

Article

Species-Specific Responses of Bloom-Forming Algae to the Ocean Warming and Acidification

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Abstract: Macroalgal biomass blooms, including those causing the green and golden tides, have been rising along Chinese coasts, resulting in considerable social impacts and economic losses. To understand the links between the ongoing climate changes (ocean warming and acidification) and algal tide formation, the effects of temperature (20 and 24 °C), pCO₂ concentration (Partial Pressure of Carbon Dioxide, 410 ppm and 1000 ppm) and their interaction on the growth of *Ulva prolifera* and *Ulva lactuca* (green tide forming species), as well as *Sargassum horneri* (golden tide forming species) were investigated. The results indicate that the concurrent rises in temperature and pCO₂ level significantly boosted the growth and nutrient uptake rates of *U. lactuca*. For *U. prolifera*, the heightened growth and photosynthetic efficiency under higher CO₂ conditions are likely due to the increased availability of inorganic carbon. In contrast, *S. horneri* exhibited negligible responsiveness to the individual and combined effects of the increased temperature and CO₂ concentration. These outcomes indicate that the progressive climate changes, characterized by ocean warming and acidification, are likely to escalate the incidence of green tides caused by *Ulva* species, whereas they are not anticipated to precipitate golden tides.

Keywords: bloom-forming macroalgae; growth; increased temperature; enriched pCO₂; physiological responses



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1. Introduction

The ongoing emission of greenhouse gases due to human activities such as burning fossil fuels and deforestation has raised the atmospheric CO₂ concentration from 280 ppm in pre-industrial times to 410 ppm at present, with the prediction that it will be more than twice the pre-industrial concentration by 2100 [1]. The increase not only enriches dissolved CO₂ in the ocean, resulting in a pH reduction (ocean acidification, OA), but also elevates the average seawater surface temperature, accelerating ocean warming [2,3]. Alterations in the physical and chemical properties of seawater, driven by climate changes, influence the calcification rates, demography, abundance, distribution and adaptability of marine organisms [4,5]. Macroalgae serve a crucial role in maintaining ocean ecosystem biodiversity and stability since they offer food, habitat, and nursery for other marine life [6–8]. Exposure to fluctuating environments typically affects their growth, photosynthetic activity, biochemical composition and even reproductive pattern [9,10].

Most algae are sensitive to the changes in CO₂ availability, as both the dissolved inorganic carbon (DIC) and pH can impact their photosynthetic performance [11]. Indeed, photosynthetic carbon (C) fixation is primarily regulated by the enzyme Ribulose-1,5-biophosphate carboxylase/oxygenase (Rubisco), which exclusively utilizes CO₂ [12] and is pH dependent [13]. The species-specific responses of marine algae to CO₂ have been extensively documented and are largely attributed to their diverse strategies for inorganic

carbon (Ci) acquisition [14]. Regarding most algae, the current CO₂ level saturates their photosynthesis because they may actively use bicarbonate (HCO₃⁻) or directly uptake CO₂ using carbon-concentration mechanisms (CCMs) [15]. Nevertheless, insufficient HCO₃⁻ usage or CO₂ acquisition would restrict the photosynthetic rate, consequently slowing down algal growth.

Some seaweeds, such as *Saccharina japonica* [7,11] and *Ulva* species [16], benefit from enriched pCO₂. The mechanism rates of these algae under the current CO₂ levels were limited by insufficient carbon acquisition but accelerated by the enriched conditions. On the other hand, algae such as *Gracilaria lemaneiformis* [17] and *Pyropia haitanensis* [14] were negatively impacted by increased CO₂ levels. The decreased pH is typically attributed to the adverse impacts because it may induce the ROS (reactive oxygen species) that damage photosystems and inhibit the activity of the enzymes that are involved in carbon assimilation [11,18]. Furthermore, other algae such as *Alaria esculenta* and *Sargassum horneri* have been reported to be unaffected by elevated pCO₂, and the neutral effect could be linked to their long-term adaptation to the acidification stress [19,20].

Temperature is another crucial factor affecting algal growth and physiology. Several primary sites of temperature sensitivity, such as PSII (in particular the D1 protein), the thylakoid membrane and the photosynthetic apparatus (the attacking sites of ROS), as well as the enzymes involved in the Calvin–Benson cycle, have been proposed [21]. For instance, an increased temperature may induce the incomplete oxidation of water at the electron donor side of PSII, resulting in an accumulation of H₂O₂, which would be then reduced by manganese to a highly oxidizing HO· through the Fenton reaction [22]. Nevertheless, previous studies have observed that elevated temperatures had no influence or positively enhanced algal photosynthesis, respiration and growth at elevated temperatures, indicating that the temperatures studied are still within the sub-optimal range for these photoautotrophs [2,14,19,20].

Ulva species and *S. horneri* have been identified as key contributors to global green and golden tide events, respectively [23]. Their remarkable tolerance to diverse environmental conditions, coupled with the rapid growth rates, has made them global problematic agents [24,25]. These macroalgal blooms pose a multifaceted threat to marine and estuarine ecosystems, leading to ecological disturbances such as hypoxia, the emission of toxic hydrogen sulfide harmful to aquatic life and humans, and the loss of species that are crucial for ecological balance and economic value [23].

Elevated temperatures increase the CO₂ diffusion coefficient and reduce its viscosity, both of which elevate the Ci availability for algae [19]. Therefore, it is imperative to examine the synergistic effects of these factors on macroalgae. Although studies have shown the separate and combined effects of CO₂ and temperature on macroalgae [2,7,14,20], the specific impacts of these factors on bloom-forming genera like *Ulva* and *Sargassum* are not fully understood. This study, accordingly, investigated the independent and interactive impacts of CO₂ and temperature on the growth, physiological traits and bio-composition of *U. prolifera*, *U. lactuca* and *S. horneri*, aiming to clarify the link between the outbreaks of green/golden tides and the progressive climate changes.

