



# **Weed Role for Pollinator in the Agroecosystem: Plant–Insect Interactions and Agronomic Strategies for Biodiversity Conservation**

**Stefano Benvenuti**

*Review*

Department of Agriculture, Food and Environment, University of Pisa, Via del Borghetto, 80, 56124 Pisa, Italy; stefano.benvenuti@unipi.it

**Abstract:** The growing interest in safeguarding agroecosystem biodiversity has led to interest in studying ecological interactions among the various organisms present within the agroecosystem. Indeed, mutualisms between weeds and pollinators are of crucial importance as they influence the respective survival dynamics. In this review, the mutualistic role of flower visitors and the possible (often predominant) abiotic alternatives to insect pollination (self- and wind-pollination) are investigated. Mutualistic relations are discussed in terms of reward (pollen and/or nectar) and attractiveness (color, shape, scent, nectar quality and quantity), analyzing whether and to what extent typical weeds are linked to pollinators by rigid (specialization) or flexible (generalization) mutualistic relations. The entomofauna involved is composed mainly of solitary and social bees, bumblebees, Diptera, and Lepidoptera. While some of these pollinators are polylectic, others are oligolectic, depending on the shape of their mouthparts, which can be suited to explore the flower corollas as function of their depths. Consequently, the persistence dynamics of weed species show more successful survival in plants that are basically (occasional insect pollination) or totally (self and/or wind pollination) unspecialized in mutualistic relations. However, even weed species with typical abiotic pollination are at times visited during periods such as late summer, in which plants with more abundant rewards are insufficiently present or completely absent. Many typically insect-pollinated weeds can represent a valid indicator of the ecological sustainability of crop management techniques, as their survival dynamics are closely dependent on the biodiversity of the surrounding entomofauna. In particular, the presence of plant communities of species pollinated above all by butterflies (e.g., several Caryophyllaceae) gives evidence to the ecological compatibility of the previous agronomic management, in the sense that butterflies require certain weed species for oviposition and subsequent larva rearing and, therefore, provide further evidence of plant biodiversity in the environment.

**Keywords:** biological conservation; functional biodiversity; weed management; wildflowers; sustainability

# **1. Introduction**

Weeds are predominantly self-pollinated [\[1\]](#page-16-0); insect-pollinated weeds are also frequently found in agricultural ecosystems [\[2\]](#page-16-1). Self-pollination is of crucial importance for rapid seed formation; in accordance with the time-limitation hypothesis [\[3\]](#page-17-0), weeds require a certain degree of allogamy to maintain a genetic base capable of adapting to the dynamics of agronomic disturbance. Cross-pollination of angiosperms evolved in ancient natural ecosystems, from entomophily to anemophily [\[4\]](#page-17-1), probably to reduce dependence on biotic factors whose presence is affected by erratic climatic conditions [\[5\]](#page-17-2). This widely accepted hypothesis is supported by the evidence of rudimental and inefficient that nectaries are often present even in typically wind-pollinated species [\[6\]](#page-17-3). However, despite this evolutionary trend, many agroecosystem plants base their survival dynamics on insect pollination. This mutualistic component of a part of the agroecosystem weed communities assumes



**Citation:** Benvenuti, S. Weed Role for Pollinator in the Agroecosystem: Plant–Insect Interactions and Agronomic Strategies for Biodiversity Conservation. *Plants* **2024**, *13*, 2249. [https://doi.org/10.3390/](https://doi.org/10.3390/plants13162249) [plants13162249](https://doi.org/10.3390/plants13162249)

Academic Editor: Francisco Rubén Badenes-Pérez

Received: 10 July 2024 Revised: 7 August 2024 Accepted: 7 August 2024 Published: 13 August 2024



**Copyright:** © 2024 by the author. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license [\(https://](https://creativecommons.org/licenses/by/4.0/) [creativecommons.org/licenses/by/](https://creativecommons.org/licenses/by/4.0/)  $4.0/$ ).

a crucial ecological importance in terms of pollinator biodiversity even in this highly anthropized environment. The insects involved are often defined as "flower visitors" rather than pollinators, as their ecological role has not yet been fully clarified. It is frequently unclear whether the insect activity on the flowers allowing a contact pollen-gynoecium is accidental or mutualistic. However, the high frequency of flower visits constitutes a valid parameter for the almost certainty of this plant–insect mutualism [\[7\]](#page-17-4). Thus, for many insects and pollinated weeds, basically wildflowers, an evolution strategy towards flowers attractiveness can be discerned [\[8\]](#page-17-5). However, there are numerous cases of predominantly self- or wind-pollinated weeds that are visited by insects virtually capable of bringing about mutualistic or accidental gamy [\[9\]](#page-17-6). This ambophily is regarded as a transitional state intermediate between biotic and/or abiotic pollen movement [\[10\]](#page-17-7). This dual strategy allows the risk that erratic biotic and/or abiotic conditions may reduce the gene flow essential to evolve biotypes suitable for surviving in the agroecosystem. Such a strategy allows a shift from predominant self-pollination to predominant cross-pollination, as observed in numerous species [\[11\]](#page-17-8). A good example is offered by *Solanum ptycanthum* (Solanaceae), whose pollination strategy depends on the extent of disturbance in its growth environment, with predominantly insect-pollinated biotypes in natural ecosystems and predominantly self-pollinated biotypes in agricultural ecosystems [\[12\]](#page-17-9). It is not clear whether in this and other similar cases the greater attractiveness of the wild biotype is due to greater or lesser development and functioning of the nectaries. But it is worth noting that the presence of nectaries is not strictly necessary to induce insect flower visits, since pollen also constitutes a food source for a vast range of insects. Scanty or absent nectar production, thus, does not rule out the possibility that a species may be insect-pollinated, as it is pollen grain size that makes a species suited to insect pollination [\[13\]](#page-17-10). But it has been found that self-compatibility and self-pollination are associated with reduced pollen limitation, presumably because the capacity for self-fertilization decreases reliance on cross-pollination by pollinators [\[14\]](#page-17-11).

Recent years have seen increasing interest in gene flow mechanisms (biotic and/or abiotic) between the various weed species, partly for agronomic reasons, such as herbicide resistance [\[15\]](#page-17-12), and due to environmental concerns, as in the case of pollen transfer between genetically modified crops and potentially hybridizable weeds [\[16\]](#page-17-13). In addition, the plant and flower visitor interaction arising from insect pollination has aroused concern on account of increased awareness of the concept of safeguarding biodiversity in the agroecosystem [\[17\]](#page-17-14) and in other anthropized ecosystems [\[18\]](#page-17-15). Attention is focusing on a possible cause–effect relationship between the rarefaction and/or disappearance of some species and their dependence on entomofauna, whose ecological role is often overlooked.

The purpose of this study was to survey the state of the art of flora–fauna interaction in weed communities in terms of insect pollination, investigate the ecological importance of these mechanisms in survival dynamics, and determine agronomic strategies that can be adopted to safeguard biodiversity in the agricultural environment.

### **2. Pollinator Biodiversity and Reward**

Most flower visitors are social and solitary bees, bumblebees, Diptera, and Lepidoptera, as shown in Table [1.](#page-3-0) Each insect species feeds on pollen and/or nectar of given plant species as a function of its respective mouthparts [\[19\]](#page-17-16), which, in many cases, have evolved in a manner that enables the insect to reach and feed on solid (pollen) or liquid (nectar) food. These food resources are produced in specific structures that are highly diversified among the various plant species [\[20\]](#page-17-17).



**Table 1.** Flower visitor observations on several weed species of the agroecosystem.



#### <span id="page-3-0"></span>**Table 1.** *Cont.*

Among flower visitors, honeybees (*Apis mellifera*, Figure [1\)](#page-4-0) and solitary bees (Figure [2\)](#page-4-1) are predominant. Within the Mediterranean environment, solitary bees are represented above all by Andrenidae, Anthophoridae, Apidae, Melittidae, Colletidae, Halictidae, and Megachilidae [\[35\]](#page-18-0). The reward consists of nectar and/or pollen, with the latter being transported in different body places and after being packed in special pollen baskets situated on the insects' legs. But it is rare for bees to collect both pollen and nectar simultaneously, as energy economy prompts bees to visit species with a predominance of one or the other reward [\[36\]](#page-18-1). Natural ecosystems have an abundance of wildflowers, which have typically evolved nectar production as a reward. On the contrary, the conventional agroecosystem has a predominance of species poor of nectar, so that insects are rewarded with pollen. For example, *Papaver rhoeas*, in spite of their flower appearance, has no nectaries, and pollen is the only food source for pollinators, the latter being indispensable for seed set as this species is completely self-incompatible [\[37\]](#page-18-2).

However, although species with brightly colored flowers are the most frequently visited, even many common weeds with less gaudy flowers also constitute a useful food source, especially during periods when the natural environment offers fewer species in flower [\[38\]](#page-18-3). Large numbers of species flower in spring [\[39\]](#page-18-4), and they do compete with one another in producing nectar [\[40\]](#page-18-5), while in the subsequent months, late-flowering species with less noticeable flowers may be of interest to the pollinator, even though the reward is less advantageous. Above all, in late summer, flower visitors may be observed on species that previously exerted poor visual attraction, as occurs in many Asteraceae, such as *Senecio vulgaris*, *Sonchus* spp., *Aster squamatus*, *Conyza canadensis*, and even *Xanthium strumarium*. The latter species, which is typically wind- and/or self-pollinated, constitutes a curious case in that the ecological role of flower visitors on pollination can be considered negligible, as the position of its male flowers, separate from the female flowers, suggests that pollen movement towards the gynaeceum may be purely accidental. The presence of flower visitors (in particular bees and bumblebees) has also been noted almost exclusively in late summer on other species with small and/or poorly attractive flowers, such as *Polygonum laphatipholium*, *P. aviculare*, *Cuscuta campestris*, *Portulaca oleracea*, *Stellaria media*, and *Abutilon theophrasti*. Similar phenomena have been observed on species with gaudy but usually self-pollinated flowers, such as *Convolvolus arvensis* and *Calystegia saepium* [\[41\]](#page-18-6) and *Datura stramonium* [\[42\]](#page-18-7).

<span id="page-4-0"></span>

len is the only food source for pollinators, the latter being indispensable for seed set as

**Figure 1.** Honeybees observed on the flowers of common weeds of the agroecosystem: 1 = *Euphorbia* helioscopia, 2 = Polygonum laphatifolium, 3 = Linaria vulgaris, 4 = Nigella damascena, 5 = Lamium purpureum,  $6 =$  Picris echioides,  $7 =$  Galinsoga parviflora,  $8 =$  Malva sylvestris,  $9 =$  Cichorium inthybus,  $10 =$  Papaver *rhoeas*, 11 = *Cirsium arvense*, 12 = *Crepis vesicaria*.

<span id="page-4-1"></span>

**Figure 2.** Bumblebees (2, 3, 8, 11), solitary bees (1, 6, 7, 9, 12), and wasps (4, 5, 10) observed on common weeds of the agroecosystem:  $1 = Borago$  officinalis,  $2 = Datura$  stramonium,  $3 = Verbascum$ *columbaria*, 9 = *Malva sylvestris*, 10 = *Daucus carota*, 11 = *Lavatera punctata*, 12 = *Chrysanthemum coronarium*. *sinuatum*, 4 = *Orobanche crenata*, 5 = *Cuscuta campestris*, 6 = *Echium vulgare*, 7 = *Anagallis arvensis*, 8 = Scabiosa columbaria, 9 = Malva sylvestris, 10 = Daucus carota, 11 = Lavatera punctata, 12 = Chrysanthe-<br>. *mum coronarium*.

A less important, but still underestimated, role is played by Diptera (Figure [3\)](#page-5-0) (overall Syrphidae, Tachinidae, Sarcophagidae, and Bombyliidae) [\[43\]](#page-18-8). In Bombyliidae, on the other A less important of the mouthparts appear to have evolved to allow utilization of nectar by means of hand, the mouthparts appear to have evolved to allow utilization of nectar by means of a long proboscis that can penetrate inside small flowers [\[44\]](#page-18-9). Visits by Bombyliidae have been noted, above all, on flowers that would be difficult for other insects to reach due to their small and elongated floral calyx, as in the case of Gentianaceae (Centaurium erythraea and *Blackstonia perfoliata*), Campanulaceae (L*egousia speculum veneris* and J*asione montana*), Lamiaceae (Lamium purpureum and L. amplexicaule), Primulaceae (Anagallis arvensis and A. *foemina*), and other species. *foemina*), and other species.  $s$  appear to have evolved to anow utilization of hectar by filearly on  $s$ their small and elongated floral calyx, as in the case of Gentianaceae (*Centaurium erythraea* 

<span id="page-5-0"></span>

**Figure 3.** Diptera Empididae (1, 2), Tachinidae (3, 12), Syrphidae (4, 5, 6, 7, 8, 10), and Bombyiliidae **Figure 3.** Diptera Empididae (1, 2), Tachinidae (3, 12), Syrphidae (4, 5, 6, 7, 8, 10), and Bombyiliidae  $\mu$   $\mu$   $\alpha$   $\beta$  of the agroecosystem:  $1 -$  *Geranium molle*,  $2 -$ *Linum perenne*  $(9, 11)$  observed on common weeds of the agroecosystem:  $1 =$  Geranium molle,  $2 =$  Linum perenne, 3 = Inula viscosa, 4 = Silene alba, 5 = Papaver hybridum, 6 = Anthemis arvensis, 7 = Crysanthemum segetum, *chamomilla*, 12 = *Tordylium apulum*. 8 = *Scandix pecten-veneris*, 9 = *Cruysanthemum coronarium*, 10 = *Cephalaria transsylvanica*, 11 = *Matricaria*  $S$ utum,  $\frac{1}{2}$ *chamomilla*, 12 = *Tordylium apulum*.

