

Review

Revisiting the Domestication Process of African *Vigna* Species (Fabaceae): Background, Perspectives and Challenges

Davide Panzeri ^{1,*} , Werther Guidi Nissim ^{1,2}, Massimo Labra ¹  and Fabrizio Grassi ^{1,*}

¹ Department of Biotechnology and Bioscience, University of Milan-Bicocca, Piazza della Scienza 2, 20126 Milano, Italy; werther.guidi.nissim@gmail.com (W.G.N.); massimo.labra@unimib.it (M.L.)

² Department of Agriculture, Food, Environment and Forestry (DAGRI), University of Florence, Viale delle Idee 30, 50019 Sesto Fiorentino, Italy

* Correspondence: davide.panzeri@unimib.it (D.P.); fabrizio.grassi@unimib.it (F.G.)

Abstract: Legumes are one of the most economically important and biodiverse families in plants recognised as the basis to develop functional foods. Among these, the *Vigna* genus stands out as a good representative because of its relatively recent African origin as well as its outstanding potential. Africa is a great biodiversity centre in which a great number of species are spread, but only three of them, *Vigna unguiculata*, *Vigna subterranea* and *Vigna vexillata*, were successfully domesticated. This review aims at analysing and valorising these species by considering the perspective of human activity and what effects it exerts. For each species, we revised the origin history and gave a focus on where, when and how many times domestication occurred. We provided a brief summary of bioactive compounds naturally occurring in these species that are fundamental for human wellbeing. The great number of wild lineages is a key point to improve landraces since the domestication process caused a loss of gene diversity. Their genomes hide a precious gene pool yet mostly unexplored, and genes lost during human activity can be recovered from the wild lineages and reintroduced in cultivated forms through modern technologies. Finally, we describe how all this information is game-changing to the design of future crops by domesticating *de novo*.

Keywords: *Vigna* genus; introgression; hybridisation; phylogeny; *de novo* domestication; feralisation; bioactive compounds



Citation: Panzeri, D.; Guidi Nissim, W.; Labra, M.; Grassi, F. Revisiting the Domestication Process of African *Vigna* Species (Fabaceae): Background, Perspectives and Challenges. *Plants* **2022**, *11*, 532. <https://doi.org/10.3390/plants11040532>

Academic Editors: Milan S. Stankovic, Paula Baptista and Petronia Carillo

Received: 25 January 2022

Accepted: 13 February 2022

Published: 16 February 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Legumes (Fabaceae) are considered one of the most important families of plants for human nutrition, especially considering the rapid growth rate of the world population [1]. However, almost all the efforts and resources invested in agriculture during the last century were focused on improving the yield, resistance and quality of a few specific staple crops. Neglected landraces are regarded as having interesting potential, and recent studies have demonstrated that some wild legumes can be an important target to develop modern functional foods because they possess various bioactive molecules that interact positively with human health [2–5]. Among these, members of the *Vigna* genus show a growing social and economic importance in several African regions, especially where the local population is not able to afford animal proteins [6–8]. Their seeds are rich in essential amino acids and contain a high concentration of minerals, lipids and vitamins [9,10].

The genus *Vigna* (Savi, 1824), which belongs to the tribe Phaseoleae of the family Fabaceae, includes over 100 species [11] distributed in the tropical and subtropical areas of the world [12] grouped in five subgenera: *Vigna*, *Ceratotropis*, *Plectotropis*, *Lasiosporon* and *Haydonia* [13–15]. Phylogenetic findings propose the age of split between *Phaseolus* and *Vigna* genera at about 8–10 million years (Mya) and the age of split between *Ceratotropis* and *Vigna* subgenera at about 3–4 Mya [13–17], but the genetic relationships between subgenera are particularly complex and far from being completely solved. Although most

domesticated or semi-domesticated species are distributed in Asia, the greatest diversity of the *Vigna* genus is located in Sub-Saharan Africa [14,18]. *Vigna* subgenus, distributed in Africa, includes about 40 wild and 2 domesticated species, namely cowpea (also called black-eyed peas, chawli and kunde) (*Vigna unguiculata* L.) and Bambara groundnut (*V. subterranea* L.) [19] while *Ceratotropis* (Piper) Verdc., distributed in Asia, contains 21 wild and 7 domesticated species used widely for food and forage, namely mungbean or green gram (*V. radiata* L. Wilczek), black gram (*V. mungo* L. Hepper), moth bean (*V. aconitifolia* Jacq. Maréchal), rice bean (*V. umbellata* Thunb. Ohwi and Ohashi), adzuki bean (*V. angularis* L. Ohwi and Ohashi), creole bean (*V. reflexo-pilosa* Hayata), jungli bean (*V. trilobata* L. Verdc.). [15,20–22]. Moreover, three species belonging to *Plectrotropis* (Schumach.) are distributed in Africa, including tuber cowpea (*V. vexillata* L.) [23]. Most of the African *Vigna* germplasm is based on wild plants and neglected or underutilized landraces, and many of these lineages are declining with a high risk of extinction. The recovery of wild accessions and research devoted to the phylogeny of the genus is therefore essential to prevent genetic erosion and the loss of *Vigna* diversity.

Plant domestication is widely recognised as an accelerated evolutionary process driven by a synergistic impact of human and natural selection, occurring in geographically restricted areas from wild progenitors. In legumes, the main modification is the loss of seed pod dehiscence or shattering [24,25]. The split at the dorsal and ventral sutures of the dry pod and successive release of the seeds occurs due to the desiccation of lignified cells in the pods [26]. The shattering habit is related to environmental aridity and persists in many varieties of domesticated *Vigna* species, thereby determining severe yield losses [27,28]. Additional implementations in *Vigna* domesticated species include an increase in seed or fruit size, change in seed colour, loss of seed dormancy, apical dominance and change in flowering timing [29–33]. These modifications were inherited more or less effectively in the various vine species currently cultivated, and this is the basis of the agrobiodiversity of this genus.

Generally, the current existing crops show lower resistance to biotic and abiotic stress compared to wild relatives, and often they have reached their full yield. The selection of desirable traits and breeding processes to improve crop productivity have caused the depletion of diversity and the increase in the frequency of deleterious genetic variants that are fixed in the genomes of crops [34–36]. These constraints have a serious impact on agriculture, limiting the possibility to grow such crops under more extreme environmental conditions. Thanks to this residual genetic diversity and also to studies performed on *Vigna* species, most of the accessions are well adapted to a wide range of extreme environmental conditions, such as sandy beaches, arid lands and wetlands, harbouring tolerance and resistance genes towards biotic and abiotic stresses. These genetic traits are used for developing new stress-tolerant crops [37–43]. By contrast, less is known about the effects of domestication on the nutritional value of seeds [7] even if recent studies have reported that cultivated legumes show a lower carotenoid and protein content in seeds compared with the wild relatives [44,45]. Where, when and how many times the domestication process of African *Vigna* crops occurred continues to be debated among researchers. Although archaeological remains of *Vigna* indicate that the domestication process in Africa was started recently compared to other field crops [46,47]. Modern evolutionary models proposed for other crops suggest that the predomestication phase may have lasted several thousands of years [48,49]. Generally, the centres of origin are also recognized as centres of diversity, and thus these areas require special precautionary measures of conservation [50]. Although for many crops the single-origin model is usually the most parsimonious, the hypothesis that provides multiple origins starting from independent founder lineages seems well suited for the crops of *Vigna* originated in Africa [51,52]. Moreover, despite whether and to what extent introgression influences the domestication process is still underexplored, some studies already show that gene flow between cowpea and its wild relatives may occur. Pervasive introgression can also intensify the feralisation process, promoting the

crops to return to a wild environment and causing serious problems for the conservation of biodiversity [53].

In this review, we re-examine the available scientific information on the domestication process of three African *Vigna* crops and discuss the future perspectives and challenges in the light of modern technologies in the time of climate change and new parading of conservative agriculture strategies. Another crucial point in exploring natural biodiversity is not only a matter of sustainability but also a matter of human health. A balanced diet gives extreme benefits to people's wellbeing by properly assuming the correct amount of micro and macro nutrients as well as useful, healthy bioactive compounds. Finding and characterising these compounds is an ambitious challenge for researchers thus we briefly summarise the bioactivity of some compounds, and we discuss how human activity and breeding has impacted the variability of molecules.

Although recent genetic studies have led to a deeper understanding of these crops, the continuation of investigating the domestication process through a multidisciplinary approach which includes genomic, transcriptomic, metabolomic and epigenomic analyses is needed to highlight the wide agronomic opportunities related to these species. Moreover, recent techniques of gene editing have opened new and crucial ways to re-design modern crops because traditional genetic improvement is generally limited by the cross-compatibility between species. Thus, because the *de novo* domestication process may represent a turn toward more modern and sustainable agriculture, further efforts are needed to explore the genome diversity of wild germplasm.

2. *Vigna unguiculata* (L.) Walp.

V. unguiculata, which was considered an orphan crop for several decades, has recently become one of the most important legumes in the world. Its name derives from Latin and means "with a small claw", referring to the size of the claw of the petals [54] or commonly named as "cowpea" because of its use as fodder for cows [55] and black-eyed pea/bean for the black hilum. This crop is largely cultivated, especially in semiarid regions of Africa and Asia where other crops fail to grow [10]. Currently, on a global scale, about 15 million hectares are dedicated to *V. unguiculata*, with an annual production of 7 million Mg and an average yield of 0.6 Mg ha⁻¹ [56]. The most interesting environmental traits of this species are represented by the generalized low agrochemical input requirements. In fact, this crop shows relatively high adaptation to drought, especially in comparison to other legumes [57] and can fix up to 200 kg N ha⁻¹ [58] with a positive soil N balance of up to 92 kg ha⁻¹ [59]. Nevertheless, several abiotic and biotic constraints (i.e., low soil fertility, pests, diseases, parasitic weeds, and nematodes) limit the yield [43,60,61]. Moreover, low productivity is often associated with the use of traditional and unimproved varieties, still widely cultivated in Africa [62]. This crop has a fundamental role in human nutrition, showing seeds rich in proteins and essential amino acids (i.e., tryptophan and lysine), carbohydrates, folic acid and minerals. Recent studies carried on a large sampling have shown high variability in protein and mineral concentrations, suggesting that some lineages could be potential sources of genes useful to produce new varieties [63–66].

The high number of wild subspecies found exclusively in Africa strongly supports the idea of an African origin. However, intraspecies phylogeny remains far from being completely elucidated [67]. The centre of origin of the species is probably located in the southernmost regions of Africa, where most subspecies are found and where most genetic diversity could be still hidden [68]. Several taxonomic revisions based on morphological and molecular traits permitted to identify 10 perennial and 1 annual subspecies, the latter split into two varieties: ssp. *unguiculata* var. *unguiculata* (domesticated cowpea) and ssp. *unguiculata* var. *spontanea* (Schweinf.) Pasquet., also known as subsp. *dekindtiana* sensu Verdcourt non Harms [69–76]. Besides the domesticated cowpea, the *dekindtiana* group includes some obligate short-day wild forms, well adapted to arid environments. While the var. *spontanea* grows especially around cultivated fields and roadsides, and it is recognized as the progenitor of domesticated cowpea [75,77–79], the subspecies *alba*, *pubescens*, *tenuis*,

stenophylla and *dekintiana* are perennial plants [75,76]. The development of new molecular tools to discriminate among wild, weedy, and cultivated accessions is considered a modern and fundamental target, particularly needed for disentangling the complex taxonomic relationships among subspecies and to discriminate between true wild plants and ferals.

Although little is known about the domestication process, some scientists have hypothesized that ancient cowpea progenitors, such as the modern wild forms, were adapted to dry habitats and grew spontaneously south of the Sahara Desert [80]. These plants were gathered, cultivated and dispersed by men near the villages, but they were unsuited for cultivation. Although they did not show high yield, the wild lineages were spread in the humid zones thanks to their pods that remained closed for the humid atmosphere. Through several generations of cultivation, new mutants have arisen, showing interesting domestic traits, including resistance to shattering. Subsequently, humans have selected and helped spread these landraces by exchange and trade activities. Since the oldest archaeological records of domesticated forms found in central Ghana are dated around 1500 BC, the domestication process likely started before that period (Figure 1) [47,81]. However, the precise origin is widely debated, and two independent domestication centres in West and East Africa are proposed by different authors [68,74,79,82–86].

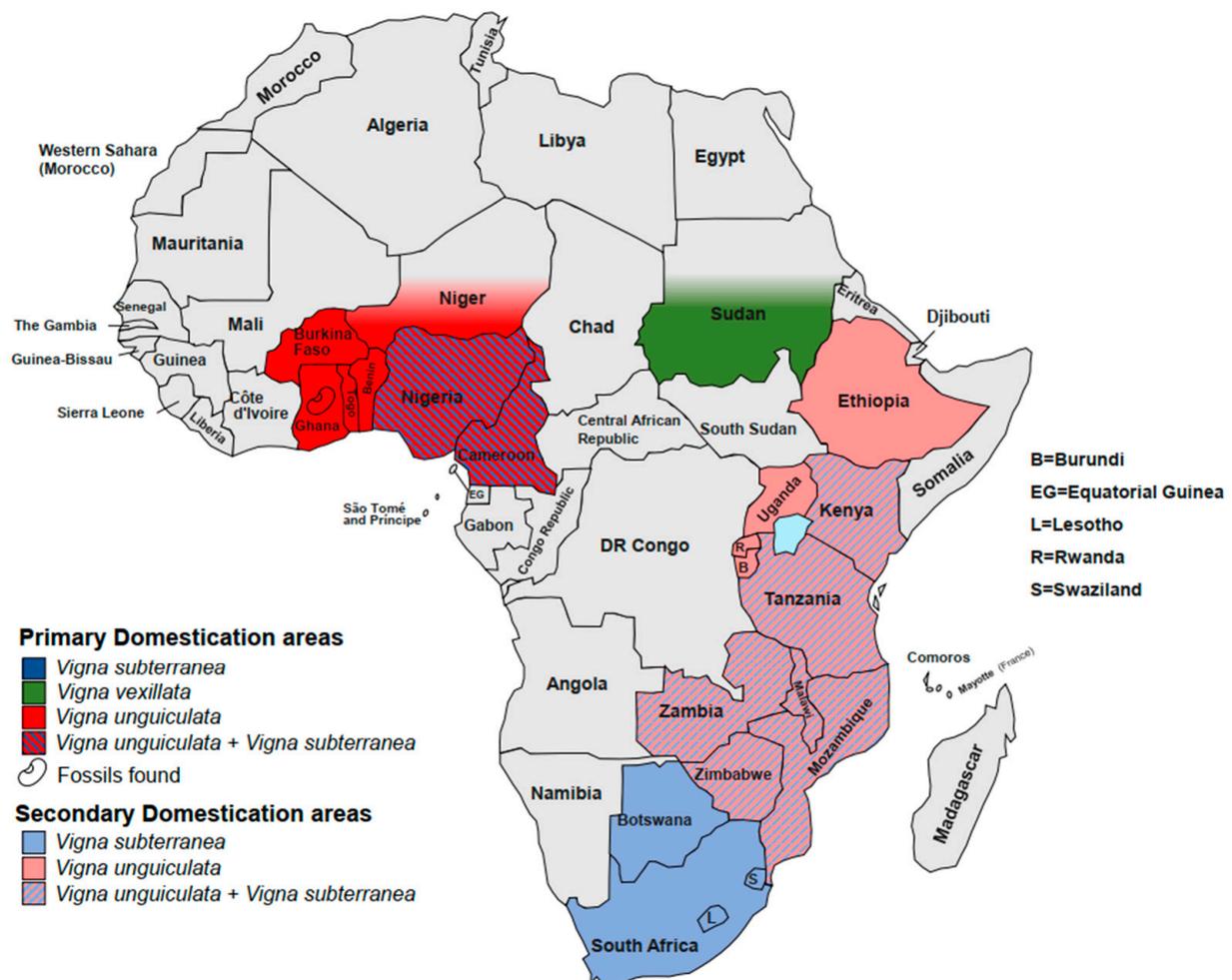


Figure 1. Primary and secondary domestication sites in Africa.

