z.; Huang, J. Effects of abiotic stress and crop management on cereal grain composition: Implications for food quality and safety. *J. Exp. Bot.* **2014**, *66*, 1145–1156. [CrossRef]
- 9. Takeda, S.; Matsuoka, M. Genetic approaches to crop improvement: Responding to environmental and population changes. *Nat. Rev. Genet.* **2008**, *9*, 444–457. [CrossRef]
- 10. Newton, A.C.; Johnson, S.N.; Gregory, P.J. Implications of climate change for diseases, crop yields and food security. *Euphytica* **2011**, *179*, 3–18. [CrossRef]
- 11. Mohan, V.; Tresina, P.; Daffodil, E. Antinutritional Factors in Legume Seeds: Characteristics and Determination. *Encycl. Food Health* **2016**, 211–220. [CrossRef]
- 12. Piltz, J.W.; Rodham, C.A.; Wilkins, J.F.; Hackney, B.F. A Comparison of Cereal and Cereal/Vetch Crops for Fodder Conser-vation. *Agriculture* **2021**, *11*, 459. [CrossRef]
- 13. Chaudhari, P.R.; Tamrakar, N.; Singh, L.; Tandon, A.; Sharma, D. Rice nutritional and medicinal properties. *J. Pharmacogn. Phytochem.* **2018**, *7*, 150–156.
- 14. Muraki, I.; Wu, H.; Imamura, F.; Laden, F.; Rimm, E.B.; Hu, F.B.; Willett, W.C.; Sun, Q. Rice consumption and risk of cardio-vascular disease: Results from a pooled analysis of 3 US cohorts. *Am. J. Clin. Nutr.* **2015**, *101*, 164–172. [CrossRef] [PubMed]
- 15. Tan, B.L.; Norhaizan, M.E. Scientific evidence of rice by-products for cancer prevention: Chemopreventive properties of waste products from rice milling on carcinogenesis in vitro and in vivo. *Biomed. Res. Int.* **2017**, 2017, 9017902. [CrossRef]
- 16. Okuda, M.; Fujita, Y.; Katsube, T.; Tabata, H.; Yoshino, K.; Hashimoto, M.; Sugimoto, H. Highly water pressurized brown rice improves cognitive dysfunction in senescence-accelerated mouse prone 8 and reduces amyloid beta in the brain. *BMC Complement. Altern. Med.* **2018**, *18*, 110. [CrossRef] [PubMed]
- 17. Almeida, D.M.; Almadanim, M.C.; Lourenço, T.; Abreu, I.A.; Saibo, N.J.M.; Oliveira, M.M. Screening for Abiotic Stress Tolerance in Rice: Salt, Cold, and Drought. In *Environmental Responses in Plants*; Humana Press: New York, NY, USA, 2016; pp. 155–182.
- 18. Hasanuzzaman, M.; Nahar, K.; Fujita, M.; Ahmad, P.; Chandna, R.; Prasad, M.N.V.; Ozturk, M. Enhancing plant productivity under salt stress: Relevance of poly-omics. In *Salt Stress in Plants*; Springer: New York, NY, USA, 2013; pp. 113–156.
- 19. Lee, K.S.; Choi, W.Y.; Ko, J.C.; Kim, T.S.; Gregoria, G.B. Salinity tolerance of japonica and indica rice (*Oryza sativa* L.) at the seedling stage. *Planta* 2003, 216, 1043–1046. [CrossRef]
- 20. Todaka, D.; Nakashima, K.; Shinozaki, K.; Yamaguchi-Shinozaki, K. Towards understanding transcriptional regulatory net-works in abiotic stress responses and tolerance in rice. *Rice* **2012**, *5*, 1–9. [CrossRef] [PubMed]
- 21. Das, P.; Nutan, K.K.; Singla-Pareek, S.N.; Pareek, A. Understanding salinity responses and adopting 'omics-based' approaches to generate salinity tolerant cultivars of rice. *Front. Plant Sci.* **2015**, *6*, 712. [CrossRef]
- 22. Alam, M.; Bell, R.W.; Hasanuzzaman, M.; Salahin, N.; Rashid, M.H.; Akter, N.; Akhter, S.; Islam, M.S.; Islam, S.; Naznin, S.; et al. Rice (*Oryza sativa* L.) establishment techniques and their implications for soil properties, global warming potential mitigation and crop yields. *Agronomy* **2020**, *10*, 888. [CrossRef]
- 23. Macauley, H.; Ramadjita, T. Cereal crops: Rice, maize, millet, sorghum, wheat. Feed. Afr. 2015, 36.
- 24. Sandhu, K.S.; Singh, N.; Malhi, N.S. Some properties of corn grains and their flours I: Physicochemical, functional and chapatimaking properties of flours. *Food Chem.* **2007**, *101*, 938–946. [CrossRef]
- 25. Shah, T.R.; Prasad, K.; Kumar, P. Maize—A potential source of human nutrition and health: A review. *Cogent Food Agric.* **2016**, 2, 1166995.
- 26. Liu, R.H. Potential synergy of phytochemicals in cancer prevention: Mechanism of action. J. Nutr. 2004, 134, 3479–3485. [CrossRef]
- 27. Díaz-Gómez, J.L.; Castorena-Torres, F.; Preciado-Ortiz, R.E.; García-Lara, S. Anti-cancer activity of maize bioactive peptides. *Front. Chem.* **2017**, *5*, 44. [CrossRef]
- 28. Iqbal, S.; Hussain, S.; Qayyaum, M.A.; Ashraf, M. The Response of Maize Physiology under Salinity Stress and Its Coping Strategies. In *Plant Stress Physiology*; IntechOpen: London, UK, 2020.