Some insects, above all, Lepidoptera, have a long proboscis that enables them to visit flowers with an elongated calyx even when the nectaries are hidden at the base of the have a particular shape, such as Dipsaceae (*Knautia arvensis* and *Dipsacus fullonum*) and calyx (Figure [4\)](#page-6-0). Such insects include Lycaenidae, Pieridae and Sphingidae, Papilionidae, Nymphalidae, and Satyrids (Figure [5\)](#page-6-1). Visits are frequent on flower species whose flowers have a particular shape, such as Dipsaceae (*Knautia arvensis* and *Dipsacus fullonum*) and Caryophyllaceae (*Agrostemma githago*, *Lychnis flos-cuculi*, *Silene* spp., etc.). However, pollen transport by butterflies is decidedly less efficient in comparison to bees [\[45\]](#page-18-10). Species belonging to the order of Coleoptera (Figure [6\)](#page-7-0) are even less efficient, as such species, similarly to those of the order of Thysanoptera, often lack pollen transport specialization and appear to act more as pollen predators rather than potential pollinators [\[46\]](#page-18-11). Similarly poor efficiency is seen in ants, even though they are flower visitors of many species [\[47\]](#page-18-12), and this appears to be due to the low pollen germination after ant contact [\[48\]](#page-18-13). Their lack of hair and their very limited plant–plant movement on account of their inability to fly, at least in most species, suggests that these Hymenoptera are likely to be only occasional pollinators. Indeed, it was observed that ants may negatively affect plant fitness by reduced intensity of pollinator visits and that ants are repelled from flowers of many plant species (overall in tropical environments), although this repellence is clearly not ubiquitous [\[49\]](#page-18-14).

<span id="page-6-0"></span>

holes into the corolla without penetrating inside it [50], and are likely to be of equally

Figure 4. Butterfly (*Gonepteryx rhamni*) observed on *Dianthus cartusianorum* flower during nectar lyxes. suction through their long-proboscid, evolved to be able to reach the nectaries of elongated floral *Plants* **2024**, *13*, x FOR PEER REVIEW 8 of 24 calyxes.

<span id="page-6-1"></span>

**Figure 5.** Lepidoptera observed on common wildflowers of the agroecosystem: 1 = C*rysanthemum* segetum, 2 = Dianthus carthusianorum, 3 = Crepis vesicaria, 4 = Anthemis arvensis, 5 = Cephalaria trans-.<br>sylvanica, 6 = Senecio erraticus, 7 = Cirsium arvense, 8 = Dipsacus fullonum, 9 = Ranunculus arvensis, 10 = *Orlaya grandiflora*, 11 = *Sinapis arvensis*, 12 = *Lavatera punctata*.

Similar scarce pollination-efficiency is evidenced by some nectar-robbing species such as *Bombus occidentalis,* which visits *Linaria vulgaris* where it collects nectar by poking holes into the corolla without penetrating inside it [\[50\]](#page-18-15), and are likely to be of equally negligible ecological impact, since they are cheaters rather than mutualists [\[51\]](#page-18-16). *Orlaya grandiflora*, 11 = *Sinapis arvensis*, 12 = *Lavatera punctata*. *vanica*, 6 = *Senecio erraticus*, 7 = *Cirsium arvense*, 8 = *Dipsacus fullonum*, 9 = *Ranunculus arvensis*, 10 =

<span id="page-7-0"></span>

Figure 6. Coleoptera observed on common weeds of the agroecosystem: 1 = *Ornithogalum umbella*tum, 2 = Ammi majus, 3 = Mentha suaveolens, 4 = Crysanthemum segetum, 5 = Campanula rapunculus, 6 = Lavatera punctata, 7 = Plantago lanceolata, 8 = Dianthus cartusianorum, 9 = Daucus carota, 10 = Malva **3. Generalization or Specialization?**  *sylvestris*, 11 = *Anagallis arvensi*, 12 = *Ornithogalum umbellatum*.

#### It is widely believed that common weeds owe their time and space persistence to their lack of specialization [52], except for some species that are increasingly rare in con-**3. Generalization or Specialization?**

ved that common weeds owe their time and space persistence to the It is widely believed that common weeds owe their time and space persistence to their lack of specialization [\[52\]](#page-18-17), except for some species that are increasingly rare in conventional agroecosystems [\[53\]](#page-18-18). The evolutionary trend from generalization to specialization noted in natural ecosystems [\[54\]](#page-18-19) does not appear to be suited to the requirements of weeds specialization of some plant species towards certain political political political political explanators could explain why they they they they are  $\frac{1}{2}$ in an agricultural environment, where "plasticity" (despecialization) seems to be more important [\[55\]](#page-18-20). Indeed, it implies a lower risk of pollinator lack because of the high degree of disturbance dynamics, as typically occurs in the agroecosystem. Therefore, the Indeed, "conventional" agroecosystems are characterized by the dominance of selfspecialization of some plant species towards certain pollinators could explain why they are increasingly uncommon [\[56\]](#page-18-21). Pesticide application, which is typical of conventional agricultural systems, can interfere with the efficacious but fragile mechanisms that involve  $s_{\rm relismes}$ as floral symmetry plays and important role in plant–pollinator systems  $\mathcal{S}$ . Zygomorphic systems  $\mathcal{S}$ rigid flora–fauna mutualisms.

Indeed, "conventional" agroecosystems are characterized by the dominance of selfand wind-pollination, while, on the contrary, insect pollination is more frequent in natural ecosystems as a function of the greater abundance and biodiversity of pollinators typically Action or the greater as the thanks of the specific flowers, or point theory available in undisturbed environments. However, even predominantly insect-pollinated species have different degrees of specialization depending on the possible pollinators [\[57\]](#page-18-22), as floral symmetry plays an important role in plant-pollinator systems [\[58\]](#page-18-23). Zygomor- $\begin{bmatrix} 1 & 1 & 1 & 1 & 1 & 1 \\ 0 & 1 & 1 & 0 & 1 & 1 \\ 0 & 0 & 0 & 0 & 0 & 1 \end{bmatrix}$ phic flowers, such as *Consolida regalis, Echium vulgaris, Lamium amplexicaule,* and *Stachys arvensis,* are visited mainly by specialized long-tongued bees (Melittidae, Megachilidae, An-throphoridae, and Apidae) as a consequence of the particular position of the nectaries [\[59\]](#page-18-24). may represent a characteristic of the pollinators themselves, as observed in Italy for *Heri-*Actinomorphic flowers, on the other hand, are visited by a higher number of visitor species.<br> For example, almost all of the Asteraceae species (e.g., *Centaurea cyanus, Chrysanthemum* 

*myconis*, and *Cirsium arvense*) showed a higher degree of unspecialized visitors, such as short-tongued bees (Colletidae, Andrenidae, and Halictidae) and flies (personal observation). Another example is that of *Raphanus raphanistrum* (Brassicaceae), which exhibits traits typical of generalized pollination, including radially symmetric flowers, exposed reproductive organs, and an upright flower [\[60\]](#page-18-25).

A further type of specialization, with visits limited to a restricted botanical group, may represent a characteristic of the pollinators themselves, as observed in Italy for *Heriades truncorum* (Megachilidae), which is almost always seen on Asteraceae weeds [\[61\]](#page-18-26). But it cannot necessarily be assumed that the insects most frequently observed on flowers are the most efficient pollinators, since pollen transport is strongly dependent on the shape and hair of the insect but also on the speed of visits, with rapid speed proving to be less efficient. For example, the typical rapid visits by long-tongued bees may result in reduced pollen transport [\[62\]](#page-18-27). Furthermore, efficiency of pollination is also influenced by the electrostatic forces of pollen, which can assure adhesion to the pollinator even if the insect may lack hair [\[63\]](#page-18-28).

Overall, however, specialization is indisputably linked to flowering phenology [\[64\]](#page-18-29), which may or may not be compatible with the biological cycle of the pollinator. Thus, with cool season flowering species, potential pollinators are represented by insects that are capable of maintaining a certain degree of activity even at low temperatures, as in the case of early Amaryllidaceae pollinated by cold-tolerant Andrenidae [\[65\]](#page-18-30). The role of temperature as a limiting factor is confirmed by observations on various species of Campanulaceae, which show decreasing frequency of visits with increasing altitude [\[66\]](#page-18-31). In addition, the literature indicates that pollinators tend to favor peak or earlier flowering, whereas predispersal seed predators tend to favor off-peak or later flowering [\[67\]](#page-18-32).

An interesting example of plant–pollinator mutualistic specialization is found in *Silene noctiflora*, a gynomonoecious annual whose individuals produce both hermaphroditic and pistillate flowers. It flowers only during the night and is pollinated exclusively by nocturnal moths [\[68\]](#page-19-0). A similar system, albeit less exclusive, is seen in *Silene alba* [\[69\]](#page-19-1), which tends to open its flowers at the end of the day, thereby allowing pollination both by diurnal (bees, flies, and wasps) and nocturnal visitors (the latter being mostly Sphyngid and noctuid moths). This mixed system is typical of numerous other species belonging to the same family of Caryophyllaceae, as in the genera *Agrostemma*, *Saponaria*, *Dianthus*, and *Vaccaria* [\[21\]](#page-17-18).

Flora–fauna specialization does not depend only on the shape and manner of opening of the flowers but also on nectar composition, in terms of sugars and amino acids, as well as the nectar secretion rate, which is measurable in the field with various techniques [\[70\]](#page-19-2). For example, butterflies are attracted by the flowers richest in amino acids, as their diet is based exclusively on nectar and must therefore allow sufficient protein synthesis. Solitary and social bees are more attracted by an elevated sugar content, as they also feed on pollen and therefore do not need an additional protein supply (Gardener and Gilman, 2002). It has also been hypothesized that specialist nectarivores can assimilate sucrose, whereas some opportunistic nectar feeders digest only the simple exoses [\[71\]](#page-19-3). A crucial role is also played by amino acid typology. The predominance of phenylalanine and/or gamma-aminobutyric acid tends to attract long-tongued bees and flies (overall Syrphidae), whereas asparagine and tryptophane are rather repellent to these insect species [\[72\]](#page-19-4). Investigations aiming to obtain experimental evidence of an ecological function of nectar composition have been conducted on a vast range of species [\[73\]](#page-19-5), with results suggesting that some amino acids elicit different responses in insect receptors. However, amino acid concentration in nectar is not exclusively a function of the genotype. It can be influenced by agronomic management, as in the case of nitrogen fertilization, which has been shown to increase nectar amino acid concentration in *Agrostemma githago* [\[74\]](#page-19-6). Natural factors such as arbuscular mycorrhizal fungi can, likewise, increase flower visitor numbers (overall visits by Diptera and Hymenoptera) in *Centaurea cyanus* [\[75\]](#page-19-7). Finally, the ecological significance of the toxic nectar secreted, for example, by *Heliotropium europaeum*, *Cuscuta* spp., *Solanum nigrum*, and *Euphorbia* spp. is still poorly understood. It has been hypothesized that bees are more resistant to alkaloids than adult Lepidoptera, and that alkaloids in nectar encourage pollination by specialist rather "flower-inconstant" pollinators [\[76\]](#page-19-8).

Pollinator attraction is linked to the mechanisms involved in recognition of appropriate flowers. Recognition is crucial in that it avoids confusion in pollen transfer, which must take place within the same species as far as possible [\[77\]](#page-19-9). Flower shape and color both play an important role in facilitating recognition. Color is perceived differently by the insect as compared to the human eye, and light reflectance at wavelengths invisible to humans (roughly 300–400 nm) is well perceived [\[78\]](#page-19-10). Many Brassicaceae reflect ultraviolet color in order to attract the attention of pollinators [\[79\]](#page-19-11). Some flower colors appear to be correlated with certain categories of pollinators, although this cannot always be generalized due to poor convergence of data from different environments [\[80\]](#page-19-12). Specialist bumblebees have been noted to show a preference for purple, and this example would appear to confirm the so-called "pollination syndrome" theory [\[81\]](#page-19-13). In some cases, recognition is facilitated by color patterns: thus, the "search images" system possessed by insects [\[82\]](#page-19-14) can be aided by characteristic black spots at the base of the petal, functioning as a "nectar guide" [\[83\]](#page-19-15), as observed, for example, in *Papaver rhoeas*. But bright and gaudy colors are not always an indispensable condition for attracting flower visitors. Some Euforbiaceae have pale green flowers that do not stand out within the surrounding vegetation, yet they are frequently visited, as in the case of *Euphorbia esula* [\[84\]](#page-19-16). Moreover, a further and often decisive mechanism for identification and recognition of flowers consists of production and emission of fragrant molecules composed essentially of terpenoids and benzenoids [\[85\]](#page-19-17).

