



# *Article* **Modeling the Effects of Temperature and Limiting Nutrients on the Competition of an Invasive Floating Plant,** *Pontederia crassipes***, with Submersed Vegetation in a Shallow Lake**

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**Abstract:** The potential for a non-native plant species to invade a new habitat depends on broadscale factors such as climate, local factors such as nutrient availability, and the biotic community of the habitat into which the plant species is introduced. We developed a spatially explicit model to assess the risk of expansion of a floating invasive aquatic plant species (FAV), the water hyacinth (*Pontederia crassipes*), an invader in the United States, beyond its present range. Our model used known data on growth rates and competition with a native submersed aquatic macrophyte (SAV). In particular, the model simulated an invasion into a habitat with a mean annual temperature different from its own growth optimum, in which we also simulated seasonal fluctuations in temperature. Twenty different nutrient concentrations and eight different temperature scenarios, with different mean annual amplitudes of seasonal temperature variation around the mean of the invaded habitat, were simulated. In each case, the ability of the water hyacinth to invade and either exclude or coexist with the native vegetation was determined. As the temperature pattern was changed from tropical towards increasingly cooler temperate levels, the competitive advantage shifted from the tropical FAV to the more temperate SAV, with a wide range in which coexistence occurred. High nutrient concentrations allowed the coexistence of FAV, even at cooler annual temperatures. But even at the highest nutrient concentrations in the model, the FAV was unlikely to persist under the current climates of latitudes in the Southeastern United States above that of Northern Alabama. This result may have some implications for where control efforts need to be concentrated.

**Keywords:** spatially explicit model; water hyacinth; Nuttall's waterweed; temperature seasonality; nutrient limitation; bifurcation diagram

# **1. Introduction**

Invasive plant species threaten native vegetation and agricultural systems around the world [\[1\]](#page-11-0) and can cause significant economic harm to native biodiversity [\[2](#page-11-1)[–4\]](#page-12-0). Success in a new environment includes the ability to escape from specialist natural enemies [\[5,](#page-12-1)[6\]](#page-12-2), wherein invasive species may grow and reproduce in an unchecked fashion. It is also believed that native species are weaker competitors than invaders in a niche, though this has not been proven [\[7,](#page-12-3)[8\]](#page-12-4). This can lead to regime shifts that can be hard to reverse due to hysteresis effects, even with control efforts [\[9\]](#page-12-5). The problems posed by invasive species will be exacerbated by climate change [\[10](#page-12-6)[,11\]](#page-12-7).

Water hyacinth ((*Pontederia crassipes*) (Mart.) Solms (Pontederiaceae), formerly *Eichhornia crassipes*) native to the Amazon Basin of South America [\[12](#page-12-8)[–14\]](#page-12-9), is considered invasive in over 50 countries in North America, Africa, Europe, Asia, and Australia [\[15](#page-12-10)[,16\]](#page-12-11) and has been called one of the ten worst weeds in the world [\[17,](#page-12-12)[18\]](#page-12-13). In Southern Florida, water hyacinth is a threat to several natural wetland habitats, including the Everglades. Water hyacinth prefers slow-moving water, full sun, warm temperature, and high nutrient concentrations, particularly regarding nitrogen. At maturity, it spreads via seeds and stolons [\[19\]](#page-12-14).



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Water hyacinth reduces oxygen exchange in waterbodies by forming thick mats over water surfaces [\[20\]](#page-12-15), excluding native vegetation such as submersed macrophytes [\[21\]](#page-12-16), and disrupting food webs [\[22\]](#page-12-17). For the manual removal of water hyacinth, chemical and biological control methods have been utilized, with varying success [\[23\]](#page-12-18). For biological control, the mottled water hyacinth weevil, *Neochetina eichhorniae* Warner (Coleoptera: Curculionidae), and the chevron water hyacinth weevil, *N. bruchii*, were released in Southern Florida in the late 1970s and managed to significantly reduce biomass and reproduction, lowering the fitness of water hyacinth overall in many habitats but only partly reducing surface coverage [\[24\]](#page-12-19). The development of effective sustainable control measures for water hyacinth is essential both economically and for environmental conservation. Large information gaps remain concerning the most effective means for control, so further empirical and modeling research is needed.

Due to the current spread of invasive species and their projected spread from the worldwide transport of biological matter and changing habitat conditions due to climate change, modeling approaches have been developed to estimate the potential spread of species beyond their native ranges [\[25–](#page-12-20)[29\]](#page-12-21). The two major types of models in this regard are mechanistic and correlative. Both are related to the idea of a species' 'fundamental niche', a hypervolume of all the resource requirements and tolerances to stresses of the species [\[30\]](#page-12-22). A species can survive and reproduce in geographic areas where sufficient resources and stress conditions are within that hypervolume, termed a 'potential niche' [\[31\]](#page-12-23). The 'realized niche' of geographic area in which a species can actually survive may be smaller due to other environmental factors such as competitors and predators [\[32\]](#page-12-24).

Mechanistic modeling, in which the biophysical and behavioral characteristics of a species are taken into account, can describe the fundamental niche of a species [\[33\]](#page-13-0) and is an active research area, but it can involve high data requirements and complex models. Therefore, there has been a heavy emphasis on correlative models, which have been given various names, such as species distribution models (SDMs), ecological niche models, and bioclimate envelopes [\[34\]](#page-13-1). For example, SDMs, using data from remote sensing, spatial analysis, and computer models such as MaxEnt (e.g., latest version 3.3.4), are widely used in conservation [\[35–](#page-13-2)[38\]](#page-13-3). These models can map ecologically suitable habitats through estimating the relationship between species presence data and the drivers through statistical algorithms under comprehensive environmental conditions [\[39\]](#page-13-4) and thus have been widely used to predict potentially suitable habitats for species. The conditions used to assess the probability of establishment usually include a number of coarse-scale indices mostly representing climatic conditions. But they can also include spatially finerscale variables, such as soil type, elevation, slope, distance to rivers, urban area fraction, etc., for terrestrial ecosystems [\[25](#page-12-20)[,40\]](#page-13-5) or nutrient concentrations in aquatic ecosystems [\[41\]](#page-13-6). Other correlative models include generalized linear models, non-parametric multiplicative regression, decision trees, boosted regression trees, and artificial neural nets [\[34\]](#page-13-1).

A further approach is that of the CLIMEX model (e.g., latest version Climex-Dymex 4.0.2), which was developed in the 1980s for predicting habitat suitability for invasive species. As described by Elith (2017) [\[34\]](#page-13-1), this model is oriented around indices relevant to population growth (temperature, moisture, radiation, substrate, and behaviors like diapause), resistance to stresses (eight stressor indices based on temperature and moisture), and a few other indices related to being able to persist, such as the length of the growing season. The environmental data largely constitute weekly climatic conditions. This model can be regarded as largely mechanistic but also uses some particular information on a species that is not simply based on physiology.