2. Results

2.1. Growth

The growth of the three bloom-forming macroalgae, including two green algae, *U. prolifera* and *U. lactuca*, and one brown alga, *S. horneri*, was markedly affected by pCO₂ ($p < 0.001$, three-way ANOVA, Table S1, Supplementary Materials). The temperature and its combination with pCO₂ had no effect on the RGR ($p > 0.05$). The RGR varied among species ($p < 0.001$) and donated the maximum effect size ($\eta^2 = 0.638$, $p < 0.001$, Table S1, Supplementary Materials) and had a significant influence with pCO₂ and temperature ($p < 0.01$). Overall, the two green algae grew faster than *S. horneri* (Figure 1). At 20 °C, *U. prolifera* showed a higher RGR (1.4 times, Figure 1a) under the enriched CO₂ level than that under ambient conditions. However, there was little significance at 24 °C. A

remarkable increase in the RGR was found in *U. lactuca* when both the temperature and CO₂ (greenhouse conditions, GH conditions) were simultaneously increased (Figure 1b). The growth of *S. horneri* was unaffected by the two environmental factors and their interaction (Figure 1c).

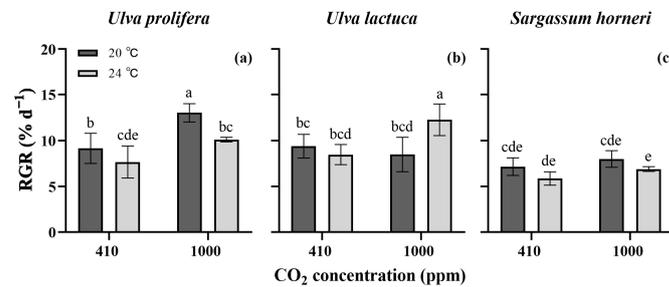


Figure 1. Relative growth rates (RGRs) of *Ulva prolifera* (a), *Ulva lactuca* (b) and *Sargassum horneri* (c) grown at different pCO₂ levels and temperatures. All the results are shown as mean value ± SD ($n = 3$). Different letters indicate significant differences ($p < 0.05$) using Scheffe's post-hoc test.

2.2. Pigment Contents

Both the Chl *a* and Car contents varied among the three algae ($p < 0.001$) and were significantly affected by the pCO₂, temperature and the interactions between species and the other two factors ($p < 0.05$, three-way ANOVA, Table S1, Supplementary Materials), whereas the interplay between pCO₂ and temperature, as well as their combination with species, had no impact on pigment contents ($p > 0.05$). The algal species exhibited the maximum effects ($\eta^2 = 0.734$ and 0.881 , respectively) on Chl *a* and Car contents. *S. horneri* had low pigment contents (around $0.24 \text{ mg Chl } a \text{ g}^{-1} \text{ FW}$ and $0.06 \text{ mg Car g}^{-1} \text{ FW}$) and no significance was observed among all environmental treatments ($p > 0.05$, Figure 2). The higher temperature decreased the pigment contents of *U. prolifera* under the enriched CO₂ conditions, in particular under the GH condition. Contrarily, the higher temperature obviously raised the pigment contents of *U. lactuca* under the two CO₂ concentrations, but the Chl *a* and Car contents showed decreases when the pCO₂ level increased. The highest pigment amounts were observed in *U. lactuca* grown at 24 °C under the ambient pCO₂ conditions, which were $0.66 \text{ mg Chl } a \text{ g}^{-1} \text{ FW}$ and $0.21 \text{ mg Car g}^{-1} \text{ FW}$.

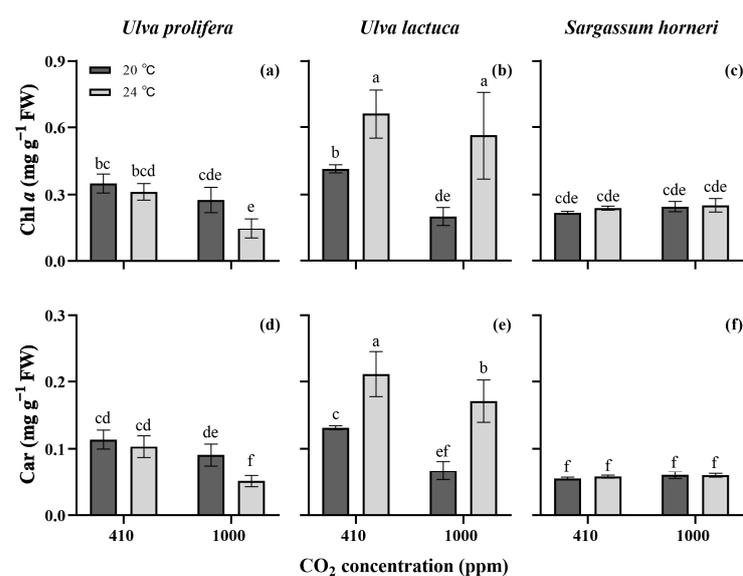


Figure 2. Pigment contents of *Ulva prolifera* (a,d), *Ulva lactuca* (b,e) and *Sargassum horneri* (c,f) grown at different pCO₂ levels and temperatures. All the results are shown as mean value ± SD ($n = 3$). Different letters indicate significant differences ($p < 0.05$) using Scheffe's post-hoc test.

2.3. Photosynthetic Rate

Consistent with the other parameters, the species had a high significant effect on the photosynthetic rate ($\eta^2 = 0.817$, $p < 0.001$, three-way ANOVA, Table S1, Supplementary Materials). The CO₂, temperature, their interactions and the interaction with species impaired the P_n significantly ($p < 0.05$). *U. prolifera* showed a higher net photosynthetic rate at 20 °C than at elevated temperatures ($p < 0.05$, Figure 3), and the maximal P_n was under the higher CO₂ level conditions and was 35.7 $\mu\text{mol O}_2 \text{ g}^{-1} \text{ FW h}^{-1}$. The enriched pCO₂ increased the P_n of *U. lactuca* at 20 °C but reduced it at 24 °C ($p < 0.05$). Under atmospheric CO₂ levels, the elevated temperature increased the P_n , whereas under the enriched CO₂ conditions it decreased the P_n . The P_n of *S. horneri* displayed no variations between the two CO₂ levels ($p > 0.05$) but decreased when the temperature was elevated ($p < 0.05$).

