

Supplementary Material

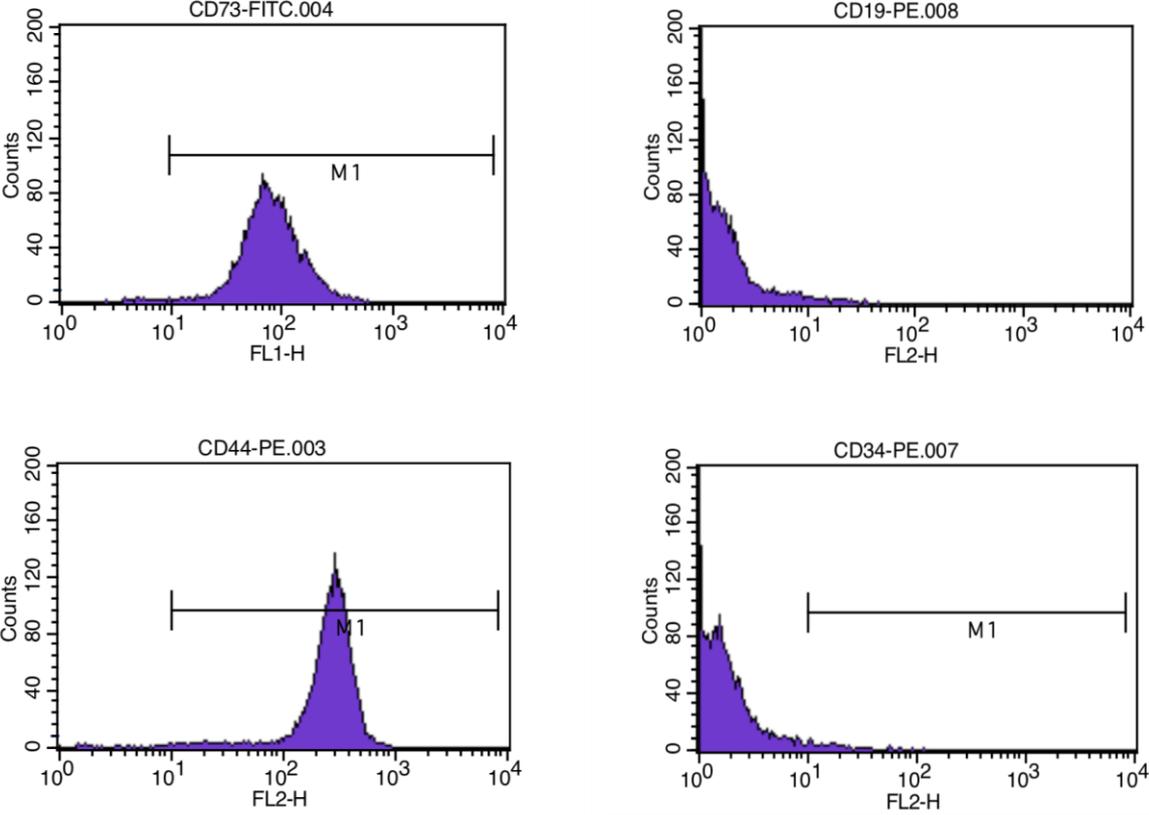


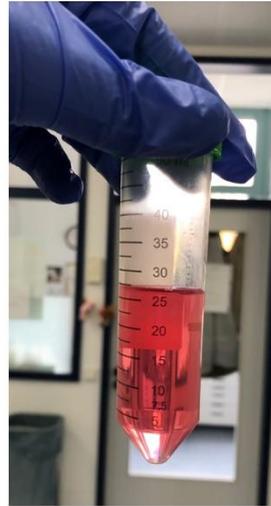
Figure S1. Flow cytometric analysis of hBMSC with specific antibodies against positive marker (CD44, CD73) and negative marker (CD19, CD34).