Plants 2021, 10, 1472 15 of 19

29. Sade, N.; del Mar Rubio-Wilhelmi, M.; Umnajkitikorn, K.; Blumwald, E. Stress-induced senescence and plant tolerance to abiotic stress. *J. Exp. Bot.* **2018**, *69*, 845–853. [CrossRef] [PubMed]

- 30. Shewry, P.R.; Hey, S.J. The contribution of wheat to human diet and health. Food Energy Secur. 2015, 4, 178–202. [CrossRef]
- 31. Hura, T. Wheat and Barley: Acclimatization to Abiotic and Biotic Stress. Int. J. Mol. Sci. 2020, 21, 7423. [CrossRef]
- 32. Luthria, D.L.; Lu, Y.; John, K.M. Bioactive phytochemicals in wheat: Extraction, analysis, processing, and functional properties. *J. Funct. Foods* **2015**, *18*, 910–925. [CrossRef]
- 33. Barros, L.; Fernandes, Â.; C.F.R. Ferreira, I.; Callejo, M.; Matallana-González, M.; Fernández-Ruiz, V.; Morales, P.; Carrillo, J.M. Potential health claims of durum and bread wheat flours as functional ingredients. *Nutrients* **2020**, 12, 504.
- 34. Aune, D.; Keum, N.; Giovannucci, E.; Fadnes, L.T.; Boffetta, P.; Greenwood, D.C.; Tonstad, S.; Vatten, L.J.; Riboli, E.; Norat, T. Whole grain consumption and risk of cardiovascular disease, cancer, and all cause and cause specific mortality: Systematic review and dose-response meta-analysis of prospective studies. *BMJ* 2016, 353, i2716. [CrossRef] [PubMed]
- 35. Della Pepa, G.; Vetrani, C.; Vitale, M.; Riccardi, G. Wholegrain intake and risk of type 2 diabetes: Evidence from epidemiological and intervention studies. *Nutrients* **2018**, *10*, 1288. [CrossRef] [PubMed]
- 36. Bedő, Z.; Láng, L.; Rakszegi, M. Breeding for grain-quality traits. In *Cereal Grains*; Woodhead Publishing: Cambridge, UK, 2017; pp. 425–452.
- 37. Loskutov, I.G.; Khlestkina, E.K. Wheat, Barley, and Oat Breeding for Health Benefit Components in Grain. *Plants* **2021**, *10*, 86. [CrossRef] [PubMed]
- 38. Schapendonk, A.H.C.M.; Xu, H.Y.; Van Der Putten, P.E.L.; Spiertz, J.H.J. Heat-shock effects on photosynthesis and sink-source dynamics in wheat (*Triticum aestivum* L.). *NJAS-Wagening*. *J. Life Sci.* **2007**, *55*, 37–54. [CrossRef]
- Srivastava, N. Biochemical and molecular responses in higher plants under salt stress. In Plant Adaptation Strategies in Changing Environment; Springer: Singapore, 2017; pp. 117–151.
- Shah, T.; Xu, J.; Zou, X.; Cheng, Y.; Nasir, M.; Zhang, X. Omics approaches for engineering wheat production under abiotic stresses. Int. J. Mol. Sci. 2018, 19, 2390. [CrossRef]
- 41. Calone, R.; Sanoubar, R.; Lambertini, C.; Speranza, M.; Antisari, L.V.; Vianello, G.; Barbanti, L. Salt tolerance and Na allocation in Sorghum bicolor under variable soil and water salinity. *Plants* **2020**, *9*, 561. [CrossRef] [PubMed]
- 42. Ramatoulaye, F.; Mady, C.; Fallou, S. Production and use sorghum: A literature review. J. Nutr. Health Food Sci. 2016, 4, 1-4.
- 43. Rao, S.; Santhakumar, A.B.; Chinkwo, K.A.; Wu, G.; Johnson, S.K.; Blanchard, C.L. Characterization of phenolic compounds and antioxidant activity in sorghum grains. *J. Cereal Sci.* **2008**, *84*, 103–111. [CrossRef]
- 44. Xiong, Y.; Zhang, P.; Warner, R.D.; Fang, Z. Sorghum grain: From genotype, nutrition, and phenolic profile to its health benefits and food applications. *Compr. Rev. Food Sci. Food Saf.* **2019**, *18*, 2025–2046. [CrossRef] [PubMed]
