



Review

A Review of Fetal Development in Pregnancies with Maternal Type 2 Diabetes Mellitus (T2DM)-Associated Hypothalamic-Pituitary-Adrenal (HPA) Axis Dysregulation: Possible Links to Pregestational Prediabetes

Mathuli Ngema , Nombuso D. Xulu, Phikelelani S. Ngubane and Andile Khathi *

School of Laboratory Medicine & Medical Sciences, University of KwaZulu-Natal, Private Bag X54001, Durban 4001, South Africa; 218022309@stu.ukzn.ac.za (M.N.); 215019278@stu.ukzn.ac.za (N.D.X.); ngubanep1@ukzn.ac.za (P.S.N.)

* Correspondence: khathia@ukzn.ac.za; Tel.: +27-31-260-7585

Abstract: Research has identified fetal risk factors for adult diseases, forming the basis for the Developmental Origins of Health and Disease (DOHaD) hypothesis. DOHaD suggests that maternal insults during pregnancy cause structural and functional changes in fetal organs, increasing the risk of chronic diseases like type 2 diabetes mellitus (T2DM) in adulthood. It is proposed that altered maternal physiology, such as increased glucocorticoid (GC) levels associated with a dysregulated hypothalamic-pituitary-adrenal (HPA) axis in maternal stress and T2DM during pregnancy, exposes the fetus to excess GC. Prenatal glucocorticoid exposure reduces fetal growth and programs the fetal HPA axis, permanently altering its activity into adulthood. This programmed HPA axis is linked to increased risks of hypertension, cardiovascular diseases, and mental disorders in adulthood. With the global rise in T2DM, particularly among young adults of reproductive age, it is crucial to prevent its onset. T2DM is often preceded by a prediabetic state, a condition that does not show any symptoms, causing many to unknowingly progress to T2DM. Studying prediabetes is essential, as it is a reversible stage that may help prevent T2DM-related pregnancy complications. The existing literature focuses on HPA axis dysregulation in T2DM pregnancies and its link to fetal programming. However, the effects of prediabetes on HPA axis function, specifically glucocorticoid in pregnancy and fetal outcomes, are not well understood. This review consolidates research on T2DM during pregnancy, its impact on fetal programming via the HPA axis, and possible links with pregestational prediabetes.

Keywords: type 2 diabetes mellitus; prediabetes; pregnancy; maternal HPA axis; fetal HPA axis; programming; fetal development; placenta; glucocorticoids; metabolic diseases



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1. Introduction

Fetal programming occurs during embryonic and fetal development, a vital stage during which tissues and organs are formed [1–3]. Many environmental cues, such as excess glucocorticoid exposure in utero, can contribute to various changes that include changes in molecular biological functions, such as receptor cell density or sensitivity, as well as alterations in metabolism or responses to physiological stressors [4,5]. Essentially, fetal programming refers to the process of sustaining or affecting a stimulus or impairment that occurs at a crucial point in its development [6,7]. Studies show that maternal diabetes, particularly type 2 diabetes mellitus (T2DM), with increased glucocorticoid (GC) levels, may be one of the common mechanisms through which glucocorticoid insults exert their programming effects [8,9]. Rapid economic development and urbanization, sedentary lifestyles, and the Westernized diet have led to a rising burden of 463 million (aged 20–79 years) adults living with T2DM in many parts of the world, especially in developing countries [10]. Although the weights of infants of diabetic mothers are generally skewed

into the upper range, intrauterine growth restriction (IUGR), commonly diagnosed as low birth weight, occurs with concerning frequency in diabetic women, especially those with underlying hypertension, uncontrolled blood glucose levels, and vascular diseases [11,12]. T2DM has been shown to account for 30–50% of cases of pregestational diabetes during pregnancy [13].

Glucocorticoids (GC), such as cortisol in humans and corticosterone in rodents, are well-known for their role in glucose homeostasis in adult life [14]. The HPA axis regulates GC production through a feedback loop involving glucocorticoid receptors (GR) and mineralocorticoid receptors (MR) [14]. During pregnancy, this feedback mechanism ensures that GC levels are kept within a range that supports pregnancy while avoiding the adverse effects of hypercortisolism [15]. GC in pregnancy has also been shown to be essential in fetal maturation [5,16]. However, fetal GC load is usually regulated by 11 β -hydroxysteroid dehydrogenase type-2 (11 β -HSD2), a placental enzyme that inactivates GCs [17,18]. Increased maternal GC levels observed in T2DM and attenuating 11 β -HSD2 expression potentially increase fetal exposure to GCs, slowing fetal growth and altering the gestational period [19–21]. Excessive glucocorticoid exposure in utero goes as far as altering the set-point and development of the offspring's HPA axis that alternately reprograms the HPA axis, thus compromising its function after birth [22–24]. In addition, excess maternal or fetal corticosterone causes the downregulation of fetal GR and MR and impairs the feedback regulation of the HPA axis in both infancy and adulthood [25,26]. Cross-sectional research has also indicated a connection between lower birth weights, elevated cortisol levels, catch-up growth in the neonatal period, and adult obesity, which may be an indication of unfavorable adaptive responses until birth [27,28]. The association with low birth weight was first reported as a several-fold increase in the incidence of glucose intolerance and T2DM in adult men compared with those born with normal birth weight [29,30]. Furthermore, studies demonstrate that low birth weight has been associated with high risks of other non-communicable diseases (NCDs) such as hypertension, cardiovascular diseases, and mental disorders in adulthood, correlating with the concept of the Developmental Origins of Health and Disease (DOHaD) hypothesis [31,32]. The DOHaD, which emerged as a broadening of the "Barker hypothesis" and was named after epidemiologist David Barker, explains the scenario in which in utero maternal insults cause structural and functional alterations in fetal organs, extending postnatal life and increasing susceptibility to chronic disease in adulthood [33,34].

Prediabetes is characterized by impaired glucose metabolism, with glucose concentration above the optimal value but still below the diagnostic levels for T2DM [35]. In 2019, the prevalence of prediabetes was 373.9 million, with 15.3% undiagnosed, and it is expected to increase to 453.8 million by 2030 in parallel with increasing T2DM prevalence [36,37]. Studies show that prediabetes precedes T2DM, and it has been suggested that the onset of complications associated with T2DM begins during the prediabetic state, including myocardial injury, renal dysfunction, hormonal dysfunction, and dysregulation in the HPA axis function, among others [38–41]. The literature primarily reports alterations that occur in pre-existing T2DM pregnancies and fetal programming, while the changes in maternal pregestational prediabetes HPA axis function, specifically glucocorticoid and its influence on fetal outcomes, have not yet been explored. Therefore, this review consolidates research on T2DM during pregnancy, its impact on fetal programming via the HPA axis, and its possible links with pregestational prediabetes. The following section describes fetal programming, theories, and associated diseases.

2. Fetal Programming

According to Barker (1995), the early life environment affects fetal growth and adds to disease susceptibility [42,43]. The developing baby adapts to an insult in utero, leading to long-term changes in form, physiology, and metabolism that are beneficial for survival [33,34]. Gluckman discovered that mismatches between early and later life circumstances might cause maladaptive alterations that raise the risk of a variety of car-

diometabolic and psychiatric disorders as well as vulnerability factors, pertaining to the phenomena known as fetal programming [44,45]. Fetal programming occurs when the normal pattern of fetal development is disrupted by an abnormal stimulus or ‘insult’ applied at a critical point in in utero development [46–48]. According to the evidence for the Developmental Origins of Health and Disease (DOHaD) hypothesis, the antenatal period is a particularly vulnerable period of development in which exposure to adverse environments, such as glucocorticoid exposure, can have long-term or permanent effects on the offspring’s health trajectory [49,50]. Studies have shown that maternal HPA axis is crucial during fetal development [51,52]. However, maternal dysregulation in the HPA axis during pregnancy or before pregnancy has been shown to exert its programming effect, especially in the brain, notably in the HPA axis [53–55]. The following section details the physiological role of the maternal HPA axis in pregnancy, and its role in fetal development.

3. Role of the Maternal Hypothalamic–Pituitary–Adrenal (HPA) Axis in Pregnancy

The hypothalamic–pituitary–adrenal axis is a complex system of neuroendocrine pathways and feedback loops that functionally maintain physiological homeostasis through a synthesis of glucocorticoids (GCs). Active GCs are known as cortisol in humans and corticosterone in rodents [56]. The maternal HPA axis adapts during pregnancy, and regulates stress-related deleterious effects on the mother and offspring [57,58]. A non-diabetic pregnancy is a state of hyperactivity in the HPA axis and is also a state of hypercortisolism, especially towards late gestation [59–61]. The increased cortisol in late gestation is regulated by the placenta, an important source secreting the corticotropin-releasing hormone (CRH), which further enters the maternal pituitary gland via the hypophyseal portal circulation and enhances adrenocorticotropin (ACTH) synthesis and secretion into the peripheral circulation (Figure 1) [62,63]. ACTH increases glucocorticoid synthesis and secretion through the adrenal cortex in the kidney into the bloodstream in the course of pregnancy [64,65]. GC levels influence the hypothalamic CRH in a negative feedback loop, while the placental CRH is strongly stimulated by GC in a mechanism of a positive feedback loop [66,67]. In addition, studies show that high GC in pregnancy also plays a primary role in regulating fuel homeostasis. After the uptake of free cortisol from the circulation, cortisol increases the availability of potential fuel substrates by the mobilization of glucose, free fatty acids, and amino acids through the enhancement of hepatic gluconeogenesis and glycogenolysis [68,69]. Hence, research shows that GC contributes to insulin resistance, which is necessary to ensure that an adequate amount of glucose reaches the fetus for its growth and development [70].

Research shows that there are two types of corticosteroid receptors in the brain, namely glucocorticoid (GR) and mineralocorticoid (MR) receptors involved in the feedback regulation of the HPA axis [15,71]. The proper functioning of these receptors ensures that glucocorticoid levels remain within a range that supports pregnancy without causing undue stress to the mother or the fetus [72]. In the brain, MR binds to endogenous glucocorticoid with a higher affinity than GR, and, at basal concentrations of cortisol and corticosterone, MR is occupied while the GR remains largely unoccupied [73,74]. During times of elevated plasma glucocorticoid levels, such as during stress, increased occupation of GR helps to reduce the release of CRH and ACTH, ultimately lowering glucocorticoid production, and thereby regulating the function of the HPA axis. [75,76]. In pregnancy, MR mRNA expression in the hippocampus is unaltered, and GR gene expression is only modestly increased, which promotes negative feedback, maintaining HPA axis activity (Figure 1) and, hence, the diurnal secretion of cortisol is maintained throughout pregnancy [16,77]. Furthermore, studies show that late pregnancy (the last week, in the rat) is associated with a substantial reduction in HPA axis responses to both psychological and physical stressors in several species [77–79]. This adaptation is considered to buffer the impact of stress by reducing fetal exposure to maternal glucocorticoid, thus minimizing the risk of detrimental glucocorticoid programming [16,80].

