






Review

# NF- $\kappa$ B Pathway and Its Inhibitors: A Promising Frontier in the Management of Alzheimer's Disease

Bhagavathi Sundaram Sivamaruthi <sup>1,2</sup>, Neha Raghani <sup>3</sup>, Mehul Chorawala <sup>3</sup>, Sankha Bhattacharya <sup>4</sup>,  
Bhupendra G. Prajapati <sup>5,\*</sup>, Gehan M. Elossaily <sup>6</sup> and Chaiyavat Chaiyasut <sup>2,\*</sup>

<sup>1</sup> Office of Research Administration, Chiang Mai University, Chiang Mai 50200, Thailand; sivamaruthi.b@cmu.ac.th

<sup>2</sup> Innovation Center for Holistic Health, Nutraceuticals, and Cosmeceuticals, Faculty of Pharmacy, Chiang Mai University, Chiang Mai 50200, Thailand

<sup>3</sup> Department of Pharmacology and Pharmacy Practice, L. M. College of Pharmacy, Ahmedabad 380009, India

<sup>4</sup> School of Pharmacy & Technology Management, SVKM'S NMIMS Deemed-to-be University, Shirpur 425405, India

<sup>5</sup> Shree S. K. Patel College of Pharmaceutical Education and Research, Ganpat University, Mehsana 384012, India

<sup>6</sup> Department of Basic Medical Sciences, College of Medicine, AlMaarefa University, P.O. Box 71666, Riyadh 11597, Saudi Arabia

\* Correspondence: bhupen27@gmail.com (B.G.P.); chaiyavat@gmail.com (C.C.)

**Abstract:** The nuclear factor kappa B (NF- $\kappa$ B) pathway has emerged as a pivotal player in the pathogenesis of various diseases, including neurodegenerative illnesses like Alzheimer's disease (AD). The involvement of the NF- $\kappa$ B pathway in immune system responses, inflammation, oxidative stress, and neuronal survival highlights its significance in AD progression. We discuss the advantages of NF- $\kappa$ B pathway inhibition, including the potential to mitigate neuroinflammation, modulate amyloid beta (A $\beta$ ) production, and promote neuronal survival. However, we also acknowledge the limitations and challenges associated with this approach. Balancing the fine line between dampening inflammation and preserving physiological immune responses is critical to avoid unintended consequences. This review combines current knowledge on the NF- $\kappa$ B pathway's intricate involvement in AD pathogenesis, emphasizing its potential as a therapeutic target. By evaluating both advantages and limitations, we provide a holistic view of the feasibility and challenges of NF- $\kappa$ B pathway modulation in AD treatment. As the quest for effective AD therapies continues, an in-depth understanding of the NF- $\kappa$ B pathway's multifaceted roles will guide the development of targeted interventions with the potential to improve AD management.

**Keywords:** Alzheimer's disease; NF- $\kappa$ B; amyloid beta plaques; neuroinflammation; phytochemicals



**Citation:** Sivamaruthi, B.S.; Raghani, N.; Chorawala, M.; Bhattacharya, S.; Prajapati, B.G.; Elossaily, G.M.; Chaiyasut, C. NF- $\kappa$ B Pathway and Its Inhibitors: A Promising Frontier in the Management of Alzheimer's Disease. *Biomedicines* **2023**, *11*, 2587. <https://doi.org/10.3390/biomedicines11092587>

Academic Editor: Cristina Carvalho

Received: 28 August 2023

Revised: 14 September 2023

Accepted: 19 September 2023

Published: 21 September 2023



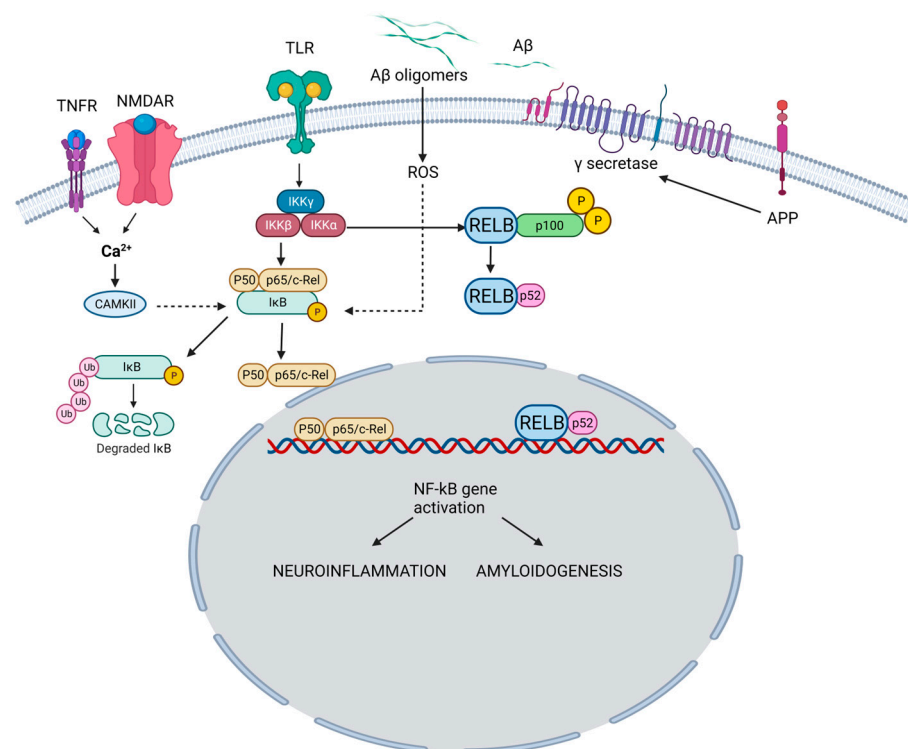
**Copyright:** © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

Alzheimer's disease (AD), a neurodegenerative disease, is the most common type of dementia, responsible for up to 75% of all cases [1]. It manifests as a clinical syndrome presenting symptoms like impairment of memory, language, and other cognitive functions, and behavioral changes that severely affect the activities of daily living. The worldwide prevalence of dementia is thought to be around 3.9%, which is estimated to escalate with the growing size of the population aged 65 years and older. The fundamental histopathological feature of AD has been considered to be aggregates of amyloid beta (A $\beta$ ) plaques in the cortical and limbic regions of the brain [2]. Despite offering a comprehensive framework for understanding AD pathogenesis, certain observations do not seamlessly align with the most straightforward rendition of the hypothesis, leading to its existing lack of intricate explanations [3–5]. It is noteworthy that, intriguingly, no treatment closely tied to the proposed amyloid hypothesis has achieved success thus far. Thus, ineffective attempts to manage the progressive symptoms have compelled new insights into pathogenesis.

Post-translational modifications (PTMs) are pivotal in AD as they influence the aggregation of pathogenic proteins implicated in the disease [6]. Aberrant PTMs, such as the hyperphosphorylation of tau protein and A $\beta$ , glycation, nitration, and ubiquitination, contribute to the misfolding and aggregation of these proteins. The phosphorylation of tau disrupts its normal function, leading to neurofibrillary tangles (NFTs). A $\beta$ , through PTMs like glycation and nitration, adopts toxic conformations, forming amyloid plaques. Ubiquitination may impair the clearance of misfolded proteins. These PTMs disrupt protein homeostasis, impair cellular functions, and trigger neuroinflammation. The interplay between PTMs and nuclear factor kappa B (NF- $\kappa$ B)-driven inflammation thus contributes to protein aggregation, neurodegeneration, and AD progression.

The NF- $\kappa$ B family is a group of transcription factors that play a pivotal role in regulating various biological processes, including immune responses, inflammation, cell survival, and cellular differentiation [7]. The family encompasses five members. NF- $\kappa$ B1 regulates immune responses and inflammation (Figure 1).



**Figure 1.** NF- $\kappa$ B pathway and its role in the pathogenesis of AD. In the canonical pathway, TNFR and NMDAR activation raises  $\text{Ca}^{2+}$ , triggering CaMKII-mediated  $\text{Ca}^{2+}$ -NF- $\kappa$ B linkage. Simultaneously, TLR activation acts on IKK subunits.  $\text{Ca}^{2+}$  leads to I $\kappa$ B kinase activation, I $\kappa$ B breakdown, and p65/p50 dimer formation. NF- $\kappa$ B enters the nucleus, binding neuronal gene targets (e.g., APP) for synaptic function, memory, and amyloid processing. A $\beta$ , resulting from APP cleavage, activates ROS-dependent NF- $\kappa$ B, intensifying amyloid dysregulation. Pro-inflammatory agents (e.g., TNF- $\alpha$ , IL-1 $\beta$ ) activate TLR through canonical (IKK $\beta$ -NEMO) and non-canonical (IKK $\gamma$ , IKK- $\alpha$ ) pathways. Both drive NF- $\kappa$ B-mediated neuroinflammation and neurodegeneration in AD. TNFR: Tumor necrosis factor receptor; NMDAR: N-Methyl-D-Aspartate Receptor; TLR: Toll-like receptor; A $\beta$ : Amyloid beta; APP: Amyloid precursor protein; CaMKII: Calcium-calmodulin (CaM)-dependent protein kinase I; IKK $\gamma$ : Inhibitor of nuclear factor- $\kappa$ B Kinase gamma (also known as NEMO: Nuclear factor- $\kappa$ B (NF- $\kappa$ B) essential modulator); IKK $\alpha$ : Inhibitor of nuclear factor- $\kappa$ B Kinase alpha; IKK $\beta$ : Inhibitor of nuclear factor- $\kappa$ B kinase beta; I $\kappa$ B: Inhibitor of  $\kappa$ B; ROS: Reactive oxygen species. TNF- $\alpha$ : Tumor necrosis factor-alpha, IL-1 $\beta$ : Interlukin-1 beta; RELB: Transcription factor.