- 45. Ba, K.; Tine, E.; Destain, J.; Cisse, N.; Thonart, P. Comparative study of phenolic compounds, the antioxidant power of various Senegalese sorghum varieties and aamylolytic enzymes of their malt. *Biotechnol. Agron. Société Environ.* **2010**, *14*, 131–139.
- 46. Djanaguiraman, M.; Prasad, P.V.; Ciampitti, I.A.; Talwar, H.S. Impacts of Abiotic Stresses on Sorghum Physiology. In *Sorghum in the 21st Century: Food–Fodder–Feed–Fuel for a Rapidly Changing World*; Springer: Singapore, 2020; pp. 157–188.
- 47. Abdel-Ghany, S.E.; Ullah, F.; Ben-Hur, A.; Reddy, A.S. Transcriptome analysis of drought-resistant and drought-sensitive sorghum (Sorghum bicolor) genotypes in response to peg-induced drought stress. *Int. J. Mol. Sci.* **2020**, *21*, 772. [CrossRef]
- 48. Muthuramalingam, P.; Jeyasri, R.; Kalaiyarasi, D.; Pandian, S.; Krishnan, S.R.; Satish, L.; Pandian, S.K.; Ramesh, M. Emerging advances in computational omics tools for systems analysis of gramineae family grass species and their abiotic stress responsive functions. *OMICS-Based Approach Plant Biotechnol.* **2019**, *185*, 185.
- 49. Sahu, M.; Dehury, B.; Modi, M.K.; Barooah, M. Functional Genomics and Bioinformatics Approach to Understand Regulation of Abiotic Stress in Cereal Crops. In *Crop Improvement in the Era of Climate Change*; I.K. International Publishing House Pvt. Ltd.: Delhi, India, 2014; p. 205.
- 50. Gong, Y.; Hao, Y.; Li, J.; Li, H.; Shen, Z.; Wang, W.; Wang, S. The effects of rainfall runoff pollutants on plant physiology in a bioretention system based on pilot experiments. *Sustainability* **2019**, *11*, 6402. [CrossRef]
- 51. Tomás, D.; Rodrigues, J.C.; Viegas, W.; Silva, M. Assessment of high temperature effects on grain yield and composition in bread wheat commercial varieties. *Agronomy* **2020**, *10*, 499. [CrossRef]
- 52. Kai, H.; Iba, K. Temperature stress in plants. In eLS; John Wiley & Sons, Ltd: Chichester, UK, 2014.
- 53. Beck, E.H.; Heim, R.; Hansen, J. Plant resistance to cold stress: Mechanisms and environmental signals triggering frost hardening and dehardening. *J. Biosci.* **2004**, *29*, 449–459. [CrossRef]
- 54. Baek, K.H.; Skinner, D.Z. Production of reactive oxygen species by freezing stress and the protective roles of antioxidant enzymes in plants. *J. Agric. Chem. Environ.* **2012**, *1*, 34–40. [CrossRef]
- 55. Cassia, R.; Nocioni, M.; Correa-Aragunde, N.; Lamattina, L. Climate change and the impact of greenhouse gasses: CO2 and NO, friends and foes of plant oxidative stress. *Front. Plant Sci.* **2018**, *9*, 273. [CrossRef] [PubMed]
- 56. Foroughbakhch Pournavab, R.; Bacópulos Mejía, E.; Benavides Mendoza, A.; Salas Cruz, L.R.; Ngangyo Heya, M. Ultraviolet radiation effect on seed germination and seedling growth of common species from Northeastern Mexico. *Agronomy* **2019**, *9*, 269. [CrossRef]
- 57. Metwally, S.A.; Shoaib, R.M.; Hashish, K.I.; El-Tayeb, T.A. In vitro ultraviolet radiation effects on growth, chemical constituents and molecular aspects of Spathiphyllum plant. *Bull. Natl. Res. Cent.* **2019**, *43*, 94. [CrossRef]