Furthermore, studies show that glucocorticoids are lipophilic and can readily pass through the placental barrier by simple diffusion [81,82]. While glucocorticoid's most well-known function is to stimulate differentiation and functional development of the lungs, glucocorticoids also play crucial roles in the development of several other organ systems, including the HPA axis [51,52]. However, 11 β -hydroxysteroid dehydrogenase type 2 (11 β -HSD2) acts as a barrier enzyme to control the passage of glucocorticoids from the mother to the fetus and protects the fetus from the much higher maternal levels of glucocorticoids [83,84]. This enzyme is found on both placental sides of the syncytiotrophoblast [84,85]. It metabolizes active glucocorticoids (cortisol in humans, and corticosterone in rats) into inactive glucocorticoids, thereby shielding the fetus against excessive glucocorticoid exposure from the mother, as shown in (Figure 1) [85,86]. Although the placenta metabolizes a significant proportion of cortisol (80–90% during gestation), excess cortisol may reach the fetus, and the 'barrier' can be further weakened by maternal high maternal glucocorticoid or placental dysfunction, which is commonly caused by increased oxidative stress, resulting in hypoxia, allowing for the increased transfer of glucocorticoids from the mother to the fetus [16,54,87]. The following section describes T2DM, prevalences, pregestational consequences associated with fetal programming, and associated diseases in adulthood.

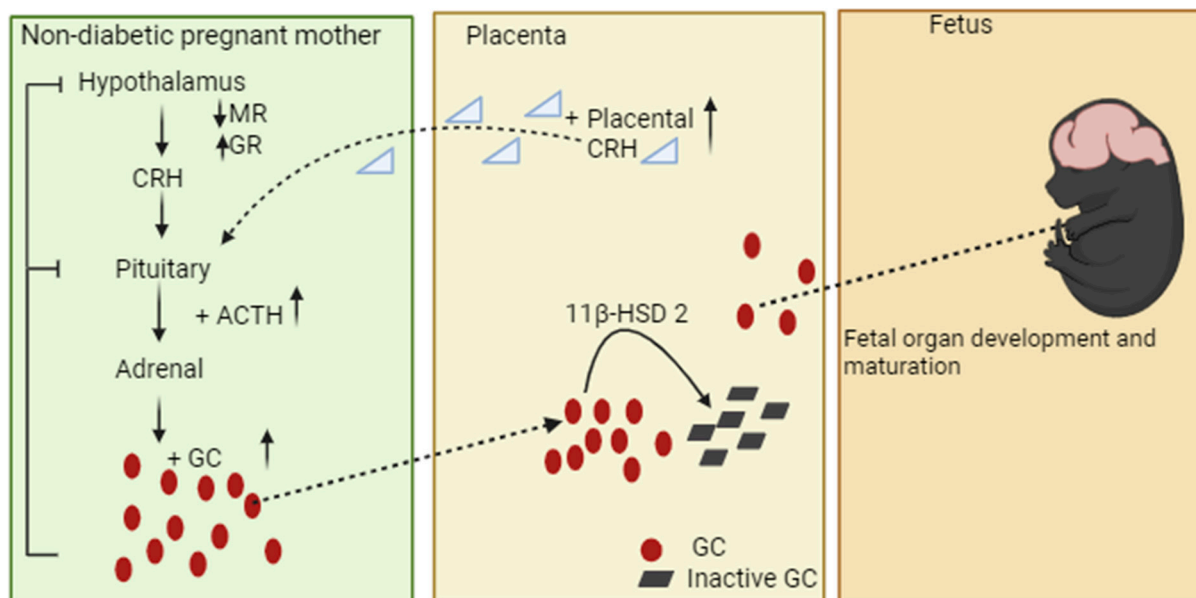


Figure 1. A schematic presentation of maternal HPA axis and GC signalling between mother, placenta, and fetus. Glucocorticoids tightly control HPA axis activity through glucocorticoid receptors (GR) and mineralocorticoid receptors (MR) in the pituitary and hypothalamus to inhibit CRH release, ACTH, and its own secretion [83,88]. In pregnancy, the placenta secretes large quantities of CRH into the maternal bloodstream as the pregnancy progresses, which promotes the production of GC [16,77]. Increased placental CRH secretion and GC also increase GR, promoting negative feedback and, therefore, maintaining the HPA axis activity in pregnancy. Nevertheless, the fetus is shielded from excess maternal GC exposure by the increased activity of 11 β -hydroxysteroid dehydrogenase type 2 (11 β -HSD2). The minimal transfer of GC from the placenta to the fetal compartment plays a vital role in the development of fetal organs, particularly the brain's HPA axis and the maturation of the lungs.

4. Changes in the HPA Axis in T2DM Pregnancies Associated with Fetal Programming

T2DM accounts for 90–95% of all diagnosed diabetes mellitus (DM) cases and is regarded as a complicated and multifaceted illness caused by a mix of genetic and environmental risk factors [89]. T2DM is characterized by insulin resistance and inadequate β -cell responsiveness to glucose stimulation [90]. Globally, the International Diabetes Federation (IDF) estimated that by 2045, 629 million are expected to have T2DM aged 20–70 years [91].

Swift urbanization, marked by the uptake of unhealthy, calorie-rich diets and sedentary lifestyles, has played a role in the progressively rising prevalence of T2DM, particularly among females compared to males, and the prevalence rises with age [92,93]. Research suggests that, while T2DM is often associated with macro- and microvascular complications, individuals with poor management of everyday stress is also associated, in diabetic patients, with the constant activation and disrupted regulation of the HPA axis, showing a similar resemblance to maternal stress, accompanied by high levels of glucocorticoids [94]. Champaneri et al. similarly found high cortisol (hypercortisolism) levels throughout the day in diabetic women [95]. Established diabetes mellitus, either type 1 or 2, is the most common pre-existing medical condition in pregnant women at younger ages, resulting in an increasing proportion of pregnancies complicated by diabetes [96–98]. In some areas, pregnant women with T2DM now outnumber those with type 1 diabetes (T1DM) [99]. Research indicates that pregnant women with T2DM exhibit comparable patterns to those observed in maternal obesity and in depressed and stressed pregnant women [100,101]. These patterns involve the prolonged activation and dysregulated function of the HPA axis with elevated glucocorticoid levels [100,101].

The pathophysiology of fetal growth in the context of T2DM pregnancy is intricate and multifaceted [102,103]. However, the complications of diabetes affecting the mother and fetus are well-known [104–106]. Maternal complications include preterm labour, nephropathy, vascular diseases, caesarean section, postoperative wound complications, uncontrolled hyperglycemia, and increased oxidative stress, among others [107]. Fetal complications include fetal wastage from early pregnancy loss or congenital anomalies, macrosomia, shoulder dystocia, stillbirth, and intrauterine growth restriction (IUGR), among others [108,109].

Approximately 20% of pregnant women with diabetes experience gestational hypertension and/or preeclampsia [110]. The individuals most susceptible to these conditions are those who have pre-existing microvascular complications such as microangiopathy, hypertension, or inadequate control of blood glucose levels, which also contribute to endothelial dysfunction [110,111]. These complications have been shown to induce a reduction in trophoblast proliferation, delaying placental growth and development, particularly in the first few weeks of gestation [112]. This mechanism suggests the presence of dysregulation of trophoblast invasion by the diabetic environment, leading to decreased placental perfusion, which results in placental dysfunction [113].

Studies show that placental dysfunction is associated with relatively low placental 11 β -HSD2 activity, therefore increasing active maternal GC to the fetus's bloodstream [114,115]. Overexposure to glucocorticoids during fetal development causes modifications in the expression of various cytostructural proteins, receptors, enzymes, ion channels, and growth factors [116]. These modifications result in changes in tissue structure, biochemical composition, metabolism, and hormone responsiveness, impacting the functionality of several fetal organ systems [117,118]. A study has discovered a correlation between maternal diabetes accompanied by elevated cortisol levels and alterations in the development of the brain's cortical neuroendocrine system by reducing the number of hippocampal neurons [119]. Nevertheless, the precise molecular and cellular process via which diabetes during pregnancy impacts brain development remains unknown [120]. Consequently, glucocorticoids trigger physiological processes that have little or insignificant roles in utero but which become crucial at birth, such as the HPA axis [121]. The HPA axis and its key limbic regulator, the hippocampus, are particularly sensitive to glucocorticoids and their perinatal programming actions [52,122]. Previous studies show that glucocorticoid excess exposure during fetal development programs has specific effects on the brain, notably the HPA axis [123,124]. This exposure changes its development, sensitivity, and activity in utero, relatively stressing its growth as the HPA axis begins to develop during the embryonic stage and continues to mature throughout pregnancy [25,125]. As a result, studies show that prenatal glucocorticoid exposure permanently increases basal plasma corticosterone levels in adult rats [122,126]. This was because the density of both types of corticosteroid

receptors, GRs and MRs, are permanently reduced in the hippocampus, changes which are anticipated to attenuate HPA axis feedback sensitivity from maternal stress shown in Figure 2 [122,126,127].