A **Conditioned medium (CM)**



Hypoxia Normoxia

Osteogenic medium (OM) under hypoxia (no cells) **Osteogenic medium (OM) under normoxia (no cells)**



B

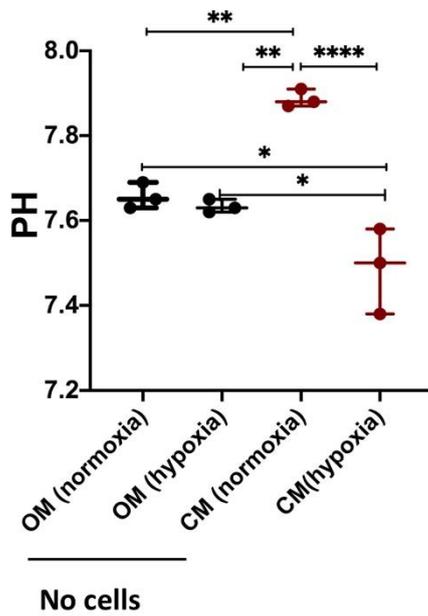


Figure S2. Conditioned medium from hypo-osteo EVs and norm-osteo EVs.

(A) Conditioned medium from hBMSCs undergoing osteogenic differentiation at 33-35 days under hypoxia and normoxia (CM). Osteogenic medium (OM, without subjecting to cell culture) was kept for two days under hypoxic and normoxic conditions.

(B) PH determination of the two conditioned media (CM) and the two osteogenic media (OM); n=3.