Plants 2021, 10, 1472 16 of 19

58. Rao, M.P.N.; Dong, Z.Y.; Xiao, M.; Li, W.J. Effect of salt stress on plants and role of microbes in promoting plant growth under salt stress. In *Microorganisms in Saline Environments: Strategies and Functions*; Springer: Cham, Switzerland, 2019; pp. 423–435.

- 59. Borsani, O.; Dıaz, P.; Agius, M.F.; Valpuesta, V.; Monza, J. Water stress generates an oxidative stress through the induction of a specific Cu/Zn superoxide dismutase in Lotus corniculatus leaves. *Plant Sci.* **2001**, *161*, 757–763. [CrossRef]
- 60. Tarakcioglu, C.; Inal, A. Changes induced by salinity, demarcating specific ion ratio (Na/Cl) and osmolality in ion and proline accumulation, nitrate reductase activity, and growth performance of lettuce. *J. Plant Nutr.* **2002**, 25, 27–41. [CrossRef]
- 61. Eraslan, F.; Inal, A.; Gunes, A.; Alpaslan, M. Impact of exogenous salicylic acid on the growth, antioxidant activity and physiology of carrot plants subjected to combined salinity and boron toxicity. *Sci. Hortic.* **2007**, *113*, 120–128. [CrossRef]
- 62. Ahmad, P.; Prasad, M.N.V. *Abiotic Stress Responses in Plants: Metabolism, Productivity and Sustainability*; Springer Science & Business Media: New York, USA, 2011.
- 63. Yildiz, M.; Poyraz, İ.; Çavdar, A.; Özgen, Y.; Beyaz, R. Plant Responses to Salt Stress. In *Plant Breeding-Current and Future Views*; IntechOpen: London, UK, 2020.
- 64. Shelden, M.C.; Roessner, U. Advances in functional genomics for investigating salinity stress tolerance mechanisms in cereals. *Front. Plant Sci.* **2013**, *4*, 123. [CrossRef]
- 65. Rajendran, K.; Tester, M.; Roy, S.J. Quantifying the three main components of salinity tolerance in cereals. *Plant Cell Environ.* **2009**, 32, 237–249. [CrossRef] [PubMed]
- 66. Roy, S.J.; Negrão, S.; Tester, M. Salt resistant crop plants. Curr. Opin. Biotechnol. 2014, 26, 115-124. [CrossRef]
- 67. Reddy, I.N.B.L.; Kim, B.K.; Yoon, I.S.; Kim, K.H.; Kwon, T.R. Salt tolerance in rice: Focus on mechanisms and approaches. *Rice Sci.* **2017**, *24*, 123–144. [CrossRef]
- 68. Emamverdian, A.; Ding, Y.; Mokhberdoran, F.; Xie, Y. Heavy metal stress and some mechanisms of plant defense response. *Sci. World J.* 2015, 2015, 75612. [CrossRef] [PubMed]
- 69. Shah, F.; Wu, W. Soil and crop management strategies to ensure higher crop productivity within sustainable environments. *Sustainability* **2019**, *11*, 1485. [CrossRef]
- 70. Oladosu, Y.; Rafii, M.Y.; Samuel, C.; Fatai, A.; Magaji, U.; Kareem, I.; Kamarudin, Z.S.; Muhammad, I.I.; Kolapo, K. Drought resistance in rice from conventional to molecular breeding: A review. *Int. J. Mol. Sci.* **2019**, 20, 3519. [CrossRef]
- 71. Rosero, A.; Berdugo-Cely, J.A.; Šamajová, O.; Šamaj, J.; Cerkal, R. A Dual Strategy of Breeding for Drought Tolerance and Introducing Drought-Tolerant, Underutilized Crops into Production Systems to Enhance Their Resilience to Water Deficiency. *Plants* 2020, *9*, 1263. [CrossRef]
- 72. Liliane, T.N.; Charles, M.S. Factors Affecting Yield of Crops. In *Agronomy-Climate Change & Food Security*; IntechOpen: London, UK, 2020; p. 9.
- 73. Chaudhary, J.; Khatri, P.; Singla, P.; Kumawat, S.; Kumari, A.; Vikram, A.; Jindal, S.K.; Kardile, H.; Kumar, R.; Sonah, H.; et al. Advances in omics approaches for abiotic stress tolerance in tomato. *Biology* **2019**, *8*, 90. [CrossRef] [PubMed]
- 74. Razzaq, M.K.; Aleem, M.; Mansoor, S.; Khan, M.A.; Rauf, S.; Iqbal, S.; Siddique, K.H. Omics and CRISPR-Cas9 Approaches for Molecular Insight, Functional Gene Analysis, and Stress Tolerance Development in Crops. *Int. J. Mol. Sci.* **2021**, 22, 1292. [CrossRef] [PubMed]
- 75. Ismail, A.M.; Horie, T. Genomics, physiology, and molecular breeding approaches for improving salt tolerance. *Annu. Rev. Plant Biol.* **2017**, *68*, 405–434. [CrossRef] [PubMed]
- 76. Tiwari, J.K.; Plett, D.; Garnett, T.; Chakrabarti, S.K.; Singh, R.K. Integrated genomics, physiology and breeding approaches for improving nitrogen use efficiency in potato: Translating knowledge from other crops. *Funct. Plant Biol.* **2018**, 45, 587–605. [CrossRef]
- 77. Chantre Nongpiur, R.; Lata Singla-Pareek, S.; Pareek, A. Genomics approaches for improving salinity stress tolerance in crop plants. *Curr. Genom.* **2016**, *17*, 343–357. [CrossRef] [PubMed]
- 78. Muthuramalingam, P.; Krishnan, S.R.; Saravanan, K.; Mareeswaran, N.; Kumar, R.; Ramesh, M. Genome-wide identification of major transcription factor superfamilies in rice identifies key candidates involved in abiotic stress dynamism. *J. Plant Biochem. Biotechnol.* **2018**, 27, 300–317. [CrossRef]
- 79. Muthuramalingam, P.; Jeyasri, R.; Bharathi, R.K.A.S.; Suba, V.; Pandian, S.T.K.; Ramesh, M. Global integrated omics expression analyses of abiotic stress signaling HSF transcription factor genes in *Oryza sativa* L.: An in silico approach. *Genomics* **2020**, 112, 908–918. [CrossRef]
- 80. Le, T.D.; Gathignol, F.; Vu, H.T.; Nguyen, K.L.; Tran, L.H.; Vu, H.T.T.; Dinh, T.X.; Lazennec, F.; Pham, X.H.; Véry, A.-A.; et al. Genome-Wide Association Mapping of Salinity Tolerance at the Seedling Stage in a Panel of Vietnamese Landraces Reveals New Valuable QTLs for Salinity Stress Tolerance Breeding in Rice. *Plants* **2021**, *10*, 1088. [CrossRef]
- 81. Chan, A.P.; Pertea, G.; Cheung, F.; Lee, D.; Zheng, L.; Whitelaw, C.; Pontaroli, A.C.; San Miguel, P.; Yuan, Y.; Bennetzen, J.; et al. The TIGR maize database. *Nucleic Acids Res.* **2006**, 34, D771–D776. [CrossRef]
- 82. Altschul, S.F.; Gish, W.; Miller, W.; Myers, E.W.; Lipman, D.J. Basic local alignment search tool. *J. Mol. Biol* **1990**, 215, 403–410. [CrossRef]
- 83. Bairoch, A.; Apweiler, R. The SWISS-PROT protein sequence database and its supplement TrEMBL in 2000. *Nucleic Acids Res.* **2000**, *28*, 45–48. [CrossRef]

Plants 2021, 10, 1472 17 of 19

84. Jeyasri, R.; Muthuramalingam, P.; Satish, L.; Adarshan, S.; Aishwarya Lakshmi, M.; Pandian, S.K.; Chen, J.T.; Ahmar, S.; Wang, X.; Freddy, M.P.; et al. The role of *OsWRKY* genes in rice when faced single and multiple abiotic stresses. *Agronomy* **2021**, *11*, 1301. [CrossRef]