Upon proteolytic processing, NF- $\kappa$ B1 generates the active p50 subunit. Similarly, NF- $\kappa$ B2 undergoes proteolytic processing to produce the active p52 subunit, which is involved

in immune responses and lymphoid organ development. The p65 subunit aids in functional NF- $\kappa$ B dimer formation and is pivotal in regulating inflammation, immune responses, and cellular survival. The transcription factor, RelB, contributes to NF- $\kappa$ B dimer assembly and is particularly associated with immune responses, especially in lymphoid organ development. The c-Rel subunit modulates immune responses and gene expression related to inflammation and lymphoid organ development. These NF- $\kappa$ B family members form homo- or heterodimers, dictating target gene specificity and ensuing biological impacts [8].

Within neuroinflammation, NF- $\kappa$ B occupies a central role in coordinating the expression of genes that contribute to both protective and detrimental aspects of the inflammatory response in the nervous system. Primarily sequestered in an inactive state within the cytoplasm, NF- $\kappa$ B is bound to inhibitory proteins named I $\kappa$ Bs. Upon activation, typically induced by a spectrum of stimuli encompassing pro-inflammatory cytokines, pathogens, or cellular stressors, these I $\kappa$ Bs undergo phosphorylation and consequent degradation, facilitating the translocation of NF- $\kappa$ B to the nucleus. This nuclear migration empowers NF- $\kappa$ B to engage specific DNA sequences, thereby orchestrating the modulation of an extensive array of target genes. NF- $\kappa$ B signaling also influences the integrity of the blood–brain barrier (BBB), governing the passage of molecules between the bloodstream and the brain milieu. The activation of NF- $\kappa$ B in the endothelial cells of the BBB provokes heightened permeability, allowing immune cells and molecules to enter the brain [9].

Further, NF- $\kappa$ B activation in microglia triggers their transition from resting to activated states, releasing pro-inflammatory cytokines, chemokines, and reactive oxygen species (ROS). NF- $\kappa$ B is pivotal for initiating protective immune responses against pathogens and facilitating tissue repair. However, prolonged NF- $\kappa$ B activation within neurons can result in neurotoxicity, contributing to oxidative stress, excitotoxicity, and inflammation, all of which undermine neuronal well-being. NF- $\kappa$ B also influences the balance between neuronal survival and apoptosis. In some cases, NF- $\kappa$ B activation can promote the expression of anti-apoptotic genes, supporting neuronal survival [10]. However, chronic NF- $\kappa$ B activation can lead to a pro-apoptotic response in other situations.

This review summarizes the role of the NF- $\kappa$ B pathway as a potential target in treating AD, along with the prospects and challenges of its modulation as a therapeutic strategy. Also, the manuscript discusses the interplay between inflammation and neurodegeneration.

## 2. Role of NF- $\kappa$ B Pathway Inhibitors in AD Treatment

The NF- $\kappa$ B pathway plays a significant role in neuroinflammation and other processes implicated in AD pathogenesis. Consequently, it has been considered a potential therapeutic target for the management of AD [11]. However, targeting the NF- $\kappa$ B pathway is complex due to its multifaceted role in promoting and regulating inflammation and immune responses. NF- $\kappa$ B pathway inhibitors offer the means to modulate this intricate signaling network and counteract neuroinflammation. By inhibiting NF- $\kappa$ B activation, these inhibitors can dampen the excessive release of pro-inflammatory molecules, thereby reducing the harmful impact of inflammation on neurons [12]. They hold the potential to alleviate neuronal damage and promote their survival in the face of AD-associated challenges.

Furthermore, NF- $\kappa$ B inhibitors may influence key pathological hallmarks of AD: A $\beta$  plaques and tau tangles. NF- $\kappa$ B activation regulates enzymes involved in A $\beta$  production and clearance [13]. Inhibiting this pathway could potentially modify A $\beta$  metabolism, impacting the deposition and accumulation of A $\beta$  plaques. Similarly, NF- $\kappa$ B activation has been linked to tau hyperphosphorylation and tangle formation. Inhibitors might intervene in these processes, attenuating tau pathology and promoting neuronal stability. However, it is essential to tread cautiously in inhibiting NF- $\kappa$ B, as this pathway also plays crucial roles in immune responses and cell survival [14]. A complete blockade could compromise the immune system's ability to defend against pathogens and maintain tissue integrity. Striking the right balance between dampening neuroinflammation and preserving necessary immune functions is critical. The direct inhibition of NF- $\kappa$ B activation is a strategy under

investigation for its therapeutic potential in various diseases, including AD [15]. Directly inhibiting NF- $\kappa$ B activation modulates the aberrant inflammatory response, potentially slowing disease progression. Researchers are exploring small molecules and compounds that directly target components of the NF- $\kappa$ B signaling pathway. These agents can interfere with the activation steps that lead to NF- $\kappa$ B translocation into the nucleus, where it regulates gene expression.

Advancements in NF- $\kappa$ B pathway inhibitor research are multifaceted. Scientists are striving to develop selective inhibitors that target specific pathway components, minimizing off-target effects. A study conducted by Elzayat and colleagues investigates mesenchymal stem cells (MSCs) and acitretin for AD treatment, focusing on the NF- $\kappa$ B pathway and miRNA regulation. Using a rat model, miR-146a, miR-155, necrotic, growth, and inflammatory genes were analyzed. Results showed that MSCs and acitretin restored normal levels, indicating potential therapeutic benefits. MiR-146a and miR-155 are proposed as AD biomarkers. This study underscores MSCs and acitretin's capacity to modulate miRNAs and related genes in the NF- $\kappa$ B pathway, offering insights for future AD interventions [16].

Melatonin, a hormone the pineal gland produces, has been recognized for its role in regulating sleep–wake cycles and its potential anti-inflammatory properties. While melatonin's direct impact on the NF- $\kappa$ B pathway is limited, it has been suggested that melatonin can indirectly modulate NF- $\kappa$ B activation through various mechanisms. Oxidative stress can activate NF- $\kappa$ B, leading to the expression of pro-inflammatory genes. Melatonin's antioxidant properties indirectly inhibit NF- $\kappa$ B activation by reducing the triggering oxidative signals [17]. Melatonin can influence molecules downstream of the NF- $\kappa$ B pathway. For instance, it has been reported to downregulate COX-2 (Cyclooxygenase-2) expression, an inflammation enzyme that NF- $\kappa$ B often regulates. Melatonin's neuroprotective effects may indirectly impact NF- $\kappa$ B activation. By preserving neuronal health and reducing neuroinflammation, melatonin can contribute to an environment where NF- $\kappa$ B activation is less pronounced. Merlo et al. demonstrated the counteracting effects of melatonin on the pro-inflammatory properties of lipopolysaccharides in *in vitro* models [18]. Another study investigated a phosphodiesterase 5 inhibitor (PDE5I) with dual antagonistic action on IKKB and TNFR1 to inhibit NF- $\kappa$ B and neuroinflammation. *In silico* docking with FDA-approved compounds identified avanafil with optimal IKKB and TNFR1 binding. Molecular dynamic studies confirmed avanafil's stability with these targets. In a mouse model of lipopolysaccharide (LPS)-induced neuroinflammation and cognitive decline, avanafil at 6 mg/kg improved cognitive performance, reduced A $\beta$  levels, and lowered inflammatory markers and oxidative parameters [19].

Bay 11-7082 is a synthetic small molecule that has gained attention for its potential as an NF- $\kappa$ B pathway inhibitor. This compound has shown inhibitory effects on I $\kappa$ B kinase (IKK), allowing NF- $\kappa$ B to translocate into the nucleus and initiate the expression of target genes involved in inflammation and immune responses. By inhibiting IKK and I $\kappa$ B degradation, Bay 11-7082 effectively interferes with interleukin-6 (IL-6) secretion and the subsequent activation of NF- $\kappa$ B, leading to decreased pro-inflammatory gene expression [20]. For Bay 11-7082 to effectively modulate NF- $\kappa$ B activity within the brain, it must cross the BBB. Developing strategies to enhance its delivery and distribution to the central nervous system while minimizing systemic effects is still a significant challenge [18]. Additionally, a low dose of recombinant IL-2 (aldesleukin) is currently under clinical trial for its clinical application in AD (NCT05821153; NCT05468073).

The SN50 analog is a cell-permeable, synthetic peptide derived from the NF- $\kappa$ B nuclear localization sequence, and it holds potential in AD treatment due to its ability to inhibit NF- $\kappa$ B nuclear translocation. A study conducted by Lin and colleagues suggests that the inhibitory effect is not due to disrupting the binding activity of NF- $\kappa$ B complexes but rather its ability to enter the cell and compete with NF- $\kappa$ B complexes for the cellular machinery responsible for nuclear translocation [21,22]. A study by Pogue and team reveals that upregulated miRNA-30b, associated with neuropathology in AD brain and LPS-stressed human neuronal–glial cells, targets neurofilament light (NF-L)-chain mRNA's 3'-UTR. This

results in the post-transcriptional downregulation of NF- $\kappa$ B expression, observed in AD and LPS-treated cells, leading to neuronal cytoskeleton atrophy and synaptic disruption. MiRNA-30b is highly expressed in A $\beta$ -treated models, facilitating LPS entry into neurons. Elevated miRNA-30b induces neuro-injury, inflammation, synaptic impairment, and neuron loss. This gut-microbiota-derived LPS-NF- $\kappa$ B-miRNA-30b-NF- $\kappa$ B pathway links GI tract microbes' LPS to AD's neuropathology and synaptic disruption, offering insights into neurodegeneration mechanisms [23]. Optimizing dosages to achieve the desired therapeutic outcomes while avoiding potential side effects is an ongoing challenge. Table 1 enlists molecules being researched for their potential role in the NF- $\kappa$ B pathway in AD.