#### **4. Weed Pollination Strategies**

The main characteristic of arable weeds is their ability to persist despite the vast range of agronomic disturbances. One of the crucial strategies adopted by weeds is rapid seed set. Rapidity is maximized with self-pollination, as autogamous seed set is not dependent on the occurrence of any (biotic or abiotic) event [\[86\]](#page-19-18). Weeds are regarded as pioneer flora of early stages of secondary successions [\[87\]](#page-19-19), and their frequent annual cycle is often correlated with self-pollination [\[88\]](#page-19-20) according to the "time-limitation" hypothesis of crucial importance in the typically highly disturbed agroecosystem. For example, *Amaranthus retroflexus*, characterized by flowers devoid of attractiveness and by almost total selfpollination [\[89\]](#page-19-21), represents the ideotype of weeds as it already produces mature seeds just a few weeks after emergence [\[90\]](#page-19-22). Species whose corolla is a marked attractant (in terms of size and/or color) may also be mainly self-pollinated, as in *Convolvulus arvensis* [\[41\]](#page-18-6), *Stellaria media* [\[91\]](#page-19-23), *Portulaca oleracea* [\[92\]](#page-19-24), and *Anagallis arvensis* [\[93\]](#page-19-25). However, it is likely that this predominance of self-fertilization is found in biotypes present in agricultural environments, while outcrossing is more widespread in biotypes present in less disturbed environments. This means that the frequent disturbances of the agroecosystem probably favored coevolution towards predominantly self-pollinated biotypes. This is the case of *Datura stramonium*, which is pollinated in its original environments by hawkmoths (*Manduca quinquemaculata*), Halictids, and honeybees (*Apis mellifera*) [\[94\]](#page-19-26), whereas the biotypes present in cultivated fields are almost totally autogamous [\[95\]](#page-19-27). This coevolution with agronomic disturbances has led to a decrease in nectary function and a corresponding elongation of the androecium and gynoecium to allow contact between anthers and stigma (herkogamy), thereby favoring self-pollination [\[96\]](#page-19-28). Despite this, some exceptions that evolved towards self-incompatibility are observed, as in the case of *Papaver rhoeas* [\[97\]](#page-19-29) and *Ranunculus repens* [\[98\]](#page-19-30). But, in general, the absolute self-incompatibility in the agroecosystem plant communities is unusual. Indeed, a variable frequency of distribution between self- and cross-pollination is more frequently found, as in the cases of *Anthemis cotula* [\[99\]](#page-19-31), *Sinapis arvensis*, [\[100\]](#page-19-32), *Raphanus raphanistrum* [\[101\]](#page-20-0), and other insect-pollinated species typically widespread in the agroecosystem [\[102\]](#page-20-1). In the case of *Raphanus raphanistrum*, flower color (typically white or yellow) plays an ecological role in favoring or discouraging crosspollination despite partial self-compatibility. Thus, one important visitor is a lepidopteran

(*Pieris rapae*), which mainly visits yellow flowers, resulting in a predominance of crosspollination in these populations [\[103\]](#page-20-2), whereas self-pollination is predominant in white flower varieties. This dual typology of biotypes could represent an example of optimal trade-off between the advantages of one or the other gene flow mechanism, highlighting plastic and evolutionary changes in floral traits. Such a trade-off is not exclusive to this weed species but is widespread in many other species as well, and can probably be interpreted as a strategy for maintaining populations with diversified biological characteristics [\[104\]](#page-20-3).

Overall, pollen self-incompatibility is one of the various strategies adopted by plant species to avoid the pollination of different flowers growing on the same plant, especially in the case of individuals bearing numerous blooms [\[105\]](#page-20-4). Such pollination, which would be pointless in terms of gene flow, is known as geitonogamy [\[106\]](#page-20-5). Other geitonogamy avoidance strategies include spatial and/or temporal separation of pollen and stigma. In general, geitonogamy avoidance is advantageous in favoring adaptability to dynamic environmental conditions [\[107\]](#page-20-6). A trade-off between geitonogamy and xenogamy (cross-pollination) is observed in *Daucus carota*, an andromonoecious and protandrous species [\[108\]](#page-20-7). In this case, separation of the male and female phases is complete at the level of the flower and umbel, but the two phases overlap at the level of the full plant, creating conditions for geitonogamy even if insect visits between the umbels of adjacent plants lead to xenogamy. Another curious characteristic of this species is the possible ecological role of the dark central floret of the inflorescences, for which a possible "fly catcher" role has been suggested, although its function has not yet been fully clarified [\[108\]](#page-20-7). Additional examples of a balance between self-pollination and insect pollination are observed in *Cynoglossum officinale*, *Echium vulgare* [\[109\]](#page-20-8), and some species belonging to the botanical genus *Delphinum* [\[110\]](#page-20-9), in which the extremely variable number of flowers results in diversified probability of geitonogamy. Plants with a greater number of flowers tend to favor self-pollination as there is an increased probability that the pollinators will sequentially visit (geitonogamy) flowers of the same plant. A further diversification of breeding frequency is found in *Echium vulgare*, as the protandrous flowers produce more nectar and receive higher rates of visitation during their male than during their female phase [\[111\]](#page-20-10), although nectar production in this species is also closely dependent on environmental conditions [\[112\]](#page-20-11).

Among geitonogamy avoidance strategies, a particularly drastic mechanism is displayed by dioecious species, in which the separation of individuals into different sexes makes self-fertilization impossible. An example is seen in *Silene dioica*, which is visited by bumblebees, hoverflies (Syrphidae), butterflies (mainly Pieridae), and honeybees [\[113\]](#page-20-12). But since the invasiveness of this species in the agroecosystem is negligible, as compared to other monoecious species belonging to the same genus, it can be concluded that such a strategy is unsuccessful [\[114\]](#page-20-13).

## **5. Insect-Pollinated Weeds as Indicators of Biodiversity and Agroecosystem Health**

A clear-cut distinction between entomopollinated and nonentomopollinated species cannot easily be drawn, as the true ecological role of insect visits has not yet been clarified for each individual species. This uncertainty is aggravated by the above-described differences among biotypes present in the wild (more dependent) versus those in the agroecosystem (less dependent). However, it is generally agreed that while the reward may consist of pollen (Figure [7\)](#page-11-0), both quantitative and qualitative (sugar concentration) nectar production is linked to entomofauna through mutualistic specialization. Since nectar production requires considerable energy requirement, in short-lived weeds, characterized by annual cycle, nectar is less abundant than in perennial species [\[115\]](#page-20-14), and if it is not collected, it is reabsorbed by the plant for its own metabolism [\[116\]](#page-20-15). It can, therefore, be stated that species with a well-developed nectary rely mainly on insect visits for their survival, with the visits being crucial for seed set. Thus, weed species characterized by this feature (essentially, wildflowers) face a greater risk within the agroecosystem, because the high level of disturbance of the agricultural environment tends to restrict the availability of their pollinators. Pesticide toxicity and its residues play a crucial role in this regard by severely

<span id="page-11-0"></span>

affecting the chance for survival of entomofauna and, consequently, of insect-pollinated flora.

**Figure 7.** Honeybee observed during a visit to an inflorescence of *Daucus carota*: note the balls of Figure 7. Honeybee observed during a visit to an inflorescence of *Daucus carota*: note the balls of pollen accumulated on the hind legs, typically yellow in this species.

The frequency of wildflower species in the agroecosystem plant communities can,

 $t_{\text{tril}}$  definition appears in the economic plant communities can The frequency of wildflower species in the agroecosystem plant communities can, therefore, represent a valid indicator of their ecological sustainability. It is now recognized that the floristic diversity of the agroecosystem provides an assessment of the agroecological impact [\[117\]](#page-20-16). This is particularly true about entomopollinated species, as their presence  $\sum_{i=1}^{n}$  in  $\sum_{i=1}^{n}$  and a boat of  $\sum_{i=1}^{n}$ . presupposes a level of biodiversity extended to the animal kingdom, and in this context, it should not be forgotten that evolution towards self-pollination occurred precisely in situations of a lack of pollinators [\[118\]](#page-20-17).  $t_{\rm{max}}$  decreased presence or disappearance or di

The mutualistically more specialized species, such as wildflowers, constitute the most reliable ecological assessment parameters since these are the species whose presence is most severely threatened by disruption of the balance of the agroecosystem. Although  $\mathbf{r}$  decades, the decades, the decades, the decades many previously widethe decreased presence or disappearance of some species may be due to other agronomic causes (herbicides, heightened aggression by more competitive weeds, crop seed selection, etc. [\[119\]](#page-20-18)), it is highly probable that the declining numbers of pollinators have been a  $f(x) = \frac{1}{2}$ contributing factor in the increasing rarity of some species. As has been widely noted, in the past few decades, the decline of biodiversity has involved many previously widespread plant and animal organisms [\[120\]](#page-20-19). Throughout Europe, only plant species whose persistence dynamics do not rely on biotic action for pollination are only occasionally cited as rare weeds [\[121\]](#page-20-20). On the contrary, lists of declining species include numerous wildflowers, which are threatened by their dependence on flower visitors for seed set [\[122\]](#page-20-21). It has also been shown that the frequency of flower visitors on wildflowers is closely related to the quantity of viable seeds produced [\[123\]](#page-20-22). Weed species that are only scantily present have difficulty in attracting insects, as it has been noted both in natural ecosystems [\[124\]](#page-20-23) and in the agroecosystem [\[125\]](#page-20-24) that insects prefer to visit more numerous species. In arable fields, a large number of insect-pollinated wildflowers have now become rare or are in decline, such as *Agrostemma githago*, *Centaurea cyanus*, *Papaver argemone*, *Ranunculus ar-* *vensis*, [\[126\]](#page-20-25), *Chrysanthemum sagetum*, *Matricaria recutita*, *Legousia hybrida*, *Silene alba*, *Viola arvensis* [\[127\]](#page-20-26), *Consolida regalis*, *Silene noctiflora*, *Lamium amplexicaule*, [\[128\]](#page-20-27), *Myosotis arvensis*, *Viola tricolor* [\[129\]](#page-20-28), *Legousia speculum veneris*, *Anchusa arvensis*, [\[130\]](#page-20-29), *Nigella arvensis*, and *Ornithogalum umbellatum* [\[131\]](#page-20-30).

As stated above, the risk of decline is greatest when mutualistic interaction is specialized. Thus, many of the abovementioned species belong to the Cariophyllaceae, a botanical family often characterized by rigid mutualistic interactions set in motion by butterflies [\[132\]](#page-21-0). This type of mutualism is highly fragile, because Lepidoptera require a twofold plant-related availability: food source (visitable flora endowed with nectaries), and suitable conditions for reproduction (appropriate flora for oviposition and feeding of larval forms). Each butterfly species is dependent on restricted plant groups, often belonging to a single botanical family, a single genus, or even a single species (Table [2\)](#page-13-0). Generally, reference is made to a hierarchy of preferences since some species may be preferred to others within a given botanical grouping, as in the case of *Papilio machaon*, which oviposits exclusively on Apiaceae (Figure [8\)](#page-14-0). If certain host plants have poor invasiveness within the various ecosystems, this inevitably leads to very scanty presence not only of the respective plant species but also of the correlated butterfly species [\[133\]](#page-21-1).



**Table 2.** Some examples of weed host selected by several butterfly species.



## <span id="page-13-0"></span>**Table 2.** *Cont.*

While specificity between pollinator and host weed is may be variable, it is quite typical of the different families of Lepidoptera, being found preferentially or obligatorily linked to restricted botanical groupings. For example, with Satytiridae (e.g., *Brintesia circe*), there is only scanty specialization as the pollinator/host-plant relation is observed in many ubiquitarian species of Graminaceae, but requirements are more stringent for *Macroglossa stellatarum* (Sphyngidae), which needs one of the various species of the genus *Galium* (Rubiaceae). The risk of butterfly/plant-host coextinction cannot be disregarded and has already been reported in some parts of the world [\[143\]](#page-21-11). It is also interesting to note that numerous Lepidoptera, known as myrmecophilous butterflies, have mutualistic relations with ants, as ants defend butterfly eggs and the subsequent caterpillars against predator attack [\[144\]](#page-21-12). It has been observed that oviposition of myrmecophilous butterflies takes place preferentially on plants most frequently visited by ants [\[145\]](#page-21-13), showing that the presence of butterflies testifies to an even more extensive level of biodiversity [\[146\]](#page-21-14). In contrast, bumblebee reproduction, while similarly limited by scanty availability of undisturbed environments, is less specialized as it generally takes place in soil [\[147\]](#page-21-15). Lack of specificity is also noted in most solitary bees, with some species nesting in soil while others also nest in plant residue cavities [\[148\]](#page-21-16).

<span id="page-14-0"></span>

*Veronica* spp. *Stenoptilia pterodactyla* [139]

**Figure 8.** Caterpillar of the *Papilio machaon* butterfly specialized to lay eggs on plants of the Apiaceae family (in this case, the toxic *Conium maculatum*).

A special form of ecological interaction is found in pollinators that require the presence of other pollinators for their reproduction. For example, in many species of Diptera Bombylidae, which have a very thin and elongated mouthpiece allowing them to collect nectar from small flowers such as *Anagallis arvensis*, *Legousia speculum veneris*, *Centaurium erytrea*, and *Lamium purpureum* (Benvenuti, personal observation), parasitic oviposition takes place in the nest of several species of solitary bees [\[149\]](#page-21-17), where the larvae feed on resources intended to be food for the bees. Indeed, Diptera Bombylidae exhibit mutualistic behavior towards insect-pollinated plant species but have parasitic behavior towards other pollinators.