Modelers of the invasive water hyacinth have used both modeling approaches. Modeling invasion potential at the global level, Cordeiro et al. (2009) [\[42\]](#page-13-7) used MaxEnt version 3.3.3, with 19 bioclimatic variables. Kriticos and Brunei (2016) [\[11\]](#page-12-7), using CLIMEX version 4, with a set of ten CLIMEX climatic variables, also addressed potential invasion at the global level, with an emphasis on water security. Several machine learning approaches (support vector, random forest, boosted regression tree) were used by Belayhun and Mekuriaw

(2024) [\[43\]](#page-13-8), with 10 vegetation and water indices, to predict the spread of water hyacinth in Lake Tana, Ethiopia. A classification tree model was used by Zarkami et al. (2002) [\[44\]](#page-13-9) to predict water hyacinth spread in wetlands in Iran. Baker et al. used CLIMEX version 4 to develop a climatic suitability map to identify areas of high risk of water hyacinth invasion in Europe [\[45\]](#page-13-10).

A weakness of the above modeling approaches, even those using CLIMEX, is that they cannot easily take into account possible interactions of the invasive species with native competitors [\[8\]](#page-12-4) or natural enemies [\[7\]](#page-12-3) that might limit its realized niche. Accounting for such biotic interactions is important, as doing so may limit range expansion. Interactions between native and invasive species can determine the success of an invasion. Native species often have already established competitive strategies and predator–prey relationships that can either hinder or facilitate the invasion process. For instance, if an invasive plant is highly palatable to native herbivores, it might spread more rapidly [\[46\]](#page-13-11). This is best determined through mechanistic modeling [\[47](#page-13-12)[,48\]](#page-13-13) at the level of population interactions. Mechanistic models of water hyacinth interactions with a biocontrol agent, the weevil *Neochetina crassipes*, exist [\[49\]](#page-13-14), as well as a model of water hyacinth competing with a submersed aquatic macrophyte [\[50\]](#page-13-15). Here, we expand on these models to take into account several key environmental characteristics [\[51\]](#page-13-16) that influence the ability of water hyacinth to successfully invade and, in particular, either exclude or coexist with native aquatic vegetation. We simulate the potential invasion of water hyacinth in a new habitat as a function of limiting nutrients, seasonally varying temperature [\[52\]](#page-13-17), a competing submersed aquatic macrophyte, and aspects of spatial heterogeneity. We integrate and elaborate on components of existing models to achieve this.

In an earlier study, a spatially implicit model was developed to describe competition between a type of generic floating aquatic vegetation (FAV) and submersed aquatic vegetation (SAV) in a shallow waterbody [\[53\]](#page-13-18) where the FAV was limited by a nutrient (generally nitrogen) in the water column but could limit solar radiation reaching the SAV. This model was parameterized for two aquatic vegetation species, floating duckweed (*Lemna gibba*) and submersed Nuttall's waterweed (*Elodea nuttallii*) and tested experimentally. A subsequent extension of this model created to consider the effects of spatial extent and heterogeneity (spatially varying water depths) was developed by van Nes and Scheffer [\[54\]](#page-13-19) using a spatially explicit lattice model with a 50  $\times$  50 pixel grid. Van Nes and Scheffer's model [\[53,](#page-13-18)[54\]](#page-13-19) showed that spatial heterogeneity in the form of varying water depths could extend the range of values of the limiting nutrient over which the FAV and SAV could coexist.

Temperature is also an important factor for the spread of aquatic plant species [\[55](#page-13-20)[,56\]](#page-13-21), including water hyacinth, which has a tropical origin. The effect of temperature, along with nitrogen, phosphorus, and other factors, on the growth rate of water hyacinth was studied by Wilson et al. (2005) [\[57\]](#page-13-22) through data synthesis and modeling. Temperature is important in regard to the ability of water hyacinth to invade an aquatic system and compete, as temperature affects growth; in particular, frost has been shown to be a major cause of mortality of the leaves of water hyacinth. Therefore, in temperate areas, the competitive ability of water hyacinth fitness could be less than that of the native vegetation.

Xu and DeAngelis (2024) [\[50\]](#page-13-15) used a spatially explicit model that is a variation of the spatial models created by van Nes and Scheffer (2005) [\[53](#page-13-18)[,54\]](#page-13-19) and McCann (2016) [\[58\]](#page-13-23) to simulate competition between FAV and SAV, particularly competition between water hyacinth and Nuttall's waterweed. The cited study was specific to Southern Florida, where the water temperature is favorable to water hyacinth. In this current study, we included the effects of temperature, both annual mean and seasonal variation, on the growth rate of water hyacinth using the equations for temperature effects developed by Wilson et al. (2005) [\[57\]](#page-13-22) in the model. The purpose was to assess the risk of invasion of water hyacinth outside of its current range, specifically for areas with a lower mean annual temperature and larger seasonal variation, to mimic a possible spread northward. In addition to fluctuations in temperature, some aspects of spatial heterogeneity were built into the modeling. The overarching question is as follows: what conditions of the total

limiting nutrients and annual temperature variation are suitable for the invasion of water hyacinth and, subsequently, its coexistence with native SAV (using Nuttall's waterweed as a proxy in this case)?

### **2. Results**

# *2.1. Effects of Temperature and Nutrients on Invasion and Coexistence*

The model simulations showed three possible qualitative outcomes: a pure state for FAV, pure state for SAV, or a state of coexistence of the two species. Both the limiting nutrient *N* and temperature had an effect; high values of *N* and high values of mean annual temperature favored the FAV, while SAV was favored at the other ends of the spectrum of those values, and coexistence occurred for a range of intermediate values (the eight temperature scenarios of Table [1\)](#page-3-0). The results for temperature scenario 3 (see Section [5\)](#page-7-0), with a mean temperature of  $\theta_{mean} = 24 \degree C$  and a seasonal amplitude of  $\theta_{amplitude} = 7.5 \degree C$ , were close to the results obtained in [\[53\]](#page-13-18). In the cited authors' spatially implicit model, coexistence for 24◦C to 25 ◦C occurred between about nutrient *N* = 0.55 and *N* = 1.35, while for our temperature scenario 3, coexistence occurs across the discrete nutrient values from  $N = 0.7$  to  $N = 1.2$ . In this scenario, the two species were approximately equal in terms of growth rates, a situation similar to that shown in Figure 1 (derived from [\[53\]](#page-13-18)). As in [\[53\]](#page-13-18), in our model scenario 3, for larger values of *N*, only FAV persisted, while for smaller *N* values, only SAV persisted. In Table [1,](#page-3-0) the mean temperature, *θmean*, decreases and the temperature fluctuation amplitude, *θamplitude*, increases as the temperature scenario number increases to the right in the table from a tropical to a more temperate climate.

<span id="page-3-0"></span>**Table 1.** Results regarding the invasion potential of FAV and possible coexistence with SAV.



F = only invader FAV persists; S = only native SAV persists; C = coexistence. <sup>†</sup> mg L<sup>-1</sup>.