**Table 1.** The list of representative molecules targeting the NF- $\kappa$ B pathway to treat and manage Alzheimer's disease.

Molecule	NF- $\kappa$ B Target	Experimental Setting	Mechanism of Action	References
Alogliptin	Direct	In vitro	Modulates TLR4/MYD88/NF- $\kappa$ B and miRNA-155/SOCS-1 signaling pathways	[24]
AS62868	Direct	In vitro	Inhibits IKK $\beta$	[25]
Docosahexaenoic acid	Indirect	In vitro	Precursor to produce neuroprotection D1, a neuro-protecting agent in the CNS	[26]
Etanercept	Direct	RCT	TNF- $\alpha$ activity inhibitor	[27,28]
Forsythoside B	Direct	In vivo and in vitro	Reduces the serine 536 phosphorylation of IKK $\alpha/\beta$ , I $\kappa$ B $\alpha$ , and p65	[29]
Minocycline	Direct	In vivo and in vitro	Reduction in IL-6; BACE inhibition	[30,31]
Pioglitazone	Indirect	In vitro	Downregulates glycogen synthase kinase 3 beta and cyclin-dependent kinase in microglia cells	[32,33]
PUFA-Plasmalogens	Direct	In vitro	Inhibits NF- $\kappa$ B, p38MAPK, and JNK pathways	[34]
Simufilam	Indirect	Inpatient	Reduces mTOR basal overactivity; restores the normal shape and function of Filamin A	[35]
Telmisartan	Indirect	In vivo and in vitro	AT <sub>1</sub> blocking reduces IL-1 $\beta$ levels, resulting in anti-neuroinflammatory effects through JNK/c-Jun and NADPH oxidase pathways; partial PPAR-gamma-stimulating activity	[36–38]
TPCA-1	Direct	In vivo	Inhibits IKK $\beta$	[39]
VX-745	Indirect	In vivo	p38 MAPK $\alpha$ inhibitor	[40]

mTOR: Mammalian target of rapamycin; IKK $\beta$ : Nuclear factor kappa B kinase subunit beta; TNF- $\alpha$ : Tumor necrosis factor alpha; IL-6: Interleukin 6; BACE:  $\beta$ -Site amyloid precursor protein-cleaving enzyme; CNS: Central nervous system; NF- $\kappa$ B: Nuclear factor kappa B; p38MAPK: p38 mitogen-activated protein kinases; JNK: c-Jun N-terminal kinase; MYD88: Myeloid differentiation primary response protein 88; miRNA-155: MicroRNA-155; SOCS-1: Suppressor of cytokine signaling 1; I $\kappa$ B $\alpha$ : Nuclear factor of kappa light polypeptide gene enhancer in B-cells inhibitor, alpha; p38 MAPK $\alpha$ : p38 mitogen-activated protein kinase alpha. RCT: Randomized clinical trials; AT<sub>1</sub>: Angiotensin 1; NADPH: Nicotinamide Adenine Dinucleotide Phosphate Hydrogen.

In practical terms, combining NF- $\kappa$ B inhibition with other therapeutic approaches may yield enhanced results. Research conducted by Shehata et al. aimed to formulate nanostructured lipid carriers (NLCs) that co-encapsulate donepezil and astaxanthin (DPL/AST-NLCs) and assess their effectiveness in AD-like rat models following daily intranasal administration [41]. AD is a complex disease with multiple underlying mechanisms, in-

cluding A $\beta$  aggregation, tau pathology, and synaptic dysfunction. Integrating NF- $\kappa$ B inhibitors with interventions targeting these aspects could offer a comprehensive treatment strategy, potentially addressing symptom relief and disease modification.

To translate NF- $\kappa$ B inhibition into clinical practice, robust clinical trials are imperative. These trials should involve diverse patient populations, incorporate appropriate outcome measures, and assess long-term safety. Furthermore, identifying reliable biomarkers that reflect NF- $\kappa$ B activity and its downstream effects will aid in monitoring treatment efficacy and patient response.

### 3. Factors Affecting NF- $\kappa$ B Inhibitors-Based AD Treatments

Inhibiting NF- $\kappa$ B in the context of AD presents a formidable scientific challenge due to the intricate and multifaceted nature of the disease and the pivotal role of NF- $\kappa$ B in immune responses and inflammation. NF- $\kappa$ B serves a dual role, exhibiting both pro-inflammatory and anti-inflammatory functions. It plays a crucial part in the innate immune response, essential for pathogen clearance and tissue homeostasis. For instance, NF- $\kappa$ B activation in microglial cells can release pro-inflammatory cytokines, such as TNF- $\alpha$  and IL-1 $\beta$ , contributing to neuroinflammation in AD [42]. However, NF- $\kappa$ B also has anti-inflammatory functions by promoting the expression of anti-inflammatory cytokines like IL-10. Thus, the complete inhibition of NF- $\kappa$ B may risk suppressing these protective anti-inflammatory responses, vital for maintaining brain homeostasis [43].

Consequently, attempting to completely inhibit NF- $\kappa$ B raises concerns regarding impaired immune responses, necessitating a delicate balance between mitigating its detrimental effects in AD and preserving its protective capabilities. One instance highlighting this challenge is that a non-specific inhibition of NF- $\kappa$ B could disturb this control mechanism, resulting in unregulated NF- $\kappa$ B activation and consequently harming neurons [44]. Furthermore, NF- $\kappa$ B is rigorously controlled through various mechanisms, including inhibitory proteins (I $\kappa$ Bs) and post-translational modifications, making precise and specific inhibition challenging. NF- $\kappa$ B operates within diverse cell types in the central nervous system, including neurons and glial cells, with its effects contingent upon cell type, rendering it problematic to target NF- $\kappa$ B without unintentional consequences in various cell populations. In astrocytes, NF- $\kappa$ B can have protective effects by regulating the release of neurotrophic factors. These beneficial effects might be compromised if NF- $\kappa$ B is inhibited without cell type specificity [45].

Additionally, AD is a progressive ailment encompassing distinct stages, and the contribution of NF- $\kappa$ B may evolve throughout the disease course, necessitating the consideration of temporal factors for therapeutic intervention. Furthermore, the limited capacity of numerous potential NF- $\kappa$ B inhibitors to penetrate the blood–brain barrier (BBB) effectively poses a significant challenge, as inhibitors must adequately reach neural tissues to modulate NF- $\kappa$ B activity. The potential for off-target effects, impacting alternate signaling pathways or cellular functions, further complicates NF- $\kappa$ B inhibition. Moreover, AD's heterogeneity, characterized by diverse underlying mechanisms and pathological manifestations, results in NF- $\kappa$ B potentially exerting differential impacts on disease progression among individuals, augmenting the complexity of targeted inhibition. Furthermore, the intricate web of interactions between NF- $\kappa$ B and other signaling pathways, including those associated with amyloid-beta and tau pathology, further complicates the potential outcomes of NF- $\kappa$ B inhibition. Inhibiting NF- $\kappa$ B, for example, may disrupt its interaction with the vital Notch signaling pathway, with potential consequences for neuronal development and function, thereby impacting neuronal health [46]. Lastly, the inhibition of NF- $\kappa$ B may trigger adaptive or compensatory cellular responses, potentially leading to the activation of alternative pro-inflammatory pathways, posing additional challenges in achieving effective and durable therapeutic outcomes [47].

#### 4. Insights and Challenges to Targeting the NF- $\kappa$ B Pathway

In the context of AD, the inhibition of NF- $\kappa$ B holds substantial promise as a therapeutic target. As a central regulator of neuroinflammation, NF- $\kappa$ B inhibition can mitigate the excessive inflammatory responses witnessed in AD brains, potentially alleviating the harmful consequences of chronic inflammation on neuronal health. By influencing A $\beta$  metabolism and accumulation, the engagement of NF- $\kappa$ B establishes the possibility of utilizing its inhibition to regulate A $\beta$  production and clearance [48]. This modulation could potentially mitigate A $\beta$  buildup and delay the development of plaques. The intricate role of NF- $\kappa$ B in neuronal survival is noteworthy; it can induce the expression of anti-apoptotic genes, thereby offering potential neuroprotective effects upon inhibition [49]. Notably, targeting NF- $\kappa$ B addresses symptomatic manifestations and certain fundamental mechanisms underpinning AD pathogenesis, suggesting a capacity to modify disease progression. Integrating NF- $\kappa$ B inhibitors with other therapeutic strategies aimed at various aspects of AD pathology holds promise for comprehensive treatment effects. The wealth of research on NF- $\kappa$ B inhibitors in diverse disease contexts provides a valuable foundation for AD treatment development. Leveraging this existing knowledge accelerates the translational potential of NF- $\kappa$ B inhibition as a viable strategy for AD. The well-established role of NF- $\kappa$ B in AD pathogenesis solidifies its standing as a rational and scientifically grounded therapeutic target, bolstering the credibility of NF- $\kappa$ B inhibition as a plausible avenue for effective AD treatment.