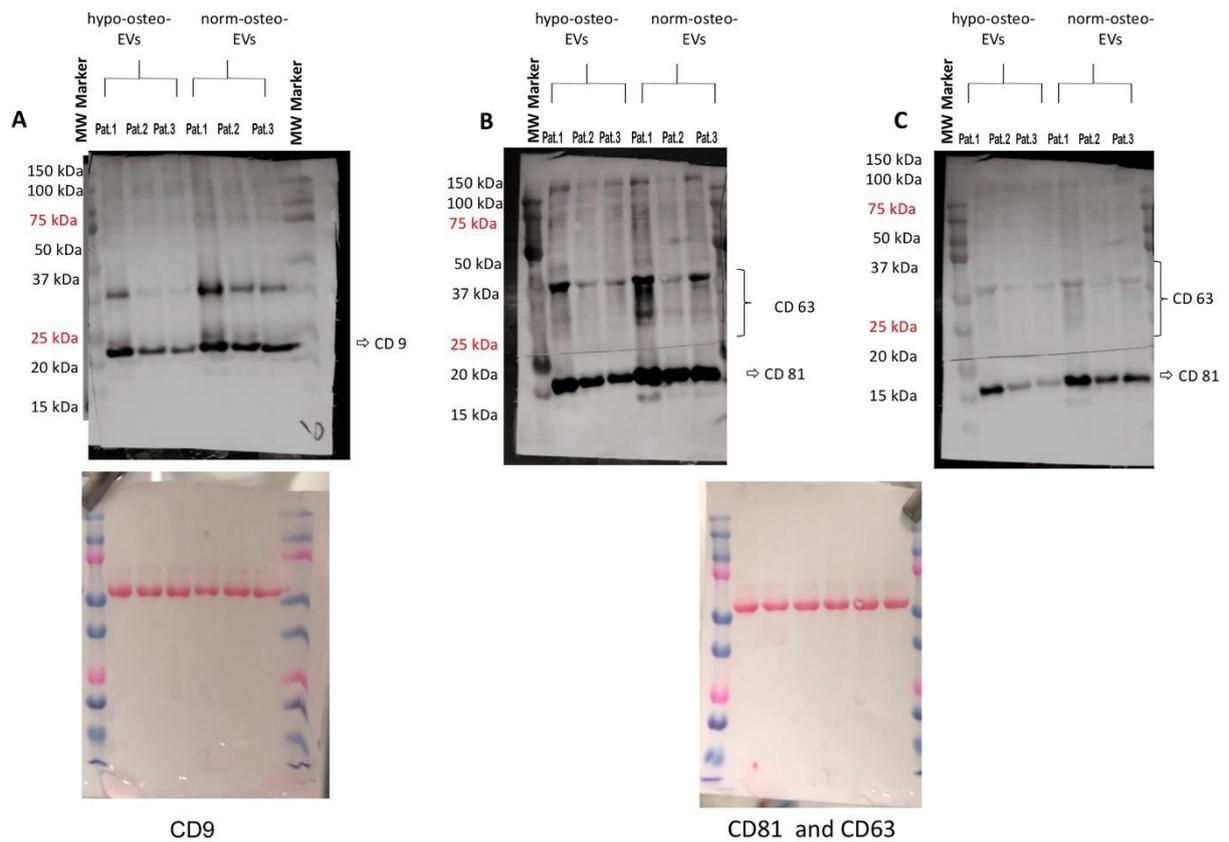


Figure S3. The uncropped western blot membranes of figure 2G and respective Ponceau Red stained loading control images below; n=3.

(A, C) Protein bands were detected using ECL detection reagents. (B) Protein bands were detected using SuperSignal West Femto Maximum Sensitivity Substrate.