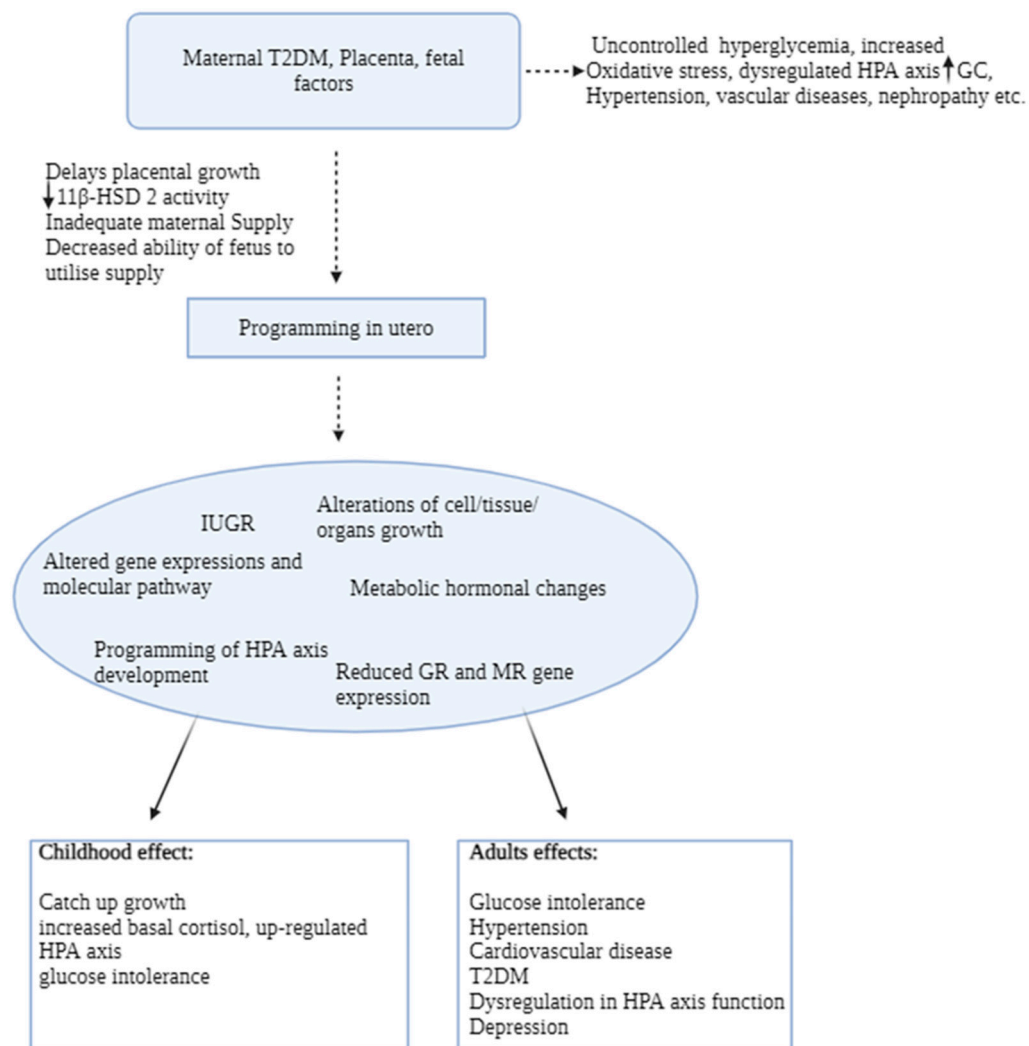


Figure 2. The schematic diagram presents the summary of maternal T2DM pregnancy complications, leading to fetal consequences in utero that persist until adulthood.

In addition, studies show that fetal exposure to excess maternal GC relative to early increases in the fetal GC concentration also triggers tissue differentiation and reduces accretion in the fetus [5]. As a result, the overall rate of maturation and growth declines as GC concentrations rise in the fetus toward term and in response to adverse intrauterine conditions, resulting in growth-retarded fetuses recognized as having IUGR [12,128]. The term intrauterine growth restriction (IUGR) refers to neonates whose birth weight and length fall below the tenth percentile for their gestational age [129,130]. IUGR is a common antenatal diagnosis; nevertheless, some of these fetuses, particularly those who were not checked during pregnancy, may be discovered only after birth [129,131]. The primary diagnostic criteria for IUGR include low birth weights (LWs), a surrogate marker of an adverse intrauterine environment, and subsequent cardio-metabolic disease and mental health problems [132,133].

On the other hand, the brain is heavily reliant on glucose for energy, and mammals have redundant systems for controlling glucose production [134]. As a result, it is possible that altered hypothalamus function may promote the dysregulation of peripheral glucose metabolism, leading to insulin resistance or T2DM in adulthood [135]. Research conducted

by Hales et al. revealed a several-fold higher incidence of glucose intolerance and T2DM in adult men who were born with low birth weight as opposed to those who were born with normal birth weight, which established the first link between low birth weight and the development of T2DM [29]. A study in rats showed that the smallest fetuses with the largest placentas had lower placental 11 β -HSD2 activity and were projected to have the highest adult blood pressures [17,136]. Heightened HPA axis activity, particularly with increased ACTH and high plasma glucocorticoid levels, is seen in children and adults who were born underweight [137,138]. In addition, previous research has indicated that infants born with lower birth weights undergo catch-up growth within the initial two years of life [139,140]. This process is seen as a means to offset their genetically predetermined growth patterns [140]. Catch-up growth is also observed in other aspects of growth, such as changes in body weight and body composition [141]. As per the theory of DOHaD, the rapid catch-up growth experienced by low-birth-weight infants in their early years is associated with various metabolic conditions such as obesity, hypertension, cardiovascular diseases, metabolic syndrome, and endothelial dysfunction later in adulthood shown in Figure 2 above [142,143]. Men were found to be more likely to develop cardiovascular disorders than women born with IUGR due to hormonal differences, as men had lower levels of estrogen, which has protective effects on the circulatory system and may contribute to women's decreased risk of cardiovascular disease [144,145]. A prior study discovered that a combination of placental weight and birth weight predicts the risk of high blood pressure and hypertension in men and women around the age of 50 [137,146]. People who were babies with large placentas had the highest blood pressure and a higher risk of hypertension [147]. Both the Barker and DOHaD hypotheses support these theories.

5. Prediabetes

Prediabetes is a condition in which blood glucose levels are abnormally high, but do not meet the diagnostic criteria for type 2 diabetes mellitus (T2DM) [148]. Prediabetes can be identified by at least two of these characteristics: impaired fasting glucose (IFG), impaired glucose tolerance (IGT), or high glycated hemoglobin A1c (HbA1c) [149]. IFG patients have significant hepatic insulin resistance with normal skeletal muscle values and poor glucose suppression, which causes hyperglycemia during fasting due to impaired insulin secretion or reduced sensitivity of β -cells to glucose stimulation [150]. IGT mainly impacts muscle insulin resistance, with minimal effects on liver insulin sensitivity. The reduced glucose absorption observed in individuals with impaired glucose tolerance (IGT) contributes to postprandial hyperglycemia [151,152]. This is primarily due to pancreatic β -cell dysfunction, resulting in inadequate secretion of insulin to counteract elevated glucose levels and stimulate a response in insulin-targeted peripheral tissues [153,154]. Lastly, IFG individuals have a poor early insulin response during the oral glucose tolerance test (OGTT), but improve insulin secretion during the second phase, which is one of the reasons why prediabetes is frequently undetectable [155]. As a result, the American Diabetes Association (ADA) recommendations were changed in 2003 to identify patients with prediabetes based on the following values. Fasting plasma glucose levels range from 5.6 mmol/L to 6.9 mmol/L, whereas IGT values recorded after OGTT range from 7.8 mmol/L to 11.0 mmol/L [156,157]. Glycated haemoglobin (HbA1c) levels between 5.7% and 6.4% are used as an additional diagnostic criterion for prediabetes [158,159]. Prior to the diagnosis of pre-diabetes, there is a presence of insulin resistance and malfunctioning of pancreatic β -cells [153,154]. Studies show that a diet high in saturated fats, high in carbohydrates, or high in fructose contributes to the development of intermediate hyperglycemia [160,161]. In addition, studies also show that these caloric foods lead to elevated triglycerides and increased release of free fatty acids (FFA) from adipocytes into circulation, which is accompanied by decreased FFA uptake by adipocytes in insulin-dependent tissues, promoting insulin resistance and dyslipidemia [162,163]. These actions result in increased circulating FFA levels and FFA flux to the liver, which stimulates increased production and secretion of atherogenic very-low-density lipoprotein and small dense low-density lipoprotein particles,

and reduced high-density lipoprotein cholesterol (HDL-C) levels, increasing the risk of microvascular and macrovascular diseases [162,164]. In addition, in the condition of insulin resistance, normal levels of insulin in the blood would be unable to elicit a reaction in the peripheral tissues that are targeted by insulin due to a decrease in the number of insulin receptors on the surface of cells, including muscle cells [165,166]. With fewer receptors available, the cells become less responsive to insulin, reducing their ability to take up glucose [90]. Consequently, the β -cells of the pancreas react by producing additional insulin to counterbalance the increased glucose levels [167]. When the β -cells fail to secrete sufficient insulin to counteract insulin resistance, the blood glucose levels commence fluctuating, leading to intermediate hyperglycemia and hyperinsulinemia, leading to further alterations in β -cell function [168].

5.1. Prevalence of Prediabetes

The prevalence of prediabetes has grown worldwide, and in 2019, the International Diabetes Federation estimated the worldwide prevalence of prediabetes to be 373.9 million, with 15.3% undiagnosed according to studies [36,37]. It is also projected that by 2030, approximately 453.8 million people will have prediabetes [37,169]. The prevalence of prediabetes is anticipated to increase to 8.3% of the global adult population, equivalent to an estimated 587 million individuals by the year 2045 [170]. Studies show that prediabetes is frequently undetected due to its often-asymptomatic nature in its early stages; hence, most humans tend to unknowingly bypass the prediabetes stage to overt T2DM [159,171]. In addition, studies show that the increase in prediabetes prevalence is due to rapid urbanization, increasingly sedentary lifestyles, and unhealthy eating habits [172,173]. As a result, a retrospective study in a rapidly urbanizing area such as Durban, South Africa, indicated that 68% of the individuals are prediabetic in the sample population between the ages of 20–45 years, with 51.0% of the study population being women [174]. This suggests that women of childbearing age are also affected by the global rise in prediabetes [175]. Furthermore, studies show that people with prediabetes have a two-fold increased likelihood of developing T2DM [176,177]. Moreover, studies show that complications associated with T2DM are already evident in some people with prediabetes; these complications include myocardial injury, renal dysfunction, hormonal dysfunction, and dysregulation in HPA axis function, among others [38–41].

5.2. HPA Axis Function in Prediabetes

Animal models have been observed to mirror human disease conditions, making them extensively utilizable for studying physiological systems and human disease states [178,179]. A high-fat, high carbohydrate and 15% fructose diet-induced animal model of prediabetes has been found to mimic the human condition [38]. In addition, this animal model showed dysregulation in the functioning of the HPA axis in the prediabetic state, as shown by elevated basal corticosterone and impaired regulation of their glucocorticoid receptor (GR) and mineralocorticoid receptor (MR) in male prediabetic rats [39]. At present, there have been no investigations to show whether this phenomenon also exists in female prediabetic rats. Furthermore, if present, this raised the question of whether maternal basal corticosterone and ACTH levels in prediabetic dams may impact fetal HPA axis development.

