



Article Supplementary Far-Red Light for Photosynthetic Active Radiation Differentially Influences the Photochemical Efficiency and Biomass Accumulation in Greenhouse-Grown Lettuce

Haijie Dou^{1,†}, Xin Li^{2,†}, Zhixin Li², Jinxiu Song³, Yanjie Yang² and Zhengnan Yan^{2,*}

- ¹ College of Intelligent Science and Engineering, Beijing University of Agriculture, Beijing 102206, China; haijiedou@bua.edu.cn
- ² College of Horticulture, Qingdao Agricultural University, Qingdao 266109, China
- ³ College of Agricultural Engineering, Jiangsu University, Zhenjiang 212013, China
 - * Correspondence: yanzn@qau.edu.cn
- ⁺ These authors contributed equally to this work.

Abstract: Adding far-red (FR, 700-800 nm) light to photosynthetic active radiation (400-700 nm) proved to be a possible approach to increasing plant biomass accumulation for lettuce production in indoor vertical farms with artificial lighting as a sole-source lighting. However, how FR light addition influences plant growth, development, and metabolic processes and the optimal value of FR photon flux density for greenhouse-grown lettuce under sunlight are still unclear. This work aims to quantify the value of supplementary FR light with different intensities on lettuce morphological and physiological characteristics in a greenhouse. Lettuce 'Dasusheng' (Lactuca sativa L.) was grown in a greenhouse under seven light treatments, including white plus red LEDs with FR photon flux density at 0, 10, 30, 50, 70, and 90 μ mol m⁻² s⁻¹ (WR, WR + FR10, WR + FR30, WR + FR50, WR + FR70, and WR + FR90, respectively), and lettuce grown with sunlight only was marked as natural light (NL). FR light addition improved the electron transport flux per cross section and performance index (PI_{abs}, PI_{total}) and decreased the changes in relative variable fluorescence of lettuce leaves compared to plants under NL. Specifically, the PI_{abs} of lettuce leaves were 41%, 41%, 38%, 33%, 26%, and 25% lower under control than in plants under treatments WR + FR90, WR + FR70, WR + FR50, WR + FR30, WR + FR10, and WR, respectively. Leaf number, leaf area, and biomass accumulation of lettuce followed a quadratic function with increasing FR light intensity and were the highest under treatment WR + FR50. The shoot fresh weight and dry weight of lettuce were increased by 111% and 275%, respectively, under treatment WR + FR50 compared to NL. The contents of vitamin C, reducing sugar, total soluble sugar, and starch in lettuce showed a similar trend with biomass accumulation. In conclusion, with commonly used photosynthetic photon flux density (PPFD, 400–700 nm) around 200 μ mol m⁻² s⁻¹, supplementary FR light intensity of 30~50 μ mol m⁻² s⁻¹ was suggested to enhance the photochemistry efficiency, biomass accumulation, and carbohydrates' contents in greenhouse-grown lettuce.

Keywords: far-red; Lactuca sativa; leaf expansion; performance index

1. Introduction

In the past decade, a number of studies have focused on the influences of different light intensities or light spectra within photosynthetic active radiation (PAR, ranging from 400 nm to 700 nm) on plant growth in controlled environments [1–4]. On the contrary, far-red (FR, 700–800 nm) photons, which are out of PAR range, have received little attention and are often presumed to be less efficient for plant photosynthetic functioning compared to shorter wavelengths such as blue (B), green (G), or red (R) light [5–7]. However, in recent years, an increasing body of empirical evidence suggested that adding FR to PAR wavelengths improved lettuce (*Lactuca sativa* L.) biomass accumulation through stimulation



Citation: Dou, H.; Li, X.; Li, Z.; Song, J.; Yang, Y.; Yan, Z. Supplementary Far-Red Light for Photosynthetic Active Radiation Differentially Influences the Photochemical Efficiency and Biomass Accumulation in Greenhouse-Grown Lettuce. *Plants* 2024, *13*, 2169. https://doi.org/ 10.3390/plants13152169

Academic Editors: Barbara Frąszczak and Anita Schroeter-Zakrzewska

Received: 30 June 2024 Revised: 2 August 2024 Accepted: 2 August 2024 Published: 5 August 2024



Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). of leaf expansion and increased the amount of light energy captured by leaves, as well as a better excitation between the two photosystems, photosystem I (PSI) and photosystem II (PSII) [6–11]. Zhen and van Iersel reported that the quantum yield of PSII (φ PSII) and net photosynthetic rate (P_n) in lettuce increased immediately by adding FR photons to combined R&B photons [R₇₆G₁B₂3, subscript numbers indicate the photon percentage of R, G, and B wavelengths in the total photon flux density (TPFD, 400–780 nm), respectively] and white (W) photons (B₁₂G₄₃R₄₁FR₄), owing to a more balanced excitation of PSI and PSII [6]. Further, FR light is equally efficient at driving canopy photosynthesis when acting synergistically with PAR wavelengths among 14 plant species, while the magnitude of the effect was less at longer wavelengths (711, 723, and 746 nm) [7]. However, it was also reported that FR addition to combined R&B photons showed no influence on leaf P_n and decreased the plant's photosynthetic capacity in the long term due to a lower chlorophyll and total nitrogen content, which reduced leaf absorbance [12,13]. Thus, it was postulated that the acclimation process of plant morphology triggered by FR addition plays a major role in improving the yield of indoor cultivated lettuce [9,13]. Legendre and van Iersel compared the efficiency of FR addition to PAR and PAR wavelengths related to light interception and biomass accumulation, and results indicated that FR light was 57% and 183% more effective at increasing the light interception received by the plant, as well as 92% and 162% more effective at increasing plant biomass at the early and late harvests, respectively, as compared to PAR wavelengths [10]. Accordingly, the enhanced crop production by FR addition, which was mainly caused by plant morphological responses, cannot be achieved by adding similar amounts of shorter wavelengths [10].

Although increasing the FR photon flux density may have beneficial effects on photosynthesis and biomass accumulation, this can happen at the expense of the contents of photosynthetic pigments. Lower levels of chlorophyll and carotenoids are often reported when plants are grown under FR addition [14,15]. Zou et al. reported that supplementary FR light during cultivation, especially under FR light during the day (FR photon flux density of 90 µmol m⁻² s⁻¹, PPFD of 200 µmol m⁻² s⁻¹ provided by a combination of red and blue light, photoperiod of 16 h), is associated with reduced nutraceutical quality at harvest and potentially shorter shelf-life, which was related to elevated O²⁻ content along with decreased activity of enzymatic (superoxide dismutase) and reduced levels of non-enzymatic (carotenoids, total phenolics, and flavonoids) antioxidants in lettuce plants [16]. The sugar content, on the other hand, is often increased when plants are grown under higher FR fractions, due to enhanced photochemistry efficiency and relative CO₂ assimilation [8,16]. However, the regulatory mechanism still needs to be further studied.

Lettuce, the main ingredient in many salads and a popular sandwich topper, is one of the most widely consumed leafy greens around the world [17–19]. According to the Food and Agriculture Organization of the United Nations (FAO), China is the world's largest lettuce producer, with an estimated 17.9 million metric tons to be produced in 2020, followed by the United States [20]. With a rising demand for lettuce production, production occurs year-round, especially with an increasing amount grown in controlled environments (e.g., greenhouses and indoor vertical farms, IVF). However, lettuce growth can be slow when the daily light integral (DLI, product of PPFD and photoperiod) is low, especially in greenhouses during early spring, late autumn, and winter time in northern latitudes [21]. Therefore, supplemental lighting, especially provided by light-emitting diodes (LEDs), is often used to increase the DLI for greenhouse lettuce production and obtain high-quality produce [21–23].