The possibility of FAV excluding SAV disappeared by temperature scenario 4, with a mean temperature of  $\theta_{mean} = 22 \degree C$  and a seasonal amplitude of  $\theta_{amplitude} = 11 \degree C$ . However, coexistence was possible until scenario 8, with a mean temperature of *θmean* = 16 ◦C and a seasonal amplitude of *θamplitude* = 19 ◦C. This shows the significant effect of temperature on the growth rates of the two species and, consequently, on the competitive outcome.

# 2.2. Bifurcation Diagrams

*2.2. Bifurcation Diagrams* 

Table [1](#page-3-0) shows only the three qualitative states of competition between FAV and SAV. The quantitative values of biomasses are given in the bifurcation diagrams in Figure [1.](#page-4-0) The In the quantum vertices of *clondises* are given in the stratedient diagrams in Figure 1. The lines represent averages over both the whole spatial lattice as well as over the period of time after which the system reached a final state. Here, temperature scenarios 1 through time after which the system reached a final state. Here, temperature scenarios 1 through 4 are shown. The mean temperature decreases and seasonal fluctuations increase when moving from panels (a) to (d). As the mean temperature decreases, the range of coexistence (where the red and blue lines overlap) shifts to the right (i.e., higher nutrient levels), and the final mean value of the FAV biomass substantially decreases. In these plots, the solid lines represent the biomasses of FAV (red) and SAV (blue) for simulations in which seasonal temperature fluctuations are included, while the dashed lines represent simulations where only the mean annual temperature was included.

<span id="page-4-0"></span>

**Figure 1.** Bifurcation diagrams showing the biomasses (g dW m<sup>−</sup>2) of FAV (red lines) and SAV (blue **Figure 1.** Bifurcation diagrams showing the biomasses (g dW m−<sup>2</sup> ) of FAV (red lines) and SAV (blue lines) as a function of total nutrient concentration (mg  $L^{-1}$ ). (a) temperature scenario 1 ( $\theta_{mean}$  = 27 °C with a seasonal amplitude of  $\theta_{amplitude} = 4 °C$ ), (b) temperature scenario 2 ( $\theta_{mean} = 25.75 °C$  with a seasonal amplitude of  $\theta_{amplitude} = 6^{\circ}C$ , (c) temperature scenario 3 ( $\theta_{mean} = 24^{\circ}C$  with a seasonal amplitude of *<sup>θ</sup>amplitude* = 7.5 ◦C), and (**d**) temperature scenario 4 (*θmean* = 22 ◦C with a seasonal amplitude of  $\theta_{amplitude} = 11 \degree C$ ). The dashed lines represent biomasses at only the mean annual temperatures, without fluctuations.

# 2.3. Temporal Variability of the biomasses and time. The biomasses are in Figure 2 for two temperature 2 for two The bifurcation diagrams show the bifurcation diagrams show the biomasses averaged over both space and time.

*2.3. Temporal Variability* 

<span id="page-5-0"></span>The bifurcation diagrams show the biomasses averaged over both space and time. There is temporal variability of the biomasses, as shown in Figure 2 for two temperature scenarios in which coexistence occurred, namely, temperature scenarios 3 and 6. The cooler part of the season favored SAV, while the warmer favored FAV; thus, seasonal fluctuations occurred for the two species that were 180 degrees out of phase.



Figure 2. Variation in biomasses (g dW  $m^{-2}$ ) of SAV (blue lines) and FAV (red lines) over time. (Left) Temperature scenario 3 ( $\theta_{mean} = 24$  °C with seasonal amplitude of  $\theta_{amplitude} = 7.5$  °C), with  $N = 1.2$ ; (Right) temperature scenario 6 ( $\theta_{mean}$  = 18.5 °C with seasonal amplitude of  $\theta_{amplitude}$  = 16 °C), with  $N = 1$ .

#### *2.4. Spatial Heterogeneity 2.4. Spatial Heterogeneity* Spatial variability was interested in the model in the model in the model in the model of FAV are proposed of F

<span id="page-5-1"></span>randomly deposited in time in random spatial pixels in one of the scenarios. When conditions are favorable, the FAV propagules grow at the expense of SAV, as shown  $\frac{1}{2}$  in Figure 3. Spatial variability was included in the model, as vegetative propagules of FAV are in Figure [3.](#page-5-1)



**Figure 3.** The spatial distribution of vegetation biomasses is shown for SAV (**left**) and FAV (**right**) sometime after two propagules of FAV had been deposited in random pixels and started to spread.

# **3. Discussion**

Predictive models of potential invasions are needed to lay the groundwork for management actions [\[1\]](#page-11-0). The array of modeling approaches that have been applied to predicting invasions has been reviewed [\[1\]](#page-11-0), and these approaches range from SDMs and statistical models [\[25](#page-12-20)[,29\]](#page-12-21) to population dynamics models [\[28\]](#page-12-25) and cellular automata models [\[59\]](#page-13-24). As early as almost three decades ago, [\[60\]](#page-13-25) developed a spatially explicit individual-based model for invasive plant spread, noting that such models can incorporate details of spatial heterogeneity and individual variability.

Our approach presented herein is spatially explicit, although it is not individual-based but follows biomasses in a  $50 \times 50$  array of grid cells. The model allowed the incorporation of several key elements related to the question of whether an introduced species can successfully invade. It combined a climatic factor, temperature, with a factor that was more variable at a local scale, that is, the nutrient concentration in a particular waterbody. The model was oriented toward competition between an invader (FAV) and a native resident species (SAV), as competition with native species is one of the possible components of resistance to invasion that a native community can offer [\[61\]](#page-13-26). Because temporal variability is one of the environmental conditions that can affect population interactions  $[62]$ , it was included through the simulation of seasonal variations in temperature (Figure [2\)](#page-5-0). Finally, spatial heterogeneity and stochasticity were included by allowing propagules of the invasive species to be deposited at random times and places in the model grid that represents a shallow waterbody (Figure [3\)](#page-5-1).

The results of the simulations showed that when the competitive abilities of the competing FAV and SAV were similar, a bifurcation diagram similar to that shown in Figure 2 (corresponding to the material from [\[53\]](#page-13-18)) was the result; that is, the nutrient concentration determining which species was dominant or whether coexistence can occur depends on nutrient concentration in the same way. As the simulated temperature scenario became more temperate and thus further from the optimal temperature for the invader, the bifurcation diagram became distorted in favor of the native species, although coexistence was possible over almost the whole range of temperature scenarios if the nutrient level was high enough. However, coexistence virtually stopped in the temperature scenario corresponding to the seasonal temperature pattern of temperature scenario 8, close to the temperature patterns of Birmingham, Alabama. Although we did not make extensive comparisons with empirical data, it is important to note that the empirical data shown in Figure 2 (obtained by Kriticos and Brunel (2016) [\[11\]](#page-12-7)) show the range of permanent occupations of water hyacinth in the Southeastern United States has an upper limit in Northern Alabama.

An important result shown in the bifurcation diagrams is that the seasonal temperature fluctuations can have a large effect on the mean biomasses of the examined species (Figure [1\)](#page-4-0). This is especially true for the more temperate scenarios (Figure [1c](#page-4-0),d), in which the seasonal fluctuations were relatively large. The fluctuations clearly promoted higher FAV biomass and lower SAV biomass, though they do not appear to affect the qualitative feature of whether invasion is possible.