The exploration of NF- $\kappa$ B as a therapeutic target in AD holds significant promise, but several challenges must be addressed for successful clinical applications. Subsequent research should emphasize formulating strategies for the selective modulation of NF- $\kappa$ B activity within distinct cell types, encompassing neurons, microglia, and astrocytes [50]. This precise targeting holds the potential to avert undesired effects while achieving therapeutic objectives. Precisely regulating the extent of NF- $\kappa$ B inhibition stands as a critical imperative. Total abrogation might compromise NF- $\kappa$ B's advantageous contributions to immune responses and cellular survival [51]. There is a need to devise methodologies that establish an equilibrium between attenuating neuroinflammation and safeguarding fundamental cellular functions. Given the intricate nature of AD, amalgamating therapies focused on NF- $\kappa$ B with complementary interventions could yield enhanced outcomes [52].

Integrating NF- $\kappa$ B inhibition with strategies addressing amyloid-beta accumulation, tau pathology, or synaptic dysfunction could yield a holistic therapeutic approach. Acknowledging the heterogeneous nature of AD patients and the diverse mechanisms underlying the ailment, prospective treatments could necessitate customization based on individual molecular profiles and disease progression [53,54]. Pioneering the development of dependable biomarkers reflecting NF- $\kappa$ B activity and downstream ramifications within the brain could facilitate gauging treatment efficacy and patient responsiveness to NF- $\kappa$ B-targeted interventions. Overcoming the considerable challenge of designing therapeutic agents capable of crossing the BBB to access the central nervous system requires concentrated research on enhancing drug delivery methodologies. Safeguarding the long-term safety of NF- $\kappa$ B-targeted therapies holds paramount importance. Prolonged NF- $\kappa$ B inhibition may engender off-target ramifications, potentially influencing immune responses and broader physiological processes [55]. A comprehensive evaluation of safety implications is imperative. Methodical and meticulously designed clinical trials are necessary to systematically assess the effectiveness and safety of NF- $\kappa$ B-targeted therapies. Such trials should incorporate diverse patient cohorts, employ pertinent outcome measures, and delineate well-defined endpoints. The disparate impacts of distinct NF- $\kappa$ B inhibitors on neuroprotection, inflammation, and immune responses necessitate further exploration. A comprehensive investigation is warranted to discern the potential of these inhibitors in addressing specific facets of AD pathogenesis. Beyond alleviating symptomatic manifestations, the trajectory of NF- $\kappa$ B-targeted therapies should encompass the development of interventions capable of arresting or decelerating the inherent progression of the disease, thereby confronting the fundamental causes of AD [42].

## 5. Future Directions

The prospects of NF- $\kappa$ B as an AD therapeutic target lie in refining its modulation to culminate in optimal therapeutic consequences. The convergence of advancements in precision medicine, biomarker elucidation, drug delivery modalities, and an enriched comprehension of NF- $\kappa$ B's intricate roles in AD pathogenesis is poised to catalyze the successful transition of this strategy from preclinical research to efficacious clinical treatments.

The future direction of targeting NF- $\kappa$ B inhibition through phytochemicals represents a promising and innovative avenue for AD treatment [56]. This approach capitalizes on the potential of natural compounds derived from plants to modulate NF- $\kappa$ B activity, offering a novel way forward in AD management. Several key directions and strategies can shape the progression of NF- $\kappa$ B inhibition using phytochemicals as a target for AD. Future research should prioritize the identification of phytochemicals with robust NF- $\kappa$ B-inhibitory properties. Screening and evaluating various plant-derived compounds can uncover novel candidates that effectively modulate NF- $\kappa$ B signaling pathways. Curcumin has demonstrated potent anti-inflammatory properties by inhibiting NF- $\kappa$ B activation. It may also impact amyloid-beta aggregation and tau phosphorylation, key factors in AD pathogenesis [57]. Epigallocatechin gallate (EGCG) is a bioactive polyphenol found abundantly in green tea, and it has gained attention for its potential benefits in AD management. EGCG has been shown to suppress the activation of NF- $\kappa$ B, a key regulator of inflammation. By inhibiting NF- $\kappa$ B, EGCG can mitigate the production of pro-inflammatory cytokines, chemokines, and other molecules involved in neuroinflammation, a prominent feature of AD [58]. *Salvia miltiorrhiza* is a traditional Chinese herbal medicine that has garnered attention for its potential benefits in AD management. The primary active compounds (Tanshinones and salvianolic acids) in *S. miltiorrhiza* have been investigated for their multifaceted effects, including their potential interaction with NF- $\kappa$ B. *S. miltiorrhiza* has been explored for its potential to influence A $\beta$  metabolism. By modulating NF- $\kappa$ B, *S. miltiorrhiza*'s compounds might impact A $\beta$  production, clearance, and aggregation, key factors in AD pathogenesis. *S. miltiorrhiza* compounds have shown promise in reducing tau hyperphosphorylation, a key feature of neurofibrillary tangles in AD [59].

Ashwagandha (*Withania somnifera*), an adaptogenic herb with a long history in traditional medicine, has gained attention for its potential benefits in AD management. The active compounds in ashwagandha, known as withanolides, have been studied for their diverse effects, including their potential interaction with NF- $\kappa$ B signaling. Ashwagandha has been suggested to enhance the expression of neurotrophic factors that support neuronal growth and plasticity. Its interaction with NF- $\kappa$ B may play a role in these effects. Using a *Drosophila melanogaster* AD model, a study explored Ashwagandha's impact on beta-amyloid toxicity and longevity. At 20 mg/mL, Ashwagandha rescued AD-related phenotypes and enhanced longevity in AD and wild-type *Drosophila*. These findings suggest Ashwagandha's potential as a potent AD treatment and cellular well-being enhancer [60]. Turner et al. conducted a 52-week randomized, placebo-controlled study investigating resveratrol's effects on mild to moderate AD. Resveratrol, well tolerated and able to cross the BBB, showed altered biomarker trajectories. Notably, CSF A $\beta$ 40 and plasma A $\beta$ 40 levels decreased less in the resveratrol group compared to placebo after 52 weeks. It exhibits antioxidant activity, which can counteract oxidative-stress-induced NF- $\kappa$ B activation. While providing class II evidence for resveratrol's safety and influence on AD biomarkers, further research is necessary to understand these changes [61]. While *Ginkgo biloba* is generally considered safe for short-term use, its efficacy in treating AD remains debatable [62]. Clinical trials examining the effects of *G. biloba* in AD have yielded mixed results. While some studies have reported modest cognitive improvements and slowed decline, others have found no significant benefit compared to placebo. Table 2 lists the representative phytochemicals demonstrating promising bioactivity against AD encompassing the NF- $\kappa$ B signaling pathway. A comprehensive understanding of how specific phytochemicals modulate NF- $\kappa$ B activation is crucial. Research efforts should elucidate the molecular



mechanisms underlying their effects, potentially leading to the development of targeted therapies with minimized off-target effects.

**Table 2.** Phytochemicals targeting the NF- $\kappa$ B pathway as therapeutic agents for Alzheimer's disease.

Phytochemicals	Origin	Mechanism of Action	Experimental Setting	References
Kai-Xin-San (KXS)	<i>Radix et Rhizoma Ginseng</i> <i>Radix Polygalae</i> , <i>Acori tatarinowii Rhizoma</i> , <i>Wolfiporia extensa</i>	Wnt/ $\beta$ -catenin signaling activation; IRE1/XBP1s pathway inhibition	In vivo	[63]
Scoparone	<i>Artemisia scoparia</i>	TLR4/MyD88/TRAF-6/TAK-1/NF- $\kappa$ B axis inactivation	In vivo	[64]
Moxibustion	<i>Artemisia vulgaris</i>	Reduces the expression of CD206 and secretion of IL-10; increases microglial polarization (M1 to M2)	In vivo	[65]
Citropten	<i>Citrus aurantifolia</i>	MAPK and PLC $\gamma$ /Ca <sup>2+</sup> pathway modulation	In vitro	[66]
Diterpenoids (Caeminaxin A, B, and others)	<i>Caesalpinia minax</i> Hance	Inhibition of iNOS, COX-2 expression; suppresses phosphorylation of MAPK; suppresses activation of NF- $\kappa$ B pathway	In vitro	[67]
Icariin (ICA)	<i>Epimedium koreanum</i>	Upregulates PPAR $\gamma$ , TAK1/IKK/NF- $\kappa$ B, and JNK/p38 MAPK signaling pathways; PSD-95 regulation	In vivo	[68]
Andrographolide	<i>Andrographis paniculata</i>	Enhances the expression of LRP-1, NF- $\kappa$ B; decreases IL-1 $\beta$ , IL-6, and TNF- $\alpha$ levels	In vitro	[69]
Biochanin-A	<i>Trifolium pretense</i> <i>Medicago sativa</i>	Increases phosphorylation of PI3K and Akt	In vivo	[70,71]
Malvidin-3-O-glucoside ( <i>B. atropurpurea</i> anthocyanin)	<i>Berberis atropurpurea</i> Schneid.	NF- $\kappa$ B, I $\kappa$ B, TLR4, and MyD88 downregulation	In silico, in vitro	[72]
Erjing Pills	<i>Polygonatum sibiricum</i> <i>Lycium chinense</i>	IL-1 $\beta$ , TNF- $\alpha$ , and IL-6 reduction; down regulated TLR4, p-NF- $\kappa$ B P65/NF- $\kappa$ B P65, p-I $\kappa$ B $\alpha$ /I $\kappa$ B $\alpha$ , and NLRP3 expression levels	In vivo	[73]
Licochalcone A	<i>Glycyrrhiza glabra</i>	Decrease in 8-iso-PGF <sub>2<math>\alpha</math></sub> , IL-6 and TNF $\alpha$ levels	In vitro	[74]
Honokiol	<i>Magnolia officinalis</i>	NF- $\kappa$ B inhibition, decrease in A $\beta$ plaques	In vivo	[75]
Shi chang pu (SCP) essential oil	<i>Acorus tatarinowii</i> Schott	Inhibits phosphorylation of IKK $\beta$ , NF- $\kappa$ B, and NLRP3	In vivo	[76]
Cardamonin (CD)	<i>Alpinia katsumadai</i> <i>Alpinia conchigera</i>	Decreases TNF- $\alpha$ , IL-1, and IL-6 levels; NF- $\kappa$ B and STAT3 modulation	In vitro	[77,78]

Table 2. Cont.