FR light is a critical signal that influences plant growth, development, and metabolic processes. To date, most studies have focused on the effect of FR light addition on lettuce morphological characteristics and photosynthetic indicators in a fully controlled environment (e.g., IVF) with artificial lighting as sole-source lighting [6–11,13,16], which may not be applicable to semi-closed agricultural facilities (e.g., greenhouses), one vital form for lettuce production. Further, the lighting environment in greenhouses is different from fully controlled facilities, in which few FR photons are provided by sunlight, as well as varying

light intensity, photoperiods, and DLI, which may not be suitable for lettuce cultivation. Thus, this work aims to explore the influence of supplementary FR light with different intensities on lettuce morphological and physiological characteristics in greenhouses during low DLI periods. We hypothesized that in greenhouses with sunlight at a low DLI, supplementary PAR with FR photons would be effective in increasing lettuce growth, biomass accumulation, and sugar contents, and these morphological and growth responses should have a qualitative threshold.

2. Results

2.1. Chlorophyll: Fluorescence Parameters of Greenhouse-Grown Lettuce

Supplementary light significantly reduced the changes in the relative variable fluorescence (ΔV_t) value of lettuce leaves compared to plants under natural light (NL), especially at the J and I steps (Figure 1). Specifically, the magnitude of the reduced amplitude of ΔV_t in lettuce leaves increased with increasing FR light intensity from 0 to 70 µmol m⁻² s⁻¹, while treatment WR + FR90 reduced the ΔV_t value by 12% and 59% compared to plants under treatment WR + FR70 at the J and I steps, respectively. From I to P, FR light addition with the highest intensity of 90 µmol m⁻² s⁻¹ increased the ΔV_t value significantly, whereas other treatments decreased the ΔV_t value in lettuce leaves.



Figure 1. Induction curve and curve of ΔV_t of chlorophyll a fluorescence in lettuce leaves under different treatments, including white plus red LEDs with FR photon flux density at 0, 10, 30, 50, 70, and 90 µmol m⁻² s⁻¹ (WR, WR + FR10, WR + FR30, WR + FR50, WR + FR70, and WR + FR90, respectively), and lettuce grown with natural light only was marked as NL (three replicates).

The chlorophyll fluorescence characteristic parameters of lettuce leaves are presented in Figure 2. Results indicated that supplementary light had no influence on the F_v/F_m of lettuce plants in comparison to plants under NL, while supplementary light reduced the absorption flux per reaction center (ABS/RC), trapped energy flux per reaction center (TR_o/RC), electron transport flux per reaction center (ET_o/RC), and DI_o/RC in lettuce. Conversely, the electron transport flux and dissipation of energy per cross section (ET_o/CS_m , DI_o/CS_m) of lettuce plants were increased by supplementary light treatments, which increased with increasing FR light intensity. The PI_{abs} of lettuce plants showed a similar trend, which was 41%, 41%, 38%, 33%, 26%, and 25% lower under NL than in plants under treatments WR + FR90, WR + FR70, WR + FR50, WR + FR30, WR + FR10, and WR, respectively. The PI_{total} of lettuce plants was 10% and 13% lower under treatment WR + FR90 compared to plants under treatments WR + FR70, of lettuce decreased with the increase in FR photon flux density (Figure 3). The value of Q_B-NRC in lettuce leaves was the lowest in plants under treatments WR + FR90, WR + FR70, and WR + FR50.



Figure 2. The absorption of flux per cross section (ABS/CS_m), trapped energy flux per cross section (TR_o/CS_m), electron transport flux per cross section (ET_o/CS_m), dissipation of energy per cross section (DI_o/CS_m), absorption flux per reaction center (ABS/RC), trapped energy flux per reaction center (TR_o/RC), electron transport flux per reaction center (ET_o/RC), dissipation of energy per reaction center (DI_o/RC), the maximum photochemical quantum yield (F_v/F_m), performance index based on the absorbed light energy (PI_{abs}), and performance index (PI_{total}) of lettuce leaves under different treatments. The light treatments included white plus red LEDs with FR photon flux density at 0, 10, 30, 50, 70, and 90 µmol m⁻² s⁻¹ (WR, WR + FR10, WR + FR30, WR + FR50, WR + FR70, and WR + FR90, respectively), and lettuce grown with natural light only was marked as NL. Means followed by the same lowercase letters and NS are not significantly different for each measured parameter, according to the least-significant difference test ($p \le 0.05$). Error bars indicate the standard deviation (three replicates).



Figure 3. Q_B-non-reducing centers of greenhouse-grown lettuce under different treatments, including white plus red LEDs with FR photon flux density at 0, 10, 30, 50, 70, and 90 µmol m⁻² s⁻¹ (WR, WR + FR10, WR + FR30, WR + FR50, WR + FR70, and WR + FR90, respectively), and lettuce grown with natural light only was marked as NL. Means followed by the same lowercase letters are not significantly different, according to the least-significant difference test ($p \le 0.05$). Error bars indicate the standard deviation (three replicates).

2.2. Photosynthetic Characteristics and Chlorophyll Content of Greenhouse-Grown Lettuce

The maximum net photosynthetic rate ($P_{n max}$) of lettuce leaves was the lowest in plants under NL and showed no difference among supplementary light treatments with different FR light intensity except plants under treatment WR + FR90 (Table 1). The dark respiration rate (R_d) of lettuce leaves was the highest under treatments WR + FR30 and WR + FR50 and showed no difference among other treatments. The light compensation point (L_c) of lettuce leaves was the highest in treatments WR + FR30 and WR + FR50, followed by treatments WR + FR10, WR + FR70, and WR + FR90, and the lowest in treatments WR and NL.

Table 1. The maximum net photosynthetic rate ($P_{n max}$), dark respiration rate (R_d), and light compensation point (L_c) of greenhouse-grown lettuce leaves under different treatments, including white plus red LEDs with FR photon flux density at 0, 10, 30, 50, 70, and 90 µmol m⁻² s⁻¹ (WR, WR + FR10, WR + FR30, WR + FR50, WR + FR70, and WR + FR90, respectively), and lettuce grown with natural light only was marked as NL. Means followed by the same lowercase letters within the column are not significantly different, according to the least-significant difference test ($p \le 0.05$). Errors indicate the standard deviation (three replicates).

Treatments	$P_{n max}$ (µmol m ⁻² s ⁻¹)		R _d (µmol m ⁻	² s ⁻¹)	L _c (μmol m ⁻² s ⁻¹)		
NL	6.5 ± 1.1	с	0.84 ± 0.09	b	12.6 ± 2.3	С	
WR	10.2 ± 0.9	ab	0.88 ± 0.16	b	12.7 ± 1.8	с	
WR + FR10	10.7 ± 2.0	а	1.01 ± 0.05	b	17.2 ± 3.1	b	
WR + FR30	11.6 ± 2.6	а	1.73 ± 0.34	а	20.1 ± 4.5	ab	
WR + FR50	11.7 ± 1.5	а	1.53 ± 0.14	а	21.3 ± 1.3	а	
WR + FR70	10.5 ± 0.1	ab	1.09 ± 0.14	b	16.5 ± 1.7	bc	
WR + FR90	8.0 ± 1.3	bc	1.01 ± 0.18	b	16.1 ± 2.9	bc	

According to the light response curve, supplementary light showed a significantly positive effect on the P_n of lettuce leaves (Supplementary Figure S1). Specifically, P_n of lettuce leaves were in the order as below: WR + FR30 and WR + FR50 > WR > WR + FR10 > WR + FR70 > WR + FR90 > NL. When the light intensity was below 1000 µmol m⁻² s⁻¹, P_n in treatment WR + FR50 was lower than plants in treatment WR + FR30 and gradually increased over the one in WR + FR30 when the light intensity exceeded 1000 µmol·m⁻²·s⁻¹.