The results of this study cannot in themselves be used to predict how far into regions with temperate climates water hyacinth might spread. This study is intended to simply predict the likelihood of water hyacinth invading a shallow lake in the presence of a particular submersed macrophyte that has a comparable growth rate but at a lower optimum temperature. Water hyacinth is known to occur transiently in areas of the Eastern United States with mean annual temperatures that are cooler than the temperature scenarios in our model. Our model only shows that temperatures lower than the optimum growth temperature for water hyacinth will slow its growth and decrease its competitive ability against better-adapted submersed vegetation. The range of nutrient levels, *N*, extended only to  $N = 2$  mg  $\bar{L}^{-1}$ , so it did not include more eutrophic conditions. This study also suffers from the limitation that while the temperature dependance of the growth of the FAV, water hyacinth, is based on data, that of the native, more temperate species is simply

assumed, though the other parameters for the species are similar to those of Nuttall's waterweed. Therefore, the model study should be regarded as a theoretical exploration of how the combination of factors included in the model might affect water hyacinth's ability to invade cooler habitats.

Our modeling approach consists of the use of a mechanistic population competition model and so is different from approaches employing the many correlation-type models or the CLIMEX model that have been in standard use over the last two decades. However, mechanistic approaches are frequently used. Higgins and Richardson (1996) [\[60\]](#page-13-25) provided a review of the types of models of invasive organisms, including simple-demographic, spatial–phenomenological, or spatial–mechanistic models, including reaction diffusion models. Such mechanistic approaches will continue to be important in studying the details of the invasive process and how to control it. Spatially explicit individual-based models have been used to simulate interactions of the invasive *Melaleuca quinquenervia* tree in Southern Florida [\[63\]](#page-14-1). These models projected the outcome of the invader competing against five native species, slash pine, pond cypress, dahoon holly, sweet bay, and loblolly bay, along with the effects of a biocontrol agent on the invader.

The effects of temperature on the spread of water hyacinth in the Southeastern United States have also been studied using simpler models. For a site in Louisiana, Nesslage et al. (2016) [\[13\]](#page-12-26) used a logistic model with a temperature-dependent seasonal mortality parameter and included a biological control in the model. This model projected a decline in the growth rate and survival of water hyacinth of 84% between 1976 and 2013 resulting from the combination of low winter temperatures and biocontrol measures. This shows that simple modeling approaches can be effective. Our model is more complex and thus requires more assumptions, which are rough estimates but are able to describe details involving the combination of temperature, nutrients, and competition, which may be useful for management.

# **4. Conclusions**

We developed a spatially explicit model to explore the potential of an FAV, the water hyacinth, to spread to more-temperate areas. This model includes the effects of temperature and nutrients on the competition between water hyacinth and a submersed macrophyte. Although the ability of water hyacinth to invade decreases as the temperature scenario becomes less tropical and more temperate, for high levels of a given nutrient, invasion is possible over a wide range of temperatures. As the temperature pattern changed from tropical to cooler, more temperate levels, the competitive advantage shifted from the tropical FAV to the more temperate SAV, with a wide range across which coexistence occurred, which depended on the nutrient concentration. But even at the highest nutrient concentrations in the model, the FAV was unlikely to persist at current climates of latitudes in the Southeastern United State above that of Northern Alabama. This result may have some implications for where control efforts need to be concentrated. However, as the model used here was based on assumptions concerning the SAV that was competing with the FAV, the results should be seen as providing rough projections that should be tested empirically. Further work will involve surveying relevant empirical work for such tests.

#### <span id="page-7-0"></span>**5. Materials and Methods**

Although the FAV in our model, *P. crassipes*, can generally be limited by either nitrogen or phosphorus, experiments conducted at the U.S. Department of Agriculture's Invasive Plant Research Laboratory in Davie, Florida, show that *P. crassipes* growth responds strongly to nitrogen, which we assume is the limiting factor in the model. We follow Scheffer et al. (2003) [\[53\]](#page-13-18) in assuming that SAV is light-limited. Figure [4](#page-8-0) shows a schematic of the relationships. The inset represents the effect of temperature on the two species.

<span id="page-8-0"></span>

(nitrogen), but the FAV can shade the submerged plant. The inset represents the effect of temperature on bout plant species. **Figure 4.** Competition between FAV and SAV. The SAV is a better competitor for the limiting nutrient on both plant species.

# 5.1. Model Developed by Scheffer et al. (2003)

*5.1. Model Developed by Scheffer et al. (2003)*  using the equations Competition between two aquatic species, one SAV and one FAV, was modeled by [\[53\]](#page-13-18)

$$
\frac{dS}{dt} = r_s * S * \frac{n}{n + h_S} * \frac{1}{1 + a_S S + bF + W} - l_S * S,
$$
(1)

$$
\frac{dF}{dt} = r_F * F * \frac{n}{n + h_F} * \frac{1}{1 + a_F * F} - l_F * F,
$$
\n(2)

where *S* and *F* are variables of the dry weight biomasses (g dW m<sup>-2</sup>) for SAV and FAV, respectively, and *n* is the nutrient concentration in the water column (mg L<sup>-1</sup>). The parameters  $r_S$  and  $r_F$  are the maximum growth rates, and  $l_S$  and  $l_F$  are the loss rates, including respiration and mortality, while *h<sub>S</sub>* and *h<sub>F</sub>* are the half-saturation values for the particular to  $\frac{1}{2}$ . The particular to  $\frac{1}{2}$  and  $\frac{1}{2}$  and  $\frac{1}{2}$  are the interconstitution of the particu effects of SAV and FAV, and *b* is the shading effect of FAV on SAV. Parameter *W* is the effect of the surrounding water's light absorption on submersed plant growth. nutrient uptake of SAV and FAV. Parameters  $a<sub>S</sub>$  and  $a<sub>F</sub>$  are the intraspecific competitive

Scheffer et al. (2003) [53] defined the total limiting nutrient concentration in the system as *N,* which includes this nutrient both in vegetation and as a solute in the water surrounding water-light absorption on submersed plant growth absorption on submersed plant growth. The column of  $\sim$ 

$$
N = n + nq_S S + nq_F F, \tag{3}
$$

where  $q_s$  and  $q_f$  are coefficients of the effect of submerged and floating plants on the nitrogen concentration in the water column; that is, they represent the fractions of this nutrient therefore, changes as vegetation biomasses changes: tied up in the vegetation per unit dry weight biomass. The concentration, *n*, in the water,

$$
n = \frac{N}{1 + q_s S + q_F F} \tag{4}
$$

In their model, [\[53\]](#page-13-18) sought to examine the impact of varying *N* on the competition of SAV and FAV, so *N* was varied over a range of values. We used the equations of the model developed by [\[10\]](#page-12-6), changing the growth of FAV from that of duckweed given

in [\[53\]](#page-13-18) ( $r_F$  = 0.5) to  $r_F$  = 0.1, which is consistent with many of the studies of water hyacinth surveyed in Wilson et al.'s work (2005) [\[57\]](#page-13-22). We changed the shading effect of the FAV on SAV from  $b = 0.01$ , which was appropriate for duckweed in [\[53\]](#page-13-18), to  $b = 0.04$ , which reflects the higher density of *P. crassipes* vegetation. The SAV was again Nuttall's waterweed.