Morphological and DNA markers support the idea that domestication occurred only once, but analyses on whole genomes provide evidence for more independent domestication events in Africa and diversification events out of Africa [51,87]. Analyses of genetic variability are generally applied to identify the origin of species and the groups of acces-

sions that show high variability in certain geographic areas and are interpreted as the most ancient populations. Although cowpea from West Africa showed a high genetic variability [88], cultivated accessions grown in East and West Africa were shown to be most closely related to the respective local wild lineages [52,89], thereby indicating that domestication could have occurred in both regions. Outside Africa, cultivated cowpea was exposed to different ecological conditions, including new biotic and abiotic stresses that probably have contributed to shaping the genetic structure of landraces. When cowpea moved through Asian regions (especially in Thailand, China, the Philippines and India), it encountered environments with more humidity and less brightness where the drying of pods and grains was hindered. Some accessions were selected for the use of the immature pods to produce a peculiar form of vegetable called yardlong bean (*V. unguiculata* ssp. *unguiculata* cv. *sesquipedalis*) [51,90,91]. Although Chinese accessions show lower genetic diversity compared to African cowpea, signals of genetic bottlenecks lead to the conclusion that a limited number of relatively recent selection events occurred; however, where the selection process arose is still unknown [92]. Moreover, other cultivar groups (e.g., 'Textilis', 'Biflora' or 'Cylindrica', 'Melanophthalmus') are classified by morphological traits [75,93]. Still, additional genomic analyses should be performed to confirm the genetic relationships and understand how and where these accessions originated [67,85,88,94,95].

3. *Vigna subterranea* (L.) Verdc.

Vigna subterranea, also named Bambara groundnut, is an indigenous African grain legume. Its common name derives from the groundnut (*Arachis hypogaea* L.) due to the hypogean pods' growth, whereas the "Bambara" name is derived from a Malian tribe [96]. Despite its potential in terms of nutritional value and resistance to biotic and abiotic stresses [97,98], Bambara is cultivated mainly in small farms or in families as a subsistence crop [99], and naturally grows in semi-arid regions in Africa. Regarding the origin of the species itself, the domesticated or semi-domesticated *Vigna subterranea* var. *subterranea* was most likely generated from its wild counterpart *Vigna subterranea* var. *spontanea* using both morphological and isozyme data [100,101].

The origin of this species is hypothesised to be in Mali, in the Timbuktu region [102], but the precise centre of origin is still unknown. In fact, there is no evidence of wild lineages in Mali [103]. Dalziel, Begemann and Goli [104–106] analysed a lot of morphologic traits such as seed morphology, seed pattern diversity and other diversity indices (number of days to maturity, pod length, number of stems per plant and internode length). They found that the most diversity is located in an area that spans from Jos Plateau and Yola Adamawa (Nigeria) to Garoua (Cameroon). Somta and Olukolu [107,108] evaluated the phylogeography of several accessions spread in Africa. The markers used (i.e., SSR and DaRT) showed a cluster with higher diversity in the area between Nigeria and Cameroon. The authors confirm the area of origin while suggesting a possible subsequent introduction of Western domesticated accessions in East Africa (Figure 1). In contrast, Aliyu et al. [97], in an overview of the past two decades of genetic diversity analysis, also proposed that the Southern African region could constitute a divergent time-spaced domestication event. However, the authors suggest that these hypotheses need further examination.

In terms of genetic diversity, Bambara has a peculiar behaviour. In fact, many authors studied Bambara's genetics with different techniques to clarify how wide the genetic pool is and how homogeneous the single landraces are. Molosiwa et al. [109] evaluated genetic distances between 24 landraces with phenotypic and genetic markers (i.e., SSR and DaRT). The main results report that landraces are different to each other, suggesting the existence of great allelic diversity among the various populations. At the same time, though, single landraces tend to be very homogeneous, and in three generations of inbreeding became pure lines. This is due mainly to its self-pollinating nature [110] but also small farmers, who, by breeding the same landraces, also acted as selection drivers [111,112]. Molosiwa [113] selected 12 SSR markers and 5 Bambara accessions to evaluate the potential for creating

pure lines, finding that these accessions at the second cycle of selection completely have lost the heterozygosity.

All these findings suggest that Bambara has incredible genetic potential. The genetic screening through the different lineages and the consequent discovery of peculiar sites of interest could be the basis for an improvement of crop programs. Moreover, the use of pure lines in agriculture is fundamental not only for the optimisation and standardisation of agricultural practices but also for the development of breeding programs. Currently, to the best of our knowledge, there are no reports of ongoing improvement or breeding programs for this species. The extremely wide pool of wild and domesticated accessions can be used to create ideal crops that can better withstand climate change as well as being able to grow with low agronomic inputs.

4. *Vigna vexillata* (L.) A. Rich.

Widely distributed in Africa, Asia, America and Australia, *V. vexillata* (Zombi pea) is one of the least known and underutilized *Vigna* crops. Likewise, *V. unguiculata*, Zombi pea shows a high morphological diversity probably determined by geological, ecological, climatic and anthropomorphic constraints that also determined exceptional patterns of genetic variability [19,71]. Eight varieties including *vexillata*, *angustifolia*, *ovata*, *dolichomena*, *yunnanensis*, *pluriflora*, *lobatiflora* and *macrosperma* are recognized [12,19,23,114,115]. Var. *macrosperma* shows typical traits associated with domestication syndrome such as bush-like habit, early flowering and higher seed yield [116,117]. Moreover, loss of seed dormancy and various degrees of pod shattering were detected in different crop accessions while the wild seeds remained intrinsically dormant [118,119]. Several authors reported that two forms were domesticated independently (i.e., seed type and tuber type), and some evidence lines suggest that the seed type was domesticated in Sudan, whereas the tuber type was domesticated in India (Figure 1) [120–124]. However, molecular analyses were performed on a limited number of accessions and loci [124], and the phylogenetic intra-specific delimitation has resulted in much more complexity than that of other *Vigna* crops [125]. Thus, modern genomic analyses are needed to resolve the genetic relationships and confirm the origin of the two forms.

Several studies have also shown that the Zombi pea is the result of a long adaptation process to different environmental stress, including acid, alkaline, saline, drought and wet soils [115,117,126–128]. Moreover, since some accessions were found to be resistant to different viral diseases and parasite insects, widely recognized as major pests of cowpea, this species is an important harbour of resistances to various biotic stresses, particularly useful to improve modern *Vigna* crops [129–134].

5. Healthy Natural Compounds for Designing Sustainable Crops

The process of domestication was selected during the early millennia due to all the characteristics that made a species very productive or easier to harvest. Nowadays, a lot of crops varieties that have a great yield and high contents of macronutrients exist, such as carbohydrates or proteins. However, bioactive compounds that are naturally present in the *Vigna* genus were never taken into account. In a world where the main concern is no more denutrition but instead malnutrition, the adoption of crops with high-value nutraceutical compounds becomes a challenge for the next generation. The *Vigna* genus is a great source of small proteins and secondary metabolites with nutraceutical roles in everyday diet.

Often agricultural practices themselves could stimulate the production of these compounds, such as hydric stress or no tillage with cover crops fields. However, they could not be sufficient to enhance the output of bioactive molecules. In this perspective, *de novo* domestication programs should consider these compounds to develop future healthy crops. In the next paragraph, we listed and discussed some of these molecules based on the nutraceutical activity they exert against three great world concerns.

5.1. Antioxidant and Anti-Inflammatory Activity

Nowadays, inflammation and oxidative stress are becoming great concerns due to detrimental effects on human health, and diet is a powerful way to protect cells from the rise of reactive oxygen species (ROS) as well as inflammatory processes. In this view, seeds of cowpea contain different phenols, and other pigments present in the seed coat [135] are able to promote antioxidant action; among these, quercetin and flavonols are very well represented [136]. Different works [137,138] demonstrated a clear correlation between antioxidant properties and the colour of the seed coat in different accessions of *Vigna vexillata* and *Vigna subterranea*. Sowndhararajan and Leu [139,140] identified that *Vigna vexillata* extracts three molecules with strong antioxidant properties. Daidzein, abscisic acid and quercetin were highly active and displayed a pivotal capacity to deny the inflammation pathway.

Studies performed on protein extracts of *Vigna subterranea* suggest that different protein fractions exert crucial properties against ROS relevant in cellular metabolism [141]. Furthermore, a review by Quan et al. [142] summarised how polyphenols and proteins naturally interact, providing a higher antioxidant and anti-inflammatory capacity as a result.

The presence of antioxidant compounds is clearly a good starting point for the bio-prospecting approach. The research could start from accessions already studied and kept in germplasm banks, with the aim of breeding the most promising ones (e.g., more colourful, thicker coat or better nutrient profile) with domesticated landraces to create variants that are, at the same time, easy to cultivate but with the most interesting characteristics found from the available natural pool. In addition, this could lead to new experiments to understand better the synergic role of the phenolic fraction with bioactive proteins.

5.2. Anti-Tumor Compounds

Concerning the anti-cancer activity, Bowman–Birk inhibitors (BBI), present exclusively in the Fabaceae family and some cereals [143], have proven anticancer effects [144,145]. Panzeri et al. [146] demonstrated that aqueous extracts from boiled seeds containing BBI are, as expected, effective against some colorectal cancer cell lines, but the healthy line was not hit by the treatment. Mehdad et al. [147] proved its activity on breast cancer lines, and they were the first to discover a potential intracellular target, the proteasome 20S. Furthermore, they demonstrated cytostatic activity and increased apoptosis in cancer lines, but BBI was ineffective on the healthy mammary epithelial line. It is important to underline that this protein is kept by evolution due to its defensive role; in fact, it inhibits herbivores' digestion by interacting negatively with trypsin and chymotrypsin. Preliminary results obtained via the alignment of sequences downloaded from genebanks (NCBI) showed a high variability of BBI gene in some cowpea accessions, confirming the greatness of natural biodiversity. Unfortunately, little is known about the impact of domestication on the variability of the BBI gene. The domestication process can have acted as a strong constraint causing a bottleneck in the gene pool and reducing the variability of genes and exchange of alleles between cultivated and wild accessions. However, the exploration of haplotypes by sequencing several accessions is needed to verify the effective impact of human activity on gene diversity. Moreover, methods of ancestral sequence reconstruction (ASR) based on phylogenetic inference can predict the existence of stable, soluble, and active variants of proteins. The comparison of the structure of modern proteins with the corresponding ancestral intermediates can highlight functionally important substitutions within proteins and consequently drive the protein engineers to design variants that confer novel or more efficient activities (Figure 2). While different case studies are discussed in the literature where ancestral reconstructions were applied in eukaryotes, few instances are available in plants. Since ASR can be used to explore the remote evolutionary past as well as to investigate molecular evolution on shorter timescales, we argue that the proteins expressed in different genera of legumes are particularly well suited for ancestral reconstruction studies.

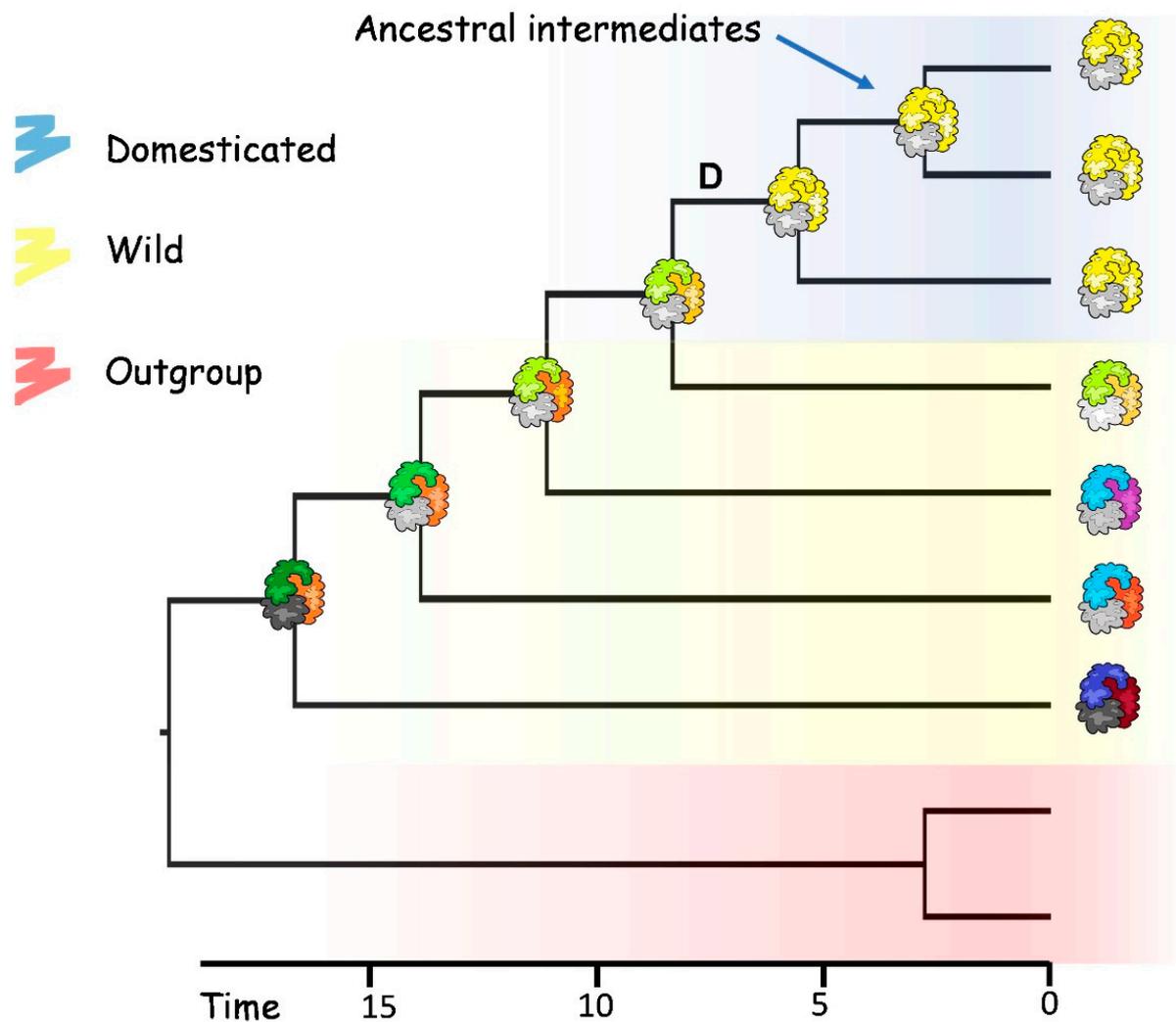


Figure 2. An example of reconstruction of putative ancestral intermediates by inferring the phylogenetic relationship between modern homologs. ASR studies can explore biodiversity to infer the historical evolution of natural proteins. Statistical models of amino acid substitution can be applied to calculate the sequences at internal nodes. Although domestication (D) has produced bottlenecks and reduced the genetic variability, ASR analyses can be applied to wild crop relatives.