Phytochemicals	Origin	Mechanism of Action	Experimental Setting	References
Platycodin D	<i>Platycodon grandiflorus</i>	TLR4 and p-p65 expression attenuation; decrease in ROS and MDA production	In vitro	[79]
Baicalin (BA)	<i>Scutellaria baicalensis</i>	Blocking signal transduction via TLR4, facilitated by the TLR4/MyD88/NF-κB and MAPK pathways; CX3CR1/NF-κB signaling pathway modulation	In vitro	[80,81]
Isorhamnetin	<i>Oenanthe javanica</i>	Downregulation of IBA1, NF-κB and CD11b expression	In vitro	[82]
Esculetin	<i>Artemisia capillaris</i> Thunb	Inhibits NO production and iNOS expression; inhibits nuclear translocation of NF-κB p65	In vitro	[83]

Wnt/ $\beta$ -catenin: Wingless-related integration site/ $\beta$ -catenin; IRE1/XBP1s: Inositol-requiring enzyme 1/X-box binding protein 1; TRAF-6: Tumor necrosis factor receptor (TNFR)-associated factor 6; TAK-1: Transforming growth factor- $\beta$  (TGF- $\beta$ )-activated kinase 1; NF- $\kappa$ B: Nuclear factor kappa B; CD206: Cluster of differentiation 206; IL-10: Interleukin 10; MAPK: Mitogen-activated protein kinases; PLC $\gamma$ /Ca<sup>2+</sup>: Phospholipase C $\gamma$ /Calcium<sup>2+</sup>; iNOS: Inducible nitric oxide synthase; COX-2: Cyclooxygenase-2; PPAR $\gamma$ : Peroxisome proliferators-activated receptor gamma; IKK: Nuclear factor kappa B kinase; JNK/p38 MAPK: c-Jun N-terminal kinase/p38 mitogen-activated protein kinases; PSD-95: Postsynaptic density protein 95; LRP-1: Low-density lipoprotein (LDL)-related protein-1; IL-1 $\beta$ : Interleukin-1 beta; IL-6: Interleukin 6; TNF- $\alpha$ : Tumor necrosis factor alpha; PI3K: Phosphoinositide 3-kinases; Akt: Protein kinase B; p-I $\kappa$ B $\alpha$ /I $\kappa$ B $\alpha$ : Phosphorylated nuclear factor of kappa light polypeptide gene enhancer in B-cells inhibitor/nuclear factor of kappa light polypeptide gene enhancer in B-cells inhibitor; NLRP3: Nucleotide-binding domain, leucine-rich-containing family, pyrin domain-containing-3; 8-iso-PGF<sub>2 $\alpha$</sub> : 8-iso-Prostaglandin F<sub>2 $\alpha$</sub> ; A $\beta$  plaques: Amyloid beta plaques; IKK $\beta$ : Nuclear factor kappa B kinase subunit beta; STAT3: Signal transducers and activators of transcription 3; ROS: Reactive oxygen species; MDA: Malondialdehyde; CX3CR1: C-X3-C Motif chemokine receptor 1; IBA1: Ionized calcium-binding adapter molecule 1; NO: Nitric oxide; iNOS: Inducible nitric oxide synthase.

## 6. Conclusions

NF- $\kappa$ B emerges as a pivotal influencer over both the onset and progression of AD. As elucidated, the dual nature of NF- $\kappa$ B becomes evident—a guardian of immune responses and cell survival, yet also a potential instigator of neuroinflammation and neuronal compromise. The multifaceted contributions of NF- $\kappa$ B in AD pathogenesis have stimulated investigations into harnessing its potential as a therapeutic target. Efforts to manipulate NF- $\kappa$ B, both in preclinical models and clinical contexts, present a compelling approach to mitigating the inflammatory milieu characteristic of AD. Yet, this endeavor is not devoid of challenges. The intricate balance between dampening neuroinflammation and preserving essential cellular functions underscores the delicate nature of NF- $\kappa$ B modulation. Its intricate interplay with other molecular players, such as  $\beta$ -secretase and amyloid-beta, demands a nuanced understanding to untangle the intricacies of AD pathogenesis. The evolving trajectory of NF- $\kappa$ B-targeted therapies hinges on precision—selectively addressing specific cell types, customizing interventions based on individual profiles, and optimizing drug delivery methodologies. The amalgamation of NF- $\kappa$ B-focused strategies with complementary approaches offers a comprehensive approach to tackling the multifaceted nature of AD. These considerations and the imperative of safety assessments and robust clinical trials aid in developing or improving AD treatments. Further studies are required to understand the potential of the NF- $\kappa$ B pathway and its inhibitors in managing AD.

**Author Contributions:** Conceptualization, B.G.P., B.S.S. and C.C.; methodology, N.R., M.C. and B.S.S.; software, N.R., M.C. and S.B.; validation, N.R., M.C., S.B. and B.S.S.; investigation, N.R., M.C., S.B. and B.S.S.; resources, C.C.; data curation M.C., G.M.E. and S.B.; writing—original draft preparation, B.S.S., N.R., M.C. and S.B.; writing—review and editing, B.G.P., B.S.S. and C.C.; supervision, B.G.P.

and C.C.; project administration, B.G.P. and C.C.; funding acquisition, C.C. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by Fundamental Fund Research, Chiang Mai University, Chiang Mai, Thailand.

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** No new data were created or analyzed in this study. Data sharing does not apply to this article.

**Acknowledgments:** The authors gratefully acknowledge Chiang Mai University, Chiang Mai, for its support.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Tahami Monfared, A.A.; Byrnes, M.J.; White, L.A.; Zhang, Q. Alzheimer's Disease: Epidemiology and Clinical Progression. *Neurol. Ther.* **2022**, *11*, 553–569. [[CrossRef](#)] [[PubMed](#)]
2. Nelson, P.T.; Brayne, C.; Flanagan, M.E.; Abner, E.L.; Agrawal, S.; Attems, J.; Castellani, R.J.; Corrada, M.M.; Cykowski, M.D.; Di, J.; et al. Frequency of LATE neuropathologic change across the spectrum of Alzheimer's disease neuropathology: Combined data from 13 community-based or population-based autopsy cohorts. *Acta Neuropathol.* **2022**, *144*, 27–44. [[CrossRef](#)] [[PubMed](#)]
3. Uddin, M.S.; Kabir, M.T.; Jalouli, M.; Rahman, M.A.; Jeandet, P.; Behl, T.; Alexiou, A.; Albadrani, G.M.; Abdel-Daim, M.M.; Perveen, A.; et al. Neuroinflammatory Signaling in the Pathogenesis of Alzheimer's Disease. *Curr. Neuropharmacol.* **2022**, *20*, 126–146. [[CrossRef](#)] [[PubMed](#)]
4. Hong, S.; Beja-Glasser, V.F.; Nfonoyim, B.M.; Frouin, A.; Li, S.; Ramakrishnan, S.; Merry, K.M.; Shi, Q.; Rosenthal, A.; Barres, B.A.; et al. Complement and microglia mediate early synapse loss in Alzheimer mouse models. *Science* **2016**, *352*, 712–716. [[CrossRef](#)] [[PubMed](#)]
5. Czirr, E.; Castello, N.A.; Mosher, K.I.; Castellano, J.M.; Hinkson, I.V.; Lucin, K.M.; Baeza-Raja, B.; Ryu, J.K.; Li, L.; Farina, S.N.; et al. Microglial complement receptor 3 regulates brain A $\beta$  levels through secreted proteolytic activity. *J. Exp. Med.* **2017**, *214*, 1081–1092. [[CrossRef](#)]
6. Deolankar, S.C.; Patil, A.H.; Koyangana, S.G.; Subbannayya, Y.; Prasad, T.S.K.; Modi, P.K. Dissecting Alzheimer's Disease Molecular Substrates by Proteomics and Discovery of Novel Post-translational Modifications. *OMICS* **2019**, *23*, 350–361. [[CrossRef](#)]
7. Fathi, N.; Mojtahedi, H.; Nasiri, M.; Abolhassani, H.; Yousefpour Marzbali, M.; Esmaeili, M.; Salami, F.; Biglari, F.; Rezaei, N. How do nuclear factor kappa B (NF- $\kappa$ B)1 and NF- $\kappa$ B2 defects lead to the incidence of clinical and immunological manifestations of inborn errors of immunity? *Expert Rev. Clin. Immunol.* **2023**, *19*, 329–339. [[CrossRef](#)]
8. Pan, W.; Deng, L.; Wang, H.; Wang, V.Y.F. Atypical I $\kappa$ B Bcl3 enhances the generation of the NF- $\kappa$ B p52 homodimer. *Front. Cell Dev. Biol.* **2022**, *10*, 930619. [[CrossRef](#)]
9. Yang, G.; Hu, Y.; Qin, X.; Sun, J.; Miao, Z.; Wang, L.; Ke, Z.; Zheng, Y. Micheliolide attenuates neuroinflammation to improve cognitive impairment of Alzheimer's disease by inhibiting NF- $\kappa$ B and PI3K/Akt signaling pathways. *Heliyon* **2023**, *9*, e17848. [[CrossRef](#)]
10. Molina-Salinas, G.; Rodríguez-Chávez, V.; Langley, E.; Cerbon, M. Prolactin-induced neuroprotection against excitotoxicity is mediated via PI3K/AKT and GSK3 $\beta$ /NF- $\kappa$ B in primary cultures of hippocampal neurons. *Peptides* **2023**, *166*, 171037. [[CrossRef](#)]
11. Sato, S.; Sanjo, H.; Takeda, K.; Ninomiya-Tsuji, J.; Yamamoto, M.; Kawai, T.; Matsumoto, K.; Takeuchi, O.; Akira, S. Essential function for the kinase TAK1 in innate and adaptive immune responses. *Nat. Immunol.* **2005**, *6*, 1087–1095. [[CrossRef](#)] [[PubMed](#)]
12. Zhu, H.; Bai, Y.; Wang, G.; Su, Y.; Tao, Y.; Wang, L.; Yang, L.; Wu, H.; Huang, F.; Shi, H.; et al. Hyodeoxycholic acid inhibits lipopolysaccharide-induced microglia inflammatory responses through regulating TGR5/AKT/NF- $\kappa$ B signaling pathway. *J. Psychopharmacol.* **2022**, *36*, 849–859. [[CrossRef](#)] [[PubMed](#)]
13. Yang, S.; Magnutzki, A.; Alami, N.O.; Lattke, M.; Hein, T.M.; Scheller, J.S.; Kröger, C.; Oswald, F.; Yilmazer-Hanke, D.; Wirth, T.; et al. IKK2/NF- $\kappa$ B Activation in Astrocytes Reduces amyloid  $\beta$  Deposition: A Process Associated with Specific Microglia Polarization. *Cells* **2021**, *10*, 2669. [[CrossRef](#)]
14. Hayden, M.S.; West, A.P.; Ghosh, S. NF- $\kappa$ B and the immune response. *Oncogene* **2006**, *25*, 6758–6780. [[CrossRef](#)] [[PubMed](#)]
15. Singh, S.; Singh, T.G. Role of Nuclear Factor Kappa B (NF- $\kappa$ B) Signalling in Neurodegenerative Diseases: An Mechanistic Approach. *Curr. Neuropharmacol.* **2020**, *18*, 918–935. [[CrossRef](#)] [[PubMed](#)]
16. Elzayat, E.M.; Shahien, S.A.; El-Sherif, A.A.; Hosney, M. Therapeutic potential of stem cells and acitretin on inflammatory signaling pathway-associated genes regulated by miRNAs 146a and 155 in AD-like rats. *Sci. Rep.* **2023**, *13*, 9613. [[CrossRef](#)] [[PubMed](#)]
17. Korkmaz, A.; Reiter, R.J.; Topal, T.; Manchester, L.C.; Oter, S.; Tan, D.X. Melatonin: An established antioxidant worthy of use in clinical trials. *Mol. Med.* **2009**, *15*, 43–50. [[CrossRef](#)]