Often, rare weeds are pollinated above all by Lepidoptera, as in the case of many Caryophyllaceae [\[150\]](#page-21-18). Consequently, they represent the most valid indicator of the biodiversity of the agroecosystem since their presence testifies to a complex level of flora–fauna interactions. But exceptions to this rule are found for some butterflies, such as *Pieris rapae*, which often choose the same species both for pollinating and for oviposition as well as for subsequent rearing of larvae. This exemplifies mutualism and parasitism simultaneously [\[151\]](#page-21-19), which may result in a sort of conflict of interest [\[152\]](#page-21-20) for the pollinator itself. However, apart from these exceptions, butterfly presence and diversity depend on the landscape context in that their survival dynamics are linked to availability of the required host species in the environment [\[153\]](#page-21-21). Analogously, the widespread presence of Syrphid (Diptera) has also been considered a good indicator of plant biodiversity [\[154\]](#page-21-22), on account of the abundance of different environments suitable for their reproduction. Some species of spider can also be considered as a further parameter for assessment of ecosystem integrity, as they hide on flowers to prey on pollinators. Such a phenomenon has been observed on *Aslepias syriaca* [\[155\]](#page-21-23), *Leucanthemum vulgare* [\[156\]](#page-21-24), and many other species. Thus, spiders represent the tip of the ecological pyramid of this flower–pollinator–predator food chain, and this ecosystem appears be particularly vulnerable [\[157\]](#page-21-25).

Confirmation of the reliability of insect-pollinated flora as an indicator of ecological sustainability of the agronomic cultural practices adopted comes from the observation that organic agricultural systems lead to an increase in insect-pollinated species [\[158\]](#page-21-26). Future

ecological assessments could be based on monitoring the spider species known to be the preferential predators of pollinating insects.

## **6. Long-Term Plant–Animal Biodiversity Sustainability**

Weed management is crucial for biodiversity sustainability [\[159\]](#page-21-27), as also highlighted by organic agricultural systems [\[160\]](#page-21-28). Perhaps the starting point is diversification of land management, since species richness, genetic variability, and extinction probability are closely linked to landscape traits such as habitat diversity, structural heterogeneity, patch dynamics, and perturbations [\[161\]](#page-21-29). In other words, arable weed diversity increases with landscape complexity [\[162\]](#page-21-30). Landscape planning is crucial for biodiversity [\[163\]](#page-21-31). It is important to keep in mind that distances between nesting environments and food sources must not be excessive, since beyond a certain distance, the trip energy will no longer be advantageous due to excessive energy consumption for flight between the nest and flora to be visited. Other, more suitable environments will therefore be sought [\[164\]](#page-21-32). This implies that the geometry of the agroecosystem plays an important role in ensuring that insect-pollinated weeds achieve sufficient seed set.

Woods represent a fundamental reserve of environments suitable for pollinator survival, especially if they are established in a mosaic pattern [\[165\]](#page-22-0) within an agricultural setting. A similar positive effect is produced by cattle grazing [\[166\]](#page-22-1) because this kind of land use presupposes both forage resources and nesting resources [\[167\]](#page-22-2). With regard to the potential for nesting, at least as far as numerous species of solitary bees and bumblebees, it is important to plan uncropped areas near the crops in such a way as to allow undisturbed aboveground nesting [\[168\]](#page-22-3). In addition, the introduction of long-term crops such as Lucerne (*Medicago sativa*) can guarantee prolonged periods (3–4 years) free from soil disturbance. Honeybees are severely damaged by microencapsuled pesticides, whose microgranules adhere to the insect's hair and are thereby transferred into the hives, where their toxic effects can kill the larvae [\[169\]](#page-22-4). Therefore, the use of such pesticides should be strictly avoided.

The likelihood of pollinator survival can be increased by the presence of field margins, hedges [\[170\]](#page-22-5), and other buffer zones [\[171\]](#page-22-6) or set-aside fields [\[172\]](#page-22-7). Such areas not only offer a suitable environment for soil-nesting pollinators, but also for Lepidoptera that require certain weeds on which to oviposit [\[173\]](#page-22-8). Wildflowers linked to mutualistic relations with the pollinators represent the ideotype of field margins as they not only provide a suitable ecological niche for an elevated number of pollinators, but they also ensure positive benefits for the agricultural landscape. Thus, it has been shown that the introduction of complex mixed wildflower strips leads to an increase in butterflies, which are drawn by the presence of host plants for oviposition and nectar as a food source [\[174\]](#page-22-9). The use of native wildflowers achieves the best ecological response in safeguarding pollinator biodiversity, above all, with regard to specialized pollinators (i.e., short- or long-tongued bees) linked to the wildflowers involved [\[175\]](#page-22-10). The introduction of exotic pollinators [\[176\]](#page-22-11) tends to impair the plant–pollinator balance of the agroecosystem overall following increasingly evident climate changes [\[177\]](#page-22-12). This occurs because the balance of weed–pollinator competition is disrupted by altered mutualistic plant–animal interactions [\[178\]](#page-22-13). Even though it is not always easy to distinguish the cause from the effect, i.e., to determine whether the imbalance is triggered by a lack of wildflowers or pollinators [\[179\]](#page-22-14), limiting or preventing the introduction of non-native animal or plant organisms into such environments represents a strategy of paramount importance. It may, therefore, help to avert the tendency to "biological globalization" and the ensuing genetic erosion or loss of native insect-pollinated plants and/or the relative flower visitors.

# **7. Conclusions**

Every weed species is characterized by a particular survival strategy and ruderal species (early flowering, abundant and prolonged seed production) derived from an evolutionary direction capable of giving them ideal weed traits [\[180\]](#page-22-15) to persist in the agroecosystem. In this context, the rigid plant–pollinator mutualisms are undoubtedly a disadvantage since they presuppose the presence of a consistent pollinator quantity and biodiversity. Despite this, in many cases, there is a transition towards weed communities that display an increasing presence of species whose survival strategies depend on pollinators with varying degrees of specialization. The growing need to maintain and/or restore agroecosystem biodiversity has focused attention on the insect-pollinated weeds that are among the first to decline or even disappear in the agricultural landscape. Paradoxically, it is now widely believed that even weeds perform an "ecosystem service" dedicated to the survival of pollinators essential for the productivity of the various insect-pollinated crops [\[181\]](#page-22-16). Within pollinators, butterflies are particularly subjected to rarefaction since their survival does not depend exclusively on the presence of pollinated plants but also on the further availability of host plants essential for their oviposition. Thus, it is important to identify valid ecological indicators to monitor the health of the agroecosystem. The presence of crab vand ecological molcators to monitor the health of the agroecosystem. The presence of crab<br>spiders has been proposed as a valid indicator of the level of the agroecosystem biodiversity. Indeed, these arthropods feed on the pollinators that wait camouflaged on the flowers, mated, these arallopeds reed on the pollinations that want called analyzed on the howers,<br>thus highlighting the biodiversity of both pollinated plants and pollinators [\[182\]](#page-22-17) (Figure [9\)](#page-16-2). However, further studies are required to determine more precisely, for each species, the pollinator requirement for seed set, especially for specialized weeds. This improved knowledge would not only aid research based on biological indicators, but it would also help to optimize the biodiversity restoration programs of degraded agroecosystems. store are productivity of the various model pollitated crops  $[101]$ representation in the proposition. Thus, it is important to ractivity iodiversity of both pollinated plants and pollinators  $[182]$  (Figure 9) precisely, expeciency for specialized meetic. This improved nicon-

<span id="page-16-2"></span>

**Figure 9.** Observed crab spider with their evident success in catching a solitary bee on *Onobrychis*  **Figure 9.** Observed crab spider with their evident success in catching a solitary bee on *Onobrychis viciifolia* inflorescence.