Phenolic acids (e.g., gallic acid, ferulic acid, caffeic acid and chlorogenic acid) and flavonols (catechins, kaempferols and quercetins) are groups of molecules that are very active against cancer. Teixeira-Guedes et al. [148] found some of these molecules in the phenolic fraction of cowpea sprouts. Sprouting is an alternative method to consume food, especially seeds, grains and pulses. As a matter of fact, sprouting refers activating the metabolism of the dormant seeds and this way, complex reserve molecules are degraded into simpler ones, releasing other molecules and secondary metabolites [149]. The authors demonstrated at first the efficacy of the extracts against CRC cell lines; then combined it with 5-Fluorouracil (5-FU). This drug is potent but is susceptible to the occurrence of resistance by the tumour mass [150]. Among all these compounds, quercetin is one of the most common, was found to be the main representative of extracts and is well known to be active against different cancer lines [151–153].

The capacity to exert different kinds of bioactivities appoints phenolics and small proteins as very potential phyto complexes with an extreme wideness of possible applications. In this paragraph, the fact that extracts can be much more effective than single drugs is underlined. The use of a mixture of bioactive compounds in addition to the chosen drug could help in the treatment of many diseases.

5.3. Anti Hypercholesterolemic

One of the major world concerns is the role of the diet for healthy living. In particular, the main problem is malnutrition, 1.9 billion adults are overweight, and 452 million are underweight [154]. These numbers are going to increase during the next few years, so a healthy diet must become a worldwide topic. One way to prevent obesity is to find food or molecules that can lower LDL cholesterol concentration or production. Legumes are known to have a good nutritional profile and possess some interesting anti hypercholesterolemic capacities. For example, in the work of Tan et al. [155], *Vigna subterranea* was the object of study to create a powdered drink mix. The authors managed to characterise the extracts and proved the ability to lower the total cholesterol content in a population of rats. The observed effects were comparable to those given by the commercial drug simvastatin, demonstrating a potential commercial formulation usable in everyday life. In addition, Bambara powder fat content was lower than the soybean, while it had more proteins. Regarding *Vigna unguiculata*, Kanetro [156] studied the hypocholesterolemic feature of protein extracts from the sprouts. The tests were performed on rats that mimicked a diabetic condition. This kind of extract established the potential of *Vigna unguiculata* in fighting high cholesterol concentrations. *Vigna unguiculata* was also studied in rabbit models by Janeesh and Abraham [157]. Rabbits received a rich fraction polyphenols and flavonoids extracted from the leaves that showed antioxidant capacity, hypolipidemic and anti-atherogenic properties in ill animals. The road opened by the studies reported here is encouraging and already tending to practical applications usable worldwide by combining the natural nutritive features to bioactive compounds present in the seeds.

Although many important bioactivities are reported in this paragraph, the actual knowledge is still incomplete. Small proteins and polyphenols were objects of these studies, and their versatility in terms of the panel of bioactivities exerted was highlighted and valorised over and over. A topic that we would like to stress more and encourage research on is the variability of seed coat colours. In fact, human activity has selected a wide range of shapes, textures and pigments in coats (including eye shapes and sizes), allowing us to clearly distinguish seeds of domestication accessions from unattractive seeds of wild lineages. The seed colours are correlated to the presence of tannins and flavonoids [158,159], and phenolic profiles showed that seed coats contain up to 10 times more flavonoids if compared to whole seeds [160]. The seed coat pattern is a fundamental aspect of consumer preference, but in different regions, only some patterns are preferred. On the other hand, local landraces contain a great variability of colours, selected through centuries by human activity, but often this richness remains undervalued [135,137,161]. Our suggestion is to use this kind of information to correlate the colours of seed coats with the proper chemical characterisation regarding previously mentioned bioactivities. Moreover, modern experimental planes should include wild accessions/species and underused landraces because these mostly unexplored taxa could hide important micronutrients. Finally, we underline that the introduction of new dishes based on a mix of seeds that show different colours could be a new way to assimilate a great variety of nutrients into the diet.

6. Introgression and Feralisation Processes

Through the domestication process, one or more populations that showed desirable traits are selected by humans producing new independent lineages. Farmers have strongly influenced the survival of these cultivated lineages that continued to diverge from wild ancestors because they were affected by different selective pressures. However, crops and their wild relatives can exchange genetic information spontaneously or through human activity. Generally, wild relatives of legumes show undesirable traits, but their genomes can hide a precious gene pool that is mostly untapped that can be recovered and reintroduced in cultivated forms.

Introgression of useful genes remains a fundamental way to improve the cultivar [162], and successful crosses mediated by humans were acquired, especially in cowpea [163]. Although domesticated cowpea is known as an inbred crop, outcrossing is reported sug-

gesting that frequency and distance can vary depending on the environment, climate, subspecies, genotype and insect involved [73,164,165]. In *Vigna unguiculata*, spontaneous introgressive events between wild perennial subspecies of the *dekindtiana* group, including accessions of var. *spontanea*, are widely described observing different morphological traits [71,84,166–168]. Molecular analysis using AFLP [74] and internal transcribed spacers [53,169,170] have confirmed the natural propensity to hybridisation between subspecies and have revealed intricate intraspecific phylogenetic relationships. Intragenomic 5S rRNA repeat unit heterogeneity was interpreted as the consequence of extensive hybridisation events [170], and recently, plastid DNA sequences have confirmed chloroplast capture events [76].

In recent decades, several researchers have tried to produce introgressive lineages obtaining interesting results and showing that the most important gene pool for breeding programs could be harboured in wild subspecies. Intraspecific hybrids obtained crossing ssp. *unguiculata* with ssp. *pubescens* and cv. *sesquipedalis* with ssp. *tenuis* have shown vigorous growth and partial fertility [171–173]. Some authors attributed the incomplete success to chromosomal disturbances that ensue in endosperms and embryos during early seed development when crosses between wild perennial accessions and domesticated cowpeas are performed [174]. However, different accessions showed diverse propensity to hybridize, and a recent study suggests that temperature and humidity also have a prevalent role in increasing the success of hybridisation [175]. A wild lineage of cowpea (TVNu-1158) collected in the Republic of Congo showed resistance to *Aphis craccivora*, surviving long after infestation [176], and was successively crossed with cowpea to produce new lineages [177]. Moreover, resistance to *Maruca vitrata* was observed in the wild lineage of ssp. *dekindtiana* (Tvnu 863) from Zimbabwe and resistance to *Clavigralla tomentosicollis* was observed in ssp. *dekindtiana* (TVnu 151) from Ghana; however, literature about their use to produce new cultivars is missing [178,179].

Limited information is available about the intraspecific introgression of *V. subterranea* and *V. vexillata*. The success of the artificial cross of Bambara is constrained by scarce pollen viability, the small size of the flower and the reduced stigma–anther separation, which improves the transfer of pollen to the stigma but at the same time complicates the emasculation process [180–183]. However, F1 and F2 lines were obtained by crossing *Vigna subterranea* var. *spontanea* (Harms) Pasquet and *Vigna subterranea* var. *subterranea* (L.) Verdc. varieties, allowing us to identify that the main morphological traits to distinguish the two forms (internode length and stems per plant) are regulated by relatively limited numbers of genes [184]. Intraspecific introgression success was also obtained by James and Lawn [185] who crossed African and Australian accessions of *V. vexillata* with the aim to explain the resistance to mottle carmovirus (CPMoV). Recently, modern hybridisation techniques were applied to cross var. *macrosperma* cultivated and wild accessions obtaining encouraging results [186,187]. Unfortunately, scarce findings are achieved by interspecific hybridisation. Differently from Asian taxa, where the compatibility was confirmed in different studies, the African taxa show a cross incompatibility barrier that has so far prevented the introgression of useful genes (e.g., *V. vexillata* × *V. unguiculata*) [168,188–190].

In recent years, advances in sequencing technologies have allowed the generation of a large number of genomic resources that, if combined with approaches that estimate the rate of gene flow, enable us to detect which lineages are prone to hybridisation. Screening the level of introgression already existing in nature is an important opportunity that can help us to obtain advanced information useful in breeding activity. For example, natural hybrid zones harbour genetic variance and, pervasive and occasional introgressive events are identified in several crops such as kiwi, common bean, soybean, sunflower and grape [191–196]. Differently from neutral introgression, which could be lost during successions of generations, adaptive introgression events are maintained by selection, and foreign gene variants introduced by gene flow can increase the fitness of receiver populations as observed in potato, rice and millet [197–199].

African *Vigna* species have a potential for introgression that today remains mostly unexplored. *V. unguiculata* and *V. vexillata* show an elevated number of wild lineages that probably have diverged well before the Pleistocene due to climate changes [71,125]. Several subspecies are adapted to different environments, and Padulosi and Ng [68] proposed that the southernmost region of Africa is presumably the origin center for *V. unguiculata* where most subspecies grow, while Pasquet [71] indicated that some lineages from Namibia to Zimbabwe are the result of spontaneous introgression events. However, genomic studies are needed to confirm these hypotheses, including in the analyses of populations spread at the margins of species distribution that could hide local adaptation to extreme conditions. Principal component analysis, Bayesian clustering methods (e.g., NEWHYBRIDS, STRUCTURE and ADMIXTURE) and divergence statistics such as F_{ST} are used to explore patterns of divergence in *Vigna* species, but they manifest shortness to provide the effective migration rate. To overcome this limitation, different probabilistic approaches recently developed are able to identify recent and ancient signals of introgression such as tree-based methods (e.g., Treemix), coalescent-with-introgression simulations (e.g., MSci model implemented in BPP), composite-likelihood test (e.g., VolcanoFinder), site frequency spectrum to explicitly model migrations (e.g., ∂adi), gene genealogies (e.g., Twisst) and ABBA-BABA statistics [200–204]. Moreover, only some genomic regions could be involved in gene flow, and thus introgression might be localised in specific chromosomes [205]. Since alleles shared through incomplete lineage sorting remain complex to distinguish from alleles shared through introgression and none of the measures described above is without simplifying assumptions, we suggest that different methods should be applied to ascertain the origin of introgression.

Although introgression from wild to crops has important economic consequences and many attempts are made to understand the evolutionary dynamics, in recent years, attention to the gene flow from crop to wild is rapidly increasing. Introgression of domesticated alleles can stimulate the evolution of weeds or increase the risk of extinction of wild populations with dramatic evolutionary consequences, as demonstrated in several annual and perennial plants [206–211]. Moreover, under specific circumstances, the spread of ferals escaped from cultivation and adapted to wild environments can hardly be contained. Although several authors consider feralisation the opposite process of domestication, few population genomics studies show how these genetic changes occurred in plants [212]. Some authors show that multiple de-domestication events have occurred in rice, highlighting that some crops are exceptionally prone to feralisation [213,214]. The introgression process is probably improved when the wild forms grow along the road margins, villages and fields where domesticated forms are cultivated. To date, few studies have investigated the introgression effects on the wild populations of African *Vigna* species. Some researchers have proposed that alleles from cowpea may be incorporated into wild forms especially improved by their cohabitation, replacing the original alleles and making new lineages well adapted to wild environments [215]. A molecular study based on analysis of isozyme loci showed that outcrossing rates in West Africa range from 1% to 9.5%, confirming possibilities of gene flow from domesticated cowpea to var. *spontanea* [216]. The distinction between feral and truly wild lineages is ever more complicated because introgression produces fertile offspring and the small seed-size typical of wild forms is dominant to large seed size [76,217]. Moreover, var. *spontanea* is represented by both annual and perennial plants, and it is acknowledged that while the annual and inbred habit is an adaptive strategy in dry and warm environments (e.g., in warm tropical savannas), perennials tend to grow in mountainous regions where the environment is often cooler and wetter [76]. Annual inbred plants produce more seeds and show a competitive advantage on perennials when they are sympatric in environments. Although few data about introgression are available, we cannot exclude that perennial outcrossed subspecies can be fertilized by cowpea pollen, and consequently, domesticated alleles can be introgressed. Moreover, feralisation can involve adaptive changes in genes related to flowering timing, dormancy and metabolic pathways, which are also unknown. Therefore, several aspects of the feralisation process, including the

ability to spread domesticated alleles across long distances by seeds and pollen through mammals or birds and the predisposition to invade territories where perennial subspecies grow, should be further explored.

7. Domestication-Related Traits and *De Novo* Domestication

As described by Darwin [218], most plants subjected to intensive domestication have lost the ability to survive in the wild environment for more than a few generations. Traits selected by humans allow us to clearly distinguish a domesticated plant from its wild progenitor, and several studies were recently proposed to highlight the genes at the base of these changes. In recent years, modern genomic techniques were applied to *Vigna* germplasm, accelerating research activity and opening new avenues to identify domestication-related traits [33,161,219–221].

Among the main domesticated traits in legumes, the loss of pod shattering and increase of organ size are most relevant for breeding. In cowpea, two main quantitative trait loci (QTLs) were identified for pod shattering, whereas QTLs identified for seed weight, leaf length, leaf width and pod length were located in the same region, suggesting a potential pleiotropy that controls the organ size [177]. Lonardi et al., 2019 [222] managed to obtain the entire genome sequence in order to analyse and identify the eventual putative syntelog for organ gigantism. They found a region containing a cluster of QTLs controlling pod length, seed size, leaf length and leaf width (CPod18, CSw8, CL18, CLw8). Similar results were also observed in *V. vexillata* where the main domestication traits, including seed size, pod size and leaf size, were controlled only by one or a major QTL and some minor QTLs [33,90,124]. More complex is the control of seed dormancy, which is generally managed by water-impermeable layers of cells of the seed coat. In yardlong bean, a vegetable crop that has experienced divergent domestication from cowpea, six QTLs were detected for seed dormancy-related traits [90]. The seed coat pattern is an essential trait in cowpea, intensely selected by human preferences that change in the different areas of Africa. For example, pigmentation displays high variability of colours, including varied eye shapes and sizes. Recent studies show that the colour and position of the pigmentation can be defined by expression patterns, and some genes that encode for proteins involved in the flavonoid biosynthesis pathway were identified [161]. Moreover, phenotypic observations show that a lack of pigment in flowers is often correlated with a lack of pigment in the seed coat, and a gene was recently proposed to have a dual function in cowpea controlling the colour in both organs [177]. As observed for several species, the flower was involved intensely in the domestication process, and it has a fundamental trait that allows us to distinguish domesticated cowpea from their wild relatives. Recently, innovative studies focused on exploring the genetic basis of floral scent. A group of five O-methyltransferase genes involved in the biosynthesis of melatonin and located within the floral scent QTL region was identified [221]. Melatonin is recognized as an essential molecule in several plants used to interact with pollinators. Flowering timing undoubtedly plays a key role in plant adaptation and diffusion of crops because several agronomic traits such as grain quality, plant growth and plant height are directly influenced by this characteristic [223]. However, how the domestication process has affected the timing of flowering in legumes is unclear [224]. Flowering timing is a complex trait generally regulated by genetic networks. While in *Arabidopsis thaliana* L., the existence of up to 80 loci [225] was shown, in a cowpea genome-wide association study (GWAS) seven reliable SNPs were revealed that explained phenotypic variance [220]. Important agronomic implications are expected because the candidate genes could be transferred by hybridisation in crops. Early flowering accessions can mature earlier, avoiding periods of drought stress, whereas late-flowering accessions can mature later and extend the vegetative period, thereby increasing biomass production.

It is widely recognized that the study of domestication-related traits is a fundamental step that enables us to understand how to design ideal crops for the future. Throughout the process of domestication and successive breeding phases, the genetic diversity of crops was significantly reduced, and this homogeneity is becoming a serious threat. The increase

of disease and inability of adaptation to environmental changes that consequently cause an increased use of pesticides and water with a severe impact on the environment are the main issues that affect the sustainability of modern agriculture. Fernie and Yan [226] emphasized that wild species contain less deleterious allelic variants than their crops, and Smykal et al. [227], in a recent review, reported that modern cultivars have lower levels of key vitamins and micronutrients, suggesting that several wild and semi-wild African species should be *de novo* domesticated.