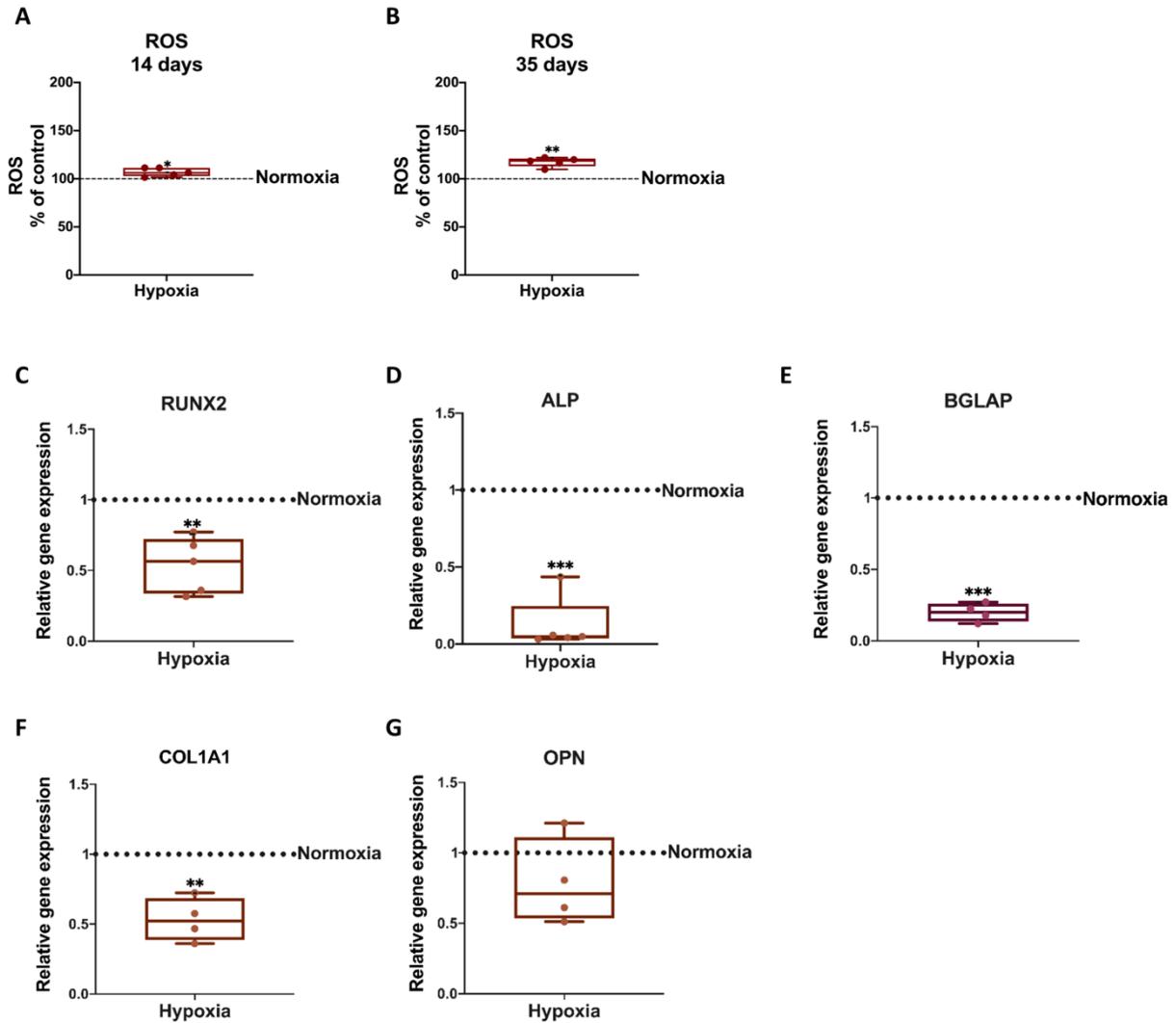


Figure S4. Evaluation of reactive oxygen species (ROS) secreted, and osteogenic marker gene expression in osteogenic differentiated hBMSCs under hypoxia and normoxia.

(A) Reactive oxygen species (ROS) secreted from hBMSCs after **14** days of osteogenic differentiation under hypoxia and normoxia; n=5.

(B) Reactive oxygen species (ROS) secreted from hBMSCs after **35** days of osteogenic differentiation under hypoxia or normoxia; n=5.

(C-G) Gene expression level of the osteogenic marker genes (OPN, BGLAP, ALP, RUNX2, and COL1A1) after 14 days of osteogenic differentiation of hBMSCs under hypoxia and normoxia; n=4-5.

Results were calculated as percentage to the control group (osteogenic differentiation of hBMSCs under normoxia, shown by the dotted line); *p < 0.05; **p < 0.01; ***p < 0.001;