**Funding:** This research received no external funding.

**Conflicts of Interest:** The author declares no conflicts of interest.

#### **References**

- <span id="page-16-0"></span>1. Sutherland, S. What makes a weed: Life story traits of native and exotic plants in the USA. *Oecologia* **2004**, *141*, 24–39. [\[CrossRef\]](https://doi.org/10.1007/s00442-004-1628-x)
- <span id="page-16-1"></span>2. Bretagnolle, V.; Gaba, S. Weeds for bees? A review. *Agron. Sustain. Dev.* **2015**, *35*, 891–909. [\[CrossRef\]](https://doi.org/10.1007/s13593-015-0302-5)
- <span id="page-17-0"></span>3. Aarssen, L.W. Why are most selfing annuals? A new hypothesis for the fitness benefit of selfing. *Oikos* **2000**, *89*, 606–612. [\[CrossRef\]](https://doi.org/10.1034/j.1600-0706.2000.890321.x)
- <span id="page-17-1"></span>4. Mitchell, R.J.; Irwin, R.E.; Flanagan, R.J.; Karron, J.D. Ecology and evolution of plant–pollinator interactions. *Ann. Bot.* **2009**, *103*, 1355–1363. [\[CrossRef\]](https://doi.org/10.1093/aob/mcp122)
- <span id="page-17-2"></span>5. Bawa, K.S. Pollination, seed dispersal and diversification of angiosperms. *Trends Ecol. Evol.* **1995**, *10*, 311–312. [\[CrossRef\]](https://doi.org/10.1016/S0169-5347(00)89116-8)
- <span id="page-17-3"></span>6. Culley, T.M.; Weller, S.G.; Sakai, A.K. The evolution of wind pollination in angiosperms. *Trends Ecol. Evol.* **2002**, *7*, 361–369. [\[CrossRef\]](https://doi.org/10.1016/S0169-5347(02)02540-5)
- <span id="page-17-4"></span>7. Ne'eman, G.; Jürgens, A.; Newstro-Lloyd, L.; Potts, S.G.; Dafni, A. A framework for comparing pollinator performance: Effectiveness and efficiency. *Biol. Rev.* **2010**, *85*, 435–451. [\[CrossRef\]](https://doi.org/10.1111/j.1469-185X.2009.00108.x)
- <span id="page-17-5"></span>8. Bronstain, J.L.; Alarcón, R.; Geber, M. The evolution of plant-insect mutualisms. *New Phytol.* **2006**, *172*, 412–425. [\[CrossRef\]](https://doi.org/10.1111/j.1469-8137.2006.01864.x)
- <span id="page-17-6"></span>9. Stephens, R.E.; Gallagher, R.V.; Dun, L.; Cornwell, W.; Sauquet, H. Insect pollination for most of angiosperm evolutionary history. *New Phytol.* **2003**, *240*, 880–891. [\[CrossRef\]](https://doi.org/10.1111/nph.18993)
- <span id="page-17-7"></span>10. Abrahamczyk, S.; Struck, J.H.; Weigend, M. The best of two worlds: Ecology and evolution of ambophilous plants. *Biol. Rev.* **2023**, *98*, 391–420. [\[CrossRef\]](https://doi.org/10.1111/brv.12911)
- <span id="page-17-8"></span>11. Holsinger, K.E. Mass-action models of plant mating systems: The evolutionary stability of mixed mating systems. *Am. Nat.* **1991**, *138*, 606–622. [\[CrossRef\]](https://doi.org/10.1086/285237)
- <span id="page-17-9"></span>12. Hermanutz, L. Outcrossing in the weed, *Solanum ptycanthum* (Solanaceae): A comparison of agrestal and ruderal populations. *Am. J. Bot.* **1991**, *78*, 638–646. [\[CrossRef\]](https://doi.org/10.1002/j.1537-2197.1991.tb12588.x)
- <span id="page-17-10"></span>13. Hao, K.; Tian, Z.X.; Wang, Z.C.; Huang, S.Q. Pollen grain size associated with pollinator feeding strategy. *Proc. R. Soc. Lond. B Biol. Sci.* **2020**, *287*, 20201191. [\[CrossRef\]](https://doi.org/10.1098/rspb.2020.1191) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32811305)
- <span id="page-17-11"></span>14. Larson, B.; Barrett, S.C.H. A comparative analysis of pollen limitation in flowering plants. *Biol. J. Linn. Soc.* **2000**, *69*, 503–520. [\[CrossRef\]](https://doi.org/10.1111/j.1095-8312.2000.tb01221.x)
- <span id="page-17-12"></span>15. Preston, C.; Powles, S.B. Evolution of herbicide resistance in weeds: Initial frequency of target site-based resistance to acetolactate synthase-inhibiting herbicides in *Lolium rigidum*. *Heredity* **2002**, *88*, 8–13. [\[CrossRef\]](https://doi.org/10.1038/sj.hdy.6800004) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11813100)
- <span id="page-17-13"></span>16. Conner, A.J.; Glare, T.R.; Nap, J.P. The release of genetically modified crops into the environment: Part II. Overview of ecological risk assessment. *Plant J.* **2003**, *33*, 19–46. [\[CrossRef\]](https://doi.org/10.1046/j.0960-7412.2002.001607.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12943539)
- <span id="page-17-14"></span>17. Gibson, R.H.; Nelson, I.L.; Hopkins, G.W.; Hamlett, B.J.; Memmott, J. Pollinators webs, plant communities and the conservation of rare plants: Arable weeds as a case study. *J. App. Ecol.* **2006**, *43*, 246–257. [\[CrossRef\]](https://doi.org/10.1111/j.1365-2664.2006.01130.x)
- <span id="page-17-15"></span>18. Benvenuti, S. Weed dynamics in the Mediterranean urban ecosystem: Ecology, biodiversity and management. *Weed Res.* **2004**, *5*, 341–354. [\[CrossRef\]](https://doi.org/10.1111/j.1365-3180.2004.00410.x)
- <span id="page-17-16"></span>19. Krenn, H.W.; Plant, J.D.; Szucsich, N.U. Mouthparts of flower visiting insect. *Arthropod Struct. Dev.* **2005**, *34*, 1–40. [\[CrossRef\]](https://doi.org/10.1016/j.asd.2004.10.002)
- <span id="page-17-17"></span>20. Pacini, E.; Nepi, M.; Vesprini, J.L. Nectar biodiversity: A short review. *Plant Syst. Evol.* **2003**, *238*, 7–21. [\[CrossRef\]](https://doi.org/10.1007/s00606-002-0277-y)
- <span id="page-17-18"></span>21. Jürgens, A. Comparative floral morphometrics in day-flowering, night-flowering and self-pollinated Caryophylloideae (Agrostemma, Dianthus, Saponaria, Silene, and Vaccaria). *Plant Syst.Evol* **2006**, *257*, 233–250. [\[CrossRef\]](https://doi.org/10.1007/s00606-005-0379-4)
- <span id="page-17-19"></span>22. Albre, J.; Quilichini, A.; Gibernau, M. Pollination ecology of *Arum italicum* (Araceae). *Bot. J. Linn. Soc.* **2003**, *141*, 205–214. [\[CrossRef\]](https://doi.org/10.1046/j.1095-8339.2003.00139.x)
- <span id="page-17-20"></span>23. Kephart, S.; Theiss, K. Pollinator-mediated isolation in sympatric milkweeds (*Asclepias*): Do floral morphology and insects behavior influence species boundaries? *New Phytol.* **2003**, *161*, 265–277. [\[CrossRef\]](https://doi.org/10.1046/j.1469-8137.2003.00956.x)
- <span id="page-17-21"></span>24. Benvenuti, S. Soil texture involvement in wildflower strip ecosystem services delivery in Mediterranean agro-environment. *Eur. J. Agron.* **2023**, *145*, 126793. [\[CrossRef\]](https://doi.org/10.1016/j.eja.2023.126793)
- <span id="page-17-22"></span>25. Brys, R.; Jacquemyn, H. 2011 Variation in the functioning of autonomous self-pollination, pollinator services and floral traits in three *Centaurium* species. *Ann. Bot.* **2011**, *107*, 917–925. [\[CrossRef\]](https://doi.org/10.1093/aob/mcr032) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21320880)
- <span id="page-17-23"></span>26. Benvenuti, S.; Mazzoncini, M.; Cioni, P.L.; Flamini, G. Wildflower-pollinator interactions: Which phytochemicals are involved? *Basic Appl. Ecol.* **2020**, *45*, 62–75. [\[CrossRef\]](https://doi.org/10.1016/j.baae.2020.03.008)
- <span id="page-17-24"></span>27. Haaland, C.; Gyllin, M. Butterflies and bumblebees in greenways and sown wildflower strips in southern Sweden. *J. Insect Conserv.* **2010**, *14*, 125–132. [\[CrossRef\]](https://doi.org/10.1007/s10841-009-9232-3)
- <span id="page-17-25"></span>28. Larson, D.L.; Royer, R.A.; Royer, M.R. Insect visitation and pollen deposition in an invaded prairie plant community. *Biol. Conserv.* **2006**, *130*, 148–159. [\[CrossRef\]](https://doi.org/10.1016/j.biocon.2005.12.009)
- <span id="page-17-26"></span>29. Newman, D.A.; Thomson, J.D. Interactions among nectar robbing, floral herbivory, and ant protection in *Linaria vulgaris*. *Oikos* **2005**, *110*, 497–506. [\[CrossRef\]](https://doi.org/10.1111/j.0030-1299.2005.13885.x)
- <span id="page-17-27"></span>30. Kawaano, S.; Odaki, M.; Yamaoka, R.; Odatanabe, M.; Takeuchi, M.; Kawano, N. Pollination biology of *Oenotera* (Onagraceae). The interplay between floral UV absorbancy patterns and floral volatiles as signals to nocturnal insects. *Plant Spec. Biol.* **1995**, *10*, 31–35. [\[CrossRef\]](https://doi.org/10.1111/j.1442-1984.1995.tb00118.x)
- <span id="page-17-28"></span>31. Young, H.J. Diurnal and nocturnal pollination of *Silene alba* (Caryophyllaceae). *Am. J. Bot.* **2002**, *89*, 433–440. [\[CrossRef\]](https://doi.org/10.3732/ajb.89.3.433) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21665639)
- <span id="page-17-29"></span>32. Kay, Q.O.N.; Lack, A.J.; Bamber, F.C.; Davies, C.R. Differences between sexes in floral morphology, nectar production and insect visit in a dioecious species, *Silene dioica*. *New Phytol.* **1984**, *98*, 515–519. [\[CrossRef\]](https://doi.org/10.1111/j.1469-8137.1984.tb04145.x)
- <span id="page-17-30"></span>33. Folke, S.H.; Delph, L.F. Environmental and physiological effects on pistillate flower production in *Silene noctiflora* L. (Caryophyllaceae). *Int. J. Plant Sci.* **1997**, *158*, 501–509. [\[CrossRef\]](https://doi.org/10.1086/297460)
- <span id="page-17-31"></span>34. Beattie, A.J. Pollination mechanism in *Viola*. *New Phytol.* **1971**, *70*, 343–346. [\[CrossRef\]](https://doi.org/10.1111/j.1469-8137.1971.tb02533.x)
- <span id="page-18-0"></span>35. Petanidou, T.; Lamborn, E. A land for flowers and bees: Studying pollination ecology in Mediterranean communities. *Plant Biosyst.* **2005**, *139*, 279–294. [\[CrossRef\]](https://doi.org/10.1080/11263500500333941)
- <span id="page-18-1"></span>36. Rasheed, S.A.; Harder, L.D. Economic motivation for plant species preferences of pollen-collecting bumble bees. *Ecol. Entomol.* **1997**, *22*, 209–219. [\[CrossRef\]](https://doi.org/10.1046/j.1365-2311.1997.t01-1-00059.x)
- <span id="page-18-2"></span>37. Thomas, S.G.; Frankin-Tong, E.F. Self-incompatibility triggers programmed cell death in *Papaver* pollen. *Nature* **2004**, *429*, 305–309. [\[CrossRef\]](https://doi.org/10.1038/nature02540) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15152254)
- <span id="page-18-3"></span>38. Dafni, A. Autumnal and winter pollination adaptations under Mediterranean conditions. *Bocconea* **1996**, *5*, 171–181.
- <span id="page-18-4"></span>39. Petanidou, T.; Vokou, D. Pollination ecology of Labiatae in a phryganic (East Mediterranean) ecosystem. *Am. J. Bot.* **1993**, *80*, 892–899. [\[CrossRef\]](https://doi.org/10.1002/j.1537-2197.1993.tb15310.x)
- <span id="page-18-5"></span>40. Levin, D.A.; Anderson, W.W. Competition for pollinators between simultaneous flowering species. *Am. Nat.* **1970**, *104*, 455–467. [\[CrossRef\]](https://doi.org/10.1086/282680)
- <span id="page-18-6"></span>41. Westwood, J.H.; Tominaga, T.; Weller, S.C. Characterization and breakdown of self-incompatibility in field bindweed (*Convolvulus arvensis* L.). *J. Hered.* **1997**, *88*, 459–465. [\[CrossRef\]](https://doi.org/10.1093/oxfordjournals.jhered.a023137)
- <span id="page-18-7"></span>42. Motten, A.F. Determinants of outcrossing rate in a predominantly self-fertilizing weed, *Datura stramonium* (Solanaceae). *Am. J. Bot.* **1992**, *79*, 419–427. [\[CrossRef\]](https://doi.org/10.1002/j.1537-2197.1992.tb14569.x)
- <span id="page-18-8"></span>43. Larson, B.; Kevan, P.; Inouye, D. Flies and flowers: Taxonomic diversity of anthophiles and pollinators. *Canad. Entomol.* **2001**, *133*, 439–465. [\[CrossRef\]](https://doi.org/10.4039/Ent133439-4)
- <span id="page-18-9"></span>44. Kearns, C.A.; Inouye, D.W.; Waser, N.M. Endangered mutualisms: The Conservation of Plant-Pollinator Interactions. *Ann. Rev. Ecol. Syst.* **1998**, *29*, 83–112. [\[CrossRef\]](https://doi.org/10.1146/annurev.ecolsys.29.1.83)
- <span id="page-18-10"></span>45. Jennersten, O. Flower visitation and pollination efficiency of some North European butterfly. *Oecologia* **1984**, *63*, 80–89. [\[CrossRef\]](https://doi.org/10.1007/BF00379789)
- <span id="page-18-11"></span>46. Sakai, S. A review of brood-site pollination mutualism: Plants providing breeding sites for their pollinators. *J. Plant Res.* **2002**, *115*, 161–168. [\[CrossRef\]](https://doi.org/10.1007/s102650200021)
- <span id="page-18-12"></span>47. Bosch, J.; Retana, J.; Cerdá, X. Flowering phenology, floral traits and pollinator composition on a herbaceous Mediterranean plant community. *Oecologia* **1997**, *101*, 583–591. [\[CrossRef\]](https://doi.org/10.1007/s004420050120)