Unfortunately, few studies of re-domestication are available, but recent advances in gene editing combined with the decryption of pan-genomes are opening new perspectives of manipulation of genes for the creation of modern crops [228,229]. Gene editing is used to modify the function of genes already existing, incorporate new genes and delete short or large DNA fragments [230,231]. For instance, undesirable traits can be reduced or removed by intervening in genes that regulate the content of secondary metabolites, accelerating the process of domestication. Otherwise, the life cycle of cowpea could be shifted coming back from annual to perennial, as occurred for *Triticum aestivum* L. [232]. Perennial cowpea crop would show deep roots, higher water and nutrients efficiency and would not need to be sown every year.

Modern techniques such as CRISPR/Cas9 are applied successfully in several staple food crops. In *Oryza sativa* L., mutations on three yield-related genes have produced more and larger grains and erect panicles [233], whereas, in *Solanum pimpinellifolium* L., eight genes were targeted improving architectural traits, day-length insensitivity, the size and shape of fruits and content of vitamins [234,235]. Moreover, the CRISPR-Cas9 system was also used in cowpea to disrupt the symbiotic nitrogen fixation by the modification of a symbiosis receptor-like kinase (SYMRLK) gene, thereby demonstrating that gene editing can be applied to the *Vigna* genus [236]. However, this technique requires that the genome is sequenced to identify the ortholog gene that controls the domestication trait [237].

African *Vigna* species are an ideal group of plants on which to apply gene-editing techniques and to produce modern crops. A great number of wild species, besides showing resistance to pests and diseases and having high nutritional values, are well adapted to diverse environmental conditions [9]. Only Angola, with 28 native *Vigna* species documented, is recognized as one of the most important sources of germplasm in the world [238]. *V. monantha* occurs in permanently dry conditions [42], whereas *V. marina* and *V. luteola* grow well in saline lands [9]. In particular, seedlings of *V. marina* can survive for several weeks in flooded conditions and high NaCl concentration [39], accumulating high levels of salt in leaves, roots and stem [41].

However, few farmers currently use these plants because of low yield and strong pod-shattering behaviour, which requires high labour during the harvest. Adaptation to extreme environments often involves multiple genes, whereas domestication-related traits are due to mutations of a single locus that affects the loss of a function. Previously reported domestication-related traits in *Vigna* seem to be controlled by a restricted number of QTLs. Thus, introducing domestication-related mutations into wild species might be preferred rather than modifying multiple genes related to complex adaptation traits. For example, the first steps of re-domestication were achieved by Takahashi [4], starting from the accessions of *Vigna stipulacea* (Lam.) Kuntze originated in Asia. The authors obtained one mutant with reduced pod shattering and three mutants with reduced seed dormancy, characterizing the respective SNPs in the candidate genes. *V. stipulacea* was selected for their fast growth, edible seeds and broad resistance to pests and diseases. Thus, *de novo* crops can be designed to preserve several traits that nature has selected in millions of years. Moreover, in the next few years, the pan-genomes of several economically important crops will be available. The investigation by sequencing multiple individuals, including wild and domesticated accessions, will allow us to acquire full knowledge of variations at the genome level. Since it is widely accepted that the use of few reference genomes is limiting, the pan-genome of the *Vigna* species should be achieved in a short time [239,240]. Consequently, given the

large diversity of wild *Vigna* germplasm spread in Africa and the modern techniques of gene editing, great margins of genetic improvement are expected in the near future.

Author Contributions: Conceptualization, D.P., F.G., W.G.N. and M.L.; investigation, D.P. and F.G.; writing—original draft preparation, D.P. and F.G.; writing—review and editing, D.P., F.G., W.G.N. and M.L.; visualization, D.P.; supervision, F.G. and M.L. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Kahane, R.; Hodgkin, T.; Jaenicke, H.; Hoogendoorn, C.; Hermann, M.; d'Arros Hughes, J.; Padulosi, S.; Looney, N. Agrobiodiversity for Food Security, Health and Income. *Agron. Sustain. Dev.* **2013**, *33*, 671–693. [CrossRef]
- Hu, X.-R.; Chou, G.-X.; Zhang, C.-G. Flavonoids, Alkaloids from the Seeds of *Crotalaria Pallida* and Their Cytotoxicity and Anti-Inflammatory Activities. *Phytochemistry* **2017**, *143*, 64–71. [CrossRef]
- Lam, S.-H.; Li, Y.-C.; Kuo, P.-C.; Hwang, T.-L.; Yang, M.-L.; Wang, C.-C.; Tzen, J.T. Chemical Constituents of *Vigna luteola* and Their Anti-Inflammatory Bioactivity. *Molecules* **2019**, *24*, 1371. [CrossRef]
- Takahashi, Y.; Sakai, H.; Yoshitsu, Y.; Muto, C.; Anai, T.; Pandiyan, M.; Senthil, N.; Tomooka, N.; Naito, K. Domesticating *Vigna stipulacea*: A Potential Legume Crop with Broad Resistance to Biotic Stresses. *Front. Plant Sci.* **2019**, *10*, 1607. [CrossRef]
- Ku, Y.-S.; Contador, C.A.; Ng, M.-S.; Yu, J.; Chung, G.; Lam, H.-M. The Effects of Domestication on Secondary Metabolite Composition in Legumes. *Front. Genet.* **2020**, *11*, 581357. [CrossRef] [PubMed]
- Marconi, E.; Ruggeri, S.; Carnovale, E. Chemical Evaluation of Wild Under-Exploited *Vigna* spp. Seeds. *Food Chem.* **1997**, *59*, 203–212. [CrossRef]
- Duranti, M. Grain Legume Proteins and Nutraceutical Properties. *Fitoterapia* **2006**, *77*, 67–82. [CrossRef] [PubMed]
- Tomooka, N.; Naito, K.; Kaga, A.; Sakai, H.; Isemura, T.; Ogiso-Tanaka, E.; Iseki, K.; Takahashi, Y. Evolution, Domestication and Neo-Domestication of the Genus *Vigna*. *Plant Genet. Resour.* **2014**, *12*, S168–S171. [CrossRef]
- Harouna, D.V.; Venkataramana, P.B.; Ndakidemi, P.A.; Matemu, A.O. Under-Exploited Wild *Vigna* Species Potentials in Human and Animal Nutrition: A Review. *Glob. Food Secur.* **2018**, *18*, 1–11. [CrossRef]
- Boukar, O.; Belko, N.; Chamarthi, S.; Togola, A.; Batiemo, J.; Owusu, E.; Haruna, M.; Diallo, S.; Umar, M.L.; Olufajo, O. Cowpea (*Vigna unguiculata*): Genetics, Genomics and Breeding. *Plant Breed.* **2019**, *138*, 415–424. [CrossRef]
- Vigna Savi | Plants of the World Online | Kew Science. Available online: <http://powo.science.kew.org/taxon/urn:lsid:ipni.org:names:325971-2> (accessed on 17 January 2022).
- Maréchal, R. Etude Taxonomique d'un Groupe Complexe d'espèces Des Genres *Phaseolus* et *Vigna* (Papilionaceae) Sur La Base de Données Morphologiques et Polliniques, Traitées Par l'analyse Informatique. *Boissiera* **1978**, *28*, 1–273.
- Thulin, M.; Lavin, M.; Pasquet, R.; Delgado-Salinas, A. Phylogeny and Biogeography of *Wajira* (Leguminosae): A Monophyletic Segregate of *Vigna* Centered in the Horn of Africa Region. *Syst. Bot.* **2004**, *29*, 903–920. [CrossRef]
- Javadi, F.; Tun, Y.T.; Kawase, M.; Guan, K.; Yamaguchi, H. Molecular Phylogeny of the Subgenus *Ceratotropis* (Genus *Vigna*, Leguminosae) Reveals Three Eco-Geographical Groups and Late Pliocene–Pleistocene Diversification: Evidence from Four Plastid DNA Region Sequences. *Ann. Bot.* **2011**, *108*, 367–380. [CrossRef] [PubMed]
- Delgado-Salinas, A.; Thulin, M.; Pasquet, R.; Weeden, N.; Lavin, M. *Vigna* (Leguminosae) Sensu Lato: The Names and Identities of the American Segregate Genera. *Am. J. Bot.* **2011**, *98*, 1694–1715. [CrossRef] [PubMed]
- Lavin, M.; Herendeen, P.S.; Wojciechowski, M.F. Evolutionary Rates Analysis of Leguminosae Implicates a Rapid Diversification of Lineages during the Tertiary. *Syst. Biol.* **2005**, *54*, 575–594. [PubMed]
- Li, H.; Wang, W.; Lin, L.; Zhu, X.; Zhu, X.; Li, J.; Chen, Z. Diversification of the Phaseoloid Legumes: Effects of Climate Change, Range Expansion and Habit Shift. *Front. Plant Sci.* **2013**, *4*, 386. [CrossRef] [PubMed]
- Singh, B. *Cowpea: The Food Legume of the 21st Century*; John Wiley & Sons: Hoboken, NJ, USA, 2020; Volume 164.
- Maxted, N.; Mabuza-Diamini, P.; Moss, H.; Padulosi, S.; Jarvis, A.; Guarino, L. *An Ecogeographic Study African Vigna*; International Plant Genetic Resources Institute (IPGRI): Rome, Italy, 2004; ISBN 978-92-9043-637-9.
- Tateishi, Y. Systematics of the Species of *Vigna* Subgenus *Ceratotropis*. *Mungbean Germplasm Collect. Util. Breed. Program.* **1996**, *9–24*.
- Tomooka, N.; Maxted, N.; Thavarasook, C.; Jayasuriya, A.H.M. Two New Species, Sectional Designations and New Combinations in *Vigna* Subgenus *Ceratotropis* (Piper) Verdc. (Leguminosae, Phaseoleae). *Kew Bull.* **2002**, *57*, 613–624. [CrossRef]
- Tomooka, N.; Yoon, M.S.; Doi, K.; Kaga, A.; Vaughan, D. AFLP Analysis of Diploid Species in the Genus *Vigna* Subgenus *Ceratotropis*. *Genet. Resour. Crop. Evol.* **2002**, *49*, 521–530. [CrossRef]
- Pienaar, B.J. The *Vigna vexillata* Complex (Fabaceae) in Southern Africa. *South Afr. J. Bot.* **1991**, *57*, 236–245. [CrossRef]
- Ladizinsky, G. Seed Dispersal in Relation to the Domestication of Middle East Legumes. *Econ. Bot.* **1979**, *33*, 284–289. [CrossRef]

25. Takahashi, Y.; Kongjaimun, A.; Muto, C.; Kobayashi, Y.; Kumagai, M.; Sakai, H.; Satou, K.; Teruya, K.; Shiroma, A.; Shimoji, M. Same Locus for Non-Shattering Seed Pod in Two Independently Domesticated Legumes, *Vigna angularis* and *Vigna unguiculata*. *Front. Genet.* **2020**, *11*, 748. [[CrossRef](#)] [[PubMed](#)]
26. Roberts, J.A.; Elliott, K.A.; Gonzalez-Carranza, Z.H. Abscission, Dehiscence, and Other Cell Separation Processes. *Annu. Rev. Plant Biol.* **2002**, *53*, 131–158. [[CrossRef](#)] [[PubMed](#)]
27. Lo, S.; Parker, T.; Muñoz-Amatriaín, M.; Berny-Mier y Teran, J.C.; Jernstedt, J.; Close, T.J.; Gepts, P. Genetic, Anatomical, and Environmental Patterns Related to Pod Shattering Resistance in Domesticated Cowpea (*Vigna unguiculata* [L.] Walp.). *J. Exp. Bot.* **2021**, *72*, 6219–6229. [[CrossRef](#)] [[PubMed](#)]
28. Parker, T.A.; Berny Mier y Teran, J.C.; Palkovic, A.; Jernstedt, J.; Gepts, P. Pod Indehiscence Is a Domestication and Aridity Resilience Trait in Common Bean. *New Phytol.* **2020**, *225*, 558–570. [[CrossRef](#)]
29. Meyer, R.S.; Purugganan, M.D. Evolution of Crop Species: Genetics of Domestication and Diversification. *Nat. Rev. Genet.* **2013**, *14*, 840–852. [[CrossRef](#)]
30. Herniter, I.A.; Muñoz-Amatriaín, M.; Lo, S.; Guo, Y.-N.; Close, T.J. Identification of Candidate Genes Controlling Black Seed Coat and Pod Tip Color in Cowpea (*Vigna unguiculata* [L.] Walp.). *G3 Genes Genomes Genet.* **2018**, *8*, 3347–3355. [[CrossRef](#)]
31. Lo, S.; Muñoz-Amatriaín, M.; Hokin, S.A.; Cisse, N.; Roberts, P.A.; Farmer, A.D.; Xu, S.; Close, T.J. A Genome-Wide Association and Meta-Analysis Reveal Regions Associated with Seed Size in Cowpea [*Vigna unguiculata* (L.) Walp.]. *Theor. Appl. Genet.* **2019**, *132*, 3079–3087. [[CrossRef](#)]
32. Seo, E.; Kim, K.; Kang, R.; Kim, G.; Park, A.; Kim, W.J.; Sun, H.; Ha, B.-K. Genome-Wide Association Study for Flowering Time in Korean Cowpea Germplasm. *Plant Breed. Biotechnol.* **2020**, *8*, 413–425. [[CrossRef](#)]
33. Amkul, K.; Somta, P.; Laosatit, K.; Wang, L. Identification of QTLs for Domestication-Related Traits in Zombi Pea [*Vigna vexillata* (L.) A. Rich.], a Lost Crop of Africa. *Front. Genet.* **2020**, *11*, 803. [[CrossRef](#)]
34. Lu, J.; Tang, T.; Tang, H.; Huang, J.; Shi, S.; Wu, C.-I. The Accumulation of Deleterious Mutations in Rice Genomes: A Hypothesis on the Cost of Domestication. *Trends Genet.* **2006**, *22*, 126–131. [[CrossRef](#)] [[PubMed](#)]
35. Zhang, H.; Mittal, N.; Leamy, L.J.; Barazani, O.; Song, B.-H. Back into the Wild—Apply Untapped Genetic Diversity of Wild Relatives for Crop Improvement. *Evol. Appl.* **2017**, *10*, 5–24. [[CrossRef](#)] [[PubMed](#)]
36. Liu, Q.; Zhou, Y.; Morrell, P.L.; Gaut, B.S. Deleterious Variants in Asian Rice and the Potential Cost of Domestication. *Mol. Biol. Evol.* **2017**, *34*, 908–924. [[CrossRef](#)] [[PubMed](#)]
37. Yokoyama, T.; Tomooka, N.; Okabayashi, M.; Kaga, A.; Boonkerd, N.; Vaughan, D.A. Variation in the Nod Gene RFLPs, Nucleotide Sequences of 16S RRNA Genes, Nod Factors, and Nodulation Abilities of Bradyrhizobium Strains Isolated from Thai *Vigna* Plants. *Can. J. Microbiol.* **2006**, *52*, 31–46. [[CrossRef](#)] [[PubMed](#)]
38. Chankaew, S.; Isemura, T.; Naito, K.; Ogiso-Tanaka, E.; Tomooka, N.; Somta, P.; Kaga, A.; Vaughan, D.A.; Srinives, P. QTL Mapping for Salt Tolerance and Domestication-Related Traits in *Vigna marina* subsp. *oblonga*, a Halophytic Species. *Theor. Appl. Genet.* **2014**, *127*, 691–702. [[CrossRef](#)]
39. Tomooka, N.; Kaga, A.; Isemura, T.; Vaughan, D.V. *Wild Crop Relatives: Genomic and Breeding Resources: Legume Crops and Forages*; Kole, C., Ed.; Springer: Berlin/Heidelberg, Germany, 2011.
40. Takahashi, Y.; Somta, P.; Muto, C.; Iseki, K.; Naito, K.; Pandiyan, M.; Natesan, S.; Tomooka, N. Novel Genetic Resources in the Genus *Vigna* Unveiled from Gene Bank Accessions. *PLoS ONE* **2016**, *11*, e0147568.
41. Yoshida, J.; Tomooka, N.; Yee Khaing, T.; Shantha, P.S.; Naito, H.; Matsuda, Y.; Ehara, H. Unique Responses of Three Highly Salt-Tolerant Wild *Vigna* Species against Salt Stress. *Plant Prod. Sci.* **2020**, *23*, 114–128. [[CrossRef](#)]
42. van Zonneveld, M.; Rakha, M.; Tan, S.y.; Chou, Y.-Y.; Chang, C.-H.; Yen, J.-Y.; Schafleitner, R.; Nair, R.; Naito, K.; Solberg, S.Ø. Mapping Patterns of Abiotic and Biotic Stress Resilience Uncovers Conservation Gaps and Breeding Potential of *Vigna* Wild Relatives. *Sci. Rep.* **2020**, *10*, 1–11.
43. Carvalho, M.d.; Halecki, W. Modeling of Cowpea (*Vigna unguiculata*) Yield and Control Insecticide Exposure in a Semi-Arid Region. *Plants* **2021**, *10*, 1074. [[CrossRef](#)]
44. Peña-Valdivia, C.B.; García-Nava, J.R.; Aguirre R, J.R.; Ybarra-Moncada, M.C.; López H, M. Variation in Physical and Chemical Characteristics of Common Bean (*Phaseolus vulgaris* L.) Grain along a Domestication Gradient. *Chem. Biodivers.* **2011**, *8*, 2211–2225. [[CrossRef](#)] [[PubMed](#)]
45. Fernández-Marín, B.; Milla, R.; Martín-Robles, N.; Arc, E.; Kranner, I.; Becerril, J.M.; García-Plazaola, J.I. Side-Effects of Domestication: Cultivated Legume Seeds Contain Similar Tocopherols and Fatty Acids but Less Carotenoids than Their Wild Counterparts. *BMC Plant Biol.* **2014**, *14*, 1–11. [[CrossRef](#)] [[PubMed](#)]
46. Larson, G.; Piperno, D.R.; Allaby, R.G.; Purugganan, M.D.; Andersson, L.; Arroyo-Kalin, M.; Barton, L.; Vigueira, C.C.; Denham, T.; Dobney, K. Current Perspectives and the Future of Domestication Studies. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, 6139–6146. [[CrossRef](#)] [[PubMed](#)]
47. D’Andrea, A.C.; Kahlheber, S.; Logan, A.L.; Watson, D.J. Early Domesticated Cowpea (*Vigna unguiculata*) from Central Ghana. *Antiquity* **2007**, *81*, 686–698. [[CrossRef](#)]
48. Allaby, R.G.; Fuller, D.Q.; Brown, T.A. The Genetic Expectations of a Protracted Model for the Origins of Domesticated Crops. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 13982–13986. [[CrossRef](#)] [[PubMed](#)]
49. Purugganan, M.D.; Fuller, D.Q. Archaeological Data Reveal Slow Rates of Evolution during Plant Domestication. *Evol. Int. J. Org. Evol.* **2011**, *65*, 171–183. [[CrossRef](#)] [[PubMed](#)]