- 85. Wang, Z.L.; Li, P.H.; Fredricksen, M.; Gong, Z.Z.; Kim, C.S.; Zhang, C.; Bohnert, H.J.; Zhu, J.K.; Bressan, R.A.; Hasegawa, P.M.; et al. Expressed sequence tags from Thellungiella halophila, a new model to study plant salt-tolerance. *Plant Sci.* **2004**, *166*, 609–616. [CrossRef]
- 86. Kawasaki, S.; Borchert, C.; Deyholos, M.; Wang, H.; Brazille, S.; Kawai, K.; Galbraith, D.; Bohnert, H.J. Gene expression profiles during the initial phase of salt stress in rice. *Plant Cell* **2001**, *13*, 889–905. [CrossRef] [PubMed]
- 87. Sreenivasulu, N.; Altschmied, L.; Radchuk, V.; Gubatz, S.; Wobus, U.; Weschke, W. Transcript profiles and deduced changes of metabolic pathways in maternal and filial tissues of developing barley grains. *Plant Jour* **2004**, *37*, 539–553. [CrossRef] [PubMed]
- 88. Lee, J.; Williams, M.E.; Tingey, S.V.; Rafalski, J.A. DNA array profiling of gene expression changes during maize embryo development. *Funct. Integr. Genom.* **2002**, *2*, 13–27. [CrossRef] [PubMed]
- 89. Buitink, J.; Leger, J.J.; Guisle, I.; Vu, B.L.; Wuilleme, S.; Lamirault, G.; Le Bars, A.; Le Meur, N.; Becker, A.; Küster, H.; et al. Transcriptome profiling uncovers metabolic and regulatory processes occurring during the transition from desiccation sensitive to desiccation-tolerant stages in Medicago truncatula seeds. *Plant J.* 2006, 47, 735–750. [CrossRef] [PubMed]
- 90. Liu, Y.; Ji, X.; Zheng, L.; Nie, X.; Wang, Y. Microarray analysis of transcriptional responses to abscisic acid and salt stress in Arabidopsis thaliana. *Int. J. Mol. Sci.* **2013**, *14*, 9979–9998. [CrossRef] [PubMed]
- 91. Rensink, W.A.; Buell, C.R. Microarray expression profiling resources for plant genomics. *Trends Plant Sci.* **2005**, *10*, 603–609. [CrossRef] [PubMed]
- 92. Li, L.; Deng, X.W. Microarray-based Approaches to Rice Transcriptome Analysis. In *Rice Biology in the Genomics Era*; Springer: Berlin/Heidelberg, Germany, 2008; pp. 37–51.
- 93. Kumar, R.; Sharma, V.; Suresh, S.; Ramrao, D.P.; Veershetty, A.; Kumar, S.; Priscilla, K.; Hangargi, B.; Narasanna, R.; Pandey, M.K.; et al. Understanding Omics Driven Plant Improvement and de novo Crop Domestication: Some Examples. *Front. Genet.* **2021**, *12*, 415. [CrossRef] [PubMed]
- 94. Yilmaz, A.; Nishiyama, M.Y., Jr.; Fuentes, B.G.; Souza, G.M.; Janies, D.; Gray, J.; Grotewold, E. GRASSIUS: A platform for comparative regulatory genomics across the grasses. *Plant Physiol.* **2009**, *149*, 171–180. [CrossRef] [PubMed]
- 95. Jin, J.; Tian, F.; Yang, D.C.; Meng, Y.Q.; Kong, L.; Luo, J.; Gao, G. PlantTFDB 4.0: Toward a central hub for transcription factors and regulatory interactions in plants. *Nucleic Acids Res.* **2016**, *45*, gkw982. [CrossRef]
- 96. Roychoudhury, A.; Paul, S.; Basu, S. Cross-talk between abscisic acid-dependent and abscisic acid-independent pathways during abiotic stress. *Plant Cell Rep.* **2013**, *32*, 985–1006. [CrossRef] [PubMed]
- 97. Czolpinska, M.; Rurek, M. Plant glycine-rich proteins in stress response: An emerging, still prospective story. *Front. Plant Sci.* **2018**, *9*, 302. [CrossRef]
- 98. Zenda, T.; Liu, S.; Dong, A.; Duan, H. Advances in Cereal Crop Genomics for Resilience under Climate Change. *Life* **2021**, *11*, 502. [CrossRef]
- 99. Koyro, H.W.; Ahmad, P.; Geissler, N. Abiotic stress responses in plants: An overview. In *Environmental Adaptations and Stress Tolerance of Plants in the Era of Climate Change*; Springer Science & Business Media: New York, NY, USA, 2012; pp. 1–28.
- 100. Seki, M.; Narusaka, M.; Abe, H.; Kasuga, M.; Yamaguchi-Shinozaki, K.; Carninci, P.; Hayashizaki, Y.; Shinozaki, K. Monitoring the expression pattern of 1300 *Arabidopsis* genes under drought and cold stresses by using a full-length cDNA microarray. *Plant Cell* 2001, 13, 61–72. [CrossRef] [PubMed]
- 101. Seki, M.; Narusaka, M.; Ishida, J.; Nanjo, T.; Fujita, M.; Oono, Y.; Kamiya, A.; Nakajima, M.; Enju, A.; Sakurai, T. Monitoring the expression profiles of 7000 *Arabidopsis* genes under drought, cold and high-salinity stresses using a full-length cDNA microarray. *Plant Jour* 2002, 31, 279–292. [CrossRef]
- 102. Fowler, S.; Thomashow, M.F. *Arabidopsis* transcriptome profiling indicates that multiple regulatory pathways are activated during cold acclimation in addition to the CBF cold response pathway. *Plant Cell* **2002**, *14*, 1675–1690. [CrossRef]
- 103. Lenka, S.K.; Katiyar, A.; Chinnusamy, V.; Bansal, K.C. Comparative analysis of drought-responsive transcriptome in Indica rice genotypes with contrasting drought tolerance. *Plant Biotechnol. J.* **2011**, *9*, 315–327. [CrossRef]
- 104. Dubouzet, J.G.; Sakuma, Y.; Ito, Y.; Kasuga, M.; Dubouzet, E.G.; Miura, S.; Seki, M.; Shinozaki, K.; Yamaguchi- Shinozaki, K. OsDREB genes in rice, *Oryza sativa* L., encode transcription activators that function in drought-, high-salt- and cold responsive gene expression. *Plant. J.* 2003, *33*, 751–763. [CrossRef]
- 105. Paul, S.; Roychoudhury, A. Transcriptome profiling of abiotic stress-responsive genes during cadmium chloride-mediated stress in two indica rice varieties. *J. Plant Growth Regul.* **2018**, *37*, 657–667. [CrossRef]
- 106. Kathuria, H.; Giri, J.; Nataraja, K.N.; Murata, N.; Udayakumar, M.; Tyagi, A.K. Glycinebetaine-induced water-stress tolerance in codA-expressing transgenic indica rice is associated with up-regulation of several stress responsive genes. *Plant Biotechnol. J.* **2009**, *7*, 512–526. [CrossRef]
- 107. Quan, R.; Shang, M.; Zhang, H.; Zhao, Y.; Zhang, J. Engineering of enhanced glycine betaine synthesis improves drought tolerance in maize. *Plant Biotechnol. J.* **2004**, *2*, 477–486. [CrossRef] [PubMed]
- 108. Maghsoudi, K.; Emam, Y.; Niazi, A.; Pessarakli, M.; Arvin, M.J. P5CS expression level and proline accumulation in the sensitive and tolerant wheat cultivars under control and drought stress conditions in the presence/absence of silicon and salicylic acid. *J. Plant Interact.* **2018**, *13*, 461–471. [CrossRef]