- <span id="page-18-13"></span>48. Beattie, A.J.; Turnbull, C.L.; Knox, R.B.; Williams, E.G. Ant inhibition of pollen function: A possible reason why ant pollination is rare. *Am. J. Bot.* **1984**, *71*, 421–426. [\[CrossRef\]](https://doi.org/10.1002/j.1537-2197.1984.tb12527.x)
- <span id="page-18-14"></span>49. Junker, R.; Chung, A.Y.C.; Blüthgen, N. Interaction between flowers, ants and pollinators: Additional evidence for floral repellence against ants. *Ecol. Res.* **2007**, *22*, 665–670. [\[CrossRef\]](https://doi.org/10.1007/s11284-006-0306-3)
- <span id="page-18-15"></span>50. Stout, J.C.; Allen, J.A.; Goulson, D. Nectar robbing, forager efficiency and seed set: Bumblebees foraging on the self incompatible plant *Linaria vulgaris* (Scrophulariaceae). *Acta Oecol.* **2000**, *21*, 277–283. [\[CrossRef\]](https://doi.org/10.1016/S1146-609X(00)01085-7)
- <span id="page-18-16"></span>51. Maloof, J.E.; Inouye, D.W. Is nectar robbers' cheaters or mutualists? *Ecology* **2000**, *81*, 2651–2661. [\[CrossRef\]](https://doi.org/10.1890/0012-9658(2000)081[2651:ANRCOM]2.0.CO;2)
- <span id="page-18-17"></span>52. Bourgeois, B.; Munoz, F.; Fried, G.; Mahaut, L.; Armengot, L.; Denelle, P.; Storkey, J.; Gaba, S.; Violle, C. What makes a weed a weed? A large-scale evaluation of arable weeds through a functional lens. *Am. J. Bot.* **2019**, *106*, 90–100. [\[CrossRef\]](https://doi.org/10.1002/ajb2.1213) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30633823)
- <span id="page-18-18"></span>53. Storkey, J.; Westbury, D.B. Managing arable weeds for biodiversity. *Pest Manag. Sci.* **2007**, *63*, 517–523. [\[CrossRef\]](https://doi.org/10.1002/ps.1375) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17437252)
- <span id="page-18-19"></span>54. Johnson, S.D.; Steiner, K.E. Generalization versus specialization in plant pollination systems. *Trends Ecol. Evol.* **2000**, *15*, 140–143. [\[CrossRef\]](https://doi.org/10.1016/S0169-5347(99)01811-X) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/10717682)
- <span id="page-18-20"></span>55. Huang, S.Q. Debates enrich our understanding of pollination biology. *Trends Ecol. Evol.* **2006**, *21*, 233–234. [\[CrossRef\]](https://doi.org/10.1016/j.tree.2005.12.004)
- <span id="page-18-21"></span>56. Sutcliffe, O.L.; Kay, Q.O.N. Changes in the arable flora of central southern England since the 1960s. *Biol. Conserv.* **2000**, *93*, 1–8. [\[CrossRef\]](https://doi.org/10.1016/S0006-3207(99)00119-6)
- <span id="page-18-22"></span>57. Aigner, P.A. Optimality modelling and fitness trade-offs: When should plants become pollinator specialist? *Oikos* **2001**, *95*, 177–184. [\[CrossRef\]](https://doi.org/10.1034/j.1600-0706.2001.950121.x)
- <span id="page-18-23"></span>58. Giurfa, M.; Dafni, A.; Neal, P.R. Floral symmetry and its role in plant-pollinator systems. *Inter. J. Plant Sci.* **1999**, *160* (Suppl. S6), 541–550. [\[CrossRef\]](https://doi.org/10.1086/314214) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/10572021)
- <span id="page-18-24"></span>59. Nilson, L.A. Deep flowers for long tongues. *Trends Ecol. Evol.* **1998**, *13*, 259–260. [\[CrossRef\]](https://doi.org/10.1016/S0169-5347(98)01359-7)
- <span id="page-18-25"></span>60. Fenster, C.B.; Armbruster, W.S.; Wilson, P.; Dudash, M.R.; Thomson, J.D. Pollination syndromes and floral specialization. *Ann. Rev. Ecol. Syst.* **2004**, *35*, 375–403. [\[CrossRef\]](https://doi.org/10.1146/annurev.ecolsys.34.011802.132347)
- <span id="page-18-26"></span>61. Konzmann, S.; Kluth, M.; Karadana, D.; Lunau, K. Pollinator effectiveness of a specialist bee exploiting a generalist plant-tracking pollen transfer by *Heriades truncorum* with quantum dots. *Apidologie* **2020**, *51*, 201–211. [\[CrossRef\]](https://doi.org/10.1007/s13592-019-00700-0)
- <span id="page-18-27"></span>62. Klumpers, S.G.; Stang, M.; Klinkhamer, P.G. Foraging efficiency and size matching in a plant–pollinator community: The importance of sugar content and tongue length. *Ecol. Lett.* **2019**, *22*, 469–479. [\[CrossRef\]](https://doi.org/10.1111/ele.13204)
- <span id="page-18-28"></span>63. Armbruster, W.S. Evolution of floral form: Electrostatic forces, pollination, and adaptative compromise. *New Phytol.* **2001**, *152*, 181–183. [\[CrossRef\]](https://doi.org/10.1046/j.0028-646X.2001.00268.x)
- <span id="page-18-29"></span>64. Fenner, M. The phenology of growth and reproduction in plants. *Perspect. Plant Ecol. Evol. Syst.* **1998**, *1*, 78–91. [\[CrossRef\]](https://doi.org/10.1078/1433-8319-00053)
- <span id="page-18-30"></span>65. Herrera, C.M. Floral biology, microclimate, and pollination by ectothermic bees in an early-blooming herb. *Ecology* **1995**, *76*, 221–228. [\[CrossRef\]](https://doi.org/10.2307/1940644)
- <span id="page-18-31"></span>66. Blionis, G.J.; Vokou, D. Pollination ecology of *Campanula* species on Mt Olympus, Greece. *Ecography* **2001**, *24*, 287–297. [\[CrossRef\]](https://doi.org/10.1034/j.1600-0587.2001.240306.x)
- <span id="page-18-32"></span>67. Elzinga, J.A.; Atlan, A.; Biere, A.; Gigord, L.; Weis, A.E.; Bernasconi, G. Time after time: Flowering phenology and biotic interactions. *Trends Ecol. Evol.* **2007**, *22*, 432–439. [\[CrossRef\]](https://doi.org/10.1016/j.tree.2007.05.006) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17573151)
- <span id="page-19-0"></span>68. Davis, S.L.; Delph, L.F. Prior selfing and gynomonoecy in *Silene noctiflora* L. (Caryophyllaceae): Opportunities for enhanced outcrossing and reproductive assurance. *Int. J. Plant Sci.* **2005**, *166*, 475–480. [\[CrossRef\]](https://doi.org/10.1086/428630)
- <span id="page-19-1"></span>69. Barthelmess, E.L.; Richards, C.M.; McCauley, D.E. Relative effects of nocturnal vs diurnal pollinators and distance on gene flow in small *Silene alba* populations. *New Phytol.* **2006**, *169*, 689–698. [\[CrossRef\]](https://doi.org/10.1111/j.1469-8137.2005.01580.x)
- <span id="page-19-2"></span>70. Corbet, A.A. Nectar sugar content: Estimating standing crop and secretion in the field. *Apidologie* **2003**, *34*, 1–10. [\[CrossRef\]](https://doi.org/10.1051/apido:2002049)
- <span id="page-19-3"></span>71. Dupont, Y.L.; Hansen, D.M.; Rasmussen, J.T.; Olsen, J.M. Evolutionary changes in nectar sugar composition with switches bird and insect pollination: The Canarian bird-flower element revisited. *Funct. Ecol.* **2004**, *18*, 670–676. [\[CrossRef\]](https://doi.org/10.1111/j.0269-8463.2004.00891.x)
- <span id="page-19-4"></span>72. Petanidou, T.; Van Laere, A.; Ellis, W.N.; Smets, E. What shapes amino acid and sugar composition in Mediterranean floral nectars? *Oikos* **2006**, *115*, 155–169. [\[CrossRef\]](https://doi.org/10.1111/j.2006.0030-1299.14487.x)
- <span id="page-19-5"></span>73. Gardener, M.C.; Gillman, P.M. The taste of nectar—A neglected area of pollination ecology. *Oikos* **2002**, *98*, 552–557. [\[CrossRef\]](https://doi.org/10.1034/j.1600-0706.2002.980322.x)
- <span id="page-19-6"></span>74. Gardener, M.C.; Gillman, P.M. The effects of soil fertilizer on amino acids in the floral nectar of corncockle, *Agrostemma githago* (Caryophyllaceae). *Oikos* **2001**, *92*, 101–106. [\[CrossRef\]](https://doi.org/10.1034/j.1600-0706.2001.920112.x)
- <span id="page-19-7"></span>75. Gange, A.C.; Smith, A.K. Arbuscolar mycorrhyzal fungi influence visitation rates of pollinating insects. *Ecol. Entomol.* **2005**, *30*, 600–606. [\[CrossRef\]](https://doi.org/10.1111/j.0307-6946.2005.00732.x)
- <span id="page-19-8"></span>76. Adler, L.S. The ecological significance of toxic nectar. *Oikos* **2000**, *91*, 409–420. [\[CrossRef\]](https://doi.org/10.1034/j.1600-0706.2000.910301.x)
- <span id="page-19-9"></span>77. Narbona, E.; Arista, M.; Whittall, J.B.; Camargo, M.G.G.; Shrestha, M. The role of flower color in angiosperm evolution. *Front. Plant Sci.* **2021**, *12*, 736998. [\[CrossRef\]](https://doi.org/10.3389/fpls.2021.736998)
- <span id="page-19-10"></span>78. Menzel, R.; Shmida, A. The ecology of flower colours and the natural colour vision of insect pollinators: The Israeli flora as a case study. *Biol. Rev.* **1993**, *68*, 81–120. [\[CrossRef\]](https://doi.org/10.1111/j.1469-185X.1993.tb00732.x)
- <span id="page-19-11"></span>79. Yoshioka, Y.; Horisaki, A.; Kobaiyashi, K.; Syfaruddin, S.; Niikura, S.; Ninomiya, S.; Ohsawa, R. Intraspecific variation in the ultraviolet colour proportion of flowers in *Brassica rapa* L. *Plant Breed.* **2005**, *124*, 551–556. [\[CrossRef\]](https://doi.org/10.1111/j.1439-0523.2005.01132.x)
- <span id="page-19-12"></span>80. Gray, L.A.; Varga, S.; Soulsbury, C.D. Floral ultraviolet absorbance area responds plastically to ultraviolet irradiance in *Brassica rapa*. *Plant-Environ. Interact.* **2022**, *3*, 203–211. [\[CrossRef\]](https://doi.org/10.1002/pei3.10091)
- <span id="page-19-13"></span>81. Nakano, C.; Washitani, I. Variability and specialization of plant-pollinator systems in a nothern maritime grassland. *Ecol. Res.* **2003**, *18*, 221–246. [\[CrossRef\]](https://doi.org/10.1046/j.1440-1703.2003.00550.x)
- <span id="page-19-14"></span>82. Goulson, D. Are insects flower constant because they use search images to find flowers? *Oikos* **2000**, *88*, 547–552. [\[CrossRef\]](https://doi.org/10.1034/j.1600-0706.2000.880311.x)
- <span id="page-19-15"></span>83. Johnson, S.D.; Dafni, A. Response of bee-flies to the shape and pattern of model flowers: Implications for floral evolution in a Mediterranean herb. *Funct. Ecol.* **1998**, *12*, 289–297. [\[CrossRef\]](https://doi.org/10.1046/j.1365-2435.1998.00175.x)
- <span id="page-19-16"></span>84. Montgomery, B.R. Effect of introduced *Euphorbia esula* on the pollination of *Viola pedatifida*. *Botany* **2009**, *87*, 283–292.
- <span id="page-19-17"></span>85. Van Schie, C.C.N.; Haring, M.A.; Schuurink, R.C. Regulation of terpenoid and benzenoid production in flowers. *Curr. Opin. Plant Biol.* **2006**, *9*, 203–206. [\[CrossRef\]](https://doi.org/10.1016/j.pbi.2006.01.001) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16458042)
- <span id="page-19-18"></span>86. Van Kleunen, M.; Manning, J.C.; Pasqualetto, V.; Johnson, S.D. Phylogenetically independent associations between autonomous self-fertilization and plant invasiveness. *Am. Nat.* **2008**, *171*, 195–201. [\[CrossRef\]](https://doi.org/10.1086/525057) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18197772)
- <span id="page-19-19"></span>87. Booth, B.D.; Swanton, C.J. Assembly theory applied to weed communities. *Weed Sci.* **2002**, *50*, 2–13. [\[CrossRef\]](https://doi.org/10.1614/0043-1745(2002)050[0002:AIATAT]2.0.CO;2)
- <span id="page-19-20"></span>88. Snell, R.; Aarssen, L.W. Life history traits in selfing versus outcrossing annuals: Exploring the 'time-limitation' hypothesis for the fitness benefit of self-pollination. *BMC Ecol.* **2005**, *5*, 1–14. [\[CrossRef\]](https://doi.org/10.1186/1472-6785-5-2) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15707481)
- <span id="page-19-21"></span>89. Brenner, D.M.; Baltensperger, D.D.; Kulakow, P.A.; Lehmann, J.W.; Myers, R.L.; Slabbert, M.M.; Sleugh, B.B. Genetic resources and breeding of *Amaranthus*. *Plant Breed. Rev.* **2000**, *19*, 227–285.
- <span id="page-19-22"></span>90. Costea, M.; Weaver, S.E.; Tardif, F.J. The biology of Canadian weeds. 130. *Amaranthus retroflexus* L., *A. powellii* S. Watson and *A. hybridus* L. *Can. J. Plant Sci.* **2004**, *130*, 631–668. [\[CrossRef\]](https://doi.org/10.4141/P02-183)
- <span id="page-19-23"></span>91. Prokop, P. Urban environment decreases pollinator availability, fertility, and prolongs anthesis in the field bindweed (*Convolvulus arvensis* Linnaeus, 1753). *Plant Sign. Behav.* **2024**, *19*, 2325225. [\[CrossRef\]](https://doi.org/10.1080/15592324.2024.2325225)
- <span id="page-19-24"></span>92. Verkleij, J.A.C.; De Boer, A.M.; Lugtenborg, T.F. On the ecogenetics of *Stellaria media* (L.) Vill. And *Stellaria pallida* (Dum.) pire from abandoned arable field. *Oecologia* **1980**, *46*, 354–359. [\[CrossRef\]](https://doi.org/10.1007/BF00346264) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28310044)