Supplementary light had a significant impact on chlorophyll content in greenhousegrown lettuce (Figure 4). The total chlorophyll content was the highest in lettuce plants under treatments WR + FR10 and WR + FR30, followed by WR, WR + FR50, WR + FR70, and WR + FR90, and the lowest under NL, which was attributed to increased chlorophyll a and b contents. The total chlorophyll content in lettuce plants was 97%, 140%, 120%, 77%, 66%, and 40% greater under treatments WR, WR + FR10, WR + FR30, WR + FR50, WR + FR70, and WR + FR90 in comparison to plants under NL, respectively. The chlorophyll a/b of lettuce was the lowest under NL and showed no difference among the six supplemental lighting treatments. The chlorophyll a/b in lettuce plants was 20%, 19%, 17%, 16%, 14%, and 7% greater under treatments WR, WR + FR10, WR + FR30, WR + FR50, WR + FR70, and WR + FR90 in comparison to plants under NL, respectively. Carotenoid content in lettuce plants was increased by 160%, 200%, 220%, 200%, 180%, and 140% under treatments WR, WR + FR10, WR + FR30, WR + FR70, and WR + FR90 compared to plants under NL, respectively.



Figure 4. Photosynthetic pigment contents and chlorophyll a/b ratio of greenhouse-grown lettuce grown under different treatments, including white plus red LEDs with FR photon flux density at 0, 10, 30, 50, 70, and 90 μ mol m⁻² s⁻¹ (WR, WR + FR10, WR + FR30, WR + FR50, WR + FR70, and WR + FR90, respectively), and lettuce grown with natural light only was marked as NL. Means followed by the same lowercase letters are not significantly different for each measured parameter, according to the least-significant difference test ($p \le 0.05$). Error bars indicate the standard deviation (three replicates).

2.3. Growth and Biomass Accumulation of Greenhouse-Grown Lettuce

Leaf number, leaf width, and leaf area of lettuce plants showed a similar trend under supplementary light treatments, which was the highest under treatment WR + FR50, the lowest under NL, and showed no difference among other treatments (Table 2). Leaf number, leaf width, and leaf area of lettuce plants were 44%, 37%, and 84% higher under treatment WR + FR50 than those in plants under NL, respectively. By contrast, supplementary light significantly decreased the leaf length and length-width ratio of lettuce leaves. The specific leaf area of lettuce plants was decreased by supplementary light treatments, no matter with or without FR light addition.

Supplementary light increased the fresh and dry weights, as well as the light use efficiency (LUE) of lettuce plants, compared to those grown with only sunlight (Figures 5 and 6). Shoot and root fresh/dry weight of lettuce plants increased first and decreased subsequently, following a quadratic function with increasing FR photon flux density (Figure 6). Lettuce plants grown under 50 μ mol m⁻² s⁻¹ FR light addition led to greater biomass accumulation than those grown with 10, 30, 70, and 90 μ mol m⁻² s⁻¹ FR light addition. Shoot fresh weight, root fresh weight, shoot dry weight, and root dry weight of lettuce plants were increased by 111%, 303%, 275%, and 462%, respectively, under treatment WR + FR50 compared with those grown under NL. Meanwhile, the LUE of lettuce plants was remarkably prompted by increasing supplementary FR light intensity from 0 to 50 μ mol m⁻² s⁻¹,

and the improvement effect was reduced with FR light intensity over 50 $\mu mol\ m^{-2}\ s^{-1}$ (Figure 7).

Table 2. Leaf morphological traits of greenhouse-grown lettuce under different treatments, including white plus red LEDs with FR photon flux density at 0, 10, 30, 50, 70, and 90 µmol m⁻² s⁻¹ (WR, WR + FR10, WR + FR30, WR + FR50, WR + FR70, and WR + FR90, respectively), and lettuce grown with natural light only was marked as NL. The means followed by the same lowercase letters within the column are not significantly different, according to the least-significant difference test ($p \le 0.05$). Errors indicate the standard deviation (three replicates).

Treatments	Leaf Number		Leaf Length (cm)		Leaf Width (cm)		Length-Width Ratio		Leaf Area (cm ²)	Specific Leaf Area (cm ² g ⁻¹)		
NL	9.0 ± 0.7	с	16.8 ± 0.8	а	12.0 ± 0.6	с	1.40 ± 0.09	а	561 ± 76	d	1121 ± 97	а
WR	11.0 ± 1.0	b	14.2 ± 0.2	b	14.8 ± 1.4	b	0.97 ± 0.09	b	678 ± 131	cd	506 ± 36	b
WR + FR10	11.4 ± 0.5	b	14.0 ± 0.6	bc	15.0 ± 1.1	b	0.94 ± 0.09	bc	745 ± 40	с	469 ± 23	b
WR + FR30	11.8 ± 0.8	b	12.8 ± 0.4	d	15.2 ± 0.6	ab	0.85 ± 0.06	cd	945 ± 73	ab	445 ± 33	b
WR + FR50	13.0 ± 1.2	а	12.6 ± 1.1	d	16.4 ± 0.5	а	0.77 ± 0.07	d	1031 ± 138	а	448 ± 45	b
WR + FR70	11.8 ± 0.8	b	13.2 ± 0.8	cd	14.8 ± 1.3	b	0.90 ± 0.11	bc	811 ± 89	bc	500 ± 17	b
WR + FR90	11.4 ± 0.5	b	13.3 ± 0.8	bcd	14.2 ± 1.2	b	0.95 ± 0.07	bc	730 ± 37	с	507 ± 39	b



Figure 5. Regressions between FR photon flux density and biomass accumulation of greenhousegrown lettuce grown under different treatments, including white plus red LEDs with FR photon flux density at 0, 10, 30, 50, 70, and 90 μ mol m⁻² s⁻¹ (WR, WR + FR10, WR + FR30, WR + FR50, WR + FR70, and WR + FR90, respectively), and lettuce grown with natural light only were marked as NL. Error bars indicate the standard deviation (three replicates).



Figure 6. Light use efficiency of greenhouse-grown lettuce under different treatments, including white plus red LEDs with FR photon flux density at 0, 10, 30, 50, 70, and 90 μ mol m⁻² s⁻¹ (WR, WR + FR10, WR + FR30, WR + FR50, WR + FR70, and WR + FR90, respectively), and lettuce grown with natural light only was marked as NL. Error bars indicate the standard deviation (three replicates).



Figure 7. Vitamin C, reducing sugar, total soluble sugar, and starch content of greenhouse-grown lettuce under different treatments, including white plus red LEDs with FR photon flux density at 0, 10, 30, 50, 70, and 90 µmol m⁻² s⁻¹ (WR, WR + FR10, WR + FR30, WR + FR50, WR + FR70, and WR + FR90, respectively), and lettuce grown with natural light only was marked as NL. Means followed by the same lowercase letters are not significantly different, according to the least-significant difference test ($p \le 0.05$). Error bars indicate the standard deviation (three replicates).

2.4. Vitamin C, Sugar, and Starch Contents of Greenhouse-Grown Lettuce

Supplementary light caused a significant rise in the contents of vitamin C, reducing sugar, total soluble sugar, and starch in lettuce plants compared to plants under NL (Figure 7). The vitamin C content in lettuce plants was the highest under treatments WR + FR10, WR + FR30, and WR + FR50, followed by treatments WR, WR + FR70, and WR + FR90, and the lowest under NL. The vitamin C content in lettuce plants under NL was 33%, 52%, 52%, 53%, 39%, and 35% lower than in plants under treatments WR, WR + FR10, WR + FR30, WR + FR70, and WR + FR90, respectively. The reducing sugar content and starch content in lettuce showed a similar trend, whereas the total soluble sugar content was lower in plants under NL and showed no differences among the six supplementary light treatments.