Plants **2021**, 10, 1472 18 of 19

109. Ma, H.; Liu, C.; Li, Z.; Ran, Q.; Xie, G.; Wang, B.; Fang, S.; Chu, J.; Zhang, J. ZmbZIP4 contributes to stress resistance in maize by regulating ABA synthesis and root development. *Plant Physiol.* **2018**, *178*, 753–770. [CrossRef]

- 110. Das, P.; Lakra, N.; Nutan, K.K.; Singla-Pareek, S.L.; Pareek, A. A unique bZIP transcription factor imparting multiple stress tolerance in Rice. *Rice* **2019**, *12*, 58. [CrossRef]
- 111. Pandey, A.S.; Sharma, E.; Jain, N.; Singh, B.; Burman, N.; Khurana, J.P. A rice bZIP transcription factor, OsbZIP16, regulates abiotic stress tolerance when over-expressed in Arabidopsis. *J. Plant Biothem. Biotechnol.* **2018**, 27, 393–400. [CrossRef]
- 112. Jiang, Z.; Song, G.; Shan, X.; Wei, Z.; Liu, Y.; Jiang, C.; Jiang, Y.; Jin, F.; Li, Y. Association analysis and identification of ZmHKT1; 5 variation with salt-stress tolerance. *Front. Plant Sci.* **2018**, *9*, 1485. [CrossRef] [PubMed]
- 113. Xu, B.; Hrmova, M.; Gilliham, M. High affinity Na + transport by wheat HKT1;5 is blocked by K +. *Plant Direct* **2020**, *4*, e00275. [CrossRef]
- 114. Basu, S.; Roychoudhury, A. Expression profiling of abiotic stress-inducible genes in response to multiple stresses in rice (*Oryza sativa* L.) varieties with contrasting level of stress tolerance. *Biomed. Res. Int.* **2014**, 2014, 706890. [CrossRef]
- 115. Muthuramalingam, P.; Jeyasri, R.; Selvaraj, A.; Kalaiyarasi, D.; Aruni, W.; Pandian, S.T.K.; Ramesh, M. Global transcriptome analysis of novel stress associated protein (SAP) genes expression dynamism of combined abiotic stresses in *Oryza sativa* (L.). *J. Biomol. Struct. Dyn.* **2020**, *39*, 2106–2117. [CrossRef]
- 116. Li, S.; Zhao, Q.; Zhu, D.; Yu, J. A DREB-like transcription factor from maize (*Zea mays*), ZmDREB4. 1, plays a negative role in plant growth and development. *Front. Plant Sci.* 2018, 9, 395. [CrossRef]
- 117. Tang, Y.; Bao, X.; Zhi, Y.; Wu, Q.; Guo, Y.; Yin, X.; Zeng, L.; Li, J.; Zhang, J.; He, W.; et al. Overexpression of a MYB family gene, OsMYB6, increases drought and salinity stress tolerance in transgenic rice. *Front. Plant Sci.* **2019**, *10*, 168. [CrossRef]
- 118. Tchagang, A.B.; Fauteux, F.; Tulpan, D.; Pan, Y. Bioinformatics identification of new targets for improving low temperature stress tolerance in spring and winter wheat. *BMC Bioinform.* **2017**, *18*, 1746. [CrossRef] [PubMed]
- 119. Iqbal, I.; Tripathi, R.K.; Wilkins, O.; Singh, J. Thaumatin-like Protein (TLP) gene family in barley: Genome-wide exploration and expression analysis during germination. *Genes* **2020**, *11*, 1080. [CrossRef]
- 120. Liu, A.L.; Zou, J.; Liu, C.F.; Zhou, X.Y.; Zhang, X.W.; Luo, G.Y.; Chen, X.B. Over-expression of OsHsfA7 enhanced salt and drought tolerance in transgenic rice. *BMB Rep.* **2013**, *46*, 31. [CrossRef]
- 121. Muthusamy, S.K.; Dalal, M.; Chinnusamy, V.; Bansal, K.C. Genome-wide identification and analysis of biotic and abiotic stress regulation of small heat shock protein (HSP20) family genes in bread wheat. *J. Plant Physiol.* **2017**, 211, 100–113. [CrossRef] [PubMed]
- 122. El-Esawi, M.A.; Alayafi, A.A. Overexpression of rice Rab7 gene improves drought and heat tolerance and increases grain yield in rice (*Oryza sativa* L.). *Genes* **2019**, *10*, 56. [CrossRef] [PubMed]