- <span id="page-19-25"></span>93. Zimmerman, C.A. A comparison of breeding systems and seed physiologies in three species of *Portulaca* L. *Ecology* **1977**, *58*, 860–868. [\[CrossRef\]](https://doi.org/10.2307/1936221)
- <span id="page-19-26"></span>94. Gibbs, P.E.; Talavera, S. Breeding system studies with three species of *Anagallis* (Primulaceae): Self-incompatibility and reduced female fertility in *A. monelli* L. *Ann. Bot.* **2001**, *88*, 139–144. [\[CrossRef\]](https://doi.org/10.1006/anbo.2001.1439)
- <span id="page-19-27"></span>95. Jiménez-Lobato, V.; Martínez-Borda, E.; Núñez-Farfán, J.; Valverde, P.L.; Cruz, L.L.; López-Velázquez, A.; Santos-Gally, R.; Arroyo, J. Changes in floral biology and inbreeding depression in native and invaded regions of *Datura stramonium*. *Plant Biol.* **2018**, *20*, 214–223. [\[CrossRef\]](https://doi.org/10.1111/plb.12658)
- <span id="page-19-28"></span>96. Motten, A.F.; Stone, J.L. Heritability of stigma position and the effect of stigma-anther separation on outcrossing in a predominantly self-fertilizing weed, *Datura stramonium* (Solanaceae). *Am. J. Bot.* **2000**, *87*, 339–347. [\[CrossRef\]](https://doi.org/10.2307/2656629) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/10718994)
- <span id="page-19-29"></span>97. Boucher, J.J.; Ireland, H.S.; Wang, R.; David, K.M.; Schaffer, R.J. The genetic control of herkogamy. *Funct. Plant Biol.* **2024**, *51*, 5. [\[CrossRef\]](https://doi.org/10.1071/FP23315) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38687848)
- <span id="page-19-30"></span>98. Wheeler, M.J.; Armstrong, S.A.; Franklin-Tong, V.E.; Franklin, F.C.H. Genomic organization of the *Papaver rhoeas* selfincompatibility S 1 locus. *J. Exp. Bot.* **2023**, *54*, 131–139. [\[CrossRef\]](https://doi.org/10.1093/jxb/erg006) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12456763)
- <span id="page-19-31"></span>99. Lundquist, A. The self-incompatibility system in *Ranunculus repens* (Ranunculaceae). *Hereditas* **1994**, *120*, 151–157. [\[CrossRef\]](https://doi.org/10.1111/j.1601-5223.1994.00151.x)
- <span id="page-19-32"></span>100. Kay, Q.O.N. Biological flora of the British Isles: *Anthemis cotula* L. *J. Ecol.* **1971**, *59*, 637–648. [\[CrossRef\]](https://doi.org/10.2307/2258337)
- <span id="page-20-0"></span>101. Stevens, J.P.; Kay, Q.O.N. The number, dominance relationships and frequencies of self-incompatibility alleles in a natural population of *Sinapis arvensis* L. in South Wales. *Heredity* **1989**, *62*, 199–205. [\[CrossRef\]](https://doi.org/10.1038/hdy.1989.29)
- <span id="page-20-1"></span>102. Sampson, D.R. Frequency and distribution of self-incompatibility alleles in *Raphanus raphanistrum*. *Genetics* **1967**, *56*, 241–251. [\[CrossRef\]](https://doi.org/10.1093/genetics/56.2.241)
- <span id="page-20-2"></span>103. Benvenuti, S.; Mazzoncini, M. Entomogamy in wildflowers: What level of pollinator biodiversity is required? *Acta Oecol.* **2021**, *111*, 103737. [\[CrossRef\]](https://doi.org/10.1016/j.actao.2021.103737)
- <span id="page-20-3"></span>104. Stanton, M.L.; Snow, A.A.; Handel, S.N.; Bereczky, J. The impact of a flower-color polymorphism on mating patterns in experimental populations of Wild Radish (*Raphanus raphanistrum* L.). *Evolution* **1989**, *43*, 335–346. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28568562)
- <span id="page-20-4"></span>105. Devaux, C.; Lepers, C.; Porcher, E. Constraints imposed by pollinator behaviour on the ecology and evolution of plant mating systems. *J. Evol. Biol.* **2014**, *27*, 1413–1430. [\[CrossRef\]](https://doi.org/10.1111/jeb.12380) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24750302)
- <span id="page-20-5"></span>106. Eckert, C.G. Contributions of autogamy and geitonogamy to self-fertilization in a mass-flowering, clonal plant. *Ecology* **2000**, *81*, 532–542. [\[CrossRef\]](https://doi.org/10.1890/0012-9658(2000)081[0532:COAAGT]2.0.CO;2)
- <span id="page-20-6"></span>107. De Jong, T.J. Geitonogamy: The neglected side of selfing. *Trends Ecol. Evol.* **1993**, *8*, 321–325. [\[CrossRef\]](https://doi.org/10.1016/0169-5347(93)90239-L)
- <span id="page-20-7"></span>108. Koul, P.; Koul, A.K.; Hamal, I.A. Reproductive biology of wild and cultivated carrot (*Daucus carota* L.). *New Phytol.* **1989**, *112*, 437–443. [\[CrossRef\]](https://doi.org/10.1111/j.1469-8137.1989.tb00335.x)
- <span id="page-20-8"></span>109. Lamborn, E.; Ollerton, J. Experimental assessment of the functional morphology of inflorescences of *Daucus carota* (Apiaceae): Testing the "fly catcher effect". *Funct. Ecol.* **2000**, *14*, 445–454. [\[CrossRef\]](https://doi.org/10.1046/j.1365-2435.2000.00440.x)
- <span id="page-20-9"></span>110. De Jong, T.J. From pollen dynamics to adaptative dynamics. *Plant Spec. Biol.* **2000**, *15*, 31–41. [\[CrossRef\]](https://doi.org/10.1046/j.1442-1984.2000.00028.x)
- <span id="page-20-10"></span>111. Ishii, H.S.; Harder, L.D. The size of individual *Delphinum* flowers and the opportunity for geitonogamous pollination. *Funct. Ecol.* **2006**, *20*, 1115–1123. [\[CrossRef\]](https://doi.org/10.1111/j.1365-2435.2006.01181.x)
- <span id="page-20-11"></span>112. Klinkhamer, P.G.L.; De Jong, T.L. Effects of plant density and sex differential reward visitation in the protandrous *Echium vulgare* (Boraginaceae). *Oikos* **1990**, *57*, 399–405. [\[CrossRef\]](https://doi.org/10.2307/3565970)
- <span id="page-20-12"></span>113. Leiss, K.A.; Klinkhammer, G.L. Genotype by environment interactions in the nectar production of *Echium vulgare*. *Funct. Ecol.* **2005**, *19*, 454–459. [\[CrossRef\]](https://doi.org/10.1111/j.1365-2435.2005.00986.x)
- <span id="page-20-13"></span>114. Westerbergh, A.; Saura, A. Gene flow and pollinator behaviour in *Silene dioica* populations. *Oikos* **1994**, *71*, 215–224. [\[CrossRef\]](https://doi.org/10.2307/3546269)
- <span id="page-20-14"></span>115. Blair, A.C.; Wolfe, L.M. The evolution of an invasive plant: An experimental study with *Silene latifolia*. *Ecology* **2004**, *85*, 3035–3042. [\[CrossRef\]](https://doi.org/10.1890/04-0341)
- <span id="page-20-15"></span>116. Petanidou, T. Sugars in Mediterranean floral nectars: An ecological and evolutionary approach. *J. Chem. Ecol.* **2005**, *31*, 1065–1088. [\[CrossRef\]](https://doi.org/10.1007/s10886-005-4248-y)
- <span id="page-20-16"></span>117. Fahn, A. Secretory tissues in vascular plants. *New Phytol.* **1988**, *108*, 229–257. [\[CrossRef\]](https://doi.org/10.1111/j.1469-8137.1988.tb04159.x)
- <span id="page-20-17"></span>118. Albrecht, H.; Mattheis, A. The effect of organic and integrated farming on rare arable weeds on the Forschungsverbund Agrarökosysteme München (FAM) research station in southern Bavaria. *Biol. Conserv.* **1998**, *86*, 347–356. [\[CrossRef\]](https://doi.org/10.1016/S0006-3207(98)00028-7)
- <span id="page-20-18"></span>119. Levin, D.A. Competition for pollinator service: A stimulus for the evolution of the autogamy. *Evolution* **1971**, *26*, 668–674. [\[CrossRef\]](https://doi.org/10.2307/2407061)
- <span id="page-20-19"></span>120. Marshall, E.J.P.; Brown, V.K.; Boatman, N.D.; Lutman, P.J.W.; Squire, G.R.; Ward, L.K. The role of weeds in supporting biological diversity within crop fields. *Weed Res.* **2003**, *43*, 77–89. [\[CrossRef\]](https://doi.org/10.1046/j.1365-3180.2003.00326.x)
- <span id="page-20-20"></span>121. Robinson, R.A.; Sutherland, W.J. Post-war changes in arable farming and biodiversity in Great Britain. *J. Appl. Ecol.* **2002**, *39*, 157–176. [\[CrossRef\]](https://doi.org/10.1046/j.1365-2664.2002.00695.x)
- <span id="page-20-21"></span>122. Munoz, F.; Fried, G.; Armengot, L.; Bourgeois, B.; Bretagnolle, V.; Chadoeuf, J.; Mahaut, L.; Plumejeaud, C.; Storkey, J.; Violle, C.; et al. Ecological specialization and rarity of arable weeds: Insights from a comprehensive survey in France. *Plants* **2020**, *9*, 824. [\[CrossRef\]](https://doi.org/10.3390/plants9070824) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32630061)
- <span id="page-20-22"></span>123. Biesmeijer, J.C.; Roberts, S.P.M.; Reemer, M.; Ohlemüller, R.; Edwards, M.; Peeters, T.; Schaffers, A.P.; Potts, S.G.; Kleukers, R.; Thomas, C.D.; et al. Parallel Declines in Pollinators and Insect-Pollinated Plants in Britain and the Netherlands. *Science* **2006**, *313*, 351–354. [\[CrossRef\]](https://doi.org/10.1126/science.1127863)
- <span id="page-20-23"></span>124. Lázaro, A.; Jakobsson, A.; Totland, Ø. How do pollinator visitation rate and seed set relate to species' floral traits and community context? *Oecologia* **2013**, *173*, 881–893. [\[CrossRef\]](https://doi.org/10.1007/s00442-013-2652-5) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23579571)
- <span id="page-20-24"></span>125. Feldman, T.S. Pollinator aggregative and functional responses to flower density: Does pollinator response to patches of plants accelerate at low densities? *Oikos* **2006**, *115*, 128–140. [\[CrossRef\]](https://doi.org/10.1111/j.2006.0030-1299.14493.x)
- <span id="page-20-25"></span>126. Pontin, D.R.; Wade, M.R.; Kehrli, P.; Wratten, S.D. Attractiveness of single and multiple species flower patches to beneficial insects in agroecosystems. *Ann. Appl. Biol.* **2005**, *148*, 39–47. [\[CrossRef\]](https://doi.org/10.1111/j.1744-7348.2005.00037.x)
- <span id="page-20-27"></span><span id="page-20-26"></span>127. Chancellor, R.J. A preliminary survey of arable weeds in Britain. *Weed Res.* **1977**, *17*, 283–289. [\[CrossRef\]](https://doi.org/10.1111/j.1365-3180.1977.tb00479.x)
- 128. Chancellor, R.J. Decline of arable weeds during 20 years in soil under grass on the periodicity of seedling emergence after cultivation. *J. Appl. Ecol.* **1983**, *23*, 631–637. [\[CrossRef\]](https://doi.org/10.2307/2404041)
- <span id="page-20-28"></span>129. Baessler, C.; Klotz, S. Effects of changes in agricultural land-use on landscape structure and arable weed vegetation over the last 50 years. *Agric. Ecosyst. Environ.* **2006**, *115*, 43–50. [\[CrossRef\]](https://doi.org/10.1016/j.agee.2005.12.007)
- <span id="page-20-29"></span>130. Andreasen, C.; Stryhn, H.; Streibig, J.C. Decline of the flora in Danish arable field. *J. Appl. Ecol.* **1996**, *33*, 619–626. [\[CrossRef\]](https://doi.org/10.2307/2404990)
- <span id="page-20-30"></span>131. Chamorro, L.; Masalles, R.M.; Sans, F.X. Arable weed decline in Northeast Spain: Does organic farming recover functional biodiversity? *Agric. Ecosys. Environ.* **2016**, *223*, 1–9. [\[CrossRef\]](https://doi.org/10.1016/j.agee.2015.11.027)
- <span id="page-21-0"></span>132. Dutoit, T.; Buisson, E.; Roche, P.; Didier, A. Land history and botanical changes in the calcareous hillsides of Upper-Normandy (north-western France): New implications for their conservation management. *Biol. Conserv.* **2004**, *115*, 1–19. [\[CrossRef\]](https://doi.org/10.1016/S0006-3207(03)00089-2)
- <span id="page-21-1"></span>133. Gu, H.; Cao, A.; Walter, G.H. Host selection and utilisation of *Sonchus oleraceus* (Asteraceae) by *Helicoverpa armigera* (Lepidoptera: Noctuidae): A genetic analysis. *Ann. Appl. Biol.* **2001**, *138*, 293–299. [\[CrossRef\]](https://doi.org/10.1111/j.1744-7348.2001.tb00114.x)
- <span id="page-21-2"></span>134. Witt, T.; Jürgens, A.; Gottsberger, G. Nectar sugar composition of European Caryophylloideae (Caryophyllaceae) in relation to flower length, pollination biology and phylogeny. *J. Evol. Biol* **2013**, *26*, 2244–2259. [\[CrossRef\]](https://doi.org/10.1111/jeb.12224)
- <span id="page-21-3"></span>135. Shapiro, A.M. The California urban butterfly is dependent on alien plants. *Diver. Distrib.* **2002**, *8*, 31–40. [\[CrossRef\]](https://doi.org/10.1046/j.1366-9516.2001.00120.x)
- <span id="page-21-4"></span>136. Wiklund, C. Generalist vs. specialist oviposition behaviour in *Papilio machaon* (Lepidoptera) and functional aspects of the hierarchy of oviposition preferences. *Oikos* **1981**, *36*, 163–170. [\[CrossRef\]](https://doi.org/10.2307/3544441)