#### *5.2. McCann's Cellular Automata (2016)*

The model developed by [\[53\]](#page-13-18) is spatially implicit. McCann (2016) [\[55\]](#page-13-20) further developed this model into a spatial CA model composed of a grid of spatial pixels that could form two-dimensional areas of different shapes. We used a spatially explicit model, as it allowed us to initiate the development of two species in a variety of ways spatially, which could have made a difference in the outcomes yielded by the model. By dispersing propagules of each species in a random fashion across the spatial grid, the simulations provided a better chance of ensuring the competition did not depend too heavily on the starting biomasses, as might be the case in a spatially implicit model. It was assumed that SAV and FAV could spread among the pixels and that the limiting nutrient in the water column could diffuse. The edges of the spatial grid were reflective so that vegetation and the limiting nutrient did not spread out from the gridded system. We adapted the two-dimensional spatial model developed by [\[55\]](#page-13-20) for the special case of a 50  $\times$  50-pixel grid, with each pixel being  $1 \text{ m} \times 1 \text{ m}$ . This scale of resolution is the same as that employed by [\[55\]](#page-13-20) and about the spatial scale of a few full-grown water hyacinth plants. Our model is not strictly a CA model, as continuous levels of biomasses of both species could occupy individual spatial cells.

#### *5.3. Temperature Effects on Growth*

The growth rates  $r_F$  of the two species were assumed to depend on temperature,  $\theta$ , in the form of a triangular function, *gF(θ)*, defined by a minimum threshold for growth, *θmin,F*, an optimal growth temperature, *θopt,F*, and a maximum temperature, *θmax,F* [\[57\]](#page-13-22).

#### *5.4. Formatting of Mathematical Components*

$$
g_F(\theta) = \begin{cases} 0 & \theta \leq \theta_{min,F} \\ \frac{\theta - \theta_{min,F}}{\theta_{opt,F} - \theta_{min,F}} \theta_{min,F} < \theta < \theta_{opt,F} \\ \frac{\theta_{max,F} - \theta}{\theta_{max,F} - \theta_{opt,F}} & \theta_{opt,F} < \theta. \end{cases} \tag{5}
$$

The function  $g(\theta)$  multiplies the  $r_F$  of the water hyacinth, so that as  $\theta$  varies, the effective growth rate varies with it. Temperate was simulated as a mean annual temperature and an amplitude of fluctuations:

$$
\theta = \theta_{mean} + \theta_{amplitude} \sin\left(\frac{2\pi t}{365}\right),\tag{6}
$$

where the effects of a variety of values of *θmean* and *θamplitude* were used to test the ability of water hyacinth to invade and possibly exclude the native species. The effective growth rate of the FAV, water hyacinth, was thus

$$
r_{F,effective} = g_F(\theta)r_F. \tag{7}
$$

We made a similar assumption of a triangular temperature dependence,  $g_S(\theta)$ , on the SAV. We did not know the temperature dependence of Nuttall's waterweed, but we assumed there was an influence of a set of parameters *θmin,S*, *θopt,S*, and *θmax,S* that describe a more sub-tropical to temperate species, such that the triangular function is similar in shape to that of the water hyacinth but shifted towards lower temperatures (Figure [5\)](#page-10-0).

<span id="page-10-0"></span>



#### The purpose of this study was to estimate the risk of water hyacinth invading regions *5.5. Seasonal Temperatures*

*5.5. Seasonal Temperatures* 

The purpose of this study was to estimate the risk of water hyacinth invading regions of climates ranging from sub-tropical, similar to Miami, Florida, to warm temperate, simwith climates outside its tropical to sub-tropical range. Therefore, we simulated a series of climates ranging from sub-tropical, similar to Miami, Florida, to warm temperate, similar to Birmingham, Alabama (Figure [6\)](#page-10-1). The climates differ both in terms of mean annual temperature and the amplitudes of seasonal temperatures, which are larger in the more temperate regions, especially those with cooler winter temperatures.  $\frac{1}{1}$  of  $\frac{1}{2}$ , which et al. (2005)  $\frac{1}{1}$ 

<span id="page-10-2"></span>**Table 2.** Values of parameters changed in the simulations. *θmean* and *θamplitude* values are ordered from tropical to warm temperate climates.



<span id="page-10-1"></span>

**Figure 6.** Eight temperature scenarios used in the simulations. The mean temperatures and ampli-**Figure 6.** Eight temperature scenarios used in the simulations. The mean temperatures and ampli- $\frac{1}{2}$  The extreme upper are given in Table 2 The extreme upper and lower curves were estimated based on graphs were estimate tudes are given in Table [2.](#page-10-2) The extreme upper and lower curves were estimated based on graphs based on a Google search of 'Climate in Miami, Florida' and 'Climate of Birmingham, Alabama'. The curve were fit besed on averages of meathly highe and lowe. The curves The curves were fit based on averages of monthly highs and lows. The curves between the extremes were found to exhibit an arbitrary series of values of means and the amplitudes of the temperatures between those.

#### *5.6. Parameterization of the Model*

The parameters of our model generally correspond to those used by Scheffer et al. (2003) [\[53\]](#page-13-18) and McCann (2016) [\[55\]](#page-13-20), though the growth rate was modeled for water hyacinth—including regarding temperature effects obtained using data from Wilson et al.'s work (2005) [\[57\]](#page-13-22)—rather than duckweed (*Lemna gibba*) as modeled in those papers to determine the spatial dynamics of FAV (duckweed) and SAV (Nuttall's waterweed). The parameter values are listed in Table [3.](#page-11-2)

<span id="page-11-2"></span>



#### *5.7. Simulations*

Simulations were performed to approximate the way that invasion might occur. As both species can propagate vegetatively, small vegetative propagules, 0.1 g dW of each species, were scattered randomly both temporally and spatially. The frequency of the invasive FAV was low, slightly less than two vegetative propagules per year over the whole lattice. Propagules of the native species were input randomly at a much higher frequency.

Twenty values of *N* were used in the simulations, ranging from 0.1 to 2.0 mg  $L^{-1}$ , identical to the range of nutrient values used by Scheffer et al. (2003) [\[53\]](#page-13-18), along with eight seasonal temperature scenarios (shown in Figure [6\)](#page-10-1). The annual means and associated seasonal fluctuations of the eight temperature scenarios are arranged in order from tropical to warm temperatures in Table [2.](#page-10-2) Simulations were run for a period corresponding to over twenty years in order for the system to reach a final state.