50. Engels, J.M.M.; Ebert, A.W.; Thormann, I.; De Vicente, M.C. Centres of Crop Diversity and/or Origin, Genetically Modified Crops and Implications for Plant Genetic Resources Conservation. *Genet. Resour. Crop. Evol.* **2006**, *53*, 1675–1688. [[CrossRef](#)]
51. Huynh, B.; Close, T.J.; Roberts, P.A.; Hu, Z.; Wanamaker, S.; Lucas, M.R.; Chiulele, R.; Cissé, N.; David, A.; Hearne, S. Gene Pools and the Genetic Architecture of Domesticated Cowpea. *Plant Genome* **2013**, *6*, 1–8. [[CrossRef](#)]
52. Herniter, I.A.; Muñoz-Amatriáin, M.; Close, T.J. Genetic, Textual, and Archeological Evidence of the Historical Global Spread of Cowpea (*Vigna unguiculata* [L.] Walp.). *Legume Sci.* **2020**, *2*, e57. [[CrossRef](#)]
53. Vijaykumar, A.; Saini, A.; Jawali, N. Assessment of Hybridization among Wild and Cultivated *Vigna unguiculata* Subspecies Revealed by Arbitrarily Primed Polymerase Chain Reaction Analysis. *AoB Plants* **2012**, *2012*, pls012. [[CrossRef](#)]
54. Small, E. *Top 100 Food Plants: The World's Most Important Culinary Crops*; NRC Research Press: Ottawa, ON, Canada, 2009.
55. Timko, M.P.; Ehlers, J.D.; Roberts, P.A. Cowpea. In *Pulses, Sugar and Tuber Crops*; Springer: Berlin/Heidelberg, Germany, 2007; pp. 49–67.
56. FAOSTAT. Available online: <https://www.fao.org/faostat/en/#home> (accessed on 17 January 2022).
57. Smith, M.R.; Veneklaas, E.; Polania, J.; Rao, I.M.; Beebe, S.E.; Merchant, A. Field Drought Conditions Impact Yield but Not Nutritional Quality of the Seed in Common Bean (*Phaseolus vulgaris* L.). *PLoS ONE* **2019**, *14*, e0217099. [[CrossRef](#)]
58. Adjei-Nsiah, S.; Kuyper, T.W.; Leeuwis, C.; Abekoe, M.K.; Cobbinah, J.; Sakyi-Dawson, O.; Giller, K.E. Farmers' Agronomic and Social Evaluation of Productivity, Yield and N₂-Fixation in Different Cowpea Varieties and Their Subsequent Residual N Effects on a Succeeding Maize Crop. *Nutr. Cycl. Agroecosyst.* **2008**, *80*, 199–209. [[CrossRef](#)]
59. Chikowo, R.; Mapfumo, P.; Nyamugafata, P.; Giller, K.E. Woody Legume Fallow Productivity, Biological N₂-Fixation and Residual Benefits to Two Successive Maize Crops in Zimbabwe. *Plant Soil* **2004**, *262*, 303–315. [[CrossRef](#)]
60. Singh, B.B.; Sharma, B. Restructuring Cowpea for Higher Yield. *Indian J. Genet.* **1996**, *56*, 389–405.
61. Boukar, O.; Fatokun, C.A.; Huynh, B.-L.; Roberts, P.A.; Close, T.J. Genomic Tools in Cowpea Breeding Programs: Status and Perspectives. *Front. Plant Sci.* **2016**, *7*, 757. [[CrossRef](#)]
62. Horn, L.; Shimelis, H.; Laing, M. Participatory Appraisal of Production Constraints, Preferred Traits and Farming System of Cowpea in the Northern Namibia: Implications for Breeding. *Legume Res. Int. J.* **2015**, *38*, 691–700. [[CrossRef](#)]
63. Boukar, O.; Massawe, F.; Muranaka, S.; Franco, J.; Maziya-Dixon, B.; Singh, B.; Fatokun, C. Evaluation of Cowpea Germplasm Lines for Protein and Mineral Concentrations in Grains. *Plant Genet. Resour.* **2011**, *9*, 515–522. [[CrossRef](#)]
64. Abadassi, J. Cowpea (*Vigna unguiculata* (L.) Walp.) Agronomic Traits Needed in Tropical Zone. *Int. J. Pure Appl. Biosci.* **2015**, *3*, 158–165.
65. Dakora, F.D.; Belane, A.K. Evaluation of Protein and Micronutrient Levels in Edible Cowpea (*Vigna unguiculata* L. Walp.) Leaves and Seeds. *Front. Sustain. Food Syst.* **2019**, *3*, 70. [[CrossRef](#)]
66. Weng, Y.; Shi, A.; Ravelombola, W.S.; Yang, W.; Qin, J.; Motes, D.; Moseley, D.O.; Chen, P. A Rapid Method for Measuring Seed Protein Content in Cowpea (*Vigna unguiculata* (L.) Walp.). *Am. J. Plant Sci.* **2017**, *8*, 2387. [[CrossRef](#)]
67. Zuluaga, D.L.; Lioi, L.; Delvento, C.; Pavan, S.; Sonnante, G. Genotyping-by-Sequencing in *Vigna unguiculata* Landraces and Its Utility for Assessing Taxonomic Relationships. *Plants* **2021**, *10*, 509. [[CrossRef](#)]
68. Padulosi, S.; Ng, N.Q. Origin, Taxonomy, and Morphology of *Vigna unguiculata* (L.) Walp. *Adv. Cowpea Res.* **1997**, 1–12.
69. Padulosi, S. Genetic Diversity, Taxonomy and Ecogeographic Survey of the Wild Relatives of Cowpea (*Vigna unguiculata* (L.) Walpers). Ph.D. Thesis, University of Louvain La Neuve, Ottignies-Louvain-la-Neuve, Belgium, 1993.
70. Pasquet, R.S. Classification Infraspécifique des Formes Spontanées de *Vigna unguiculata* (L.) Walp. (Fabaceae) à Partir de Données Morphologiques. *Bull. Du Jard. Bot. Natl. Belg. Bull. Natl. Plantentuin Van Belg.* **1993**, *62*, 127–173. [[CrossRef](#)]
71. Pasquet, R.S. Wild Cowpea (*Vigna unguiculata*) Evolution. *Adv. Legume Syst.* **1996**, *8*, 95–100.
72. Pasquet, R. A New Subspecies of *Vigna unguiculata* (Leguminosae: Papilionoideae). *Kew Bull.* **1997**, *52*, 840. [[CrossRef](#)]
73. Pasquet, R.S. Genetic Relationships among Subspecies of *Vigna unguiculata* (L.) Walp. Based on Allozyme Variation. *Theor. Appl. Genet.* **1999**, *98*, 1104–1119. [[CrossRef](#)]
74. Coulibaly, S.; Pasquet, R.S.; Papa, R.; Gepts, P. AFLP Analysis of the Phenetic Organization and Genetic Diversity of *Vigna unguiculata* L. Walp. Reveals Extensive Gene Flow between Wild and Domesticated Types. *Theor. Appl. Genet.* **2002**, *104*, 358–366. [[CrossRef](#)]
75. Pasquet, R. Genus *Vigna* and Cowpea (*V. unguiculata* [L.] Walp.) Taxonomy: Current Status and Prospects. In Proceedings of the Fifth World Cowpea Conference on Improving Livelihoods in the Cowpea Value Chain through Advancement in Science, Dakar, Senegal, 27 September–1 October 2010.
76. Pasquet, R.S.; Feleke, Y.; Gepts, P. Cowpea [*Vigna unguiculata* (L.) Walp.] Maternal Lineages, Chloroplast Captures, and Wild Cowpea Evolution. *Genet. Resour. Crop. Evol.* **2021**, *68*, 2799–2812. [[CrossRef](#)]
77. Lush, W.M.; Evans, L.T.; Wien, H.C. Environmental Adaptation of Wild and Domesticated Cowpeas (*Vigna unguiculata* (L.) Walp.). *Field Crop. Res.* **1980**, *3*, 173–187. [[CrossRef](#)]
78. Faris, D.G. The Origin and Evolution of the Cultivated Forms of *Vigna sinensis*. *Can. J. Genet. Cytol.* **1965**, *7*, 433–452. [[CrossRef](#)]
79. Vaillancourt, R.E.; Weeden, N.F. Chloroplast DNA Polymorphism Suggests Nigerian Center of Domestication for the Cowpea, *Vigna unguiculata* (Leguminosae). *Am. J. Bot.* **1992**, *79*, 1194–1199. [[CrossRef](#)]
80. Lush, W.M.; Evans, L.T. The Domestication and Improvement of Cowpeas (*Vigna unguiculata* (L.) W Alp.). *Euphytica* **1981**, *30*, 579–587. [[CrossRef](#)]