6. Possible Links of Prediabetes with T2DM Pregnancies in Association with the HPA Axis

Both T2DM and prediabetes result from insulin resistance, leading to impaired glucose tolerance and chronic hyperglycemia [154,180]. Several studies have shown that during pregnancy, increased maternal serum glucocorticoids (GC), observed in T2DM and maternal stress, may cross the placenta, overwhelming the protective placental barrier [11,30]. Conversely, research shows that pre-existing metabolic disorders associated with T2DM, such as hypertension, renal disease, and maternal microangiopathy during pregnancy, decrease trophoblast proliferation [110,111]. This delays placental growth and its ability

to supply the fetus with enough nutrients and oxygen, resulting in fetal hypoxia and inadequate nutrition supply [112]. Decreased placental function is linked to placental dysfunction and relatively low activity of placental 11 β -HSD2 [115,118]. These complications have been associated with increasing the vulnerability of the fetus during this period to unwanted programming effects such as increased transfer of active maternal cortisol to the fetal compartments [115,118]. Early maternal GC exposure to the fetus has been associated with alterations to the balance of both GR and MR development in the fetal brain, leading to changes in gene expression patterns and neural circuit formation, evidenced by low GR and MR expression in offspring after birth, even in both basal and stressed animals [126]. Moreover, studies show that excessive GC exposure during critical periods of development could program the fetal HPA axis to be hyper-responsive, beginning in utero and persisting later in life, leading to outcomes such as the prolonged increases in ACTH and GC seen in adulthood, potentially predisposing the offspring to diseases such as depression, cardiometabolic disorders, or T2DM [53]. Furthermore, studies suggest that excessive maternal GC exposure or reduced placental function during pregnancy is associated with reduced fetal growth, contributing to intrauterine growth restriction (IUGR), commonly diagnosed at birth as low birth weight [12,128]. Low-birth-weight offspring have been associated with HPA axis hyperactivity, glucose intolerance, hypertension, obesity, and greater risks for developing depression, anxiety, T2DM, and cardiovascular diseases in adulthood, especially when there was catch-up growth in the first 2 years, as supported by the DOHaD hypotheses [49,50].

There are various possible causes that contribute to IUGR or an unfavourable hostile environment during pregnancy, including preeclampsia and hypertensive disorders that have also been associated with greater risk in T2DM pregnancies [110]. Prediabetes, which often precedes the onset of T2DM, has been shown by various studies to be the genesis of complications associated with T2DM, including myocardial injury, renal dysfunction, hormonal dysfunction, and dysregulation in HPA axis function, among others [38–41]. A recent study by Ludidi and colleagues showed that prediabetes is a risk factor for developing pre-eclampsia, similar to T2DM pregnancy [181]. In addition, since prediabetes often goes undiagnosed, this suggests that there is a population of people unaware of their elevated risk of developing hypertension and preeclampsia [181]. Therefore, the presence of prediabetes in pregnancy might increase the likelihood of IUGR and impaired glucose tolerance in offspring. With the increasing prevalence of prediabetes, especially in women of childbearing age, there is an increased possibility of pre-gestational, gestational, and fetal outcome consequences. However, there have been no studies that have looked at how maternal prediabetes affects the HPA axis along with fetal outcomes (summarised in Table 1 below). Conducting preliminary investigations, generating hypotheses, carrying out invasive procedures, collecting tissue samples, and understanding the fundamental mechanism of pregnancy-related disorders would be ethically or practically challenging in humans. As a result, most primary studies are recommended to begin with animal models; hence, this study recommends that future studies focus on pregestational prediabetes, detailing its effects on maternal HPA axis function and its influences on fetal outcomes in Sprague Dawley rats.

Table 1. Summarizes possible links of prediabetes with T2DM pregnancies in association with the HPA axis.

| Prediabetes | T2DM |
|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|-------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| <ul style="list-style-type: none"> Prediabetes is characterized by the concurrent occurrence of insulin resistance and β-cell dysfunction, which are abnormalities that occur prior to the detection of changes in glucose levels [150,182]. | <ul style="list-style-type: none"> T2DM, a condition marked by deficient insulin secretion by pancreatic islet β-cells, tissue insulin resistance (IR), and an inadequate compensatory insulin secretory response [183]. |
| <ul style="list-style-type: none"> Studies show that young adults from 25–45 are diagnosed with prediabetes. The number of individuals with prediabetes is expected to rise to around 587 million by 2045 [170,174]. | <ul style="list-style-type: none"> It is projected that, by 2045, the number of people with type 2 diabetes mellitus (T2DM) will reach 629 million people aged 20–79 years, respectively [91]. |
| <ul style="list-style-type: none"> This study shows that a prediabetic diet-induced animal model showed dysregulation in the functioning of the HPA axis, associated with elevated basal corticosterone and impaired regulation of the glucocorticoid receptor (GR) and mineralocorticoid receptor (MR) in male prediabetic rats animals [39]. | <ul style="list-style-type: none"> Patients with diabetes present alterations of the HPA axis negative feedback, suggestive of the impairment of corticosteroid receptor sensitivity, which is associated with high levels of glucocorticoids and ACTH [94]. |
| <ul style="list-style-type: none"> Pre-eclampsia has been demonstrated to be associated with an increased risk in those with prediabetes [181]. Studies show that prediabetes is also linked to the early development of complications shown in T2DM. Therefore, the occurrence of prediabetes during pregnancy may increase the likelihood of exacerbated maternal dysregulation in HPA axis function, leading to fetal adverse consequences. | <ul style="list-style-type: none"> Research indicates that T2DM pregnancy with uncontrolled hyperglycemia, hypertension, elevated glucocorticoids, and an increased risk of pre-eclampsia is associated with higher rates of complications for both the mother and the baby [110,111]. The complications include IUGR, altered gene expression, HPA axis programming, glucose intolerance, increased risk of developing T2DM, and cardiovascular diseases, among others [67,137,184–186]. |

Nevertheless, no research has been conducted to investigate the impact of maternal prediabetes on the HPA axis in relation to fetal outcomes in pregnancy.

7. Conclusions

This review paper highlights the significant impact of the dysregulation of the maternal HPA axis during pregnancy, particularly elevated glucocorticoid levels, on fetal growth and programming, with potential implications for HPA axis development in the fetus seen in T2DM. It discusses the possible links between prediabetes and T2DM pregnancies relative to impaired HPA axis function. Women with type 2 diabetes represent high-risk groups during pregnancy. As the incidence of diagnosed diabetes and prediabetes continues to increase, especially at young ages, the number of women with diabetes or prediabetes in pregnancy may also continue to increase. However, further research is needed to understand the effects of pregestational prediabetes on the maternal HPA axis and its impact on fetal outcomes. This could further underscore the importance of a continued investigation into the complex interplay between maternal metabolic health, HPA axis regulation, and fetal development to inform clinical management and improve pregnancy outcomes.

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References

1. Lemley, C.O.; Littlejohn, B.P.; Burnett, D.D. Fetal programming. In *Bovine Reproduction*; John Wiley & Sons, Inc.: Hoboken, NJ, USA, 2021; pp. 339–346.
2. Öztürk, H.N.O.; Türker, P.F. Fetal programming: Could intrauterine life affect health status in adulthood? *Obstet. Gynecol. Sci.* **2021**, *64*, 473–483. [[CrossRef](#)] [[PubMed](#)]
3. Seneviratne, S.N.; Rajindrajith, S. Fetal programming of obesity and type 2 diabetes. *World J. Diabetes* **2022**, *13*, 482. [[CrossRef](#)] [[PubMed](#)]
4. Hong, J.Y. Developmental Programming by Perinatal Glucocorticoids. *Mol. Cells* **2022**, *45*, 685. [[CrossRef](#)] [[PubMed](#)]
5. Fowden, A.L.; Vaughan, O.R.; Murray, A.J.; Forhead, A.J. Metabolic consequences of glucocorticoid exposure before birth. *Nutrients* **2022**, *14*, 2304. [[CrossRef](#)] [[PubMed](#)]
6. Piro, E.; Serra, G.; Schierz, I.A.M.; Giuffrè, M.; Corsello, G. Fetal growth restriction: A growth pattern with fetal, neonatal and long-term consequences. *Euromediterr. Biomed. J.* **2019**, *14*, 038–044.
7. Raja, G.L.; Subhashree, K.D.; Kantayya, K.E. In utero exposure to endocrine disruptors and developmental neurotoxicity: Implications for behavioural and neurological disorders in adult life. *Environ. Res.* **2022**, *203*, 111829. [[CrossRef](#)] [[PubMed](#)]
8. Bashir, M.; Dabbous, Z.; Baagar, K.; Elkhatib, F.; Ibrahim, A.; Brich, S.-A.; Abdel-Rahman, M.E.; Konje, J.C.; Abou-Samra, A.-B. Type 2 diabetes mellitus in pregnancy: The impact of maternal weight and early glycaemic control on outcomes. *Eur. J. Obstet. Gynecol. Reprod. Biol.* **2019**, *233*, 53–57. [[CrossRef](#)] [[PubMed](#)]
9. Vickers, M.H. Early life nutrition and its effect on the development of obesity and type-2 diabetes. In *Early Nutrition and Long-Term Health*; Elsevier: Amsterdam, The Netherlands, 2022; pp. 281–307.
10. Aschner, P.; Karuranga, S.; James, S.; Simmons, D.; Basit, A.; Shaw, J.E.; Wild, S.H.; Ogurtsova, K.; Saeedi, P. The International Diabetes Federation's guide for diabetes epidemiological studies. *Diabetes Res. Clin. Pract.* **2021**, *172*, 108630. [[CrossRef](#)]
11. Cleary, E.M.; Thung, S.F.; Buschur, E.O. Pregestational Diabetes Mellitus. In *Endotext [Internet]*; MDText.com, Inc.: South Dartmouth, MA, USA, 2021.
12. Bedell, S.; Hutson, J.; de Vrijer, B.; Eastabrook, G. Effects of maternal obesity and gestational diabetes mellitus on the placenta: Current knowledge and targets for therapeutic interventions. *Curr. Vasc. Pharmacol.* **2021**, *19*, 176–192. [[CrossRef](#)]
13. Raets, L.; Ingelbrecht, A.; Benhalima, K. Management of type 2 diabetes in pregnancy: A narrative review. *Front. Endocrinol.* **2023**, *14*, 1193271. [[CrossRef](#)]
14. Bhaumik, S.; Lockett, J.; Cuffe, J.; Clifton, V.L. Glucocorticoids and Their Receptor Isoforms: Roles in Female Reproduction, Pregnancy, and Foetal Development. *Biology* **2023**, *12*, 1104. [[CrossRef](#)] [[PubMed](#)]
15. Pofi, R.; Tomlinson, J.W. Glucocorticoids in pregnancy. *Obstet. Med.* **2020**, *13*, 62–69. [[CrossRef](#)] [[PubMed](#)]
16. Sheng, J.A.; Bales, N.J.; Myers, S.A.; Bautista, A.L.; Roueifar, M.; Hale, T.M.; Handa, R.J. The hypothalamic-pituitary-adrenal axis: Development, programming actions of hormones, and maternal-fetal interactions. *Front. Behav. Neurosci.* **2021**, *14*, 256. [[CrossRef](#)] [[PubMed](#)]
17. Yu, P.; Zhou, J.; Ge, C.; Fang, M.; Zhang, Y.; Wang, H. Differential expression of placental 11 β -HSD2 induced by high maternal glucocorticoid exposure mediates sex differences in placental and fetal development. *Sci. Total Environ.* **2022**, *827*, 154396. [[CrossRef](#)] [[PubMed](#)]
18. Rensel, M.A.; Schlinger, B.A. 11 β hydroxysteroid dehydrogenases regulate circulating glucocorticoids but not central gene expression. *Gen. Comp. Endocrinol.* **2021**, *305*, 113734. [[CrossRef](#)] [[PubMed](#)]
19. Chavey, A.; Kioon, M.-D.A.; Bailbé, D.; Movassat, J.; Portha, B. Maternal diabetes, programming of beta-cell disorders and intergenerational risk of type 2 diabetes. *Diabetes Metab.* **2014**, *40*, 323–330. [[CrossRef](#)] [[PubMed](#)]
20. Kadayifci, F.Z.; Haggard, S.; Jeon, S.; Ranard, K.; Tao, D.; Pan, Y.-X. Early-life programming of type 2 diabetes mellitus: Understanding the association between epigenetics/genetics and environmental factors. *Curr. Genom.* **2019**, *20*, 453–463. [[CrossRef](#)]
21. Kong, L.; Chen, X.; Gissler, M.; Lavebratt, C. Relationship of prenatal maternal obesity and diabetes to offspring neurodevelopmental and psychiatric disorders: A narrative review. *Int. J. Obes.* **2020**, *44*, 1981–2000. [[CrossRef](#)]
22. Reynolds, R.M. Glucocorticoid excess and the developmental origins of disease: Two decades of testing the hypothesis—2012 Curt Richter Award Winner. *Psychoneuroendocrinology* **2013**, *38*, 1–11. [[CrossRef](#)]
23. Seckl, J.R. Glucocorticoids and Fetal Programming; Necessary and Sufficient? In *Hormones, Intrauterine Health and Programming*; Springer: Cham, Switzerland, 2014; pp. 1–15.
24. Rizzo, H.E.; Escaname, E.N.; Alana, N.B.; Lavender, E.; Gelfond, J.; Fernandez, R.; Hibbs, M.A.; King, J.M.; Carr, N.R.; Blanco, C.L. Maternal diabetes and obesity influence the fetal epigenome in a largely Hispanic population. *Clin. Epigenet.* **2020**, *12*, 34. [[CrossRef](#)]
25. Cottrell, E.C.; Seckl, J. Prenatal stress, glucocorticoids and the programming of adult disease. *Front. Behav. Neurosci.* **2009**, *3*, 707. [[CrossRef](#)] [[PubMed](#)]
26. Miranda, A.; Sousa, N. Maternal hormonal milieu influence on fetal brain development. *Brain Behav.* **2018**, *8*, e00920. [[CrossRef](#)] [[PubMed](#)]
27. Grace, C.E.; Kim, S.J.; Rogers, J.M. Maternal influences on epigenetic programming of the developing hypothalamic-pituitary-adrenal axis. *Birth Defects Res. Part A Clin. Mol. Teratol.* **2011**, *91*, 797–805. [[CrossRef](#)] [[PubMed](#)]