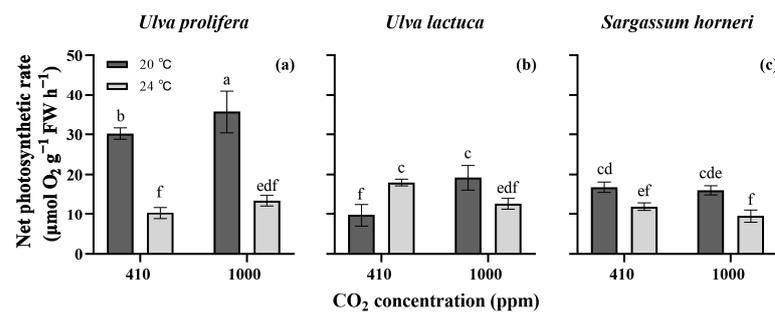


Figure 3. Net photosynthetic rates of *Ulva prolifera* (a), *Ulva lactuca* (b) and *Sargassum horneri* (c) grown at different pCO₂ levels and temperatures. All the results are shown as mean value \pm SD ($n = 3$). Different letters indicate significant differences ($p < 0.05$) using Scheffe's post-hoc test.

2.4. Nutrients Uptake Rates

The three seaweeds displayed completely different N removal behaviors ($\eta^2 = 0.945$, $p < 0.001$, three-way ANOVA, Table S1, Supplementary Materials), which were also significantly impacted by CO₂, temperature and their combinations with species ($p < 0.05$). Figure 4a–c show that the green alga *U. lactuca* absorbed nitrogen faster than *U. prolifera* and *S. horneri* ($p < 0.05$). The brown alga had the lowest N uptake rates (0.13–0.14 $\text{mg g}^{-1} \text{ FW d}^{-1}$). Specifically, there was no significant effect of CO₂ and temperature on the N uptake rates of both *U. prolifera* and *S. horneri*. Elevated pCO₂ values and temperatures accelerated the N uptake of *U. lactuca*, and the maximal rate (1.9 $\text{mg g}^{-1} \text{ FW d}^{-1}$) was observed under the GH conditions.

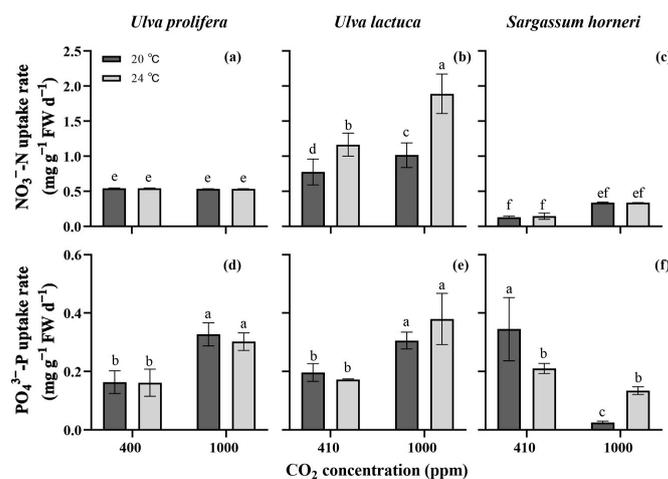


Figure 4. Nitrogen (a–c) and phosphorus uptake rates (d–f) of *Ulva prolifera*, *Ulva lactuca* and *Sargassum horneri* grown under different pCO₂ levels and temperatures. All the results are shown as mean value \pm SD ($n = 3$). Different letters indicate significant differences ($p < 0.05$) using Scheffe's post-hoc test.

Species and CO₂ level showed individual and synergistic effects on the P uptake of the three seaweeds ($p < 0.05$, three-way ANOVA, Table S1, Supplementary Materials), but the temperature and its combination with species had no significance ($p > 0.05$). In addition, pCO₂ combined with temperature, and their combination together with species revealed notable cooperative influences on P uptake ($p < 0.01$). Elevated pCO₂ levels increased the P uptake rates of green algae regardless of temperature but reduced the P uptake rate of *S. horneri* ($p < 0.05$, Figure 4d–f). Concurrently, higher temperatures were observed to decrease the P uptake rate of *S. horneri* under ambient CO₂ conditions, whereas increasing it under the enriched CO₂ conditions ($p < 0.05$, Figure 4f).

2.5. Soluble Protein Content

Figure 5 shows the soluble protein (SP) contents of *U. prolifera*, *U. lactuca* and *S. horneri*. The species and CO₂ individually and interactively affected the SP content ($p < 0.05$), whereas the temperature and its interactions with species and pCO₂ had little effect on the SP content ($p > 0.05$, three-way ANOVA, Table S1, Supplementary Materials). *U. prolifera* showed the highest SP content, followed by *U. lactuca* and then *S. horneri*. CO₂ levels and temperature exhibited no effect on the latter two algae, but the elevated pCO₂ inhibited the SP accumulation of *U. prolifera* ($p < 0.05$).

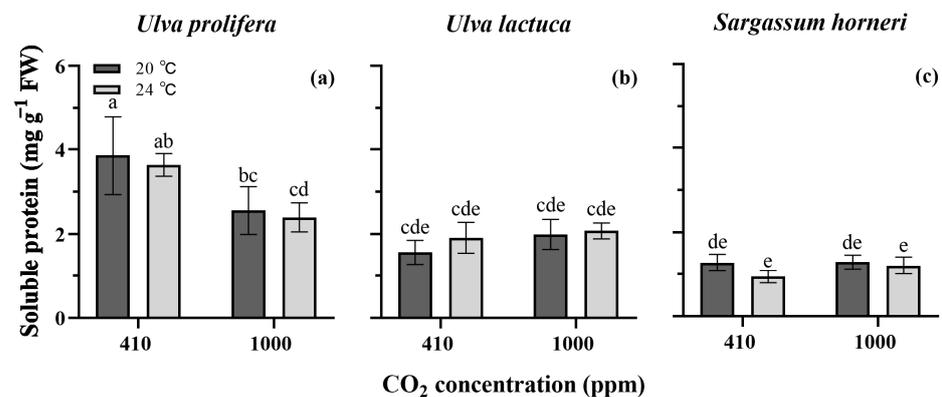


Figure 5. Soluble protein contents of *Ulva prolifera* (a), *Ulva lactuca* (b) and *Sargassum horneri* (c) grown at different pCO₂ levels and temperatures. All the results are shown as mean value \pm SD ($n = 3$). Different letters indicate significant differences ($p < 0.05$) using Scheffe's post-hoc test.