81. Champion, L.; Fuller, D.Q.; Ozainne, S.; Huysecom, É.; Mayor, A. Agricultural Diversification in West Africa: An Archaeobotanical Study of the Site of Sadia (Dogon Country, Mali). *Archaeol. Anthropol. Sci.* **2021**, *13*, 60. [[CrossRef](#)] [[PubMed](#)]
82. Vavilov, N.I. Centers of Origin of Cultivated Plants. In *Origin and Geography of Cultivated Plants*; Cambridge University Press: Cambridge, UK, 1926.
83. Faris, D.G. *Evidence for the West African Origin of Vigna sinensis (L.) Savi*; University of California: Davis, CA, USA, 1963.
84. Rawal, K.M. Natural Hybridization among Wild, Weedy and Cultivated *Vigna unguiculata* (L.) Walp. *Euphytica* **1975**, *24*, 699–707. [[CrossRef](#)]
85. Ba, F.S.; Pasquet, R.S.; Gepts, P. Genetic Diversity in Cowpea [*Vigna unguiculata* (L.) Walp.] as Revealed by RAPD Markers. *Genet. Resour. Crop. Evol.* **2004**, *51*, 539–550. [[CrossRef](#)]
86. Fang, J.; Chao, C.-C.T.; Roberts, P.A.; Ehlers, J.D. Genetic Diversity of Cowpea [*Vigna unguiculata* (L.) Walp.] in Four West African and USA Breeding Programs as Determined by AFLP Analysis. *Genet. Resour. Crop. Evol.* **2007**, *54*, 1197–1209. [[CrossRef](#)]
87. Xiong, H.; Shi, A.; Mou, B.; Qin, J.; Motes, D.; Lu, W.; Ma, J.; Weng, Y.; Yang, W.; Wu, D. Genetic Diversity and Population Structure of Cowpea (*Vigna unguiculata* L. Walp.). *PLoS ONE* **2016**, *11*, e0160941. [[CrossRef](#)]
88. Muñoz-Amatriain, M.; Lo, S.; Herniter, I.; Boukar, O.; Fatokun, C.; Carvalho, E.; Castro, I.; Guo, Y.; Huynh, B.; Roberts, P.A. The UCR Minicore: A Valuable Resource for Cowpea Research and Breeding. *Legume Sci.* **2021**, *3*, e95. [[CrossRef](#)]
89. Sarr, A.; Bodian, A.; Gbedevi, K.M.; Ndir, K.N.; Ajewole, O.O.; Gueye, B.; Foncéka, D.; Diop, E.A.; Diop, B.M.; Cissé, N. Genetic Diversity and Population Structure Analyses of Wild Relatives and Cultivated Cowpea (*Vigna unguiculata* (L.) Walp.) from Senegal Using Simple Sequence Repeat Markers. *Plant Mol. Biol. Rep.* **2021**, *39*, 112–124. [[CrossRef](#)]
90. Kongjaimun, A.; Kaga, A.; Tomooka, N.; Somta, P.; Vaughan, D.A.; Srinives, P. The Genetics of Domestication of Yardlong Bean, *Vigna unguiculata* (L.) Walp. ssp. *unguiculata* Cv.-Gr. *sesquipedalis*. *Ann. Bot.* **2012**, *109*, 1185–1200. [[CrossRef](#)]
91. Xu, P.; Wu, X.; Wang, B.; Luo, J.; Liu, Y.; Ehlers, J.D.; Close, T.J.; Roberts, P.A.; Lu, Z.; Wang, S. Genome Wide Linkage Disequilibrium in Chinese Asparagus Bean (*Vigna unguiculata* ssp. *sesquipedialis*) Germplasm: Implications for Domestication History and Genome Wide Association Studies. *Heredity* **2012**, *109*, 34–40. [[CrossRef](#)]
92. Xu, P.; Wu, X.; Wang, B.; Liu, Y.; Qin, D.; Ehlers, J.D.; Close, T.J.; Hu, T.; Lu, Z.; Li, G. Development and Polymorphism of *Vigna unguiculata* ssp. *unguiculata* Microsatellite Markers Used for Phylogenetic Analysis in Asparagus Bean (*Vigna unguiculata* ssp. *sesquipedialis* (L.) Verdc.). *Mol. Breed.* **2010**, *25*, 675–684. [[CrossRef](#)]
93. Pasquet, R.S. Morphological Study of Cultivated Cowpea *Vigna unguiculata* (L.) Walp. Importance of Ovule Number and Definition of Cv Gr *Melanophthalmus*. *Agronomie* **1998**, *18*, 61–70. [[CrossRef](#)]
94. Pasquet, R.S. Allozyme Diversity of Cultivated Cowpea *Vigna unguiculata* (L.) Walp. *Theor. Appl. Genet.* **2000**, *101*, 211–219. [[CrossRef](#)]
95. Kongjaimun, A.; Kaga, A.; Tomooka, N.; Somta, P.; Shimizu, T.; Shu, Y.; Isemura, T.; Vaughan, D.A.; Srinives, P. An SSR-Based Linkage Map of Yardlong Bean (*Vigna unguiculata* (L.) Walp. subsp. *unguiculata* Sesquipedalis Group) and QTL Analysis of Pod Length. *Genome* **2012**, *55*, 81–92. [[CrossRef](#)] [[PubMed](#)]
96. Mubaiwa, J.; Fogliano, V.; Chidewe, C.; Linnemann, A.R. Hard-to-Cook Phenomenon in Bambara Groundnut (*Vigna subterranea* (L.) Verdc.) Processing: Options to Improve Its Role in Providing Food Security. *Food Rev. Int.* **2017**, *33*, 167–194. [[CrossRef](#)]
97. Aliyu, S.; Massawe, F.; Mayes, S. Genetic Diversity and Population Structure of Bambara Groundnut (*Vigna subterranea* (L.) Verdc.): Synopsis of the Past Two Decades of Analysis and Implications for Crop Improvement Programmes. *Genet. Resour. Crop. Evol.* **2016**, *63*, 925–943. [[CrossRef](#)]
98. Olayide, O.E.; Donkoh, S.A.; Ansah, I.G.K.; Adzawla, W.; O'Reilly, P.J.; Mayes, S.; Feldman, A.; Halimi, R.A.; Nyarko, G.; Ilori, C.O. *Assessing Socioeconomic Factors Influencing Production and Commercialization of Bambara Groundnut as an Indigenous Climate Resilient Crop in Nigeria*; Springer Nature: Cham, Switzerland, 2018.
99. Azam-Ali, S.; Azam-Ali, S.N.; Aguilar-Manjarrez, J.; Bannayan-Avval, M. *A Global Mapping System for Bambara Groundnut Production*; Food and Agriculture Organization: Rome, Italy, 2001; Volume 1.
100. Pasquet, R.S.; Schwedes, S.; Gepts, P. Isozyme Diversity in Bambara Groundnut. *Crop. Sci.* **1999**, *39*, 1228–1236. [[CrossRef](#)]
101. Doku, E.V.; Karikari, S.K. Operational Selection in Wild Bambara Groundnuts. *Ghana J. Sci.* **1971**, *11*, 45–56.
102. Majola, N.G.; Gerrano, A.S.; Shimelis, H. Bambara Groundnut (*Vigna subterranea* [L.] Verdc.) Production, Utilisation and Genetic Improvement in Sub-Saharan Africa. *Agronomy* **2021**, *11*, 1345. [[CrossRef](#)]
103. Mohammed, S.M. Pre-Breeding of Bambara Groundnut (*Vigna subterranea* [L.] Verdc.). Ph.D. Thesis, Abubakar Tafawa Balewa University, Bauchi, Nigeria, 2014.
104. Dalziel, J.M. *The Useful Plants of West Tropical Africa*; Royal Botanic Gardens: London, UK, 1937.
105. Begemann, F. *Ecogeographic Differentiation of Bambara Groundnut (Vigna subterranea) in the Collection of the International Institute of Tropical Agriculture (IITA)*; Wissenschaftlicher Fachverlag; Wissenschaftlicher Fachverlag: Berlin, Germany, 1988.
106. Goli, A.E.; Begemann, F.; Ng, N.Q. *Characterisation and Evaluation of IITA'S Bambara Groundnut (Vigna subterranea (L.) Verdc.) Promoting the Conservation and Use of Underutilized and Neglected Crops*; International Plant Genetic Resources Institute: Rome, Italy, 1997; Volume 9, pp. 101–118.
107. Somta, P.; Chankaew, S.; Rungnoi, O.; Srinives, P. Genetic Diversity of the Bambara Groundnut (*Vigna subterranea* (L.) Verdc.) as Assessed by SSR Markers. *Genome* **2011**, *54*, 898–910. [[CrossRef](#)]

108. Olukolu, B.A.; Mayes, S.; Stadler, F.; Ng, N.Q.; Fawole, I.; Dominique, D.; Azam-Ali, S.N.; Abbott, A.G.; Kole, C. Genetic Diversity in Bambara Groundnut (*Vigna subterranea* (L.) Verdc.) as Revealed by Phenotypic Descriptors and DArT Marker Analysis. *Genet. Resour. Crop. Evol.* **2012**, *59*, 347–358. [[CrossRef](#)]
109. Molosiwa, O.O.; Aliyu, S.; Stadler, F.; Mayes, K.; Massawe, F.; Kilian, A.; Mayes, S. SSR Marker Development, Genetic Diversity and Population Structure Analysis of Bambara Groundnut [*Vigna subterranea* (L.) Verdc.] Landraces. *Genet. Resour. Crop. Evol.* **2015**, *62*, 1225–1243. [[CrossRef](#)]
110. Tanimu, B.; Aliyu, L. Bambara Groundnut, *Vigna subterranea* (L.) verdc. In Proceedings of the Workshop on Conservation and Improvement of Bambara Groundnut, Harare, Zimbabwe, 14–16 November 1995; Country Report: Northern Nigeria; Promoting the Conservation and Use of Underutilized and Neglected Crops. International Plant Genetic Resources Institute: Rome, Italy, 1997.
111. Muhammad, I.; Rafii, M.Y.; Ramlee, S.I.; Nazli, M.H.; Harun, A.R.; Oladosu, Y.; Musa, I.; Arolu, F.; Chukwu, S.C.; Sani Haliru, B. Exploration of Bambara Groundnut (*Vigna subterranea* (L.) Verdc.), an Underutilized Crop, to Aid Global Food Security: Varietal Improvement, Genetic Diversity and Processing. *Agronomy* **2020**, *10*, 766. [[CrossRef](#)]
112. Molosiwa, O.; Basu, S.M.; Stadler, F.; Azam-Ali, S.; Mayes, S. Assessment of Genetic Variability of Bambara Groundnut (*Vigna subterranea* (L.) Verdc.) Accessions Using Morphological Traits and Molecular Markers. In Proceedings of the II International Symposium on Underutilized Plant Species: Crops for the Future-Beyond Food Security 979, Kuala Lumpur, Malaysia, 27 June–1 July 2011; pp. 779–790.
113. Molosiwa, O.O. Genetic Diversity and Population Structure Analysis of Bambara Groundnuts (*Vigna subterranea* (L.) Verdc.) Landraces Using Morpho-Agronomic Characters and SSR Markers. Ph.D. Thesis, University of Nottingham, Nottingham, UK, 2012.
114. Verdcourt, B. Studies in the Leguminosae-Papilionoideae for the 'Flora of Tropical East Africa': III. *Kew Bull.* **1970**, *24*, 379–447. [[CrossRef](#)]
115. Vanderborght, T. Some Observations on Seedlings of *Vigna vexillata* (L.) A. Rich. (Fabaceae). *Bull. Du Jard. Bot. Natl. Belg. Bull. Natl. Plantentuin Belg.* **1989**, *59*, 179–187. [[CrossRef](#)]
116. Damayanti, F.; Lawn, R.J.; Bielig, L.M. Genotypic Variation in Domesticated and Wild Accessions of the Tropical Tuberos Legume *Vigna vexillata* (L.) A. Rich. *Crop. Pasture Sci.* **2010**, *61*, 771–784. [[CrossRef](#)]
117. Dachapak, S.; Somta, P.; Poonchaivilaisak, S.; Yimram, T.; Srinives, P. Genetic Diversity and Structure of the Zombi Pea (*Vigna vexillata* (L.) A. Rich.) Gene Pool Based on SSR Marker Analysis. *Genetica* **2017**, *145*, 189–200. [[CrossRef](#)]
118. Cosmas, P.; Agathar, K.; Ronald, M.; John, C.T.; Simon, M. Preliminary Evaluation of Different Seed Dormancy Breaking Methods In Wild Tuber Cowpea (*Vigna vexillata*). *Can. J. Agric. Crop.* **2019**, *4*, 33–40. [[CrossRef](#)]
119. Tripathi, K.; Gore, P.G.; Pandey, A.; Nayar, E.R.; Gayacharan, C.; Pamarthi, R.K.; Bhardwaj, R.; Kumar, A. Morphological and Nutritional Assessment of *Vigna Vexillata* (L.) A. Rich.: A Potential Tuberos Legume of India. *Genet. Resour. Crop. Evol.* **2021**, *68*, 397–408. [[CrossRef](#)]
120. Ferguson, H. *The Food Crops of the Sudan and Their Relationship to Environment*; McCorquodale & Co., Ltd.: London, UK, 1954.
121. Bhattacharyya, P.K.; Ghosh, A.K.; Sanyal, B.; Deb Ray, G. Grow *Vigna vexillata* for Protein-Rich Tuber-Cum-Pulse Crop in North-Eastern Hill Region. *Seeds Farms* **1984**, *10*, 33–36.
122. Wong, K.C. *Vigna vexillata* (L.) A. Richard. PROSEA, Plant Resources of South-East Asia. *Aux. Plants* **1997**, *11*, 261–263.
123. Asati, B.S.; Yadav, D.S. Diversity of Horticultural Crops in North Eastern Region. *ENVIS Bull. Himal. Ecol.* **2004**, *12*, 1.
124. Dachapak, S.; Tomooka, N.; Somta, P.; Naito, K.; Kaga, A.; Srinives, P. QTL Analysis of Domestication Syndrome in Zombi Pea (*Vigna vexillata*), an Underutilized Legume Crop. *PLoS ONE* **2018**, *13*, e0200116. [[CrossRef](#)] [[PubMed](#)]
125. Garba, M.; Pasquet, R.S. Isozyme Diversity in *Vigna vexillata* (L.) A. Rich. (Fabaceae) Complex. *S. Afr. J. Bot.* **1998**, *64*, 163–175. [[CrossRef](#)]
126. Lawn, R.J.; Watkinson, A.R. Habitats, Morphological Diversity, and Distribution of the Genus *Vigna* Savi in Australia. *Aust. J. Agric. Res.* **2002**, *53*, 1305–1316. [[CrossRef](#)]
127. Karuniawan, A.; Iswandi, A.; Kale, P.R.; Heinzemann, J.; Grüneberg, W.J. *Vigna vexillata* (L.) A. Rich. Cultivated as a Root Crop in Bali and Timor. *Genet. Resour. Crop. Evol.* **2006**, *53*, 213–217. [[CrossRef](#)]
128. Butsayawarapat, P.; Juntawong, P.; Khamsuk, O.; Somta, P. Comparative Transcriptome Analysis of Waterlogging-Sensitive and Tolerant Zombi Pea (*Vigna vexillata*) Reveals Energy Conservation and Root Plasticity Controlling Waterlogging Tolerance. *Plants* **2019**, *8*, 264. [[CrossRef](#)]
129. Chiang, H.S.; Singh, S.R. Pod Hairs as a Factor in *Vigna vexillata* Resistance to the Pod-Sucking Bug, *Clavigralla tomentosicollis*. *Entomol. Exp. Et Appl.* **1988**, *47*, 195–199. [[CrossRef](#)]
130. Jackai, L.E.N.; Oghiakhe, S. Pod Wall Trichomes and Resistance of Two Wild Cowpea, *Vigna vexillata*, Accessions to *Maruca testualis* (Geyer) (Lepidoptera: Pyralidae) and *Clavigralla tomentosicollis* Stål (Hemiptera: Coreidae). *Bull. Entomol. Res.* **1989**, *79*, 595–605. [[CrossRef](#)]
131. Thottappilly, G.; Ng, N.Q.; Rossel, H.W. Screening Germplasm of *Vigna vexillata* for Resistance to Cowpea Mottle Virus. *Int. J. Trop. Plant Dis.* **1994**, *12*, 75–80.
132. Oguniwin, E.A.; Thottappilly, G.; AkenOva, M.E.; Ekpo, E.J.A.; Fatokun, C.A. Resistance to Cowpea Mottle Carmovirus in *Vigna vexillata*. *Plant Breed.* **2002**, *121*, 517–520. [[CrossRef](#)]