28. de Mendonça, E.L.S.S.; de Lima Macêna, M.; Bueno, N.B.; de Oliveira, A.C.M.; Mello, C.S. Premature birth, low birth weight, small for gestational age and chronic non-communicable diseases in adult life: A systematic review with meta-analysis. *Early Hum. Dev.* **2020**, *149*, 105154. [[CrossRef](#)] [[PubMed](#)]
29. Hales, C.N.; Barker, D.J.; Clark, P.M.; Cox, L.J.; Fall, C.; Osmond, C.; Winter, P. Fetal and infant growth and impaired glucose tolerance at age 64. *BMJ Br. Med. J.* **1991**, *303*, 1019. [[CrossRef](#)] [[PubMed](#)]
30. Weiss, U.; Cervar, M.; Puerstner, P.; Schmut, O.; Haas, J.; Mauschitz, R.; Arikan, G.; Desoye, G. Hyperglycaemia in vitro alters the proliferation and mitochondrial activity of the choriocarcinoma cell lines BeWo, JAR and JEG-3 as models for human first-trimester trophoblast. *Diabetologia* **2001**, *44*, 209–219. [[CrossRef](#)] [[PubMed](#)]
31. Starikov, R.; Has, P.; Wu, R.; Nelson, D.M.; He, M. Small-for-gestational age placentas associate with an increased risk of adverse outcomes in pregnancies complicated by either type I or type II pre-gestational diabetes mellitus. *J. Matern.-Fetal Neonatal Med.* **2022**, *35*, 1677–1682. [[CrossRef](#)]
32. Baird, J.; Jacob, C.; Barker, M.; Fall, C.H.; Hanson, M.; Harvey, N.C.; Inskip, H.M.; Kumaran, K.; Cooper, C. Developmental origins of health and disease: A lifecourse approach to the prevention of non-communicable diseases. *Healthcare* **2017**, *5*, 14. [[CrossRef](#)] [[PubMed](#)]
33. Lampl, M.; Mummert, A.; Schoen, M. Auxological perspectives on ‘growth’ in DOHaD. *J. Dev. Orig. Health Dis.* **2015**, *6*, 390–398. [[CrossRef](#)]
34. Ventura, N.M. Developmental Origins of Cardiovascular Disease: Alterations in Gestational Hypertension and Stroke Outcome in Adult Offspring. Ph.D. Thesis, Queen’s University, Kingston, ON, Canada, 2015.
35. Baranowska-Jurkun, A.; Matuszewski, W.; Bandurska-Stankiewicz, E. Chronic microvascular complications in prediabetic states—An overview. *J. Clin. Med.* **2020**, *9*, 3289. [[CrossRef](#)]
36. Saeedi, P.; Petersohn, I.; Salpea, P.; Malanda, B.; Karuranga, S.; Unwin, N.; Colagiuri, S.; Guariguata, L.; Motala, A.A.; Ogurtsova, K. Global and regional diabetes prevalence estimates for 2019 and projections for 2030 and 2045: Results from the International Diabetes Federation Diabetes Atlas. *Diabetes Res. Clin. Pract.* **2019**, *157*, 107843. [[CrossRef](#)] [[PubMed](#)]
37. Sosibo, A.M.; Mzimela, N.C.; Ngubane, P.S.; Khathi, A. Prevalence and correlates of pre-diabetes in adults of mixed ethnicities in the South African population: A systematic review and meta-analysis. *PLoS ONE* **2022**, *17*, e0278347. [[CrossRef](#)] [[PubMed](#)]
38. Luvuno, M.; Mabandla, M.; Khathi, A. Voluntary ingestion of a high-fat high-carbohydrate diet: A model for prediabetes. *Ponte Int. Sci. Res. J.* **2018**, *74*, 119–143.
39. Mosili, P.; Mkhize, B.C.; Ngubane, P.; Sibiyi, N.; Khathi, A. The dysregulation of the hypothalamic–pituitary–adrenal axis in diet-induced prediabetic male Sprague Dawley rats. *Nutr. Metab.* **2020**, *17*, 104. [[CrossRef](#)] [[PubMed](#)]
40. Mkhize, B.C.; Mosili, P.; Ngubane, P.S.; Sibiyi, N.H.; Khathi, A. Diet-induced prediabetes: Effects on the activity of the renin–angiotensin–aldosterone system in selected organs. *J. Diabetes Investig.* **2022**, *13*, 768–780. [[CrossRef](#)] [[PubMed](#)]
41. Dimba, N.R.; Mzimela, N.; Mosili, P.; Ngubane, P.S.; Khathi, A. Investigating the association between diet-induced “leaky gut” and the development of prediabetes. *Exp. Clin. Endocrinol. Diabetes* **2023**, *131*, 569–576. [[CrossRef](#)] [[PubMed](#)]
42. Barker, D.J.; Clark, P.M. Fetal undernutrition and disease in later life. *Rev. Reprod.* **1997**, *2*, 105–112. [[CrossRef](#)] [[PubMed](#)]
43. Monahan, B.; Farland, L.V.; Shadyab, A.H.; Hankinson, S.E.; Manson, J.E.; Spracklen, C.N. Birthweight and subsequent risk for thyroid and autoimmune conditions in postmenopausal women. *J. Dev. Orig. Health Dis.* **2022**, *13*, 463–470. [[CrossRef](#)]
44. Gluckman, P.D.; Buklijas, T.; Hanson, M.A. The developmental origins of health and disease (DOHaD) concept: Past, present, and future. In *The Epigenome and Developmental Origins of Health and Disease*; Elsevier: Amsterdam, The Netherlands, 2016; pp. 1–15.
45. Poston, L.; Godfrey, K.; Gluckman, P.; Hanson, M. *Developmental Origins of Health and Disease*; Cambridge University Press: Cambridge, UK, 2022.
46. Lemley, C. Fetal programming: Maternal-fetal interactions and postnatal performance. *Clin. Theriogenology* **2020**, *12*, 252–267.
47. Cerritelli, F.; Frasca, M.G.; Antonelli, M.C.; Viglione, C.; Vecchi, S.; Chiera, M.; Manzotti, A. A review on the vagus nerve and autonomic nervous system during fetal development: Searching for critical windows. *Front. Neurosci.* **2021**, *15*, 721605. [[CrossRef](#)]
48. Gundacker, A.; Cuenca Rico, L.; Stoehrmann, P.; Tillmann, K.E.; Weber-Stadlbauer, U.; Pollak, D.D. Interaction of the pre-and postnatal environment in the maternal immune activation model. *Discov. Ment. Health* **2023**, *3*, 15. [[CrossRef](#)] [[PubMed](#)]
49. Wolford, E. Developmental Origins of Mental Health: Human Observational Studies of Preterm Birth, Antenatal Synthetic Glucocorticoid Exposure, and Maternal Depressive Symptoms during Pregnancy. Ph.D. Thesis, University of Helsinki, Helsinki, Finland, 2018.
50. Hoffman, D.J.; Powell, T.L.; Barrett, E.S.; Hardy, D.B. Developmental origins of metabolic diseases. *Physiol. Rev.* **2021**, *101*, 739–795. [[CrossRef](#)] [[PubMed](#)]
51. Morsi, A.; DeFranco, D.; Witchel, S.F. The hypothalamic-pituitary-adrenal axis and the fetus. *Horm. Res. Paediatr.* **2018**, *89*, 380–387. [[CrossRef](#)]
52. Drake, A.J.; Tang, J.I.; Nyirenda, M.J. Mechanisms underlying the role of glucocorticoids in the early life programming of adult disease. *Clin. Sci.* **2007**, *113*, 219–232. [[CrossRef](#)]
53. Braun, T.; Challis, J.R.; Newnham, J.P.; Sloboda, D.M. Early-life glucocorticoid exposure: The hypothalamic-pituitary-adrenal axis, placental function, and long-term disease risk. *Endocr. Rev.* **2013**, *34*, 885–916. [[CrossRef](#)] [[PubMed](#)]
54. Duthie, L.; Reynolds, R.M. Changes in the maternal hypothalamic-pituitary-adrenal axis in pregnancy and postpartum: Influences on maternal and fetal outcomes. *Neuroendocrinology* **2013**, *98*, 106–115. [[CrossRef](#)] [[PubMed](#)]