3. Discussion

The growth of the three algae species was differentially influenced (Figure 1), indicating the different responses of the bloom-forming algae to the ongoing climate changes. The diversity in responses may be due to their distinct acclimation strategies, such as the regulation of energy and carbon partitioning via photosynthesis and respiration. These strategies enable organisms to optimize their growth and survival under varying environmental conditions [19,26]. In our study, the green algae grew faster than the Phaeophyceae, suggesting a higher potential for green tide outbreaks caused by *Ulva* species compared to the golden tides caused by *S. horneri*. This could be linked to the higher content of antenna pigments in green algae (except the *U. prolifera* grown under the GH conditions, Figure 2), which may absorb more photons for biomass gains [27]. In addition, the higher nutrient uptake rates may accelerate the growth of *Ulva* species, as they are crucial for the synthesis of essential biomolecules such as proteins, enzymes, energy-transfer molecules (ATP, ADP), chlorophylls and genetic materials (RNA, DNA) [28]. Shahar et al. [29] also confirmed that the higher daily growth rate of *Ulva fasciata* was associated with its higher N utilization efficiency.

Under enriched CO₂ conditions, *U. prolifera* showed increased RGRs at both temperatures, mainly due to the increased DIC availability. Appropriately elevated CO₂ levels have been reported to enhance macroalgal photosynthetic activities, providing the energy

and carbon needed for biomolecule synthesis and biomass accumulation [7,14,16]. This study confirmed this finding, with an increased net photosynthetic rate (P_n) observed in *U. prolifera* under the enriched $p\text{CO}_2$ conditions (Figure 3a). In addition to the increased P_n , the enhanced P uptake rate may also contribute to the greater growth potential (Figure 4d), given that phosphorus facilitates the storage and exchange of energy and information in cells [30]. However, it is noteworthy that enriched $p\text{CO}_2$ declined the contents of photosynthetic pigments, despite only minor reductions in Chl *a* and Car being observed at 20 °C (Figure 2). A high $p\text{CO}_2$ level generally raises the HCO_3^- concentration, driving the CCMs with a lowered energy demand, which may result in a decrease in the biosynthesis of energy-capturing pigments. This phenomenon of “pigment economy” has been reported in some *Ulva* species by Gao et al. [31] and Wang et al. [16]. Consistent with those studies and same results of *U. prolifera* in this work, reductions in the pigment contents induced by the elevated $p\text{CO}_2$ were also observed in *U. lactuca* (Figure 2b,e).

At the higher temperature, *U. prolifera* exhibited reduced RGRs, although the GH conditions partly offset this decline. The lowered photosynthetic rate at 24 °C is likely a primary reason for the RGR reduction. Cui et al. [32] identified 20 °C as the optimal temperature for *U. prolifera*, noting that the growth rate decreased significantly at 25 °C. High temperatures can lead to the accumulation of ROS and oxidative stress, which may damage the photosynthetic apparatus. Muñoz et al. [33] suggested that the detrimental effects of elevated temperatures on *Lithothamnion crispatum* and *Sonderophycus capensis* may stem from a significant rise in ROS levels. Therefore, the decreased growth rate and P_n observed in this study could be attributed to the increased temperature exceeding the algal thermal tolerance threshold.

On the other hand, temperature had little influence on the soluble protein (SP) content of *U. prolifera*, whereas under the elevated CO_2 and GH conditions, the SP contents were significantly reduced (Figure 5a). This suggests that the CO_2 level exerted a more pronounced effect on SP accumulation. Suárez-Álvarez et al. [34] proposed that the reduction in soluble protein content at a high CO_2 level may be due to an uncoupling of carbon assimilation via photosynthesis with an increased nitrogen demand. In this work, although elevated $p\text{CO}_2$ increased the P_n , there was little effect on nitrogen uptake (Figure 4a), potentially leading to changes in nitrogen distribution among proteins and other nitrogen-containing compounds [34,35]. Additionally, the reduction in the soluble protein content under the enriched CO_2 conditions might also be associated with a reallocation of energy towards faster growth.

In the case of *U. lactuca*, an increased temperature alone showed no influence on algal growth. However, the RGR under GH conditions was notably higher, approximately 1.4 times greater than that under both the current conditions and the combination of enriched CO_2 with 20 °C (Figure 1b). This suggests that the predicted climate changes may potentially increase the outbreak frequency of green tides caused by *U. lactuca*. Under GH conditions, the increased temperature increases the diffusive coefficient of CO_2 , thereby facilitating greater CO_2 availability for biomass accumulation [19,36]. Similar increases in algal biomass under GH conditions have been documented in *Gracilariopsis lemaneiformis* [37], *Pycodry rubens* and *Saccorhiza dermatodea* [19].

U. lactuca exhibited a higher pigment content at 24 °C than at 20 °C under both $p\text{CO}_2$ conditions (Figure 2). The increased pigment accumulation might serve to mitigate the oxidative stress in the photosynthetic apparatus induced by the higher temperature [14,38]. The rises in the Chl *a* and Car contents were also closely correlated with the higher N uptake rate under enriched CO_2 conditions (Figure 4). This may be because cellular nitrogen plays important roles in the accumulation of photosynthetic pigments. Similar findings have been reported in *Kappaphycus alvarezii* by Peter et al. [25]. However, compared to at 24 °C under ambient CO_2 conditions, the elevated pigment contents under GH conditions appears to contradict the observed decline in the photosynthetic rate (Figures 2 and 3). We hypothesize that the further decreased intracellular pH under GH conditions might adversely interrupt the electron donor side of PSII centers, reducing the oxygen evolution rate (Figure 3b),

which is in line with the finding observed by Schlodder and Meyer [39]. However, to clearly elucidate the lack of correlation between pigment contents and photosynthetic performance, further studies should be carried out.