133. Amkul, K.; Wang, L.; Somta, P.; Wang, S.; Cheng, X. Construction of a High Density Linkage Map and Genome Dissection of Bruchid Resistance in Zombi Pea (*Vigna vexillata* (L.) A. Rich.). *Sci. Rep.* **2019**, *9*, 1–10. [CrossRef] [PubMed]
134. Sodedji, F.A.K.; Agbahoungba, S.; Nguetta, S.-P.A.; Agoyi, E.E.; Ayanan, M.A.T.; Sossou, S.H.; Mamadou, C.; Assogbadjo, A.E.; Kone, D. Resistance to Legume Pod Borer (*Maruca vitrata* Fabricius) in Cowpea: Genetic Advances, Challenges, and Future Prospects. *J. Crop. Improv.* **2020**, *34*, 238–267. [CrossRef]
135. Ojwang, L.O.; Dykes, L.; Awika, J.M. Ultra Performance Liquid Chromatography–Tandem Quadrupole Mass Spectrometry Profiling of Anthocyanins and Flavonols in Cowpea (*Vigna unguiculata*) of Varying Genotypes. *J. Agric. Food Chem.* **2012**, *60*, 3735–3744. [CrossRef]
136. González, R.; Ballester, I.; López-Posadas, R.; Suárez, M.D.; Zarzuelo, A.; Martínez-Augustin, O.; Medina, F.S.D. Effects of Flavonoids and Other Polyphenols on Inflammation. *Crit. Rev. Food Sci. Nutr.* **2011**, *51*, 331–362. [CrossRef]
137. Sombié, P.A.E.D.; Compaoré, M.; Coulibaly, A.Y.; Ouédraogo, J.T.; Tignégré, J.-B.D.L.S.; Kiendrébéogo, M. Antioxidant and Phytochemical Studies of 31 Cowpeas (*Vigna unguiculata* (L. Walp.)) Genotypes from Burkina Faso. *Foods* **2018**, *7*, 143. [CrossRef]
138. Adedayo, B.C.; Anyasi, T.A.; Taylor, M.J.; Rautenbauch, F.; Roes-Hill, L.; Jideani, V.A. Phytochemical Composition and Antioxidant Properties of Methanolic Extracts of Whole and Dehulled Bambara Groundnut (*Vigna subterranea*) Seeds. *Sci. Rep.* **2021**, *11*, 14116. [CrossRef]
139. Sowndhararajan, K.; Siddhuraju, P.; Manian, S. Antioxidant and Free Radical Scavenging Capacity of the Underutilized Legume, *Vigna vexillata* (L.) A. Rich. *J. Food Compos. Anal.* **2011**, *24*, 160–165. [CrossRef]
140. Leu, Y.-L.; Hwang, T.-L.; Kuo, P.-C.; Liou, K.-P.; Huang, B.-S.; Chen, G.-F. Constituents from *Vigna vexillata* and Their Anti-Inflammatory Activity. *Int. J. Mol. Sci.* **2012**, *13*, 9754–9768. [CrossRef] [PubMed]
141. Arise, A.K.; Alashi, A.M.; Nwachukwu, I.D.; Ijabadeniya, O.A.; Aluko, R.E.; Amonsou, E.O. Antioxidant Activities of Bambara Groundnut (*Vigna subterranea*) Protein Hydrolysates and Their Membrane Ultrafiltration Fractions. *Food Funct.* **2016**, *7*, 2431–2437. [CrossRef]
142. Quan, T.H.; Benjakul, S.; Sae-leaw, T.; Balange, A.K.; Maqsood, S. Protein–Polyphenol Conjugates: Antioxidant Property, Functionalities and Their Applications. *Trends Food Sci. Technol.* **2019**, *91*, 507–517. [CrossRef]
143. James, A.M. Evolutionary Insights into Two Plant Protein Families: Bowman-Birk Inhibitors and Asparaginyl Endopeptidases. Ph.D. Thesis, University of Victoria, Victoria, BC, Canada, 2017.
144. Clemente, A.; del Carmen Arques, M. Bowman-Birk Inhibitors from Legumes as Colorectal Chemopreventive Agents. *World J. Gastroenterol. WJG* **2014**, *20*, 10305. [CrossRef]
145. Clemente, A.; Olias, R. Beneficial Effects of Legumes in Gut Health. *Curr. Opin. Food Sci.* **2017**, *14*, 32–36. [CrossRef]
146. Panzeri, D.; Guzzetti, L.; Sacco, G.; Tedeschi, G.; Nonnis, S.; Airoidi, C.; Labra, M.; Fusi, P.; Forcella, M.; Regonesi, M.E. Effectiveness of *Vigna unguiculata* Seed Extracts in Preventing Colorectal Cancer. *Food Funct.* **2020**, *11*, 5853–5865. [CrossRef]
147. Mehdad, A.; Brumana, G.; Souza, A.A.; Barbosa, J.; Ventura, M.M.; De Freitas, S. A Bowman–Birk Inhibitor Induces Apoptosis in Human Breast Adenocarcinoma through Mitochondrial Impairment and Oxidative Damage Following Proteasome 20S Inhibition. *Cell Death Discov.* **2016**, *2*, 1–10. [CrossRef]
148. Teixeira-Guedes, C.I.; Oppolzer, D.; Barros, A.I.; Pereira-Wilson, C. Phenolic Rich Extracts from Cowpea Sprouts Decrease Cell Proliferation and Enhance 5-Fluorouracil Effect in Human Colorectal Cancer Cell Lines. *J. Funct. Foods* **2019**, *60*, 103452. [CrossRef]
149. Guo, X.; Li, T.; Tang, K.; Liu, R.H. Effect of Germination on Phytochemical Profiles and Antioxidant Activity of Mung Bean Sprouts (*Vigna radiata*). *J. Agric. Food Chem.* **2012**, *60*, 11050–11055. [CrossRef]
150. Rejhová, A.; Opatková, A.; Čumová, A.; Sliva, D.; Vodička, P. Natural Compounds and Combination Therapy in Colorectal Cancer Treatment. *Eur. J. Med. Chem.* **2018**, *144*, 582–594. [CrossRef]
151. Xavier, C.P.; Lima, C.F.; Preto, A.; Seruca, R.; Fernandes-Ferreira, M.; Pereira-Wilson, C. Luteolin, Quercetin and Ursolic Acid Are Potent Inhibitors of Proliferation and Inducers of Apoptosis in Both KRAS and BRAF Mutated Human Colorectal Cancer Cells. *Cancer Lett.* **2009**, *281*, 162–170. [CrossRef]
152. Yang, L.; Liu, Y.; Wang, M.; Qian, Y.; Dong, X.; Gu, H.; Wang, H.; Guo, S.; Hisamitsu, T. Quercetin-Induced Apoptosis of HT-29 Colon Cancer Cells via Inhibition of the Akt–CSN6–Myc Signaling Axis. *Mol. Med. Rep.* **2016**, *14*, 4559–4566. [CrossRef]
153. Reyes-Farias, M.; Carrasco-Pozo, C. The Anti-Cancer Effect of Quercetin: Molecular Implications in Cancer Metabolism. *Int. J. Mol. Sci.* **2019**, *20*, 3177. [CrossRef]
154. Fact Sheets—Malnutrition. Available online: <https://www.who.int/news-room/fact-sheets/detail/malnutrition> (accessed on 17 January 2022).
155. Tan, W.-C.; Tan, C.-H.; Nyam, K.-L.; Tan, C.-P.; Julkifle, A. Nutritive Bambara Groundnut Powdered Drink Mix: Characterization and in-Vivo Assessment of the Cholesterol-Lowering Effect. *J. Food Sci. Technol.* **2021**, *58*, 2992–3000. [CrossRef]
156. Kanetro, B. Hypocholesterolemic Properties of Protein Isolate from Cowpeas (*Vigna unguiculata*) Sprout in Normal and Diabetic Rats. *Procedia Food Sci.* **2015**, *3*, 112–118. [CrossRef]
157. Janeesh, P.A.; Abraham, A. Amelioration of Cholesterol Induced Atherosclerosis by Normalizing Gene Expression, Cholesterol Profile and Antioxidant Enzymes by *Vigna unguiculata*. *Plant Foods Hum. Nutr.* **2013**, *68*, 118–123. [CrossRef]
158. Lattanzio, V.; Cardinali, A.; Linsalata, V.; Perrino, P.; Ng, N.Q. A Chemosystematic Study of the Flavonoids of *Vigna*. *Genet. Resour. Crop. Evol.* **1996**, *43*, 493–504. [CrossRef]
159. Lattanzio, V.; Terzano, R.; Cicco, N.; Cardinali, A.; Venere, D.D.; Linsalata, V. Seed Coat Tannins and Bruchid Resistance in Stored Cowpea Seeds. *J. Sci. Food Agric.* **2005**, *85*, 839–846. [CrossRef]

160. Gutiérrez-Urbe, J.A.; Romo-Lopez, I.; Serna-Saldívar, S.O. Phenolic Composition and Mammary Cancer Cell Inhibition of Extracts of Whole Cowpeas (*Vigna unguiculata*) and Its Anatomical Parts. *J. Funct. Foods* **2011**, *3*, 290–297. [[CrossRef](#)]
161. Herniter, I.A.; Lo, R.; Muñoz-Amatriáin, M.; Lo, S.; Guo, Y.-N.; Huynh, B.-L.; Lucas, M.; Jia, Z.; Roberts, P.A.; Lonardi, S. Seed Coat Pattern QTL and Development in Cowpea (*Vigna unguiculata* [L.] Walp.). *Front. Plant Sci.* **2019**, *10*, 1346. [[CrossRef](#)]
162. Gaiero, P.; Speranza, P.; de Jong, H. Introgressive Hybridization in Potato Revealed by Novel Cytogenetic and Genomic Technologies. *Am. J. Potato Res.* **2018**, *95*, 607–621. [[CrossRef](#)]
163. Aliyu, B. Crossability of *Vigna rhomboidea* Burt. Davy with Cowpea (*V. unguiculata* (L.) Walp.). *Genet. Resour. Crop. Evol.* **2005**, *52*, 447–453. [[CrossRef](#)]
164. Fatokun, C.A.; Ng, Q. Outcrossing in Cowpea. *J. Food Agric. Environ.* **2007**, *5*, 334–338.
165. Pasquet, R.S.; Peltier, A.; Hufford, M.B.; Oudin, E.; Saulnier, J.; Paul, L.; Knudsen, J.T.; Herren, H.R.; Gepts, P. Long-Distance Pollen Flow Assessment through Evaluation of Pollinator Foraging Range Suggests Transgene Escape Distances. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 13456–13461. [[CrossRef](#)]
166. Sakupwanya, S.; Mithen, R.; Mutangandura-Mhlanga, T. *Studies on the African Vigna Genepool. II. Hybridization Studies with Vigna unguiculata var. tenuis and var. stenophylla. Bulletin des Ressources Genétiques Vegetales (CIRP/FAO); Noticiario de Recursos Genéticos Vegetales (CIRF/FAO)*: Roma, Italy, 1989.
167. Mithen, R.; Kibblewhite, H. Taxonomy and ecology of *Vigna unguiculata* (Leguminosae—Papilionoideae) in south-central africa. *Kirkia* **1993**, *14*, 100–113.
168. Fatokun, C.A. Wide Hybridization in Cowpea: Problems and Prospects. *Euphytica* **1991**, *54*, 137–140. [[CrossRef](#)]
169. Vijaykumar, A.; Saini, A.; Jawali, N. Phylogenetic Analysis of Subgenus *Vigna* Species Using Nuclear Ribosomal RNA ITS: Evidence of Hybridization among *Vigna unguiculata* Subspecies. *J. Hered.* **2010**, *101*, 177–188. [[CrossRef](#)]
170. Vijaykumar, A.; Saini, A.; Jawali, N. Molecular Characterization of Intergenic Spacer Region of 5S Ribosomal RNA Genes in Subgenus *Vigna*: Extensive Hybridization among *V. unguiculata* Subspecies. *Plant Syst. Evol.* **2011**, *294*, 39–55. [[CrossRef](#)]
171. Fatokun, C.A.; Singh, B.B. Interspecific Hybridization between *Vigna pubescens* and *V. unguiculata* (L.) Walp. through Embryo Rescue. *Plant Cell Tissue Organ Cult.* **1987**, *9*, 229–233. [[CrossRef](#)]
172. Fatokun, C.A.; Perrino, P.; Ng, N.Q. Wide Crossing in African *Vigna* Species. *Adv. Cowpea Res.* **1997**, 50–57.
173. Mohammed, M.S.; Russom, Z.; Abdul, S.D. Inheritance of Hairiness and Pod Shattering, Heritability and Correlation Studies in Crosses between Cultivated Cowpea (*Vigna unguiculata* (L.) Walp.) and Its Wild (var. *pubescens*) Relative. *Euphytica* **2010**, *171*, 397–407. [[CrossRef](#)]
174. Lelou, B.; Diawata, M.; Van Damme, P. A Study of Intraspecific Hybrid Lines Derived from the Reciprocal Crosses between Wild Accessions and Cultivated Cowpeas (*Vigna unguiculata* (L.) Walp.). *Afr. J. Plant Sci.* **2011**, *5*, 337–348.
175. Amusa, O.D.; Ogunkanmi, L.A.; Adetumbi, J.A.; Akinyosoye, S.T.; Bolarinwa, K.A.; Ogundipe, O.T. Intraspecific-Cross Compatibility in Cowpea (*Vigna unguiculata* (L.) Walp.). *J. Crop. Improv.* **2021**, *19*, 100–119. [[CrossRef](#)]
176. Souleymane, A.; Aken’Ova, M.E.; Fatokun, C.A.; Alabi, O.Y. Screening for Resistance to Cowpea Aphid (*Aphis craccivora* Koch) in Wild and Cultivated Cowpea (*Vigna unguiculata* L. Walp.) Accessions. *Int. J. Sci. Environ. Technol.* **2013**, *2*, 611–621.
177. Lo, S.; Muñoz-Amatriáin, M.; Boukar, O.; Herniter, I.; Cisse, N.; Guo, Y.-N.; Roberts, P.A.; Xu, S.; Fatokun, C.; Close, T.J. Identification of QTL Controlling Domestication-Related Traits in Cowpea (*Vigna unguiculata* L. Walp.). *Sci. Rep.* **2018**, *8*, 1–9. [[CrossRef](#)] [[PubMed](#)]
178. Ogiangbe, O.N.; Jackai, L.E.N.; Ewete, F.K.; Hughes, J.d.A.; Lajide, L. Reduced Consumption and Use of Pods of *Vigna* Species (Leguminosae) by *Maruca vitrata* (Lepidoptera: Pyralidae). *Afr. Entomol.* **2002**, *10*, 333–340.
179. Koon, P.; Osisanya, E.O.; Jackai, L.E.N.; Tamo, M.; Markham, R.H. Resistance in Accessions of Cowpea to the Coreid Pod-Bug *Clavigralla tomentosicollis* (Hemiptera: Coreidae). *J. Econ. Entomol.* **2002**, *95*, 1281–1288. [[CrossRef](#)]
180. Massawe, F.J.; Schenkel, W.; Basu, S.; Temba, E.M. Artificial Hybridisation in Bambara Groundnut (*Vigna subterranea* (L.) Verdc.). In Proceedings of the International Symposium on Bambara Groundnut, Botswana College of Agriculture, Gaborone, Botswana, 8–12 September 2003; pp. 8–12.
181. Suwanprasert, J.; Toojinda, T.; Srinives, P.; Chanprame, S. Hybridization Technique for Bambara Groundnut. *Breed. Sci.* **2006**, *56*, 125–129. [[CrossRef](#)]
182. Oyiga, B.C.; Uguru, M.I.; Aruah, C.B. Studies on the Floral Traits and Their Implications on Pod and Seed Yields in Bambara Groundnut [*Vigna subterranea* (L.) Verdc]. *Aust. J. Crop. Sci.* **2010**, *4*, 91–97.
183. Onwubiko, N.I.C.; Odum, O.B.; Utazi, C.O.; Poly-Mbah, P.C. Studies on the Adaptation of Bambara Groundnut [*Vigna subterranea* (L.) Verdc] in Owerri Southeastern Nigeria. *N. Y. Sci. J.* **2011**, *4*, 60–67. [[CrossRef](#)]
184. Basu, S.; Mayes, S.; Davey, M.; Roberts, J.A.; Azam-Ali, S.N.; Mithen, R.; Pasquet, R.S. Inheritance of ‘Domestication’ Traits in Bambara Groundnut (*Vigna subterranea* (L.) Verdc.). *Euphytica* **2007**, *157*, 59–68. [[CrossRef](#)]
185. James, A.T.; Lawn, R.J. Inheritance of Selected Traits in Accessions of *Vigna vexillata* (L.) A Rich of Australian and African Origin. *Aust. J. Bot.* **1991**, *39*, 415–429. [[CrossRef](#)]
186. Damayanti, F.; Lawn, R.J.; Bielig, L.M. Genetic Compatibility among Domesticated and Wild Accessions of the Tropical Tuberos Legume *Vigna vexillata* (L.) A. Rich. *Crop. Pasture Sci.* **2010**, *61*, 785–797. [[CrossRef](#)]
187. Damayanti, F.; Lawn, R.J.; Bielig, L.M. Expression of Qualitative and Quantitative Traits in Hybrids between Domesticated and Wild Accessions of the Tropical Tuberos Legume *Vigna vexillata* (L.) A. Rich. *Crop. Pasture Sci.* **2010**, *61*, 798–811. [[CrossRef](#)]