18. Merlo, S.; Caruso, G.I.; Korde, D.S.; Khodorovska, A.; Humpel, C.; Sortino, M.A. Melatonin Activates Anti-Inflammatory Features in Microglia in a Multicellular Context: Evidence from Organotypic Brain Slices and HMC3 Cells. *Biomolecules* **2023**, *13*, 373. [[CrossRef](#)]
19. Chowdari Gurram, P.; Satarker, S.; Kumar, G.; Begum, F.; Mehta, C.; Nayak, U.; Mudgal, J.; Arora, D.; Nampoothiri, M. Avanafil mediated dual inhibition of IKK $\beta$  and TNFR1 in an experimental paradigm of Alzheimer's disease: In silico and in vivo approach. *J. Biomol. Struct. Dyn.* **2022**, 1–19. [[CrossRef](#)]
20. Cook, M.; Lin, H.; Mishra, S.K.; Wang, G.Y. BAY 11-7082 inhibits the secretion of interleukin-6 by senescent human microglia. *Biochem. Biophys. Res. Commun.* **2022**, *617*, 30–35. [[CrossRef](#)]
21. Choi, S.; Kim, J.H.; Roh, E.J.; Ko, M.J.; Jung, J.E.; Kim, H.J. Nuclear factor-kappaB activated by capacitative Ca<sup>2+</sup> entry enhances muscarinic receptor-mediated soluble amyloid precursor protein (sAPP $\alpha$ ) release in SH-SY5Y cells. *J. Biol. Chem.* **2006**, *281*, 12722–12728. [[CrossRef](#)] [[PubMed](#)]
22. Lin, Y.Z.; Yao, S.; Veach, R.A.; Torgerson, T.R.; Hawiger, J. Inhibition of Nuclear Translocation of Transcription Factor NF-kappa B by a Synthetic Peptide Containing a Cell Membrane-permeable Motif and Nuclear Localization Sequence. *J. Biol. Chem.* **1995**, *270*, 14255–14258. [[CrossRef](#)] [[PubMed](#)]
23. Pogue, A.I.; Jaber, V.R.; Sharfman, N.M.; Zhao, Y.; Lukiw, W.J. Downregulation of Neurofilament Light Chain Expression in Human Neuronal-Glial Cell Co-Cultures by a Microbiome-Derived Lipopolysaccharide-Induced miRNA-30b-5p. *Front. Neurol.* **2022**, *13*, 900048. [[CrossRef](#)] [[PubMed](#)]
24. El-Sahar, A.E.; Shiha, N.A.; El Sayed, N.S.; Ahmed, L.A. Alogliptin Attenuates Lipopolysaccharide-Induced Neuroinflammation in Mice Through Modulation of TLR4/MYD88/NF- $\kappa$ B and miRNA-155/SOCS-1 Signaling Pathways. *Int. J. Neuropsychopharmacol.* **2020**, *24*, 158–169. [[CrossRef](#)]
25. Sarnico, I.; Boroni, F.; Benarese, M.; Alghisi, M.; Valerio, A.; Battistin, L.; Spano, P.; Pizzi, M. Targeting IKK2 by pharmacological inhibitor AS602868 prevents excitotoxic injury to neurons and oligodendrocytes. *J. Neural Transm.* **2008**, *115*, 693–701. [[CrossRef](#)]
26. Zhao, Y.; Bhattacharjee, S.; Jones, B.M.; Hill, J.; Dua, P.; Lukiw, W.J. Regulation of Neurotrophic Signaling by the Inducible, NF- $\kappa$ B-Sensitive miRNA-125b in Alzheimer's Disease (AD) and in Primary Human Neuronal-Glial (HNG) Cells. *Mol. Neurobiol.* **2014**, *50*, 97–106. [[CrossRef](#)]
27. Clark, I. Basic Scientific Evidence Consistent with Etanercept Efficacy Against Alzheimer's Disease. *Neuroscience* **2022**, *484*, 139. [[CrossRef](#)]
28. Butchart, J.; Brook, L.; Hopkins, V.; Teeling, J.; Püntener, U.; Culliford, D.; Sharples, R.; Sharif, S.; McFarlane, B.; Raybould, R.; et al. Etanercept in Alzheimer disease: A randomized, placebo-controlled, double-blind, phase 2 trial. *Neurology* **2015**, *84*, 2161–2168. [[CrossRef](#)]
29. Kong, F.; Jiang, X.; Wang, R.; Zhai, S.; Zhang, Y.; Wang, D. Forsythoside B attenuates memory impairment and neuroinflammation via inhibition on NF- $\kappa$ B signaling in Alzheimer's disease. *J. Neuroinflamm.* **2020**, *17*, 305. [[CrossRef](#)]
30. Ferretti, M.T.; Allard, S.; Partridge, V.; Ducatenzeiler, A.; Cuello, A.C. Minocycline corrects early, pre-plaque neuroinflammation and inhibits BACE-1 in a transgenic model of Alzheimer's disease-like amyloid pathology. *J. Neuroinflamm.* **2012**, *9*, 62. [[CrossRef](#)]
31. Cheng, S.; Hou, J.; Zhang, C.; Xu, C.; Wang, L.; Zou, X.; Yu, H.; Shi, Y.; Yin, Z.; Chen, G. Minocycline reduces neuroinflammation but does not ameliorate neuron loss in a mouse model of neurodegeneration. *Sci. Rep.* **2015**, *5*, 10535. [[CrossRef](#)] [[PubMed](#)]
32. Alhowail, A.; Alsikhan, R.; Alsaud, M.; Aldubayan, M.; Rabbani, S.I. Protective Effects of Pioglitazone on Cognitive Impairment and the Underlying Mechanisms: A Review of Literature. *Drug Des. Devel. Ther.* **2022**, *16*, 2919–2931. [[CrossRef](#)]
33. da Rocha, G.H.; Loiola, R.A.; de Paula-Silva, M.; Shimizu, F.; Kanda, T.; Vieira, A.; Gosselet, F.; Farsky, S.H.P. Pioglitazone Attenuates the Effects of Peripheral Inflammation in a Human In Vitro Blood-Brain Barrier Model. *Int. J. Mol. Sci.* **2022**, *23*, 12781. [[CrossRef](#)] [[PubMed](#)]
34. Youssef, M.; Ibrahim, A.; Akashi, K.; Hossain, M.S. PUFA-Plasmalogens Attenuate the LPS-Induced Nitric Oxide Production by Inhibiting the NF- $\kappa$ B, p38 MAPK and JNK Pathways in Microglial Cells. *Neuroscience* **2019**, *397*, 18–30. [[CrossRef](#)]
35. Wang, H.Y.; Pei, Z.; Lee, K.C.; Nikolov, B.; Doehner, T.; Puente, J.; Friedmann, N.; Burns, L.H. Simufilam suppresses overactive mTOR and restores its sensitivity to insulin in Alzheimer's disease patient lymphocytes. *Front. Aging* **2023**, *4*, 1175601. [[CrossRef](#)]
36. Kurata, T.; Lukic, V.; Kozuki, M.; Wada, D.; Miyazaki, K.; Morimoto, N.; Ohta, Y.; Deguchi, K.; Yamashita, T.; Hishikawa, N.; et al. Long-term effect of telmisartan on Alzheimer's amyloid genesis in SHR-SR after tMCAO. *Transl. Stroke Res.* **2015**, *6*, 107–115. [[CrossRef](#)]
37. Kishi, T.; Hirooka, Y.; Sunagawa, K. Telmisartan protects against cognitive decline via up-regulation of brain-derived neurotrophic factor/tropomyosin-related kinase B in hippocampus of hypertensive rats. *J. Cardiol.* **2012**, *60*, 489–494. [[CrossRef](#)]
38. Pang, T.; Wang, J.; Benicky, J.; Sánchez-Lemus, E.; Saavedra, J.M. Telmisartan directly ameliorates the neuronal inflammatory response to IL-1 $\beta$  partly through the JNK/c-Jun and NADPH oxidase pathways. *J. Neuroinflamm.* **2012**, *9*, 102. [[CrossRef](#)]
39. Wang, C.; Fan, L.; Khawaja, R.R.; Liu, B.; Zhan, L.; Kodama, L.; Chin, M.; Li, Y.; Le, D.; Zhou, Y.; et al. Microglial NF- $\kappa$ B drives tau spreading and toxicity in a mouse model of tauopathy. *Nat. Commun.* **2022**, *13*, 1969. [[CrossRef](#)]
40. Alam, J.J. Selective Brain-Targeted Antagonism of p38 MAPK $\alpha$  Reduces Hippocampal IL-1 $\beta$  Levels and Improves Morris Water Maze Performance in Aged Rats. *J. Alzheimer's Dis.* **2015**, *48*, 219–227. [[CrossRef](#)]
41. Shehata, M.K.; Ismail, A.A.; Kamel, M.A. Combined Donepezil with Astaxanthin via Nanostructured Lipid Carriers Effective Delivery to Brain for Alzheimer's Disease in Rat Model. *Int. J. Nanomed.* **2023**, *18*, 4193–4227. [[CrossRef](#)] [[PubMed](#)]