S. horneri demonstrated lower growth rates than the two *Ulva* species, and the elevated temperature and enriched pCO₂, both independently and interactively, had little impact on the growth, pigment contents, N uptake and soluble protein content, though the increased temperature slightly decreased the P_n . This may suggest that compared to the *Ulva* species, *S. horneri* is less sensitive to climate changes. The *Ulva* genus tends to bloom in the photic zone of eutrophic coasts and estuaries, whereas the pelagic *Sargassum* genus is widespread in the ocean. In recent years, however, unusual bimaerial blooms have been observed in the Yellow Sea of China [23]. Given the proximity of the sampling sites for the three algae, the possibility of green and golden tides occurring simultaneously in the maritime area of Jiangsu Province could not be ruled out (See Section 4.1). Yet, the findings revealed the green algae showed incomparable performance in growth and physiological characteristics compared to the brown alga; therefore, it appears more likely that the outbreak frequency of green tides at the studied location will be higher.

In our previous study [20], however, the increased temperature alone and its combination with the elevated pCO₂ enhanced the growth, photosynthesis and carbon assimilation of *S. horneri*. The different responses in the two assays may be linked to the sampling of *S. horneri* at different growth stages. In the previous assay, *S. horneri* was collected in March, during its rapid growth phase, whereas in the current study, the sampling was performed in June, corresponding to the maturation phase of this alga [40]. Mature *S. horneri* exhibits greater resilience to the fluctuating climate changes, suggesting that the greenhouse effect may exacerbate golden tides during the algal growth stage but may have negligible influence during its maturation stage.

On the whole, the experimental results provide evidence that the three bloom-forming algae investigated here displayed distinct responses to the ongoing climate changes in terms of the increasing atmospheric CO₂ and temperature. The enriched pCO₂ and elevated temperature synergistically raised the growth rate, pigment content and nutrient uptake rates of *U. lactuca*, and higher pCO₂ concentrations increased the growth and photosynthetic rates of *U. prolifera*. However, those factors showed little significance in *S. horneri*. We propose that the combined increase in temperature and CO₂ concentration would aggravate the outbreaks of green tides formed by *U. lactuca*, while CO₂ enrichment may be specifically associated with the blooms of *U. prolifera*, and that the factors tested may have no link to the biomass blooms of *S. horneri*.

4. Materials and Methods

4.1. Sample Collection

U. prolifera and *U. lactuca* were collected in June 2021 from Gaogong Island (34°54'31" N; 119°31'57" E), Lianyungang, Jiangsu Province, China; *S. horneri* was obtained from Qidong, Nantong (31°41'6" N; 121°25'40" E), Jiangsu Province, China. The temperatures at the sampling sites were 18–20 °C. The samples were placed in a closed tank containing ice packs and transferred to the laboratory within 2 h. Healthy thalli were selected and rinsed with sterilized seawater and, in particular, the secondary branches of *S. horneri* were cut into around 2.0 cm. The thalli were stock-cultured in three Erlenmeyer flasks (5 L) containing 5 L autoclaved seawater in the incubators (GXZ-500C, Ningbo Jiangnan instrument factory, China) and maintained at 20 °C with air-filtered aeration and an irradiance of 100 μmol photon m⁻² s⁻¹ (12:12 h light/dark cycle) for 3 days to reduce the negative effects of cutting.

4.2. Experimental Design

Under the global warming process, it is projected that temperature will rise by 4 °C and the atmospheric CO₂ concentration will reach 1000 ppm by the end of 21st century [41]. Therefore, two temperatures (20 °C and 24 °C) and two CO₂ concentrations (410 ppm and 1000 ppm) were maintained by the incubators to study the effects of temperature, CO₂

and their interaction on the growth and physiological traits of the three bloom-forming macroalgae. Herein, four treatments, including 20 °C + 410 ppm pCO₂, 20 °C + 1000 ppm pCO₂, 24 °C + 410 ppm pCO₂ and 24 °C + 1000 ppm pCO₂, were investigated. Each treatment was performed in three replicates. Uniformly growing and healthy 0.5 g thalli were introduced into round bottles that contained 500 mL autoclaved seawater (AS, around 30 µM N, ambient level; enriched with 8 µM P to avoid P restriction) at a starting biomass density of 1 g L⁻¹. The seaweeds were continuously illuminated with 100 µmol photon m⁻² s⁻¹ (12:12 h light/dark cycle), and the medium was aerated and replaced with fresh AS every three days. In total, the seaweeds were cultured for 12 days. The fresh thalli were harvested when renewing the medium to characterize the growth rate, pigment content, nutrient uptake rates, photosynthesis and respiration rates and soluble protein content.

4.3. Growth Rate

The relative growth rate (RGR) was calculated using the formula $RGR (\% d^{-1}) = 100 \times \ln(W_t/W_0)/t$, where W_t and W_0 are the fresh weight (FW, g) measured at day t and the beginning of experiment, respectively [42].

4.4. Biochemical Components

Chlorophyll *a* (Chl *a*) and Carotenoids (Cars) were extracted using approximately 0.02 g fresh thalli. The tissue was fully submerged in 5 mL methanol and then incubated at 4 °C for 24 h in darkness. The extracts were spectrophotometrically detected at 470, 652 and 665 nm using an ultraviolet absorption spectrophotometer (U-2900, HITACHI, Tokyo, Japan). The pigment contents (mg g⁻¹ FW) were estimated using the formula reported by Wellburn [43].

The soluble protein (SP) content (mg g⁻¹ FW) was quantified using Coomassie Brilliant Blue G-250 dye according to Kochert [44]. At the end of the cultivation, 0.02 g tissue was ground in a cold mortar with phosphoric acid buffer (PBS, stored at 4 °C). The homogenate was diluted to 10 mL with PBS and centrifuged at 5000 rpm for 15 min (4 °C). A total of 1 mL supernatant was mixed with 4 mL G-250 dye solution and, after 5 min, the absorbance at 595 nm was recorded using the spectrophotometer. Bovine serum albumin was adapted as the standard ($y = 0.0014x - 0.0036$, $R^2 = 0.9928$).