3. Discussion

Light energy that is absorbed by chlorophyll in photosynthetic systems can undergo three fates: photochemistry, heat dissipation, and fluorescence emission [24]. Thus, chlorophyll fluorescence is commonly used as a tool to provide precise and objective information regarding photochemical efficiency and non-photochemical de-excitation involved in the conversion of light energy under different conditions [25,26]. In the present study, as FR light intensity increased, the value of ΔV_I and ΔV_I of lettuce leaves decreased gradually except for treatment WR + FR90 (Figure 1), indicating less accumulation of PSII acceptor quencher (QA) in the photosynthetic electron transport chain under FR light addition, which promoted electron transport capacity of PSII and CO_2 assimilation [24,27,28]. In the case of the increased chlorophyll fluorescence value of lettuce leaves at the O-I-P point under treatment WR + FR90 (Figure 1), a higher FR photon intensity, or FR proportion, reduced the electron transport capacity of PSII and the proportion of absorbed light energy used for photochemical reactions [27,29]. Furthermore, supplementary light increased the number of active reaction centers as well as the electron transfer efficiency per cross section, which was explained by the decreased ABS/RC and TRo/RC values with similar ABS/CSm and TR_o/CS_m values (Figure 2). This was consistent with the decreased value of Q_B -NRC with increasing FR photon flux density (Figure 3), suggesting an increase in the number of PSII photochemically active reaction centers [30]. The smaller reduction of ET_0/RC compared to ABS/RC and TRo/RC suggested that FR light addition promoted electron transfer efficiency per active reaction center, and its lifting effect increased with FR photon flux density (Figure 2). The photosynthetic performance index PI_{abs} is a performance index based on the absorbed light energy, which mainly reflects the efficiency of the reaction center of PSII, while PI_{total} can further reflect the ability of electron transport between PSII and PSI and the related properties of PSI [31]. The greater increase of PI_{total} than PI_{abs} in lettuce leaves under supplementary light in comparison to NL suggested higher electron transfer efficiency within the photosynthetic electron transport chain, which is positively correlated with FR light intensity except for the FR photon flux density of 90 μ mol m⁻² s⁻¹, due to an overexcitation of PSI to PSII [6,32]. Taken together, enhanced cyclic electron transport and photochemical efficiency by FR light addition exhibited a saturation response to the dose of FR light, and a high FR light intensity of 90 μ mol \cdot m⁻² \cdot s⁻¹ inhibited plant photochemistry efficiency, which is due to an unbalanced excitation between the two photosystems [6,33].

Photosynthesis is the basis for material accumulation, and recent research indicated that adding FR light to PAR wavelengths in both fully controlled environments with artificial lighting and semi-closed facilities with sunlight has a positive influence on leaf and canopy photosynthesis in lettuce, sunflower (*Helianthus annuus* L.), corn (*Zea mays* L.), burdock (*Arctium minus* Bernh.), and Norway maple (*Acer platanoides* L.) plants [7,34]. The $P_{n max}$ represents the photosynthetic potential of a plant, and the larger the value, the more photosynthetic products are synthesized by plants under the same light conditions [27,35]. In the present study, the promotion of $P_{n max}$, R_d , and L_c in lettuce leaves by supplementary light indicated an improved plant's ability to utilize strong light. Although $P_{n max}$ showed no significant difference among different FR light intensities, there was a trend that $P_{n max}$

increased with increasing FR photon intensity from 0 to 50 µmol m⁻² s⁻¹ and decreased gradually over 50 µmol m⁻² s⁻¹ (Table 1, Figure 3). This agreed with previous studies conducted in IVF that at a given level of PAR (PPFD around 200 µmol m⁻² s⁻¹ provided by a combination of red and blue light, photoperiod of 16 h), only a certain amount of FR photons (50 µmol m⁻² s⁻¹) increased the P_n and φ PSII in lettuce plants through better balanced excitation of the two photosystems, and increasing FR photons showed no further improvement or even deterioration [6,8]. The consistent conclusion between our experiment in greenhouses and previous studies in IVFs might be contributed to the similar lighting environment (PPFD around 200 µmol·m⁻²·s⁻¹ with FR photon flux density about 30 to 50 µmol·m⁻²·s⁻¹) among different experiments [6,8,9]. In the present study, the lighting environment in the greenhouse during FR light application was mainly provided by supplementary artificial lighting, which was applied for a photoperiod of 12 h d⁻¹ (04:00–10:00 and 14:00–20:00) from late Nov. to Dec. (sunrise around 07:00 and sunset around 16:40), during which the sunlight intensity in the greenhouse (including PAR and FR wavelength) is mostly lower than 50 µmol m⁻² s⁻¹.

FR light addition adjusts light-harvesting structure to increase light harvesting ability and carbon assimilation of plants, which was attributed to plant shade-avoidance or shade-tolerance responses [36,37]. Shade-avoidance responses often manifest as greater hypocotyl/stem/petiole elongation and reduced branching under shade, while shadetolerance responses show leaf expansion with reduced leaf thickness [38]. A significant shade-tolerance response to leaf expansion by supplementary FR light was observed in this study, which was mainly caused by increased leaf number and/or leaf width instead of leaf length (Table 2). Similar effects by FR light addition as photoperiodic lighting in lettuce and other plant species such as geranium (*Pelargonium* \times *hortorum*), snapdragon (Antirrhinum majus), radish (Raphanus sativus), and kale (Brassica napus) were observed in previous studies in IVF, which facilitated better light interception and led to higher plant biomass accumulation [13,37-40]. Jin et al. investigated the underlying components of FR effects (52 μ mol m⁻² s⁻¹ FR light as photoperiodic lighting added to R&B light of 218 μ mol $m^{-2} s^{-1}$) on lettuce growth, and results indicated that FR increased plant dry weight by 46–77%, which was mainly due to a higher canopy-intercepted PPFD caused by increased leaf area, and to a smaller extent (8-23%) by higher intercepted LUE [9]. As Legendre and van Iersel stated, each 1 m² increase in lettuce leaf area was associated with a 59 mol increase in incident light after 25 days of growth [10]. Thus, this can be a self-reinforcing process: plants under FR light treatment developed a larger leaf area to absorb more light, grow faster, and produce additional leaf area faster than plants under treatment without FR light [41,42]. Specific leaf area has often been found to increase with additional FR light at a shorter wavelength [13,43], but not in the current research or study by Jin et al., which may be due to varied FR effects on different lettuce species and cultivars [9,44]. In this study, supplementary light was used during 04:00–10:00 and 14:00–20:00 (sunrise around 07:00 and sunset around 16:40 during the experiment), which meant the extended photoperiod was applied and the natural photoperiod was extended by approximately 3 h in the beginning and at the end of the natural daylight period, respectively, leading to the bigger leaf area and higher dry weight of lettuce (Table 2 and Figure 5). Zou et al. reported that adding FR to red plus blue light either during the day (16 h) or end-of-day (EOD, 1 h) improved biomass production of lettuce compared with those grown without FR; however, additional FR light at EOD led to lower leaf area and lower dry weight of lettuce than those grown under FR during the day [13]. The differences may be related to the supplementary light duration. In addition, with the same light intensity and duration (50 μ mol m⁻² s⁻¹ for 1 h), applying FR at the EOD without PAR resulted in a bigger leaf area and higher biomass of lettuce compared with those FR lights applied at the EOD with PAR, indicating that FR light, with the appropriate supplementary time, proved to be more effective in enhancing growth and improving radiation use efficiency [6].

Notably, supplementary FR light to short wavelengths means a decreased ratio of R:FR, which is linked to many light effects on plant morphogenesis, growth, and metabolic