55. Xiong, F.; Zhang, L. Role of the hypothalamic–pituitary–adrenal axis in developmental programming of health and disease. *Front. Neuroendocrinol.* **2013**, *34*, 27–46. [[CrossRef](#)]
56. Matthews, S.G.; McGowan, P.O. Developmental programming of the HPA axis and related behaviours: Epigenetic mechanisms. *J. Endocrinol.* **2019**, *242*, T69–T79. [[CrossRef](#)]
57. Maniam, J.; Antoniadis, C.; Morris, M.J. Early-life stress, HPA axis adaptation, and mechanisms contributing to later health outcomes. *Front. Endocrinol.* **2014**, *5*, 73. [[CrossRef](#)]
58. Brunton, P.J. Effects of maternal exposure to social stress during pregnancy: Consequences for mother and offspring. *Reproduction* **2013**, *146*, R175–R189. [[CrossRef](#)]
59. Tal, R.; Taylor, H.S. Endocrinology of pregnancy. In *Endotext [Internet]*; MDText.com, Inc.: South Dartmouth, MA, USA, 2021.
60. O'Reilly, J.R. Effects of Maternal Stress and Obesity on Human Feto-Placental Glucocorticoid Exposure. Ph.D. Thesis, The University of Edinburgh, Edinburgh, UK, 2014.
61. Ying, S. Investigating the Mechanisms Mediating the Outcomes of Prenatal Stress. Ph.D. Thesis, The University of Edinburgh, Edinburgh, UK, 2020.
62. Erhuma, A.M. The interaction between maternal and fetal hypothalamic–pituitary–adrenal axes. In *Corticosteroids—A Paradigmatic Drug Class*; IntechOpen Limited: London, UK, 2021; Volume 6.
63. Valsamakis, G.; Chrousos, G.; Mastorakos, G. Stress, female reproduction and pregnancy. *Psychoneuroendocrinology* **2019**, *100*, 48–57. [[CrossRef](#)] [[PubMed](#)]
64. Opichka, M.A.; Livergood, M.C.; Grobe, J.L.; McIntosh, J.J. Cardiovascular Neuroendocrinology of Pregnancy. In *Cardiovascular Neuroendocrinology*; Springer: Cham, Switzerland, 2023; pp. 111–135.
65. Gonzalez-Iglesias, A.E.; Freeman, M.E. Brain control over pituitary gland hormones. In *Neuroscience in the 21st Century: From Basic to Clinical*; Springer: Cham, Switzerland, 2022; pp. 2291–2344.
66. Kota, S.K.; Gayatri, K.; Jammula, S.; Kota, S.K.; Krishna, S.V.; Meher, L.K.; Modi, K.D. Endocrinology of parturition. *Indian J. Endocrinol. Metab.* **2013**, *17*, 50–59. [[CrossRef](#)] [[PubMed](#)]
67. Gans, I.M.; Coffman, J.A. Glucocorticoid-mediated developmental programming of vertebrate stress responsivity. *Front. Physiol.* **2021**, *12*, 812195. [[CrossRef](#)] [[PubMed](#)]
68. Kuo, T.; McQueen, A.; Chen, T.-C.; Wang, J.-C. Regulation of glucose homeostasis by glucocorticoids. In *Glucocorticoid Signaling: From Molecules to Mice to Man*; Springer: New York, NY, USA, 2015; pp. 99–126.
69. Mourtzi, N.; Sertedaki, A.; Charmandari, E. Glucocorticoid signaling and epigenetic alterations in stress-related disorders. *Int. J. Mol. Sci.* **2021**, *22*, 5964. [[CrossRef](#)]
70. Grilo, L.F.; Tocantins, C.; Diniz, M.S.; Gomes, R.M.; Oliveira, P.J.; Matafome, P.; Pereira, S.P. Metabolic disease programming: From mitochondria to epigenetics, glucocorticoid signalling and beyond. *Eur. J. Clin. Investig.* **2021**, *51*, e13625. [[CrossRef](#)] [[PubMed](#)]
71. Künzel, R.G.; Elgazzar, M.; Bain, P.A.; Kirschbaum, C.; Papatheodorou, S.; Gelaye, B. The Association Between Maternal Prenatal Hair Cortisol Concentration and Preterm Birth: A Systematic Review and Meta-Analysis. *Psychoneuroendocrinology* **2024**, *165*, 107041. [[CrossRef](#)] [[PubMed](#)]
72. Solano, M.E.; Arck, P.C. Steroids, pregnancy and fetal development. *Front. Immunol.* **2020**, *10*, 477454. [[CrossRef](#)] [[PubMed](#)]
73. Groeneweg, F.L.; Karst, H.; de Kloet, E.R.; Joëls, M. Rapid non-genomic effects of corticosteroids through the membrane-associated MR and GR and their role in the central stress. *J. Endocrinol.* **2011**, *2*, 153–167. [[CrossRef](#)] [[PubMed](#)]
74. Vyas, S.; Maatouk, L. Contribution of glucocorticoids and glucocorticoid receptors to the regulation of neurodegenerative processes. *CNS Neurol. Disord. Drug Targets* **2013**, *12*, 1175–1193. [[CrossRef](#)]
75. de Kloet, E.R.; Schmidt, M.; Meijer, O.C. Corticosteroid receptors and HPA-axis regulation. In *Techniques in the Behavioral and Neural Sciences*; Elsevier: Amsterdam, The Netherlands, 2005; Volume 15, pp. 265–294.
76. de Kloet, E.R.; Joëls, M. Mineralocorticoid receptors and glucocorticoid receptors in HPA stress responses during coping and adaptation. In *Oxford Research Encyclopedia of Neuroscience*; Oxford University Press: Oxford, UK, 2020.
77. Brunton, P. Resetting the dynamic range of hypothalamic-pituitary-adrenal axis stress responses through pregnancy. *J. Neuroendocrinol.* **2010**, *22*, 1198–1213. [[CrossRef](#)]
78. Brunton, P.; Russell, J.; Douglas, A. Adaptive responses of the maternal hypothalamic-pituitary-adrenal axis during pregnancy and lactation. *J. Neuroendocrinol.* **2008**, *20*, 764–776. [[CrossRef](#)]
79. Azizi, N.; Heidari, M.; Saboory, E.; Abdollahzade, N.; Roshan-Milani, S. Investigating the effect of parental pre-gestational stress on ethological parameters in male rat offspring. *J. Vet. Behav.* **2024**, *73*, 31–40. [[CrossRef](#)]
80. Ruffaner-Hanson, C.; Noor, S.; Sun, M.S.; Solomon, E.; Marquez, L.E.; Rodriguez, D.E.; Allan, A.M.; Caldwell, K.K.; Bakhireva, L.N.; Milligan, E.D. The maternal-placental-fetal interface: Adaptations of the HPA axis and immune mediators following maternal stress and prenatal alcohol exposure. *Exp. Neurol.* **2022**, *355*, 114121. [[CrossRef](#)]
81. Stirrat, L.I.; Sengers, B.G.; Norman, J.E.; Homer, N.Z.; Andrew, R.; Lewis, R.M.; Reynolds, R.M. Transfer and metabolism of cortisol by the isolated perfused human placenta. *J. Clin. Endocrinol. Metab.* **2018**, *103*, 640–648. [[CrossRef](#)]
82. Fowden, A.L.; Forhead, A.J. Glucocorticoids as regulatory signals during intrauterine development. *Exp. Physiol.* **2015**, *100*, 1477–1487. [[CrossRef](#)]
83. Chapman, K.; Holmes, M.; Seckl, J. 11 β -hydroxysteroid dehydrogenases: Intracellular gate-keepers of tissue glucocorticoid action. *Physiol. Rev.* **2013**, *93*, 1139–1206. [[CrossRef](#)]