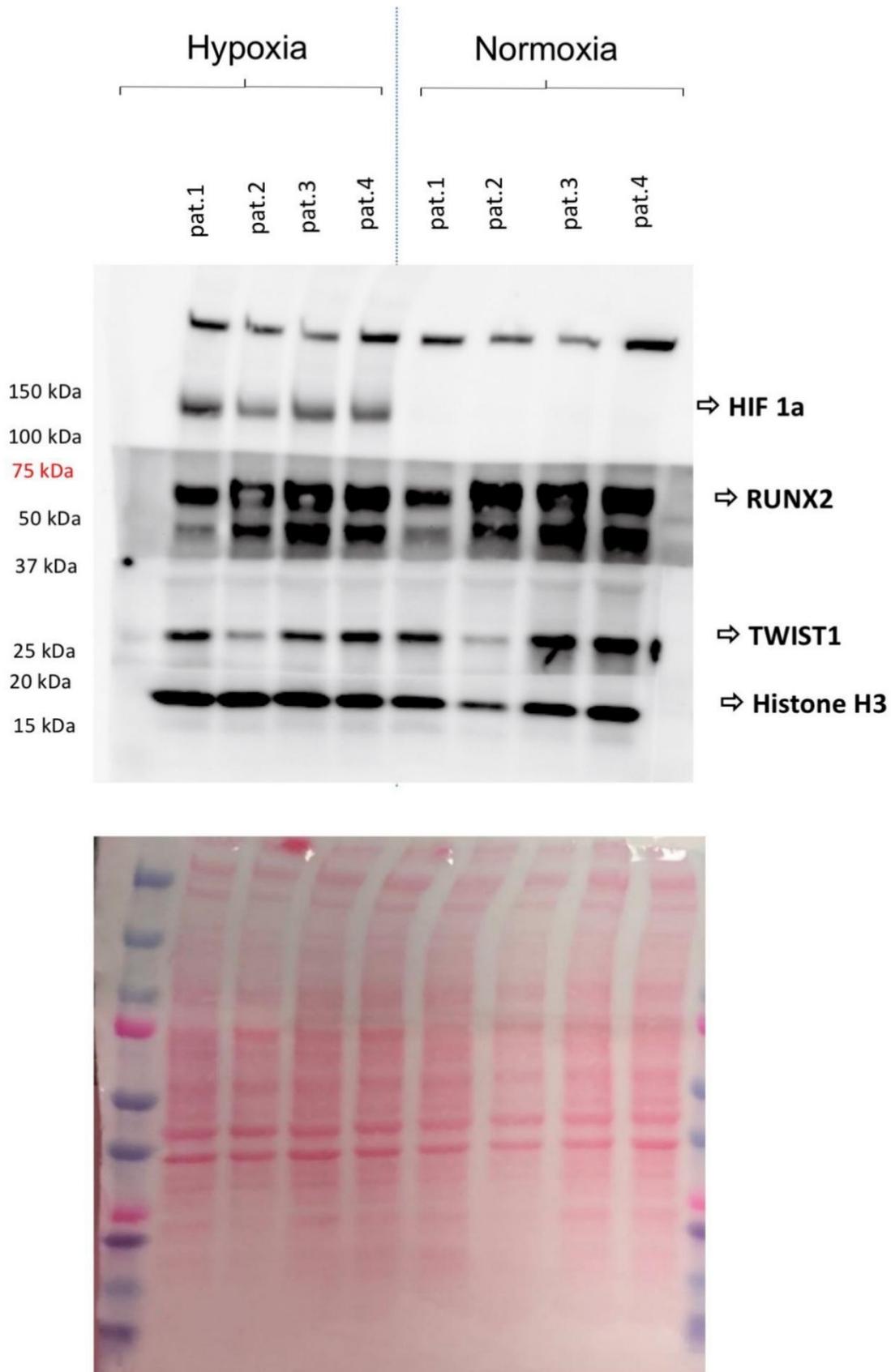


Figure S5. The uncropped western blot membrane of figure 4F and respective Ponceau Red stained loading control images below; n=4.