processes mediated by phytochromes [45–47]. Phytochromes, which exist in two photointerconvertible isomeric forms: a red-light-absorbing form (Pr, $\lambda max = 660$ nm, biologically inactive form) and a FR-light-absorbing form (Pfr, $\lambda max = 730$ nm, biologically active form), enable plants to sense and adapt to the light environment, thereby improving energy efficiency, growth patterns, and resilience to abiotic stresses [37,45,47]. For example, a lower R:FR condition improved tomato salinity stress tolerance, and phytochrome B1 plays an important role in this process [47]. In the present study, the R:FR ratio provided by treatments WR, WR + FR10, WR + FR30, WR + FR50, WR + FR70, and WR + FR90 was 15.72, 3.75, 1.49, 0.93, 0.67, and 0.53, respectively. It clearly indicated that a decreased R:FR ratio (no lower than 0.93) boosted lettuce growth and biomass accumulation through both morphological and photochemical effects. Interestingly, this R:FR ratio of 0.93 (WR + FR50) is similar to the R:FR ratio of sunlight at noon (around 1.0 to 1.3), which is proven suitable for plant growth as it supports optimal phytochrome activity, efficient photosynthesis, and favorable morphological traits [48]. Further, this was consistent with the suggestion by Zhen and Bugbee that a FR proportion of 35% (the R:FR ratio was not provided) increased lettuce photosynthetic rates [7]. However, Kusuma and Bugbee found a positive effect of FR on leaf area and dry mass accumulation at high PPFD but a negative effect on these growth parameters at lower PPFD, suggesting interactive effects of PPFD and FR light addition on lettuce growth, which have previously been described for other crops [11,29]. Furthermore, Zou et al. stated that the acclimation process of plant morphology triggered by FR addition made a major contribution to the yield improvement of lettuce in IVF since FR light decreased the plant's photosynthetic capacity in the long term due to a lower chlorophyll and total nitrogen content, along with decreased leaf light absorption [13]. The decreased chlorophyll content by FR light addition was largely reported in previous literature [15,39,40,43], while in the present study, this reduction effect on chlorophyll contents was only observed with FR photon flux density over 50 μ mol·m⁻²·s⁻¹ (Figure 4). This might be the reason for the inconsistency of the optimal FR photon flux density for plant photochemical efficiency and biomass accumulation, which was 70 μ mol m⁻² s⁻¹ and 30~50 μ mol m⁻² s⁻¹, respectively, since high FR photon flux density over 50 μ mol $m^{-2} s^{-1}$ significantly decreased leaf chlorophyll content, thus leaf light absorption and LUE. Taken together, the varied background light intensity, different spectral compositions, and proportion of FR photons in total PPFD, as well as lettuce species and cultivars, all affect plant responses to supplementary FR light. According to the results of current research, in the case of greenhouse-grown lettuce produce with commonly used PPFD of 200 μ mol·m⁻²·s⁻¹, FR light intensity of 30~50 μ mol m⁻² s⁻¹ or a FR proportion of 15~25% was suggested.

It has been widely reported that FR light addition or a low R:FR ratio would increase plant carbohydrate content, such as starch and sucrose content, mainly due to improved photochemical efficiency [8,12,49]. The changes in trends of vitamin C, reducing sugar content, and starch content were similar to the $P_{n max}$, light use efficiency, and total chlorophyll content of lettuce in this study (Figure 7), indicating that improved photosynthetic capacity and relative CO₂ assimilation by FR light addition possibly contributed to increased carbohydrate contents. Further, high levels of carbohydrates improve the sweetness and crispness of lettuce, which may enhance plant sensory quality and shelf life after harvest [50,51]. However, a decreased content of carotenoids and total phenolics under FR light addition was also reported in previous studies [39,52,53]. This reduction might cause crops to be more susceptible to pests and pathogens, and fungi [43,52]. This might be a result of plants investing resources in the most efficient way into plant expansion growth instead of phytochemical synthesis [38,54]. Further experiments are required to clarify how FR light influences the synthesis of these secondary metabolites.

4. Materials and Methods

4.1. Plant Materials

Lettuce 'Dasusheng' (*Lactuca sativa* L.) was selected as the experimental material. All lettuce seeds were grown in 72-cell plug trays filled with mixed peat (The Pindstrup Group, Kongersle, Denmark), vermiculite (Shandong Lige Technology Co., Ltd., Jinan, China), and perlite (Shandong Lige Technology Co., Ltd., Jinan, China) (3:1:1, v/v/v) in a commercial greenhouse. Lettuce seedlings with four true leaves were selected for uniformity and transplanted into 1.8 L plastic pots (diameter, 16.8 cm; depth, 14 cm) one day before treatment. The plastic pots were filled with mixed peat, vermiculite, and perlite (3:1:1, v/v/v) and kept in a Venlo-type greenhouse at Qingdao Agricultural University (36°19' N, 120°23' E), Qingdao, China. The experiment was conducted for 35 days (from 21 November to 25 December 2023), and the air temperature was maintained at 22 ± 3 °C/18 ± 3 °C (day/night), while the relative humidity was maintained at 60~70%. Lettuce was cultivated using Hoagland's nutrient solution according to a previously reported method [55].

4.2. Light Treatment Design

A randomized complete block design was implemented to evaluate the effect of supplementary FR light at shorter wavelengths on greenhouse-grown lettuce. According to our previous study, white plus red LEDs (WR, color temperature of 6500 K, provided by Zhongshan Aier Lighting Technology Co., Ltd., Zhongshan, China) with a DLI of $6.52 \text{ mol m}^{-2} \text{ d}^{-1}$ were suitable for lettuce growth [55]. Therefore, in this experiment, six supplemental light treatments with different FR light intensities: WR LEDs without FR light addition and WR LEDs with FR (Xiamen Lumigro Technology Co., Ltd., Xiamen, China) photon flux density at 10, 30, 50, 70, and 90 μ mol m⁻² s⁻¹ (WR + FR10, WR + FR30, WR + FR50, WR + FR70, and WR + FR90, respectively) were implemented. Lettuce grown with only natural light was marked as NL. The PPFD provided by WR LEDs was set at 151 μ mol m⁻² s⁻¹, and the supplementary lighting was carried out in two time slots, 04:00-10:00 and 14:00-20:00 (sunrise around 07:00 and sunset around 16:40), during which the sunlight intensity in the greenhouse is mostly lower than 50 μ mol m⁻² s⁻¹. The average DLI of solar light inside the greenhouse was 5.0 mol m⁻² d⁻¹, while the DLI of supplemental light treatments was 6.52 mol m⁻² d⁻¹. For treatment WR, WR + FR10, WR + FR30, WR + FR50, WR + FR70, and WR + FR90, the R:FR ratio provided by LEDs was 15.72, 3.75, 1.49, 0.93, 0.67, and 0.53, respectively. The spectral distribution of each treatment was measured with a spectrometer (AvaSpec-ULS2048-USB2, Avantes Inc., Apeldoorn, The Netherlands) above the plant canopy (Table 3 and Figure 8). The peak wavelengths of the WR LEDs and FR light lamps are 446 nm (B), 633 nm (R), and 730 nm (FR), respectively. In this experiment, each treatment was replicated in three blocks. For each block, 20 lettuce plants were cultivated with a planting density of 25 plants m⁻². At harvest, three uniform plants were randomly selected in each block for measurement.

Table 3. The photosynthetic photon flux density (PPFD, 400–700 nm), ratio of blue (B, 400–499 nm), green (G, 500–599 nm), and red (R, 600–699 nm) photon flux density, far-red photon flux density (FR, 700–800 nm), and R:FR ratio of different supplementary lighting treatments, including white plus red LEDs with FR photon flux density at 0, 10, 30, 50, 70, and 90 μ mol m⁻² s⁻¹ (WR, WR + FR10, WR + FR30, WR + FR50, WR + FR70, and WR + FR90, respectively).

Treatments	PPFD (µmol m ⁻² s ⁻¹)	Ratio of B:G:R	FR Photon Flux Density (μmol m ⁻² s ⁻¹)	Ratio of R:FR
WR	151		3	15.72
WR + FR10		110/114	13	3.75
WR + FR30			33	1.49
WR + FR50		1:1.36:1.14	53	0.93
WR + FR70			73	0.67
WR + FR90			93	0.53



Figure 8. Spectral distribution of supplementary white-red (WR) light-emitting diodes (LEDs) and far-red LEDs (FR).

4.3. Growth Measurement

4.3.1. Photosynthetic Characteristics, Pigments, and Chlorophyll Fluorescence

A light response curve was determined by a portable photosynthesis system (Li-6400XT, LI-COR Corporation, Lincoln, NE, USA) for lettuce leaves. Measurement was conducted in clear weather, and PPFD inside the leaf chamber was set to eight gradients of 2000, 1500, 1000, 500, 200, 100, 50, and 0 μ mol m⁻² s⁻¹, respectively. Leaf temperature, gas flow rate, and the CO₂ concentration inside the leaf chamber were set at 22 °C, 500 μ mol s⁻¹, and 400 μ mol mol⁻¹, respectively. The maximum net photosynthetic rate (P_{n max}), dark respiration rate (R_d), and light compensation point (L_c) were calculated based on the light response curve according to the Ye Model [56].