188. Barone, A.; Del Giudice, A.; Ng, N.Q. Barriers to Interspecific Hybridization between *Vigna unguiculata* and *Vigna vexillata*. *Sex. Plant Reprod.* **1992**, *5*, 195–200. [[CrossRef](#)]
189. Gomathinayagam, P.; Rathnaswamy, R.; Ramaswamy, N.M. Interspecific Hybridization between *Vigna unguiculata* (L.) Walp. and *V. vexillata* (L.) A. Rich. through in Vitro Embryo Culture. *Euphytica* **1998**, *102*, 203–209. [[CrossRef](#)]
190. Boukar, O.; Abberton, M.; Oyatomi, O.; Togola, A.; Tripathi, L.; Fatokun, C. Introgression Breeding in Cowpea [*Vigna Unguiculata* (L.) Walp.]. *Front. Plant Sci.* **2020**, *11*, 567425. [[CrossRef](#)]
191. Lexer, C.; Heinze, B.; Alia, R.; Rieseberg, L.H. Hybrid Zones as a Tool for Identifying Adaptive Genetic Variation in Outbreeding Forest Trees: Lessons from Wild Annual Sunflowers (*Helianthus* spp.). *For. Ecol. Manag.* **2004**, *197*, 49–64. [[CrossRef](#)]
192. Huang, H.; Liu, Y. Natural Hybridization, Introgression Breeding, and Cultivar Improvement in the Genus *Actinidia*. *Tree Genet. Genomes* **2014**, *10*, 1113–1122. [[CrossRef](#)]
193. Janzen, G.M.; Wang, L.; Hufford, M.B. The Extent of Adaptive Wild Introgression in Crops. *New Phytol.* **2019**, *221*, 1279–1288. [[CrossRef](#)]
194. Wang, X.; Chen, L.; Ma, J. Genomic Introgression through Interspecific Hybridization Counteracts Genetic Bottleneck during Soybean Domestication. *Genome Biol.* **2019**, *20*, 1–15. [[CrossRef](#)]
195. Zecca, G.; Labra, M.; Grassi, F. Untangling the Evolution of American Wild Grapes: Admixed Species and How to Find Them. *Front. Plant Sci.* **2020**, *10*, 1814. [[CrossRef](#)]
196. Chacón-Sánchez, M.I.; Martínez-Castillo, J.; Duitama, J.; Debouck, D.G. Gene Flow in *Phaseolus* Beans and Its Role as a Plausible Driver of Ecological Fitness and Expansion of Cultigens. *Front. Ecol. Evol.* **2021**, *9*, 312. [[CrossRef](#)]
197. Kovach, M.J.; Calingacion, M.N.; Fitzgerald, M.A.; McCouch, S.R. The Origin and Evolution of Fragrance in Rice (*Oryza sativa* L.). *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 14444–14449. [[CrossRef](#)]
198. Hardigan, M.A.; Laimbeer, F.P.E.; Newton, L.; Crisovan, E.; Hamilton, J.P.; Vaillancourt, B.; Wiegert-Rininger, K.; Wood, J.C.; Douches, D.S.; Farré, E.M. Genome Diversity of Tuber-Bearing *Solanum* Uncovers Complex Evolutionary History and Targets of Domestication in the Cultivated Potato. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, E9999–E10008. [[CrossRef](#)]
199. Burgarella, C.; Barnaud, A.; Kane, N.A.; Jankowski, F.; Scarcelli, N.; Billot, C.; Vigouroux, Y.; Berthouly-Salazar, C. Adaptive Introgression: An Untapped Evolutionary Mechanism for Crop Adaptation. *Front. Plant Sci.* **2019**, *10*, 4. [[CrossRef](#)]
200. Pickrell, J.; Pritchard, J. Inference of Population Splits and Mixtures from Genome-Wide Allele Frequency Data. *Nat. Preced.* **2012**, *8*, e1002967. [[CrossRef](#)]
201. Martin, S.H.; Davey, J.W.; Jiggins, C.D. Evaluating the Use of ABBA–BABA Statistics to Locate Introgressed Loci. *Mol. Biol. Evol.* **2015**, *32*, 244–257. [[CrossRef](#)]
202. Martin, S.H.; Van Belleghem, S.M. Exploring Evolutionary Relationships across the Genome Using Topology Weighting. *Genetics* **2017**, *206*, 429–438. [[CrossRef](#)]
203. Flouri, T.; Jiao, X.; Rannala, B.; Yang, Z. A Bayesian Implementation of the Multispecies Coalescent Model with Introgression for Phylogenomic Analysis. *Mol. Biol. Evol.* **2020**, *37*, 1211–1223. [[CrossRef](#)]
204. Setter, D.; Mousset, S.; Cheng, X.; Nielsen, R.; DeGiorgio, M.; Hermisson, J. VolcanoFinder: Genomic Scans for Adaptive Introgression. *PLoS Genet.* **2020**, *16*, e1008867. [[CrossRef](#)]
205. Hellenthal, G.; Busby, G.B.; Band, G.; Wilson, J.F.; Capelli, C.; Falush, D.; Myers, S. A Genetic Atlas of Human Admixture History. *Science* **2014**, *343*, 747–751. [[CrossRef](#)]
206. Ellstrand, N.C.; Meirmans, P.; Rong, J.; Bartsch, D.; Ghosh, A.; De Jong, T.J.; Haccou, P.; Lu, B.-R.; Snow, A.A.; Neal Stewart Jr, C. Introgression of Crop Alleles into Wild or Weedy Populations. *Annu. Rev. Ecol. Syst.* **2013**, *44*, 325–345. [[CrossRef](#)]
207. Heredia, S.M.; Ellstrand, N.C. Novel Seed Protection in the Recently Evolved Invasive, California Wild Radish, a Hybrid *Raphanus* sp. (Brassicaceae). *Am. J. Bot.* **2014**, *101*, 2043–2051. [[CrossRef](#)]
208. Page, A.M.; Daunay, M.-C.; Aubriot, X.; Chapman, M.A. Domestication of Eggplants: A Phenotypic and Genomic Insight. In *The Eggplant Genome*; Springer: Berlin/Heidelberg, Germany, 2019; pp. 193–212.
209. Grimm, A.; Sahi, V.P.; Amann, M.; Vidotto, F.; Fogliatto, S.; Devos, K.M.; Ferrero, A.; Nick, P. Italian Weedy Rice—A Case of de-Domestication? *Ecol. Evol.* **2020**, *10*, 8449–8464. [[CrossRef](#)] [[PubMed](#)]
210. Grassi, F.; De Lorenzis, G. Back to the Origins: Background and Perspectives of Grapevine Domestication. *Int. J. Mol. Sci.* **2021**, *22*, 4518. [[CrossRef](#)]
211. McAlvay, A.C.; Ragsdale, A.P.; Mabry, M.E.; Qi, X.; Bird, K.; Velasco, P.; An, H.; Pires, C.; Emshwiller, E. Brassica Rapa Domestication: Untangling Wild and Feral Forms and Convergence of Crop Morphotypes. *bioRxiv* **2021**. [[CrossRef](#)]
212. Scossa, F.; Fernie, A.R. When a Crop Goes Back to the Wild: Feralization. *Trends Plant Sci.* **2021**, *26*, 543–545. [[CrossRef](#)]
213. Cao, Q.; Lu, B.-R.; Xia, H.U.I.; Rong, J.; Sala, F.; Spada, A.; Grassi, F. Genetic Diversity and Origin of Weedy Rice (*Oryza sativa* f. *spontanea*) Populations Found in North-Eastern China Revealed by Simple Sequence Repeat (SSR) Markers. *Ann. Bot.* **2006**, *98*, 1241–1252. [[CrossRef](#)]
214. Gering, E.; Incorvaia, D.; Henriksen, R.; Conner, J.; Getty, T.; Wright, D. Getting Back to Nature: Feralization in Animals and Plants. *Trends Ecol. Evol.* **2019**, *34*, 1137–1151. [[CrossRef](#)] [[PubMed](#)]
215. Feleke, Y.; Pasquet, R.S.; Gepts, P. Development of PCR-Based Chloroplast DNA Markers That Characterize Domesticated Cowpea (*Vigna unguiculata* ssp. *unguiculata* var. *unguiculata*) and Highlight Its Crop-Weed Complex. *Plant Syst. Evol.* **2006**, *262*, 75–87. [[CrossRef](#)]

216. Kouam, E.B.; Pasquet, R.S.; Campagne, P.; Tignegre, J.-B.; Thoen, K.; Gaudin, R.; Ouedraogo, J.T.; Salifu, A.B.; Muluvi, G.M.; Gepts, P. Genetic Structure and Mating System of Wild Cowpea Populations in West Africa. *BMC Plant Biol.* **2012**, *12*, 113. [[CrossRef](#)]
217. Kouadio, D.; Echikh, N.; Toussaint, A.; Pasquet, R.S.; Baudoin, J.-P. Organization of the Gene Pool of *Vigna unguiculata* (L.) Walp.: Crosses between the Wild and Cultivated Forms of Cowpea. *Biotechnol. Agron. Soc. Environ.* **2007**, *11*, 47–57.
218. Darwin's, C. *On the Origin of Species*; John Murray: London, UK, 1859.
219. Ho, W.K.; Chai, H.H.; Kendabie, P.; Ahmad, N.S.; Jani, J.; Massawe, F.; Kilian, A.; Mayes, S. Integrating Genetic Maps in Bambara Groundnut [*Vigna subterranea* (L.) Verdc.] and Their Syntenic Relationships among Closely Related Legumes. *BMC Genom.* **2017**, *18*, 1–9. [[CrossRef](#)] [[PubMed](#)]
220. Paudel, D.; Dareus, R.; Rosenwald, J.; Munoz-Amatriaín, M.; Rios, E. Genome-Wide Association Study Reveals Candidate Genes for Flowering Time in Cowpea (*Vigna unguiculata* [L.] Walp.). *bioRxiv* **2021**, *12*, 667038. [[CrossRef](#)] [[PubMed](#)]
221. Lo, S.; Fatokun, C.; Boukar, O.; Gepts, P.; Close, T.J.; Muñoz-Amatriaín, M. Identification of QTL for Perenniality and Floral Scent in Cowpea (*Vigna unguiculata* [L.] Walp.). *PLoS ONE* **2020**, *15*, e0229167. [[CrossRef](#)] [[PubMed](#)]
222. Lonardi, S.; Muñoz-Amatriaín, M.; Liang, Q.; Shu, S.; Wanamaker, S.I.; Lo, S.; Tanskanen, J.; Schulman, A.H.; Zhu, T.; Luo, M.-C. The Genome of Cowpea (*Vigna unguiculata* [L.] Walp.). *Plant J.* **2019**, *98*, 767–782. [[CrossRef](#)]
223. González, A.M.; Yuste-Lisbona, F.J.; Saburido, S.; Bretones, S.; De Ron, A.M.; Lozano, R.; Santalla, M. Major Contribution of Flowering Time and Vegetative Growth to Plant Production in Common Bean as Deduced from a Comparative Genetic Mapping. *Front. Plant Sci.* **2016**, *7*, 1940. [[CrossRef](#)]
224. Gong, Z. Flowering Phenology as a Core Domestication Trait in Soybean. *J. Integr. Plant Biol.* **2020**, *62*, 546–549. [[CrossRef](#)]
225. Levy, Y.Y.; Dean, C. Control of Flowering Time. *Curr. Opin. Plant Biol.* **1998**, *1*, 49–54. [[CrossRef](#)]
226. Fernie, A.R.; Yan, J. *De Novo* Domestication: An Alternative Route toward New Crops for the Future. *Mol. Plant* **2019**, *12*, 615–631. [[CrossRef](#)]
227. Smýkal, P.; Nelson, M.N.; Berger, J.D.; Von Wettberg, E.J. The Impact of Genetic Changes during Crop Domestication. *Agronomy* **2018**, *8*, 119. [[CrossRef](#)]
228. Khan, A.W.; Garg, V.; Roorkiwal, M.; Golicz, A.A.; Edwards, D.; Varshney, R.K. Super-Pangenome by Integrating the Wild Side of a Species for Accelerated Crop Improvement. *Trends Plant Sci.* **2020**, *25*, 148–158. [[CrossRef](#)] [[PubMed](#)]
229. Della Coletta, R.; Qiu, Y.; Ou, S.; Hufford, M.B.; Hirsch, C.N. How the Pan-Genome Is Changing Crop Genomics and Improvement. *Genome Biol.* **2021**, *22*, 1–19. [[CrossRef](#)] [[PubMed](#)]
230. Rodríguez-Leal, D.; Lemmon, Z.H.; Man, J.; Bartlett, M.E.; Lippman, Z.B. Engineering Quantitative Trait Variation for Crop Improvement by Genome Editing. *Cell* **2017**, *171*, 470–480. [[CrossRef](#)] [[PubMed](#)]
231. Xiao, A.; Wang, Z.; Hu, Y.; Wu, Y.; Luo, Z.; Yang, Z.; Zu, Y.; Li, W.; Huang, P.; Tong, X. Chromosomal Deletions and Inversions Mediated by TALENs and CRISPR/Cas in Zebrafish. *Nucleic Acids Res.* **2013**, *41*, e141. [[CrossRef](#)]
232. DeHaan, L.; Larson, S.; López-Marqués, R.L.; Wenkel, S.; Gao, C.; Palmgren, M. Roadmap for Accelerated Domestication of an Emerging Perennial Grain Crop. *Trends Plant Sci.* **2020**, *25*, 525–537. [[CrossRef](#)]
233. Li, M.; Li, X.; Zhou, Z.; Wu, P.; Fang, M.; Pan, X.; Lin, Q.; Luo, W.; Wu, G.; Li, H. Reassessment of the Four Yield-Related Genes Gn1a, DEP1, GS3, and IPA1 in Rice Using a CRISPR/Cas9 System. *Front. Plant Sci.* **2016**, *7*, 377. [[CrossRef](#)]
234. Zsögön, A.; Čermák, T.; Naves, E.R.; Notini, M.M.; Edel, K.H.; Weinel, S.; Freschi, L.; Voytas, D.F.; Kudla, J.; Peres, L.E.P. *De Novo* Domestication of Wild Tomato Using Genome Editing. *Nat. Biotechnol.* **2018**, *36*, 1211–1216. [[CrossRef](#)]
235. Li, T.; Yang, X.; Yu, Y.; Si, X.; Zhai, X.; Zhang, H.; Dong, W.; Gao, C.; Xu, C. Domestication of Wild Tomato Is Accelerated by Genome Editing. *Nat. Biotechnol.* **2018**, *36*, 1160–1163. [[CrossRef](#)]
236. Ji, J.; Zhang, C.; Sun, Z.; Wang, L.; Duanmu, D.; Fan, Q. Genome Editing in Cowpea *Vigna unguiculata* Using CRISPR-Cas9. *Int. J. Mol. Sci.* **2019**, *20*, 2471. [[CrossRef](#)]
237. Østerberg, J.T.; Xiang, W.; Olsen, L.I.; Edenbrandt, A.K.; Vedel, S.E.; Christiansen, A.; Landes, X.; Andersen, M.M.; Pagh, P.; Sandøe, P. Accelerating the Domestication of New Crops: Feasibility and Approaches. *Trends Plant Sci.* **2017**, *22*, 373–384. [[CrossRef](#)]
238. Catarino, S.; Rangel, J.; Darbyshire, I.; Costa, E.; Duarte, M.C.; Romeiras, M.M. Conservation Priorities for African *Vigna* Species: Unveiling Angola's Diversity Hotspots. *Glob. Ecol. Conserv.* **2021**, *25*, e01415. [[CrossRef](#)]
239. Tettelin, H.; Massignani, V.; Cieslewicz, M.J.; Donati, C.; Medini, D.; Ward, N.L.; Angiuoli, S.V.; Crabtree, J.; Jones, A.L.; Durkin, A.S. Genome Analysis of Multiple Pathogenic Isolates of *Streptococcus agalactiae*: Implications for the Microbial “Pan-Genome”. *Proc. Natl. Acad. Sci. USA* **2005**, *102*, 13950–13955. [[CrossRef](#)] [[PubMed](#)]
240. Bayer, P.E.; Golicz, A.A.; Scheben, A.; Batley, J.; Edwards, D. Plant Pan-Genomes Are the New Reference. *Nat. Plants* **2020**, *6*, 914–920. [[CrossRef](#)] [[PubMed](#)]