- <span id="page-21-5"></span>137. Alam, S.M.; Bashar, A.; Aich, U.; Akand, S.; Rahman, S. Colonization and strategic stratification of butterfly *Pachliopta aristolochiae* (Lepidoptera: Papilionidae) and its relationship with the host plant *Aristolochia indica* (Piperales: Aristolochiaceae). *J. Biodiv. Conserv. Biores. Manag.* **2017**, *3*, 45–54. [\[CrossRef\]](https://doi.org/10.3329/jbcbm.v3i1.36760)
- <span id="page-21-6"></span>138. Konvicka, M.; Novak, J.; Benes, J.; Fric, Z.; Bradley, J.; Keil, P.; Hrcek, J.; Chobot, K.; Marhoul, P. The last population of the Woodland Brown butterfly (*Lopinga achine*) in the Czech Republic: Habitat use, demography and site management. *J. Insect Conserv.* **2008**, *12*, 549–560. [\[CrossRef\]](https://doi.org/10.1007/s10841-007-9087-4)
- <span id="page-21-7"></span>139. Courtney, S.P. Coevolution of pierid butterflies and their cruciferous foodplants IV. Crucifer apparency and *Anthocharis cardamines* (L.) oviposition. *Oecologia* **1982**, *52*, 258–265. [\[CrossRef\]](https://doi.org/10.1007/BF00363846)
- <span id="page-21-8"></span>140. Novak, I.; Severa, F. *Impariamo a Conoscere le Farfalle*; Istituto Geografico De Agostini: Novara, Italy, 1980; 352p.
- <span id="page-21-9"></span>141. Bowers, M.D. The role of iridoid glycosides in host-plant specificity of checkerspot butterflies. *J. Chem. Ecol.* **1983**, *9*, 475–493. [\[CrossRef\]](https://doi.org/10.1007/BF00990220)
- <span id="page-21-10"></span>142. Warren, M.S. The ecology and conservation of the heath fritillary butterfly, *Mellicta hatalia* I. host selection and phenology. *J. Appl. Ecol.* **1987**, *24*, 467–482. [\[CrossRef\]](https://doi.org/10.2307/2403887)
- <span id="page-21-11"></span>143. Dennis, R.L.H.; Hodgson, J.G.; Grenyer, R.; Shreeve, T.G.; Roy, D.B. Host plants and butterfly biology. *Ecol. Entomol.* **2004**, *29*, 12–26. [\[CrossRef\]](https://doi.org/10.1111/j.1365-2311.2004.00572.x)
- <span id="page-21-12"></span>144. Koh, L.P.; Sodhi, N.S.; Brook, B.W. Co-extinctions of tropical butterflies and their host plants. *Biotropica* **2004**, *36*, 272–274.
- <span id="page-21-13"></span>145. DeVries, P.J.; Baker, I. Butterfly exploitation of a plant–ant mutualism: Adding insult to herbivory. *J. N. Y. Entomol. Soc.* **1989**, *97*, 332–340.
- <span id="page-21-14"></span>146. Fiedler, K. Ants that associate with Lycaeninae butterfly larvae: Diversity, ecology and biogeography. *Div. Distrib.* **2001**, *7*, 45–60. [\[CrossRef\]](https://doi.org/10.1046/j.1472-4642.2001.00096.x)
- <span id="page-21-15"></span>147. Kells, A.R.; Goulson, D. Preferred nesting sites of bumblebee queens (Hymenoptera: Apidae) in agroecosystems in the UK. *Biol. Conserv.* **2003**, *109*, 165–174. [\[CrossRef\]](https://doi.org/10.1016/S0006-3207(02)00131-3)
- <span id="page-21-16"></span>148. Potts, S.G.; Petanidou, T.; Roberts, S.; O'Toole, C.; Hulbert, A.; Willmer, P. Plant-pollinator biodiversity and pollination services in a complex Mediterranean landscape. *Biol. Conserv.* **2006**, *129*, 519–529. [\[CrossRef\]](https://doi.org/10.1016/j.biocon.2005.11.019)
- <span id="page-21-17"></span>149. Ferguson, D.J.; Li, X.; Yeates, D.K. Silent, underground warfare of flies: An endoparasitic bee fly (Diptera: Bombyliidae) larva parasitising a predatory assassin fly (Diptera: Asilidae) larva. *Austral Entomol.* **2020**, *59*, 582–592. [\[CrossRef\]](https://doi.org/10.1111/aen.12482)
- <span id="page-21-18"></span>150. Kephart, S. Pollination mutualisms in Caryophyllaceae. *New Phytol.* **2006**, *169*, 637. [\[CrossRef\]](https://doi.org/10.1111/j.1469-8137.2006.01656.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16441744)
- <span id="page-21-19"></span>151. Jones, R.E. Behavioural evolution in the cabbage butterfly (*Pieris rapae*). *Oecologia* **1987**, *72*, 69–76. [\[CrossRef\]](https://doi.org/10.1007/BF00385047)
- <span id="page-21-20"></span>152. Dufaÿ, M.; Anstett, M.C. Conflicts between plants and pollinators that reproduce within inflorescences: Evolutionary variations on a theme. *Oikos* **2003**, *100*, 3–14. [\[CrossRef\]](https://doi.org/10.1034/j.1600-0706.2003.12053.x)
- <span id="page-21-21"></span>153. Rundlölf, M.; Smith, H.G. The effect of organic farming on butterfly diversity depends on landscape context. *J. Appl. Ecol.* **2006**, *43*, 1121–1127. [\[CrossRef\]](https://doi.org/10.1111/j.1365-2664.2006.01233.x)
- <span id="page-21-22"></span>154. Heigland, S.J.; Boeke, L. Relationships between the density and diversity of floral resources and flower visitor activity in a temperate grassland community. *Ecol. Entomol.* **2006**, *31*, 532–538. [\[CrossRef\]](https://doi.org/10.1111/j.1365-2311.2006.00812.x)
- <span id="page-21-23"></span>155. Dukas, R.; Morse, D.H. Crab spiders affect flower visitation by bees. *Oikos* **2003**, *101*, 157–163. [\[CrossRef\]](https://doi.org/10.1034/j.1600-0706.2003.12143.x)
- <span id="page-21-24"></span>156. Suttle, K.B. Pollinators as mediators of top-down effects on plants. *Ecol. Lett.* **2003**, *6*, 688–694. [\[CrossRef\]](https://doi.org/10.1046/j.1461-0248.2003.00490.x)
- <span id="page-21-25"></span>157. Su, Q.; Qi, L.; Zhang, W.; Yun, Y.; Zhao, Y.; Peng, Y. Biodiversity survey of flower-visiting spiders based on literature review and field study. *Environ. Entomol.* **2020**, *49*, 673–682. [\[CrossRef\]](https://doi.org/10.1093/ee/nvaa022) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32281625)
- <span id="page-21-26"></span>158. Gabriel, D.; Tscharntke, T. Local diversity of arable weeds increases with landscape complexity. *Agric. Ecosyst. Environ.* **2005**, *7*, 85–93. [\[CrossRef\]](https://doi.org/10.1016/j.ppees.2005.04.001)
- <span id="page-21-27"></span>159. Gerowitt, B. Development and control of weeds in arable farming systems. *Agric. Ecosyst. Environ.* **2003**, *98*, 247–254. [\[CrossRef\]](https://doi.org/10.1016/S0167-8809(03)00084-7)
- <span id="page-21-28"></span>160. Bengtsson, J.; Ahnström, J.; Weitbull, A.C. The effects of organic agriculture on biodiversity and abundance: A meta-analysis. *J. Appl. Ecol.* **2005**, *42*, 261–269. [\[CrossRef\]](https://doi.org/10.1111/j.1365-2664.2005.01005.x)
- <span id="page-21-29"></span>161. Barbault, R. Biodiversity dynamics: From population and community ecology approaches to a landscape ecology point of view. *Landsc. Urban Plan.* **1995**, *31*, 89–98. [\[CrossRef\]](https://doi.org/10.1016/0169-2046(94)01038-A)
- <span id="page-21-30"></span>162. Gabriel, D.; Tscharntke, T. Insect pollinated plants benefit from organic farming. *Agric. Ecosyst. Environ.* **2007**, *118*, 43–48. [\[CrossRef\]](https://doi.org/10.1016/j.agee.2006.04.005)
- <span id="page-21-31"></span>163. Rookwood, P. Landscape planning for biodiversity. *Landsc. Urban Plan.* **1995**, *31*, 379–385. [\[CrossRef\]](https://doi.org/10.1016/0169-2046(94)01064-F)
- <span id="page-21-32"></span>164. Kevan, P.G.; Baker, H.G. Insects as Flower Visitors and Pollinators. *Ann. Rev. Entomol.* **1989**, *28*, 407–453. [\[CrossRef\]](https://doi.org/10.1146/annurev.en.28.010183.002203)
- <span id="page-22-0"></span>165. Banaszak, J. Strategy for conservation of wild bees in an agricultural landscape. *Agric. Ecosyst. Environ.* **1992**, *40*, 179–192. [\[CrossRef\]](https://doi.org/10.1016/0167-8809(92)90091-O)
- <span id="page-22-1"></span>166. Zamora, J.; Verdù, J.R.; Galante, E. Species richness in Mediterranean agroecosystems: Spatial and temporal analysis for biodiversity conservation. *Biol. Conserv.* **2007**, *134*, 113–121. [\[CrossRef\]](https://doi.org/10.1016/j.biocon.2006.08.011)
- <span id="page-22-2"></span>167. Vulliamy, B.; Potts, S.G.; Willmer, P.G. The effects of cattle grazing on plant-pollinator communities in a fragmented Mediterranean landscape. *Oikos* **2006**, *114*, 529–543. [\[CrossRef\]](https://doi.org/10.1111/j.2006.0030-1299.14004.x)
- <span id="page-22-3"></span>168. Tschanz, P.; Vogel, S.; Walter, A.; Keller, T.; Albrecht, M. Nesting of ground-nesting bees in arable fields is not associated with tillage system per se, but with distance to field edge, crop cover, soil and landscape context. *J. Appl. Ecol.* **2023**, *60*, 158–169. [\[CrossRef\]](https://doi.org/10.1111/1365-2664.14317)
- <span id="page-22-4"></span>169. Atkins, E.L.; Kellum, D. Comparative morphogenic and toxicity studies on the effect of pesticides on honeybee brood. *J. Apic. Res.* **1986**, *25*, 242–255. [\[CrossRef\]](https://doi.org/10.1080/00218839.1986.11100725)
- <span id="page-22-5"></span>170. Marshall, E.J.P.; Arnold, G.M. Factors affecting field weed and field margin flora on a farm in Essex, UK. *Landsc. Urban Plan.* **1995**, *31*, 205–216. [\[CrossRef\]](https://doi.org/10.1016/0169-2046(94)01047-C)
- <span id="page-22-6"></span>171. Ma, M.; Tarmi, S.; Helenius, J. Revisiting the speces-area relationship in a semi-natural habitat: Floral richness in agricultural buffer zones in Finland. *Agric. Ecosyst. Environ.* **2002**, *89*, 137–148. [\[CrossRef\]](https://doi.org/10.1016/S0167-8809(01)00325-5)
- <span id="page-22-7"></span>172. Steffan-Dewenter, I.; Tscharntke, T. Early succession of butterfly and plant communities on set-aside fields. *Oecologia* **1997**, *109*, 294–302. [\[CrossRef\]](https://doi.org/10.1007/s004420050087)
- <span id="page-22-8"></span>173. Graves, S.D.; Shapiro, A.M. Exotic as host plants of the Calofornia butterfly fauna. *Biol. Conserv.* **2003**, *110*, 413–433. [\[CrossRef\]](https://doi.org/10.1016/S0006-3207(02)00233-1)
- <span id="page-22-9"></span>174. Saarinen, K.A. comparison of butterfly communities along field margins under traditional and intensive management in SE Finland. *Agric. Ecosyst. Environ.* **2002**, *90*, 59–65. [\[CrossRef\]](https://doi.org/10.1016/S0167-8809(01)00168-2)
- <span id="page-22-10"></span>175. Carvell, C.; Meek, W.R.; Pywell RFGouldson, D.; Nowakoski, M. Comparing the efficacy of agri-environment schemes to enhance bumble bee abundance and diversity on arable field margins. *J. Appl. Ecol.* **2007**, *44*, 29–40. [\[CrossRef\]](https://doi.org/10.1111/j.1365-2664.2006.01249.x)
- <span id="page-22-11"></span>176. Debnam, S.; Saez, A.; Aizen, M.A.; Callaway, R.M. Exotic insect pollinators and native pollination systems. *Plant Ecol.* **2021**, *222*, 1075–1088. [\[CrossRef\]](https://doi.org/10.1007/s11258-021-01162-0)
- <span id="page-22-12"></span>177. Ghisbain, G.; Gérard, M.; Wood, T.J.; Hines, H.M.; Michez, D. Expanding insect pollinators in the Anthropocene. *Biol. Rev.* **2021**, *96*, 2755–2770. [\[CrossRef\]](https://doi.org/10.1111/brv.12777) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34288353)
- <span id="page-22-13"></span>178. Bjerknes, A.L.; Totland, Ø.; Hegland, S.J.; Nielsen, A. Do alien plant invasions really affect pollination success in native plant species? *Biol. Conserv.* **2007**, *138*, 1–12. [\[CrossRef\]](https://doi.org/10.1016/j.biocon.2007.04.015)
- <span id="page-22-14"></span>179. Hanley, M.E.; Goulsonm, D. Introduced weeds pollinated by introduced bees: Cause or effect? *Weed Biol. Manag.* **2003**, *3*, 204–212. [\[CrossRef\]](https://doi.org/10.1046/j.1444-6162.2003.00108.x)
- <span id="page-22-15"></span>180. Lau, J.A.; Funk, J.L. How ecological and evolutionary theory expanded the 'ideal weed' concept. *Oecologia* **2023**, *203*, 251–266. [\[CrossRef\]](https://doi.org/10.1007/s00442-023-05397-8) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37340279)
- <span id="page-22-16"></span>181. Fijen, T.P.; Read, S.F.; Walker, M.K.; Gee, M.; Nelson, W.R.; Howlett, B.G. Different landscape features within a simplified agroecosystem support diverse pollinators and their service to crop plants. *Landsc. Ecol.* **2022**, *37*, 1787–1799. [\[CrossRef\]](https://doi.org/10.1007/s10980-022-01423-x)
- <span id="page-22-17"></span>182. Benvenuti, S. Wildflowers-pollinator-crab spider predator food-web as indicator of the agroecosystem biodiversity. *Ecol. Indic.* **2022**, *143*, 109272. [\[CrossRef\]](https://doi.org/10.1016/j.ecolind.2022.109272)

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.