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# **References**

- <span id="page-11-0"></span>1. Rai, P.K. Paradigm of plant invasion: Multifaceted review on sustainable management. *Environ. Monit. Assess.* **2015**, *187*, 1–30. [\[CrossRef\]](https://doi.org/10.1007/s10661-015-4934-3) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26581605)
- <span id="page-11-1"></span>2. Cox, G.W. *Alien Species in North America and Hawaii*; Island Press: Washington, DC, USA, 1999.
- 3. Mack, R.N.; Simberloff, D.; Mark Lonsdale, W.; Evans, H.; Clout, M.; Bazzaz, F.A. Biotic invasions: Causes, epidemiology, global consequences, and control. *Ecol. Appl.* **2000**, *10*, 689–710. [\[CrossRef\]](https://doi.org/10.1890/1051-0761(2000)010[0689:BICEGC]2.0.CO;2)
- <span id="page-12-0"></span>4. Pyšek, P.; Richardson, D.M. Invasive species, environmental change and management, and health. *Annu. Rev. Environ. Resour.* **2010**, *35*, 25–55. [\[CrossRef\]](https://doi.org/10.1146/annurev-environ-033009-095548)
- <span id="page-12-1"></span>5. Lawton, J.H.; Brown, K.C. The population and community ecology of invading insects. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **1986**, *314*, 607–617. [\[CrossRef\]](https://doi.org/10.1098/rstb.1986.0076)
- <span id="page-12-2"></span>6. Wolfe, L.M. Why alien invaders succeed: Support for the escape-from-enemy hypothesis. *Am. Nat.* **2002**, *160*, 705–711. [\[CrossRef\]](https://doi.org/10.1086/343872)
- <span id="page-12-3"></span>7. Vila, M.; Weiner, J. Are invasive plant species better competitors than native plant species?–evidence from pair-wise experiments. *Oikos* **2004**, *105*, 229–238. [\[CrossRef\]](https://doi.org/10.1111/j.0030-1299.2004.12682.x)
- <span id="page-12-4"></span>8. Perkins, L.B.; Hatfield, G. Competition, legacy, and priority and the success of three invasive species. *Biol. Invasions* **2014**, *16*, 2543–2550. [\[CrossRef\]](https://doi.org/10.1007/s10530-014-0684-3)
- <span id="page-12-5"></span>9. Reynolds, S.A.; Aldridge, D.C. Embracing the allelopathic potential of invasive aquatic plants to manipulate freshwater ecosystems. *Front. Environ. Sci.* **2021**, *8*, e551803. [\[CrossRef\]](https://doi.org/10.3389/fenvs.2020.551803)
- <span id="page-12-6"></span>10. Thuiller, W. Patterns and uncertainties of species' range shifts under climate change. *Glob. Change Biol.* **2004**, *10*, 2020–2027. [\[CrossRef\]](https://doi.org/10.1111/j.1365-2486.2004.00859.x)
- <span id="page-12-7"></span>11. Kriticos, D.J.; Brunel, S. Assessing and managing the current and future pest risk from water hyacinth, (*Eichhornia crassipes*), an invasive aquatic plant threatening the environment and water security. *PLoS ONE* **2016**, *11*, e0120054. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0120054)
- <span id="page-12-8"></span>12. Cerveira Junior, W.R.; de Carvalho, L.B. Control of water hyacinth: A short review. *Commun. Plant Sci.* **2019**, *9*, 129–132. [\[CrossRef\]](https://doi.org/10.26814/cps2019021)
- <span id="page-12-26"></span>13. Nesslage, G.M.; Wainger, L.A.; Harms, N.E.; Cofrancesco, A.F. Quantifying the population response of invasive water hyacinth, Eichhornia crassipes, to biological control and winter weather in Louisiana, USA. *Biol. Invasions* **2016**, *18*, 2107–2115. [\[CrossRef\]](https://doi.org/10.1007/s10530-016-1155-9)
- <span id="page-12-9"></span>14. Julien, M.H. Biological control of water hyacinth with arthropods: A review to 2000. In *Aciar Proceedings; Proceedings of the Second Meeting of the Global Working Group for the Biological and Integrated Control of Water Hyacinth, Beijing, China, 9–12 October 2000*; Julien, M.H., Hill, M.P., Center, T.D., Ding, J., Eds.; Australian Centre for International Agricultural Research: Canberra, ACT, Australia, 2001; pp. 8–20.
- <span id="page-12-10"></span>15. Villamagna, A.M.; Murphy, B.R. Ecological and socio-economic impacts of invasive water hyacinth (*Eichhornia crassipes*): A review. *Freshw. Biol.* **2010**, *55*, 282–298. [\[CrossRef\]](https://doi.org/10.1111/j.1365-2427.2009.02294.x)
- <span id="page-12-11"></span>16. Lu, J.; Wu, J.; Fu, Z.; Zhu, L. Water hyacinth in China: A sustainability science-based management framework. *Environ. Manag.* **2007**, *40*, 823–830. [\[CrossRef\]](https://doi.org/10.1007/s00267-007-9003-4) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17768654)
- <span id="page-12-12"></span>17. Aboul-Enein, A.M.; Al-Abd, A.M.; Shalaby, E.; Abul-Ela, F.; Nasr-Allah, A.A.; Mahmoud, A.M.; El-Shemy, H.A. *Eichhornia crassipes* (Mart) solms: From water parasite to potential medicinal remedy. *Plant Signal Behav.* **2011**, *6*, 834–836. [\[CrossRef\]](https://doi.org/10.4161/psb.6.6.15166)
- <span id="page-12-13"></span>18. Milićević, D.B. Modeling water hyacinth growth dynamics. *Arch. Biol. Sci.* 2023, 75, 165-185. [\[CrossRef\]](https://doi.org/10.2298/ABS230222014M)
- <span id="page-12-14"></span>19. Gupta, A.K.; Yadav, D. Biological control of water hyacinth. *ECR* **2020**, *3*, 37–39. [\[CrossRef\]](https://doi.org/10.26480/ecr.01.2020.37.39)
- <span id="page-12-15"></span>20. Uka, U.N.; Chukwuka, K.S.; Daddy, F. Effect of water hyacinth (Eichhornia crassippes) infestation on zooplankton populations in Awba Reservoir, Ibadan, south-west Nigeria. 2007. Available online: <http://hdl.handle.net/1834/37727> (accessed on 15 September 2024).
- <span id="page-12-16"></span>21. Brendonck, L.; Maes, J.; Rommens, W.; Dekeza, N.; Nhiwatiwa, T.; Barson, M.; Callebaut, V.; Phiri, C.; Moreau, K.; Gratwicke, B.; et al. The impact of water hyacinth (*Eichhornia crassipes*) in a eutrophic subtropical impoundment (Lake Chivero, Zimbabwe). II. Species diversity. *Arch. Hydrobiol.* **2003**, *158*, 389–405. [\[CrossRef\]](https://doi.org/10.1127/0003-9136/2003/0158-0389)