4.5. Photosynthetic Oxygen Evolution

A Clark-type oxygen electrode (YSI Model 5300; Yellow Springs Instrument Co., Yellow Springs, OH, USA) was adapted to quantify the photosynthetic oxygen evolution. The thalli were cut into 1 cm length pieces and re-cultured for 1 h to reduce the mechanical damage. A total of 0.02 g of small thalli were placed into a column filled with 8 mL medium. The net photosynthetic rate (P_n) was determined under the growth conditions (combined temperatures and pCO₂ levels).

4.6. Nutrient Uptake Rates

At the time of the last water change, 5 mL fresh and 5 mL algae-cultured medium were collected at day 9 and at day 12, respectively. The N and P concentrations (mg L⁻¹) in water samples were quantified using a nutrient analyzer (SEAL QuAAtro 39-SFA, Mequon, WI, USA), and the nutrient uptake rates were estimated using the formula [45]:

$$N_{\text{uptake}} (\text{mg g}^{-1} \text{FW d}^{-1}) = (N_0 - N_t) \times V/M/t \quad (1)$$

where N_{uptake} represents the N (or P) uptake rate of the algae; N_0 and N_t are the N (or P) concentrations in the fresh and algae-cultured media, respectively; V is the culture volume (here $V = 0.5$ L); M stands for biomass in the bottles; and t is the water change interval (here $t = 3$ d).

4.7. Data Analysis

All treatments were conducted in three independent biological replicates, and the data are reported as mean values with standard deviations (Mean \pm SD). The relative growth rates, pigment contents, photosynthetic rates, nutrients uptake rates and soluble protein contents between the two temperatures, two pCO₂ levels and among the three species were statistically analyzed by three-way ANOVA using IBM SPSS Statistics 26.0 software (SPSS Inc., Chicago, IL, USA). Scheffe's multiple comparisons procedure was performed after the tests of normality and variance homogeneity. Significant variations at a level of $p < 0.05$ are shown with different letters.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/plants13172433/s1>, Table S1: Analysis of three-way ANOVA showing the effects of species, pCO₂, temperature and their interactions on the relative growth rate (RGR), pigments contents, net photosynthetic rate, nitrogen and phosphorous removal rate, as well as soluble protein content of three macroalgae, *Ulva prolifera*, *Ulva lactuca* and *Sargassum horneri*.

Author Contributions: Conceptualization, H.W. and S.L.; methodology, H.W.; software, F.C.; validation, J.C., F.C. and H.L.; formal analysis, H.L. and J.C.; investigation, J.C., F.C. and H.L.; resources, J.X.; data curation, P.H.; writing—original draft preparation, S.L.; writing—review and editing, S.L. and H.W.; visualization, J.X.; supervision, H.W.; project administration, H.W.; funding acquisition, H.W. All authors have read and agreed to the published version of the manuscript.

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Data Availability Statement: The data from this study are available from the corresponding author upon reasonable request.

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References

1. IPCC The Intergovernmental Panel on Climate Change. Climate Change 2013: The Physical Science Basis. In *Summary for Policymakers*; Cambridge University Press: Cambridge, UK, 2013; pp. 24–25.
2. Liu, L.; Zou, D.H.; Jiang, H.; Chen, B.B.; Zeng, X.P. Effects of Increased CO₂ and Temperature on the Growth and Photosynthesis in the Marine Macroalga *Gracilaria lemaneiformis* from the Coastal Waters of South China. *J. Appl. Phycol.* **2018**, *30*, 1271–1280. [[CrossRef](#)]
3. Scheffer, M.; Schulz, V.; Bellerby, R.G.J.; Botros, M.; Fritsche, P.; Meyerhöfer, M.; Neill, C.; Nondal, G.; Oschlies, A.; Wohlers, J.; et al. Positive Feedback between Global Warming and Atmospheric CO₂ Concentration Inferred from Past Climate Change. *Geophys. Res. Lett.* **2006**, *33*, 229–237. [[CrossRef](#)]
4. Poloczanska, E.S.; Burrows, M.T.; Brown, C.J.; García Molinos, J.; Halpern, B.S.; Hoegh-Guldberg, O.; Kappel, C.V.; Moore, P.J.; Richardson, A.J.; Schoeman, D.S.; et al. Responses of Marine Organisms to Climate Change across Oceans. *Front. Mar. Sci.* **2016**, *3*, 62. [[CrossRef](#)]
5. Yao, C.-L.; Somero, G.N. The Impact of Ocean Warming on Marine Organisms. *Chin. Sci. Bull.* **2014**, *59*, 468–479. [[CrossRef](#)]
6. Evans, R.D.; Wilson, S.K.; Field, S.N.; Moore, J.A.Y. Importance of Macroalgal Fields as Coral Reef Fish Nursery Habitat in North-west Australia. *Mar. Biol.* **2014**, *161*, 599–607. [[CrossRef](#)]
7. Zhang, X.S.; Xu, D.; Guan, Z.; Wang, S.; Zhang, Y.; Wang, W.; Zhang, X.W.; Fan, X.; Li, F.; Ye, N. Elevated CO₂ Concentrations Promote Growth and Photosynthesis of the Brown Alga *Saccharina japonica*. *J. Appl. Phycol.* **2020**, *32*, 1949–1959. [[CrossRef](#)]
8. Fulton, C.J.; Berkström, C.; Wilson, S.K.; Abesamis, R.A.; Bradley, M.; Åkerlund, C.; Barrett, L.T.; Bucol, A.A.; Chacin, D.H.; Chong-Seng, K.M.; et al. Macroalgal Meadow Habitats Support Fish and Fisheries in Diverse Tropical Seascapes. *Fish Fish.* **2020**, *21*, 700–717. [[CrossRef](#)]