42. Park, J.; Min, J.S.; Kim, B.; Chae, U.B.; Yun, J.W.; Choi, M.S.; Kong, I.K.; Chang, K.T.; Lee, D.S. Mitochondrial ROS govern the LPS-induced pro-inflammatory response in microglia cells by regulating MAPK and NF- $\kappa$ B pathways. *Neurosci. Lett.* **2015**, *584*, 191–196. [[CrossRef](#)] [[PubMed](#)]
43. Salminen, A.; Lehtonen, M.; Suuronen, T.; Kaarniranta, K.; Huuskonen, J. Terpenoids: Natural inhibitors of NF- $\kappa$ B signaling with anti-inflammatory and anticancer potential. *Cell. Mol. Life Sci.* **2008**, *65*, 2979–2999. [[CrossRef](#)] [[PubMed](#)]
44. Yaron, A.; Gonen, H.; Alkalay, I.; Hatzubai, A.; Jung, S.; Beyth, S.; Mercurio, F.; Manning, A.M.; Ciechanover, A.; Ben-Neriah, Y. Inhibition of NF- $\kappa$ B cellular function via specific targeting of the I $\kappa$ B-ubiquitin ligase. *EMBO J.* **1997**, *16*, 6486–6494. [[CrossRef](#)] [[PubMed](#)]
45. Caccamo, D.; Campisi, A.; Marini, H.; Adamo, E.B.; Li Volti, G.; Squadrito, F.; Ientile, R. Glutamate promotes NF- $\kappa$ B pathway in primary astrocytes: Protective effects of IRFI 016, a synthetic vitamin E analogue. *Exp. Neurol.* **2005**, *193*, 377–383. [[CrossRef](#)]
46. Arumugam, T.V.; Baik, S.H.; Balaganopathy, P.; Sobey, C.G.; Mattson, M.P.; Jo, D.G. Notch signaling and neuronal death in stroke. *Prog. Neurobiol.* **2018**, *165–167*, 103–116. [[CrossRef](#)]
47. Mauro, C.; Leow, S.C.; Anso, E.; Rocha, S.; Thotakura, A.K.; Tornatore, L.; Moretti, M.; De Smaele, E.; Beg, A.A.; Tergaonkar, V.; et al. NF- $\kappa$ B controls energy homeostasis and metabolic adaptation by upregulating mitochondrial respiration. *Nat. Cell Biol.* **2011**, *13*, 1272–1279. [[CrossRef](#)]
48. Qiao, A.; Li, J.; Hu, Y.; Wang, J.; Zhao, Z. Reduction BACE1 expression via suppressing NF- $\kappa$ B mediated signaling by Tamibarotene in a mouse model of Alzheimer's disease. *IBRO Neurosci. Rep.* **2021**, *10*, 153–160. [[CrossRef](#)]
49. Bayraktar, A.; Li, X.; Kim, W.; Zhang, C.; Turkez, H.; Shoaie, S.; Mardinoglu, A. Drug repositioning targeting glutaminase reveals drug candidates for the treatment of Alzheimer's disease patients. *J. Transl. Med.* **2023**, *21*, 332. [[CrossRef](#)]
50. Hammond, S.L.; Bantle, C.M.; Popichak, K.A.; Wright, K.A.; Thompson, D.; Forero, C.; Kirkley, K.S.; Damale, P.U.; Chong, E.K.P.; Tjalkens, R.B. NF- $\kappa$ B Signaling in Astrocytes Modulates Brain Inflammation and Neuronal Injury Following Sequential Exposure to Manganese and MPTP During Development and Aging. *Toxicol. Sci.* **2020**, *177*, 506–520. [[CrossRef](#)]
51. Snow, W.M.; Albensi, B.C. Neuronal Gene Targets of NF- $\kappa$ B and Their Dysregulation in Alzheimer's Disease. *Front. Mol. Neurosci.* **2016**, *9*, 118. [[CrossRef](#)]
52. Srinivasan, M.; Lahiri, D.K. Significance of NF- $\kappa$ B as a pivotal therapeutic target in the neurodegenerative pathologies of Alzheimer's disease and multiple sclerosis. *Expert Opin. Ther. Targets* **2015**, *19*, 471–487. [[CrossRef](#)] [[PubMed](#)]
53. Uddin, M.S.; Rahman, M.A.; Kabir, M.T.; Behl, T.; Mathew, B.; Perveen, A.; Barreto, G.E.; Bin-Jumah, M.N.; Abdel-Daim, M.M.; Ashraf, G.M. Multifarious roles of mTOR signaling in cognitive aging and cerebrovascular dysfunction of Alzheimer's disease. *IUBMB Life* **2020**, *72*, 1843–1855. [[CrossRef](#)]
54. Kumar, A.; Singh, A.; Ekavali. A review on Alzheimer's disease pathophysiology and its management: An update. *Pharmacol. Rep.* **2015**, *67*, 195–203. [[CrossRef](#)]
55. Zhang, Z.; Yue, P.; Lu, T.; Wang, Y.; Wei, Y.; Wei, X. Role of lysosomes in physiological activities, diseases, and therapy. *J. Hematol. Oncol.* **2021**, *14*, 79. [[CrossRef](#)]
56. Piccialli, I.; Tedeschi, V.; Caputo, L.; D'Errico, S.; Ciccone, R.; De Feo, V.; Secondo, A.; Pannaccione, A. Exploring the Therapeutic Potential of Phytochemicals in Alzheimer's Disease: Focus on Polyphenols and Monoterpenes. *Front. Pharmacol.* **2022**, *13*, 876614. [[CrossRef](#)]
57. Ly, P.T.; Wu, Y.; Zou, H.; Wang, R.; Zhou, W.; Kinoshita, A.; Zhang, M.; Yang, Y.; Cai, F.; Woodgett, J.; et al. Inhibition of GSK3 $\beta$ -mediated BACE1 expression reduces Alzheimer-associated phenotypes. *J. Clin. Investig.* **2013**, *123*, 224–235. [[CrossRef](#)]
58. Payne, A.; Nahashon, S.; Taka, E.; Adinew, G.M.; Soliman, K.F.A. Epigallocatechin-3-Gallate (EGCG): New Therapeutic Perspectives for Neuroprotection, Aging, and Neuroinflammation for the Modern Age. *Biomolecules* **2022**, *12*, 371. [[CrossRef](#)]
59. Chong, C.M.; Su, H.; Lu, J.J.; Wang, Y. The effects of bioactive components from the rhizome of *Salvia miltiorrhiza* (Danshen) on the characteristics of Alzheimer's disease. *Chin. Med.* **2019**, *14*, 19. [[CrossRef](#)]
60. Halim, M.A.; Rosli, I.M.; Jaafar, S.S.M.; Ooi, H.; Leong, P.; Shamsuddin, S.; Najimudin, N.; Azzam, G. *Withania somnifera* showed neuroprotective effect and increase longevity in Drosophila Alzheimer's disease model. *bioRxiv* **2020**. [[CrossRef](#)]
61. Turner, R.S.; Thomas, R.G.; Craft, S.; van Dyck, C.H.; Mintzer, J.; Reynolds, B.A.; Brewer, J.B.; Rissman, R.A.; Raman, R.; Aisen, P.S.; et al. A randomized, double-blind, placebo-controlled trial of resveratrol for Alzheimer disease. *Neurology* **2015**, *85*, 1383–1391. [[CrossRef](#)] [[PubMed](#)]
62. Demarin, V.; Bašić Kes, V.; Trkanjec, Z.; Budišić, M.; Bošnjak Pašić, M.; Črnac, P.; Budinčević, H. Efficacy and safety of Ginkgo biloba standardized extract in the treatment of vascular cognitive impairment: A randomized, double-blind, placebo-controlled clinical trial. *Neuropsychiatr. Dis. Treat.* **2017**, *13*, 483–490. [[CrossRef](#)]
63. Xu, Y.M.; Lu, F.M.; Xu, H.C.; Zhang, J.; Hei, S.Y.; Qiu, Y.H.; Cai, Y.F.; Zhang, S.J.; Zhao, M. Kai-Xin-San Improves Cognitive Impairment via Wnt/ $\beta$ -Catenin and IRE1/XBP1s Signalings in APP/PS1 Mice. *Rejuvenation Res.* **2023**, *26*, 105–115. [[CrossRef](#)] [[PubMed](#)]
64. Ibrahim, W.W.; Skalicka-Woźniak, K.; Budzyńska, B.; El Sayed, N.S. NLRP3 inflammasome inhibition and M1-to-M2 microglial polarization shifting via scoparone-inhibited TLR4 axis in ovariectomy/D-galactose Alzheimer's disease rat model. *Int. Immunopharmacol.* **2023**, *119*, 110239. [[CrossRef](#)]
65. Song, Z.S.; Li, Z.; Wang, Y.; Li, M.X.; Liu, Q.; Shi, K.J.; Yao, X.W.; Ding, H.; Li, S.L.; Tang, W. Mechanisms of moxibustion preconditioning underlying improving learning-memory ability by regulating polarization of microglia via TLR4/NF- $\kappa$ B signaling pathway in AD rats. *Zhen Ci Yan Jiu* **2023**, *48*, 525–532.