- <span id="page-12-17"></span>22. Coetzee, J.A.; Jones, R.W.; Hill, M.P. Water hyacinth, *Eichhornia crassipes* (Pontederiaceae), reduces benthic macroinvertebrate diversity in a protected subtropical lake in South Africa. Biodiversity and conservation. *Biodivers. Conserv.* **2014**, *23*, 1319–1330. [\[CrossRef\]](https://doi.org/10.1007/s10531-014-0667-9)
- <span id="page-12-18"></span>23. Tipping, P.W.; Gettys, L.A.; Minteer, C.R.; Foley, J.R.; Sardes, S.N. Herbivory by biological control agents improves herbicidal control of waterhyacinth (*Eichhornia crassipes*). *Invasive Plant Sci. Manag.* **2017**, *10*, 271–276. [\[CrossRef\]](https://doi.org/10.1017/inp.2017.30)
- <span id="page-12-19"></span>24. Tipping, P.W.; Martin, M.R.; Pokorny, E.N.; Nimmo, K.R.; Fitzgerald, D.L.; Dray Jr, F.A.; Center, T.D. Current levels of suppression of waterhyacinth in Florida USA by classical biological control agents. *Biol. Control* **2014**, *71*, 65–69. [\[CrossRef\]](https://doi.org/10.1016/j.biocontrol.2014.01.008)
- <span id="page-12-20"></span>25. Bazzichetto, M.; Malavasi, M.; Bartak, V.; Acosta, A.T.R.; Rocchini, D.; Carranza, M.L. Plant invasion risk: A quest for invasive species distribution modelling in managing protected areas. *Ecol. Indic.* **2018**, *95*, 311–319. [\[CrossRef\]](https://doi.org/10.1016/j.ecolind.2018.07.046)
- 26. Kariyawasam, C.S.; Kumar, L.; Ratnayake, S.S. Invasive plant species establishment and range dynamics in Sri Lanka under climate change. *Entropy* **2019**, *21*, 571. [\[CrossRef\]](https://doi.org/10.3390/e21060571) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33267285)
- 27. Pěknicová, J.; Berchová-Bímová, K. Application of species distribution models for protected areas threatened by invasive plants. *J. Nat. Conserv.* **2016**, *34*, 1–7. [\[CrossRef\]](https://doi.org/10.1016/j.jnc.2016.08.004)
- <span id="page-12-25"></span>28. Caplat, P.; Coutts, S.; Buckley, Y.M. Modeling population dynamics, landscape structure, and management decisions for controlling the spread of invasive plants. *Ann. N. Y. Acad. Sci.* **2012**, *1249*, 72–83. [\[CrossRef\]](https://doi.org/10.1111/j.1749-6632.2011.06313.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22239680)
- <span id="page-12-21"></span>29. Srivastava, V.; Lafond, V.; Griess, V.C. Species distribution models (SDM): Applications, benefits and challenges in invasive species management. *CABI Rev.* **2019**, *14*, 1–13. [\[CrossRef\]](https://doi.org/10.1079/PAVSNNR201914020)
- <span id="page-12-22"></span>30. Hutchinson, G.E. *An Introduction to Population Ecology*; Yale Univ Press: New Haven, CT, USA, 1978.
- <span id="page-12-23"></span>31. Jackson, S.T.; Overpeck, J.T. Responses of plant populations and communities to environmental changes of the late Quaternary. *Paleobiology* **2000**, *26*, 194–220. [\[CrossRef\]](https://doi.org/10.1017/S0094837300026932)
- <span id="page-12-24"></span>32. Elith, J.; Phillips, S.J.; Hastie, T.; Dudík, M.; Chee, Y.E.; Yates, C.J. A statistical explanation of MaxEnt for ecologists. *Divers Distrib.* **2011**, *17*, 43–57. [\[CrossRef\]](https://doi.org/10.1111/j.1472-4642.2010.00725.x)
- <span id="page-13-0"></span>33. Kearney, M.R.; Porter, W.P. NicheMapR–an R package for biophysical modelling: The ectotherm and dynamic energy budget models. *Ecography* **2020**, *43*, 85–96. [\[CrossRef\]](https://doi.org/10.1111/ecog.04680)
- <span id="page-13-1"></span>34. Elith, J. Predicting distributions of invasive species. *Invasive Species Risk Assess. Manag.* **2017**, *10*, 93–129. [\[CrossRef\]](https://doi.org/10.1017/9781139019606.006)
- <span id="page-13-2"></span>35. Elith, J.; Leathwick, J.R. Species distribution models: Ecological explanation and prediction across space and time. *Ann. Rev. Ecol. Evol. Syst.* **2009**, *40*, 677–697. [\[CrossRef\]](https://doi.org/10.1146/annurev.ecolsys.110308.120159)
- 36. Guisan, A.; Thuiller, W. Predicting species distribution: Offering more than simple habitat models. *Ecol. Lett.* **2005**, *8*, 993–1009. [\[CrossRef\]](https://doi.org/10.1111/j.1461-0248.2005.00792.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34517687)
- 37. Measey, G.J.; Rdder, D.; Green, S.L.; Kobayashi, R.; Lillo, F.; Lobos, G.; Rebelo, R.; Thirion, J.M. Ongoing invasions of the African clawed frog, Xenopus laevis: A global review. *Biol. Invasions* **2012**, *14*, 2255–2270. [\[CrossRef\]](https://doi.org/10.1007/s10530-012-0227-8)
- <span id="page-13-3"></span>38. Yuan, Y.; Tang, X.; Liu, M.; Liu, X.; Tao, J. Species Distribution Models of the Spartina alterniflora Loisel in Its Origin and Invasive Country Reveal an Ecological Niche Shift. *Front. Plant Sci.* **2021**, *12*, 738769. [\[CrossRef\]](https://doi.org/10.3389/fpls.2021.738769) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34712259)
- <span id="page-13-4"></span>39. Austin, M. Species distribution models and ecological theory: A critical assessment and some possible new approaches. *Ecol. Model.* **2007**, *200*, 1–19. [\[CrossRef\]](https://doi.org/10.1016/j.ecolmodel.2006.07.005)
- <span id="page-13-5"></span>40. Augusto, L.; Achat, D.L.; Jonard, M.; Vidal, D.; Ringeval, B. Soil parent material—A major driver of plant nutrient limitations in terrestrial ecosystems. *Glob. Change Biol.* **2017**, *23*, 3808–3824. [\[CrossRef\]](https://doi.org/10.1111/gcb.13691)
- <span id="page-13-6"></span>41. Marchetto, A.; Padedda, B.M.; Mariani, M.A.; Luglie, A.; Sechi, N. A numerical index for evaluating phytoplankton response to changes in nutrient levels in deep mediterranean reservoirs. *J. Limnol.* **2009**, *68*, 106. [\[CrossRef\]](https://doi.org/10.4081/jlimnol.2009.106)
- <span id="page-13-7"></span>42. Cordeiro, P.F.; Goulart, F.F.; Macedo, D.R.; Campos, M.D.C.S.; Castro, S.R. Modeling of the potential distribution of Eichhornia crassipes on a global scale: Risks and threats to water ecosystems. *Rev. Ambient. Água* **2020**, *15*, e2421. [\[CrossRef\]](https://doi.org/10.4136/ambi-agua.2421)