84. Zhu, P.; Wang, W.; Zuo, R.; Sun, K. Mechanisms for establishment of the placental glucocorticoid barrier, a guard for life. *Cell. Mol. Life Sci.* **2019**, *76*, 13–26. [[CrossRef](#)]
85. Shallie, P.D.; Naicker, T. The placenta as a window to the brain: A review on the role of placental markers in prenatal programming of neurodevelopment. *Int. J. Dev. Neurosci.* **2019**, *73*, 41–49. [[CrossRef](#)]
86. Waldorf, K.M.A. Maternal-fetal immunology. In *Obstetrics: Normal and Problem Pregnancies*; Elsevier: Amsterdam, The Netherlands, 2016; Volume 7, pp. 64–82.
87. Volqvartz, T.; Andersen, H.H.B.; Pedersen, L.H.; Larsen, A. Obesity in pregnancy—Long-term effects on offspring hypothalamic-pituitary-adrenal axis and associations with placental cortisol metabolism: A systematic review. *Eur. J. Neurosci.* **2023**, *58*, 4393–4422. [[CrossRef](#)]
88. Koorneef, L.L.; Viho, E.M.; Wahl, L.F.; Meijer, O.C. Do corticosteroid receptor mRNA levels predict the expression of their target genes? *J. Endocr. Soc.* **2023**, *7*, bvac188. [[CrossRef](#)]
89. Lima, J.E.; Moreira, N.C.; Sakamoto-Hojo, E.T. Mechanisms underlying the pathophysiology of type 2 diabetes: From risk factors to oxidative stress, metabolic dysfunction, and hyperglycemia. *Mutat. Res./Genet. Toxicol. Environ. Mutagen.* **2022**, *874*, 503437. [[CrossRef](#)]
90. Rachdaoui, N. Insulin: The friend and the foe in the development of type 2 diabetes mellitus. *Int. J. Mol. Sci.* **2020**, *21*, 1770. [[CrossRef](#)]
91. Cho, N.H.; Shaw, J.; Karuranga, S.; Huang, Y.; da Rocha Fernandes, J.; Ohlrogge, A.; Malanda, B. IDF Diabetes Atlas: Global estimates of diabetes prevalence for 2017 and projections for 2045. *Diabetes Res. Clin. Pract.* **2018**, *138*, 271–281. [[CrossRef](#)]
92. Goedecke, J.H.; Mtintsilana, A.; Dlamini, S.N.; Kengne, A.P. Type 2 diabetes mellitus in African women. *Diabetes Res. Clin. Pract.* **2017**, *123*, 87–96. [[CrossRef](#)]
93. Nanditha, A.; Chamukuttan, S.; Raghavan, A.; Ramachandran, A. Global Epidemic of Type 2 Diabetes Mellitus: An Epidemiologist's Perspective. In *Current Trends in Diabetes*; JP Medical Publishers: Ashland, OH, USA, 2020; p. 36.
94. Kivimäki, M.; Bartolomucci, A.; Kawachi, I. The multiple roles of life stress in metabolic disorders. *Nat. Rev. Endocrinol.* **2023**, *19*, 10–27. [[CrossRef](#)]
95. Champaneri, S.; Xu, X.; Carnethon, M.R.; Bertoni, A.G.; Seeman, T.; Roux, A.D.; Golden, S.H. Diurnal salivary cortisol and urinary catecholamines are associated with diabetes mellitus: The Multi-Ethnic Study of Atherosclerosis. *Metabolism* **2012**, *61*, 986–995. [[CrossRef](#)]
96. Kitzmiller, J.L.; Ferrara, A.; Peng, T.; Cissell, M.A.; Kim, C. Preexisting diabetes and pregnancy. In *Diabetes in America*, 3rd ed.; National Institutes of Health: Bethesda, MD, USA, 2018.
97. Bapayeva, G.; Terzic, S.; Dotlic, J.; Togyzbayeva, K.; Bugibaeva, U.; Mustafinova, M.; Alisheva, A.; Garzon, S.; Terzic, M.; Laganà, A.S. Pregnancy outcomes in women with diabetes mellitus—the impact of diabetes type and treatment. *Menopause Rev./Przegląd Menopauzalny* **2022**, *21*, 37–46. [[CrossRef](#)]
98. Chivese, T.; Hoegfeldt, C.A.; Werfalli, M.; Yuen, L.; Sun, H.; Karuranga, S.; Li, N.; Gupta, A.; Immanuel, J.; Divakar, H. IDF Diabetes Atlas: The prevalence of pre-existing diabetes in pregnancy—A systematic review and meta-analysis of studies published during 2010–2020. *Diabetes Res. Clin. Pract.* **2022**, *183*, 109049. [[CrossRef](#)]
99. Thong, E.P.; Codner, E.; Laven, J.S.; Teede, H. Diabetes: A metabolic and reproductive disorder in women. *Lancet Diabetes Endocrinol.* **2020**, *8*, 134–149. [[CrossRef](#)]
100. Johns, E.C.; Denison, F.C.; Reynolds, R.M. The impact of maternal obesity in pregnancy on placental glucocorticoid and macronutrient transport and metabolism. *Biochim. Biophys. Acta (BBA)-Mol. Basis Dis.* **2020**, *1866*, 165374. [[CrossRef](#)]
101. Valsamakis, G.; Papatheodorou, D.; Chalarakis, N.; Manolikaki, M.; Margeli, A.; Papassotiriou, I.; Barber, T.M.; Kumar, S.; Kalantaridou, S.; Mastorakos, G. Maternal chronic stress correlates with serum levels of cortisol, glucose and C-peptide in the fetus, and maternal non chronic stress with fetal growth. *Psychoneuroendocrinology* **2020**, *114*, 104591. [[CrossRef](#)]
102. Basu, M.; Garg, V. Maternal hyperglycemia and fetal cardiac development: Clinical impact and underlying mechanisms. *Birth Defects Res.* **2018**, *110*, 1504–1516. [[CrossRef](#)]
103. Mukerji, G.; Bacon, S.; Feig, D.S. Gestational diabetes and type 2 diabetes during pregnancy. In *Maternal-Fetal and Neonatal Endocrinology*; Elsevier: Amsterdam, The Netherlands, 2020; pp. 371–388.
104. Vrachnis, N.; Antonakopoulos, N.; Iliodromiti, Z.; Dafopoulos, K.; Siristatidis, C.; Pappa, K.I.; Deligeorgiou, E.; Vitoratos, N. Impact of maternal diabetes on epigenetic modifications leading to diseases in the offspring. *J. Diabetes Res.* **2012**, *2012*, 538474. [[CrossRef](#)]
105. Ornoy, A.; Reece, E.A.; Pavlinkova, G.; Kappen, C.; Miller, R.K. Effect of maternal diabetes on the embryo, fetus, and children: Congenital anomalies, genetic and epigenetic changes and developmental outcomes. *Birth Defects Res. Part C Embryo Today Rev.* **2015**, *105*, 53–72. [[CrossRef](#)]
106. Ornoy, A.; Becker, M.; Weinstein-Fudim, L.; Ergaz, Z. Diabetes during pregnancy: A maternal disease complicating the course of pregnancy with long-term deleterious effects on the offspring. a clinical review. *Int. J. Mol. Sci.* **2021**, *22*, 2965. [[CrossRef](#)]
107. Aboud, F.; Torjman, F.; Sultan, M.; Benjama, A. Histopathological changes of umbilical cord blood vessels in diabetic pregnancies. *Tripolitan Med. J.* **2018**, *7*, 1–6.
108. Suma, M. Pregnancy Outcome in Gestational Diabetes Mellitus. Master's Thesis, Rajiv Gandhi University of Health Sciences, Bengaluru, India, 2011.

109. Mackin, S.T. Vascular Function in Hyperglycaemic Pregnancy: Studies into Potential Mechanisms for Adverse Obstetric Outcomes in Diabetes. Ph.D. Thesis, University of Glasgow, Glasgow, UK, 2021.
110. Sullivan, S.D.; Umans, J.G.; Ratner, R. Hypertension complicating diabetic pregnancies: Pathophysiology, management, and controversies. *J. Clin. Hypertens.* **2011**, *13*, 275–284. [[CrossRef](#)]
111. Mota, R.I.; Morgan, S.E.; Bahnson, E.M. Diabetic vasculopathy: Macro and microvascular injury. *Curr. Pathobiol. Rep.* **2020**, *8*, 1–14. [[CrossRef](#)]
112. Biesenbach, G.; Grafinger, P.; Zazgornik, J.; Stöger, H. Perinatal complications and three-year follow up of infants of diabetic mothers with diabetic nephropathy stage IV. *Ren. Fail.* **2000**, *22*, 573–580. [[CrossRef](#)]
113. Fowden, A.; Valenzuela, O.; Vaughan, O.; Jellyman, J.; Forhead, A. Glucocorticoid programming of intrauterine development. *Domest. Anim. Endocrinol.* **2016**, *56*, S121–S132. [[CrossRef](#)]
114. Lu, M.; Sferruzzi-Perri, A.N. Placental mitochondrial function in response to gestational exposures. *Placenta* **2021**, *104*, 124–137. [[CrossRef](#)]
115. Nugent, J.L. Effects of Glucocorticoids on Placental Development and Function: Implications for Fetal Growth Restriction. Ph.D. Thesis, The University of Manchester, Manchester, UK, 2012.
116. Cheong, J.N.; Wlodek, M.E.; Moritz, K.M.; Cuffe, J.S. Programming of maternal and offspring disease: Impact of growth restriction, fetal sex and transmission across generations. *J. Physiol.* **2016**, *594*, 4727–4740. [[CrossRef](#)]
117. Agorastos, A.; Pervanidou, P.; Chrousos, G.P.; Baker, D.G. Developmental trajectories of early life stress and trauma: A narrative review on neurobiological aspects beyond stress system dysregulation. *Front. Psychiatry* **2019**, *10*, 377300. [[CrossRef](#)]
118. Korgun, E.T.; Ozmen, A.; Unek, G.; Mendilcioglu, I. The effects of glucocorticoids on fetal and placental development. In *Glucocorticoids—New Recognition of Our Familiar Friend*; IntechOpen Limited: London, UK, 2012.
119. Vafaei-Nezhad, S.; Hami, J.; Sadeghi, A.; Ghaemi, K.; Hosseini, M.; Abedini, M.; Haghiri, H. The impacts of diabetes in pregnancy on hippocampal synaptogenesis in rat neonates. *Neuroscience* **2016**, *318*, 122–133. [[CrossRef](#)]
120. Sadeghi, A.; Asghari, H.; Hami, J.; Roodi, M.M.; Mostafaei, H.; Karimipour, M.; Namavar, M.; Idoon, F. Volumetric investigation of the hippocampus in rat offspring due to diabetes in pregnancy—A stereological study. *J. Chem. Neuroanat.* **2019**, *101*, 101669. [[CrossRef](#)]
121. Rakers, F.; Rupprecht, S.; Dreiling, M.; Bergmeier, C.; Witte, O.W.; Schwab, M. Transfer of maternal psychosocial stress to the fetus. *Neurosci. Biobehav. Rev.* **2020**, *117*, 185–197. [[CrossRef](#)]
122. McGowan, P.O.; Matthews, S.G. Prenatal stress, glucocorticoids, and developmental programming of the stress response. *Endocrinology* **2018**, *159*, 69–82. [[CrossRef](#)]
123. Van den Bergh, B.R.; van den Heuvel, M.I.; Lahti, M.; Braeken, M.; de Rooij, S.R.; Entringer, S.; Hoyer, D.; Roseboom, T.; Räikkönen, K.; King, S. Prenatal developmental origins of behavior and mental health: The influence of maternal stress in pregnancy. *Neurosci. Biobehav. Rev.* **2020**, *117*, 26–64. [[CrossRef](#)]
124. Manojlović-Stojanoski, M.; Nestorović, N.; Milošević, V. Prenatal glucocorticoids: Short-term benefits and long-term risks. In *Glucocorticoids—New Recognition of Our Familiar Friend*; Tech Open Access Publisher: Rijeka, Croatia, 2012; pp. 337–390.
125. Rooij, S.R.d. Prenatal Diet and Stress Responsiveness. In *Handbook of Behavior, Food and Nutrition*; Springer: New York, NY, USA, 2011; pp. 2023–2039.
126. Moisiadis, V.G.; Matthews, S.G. Glucocorticoids and fetal programming part 2: Mechanisms. *Nat. Rev. Endocrinol.* **2014**, *10*, 403–411. [[CrossRef](#)]
127. de Kloet, E.R.; Claessens, S.E.; Kentrop, J. Context modulates outcome of perinatal glucocorticoid action in the brain. *Front. Endocrinol.* **2014**, *5*, 100. [[CrossRef](#)] [[PubMed](#)]
128. Sallam, N.A.; Palmgren, V.A.; Singh, R.D.; John, C.M.; Thompson, J.A. Programming of vascular dysfunction in the intrauterine milieu of diabetic pregnancies. *Int. J. Mol. Sci.* **2018**, *19*, 3665. [[CrossRef](#)]
129. Sharma, D.; Shastri, S.; Sharma, P. Intrauterine growth restriction: Antenatal and postnatal aspects. *Clin. Med. Insights Pediatr.* **2016**, *10*, 67–83. [[CrossRef](#)]
130. Kesavan, K.; Devaskar, S.U. Intrauterine growth restriction: Postnatal monitoring and outcomes. *Pediatr. Clin.* **2019**, *66*, 403–423.
131. Parikh, N.I.; Gonzalez, J.M.; Anderson, C.A.; Judd, S.E.; Rexrode, K.M.; Hlatky, M.A.; Gunderson, E.P.; Stuart, J.J.; Vaidya, D.; American Heart Association Council on Epidemiology and Prevention; et al. Adverse pregnancy outcomes and cardiovascular disease risk: Unique opportunities for cardiovascular disease prevention in women: A scientific statement from the American Heart Association. *Circulation* **2021**, *143*, e902–e916. [[CrossRef](#)]
132. Burd, I.; Baschat, A.; Costantine, M. *Prenatal Beginnings for Better Health*; Frontiers Media SA: Lausanne, Switzerland, 2018.
133. Robinson, N. Early Life Risk Factors and Epigenetic Biomarkers of Obesity across the Life Course. Ph.D. Thesis, Newcastle University, Newcastle upon Tyne, UK, 2020.
134. López-Gamero, A.; Martínez, F.; Salazar, K.; Cifuentes, M.; Nualart, F. Brain glucose-sensing mechanism and energy homeostasis. *Mol. Neurobiol.* **2019**, *56*, 769–796. [[CrossRef](#)]
135. Nimgampalle, M.; Chakravarthy, H.; Devanathan, V. Glucose metabolism in the brain: An update. In *Recent Developments in Applied Microbiology and Biochemistry*; Elsevier: Amsterdam, The Netherlands, 2021; pp. 77–88.
136. Tartour, A.; Chivese, T.; Eltayeb, S.; Elamin, F.M.; Fthenou, E.; Ahmed, M.S.; Babu, G.R. Prenatal Psychological Distress and 11 β -HSD2 Gene Expression in Human Placentas: Systematic Review and Meta-analysis. *Psychoneuroendocrinology* **2024**, *166*, 107060. [[CrossRef](#)] [[PubMed](#)]