66. Trang, N.M.; Kim, E.N.; Pham, T.H.; Jeong, G.S. Citropten Ameliorates Osteoclastogenesis Related to MAPK and PLC $\gamma$ /Ca<sup>2+</sup> Signaling Pathways through the Regulation of Amyloid Beta. *J. Agric. Food Chem.* **2023**, *71*, 10037–10049. [[CrossRef](#)] [[PubMed](#)]
67. Lu, W.; Chen, J.T.; Shi, Y.F.; Chen, M.S.; Wang, P.P.; Zhang, X.J.; Xiao, C.J.; Li, D.; Cao, C.Y.; Li, C.H.; et al. Diversified cassane family diterpenoids from the leaves of *Caesalpinia minax* exerting anti-neuroinflammatory activity through suppressing MAPK and NF- $\kappa$ B pathways in BV-2 microglia. *J. Ethnopharmacol.* **2023**, *315*, 116653. [[CrossRef](#)] [[PubMed](#)]
68. Zheng, L.; Wu, S.; Jin, H.; Wu, J.; Wang, X.; Cao, Y.; Zhou, Z.; Jiang, Y.; Li, L.; Yang, X.; et al. Molecular mechanisms and therapeutic potential of icariin in the treatment of Alzheimer's disease. *Phytomedicine* **2023**, *116*, 154890. [[CrossRef](#)]
69. Ju, Y.; Gu, L.; Hu, M.; Zheng, M.; Zhou, X.; Li, Q.; Zhang, X. Andrographolide exerts a neuroprotective effect by regulating the LRP1-mediated PPAR $\gamma$ /NF- $\kappa$ B pathway. *Eur. J. Pharmacol.* **2023**, *951*, 175756. [[CrossRef](#)]
70. Singh, L.; Kaur, N.; Bhatti, R. Neuroprotective potential of biochanin-A and review of the molecular mechanisms involved. *Mol. Biol. Rep.* **2023**, *50*, 5369–5378. [[CrossRef](#)]
71. Biradar, S.M.; Joshi, H.; Chheda, T.K. Biochanin-A ameliorates behavioural and neurochemical derangements in cognitive-deficit mice for the betterment of Alzheimer's disease. *Hum. Exp. Toxicol.* **2013**, *33*, 369–382. [[CrossRef](#)]
72. Bao, H.Y.; Chen, L.; Yang, Y.; Li, M.; Li, H.M.; Kang, Y.Y.; Li, J.G. Mechanism of *Berberis atropurpurea* anthocyanin against Alzheimer's disease based on network pharmacology and experimental verification. *Zhongguo Zhong Yao Za Zhi* **2023**, *48*, 778–788.
73. Huang, L.P.; Lu, L.H.; Yang, X.Y.; Xie, Y.Y.; Xu, Z.W.; Zhu, X.D.; Wang, J.J.; Wu, Z.X.; Tang, J.F.; Wu, Y.; et al. Effect of Erjing Pills on alleviating neuroinflammation of AD rats based on TLR4/NF- $\kappa$ B/NLRP3 pathway and its mechanism. *Zhongguo Zhong Yao Za Zhi* **2023**, *48*, 770–777. [[PubMed](#)]
74. Bhatia, H.S.; Apweiler, M.; Sun, L.; Baron, J.; Tirkey, A.; Fiebich, B.L. Licochalcone A Inhibits Prostaglandin E(2) by Targeting the MAPK Pathway in LPS Activated Primary Microglia. *Molecules* **2023**, *28*, 1927. [[CrossRef](#)]
75. Singh, L.; Singh, S. Neuroprotective potential of Honokiol in ICV-STZ induced neuroinflammation, A $\beta$  (1–42) and NF- $\kappa$ B expression in experimental model of rats. *Neurosci. Lett.* **2023**, *799*, 137090. [[CrossRef](#)]
76. Xu, Z.; Zhou, X.; Hong, X.; Wang, S.; Wei, J.; Huang, J.; Ji, L.; Yang, Y.; Efferth, T.; Hong, C.; et al. Essential oil of *Acorus tatarinowii* Schott inhibits neuroinflammation by suppressing NLRP3 inflammasome activation in 3  $\times$  Tg-AD transgenic mice. *Phytomedicine* **2023**, *112*, 154695. [[CrossRef](#)] [[PubMed](#)]
77. Barber, K.; Mendonca, P.; Soliman, K.F.A. The Neuroprotective Effects and Therapeutic Potential of the Chalcone Cardamomin for Alzheimer's Disease. *Brain Sci.* **2023**, *13*, 145. [[CrossRef](#)]
78. Chow, Y.L.; Lee, K.H.; Vidyadaran, S.; Lajis, N.H.; Akhtar, M.N.; Israf, D.A.; Syahida, A. Cardamomin from *Alpinia rafflesiana* inhibits inflammatory responses in IFN- $\gamma$ /LPS-stimulated BV2 microglia via NF- $\kappa$ B signalling pathway. *Int. Immunopharmacol.* **2012**, *12*, 657–665. [[CrossRef](#)] [[PubMed](#)]
79. Zhang, J.; Song, N.; Liu, Y.; Guo, J. Platycodin D Inhibits  $\beta$ -Amyloid-Induced Inflammation and Oxidative Stress in BV-2 Cells Via Suppressing TLR4/NF- $\kappa$ B Signaling Pathway and Activating Nrf2/HO-1 Signaling Pathway. *Neurochem. Res.* **2021**, *46*, 638–647. [[CrossRef](#)]
80. Li, B.; Wang, M.; Chen, S.; Li, M.; Zeng, J.; Wu, S.; Tu, Y.; Li, Y.; Zhang, R.; Huang, F.; et al. Baicalin Mitigates the Neuroinflammation through the TLR4/MyD88/NF- $\kappa$ B and MAPK Pathways in LPS-Stimulated BV-2 Microglia. *BioMed Res. Int.* **2022**, *2022*, 3263446. [[CrossRef](#)]
81. Xie, X.M.; Hao, J.J.; Shi, J.Z.; Zhou, Y.F.; Liu, P.F.; Wang, F.; Zheng, X.M.; Yu, X.Y.; Wang, C.C.; Yan, Y.; et al. Baicalein ameliorates Alzheimer's disease via orchestration of CX3CR1/NF- $\kappa$ B pathway in a triple transgenic mouse model. *Int. Immunopharmacol.* **2023**, *118*, 109994. [[CrossRef](#)] [[PubMed](#)]
82. Wei, P.C.; Lee-Chen, G.J.; Chen, C.M.; Chen, Y.; Lo, Y.S.; Chang, K.H. Isorhamnetin Attenuated the Release of Interleukin-6 from  $\beta$ -Amyloid-Activated Microglia and Mitigated Interleukin-6-Mediated Neurotoxicity. *Oxid. Med. Cell. Longev.* **2022**, *2022*, 3652402. [[CrossRef](#)] [[PubMed](#)]
83. Jayakumar, T.; Huang, C.J.; Yen, T.L.; Hsia, C.W.; Sheu, J.R.; Bhavan, P.S.; Huang, W.C.; Hsieh, C.Y.; Hsia, C.H. Activation of Nrf2 by Esculetin Mitigates Inflammatory Responses through Suppression of NF- $\kappa$ B Signaling Cascade in RAW 264.7 Cells. *Molecules* **2022**, *27*, 5143. [[CrossRef](#)] [[PubMed](#)]

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.