

Table S1. Primer sequences used for quantitative RT-PCR.

Gene name	Gene ID	Forward primer (5' ==> 3')	Reverse primer (5' ==> 3')	References
<i>ABF1</i>	AT1G49720	TGCAGAAGAAACAGGCTGAA	GACCGGTAAGGGTCTTCTCA	Yoshida et al., 2015
<i>ABF3</i>	AT4G34000	CGTTCTCAACCTGCAACACA	TCATAGGATGGTTATGAATTCCAAG	Yoshida et al., 2015
<i>AREB1</i>	AT1G45249	GAGAGAAGGCAAAGGAGAATGA	CTTCAAGCTCCACGGTGTAAG	Yoshida et al., 2015
<i>AREB2</i>	AT3G19290	GCAACCTGGTGTGGTATCC	TGTTCTAGGCAACGTCAACGA	Yoshida et al., 2015
<i>DREB2A</i>	AT5G05410	CAGTGTGCCAACGGTTCAT	AAACGGAGGTATTCCGTAGTTGAG	Sakuma et al., 2006
<i>DREB2B</i>	AT3G11020	CAGGCCGGACAGTTGCA	CCCAATACTGCTGCTCAAACCTCTA	Sakuma et al., 2006
<i>RD29A</i>	AT5G52310	TGGATCTGAAGAACGAATCTGATATC	GGTCTTCCCTTCGCCAGAA	Sakuma et al., 2006
<i>RD29B</i>	AT5G52300	GCAAGCAGAAGAACCAATCA	CTTTGGATGCTCCCTTCTCA	Yoshida et al., 2015
<i>RAB18</i>	AT1G43890	TCGGTCGTGTATTGTGCTTTTT	CCAGATGCTCATTACACACTCATG	Yoshida et al., 2015
<i>SOS1</i>	AT2G01980	TGGAGGAAGCGACCGATTC	CGATAACGAGAAGAGCGACAG	Cheng et al., 2016
<i>HKT1</i>	AT4G10310	TCTGGTCTCATCTGGCTCTAATC	CACCGTCACTCCAAGAAGAACAC	Cheng et al., 2016
<i>HRE1</i>	AT1G72360	GGCCTCTGCCTTATCCCTCTGT	GCGTAAACCCGTCTCAGTGAGTG	Licausi et al., 2010
<i>HRE2</i>	AT2G47520	GAAGCGTAAACCCGTCTCAGT	TTTGCTCGGGCACGAATCT	Licausi et al., 2010
<i>RAP2.2</i>	AT3G14230	AATCCGTGGCAGAAAGC	GGATACGACGCTAGGGTCTTCT	Papdi et al., 2015
<i>RAP2.3</i>	AT3G16770	CGTCATCGCCACGATCAA	TGAGAGACAACGCAGACCTTCTT	Papdi et al., 2015
<i>RAP2.12</i>	AT1G53910	CGCTGCGGAAGGTTCAATT	CAGCCCATTTTCCCAAGGA	Paul et al., 2016
<i>ADH1</i>	AT1G77120	TATTCGATGCAAAGCTGCTGTG	CGAACTTCGTGTTTCTGCGGT	Paul et al., 2016
<i>PDF1.2</i>	AT5G44420	CACCCTTATCTTCGCTGCTCTT	GCCGGTGCGTCAAAG	Van der Does et al., 2013
<i>ACO2</i>	AT1G62380	CCTCCTCAACCACTCTATTGTC	CATCTTCATAAACATAAACACCCACAC	Dong et al., 2015
<i>ERF1</i>	AT3G23240	ATTCTTCTCATCCTCTTCTTCT	CGAATCTCTTATCTCCGCCG	Mao et al., 2016
<i>BAS1</i>	AT2G26710	CCAAGGACCATGTCGTTAAGC	CCTGAAGTATAGCAAGATTCTGACC	Schneider et al., 2012
<i>PAE8</i>	AT4G19420	GGGCATCATTACAGGAGAT	CAGCAGAACAGCCAGACAAA	Xie et al., 2015
<i>XTH33</i>	AT1G10550	ATACCTCATTTGGAACCTCCACC	CACTTAACTCCACGTAGCAACG	Dong et al., 2014
<i>Saur_AC1</i>	AT4G38850	CAGAAGAAGAAGAGATATGTGGTG	GTATTGTAAAGCCGCCATTG	Zhu et al., 2013
<i>18S RNA</i>	AT2G01010	AAACGGCTACCACATCCAAG	CCTCCAATGGATCCTCGTTA	Yoshida et al., 2015
<i>UBQ10</i>	AT4G05320	AGATCCAGGACAAGGAAGTATTC	CGCAGGACCAAGTGAAGAGTAG	Lin et al., 2013