Chlorophyll fluorescence parameters of lettuce were measured at harvest using a pocket Plant Efficiency Analyzer chlorophyll fluorimeter (PEA, Hansatech Instruments Ltd., Norfolk, UK). Leaves were darkly adapted for at least 30 min prior to a rapid chlorophyll -a fluorescence induction kinetic curve (OJIP curve) measurement. Calculation of chlorophyll a fluorescence parameters including F_v/F_m , ABS/RC, TR_o/RC , ET_o/RC , DI_o/RC , ET_o/CS_m , DI_o/CS_m , ABS/CS_m and TR_o/CS_m , PI_{abs} , and PI_{total} were based on Li et al. [57]. The proportion of QB-NRC was calculated based on the method reported by Klinkovsky and Naus [58]. To help visualize the influence of supplementary light on chlorophyll a fluorescence transients of lettuce leaves, the value of relative variable fluorescence (ΔV_t) were calculated by subtracting the values of the fluorescence recorded in plants under supplementary treatments from those recorded for NL plants.

At harvest, three plants were selected for the measurement of chlorophyll content and carotenoids' content within each block, and the third leaf from the top was selected for measurement. Fresh lettuce leaves were cut into small pieces and then extracted in 80% acetone (v/v) for 24 h. The absorbance of the extracted supernatants was measured at 663, 645, and 470 nm, respectively, using a UV-VIS spectrophotometer (1810, Shanghai Yoke Instrument Co., Ltd., Shanghai, China). The concentrations of chlorophyll a, chlorophyll b, and carotenoids were calculated according to Lichtenthaler and Wellburn [59] and expressed as mg g⁻¹ (fresh weight). The total chlorophyll content and ratio of chlorophyll a/b were calculated accordingly.

4.3.2. Plant Morphology and Growth Characteristics

Leaf number, leaf length and width of the maximum leaf blade, and shoot and root fresh weight (using an electronic analytical balance, JY20002, Shanghai Hengping Instrument Co., Ltd., Shanghai, China) were recorded at harvest (35 days after treatment). Leaf area was measured by a leaf area scanner (Yaxin-1241, Beijing Yaxin Liyi Technology Co.,

Ltd., Beijing, China). Fresh leaves and roots were dried in an oven at 105 $^{\circ}$ C for 3 h, and then dried at 80 $^{\circ}$ C for 72 h to measure the shoot and root dry weight.

4.3.3. Vitamin C, Reducing Sugar, Soluble Sugar, and Starch Content

At harvest, fresh lettuce leaves (the third leaf from the top) were selected for the measurement of vitamin C using the 2, 6-dichlorophenol indophenol titration method according to Shyamala and Jamuna [60] and expressed as mg 100g⁻¹ (fresh weight). Dried leaf samples were selected for the measurement of soluble sugar content and reducing sugar content using the anthronesulfuric acid colorimetry method and the 5-dinitrosalicylic acid colorimetric method, according to Song et al. [61] and Zhan et al. [62], respectively. The starch content of dried leaf samples was measured according to Takahashi et al. [63]. The measurement of soluble sugar, reducing sugar, and starch contents was done using a UV-VIS spectrophotometer (1810, Shanghai Yoke Instrument Co., Ltd., Shanghai, China) and was expressed as a percentage (dry weight).

4.4. Statistical Analysis

Data collected from the three blocks for each treatment were analyzed, and one-way analysis of variance (ANOVA) and the least significant difference (LSD) test ($p \le 0.05$) were carried out using SPSS 26.0 software (IBM, Inc., Chicago, IL, USA) to reveal the difference among groups. The data were expressed as the means \pm standard deviations (SD). Regressions between treatments and the morphological characteristics of lettuce were performed using Microsoft Excel 2021 software.

5. Conclusions

Our results demonstrated that the addition of FR light to the PAR wavelength differentially influenced the photochemical efficiency of PSII as well as the electron transfer efficiency between PSII and PSI, which was positively correlated with the FR photon flux density from 10 to 70 µmol m⁻² s⁻¹. However, shoot fresh weight of lettuce plants was increased by 26%, 19%, 24%, and 37%, respectively, under treatment WR + FR50 compared with those grown under 10, 30, 70, and 90 µmol m⁻² s⁻¹ FR light addition. A similar trend was observed: the contents of vitamin C and carbohydrates in lettuce were enriched the most under FR addition of 30~50 µmol m⁻² s⁻¹. This is probably because the FR photon flux density of 70~90 µmol m⁻² s⁻¹ significantly decreased leaf total chlorophyll content by 7~21% compared to treatment WR + FR50, thus decreasing leaf light absorption and LUE accordingly. Therefore, with commonly used growing light of PPFD around 200 µmol m⁻² s⁻¹, addition of FR light as photoperiodic light around 30~50 µmol m⁻² s⁻¹ was suggested to enhance the photochemistry efficiency, biomass accumulation, and carbohydrate contents in greenhouse-grown lettuce.

Supplementary Materials: The following supporting information can be downloaded at: https: //www.mdpi.com/article/10.3390/plants13152169/s1, Figure S1: Photosynthetic response curves of lettuce leaves under different treatments, including white plus red LEDs with FR photon flux density at 0, 10, 30, 50, 70, and 90 µmol m⁻² s⁻¹ (WR, WR + FR10, WR + FR30, WR + FR50, WR + FR70, and WR + FR90, respectively), and lettuce grown with natural light only was marked as NL.

Author Contributions: Conceptualization, methodology, and investigation, X.L., Z.L., H.D. and Z.Y.; resources and data curation, J.S., Y.Y. and Z.Y.; writing and editing, H.D., X.L. and Y.Y.; funding acquisition, Y.Y. and Z.Y. All authors have read and agreed to the published version of the manuscript.

Funding: This work was financially supported by the Shandong Provincial Natural Science Foundation (ZR2021QC174); the Key Research and Development Program of Shandong Province (2021TZXD007); the Modern Agricultural Industrial Technology System of Shandong Province (SDAIT-05); the Innovation and Entrepreneurship Training Program for Postgraduate in QAU (QNYCX23088).

Data Availability Statement: Data are contained in the article. Please contact the corresponding author for any additional information.

Acknowledgments: The authors thank Geng Zhang at Jiangsu Vocational College of Agriculture and Forestry for her help with the manuscript.

Conflicts of Interest: The authors declare that they have no conflicts of interest.