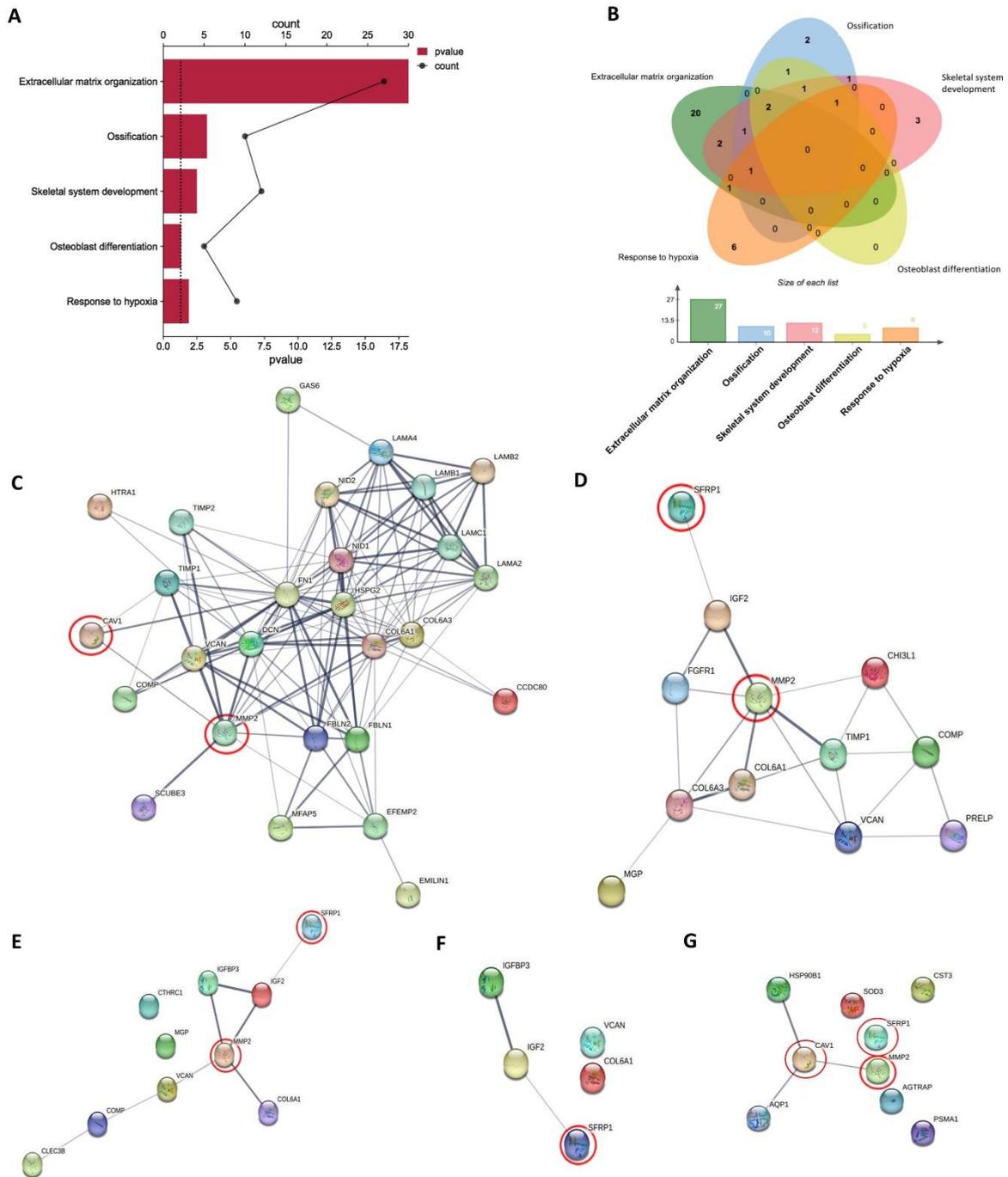


Figure S6. GO Enrichment analyses and protein-protein interaction (PPI) network analyses of norm-naïve EVs and norm-oste EVs

(A) GO analysis for the “upregulated biological processes” related to hypoxia and osteogenesis in norm-oste EVs compared with that in norm-naïve EVs.

(B) Venn diagram of ‘response to hypoxia associated’, ‘ossification’, ‘skeletal system development’, ‘osteoblast differentiation’ and ‘extracellular matrix (ECM) organization’. PPI

networks of 'ECM organization' (C), 'ossification' (D), 'skeletal system development' (E), 'osteoblast differentiation' (F), and 'response to hypoxia' (G). The three proteins of 'response to hypoxia' (SFRP1, MMP2 and CAV1) in red circles are associated with ECM synthesis and osteogenesis; n=3.