- 123. Chen, C.; Heo, J.B. Overexpression of constitutively active OsRab11 in plants enhances tolerance to high salinity levels. *J. Plant. Biol.* **2018**, *61*, 169–176. [CrossRef]
- 124. Yao, X.; Wu, K.; Yao, Y.; Li, J.; Ren, Y.; Chi, D. The response mechanism of the HVA1 gene in hulless barley under drought stress. *Ital. J. Agron.* **2017**, *12*, 804. [CrossRef]
- 125. Chen, Y.S.; Lo, S.F.; Sun, P.K.; Lu, C.A.; Ho, T.H.D.; Yu, S.M. A late embryogenesis abundant protein HVA 1 regulated by an inducible promoter enhances root growth and abiotic stress tolerance in rice without yield penalty. *Plant Biotechnol. J.* **2015**, 13, 105–116. [CrossRef]
- 126. Gürel, F.; Öztürk, Z.N.; Uçarlı, C.; Rosellini, D. Barley genes as tools to confer abiotic stress tolerance in crops. *Front. Plant Sci.* **2016**, *7*, 1137. [CrossRef] [PubMed]
- 127. Zhang, Z.; Zhang, Q.; Wu, J.; Zheng, X.; Zheng, S.; Sun, X.; Qiu, Q.; Lu, T. Gene knockout study reveals that cytosolic ascorbate peroxidase 2 (OsAPX2) plays a critical role in growth and reproduction in rice under drought, salt and cold stresses. *PLoS ONE* **2013**, *8*, e57472. [CrossRef]
- 128. Luna, C.M.; Pastori, G.M.; Driscoll, S.; Groten, K.; Bernard, S.; Foyer, C.H. Drought controls on H2O2 accumulation, catalase (CAT) activity and CAT gene expression in wheat. *J. Exp. Bot.* 2005, *56*, 417–423. [CrossRef] [PubMed]
- 129. Shiraya, T.; Mori, T.; Maruyama, T.; Sasaki, M.; Takamatsu, T.; Oikawa, K.; Itoh, K.; Kaneko, K.; Ichikawa, H.; Mitsui, T. Golgi/plastid-type manganese superoxide dismutase involved in heat-stress tolerance during grain filling of rice. *Plant Biotechnol. J.* 2015, *13*, 1251–1263. [CrossRef] [PubMed]
- 130. Guan, Q.; Liao, X.; He, M.; Li, X.; Wang, Z.; Ma, H.; Yu, S.; Liu, S. Tolerance analysis of chloroplast OsCu/Zn-SOD overexpressing rice under NaCl and NaHCO3 stress. *PLoS ONE* **2017**, *12*, e0186052. [CrossRef] [PubMed]
- 131. de Deus, K.E.; Lanna, A.C.; Abreu, F.R.M.; Silveira, R.D.D.; Pereira, W.J.; Brondani, C.; Vianello, R.P. Molecular and biochemical characterization of superoxide dismutase (SOD) in upland rice under drought. *Embrapa Arroz Feijão-Artigo Periódico Indexado* (ALICE) 2015, 9, 744–753.
- 132. Xiao, B.Z.; Chen, X.; Xiang, C.B.; Tang, N.; Zhang, Q.F.; Xiong, L.Z. Evaluation of seven function-known candidate genes for their effects on improving drought resistance of transgenic rice under field conditions. *Mol. Plant* **2009**, *2*, 73–83. [CrossRef]
- 133. Xie, G.; Kato, H.; Sasaki, K.; Imai, R. A cold-induced thioredoxinh of rice, OsTrx23, negatively regulates kinase activities of OsMPK3 and OsMPK6 in vitro. *FEBS Lett.* **2009**, *583*, 2734–2738. [CrossRef]
- 134. Menguer, P.K.; Sperotto, R.A.; Ricachenevsky, F.K. A walk on the wild side: Oryza species as source for rice abiotic stress tolerance. Genet. *Mol. Biol.* 2017, 40, 238–252. [CrossRef]

Plants 2021, 10, 1472 19 of 19

135. Mohanty, A.; Kathuria, H.; Ferjani, A.; Sakamoto, A.; Mohanty, P.; Murata, N.; Tyagi, A. Transgenics of an elite indica rice variety Pusa Basmati 1 harbouring the codA gene are highly tolerant to salt stress. *Theor. Appl. Genet.* **2002**, *106*, 51–57. [CrossRef]