References

- 1. Wang, J.; Lu, W.; Tong, Y.; Yang, Q. Leaf morphology, photosynthetic performance, chlorophyll fluorescence, stomatal development of lettuce (*Lactuca sativa* L.) exposed to different ratios of red light to blue light. *Front Plant. Sci.* **2016**, *10*, 250.
- 2. Dou, H.; Niu, G.; Gu, M.; Masabni, J.G. Responses of sweet basil to different daily light integrals in photosynthesis, morphology, yield, and nutritional quality. *HortScience* **2018**, *53*, 496–503. [CrossRef]
- Pennisi, G.; Orsini, F.; Blasioli, S.; Cellini, A.; Crepaldi, A.; Braschi, I.; Spinelli, F.; Nicola, S.; Fernandez, J.A.; Stanghellini, C. Resource use efficiency of indoor lettuce (*Lactuca sativa* L.) cultivation as affected by red:blue ratio provided by LED lighting. *Nat. Sci. Rep.* 2019, 9, 14127.
- 4. Spalholz, H.; Perkins-Veazie, P.; Hernández, R. Impact of sun-simulated white light and varied blue:red spectrums on the growth, morphology, development, and phytochemical content of green- and red-leaf lettuce at different growth stages. *Sci. Hortic.* 2020, 264, 109195. [CrossRef]
- 5. McCree, K.J. The action spectrum, absorptance and quantum yield of photosynthesis in crop plants. *Agric. Meteorol.* **1972**, *9*, 191–216. [CrossRef]
- 6. Zou, J.; Fanourakis, D.; Tsaniklidis, G.; Cheng, R.; Yang, Q.; Li, T. Lettuce growth, morphology and critical leaf trait responses to far-red light during cultivation are low fluence and obey the reciprocity law. *Sci. Hortic.* **2021**, *289*, 110455. [CrossRef]
- 7. Jin, W.; Urbina, J.L.; Heuvelink, E.; Marcelis, L.F.M. Adding far-red to red-blue light-emitting diode light promotes yield of lettuce at different planting densities. *Front. Plant Sci.* 2021, *11*, 609977. [CrossRef] [PubMed]
- Zhen, S.; Van Iersel, M.W. Far-red light is needed for efficient photochemistry and photosynthesis. J. Plant Physiol. 2017, 209, 115–122. [CrossRef] [PubMed]
- 9. Zhen, S.; Bugbee, B. Far-red photons have equivalent efficiency to traditional photosynthetic photons: Implications for redefining photosynthetically active radiation. *Plant Cell Environ.* **2020**, *43*, 1259–1272. [CrossRef] [PubMed]
- 10. Legendre, R.; van Iersel, M.W. Supplemental far-red light stimulates lettuce growth: Disentangling morphological and physiological effects. *Plants* **2021**, *10*, 166. [CrossRef] [PubMed]
- 11. Kusuma, P.; Bugbee, B. On the contrasting morphological response to far-red at high and low photon fluxes. *Front. Plant Sci.* 2023, 14, 1185622. [CrossRef]
- Ji, Y.; Ouzounis, T.; Courbier, S.; Kaiser, E.; Nguyen, P.T.; Schouten, H.J.; Visser, R.G.F.; Pierik, R.; Marcelis, L.F.M.; Heuvelink, E. Far-red radiation increases dry mass partitioning to fruits but reduces *Botrytis cinerea* resistance in tomato. *Environ. Exp. Bot.* 2019, 168, 103889. [CrossRef]
- 13. Zou, J.; Zhang, Y.; Zhang, Y.; Bian, Z.; Fanourakis, D.; Yang, Q.; Li, T. Morphological and physiological properties of indoor cultivated lettuce in response to additional far-red light. *Sci. Hortic.* **2019**, 257, 108725. [CrossRef]
- 14. Wong, C.; Teo, Z.W.N.; Shen, L.; Yu, H. Seeing the lights for leafy greens in indoor vertical farming. *Trends Food Sci. Technol.* 2020, 106, 48–63. [CrossRef]
- 15. Kong, Y.; Nemali, K. Blue and far-red light affect area and number of individual leaves to influence vegetative growth and pigment synthesis in lettuce. *Front. Plant Sci.* **2021**, *12*, 667407. [CrossRef]
- 16. Zou, J.; Fanourakis, D.; Tsaniklidis, G.; Woltering, E.J.; Cheng, R.; Li, T. Far-red radiation during indoor cultivation reduces lettuce nutraceutical quality and shortens the shelf-life when stored at supra optimal temperatures. *Postharvest Biol. Technol.* **2023**, *198*, 112269. [CrossRef]
- 17. Gao, H.; Gong, L.; Ni, J.; Li, Q. Metabolomics analysis of lettuce (*Lactuca sativa* L.) affected by low potassium supply. *Agriculture* **2022**, *12*, 1153.
- 18. Zhao, J.; Li, H.; Chen, C.; Pang, Y.; Zhu, X. Detection of water content in lettuce canopies based on hyperspectral imaging technology under outdoor conditions. *Agriculture* **2022**, *12*, 1796. [CrossRef]
- Li, Q.; Gao, H.; Zhang, X.; Ni, J.; Mao, H. Describing lettuce growth using morphological features combined with nonlinear models. *Agronomy* 2022, 12, 860. [CrossRef]
- THECORNERPLOT. Available online: https://thecornerplot.blog/2023/07/28/exploring-the-global-production-of-lettucewhere-does-it-grow/ (accessed on 15 March 2024).
- Paz, M.; Fisher, P.R.; Gómez, C. Minimum light requirements for indoor gardening of lettuce. Urban Agric. Reg. Food Syst. 2019, 4, 1–10. [CrossRef]
- 22. Zhang, M.; Whitman, C.M.; Runkle, E.S. Manipulating growth, color, and taste attributes of fresh cut lettuce by greenhouse supplemental lighting. *Sci. Hortic.* 2019, 252, 274–282. [CrossRef]
- 23. Yan, Z.; Wang, C.; Li, Z.; Li, X.; Cheng, F.; Lin, D.; Yang, Y. Supplementary white, UV-A, and far-red radiation differentially regulates growth and nutritional qualities of greenhouse lettuce. *Plants* **2023**, *12*, 3234. [CrossRef]
- 24. Misra, A.N.; Misra, M.; Singh, R. Chlorophyll fluorescence in plant biology. *Biophysics* 2012, 7, 171–192.
- Strasser, R.J.; Srivastava, A.; Tsimilli-Michael, M. The fluorescence transient as a tool to characterize and screen photosynthetic samples. *Probing Photosynth. Mech. Regul. Adapt.* 2000, 25, 445–483.