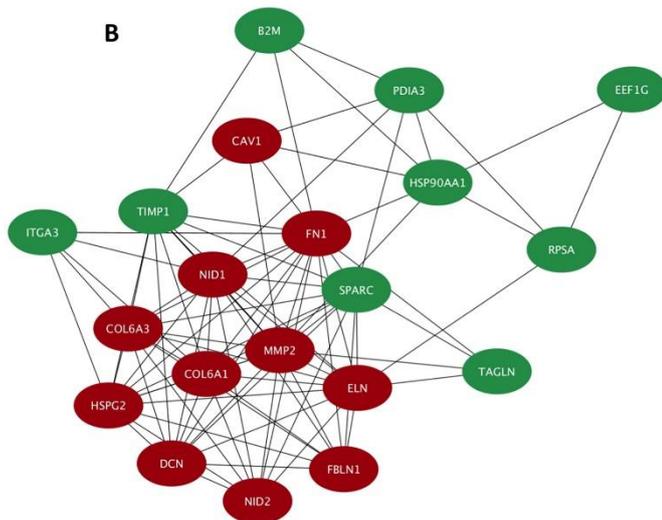
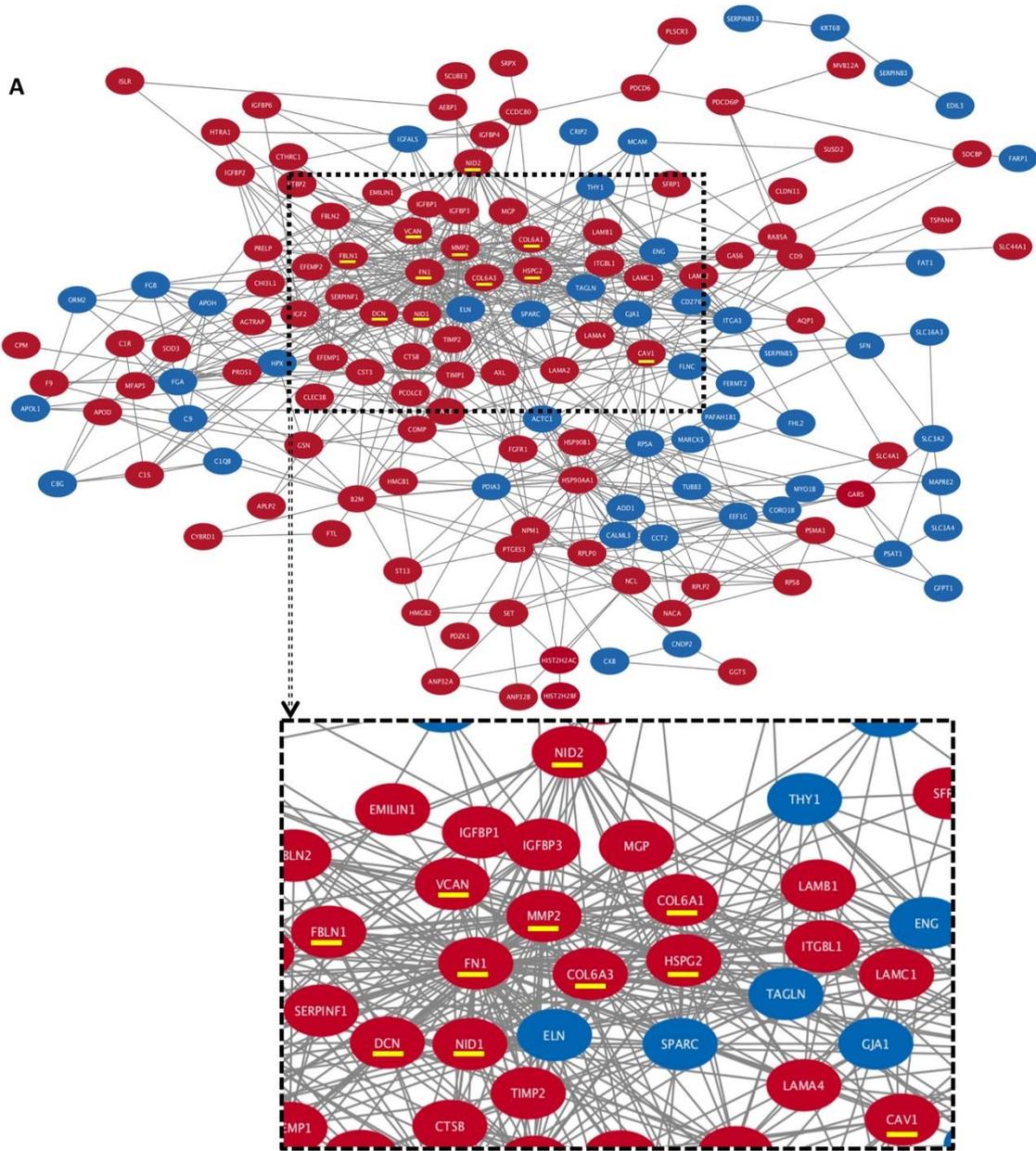


Figure S7. PPI network of the distinct proteins and hub proteins identified in norm-osteo EVs compared with that in norm-naïve EVs .

(A) Interactions between upregulated and downregulated proteins in norm-osteo EVs compared with that in norm-naïve EVs; n=3; Red nodes indicate upregulated proteins, and blue nodes indicate downregulated proteins.

(B) The 20 most highly correlated hub proteins in PPI network. The red nodes indicate the 11 hub proteins related to hypoxia or osteogenesis. The 11 hub proteins are marked with a yellow line.