- <span id="page-13-8"></span>43. Belayhun, M.; Mekuriaw, A. 2024. Modeling water hyacinth (*Eichhornia crassipes*) distribution in Lake Tana, Ethiopia, using machine learning. *RSASE* **2024**, *36*, 101273. [\[CrossRef\]](https://doi.org/10.1016/j.rsase.2024.101273)
- <span id="page-13-9"></span>44. Zarkami, R.; Esfandi, J.; Sadeghi, R. Modelling occurrence of invasive water hyacinth (Eichhornia crassipes) in Wetlands. *Wetlands* **2021**, *41*, 8. [\[CrossRef\]](https://doi.org/10.1007/s13157-021-01405-w)
- <span id="page-13-10"></span>45. Baker, R.H.; Benninga, J.; Bremmer, J.; Brunel, S.; Dupin, M.; Eyre, D.; Ilieva, Z.; Jarošík, V.; Kehlenbeck, H.; Kriticos, D.J.; et al. A decision-support scheme for mapping endangered areas in pest risk analysis. *EPPO Bull.* **2012**, *42*, 65–73. [\[CrossRef\]](https://doi.org/10.1111/j.1365-2338.2012.02545.x)
- <span id="page-13-11"></span>46. Morrison, W.E.; Hay, M.E. Herbivore preference for native vs. exotic plants: Generalist herbivores from multiple continents prefer exotic plants that are evolutionarily naïve. *PLoS ONE* **2011**, *6*, e17227. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0017227) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21394202)
- <span id="page-13-12"></span>47. Strasser, C.A.; Lewis, M.A.; DiBacco, C. A mechanistic model for understanding invasions: Using the environment as a predictor of population success. *Divers. Distrib.* **2011**, *17*, 1210–1224. [\[CrossRef\]](https://doi.org/10.1111/j.1472-4642.2011.00791.x)
- <span id="page-13-13"></span>48. Young, P.C.; Chotai, A.; Beven, K.J. Data-based mechanistic modelling and the simplification of environmental systems. *Environ. Model. Find. Simplicity Complex.* **2004**, *371*, 388.
- <span id="page-13-14"></span>49. Van Schalkwyk, H. The Development of a Spatio-Temporal Model for Water Hyacinth, *Eichhornia crassipes (Martius) Solms-Laubach (Pontederiaceae),* Biological Control Strategies. Doctoral Dissertation, Stellenbosch University, Stellenbosch, South Africa, 2016. Available online: <https://core.ac.uk/reader/188225750> (accessed on 15 September 2024).
- <span id="page-13-15"></span>50. Xu, L.; Goode, A.B.; Tipping, P.W.; Smith, M.C.; Gettys, L.A.; Knowles, B.K.; Pokorny, E.; Salinas, L.; DeAngelis, D.L. Less is more: Less herbicide does more when biological control is present in Pontederia crassipes. *Ecol. Model.* **2024**, *487*, 110566. [\[CrossRef\]](https://doi.org/10.1016/j.ecolmodel.2023.110566)
- <span id="page-13-16"></span>51. Lee, A.M.; Sæther, B.E.; Engen, S. Spatial covariation of competing species in a fluctuating environment. *Ecology* **2020**, *101*, e02901. [\[CrossRef\]](https://doi.org/10.1002/ecy.2901)
- <span id="page-13-17"></span>52. Gardner, A.S.; Maclean, I.M.; Gaston, K.J. Climatic predictors of species distributions neglect biophysiologically meaningful variables. *Divers. Distrib.* **2019**, *25*, 1318–1333. [\[CrossRef\]](https://doi.org/10.1111/ddi.12939)
- <span id="page-13-18"></span>53. Scheffer, M.; Szabo, S.; Gragnani, A.; van Nes E., H.; Rinaldi, S.; Kautsky, N.; Norberg, J.; Roijackers, R.M.; Franken, R.J. Floating plant dominance as a stable state. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 4040–4045. [\[CrossRef\]](https://doi.org/10.1073/pnas.0737918100)
- <span id="page-13-19"></span>54. van Nes, E.H.; Scheffer, M. Implications of spatial heterogeneity for catastrophic regime shifts in ecosystems. *Ecology* **2005**, *86*, 1797–1807. [\[CrossRef\]](https://doi.org/10.1890/04-0550)
- <span id="page-13-20"></span>55. McCann, M.J. Evidence of alternative states in freshwater lakes: A spatially-explicit model of submerged and floating plants. *Ecol. Model.* **2016**, *337*, 298–309. [\[CrossRef\]](https://doi.org/10.1016/j.ecolmodel.2016.07.006)
- <span id="page-13-21"></span>56. Dijkstra, J.A.; Westerman, E.L.; Harris, L.G. Elevated seasonal temperatures eliminate thermal barriers of reproduction of a dominant invasive species: A community state change for northern communities? *Divers. Distrib.* **2017**, *23*, 1182–1192. [\[CrossRef\]](https://doi.org/10.1111/ddi.12604)
- <span id="page-13-22"></span>57. Wilson, J.R.; Holst, N.; Rees, M. Determinants and patterns of population growth in water hyacinth. *Aquat. Bot.* **2005**, *81*, 51–67. [\[CrossRef\]](https://doi.org/10.1016/j.aquabot.2004.11.002)
- <span id="page-13-23"></span>58. McCann, M.J. Response diversity of free-floating plants to nutrient stoichiometry and temperature: Growth and resting body formation. *PeerJ* **2016**, *4*, e1781. [\[CrossRef\]](https://doi.org/10.7717/peerj.1781) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26989619)
- <span id="page-13-24"></span>59. Savage, D.; Renton, M. Requirements, design and implementation of a general model of biological invasion. *Ecol. Model.* **2014**, *272*, 394–409. [\[CrossRef\]](https://doi.org/10.1016/j.ecolmodel.2013.10.001)
- <span id="page-13-25"></span>60. Higgins, S.I.; Richardson, D.M.; Cowling, R.M. Modeling invasive plant spread: The role of plant-environment interactions and model structure. *Ecology* **1996**, *77*, 2043–2054. [\[CrossRef\]](https://doi.org/10.2307/2265699)
- <span id="page-13-26"></span>61. Levine, J.M.; Adler, P.B.; Yelenik, S.G. A meta-analysis of biotic resistance to exotic plant invasions. *Ecol. Lett.* **2004**, *7*, 975–989. [\[CrossRef\]](https://doi.org/10.1111/j.1461-0248.2004.00657.x)
- <span id="page-14-0"></span>62. Chesson, P. Mechanisms of maintenance of species diversity. *Ann. Rev. Ecol. Syst.* **2000**, *31*, 343–366. [\[CrossRef\]](https://doi.org/10.1146/annurev.ecolsys.31.1.343)
- <span id="page-14-1"></span>63. Zhang, B.; DeAngelis, D.L.; Rayamajhi, M.B.; Botkin, D. Modeling the long-term effects of introduced herbivores on the spread of an invasive tree. *Landsc. Ecol.* **2017**, *32*, 1147–1161. [\[CrossRef\]](https://doi.org/10.1007/s10980-017-0519-6)

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