- Zhang, C.; Akhlaq, M.; Yan, H.; Ni, Y.; Liang, S.; Zhou, J.; Xue, R.; Li, M.; Adnan, M.R.; Li, J. Chlorophyll fluorescence parameter as a predictor of tomato growth and yield under CO₂ enrichment in protective cultivation. *Agricul. Water Manag.* 2023, 284, 108333. [CrossRef]
- 27. Chen, X.; Li, Y.; Wang, L.; Yang, Q.; Guo, W. Responses of butter leaf lettuce to mixed red and blue light with extended light/dark cycle period. *Sci. Rep.* 2022, 12, 6924. [CrossRef] [PubMed]
- Shamsabad, M.R.M.; Esmaeilizadeh, M.; Roosta, H.R.; Dehghani, M.R.; Dąbrowski, P.; Kalaji, H.M. The effect of supplementary light on the photosynthetic apparatus of strawberry plants under salinity and alkalinity stress. *Sci. Rep.* 2022, *12*, 13257. [CrossRef] [PubMed]
- 29. Yuan, J.; Ma, C.; Feng, Y.; Zhang, J.; Yang, F.; Li, Y. Response of chlorophyll fluorescence transient in leaves of wheats with different drought resistances to drought stresses and rehydration. *Plant Physiol. J.* **2018**, *54*, 1119–1129.
- Khudyakova, A.Y.; Kreslavski, V.D.; Shmarev, A.N.; Lyubimov, V.Y.; Shirshikova, G.N.; Pashkovskiy, P.P.; Kuznetsov, V.V.; Allakhverdiev, S.I. Impact of UV-B radiation on the photosystem II activity, pro-/antioxidant balance and expression of lightactivated genes in *Arabidopsis thaliana* hy4 mutants grown under light of different spectral composition. *J. Photochem. Photobiol. B Biol.* 2019, 194, 14–20. [CrossRef] [PubMed]
- Stefanov, D.; Petkova, V.; Denev, I.D. Screening for heat tolerance in common bean (*Phaseolus vulgaris* L.) lines and cultivars using JIP-test. *Sci. Hortic.* 2011, 128, 1–6.
- 32. Wang, Q.; Ning, Z.; Awan, S.A.; Gao, J.; Chen, J.; Lei, Y.; Tan, X.; Wu, X.; Wu, Y.; Liu, C.; et al. Far-red light mediates light energy capture and distribution in soybeans (*Glycine max* L.) under the shade. *Plant Physiol. Biochem.* **2023**, 204, 108130.
- 33. Kono, M.; Kawaguchi, H.; Mizusawa, N.; Yamori, W.; Suzuki, Y.; Terashima, I. Far-red light accelerates photosynthesis in the low-light phases of fluctuating light. *Plant Cell Physiol.* **2019**, *61*, 192–202. [CrossRef] [PubMed]
- Zhen, S.; van Iersel, M.W.; Bugbee, B. Photosynthesis in sun and shade: The surprising importance of far-red photons. *New Phytol.* 2022, 236, 538–546. [CrossRef] [PubMed]
- Zhang, Y.; Huang, Z.; Li, Y.; Lu, X.; Li, G.; Qi, S.; Khan, I.; Li, G.; Dai, Z.; Du, D. The degradability of microplastics may not necessarily equate to environmental friendliness: A case study of cucumber seedlings with disturbed photosynthesis. *Agriculture* 2024, 14, 53. [CrossRef]
- 36. Franklin, K.A. Shade avoidance. New Phytol. 2010, 179, 930–944. [CrossRef]
- 37. Park, Y.; Runkle, E.S. Far-red radiation promotes growth of seedlings by increasing leaf expansion and whole-plant net assimilation. *Environ Exp. Bot.* 2017, 136, 41–49. [CrossRef]
- 38. Meng, Q.; Kelly, N.; Runkle, E. Substituting green or far-red radiation for blue radiation induces shade avoidance and promotes growth in lettuce and kale. *Environ. Exp. Bot.* **2019**, *162*, 383–391. [CrossRef]
- Li, Q.; Kubota, C. Effects of supplemental light quality on growth and phytochemicals of baby leaf lettuce. *Environ. Exp. Bot.* 2009, 67, 59–64. [CrossRef]
- 40. Meng, Q.; Runkle, E. Far red radiation interacts with relative and absolute blue and red photon flux densities to regulate growth, morphology, and pigmentation of lettuce and basil seedlings. *Sci. Hortic.* **2019**, 255, 269–280. [CrossRef]
- 41. Elkins, C.; van Iersel, M.W. Longer photoperiods with the same daily light integral improve growth of Rudbeckia seedlings in a greenhouse. *HortScience* **2020**, *55*, 1676–1682. [CrossRef]
- 42. Weaver, G.; van Iersel, M.W. Longer photoperiods with adaptive lighting control can improve growth of greenhouse-grown 'Little Gem' lettuce (*Lactuca sativa*). *HortScience* 2020, *55*, 573–580. [CrossRef]
- 43. Van de Velde, E.; Steppe, K.; Van Labeke, M.C. Leaf morphology, optical characteristics and phytochemical traits of butterhead lettuce affected by increasing the far-red photon flux. *Front. Plant Sci.* **2023**, *14*, 1129335. [CrossRef] [PubMed]
- 44. Liu, J.; van Iersel, M.W. Far-red light effects on lettuce growth and morphology in indoor production are cultivar specific. *Plants* **2022**, *11*, 2714. [CrossRef]
- 45. Trupkin, S.A.; Legris, M.; Buchovsky, A.S.; Rivero, M.B.T.; Casal, J.J. Phytochrome B nuclear bodies respond to the low red to far-red ratio and to the reduced irradiance of canopy shade in *Arabidopsis*. *Plant Physiol.* **2014**, *165*, 1698–1708. [CrossRef]
- 46. Shmarev, A.; Vereshagin, M.; Pashkovskiy, P.; Kreslavski, V.D.; Allakhverdiev, S.I. Influence of additional far-red light on photosynthetic and growth parameters of lettuce plants and the resistance of the photosynthetic apparatus to high irradiance. *Photosynthetica* **2024**, *62*, 180–186. [CrossRef]
- 47. Cao, K.; Yu, J.; Xu, D.; Ai, K.; Bao, E.; Zou, Z. Exposure to lower red to far-red light ratios improve tomato tolerance to salt stress. BMC Plant Biol. 2018, 18, 92. [CrossRef] [PubMed]
- 48. Holmes, M.G.; Smith, H. The function of phytochrome in the natural environment. I. Characterization of daylight for studies in photomorphogenesis and photoperiodism. *Photochem. Photobiol.* **1977**, *25*, 533–538. [CrossRef]
- Yang, F.; Liu, Q.; Cheng, Y.; Feng, L.; Wu, X.; Fan, Y.; Raza, M.A.; Wang, X.; Yong, T.; Liu, W.; et al. Low red/far-red ratio as a signal promotes carbon assimilation of soybean seedlings by increasing the photosynthetic capacity. *BMC Plant Biol.* 2020, 20, 148. [CrossRef] [PubMed]
- 50. Lin, K.; Huang, M.; Huang, W.; Hsu, M.H.; Yang, C. The effects of red, blue, and white light-emitting diodes on the growth, development, and edible quality of hydroponically grown lettuce (*Lactuca sativa* L. var. *capitata*). *Sci. Hortic.* **2013**, *150*, 86–91.
- Woltering, E.J.; Witkowska, I.M. Effects of pre- and postharvest lighting on quality and shelf life of fresh-cut lettuce. *Acta. Hortic.* 2016, 1134, 357–366. [CrossRef]
- 52. Ballaré, C.L. Light regulation of plant defense. Annu. Rev. Plant Biol. 2014, 65, 335–363. [CrossRef] [PubMed]

- Mickens, M.A.; Skoog, E.J.; Reese, L.E.; Barnwell, P.L.; Spencer, L.E.; Massa, G.D.; Wheeler, R.M. A strategic approach for investigating light recipes for 'Outredgeous' red romaine lettuce using white and monochromatic LEDs. *Life Sci. Space Res.* 2018, 19, 53–62. [CrossRef] [PubMed]
- Shibuya, T.; Endo, R.; Hayashi, N.; Kitaya, Y. High-light-like photosynthetic responses of *Cucumis sativus* leaves acclimated to fluorescent illumination with a high red:far-red ratio: Interaction between light quality and quantity. *Photosynthetic* 2012, 50, 623–629. [CrossRef]
- 55. Yan, Z.; He, D.; Niu, G.; Zhou, Q.; Qu, Y. Growth, nutritional quality and energy use efficiency of hydroponic lettuce as influenced by daily light integrals exposed to white versus white plus red light-emitting diodes. *HortScience* 2019, *54*, 1737–1744. [CrossRef]
- 56. Ye, Z.; Suggett, D.; Robakowski, P.; Kang, H. A mechanistic model for the photosynthesis light response based on the photosynthetic electron transport of photosystem II in C3 and C4 species. *New Phytol.* **2013**, *199*, 110–120. [CrossRef]
- 57. Li, Q.; Chen, L.; Jiang, H.; Tang, N.; Yang, L.; Lin, Z.; Li, Y. Effects of manganese-excess on CO₂ assimilation, ribulose-1,5bisphosphate carboxylase/oxygenase, carbohydrates and photosynthetic electron transport of leaves, and antioxidant systems of leaves and roots in Citrus grandis seedlings. *BMC. Plant Biol.* **2010**, *10*, 42. [CrossRef]
- 58. Klinkovsky, T.; Naus, J. Sensitivity of the relative Fpl level of chlorophyll fluorescence induction in leaves to the heat stress. *Photosynth. Res.* **1994**, *39*, 201–204. [CrossRef] [PubMed]
- 59. Lichtenthaler, H.K.; Wellburn, A. Determinations of total carotenoids and chlorophylls a and b of leaf extracts in different solvents. *Biochem. Soc. Trans.* **1983**, *603*, 591–592. [CrossRef]
- 60. Shyamala, B.J.; Jamuna, P. Nutritional content and antioxidant properties of pulp waste from Daucus carota and Beta vulgaris. *Malays. J. Nutr.* **2010**, *16*, 397–408. [PubMed]
- 61. Song, J.; Huang, H.; Hao, Y.; Song, S.; Liu, H. Nutritional quality, mineral and antioxidant content in lettuce affected by interaction of light intensity and nutrient solution concentration. *Sci. Rep.* **2020**, *10*, 2796. [CrossRef]
- 62. Zhan, L.; Hu, J.; Ai, Z.; Pang, L.; Li, Y.; Zhu, M. Light exposure during storage preserving soluble sugar and l-ascorbic acid content of minimally processed romaine lettuce (*Lactuca sativa* L.var. *longifolia*). *Food Chem.* **2012**, 136, 273–278.
- 63. Takahashi, K.; Fujino, K.; Kikuta, Y.; Koda, Y. Involvement of the accumulation of sucrose and the synthesis of cell wall polysaccharides in the expansion of potato cells in response to jasmonic acid. *Plant Sci.* **1995**, *111*, 11–18. [CrossRef]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.