- 136. Thomas, S.; Krishna, G.K.; Yadav, P.; Pal, M. Cloning and abiotic stress responsive expression analysis of Arginine decarboxylase genes in contrasting rice genotypes. *Indian J. Genet.* **2019**, *79*, 411–419. [CrossRef]
- 137. Lee, H.J.; Abdula, S.E.; Jang, D.W.; Park, S.H.; Yoon, U.H.; Jung, Y.J.; Kang, K.K.; Nou, I.S.; Cho, Y.G. Overexpression of the glutamine synthetase gene modulates oxidative stress response in rice after exposure to cadmium stress. *Plant Cell Rep.* **2013**, 32, 1521–1529. [CrossRef]
- 138. Garg, A.K.; Kim, J.K.; Owens, T.G.; Ranwala, A.P.; Do Choi, Y.; Kochian, L.V.; Wu, R.J. Trehalose Accumulation in Rice Plants Confers High Tolerance Levels to Different Abiotic Stresses. *Proc. Natl. Acad. Sci. USA* **2002**, *99*, 15898–15903. [CrossRef]
- 139. Abebe, T.; Guenzi, A.C.; Martin, B.; Cushman, J.C. Tolerance of mannitol-accumulating transgenic wheat to water stress and salinity. *Plant Physiol.* **2003**, *131*, 1748–1755. [CrossRef] [PubMed]
- 140. Wei, S.; Hu, W.; Deng, X.; Zhang, Y.; Liu, X.; Zhao, X.; Luo, Q.; Jin, Z.; Li, Y.; Zhou, S.; et al. A rice calcium-dependent protein kinase OsCPK9 positively regulates drought stress tolerance and spikelet fertility. *BMC Plant Biol.* **2014**, *14*, 133. [CrossRef] [PubMed]
- 141. Geda, C.K.; Repalli, S.K.; Dash, G.K.; Swain, P.; Rao, G.J.N. Enhancement of Drought Tolerance in Rice through Introgression of Arabidopsis DREB1A through Transgenic Approach. *J. Rice Res.* **2019**, 7, 2.
- 142. Amin, U.S.M.; Biswas, S.; Elias, S.M.; Razzaque, S.; Haque, T.; Malo, R.; Seraj, Z.I. Enhanced salt tolerance conferred by the complete 2.3 kb cDNA of the rice vacuolar Na+/H+ antiporter gene compared to 1.9 kb coding region with 5' UTR in transgenic lines of rice. *Front. Plant Sci.* **2016**, *7*, 14. [CrossRef] [PubMed]
- 143. Khan, I.; Mohamed, S.; Regnault, T.; Mieulet, D.; Guiderdoni, E.; Sentenac, H.; Véry, A.A. Constitutive contribution by the rice OsHKT1; 4 Na+ transporter to xylem sap desalinization and low Na+ accumulation in young leaves under low as high external Na+ conditions. *Front. Plant Sci.* 2020, *11*, 1130. [CrossRef] [PubMed]
- 144. Mohanty, B.; Takahashi, H.; Benildo, G.; Wijaya, E.; Nakazono, M.; Lee, D.Y. Transcriptional regulatory mechanism of alcohol dehydrogenase 1-deficient mutant of rice for cell survival under complete submergence. *Rice* **2016**, *9*, 51. [CrossRef]
- 145. Zang, X.; Geng, X.; Wang, F.; Liu, Z.; Zhang, L.; Zhao, Y.; Tian, X.; Ni, Z.; Yao, Y.; Xin, M.; et al. Overexpression of wheat ferritin gene TaFER-5B enhances tolerance to heat stress and other abiotic stresses associated with the ROS scavenging. *BMC Plant Biol.* **2017**, *17*, 14. [CrossRef]
- 146. Quimio, C.A.; Torrizo, L.B.; Setter, T.L.; Ellis, M.; Grover, A.; Abrigo, E.M.; Oliva, N.P.; Ella, E.S.; Carpena, A.L.; Ito, O.; et al. Enhancement of submergence tolerance in transgenic rice overproducing pyruvate decarboxylase. *J. Plant Physiol.* **2000**, *156*, 516–521. [CrossRef]
- 147. Wang, Y.; Deng, C.; Ai, P.; Zhang, Z. ALM1, encoding a Fe-superoxide dismutase, is critical for rice chloroplast biogenesis and drought stress response. *Crop J.* **2020**. [CrossRef]
- 148. Saibi, W.; Brini, F. Superoxide dismutase (SOD) and abiotic stress tolerance in plants: An overview. In *Superoxide Dismutase:* Structure, Synthesis and Applications; Magliozzi, S., Ed.; Nova Science Publishers, Inc.: Hauppauge, NY, USA, 2018; pp. 101–142.
- 149. Alqudah, A.M.; Sallam, A.; Baenziger, P.S.; Börner, A. GWAS: Fast-forwarding gene identification and characterization in temperate Cereals: Lessons from Barley–A review. *J. Adv. Res.* **2020**, 22, 119–135. [CrossRef] [PubMed]
- 150. Muthuramalingam, P.; Krishnan, S.R.; Pandian, S.; Mareeswaran, N.; Aruni, W.; Pandian, S.K.; Ramesh, M. Global analysis of threonine metabolism genes unravel key players in rice to improve the abiotic stress tolerance. *Sci. Rep.* **2018**, *8*, 9270. [CrossRef] [PubMed]
- 151. Mitchell-Olds, T. Complex-trait analysis in plants. Genome Biol. 2010, 11, 113. [CrossRef]
- 152. Atwell, S.; Huang, Y.S.; Vilhjálmsson, B.J.; Willems, G.; Horton, M.; Li, Y.; Meng, D.; Platt, A.; Tarone, A.M.; Hu, T.T.; et al. Genome-wide association study of 107 phenotypes in Arabidopsis thaliana inbred lines. *Nature* **2010**, *465*, 627–631. [CrossRef]