9. Revilla-Lovano, S.; Sandoval-Gil, J.M.; Zertuche-González, J.A.; Belando-Torrenetes, M.D.; Bernardeau-Esteller, J.; Rangel-Mendoza, L.K.; Ferreira-Arrieta, A.; Guzmán-Calderón, J.M.; Camacho-Ibar, V.F.; Muñoz-Salazar, R.; et al. Physiological Responses and Productivity of the Seaweed *Ulva ohnoi* (Chlorophyta) under Changing Cultivation Conditions in Pilot Large Land-Based Ponds. *Algal Res.* **2021**, *56*, 102316. [[CrossRef](#)]
10. Borburema, H.D.S.; Graiff, A.; Marinho-Soriano, E.; Karsten, U. Photosynthetic Performance, Growth, Pigment Content, and Photoprotective Compounds of the Mangrove Macroalgae *Bostrychia calliptera* and *Bostrychia montagnei* (Rhodophyta) under Light Stress. *Front. Mar. Sci.* **2022**, *9*, 989454. [[CrossRef](#)]
11. Zhang, W.; Shi, Y.; He, L.; Chen, X.; Hu, F.; Chen, Y.; Pang, Y.; Li, S.; Chu, Y. Decreased Salinity Offsets the Stimulation of Elevated pCO₂ on Photosynthesis and Synergistically Inhibits the Growth of Juvenile Sporophyte of *Saccharina japonica* (Laminariaceae, Phaeophyta). *Plants* **2022**, *11*, 2978. [[CrossRef](#)]
12. Raven, J.A.; Beardall, J. Carbon Acquisition Mechanisms of Algae: Carbon Dioxide Diffusion and Carbon Dioxide Concentrating Mechanisms. In *Advances in Photosynthesis and Respiration*; Kluwer Academic Publishers: Dordrecht, The Netherlands, 2003; pp. 225–244.
13. Laterre, R.; Pottier, M.; Remacle, C.; Boutry, M. Photosynthetic Trichomes Contain a Specific Rubisco with a Modified pH-Dependent Activity. *Plant Physiol.* **2017**, *173*, 2110–2120. [[CrossRef](#)]
14. Liu, C.; Zou, D. Do Increased Temperature and CO₂ Levels Affect the Growth, Photosynthesis, and Respiration of the Marine Macroalga *Pyropia haitanensis* (Rhodophyta)? An Experimental Study. *Hydrobiologia* **2015**, *745*, 285–296. [[CrossRef](#)]
15. Raven, J. Inorganic Carbon Acquisition by Marine Autotrophs. In *Advances in Botanical Research*; Elsevier: Amsterdam, The Netherlands, 1997; Volume 27, pp. 85–209. ISBN 978-0-12-005927-0.
16. Wang, Y.; Xu, D.; Ma, J.; Zhang, X.; Fan, X.; Zhang, Y.; Wang, W.; Sun, K.; Ye, N. Elevated CO₂ Accelerated the Bloom of Three *Ulva* Species after One Life Cycle Culture. *J. Appl. Phycol.* **2021**, *33*, 3963–3973. [[CrossRef](#)]
17. Zhou, W.; Wu, H.; Huang, J.; Wang, J.; Zhen, W.; Wang, J.; Ni, J.; Xu, J. Elevated-CO₂ and Nutrient Limitation Synergistically Reduce the Growth and Photosynthetic Performances of a Commercial Macroalga *Gracilariopsis lemaneiformis*. *Aquaculture* **2022**, *550*, 737878. [[CrossRef](#)]
18. Liu, T.; Chen, J.A.; Wang, W.; Simon, M.; Wu, F.; Hu, W.; Chen, J.B.; Zheng, H. A Combined Proteomic and Transcriptomic Analysis on Sulfur Metabolism Pathways of *Arabidopsis thaliana* under Simulated Acid Rain. *PLoS ONE* **2014**, *9*, e90120. [[CrossRef](#)]
19. Gordillo, F.J.L.; Carmona, R.; Viñegla, B.; Wiencke, C.; Jiménez, C. Effects of Simultaneous Increase in Temperature and Ocean Acidification on Biochemical Composition and Photosynthetic Performance of Common Macroalgae from Kongsfjorden (Svalbard). *Polar Biol.* **2016**, *39*, 1993–2007. [[CrossRef](#)]
20. Wu, H.; Feng, J.; Li, X.; Zhao, C.; Liu, Y.; Yu, J.; Xu, J. Effects of Increased CO₂ and Temperature on the Physiological Characteristics of the Golden Tide Blooming Macroalgae *Sargassum horneri* in the Yellow Sea, China. *Mar. Pollut. Bull.* **2019**, *146*, 639–644. [[CrossRef](#)] [[PubMed](#)]
21. Figueroa, F.; Bonomi Barufi, J.; Malta, E.; Conde-Álvarez, R.; Nitschke, U.; Arenas, F.; Mata, M.; Connan, S.; Abreu, M.; Marquardt, R.; et al. Short-Term Effects of Increasing CO₂, Nitrate and Temperature on Three Mediterranean Macroalgae: Biochemical Composition. *Aquat. Biol.* **2014**, *22*, 177–193. [[CrossRef](#)]
22. Pospíšil, P. Production of Reactive Oxygen Species by Photosystem II as a Response to Light and Temperature Stress. *Front. Plant Sci.* **2016**, *7*, 1950. [[CrossRef](#)]
23. Xiao, J.; Wang, Z.L.; Liu, D.Y.; Fu, M.Z.; Yuan, C.; Yan, T. Harmful Macroalgal Blooms (HMBs) in China's Coastal Water: Green and Golden Tides. *Harmful Algae* **2021**, *107*, 102061. [[CrossRef](#)] [[PubMed](#)]
24. Jiang, M.; Gao, L.; Huang, R.; Lin, X.; Gao, G. Differential Responses of Bloom-Forming *Ulva intestinalis* and Economically Important *Gracilariopsis lemaneiformis* to Marine Heatwaves under Changing Nitrate Conditions. *Sci. Total Environ.* **2022**, *840*, 156591. [[CrossRef](#)] [[PubMed](#)]
25. Peter, N.R.; Raja, N.R.; Rengarajan, J.; Radhakrishnan Pillai, A.; Kondusamy, A.; Saravanan, A.K.; Changaramkumarath Paran, B.; Kumar Lal, K. A Comprehensive Study on Ecological Insights of *Ulva lactuca* Seaweed Bloom in a Lagoon along the Southeast Coast of India. *Ocean Coast. Manag.* **2024**, *248*, 106964. [[CrossRef](#)]
26. Zhang, L.; Pei, H.; Chen, S.; Jiang, L.; Hou, Q.; Yang, Z.; Yu, Z. Salinity-Induced Cellular Cross-Talk in Carbon Partitioning Reveals Starch-to-Lipid Biosynthesis Switching in Low-Starch Freshwater Algae. *Bioresour. Technol.* **2018**, *250*, 449–456. [[CrossRef](#)]
27. Halsey, K.H.; Jones, B.M. Phytoplankton Strategies for Photosynthetic Energy Allocation. *Annu. Rev. Mar. Sci.* **2015**, *7*, 265–297. [[CrossRef](#)] [[PubMed](#)]
28. Kumar, A.; Bera, S. Revisiting Nitrogen Utilization in Algae: A Review on the Process of Regulation and Assimilation. *Bioresour. Technol. Rep.* **2020**, *12*, 100584. [[CrossRef](#)]
29. Shahar, B.; Shpigel, M.; Barkan, R.; Masasa, M.; Neori, A.; Chernov, H.; Salomon, E.; Kiflawi, M.; Guttman, L. Changes in Metabolism, Growth and Nutrient Uptake of *Ulva fasciata* (Chlorophyta) in Response to Nitrogen Source. *Algal Res.* **2020**, *46*, 101781. [[CrossRef](#)]
30. Solovchenko, A.E.; Ismagulova, T.T.; Lukyanov, A.A.; Vasilieva, S.G.; Konyukhov, I.V.; Pogosyan, S.I.; Lobakova, E.S.; Gorelova, O.A. Luxury Phosphorus Uptake in Microalgae. *J. Appl. Phycol.* **2019**, *31*, 2755–2770. [[CrossRef](#)]
31. Gao, G.; Beardall, J.; Wang, C.; Ren, W.; Xu, J. Ocean Acidification and Nutrient Limitation Synergistically Reduce Growth and Photosynthetic Performances of a Green Tide Alga *Ulva linza*. *Biogeosciences* **2018**, *15*, 3409–3420. [[CrossRef](#)]