References

- Dong, J., Tang, D., Gao, Z., Yu, R., Li, K., He, H., et al. (2014) Arabidopsis DE-ETIOLATED1 represses photomorphogenesis by positively regulating phytochrome-interacting factors in the dark. *Plant Cell* 26: 3630-3645.
- Dong, Z., Yu, Y., Li, S., Wang, J., Tang, S. and Huang, R. (2016) Abscisic Acid Antagonizes Ethylene Production through the ABI4-Mediated Transcriptional Repression of ACS4 and ACS8 in Arabidopsis. *Mol. Plant* 9: 126-135.
- Li, P.C., Huang, J.G., Yu, S.W., Li, Y.Y., Sun, P., Wu, C.A., et al. (2016) Arabidopsis YL1/BPG2 Is Involved in Seedling Shoot Response to Salt Stress through ABI4. *Sci. Rep.* 6: 30163.
- Licausi, F., van Dongen, J.T., Giuntoli, B., Novi, G., Santaniello, A., Geigenberger, P., et al. (2010) HRE1 and HRE2, two hypoxia-inducible ethylene response factors, affect anaerobic responses in Arabidopsis thaliana. *Plant J.* 62: 302-315.
- Lin, W., Lu, D., Gao, X., Jiang, S., Ma, X., Wang, Z., et al. (2013) Inverse modulation of plant immune and brassinosteroid signaling pathways by the receptor-like cytoplasmic kinase BIK1. *Proc. Natl. Acad. Sci. USA* 110: 12114-12119.
- Mao, J.L., Miao, Z.Q., Wang, Z., Yu, L.H., Cai, X.T. and Xiang, C.B. (2016) Arabidopsis ERF1 Mediates Cross-Talk between Ethylene and Auxin Biosynthesis during Primary Root Elongation by Regulating ASA1 Expression. *PLoS Genet.* 12: e1005760.

- Papdi, C., Perez-Salamo, I., Joseph, M.P., Giuntoli, B., Bogre, L., Koncz, C., et al. (2015) The low oxygen, oxidative and osmotic stress responses synergistically act through the ethylene response factor VII genes RAP2.12, RAP2.2 and RAP2.3. *Plant J.* 82: 772-784.
- Paul, M.V., Iyer, S., Amerhauser, C., Lehmann, M., van Dongen, J.T. and Geigenberger, P. (2016) Oxygen Sensing via the Ethylene Response Transcription Factor RAP2.12 Affects Plant Metabolism and Performance under Both Normoxia and Hypoxia. *Plant Physiol.* 172: 141-153.
- Sakuma, Y., Maruyama, K., Osakabe, Y., Qin, F., Seki, M., Shinozaki, K., et al. (2006) Functional analysis of an Arabidopsis transcription factor, DREB2A, involved in drought-responsive gene expression. *Plant Cell* 18: 1292-1309.
- Schneider, K., Breuer, C., Kawamura, A., Jikumaru, Y., Hanada, A., Fujioka, S., et al. (2012) Arabidopsis PIZZA has the capacity to acylate brassinosteroids. *PLoS One* 7: e46805.
- Van der Does, D., Leon-Reyes, A., Koornneef, A., Van Verk, M.C., Rodenburg, N., Pauwels, L., et al. (2013) Salicylic acid suppresses jasmonic acid signaling downstream of SCFCO11-JAZ by targeting GCC promoter motifs via transcription factor ORA59. *Plant Cell* 25: 744-761.
- Xie, Y., Straub, D., Eguen, T., Brandt, R., Stahl, M., Martinez-Garcia, J.F., et al. (2015) Meta-Analysis of Arabidopsis KANADI1 Direct Target Genes Identifies a Basic Growth-Promoting Module Acting Upstream of Hormonal Signaling Pathways. *Plant Physiol.* 169: 1240-1253.
- Yoshida, T., Fujita, Y., Maruyama, K., Mogami, J., Todaka, D., Shinozaki, K., et al. (2015) Four Arabidopsis AREB/ABF transcription factors function predominantly in gene expression downstream of SnRK2 kinases in abscisic acid signalling in response to osmotic stress. *Plant Cell Environ.* 38: 35-49.
- Zhu, W., Wang, H., Fujioka, S., Zhou, T., Tian, H., Tian, W., et al. (2013) Homeostasis of brassinosteroids regulated by DRL1, a putative acyltransferase in Arabidopsis. *Mol. Plant* 6: 546-558.