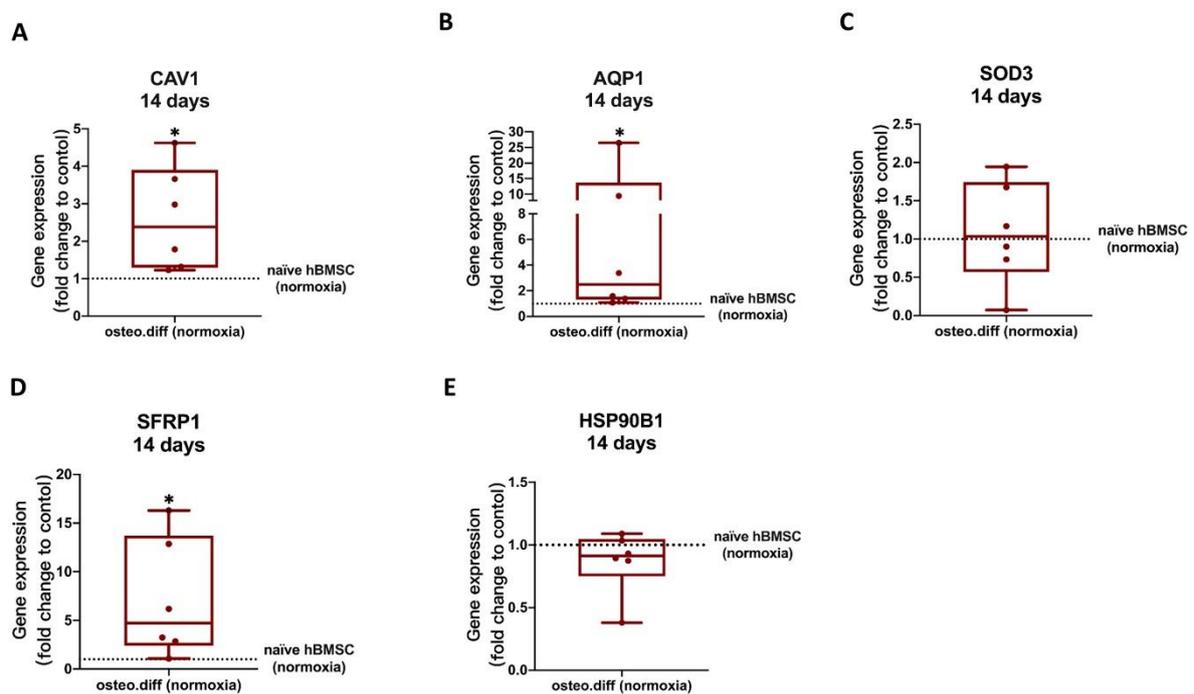


Figure S8. Validation of the proteomic data obtained from norm-naïve EVs and norm-osteo EVs by gene expression analysis in hBMSC.

(A-E) Expression level of the five anti-hypoxic genes were analysed in hBMSCs after 14 days of osteogenic differentiation under normoxia and naïve hBMSCs under normoxia; n=6.

Tables

Table S1. Distinct proteins identified in hypo-osteo EVs and norm-osteo EVs (attached excel file).

Table S2. Primer sequences for qPCR.

Gene	Primer sequences (5'–3')
GAPDH	Fwd, CTGACTTCAACAGCGACACC Rev, CCCTGTTGCTGTAGCCAAAT
TBP	Fwd, TTGTACCGCAGCTGCAAAT Rev, TATATTCGGCGTTTCGGGCA
ALP	Fwd, CCTCCTCGGAAGACACTCTG Rev, CCTCCTCGGAAGACACTCTG
BGLAP	Fwd, GTGCAGAGTCCAGCAAAGGT Rev, TCAGCCAACTCGTCACAGTC
COL1A1	Fwd, ACGTCCTGGTGAAGTTGGTC Rev, ACCAGGGAAGCCTCTCTCTC
RUNX2	Fwd, CGGAATGCCTCTGCTGTTATG Rev, GCTTCTGTCTGTGCCTTCTG
OPN	Fwd, TGAAACGAGTCAGCTGGATG Rev, TGAAATTCATGGCTGTGGAA
SOD3	Fwd, ATGCTGGCGCTACTGTGTT Rev, CTCCGCCGAGTCAGAGTTG
CAV1	Fwd, GCGACCCTAAACACCTCAAC Rev, ATGCCGTCAAAACTGTGTGTC

SFRP1	Fwd, ACGTGGGCTACAAGAAGATGG
	Rev, CAGCGACACGGGTAGATGG
AQP1	Fwd, CTGGGCATCGAGATCATCGG
	Rev, ATCCCACAGCCAGTGTAGTCA
HSP90B1	Fwd, CTGGGCATCGAGATCATCGG
	Rev, ATCCCACAGCCAGTGTAGTCA