32. Cui, J.; Zhang, J.; Huo, Y.; Zhou, L.; Wu, Q.; Chen, L.; Yu, K.; He, P. Adaptability of Free-Floating Green Tide Algae in the Yellow Sea to Variable Temperature and Light Intensity. *Mar. Pollut. Bull.* **2015**, *101*, 660–666. [[CrossRef](#)]
33. Muñoz, P.T.; Sáez, C.A.; Martínez-Callejas, M.B.; Flores-Molina, M.R.; Bastos, E.; Fonseca, A.; Gurgel, C.F.D.; Barufi, J.B.; Rörig, L.; Hall-Spencer, J.M.; et al. Eduardo Short-Term Interactive Effects of Increased Temperatures and Acidification on the Calcifying Macroalgae *Lithothamnion crispatum* and *Sonderophycus capensis*. *Aquat. Bot.* **2018**, *148*, 46–52. [[CrossRef](#)]
34. Suárez-Álvarez, S.; Gómez-Pinchetti, J.L.; García-Reina, G. Effects of Increased CO₂ Levels on Growth, Photosynthesis, Ammonium Uptake and Cell Composition in the Macroalga *Hypnea spinella* (Gigartinales, Rhodophyta). *J. Appl. Phycol.* **2012**, *24*, 815–823. [[CrossRef](#)]
35. Stitt, M.; Krapp, A. The Interaction between Elevated Carbon Dioxide and Nitrogen Nutrition: The Physiological and Molecular Background. *Plant Cell Environ.* **1999**, *22*, 583–621. [[CrossRef](#)]
36. Ahmadi, H.; Jamialahmadi, M.; Soulgani, B.S.; Dinarvand, N.; Sharafi, M.S. Experimental Study and Modelling on Diffusion Coefficient of CO₂ in Water. *Fluid Phase Equilibria* **2020**, *523*, 112584. [[CrossRef](#)]
37. Liu, C.; Zou, D.; Yang, Y. Comparative Physiological Behaviors of *Ulva lactuca* and *Gracilariopsis lemaneiformis* in Responses to Elevated Atmospheric CO₂ and Temperature. *Environ. Sci. Pollut. Res.* **2018**, *25*, 27493–27502. [[CrossRef](#)]
38. Sachdev, S.; Ansari, S.A.; Ansari, M.I. Photosynthetic Apparatus: Major Site of Oxidative Damage. In *Reactive Oxygen Species in Plants: The Right Balance*; Springer Nature: Singapore, 2023; pp. 75–92.
39. Schlodder, E.; Meyer, B. pH Dependence of Oxygen Evolution and Reduction Kinetics of Photooxidized Chlorophyll aII (P-680) in Photosystem II Particles from *Synechococcus* sp. *Biochim. Biophys. Acta BBA-Bioenerg.* **1987**, *890*, 23–31. [[CrossRef](#)]
40. Yu, J.; Li, J.; Wang, Q.; Liu, Y.; Gong, Q. Growth and Resource Accumulation of Drifting *Sargassum horneri* (Fucales, Phaeophyta) in Response to Temperature and Nitrogen Supply. *J. Ocean Univ. China* **2019**, *18*, 1216–1226. [[CrossRef](#)]
41. IPCC. *Climate Change, 2014: Synthesis Report*; Contribution of Working Groups I, II and III to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change; IPCC: Geneva, Switzerland, 2014; p. 151.
42. Li, S.F.; Liu, Y.; Gong, Q.L.; Gao, X.; Li, J.Y. Physiological and Ultrastructural Responses of the Brown Seaweed *Undaria pinnatifida* to Triphenyltin Chloride (TPTCL) Stress. *Mar. Pollut. Bull.* **2020**, *153*, 110978. [[CrossRef](#)]
43. Wellburn, A.R. The Spectral Determination of Chlorophylls *a* and *b*, as Well as Total Carotenoids, Using Various Solvents with Spectrophotometers of Different Resolution. *J. Plant Physiol.* **1994**, *144*, 307–313. [[CrossRef](#)]
44. Kochert, G. *Protein Determination by Dye Binding*; Cambridge University Press: London, UK, 1978.
45. Liu, H.; Wang, F.; Dong, S.; Tian, X. A Comparative Study of the Nutrient Uptake and Growth Capacities of Seaweeds *Caulerpa lentillifera* and *Gracilaria lichenoides*. *J. Appl. Phycol.* **2016**, *28*, 3083–3089. [[CrossRef](#)]

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