137. Reynolds, R.M.; Walker, B.R.; Syddall, H.E.; Andrew, R.; Wood, P.J.; Whorwood, C.B.; Phillips, D.I. Altered control of cortisol secretion in adult men with low birth weight and cardiovascular risk factors. *J. Clin. Endocrinol. Metab.* **2001**, *86*, 245–250. [[CrossRef](#)] [[PubMed](#)]
138. Dunlop, K.; Sarr, O.; Stachura, N.; Zhao, L.; Nygard, K.; Thompson, J.A.; Hadway, J.; Richardson, B.S.; Bureau, Y.; Borradaile, N. Differential and synergistic effects of low birth weight and western diet on skeletal muscle vasculature, mitochondrial lipid metabolism and insulin signaling in male Guinea pigs. *Nutrients* **2021**, *13*, 4315. [[CrossRef](#)]
139. Singhal, A. Should we promote catch-up growth or growth acceleration in low-birthweight infants? In *Low-Birthweight Baby: Born too Soon or too Small*; Karger: Basel, Switzerland, 2015; Volume 81, pp. 51–60.
140. Möllers, L.S.; Yousuf, E.I.; Hamatschek, C.; Morrison, K.M.; Hermanussen, M.; Fusch, C.; Rochow, N. Metabolic-endocrine disruption due to preterm birth impacts growth, body composition, and neonatal outcome. *Pediatr. Res.* **2022**, *91*, 1350–1360. [[CrossRef](#)]
141. Marcovecchio, M.L.; Gorman, S.; Watson, L.P.; Dunger, D.B.; Beardsall, K. Catch-up growth in children born small for gestational age related to body composition and metabolic risk at six years of age in the UK. *Horm. Res. Paediatr.* **2020**, *93*, 119–127. [[CrossRef](#)]
142. Dulloo, A.G. Thrifty energy metabolism in catch-up growth trajectories to insulin and leptin resistance. *Best Pract. Res. Clin. Endocrinol. Metab.* **2008**, *22*, 155–171. [[CrossRef](#)] [[PubMed](#)]
143. Kelishadi, R.; Haghdoost, A.A.; Jamshidi, F.; Aliramezany, M.; Moosazadeh, M. Low birthweight or rapid catch-up growth: Which is more associated with cardiovascular disease and its risk factors in later life? A systematic review and cryptanalysis. *Paediatr. Int. Child Health* **2015**, *35*, 110–123. [[CrossRef](#)] [[PubMed](#)]
144. Dasinger, J.H.; Alexander, B.T. Gender differences in developmental programming of cardiovascular diseases. *Clin. Sci.* **2016**, *130*, 337–348. [[CrossRef](#)] [[PubMed](#)]
145. Tarnovski, L.; Brinar, I.V.; Kirhmajer, M.V.; Vrkic, T.Z.; Laganovic, M. Sex Differences in Cardiovascular Risk Factors and Renal Function among Young Adults after Intrauterine Growth Restriction. *Acta Clin. Croat.* **2021**, *60*, 164–173.
146. Andersson, S.W.; Lapidus, L.; Niklasson, A.; Hallberg, L.; Bengtsson, C.; Hulthén, L. Blood pressure and hypertension in middle-aged women in relation to weight and length at birth: A follow-up study. *J. Hypertens.* **2000**, *18*, 1753–1761. [[CrossRef](#)] [[PubMed](#)]
147. Barker, D.; Bull, A.R.; Osmond, C.; Simmonds, S.J. Fetal and placental size and risk of hypertension in adult life. *Br. Med. J.* **1990**, *301*, 259–262. [[CrossRef](#)] [[PubMed](#)]
148. Brar, P.C. Update on the current modalities used to screen high risk youth for prediabetes and/or type 2 diabetes mellitus. *Ann. Paediatr. Endocrinol. Metab.* **2019**, *24*, 71. [[CrossRef](#)] [[PubMed](#)]
149. Tura, A.; Grespan, E.; Göbl, C.S.; Koivula, R.W.; Franks, P.W.; Pearson, E.R.; Walker, M.; Forgie, I.M.; Giordano, G.N.; Pavo, I. Profiles of glucose metabolism in different prediabetes phenotypes, classified by fasting glycemia, 2-hour OGTT, glycated hemoglobin, and 1-hour OGTT: An IMI DIRECT study. *Diabetes* **2021**, *70*, 2092–2106. [[CrossRef](#)] [[PubMed](#)]
150. Yip, W.C.; Sequeira, I.R.; Plank, L.D.; Poppitt, S.D. Prevalence of pre-diabetes across ethnicities: A review of impaired fasting glucose (IFG) and impaired glucose tolerance (IGT) for classification of dysglycaemia. *Nutrients* **2017**, *9*, 1273. [[CrossRef](#)]
151. Succurro, E.; Pedace, E.; Andreozzi, F.; Papa, A.; Vizza, P.; Fiorentino, T.V.; Perticone, F.; Veltri, P.; Cascini, G.L.; Sesti, G. Reduction in global myocardial glucose metabolism in subjects with 1-hour postload hyperglycemia and impaired glucose tolerance. *Diabetes Care* **2020**, *43*, 669–676. [[CrossRef](#)]
152. Penhaligan, J.; Sequeira-Bisson, I.R.; Miles-Chan, J.L. The role of postprandial thermogenesis in the development of impaired glucose tolerance and type II diabetes. *Am. J. Physiol.-Endocrinol. Metab.* **2023**, *325*, E171–E179. [[CrossRef](#)] [[PubMed](#)]
153. Mittendorfer, B.; Patterson, B.W.; Smith, G.I.; Yoshino, M.; Klein, S. β cell function and plasma insulin clearance in people with obesity and different glycemic status. *J. Clin. Investig.* **2022**, *132*, e154068. [[CrossRef](#)] [[PubMed](#)]
154. Papakonstantinou, E.; Oikonomou, C.; Nychas, G.; Dimitriadis, G.D. Effects of diet, lifestyle, chrononutrition and alternative dietary interventions on postprandial glycemia and insulin resistance. *Nutrients* **2022**, *14*, 823. [[CrossRef](#)] [[PubMed](#)]
155. Roncero-Ramos, I.; Alcalá-Díaz, J.F.; Rangel-Zuñiga, O.A.; Gomez-Delgado, F.; Jimenez-Lucena, R.; García-Rios, A.; Vals-Delgado, C.; Romero-Baldonado, C.; Luque, R.M.; Ordovas, J.M. Prediabetes diagnosis criteria, type 2 diabetes risk and dietary modulation: The CORDIOPREV study. *Clin. Nutr.* **2020**, *39*, 492–500. [[CrossRef](#)] [[PubMed](#)]
156. Bergman, M.; Manco, M.; Sesti, G.; Dankner, R.; Pareek, M.; Jagannathan, R.; Chetrit, A.; Abdul-Ghani, M.; Buysschaert, M.; Olsen, M.H. Petition to replace current OGTT criteria for diagnosing prediabetes with the 1-hour post-load plasma glucose \geq 155 mg/dL (8.6 mmol/L). *Diabetes Res. Clin. Pract.* **2018**, *146*, 18–33. [[CrossRef](#)] [[PubMed](#)]
157. Herman, W.H. Prediabetes Diagnosis and Management. *JAMA* **2023**, *329*, 1157–1159. [[CrossRef](#)] [[PubMed](#)]
158. Basit, A.; Fawwad, A.; Basit, K.A.; Waris, N.; Tahir, B.; Siddiqui, I.A. Glycated hemoglobin (HbA1c) as diagnostic criteria for diabetes: The optimal cut-off points values for the Pakistani population; a study from second National Diabetes Survey of Pakistan (NDSP) 2016–2017. *BMJ Open Diabetes Res. Care* **2020**, *8*, e001058. [[CrossRef](#)] [[PubMed](#)]
159. Rett, K.; Gottwald-Hostalek, U. Understanding prediabetes: Definition, prevalence, burden and treatment options for an emerging disease. *Curr. Med. Res. Opin.* **2019**, *35*, 1529–1534. [[CrossRef](#)]
160. Ekta; Gupta, M.; Kaur, A.; Singh, T.G.; Bedi, O. Pathobiological and molecular connections involved in the high fructose and high fat diet induced diabetes associated nonalcoholic fatty liver disease. *Inflamm. Res.* **2020**, *69*, 851–867. [[CrossRef](#)]

161. Aimaretti, E.; Chimienti, G.; Rubeo, C.; Di Lorenzo, R.; Trisolini, L.; Dal Bello, F.; Moradi, A.; Collino, M.; Lezza, A.M.S.; Aragno, M. Different Effects of High-Fat/High-Sucrose and High-Fructose Diets on Advanced Glycation End-Product Accumulation and on Mitochondrial Involvement in Heart and Skeletal Muscle in Mice. *Nutrients* **2023**, *15*, 4874. [[CrossRef](#)]
162. Sangrós, F.J.; Torrecilla, J.; Giraldez-García, C.; Carrillo, L.; Mancera, J.; Mur, T.; Franch, J.; Diez, J.; Goday, A.; Serrano, R. Association of general and abdominal obesity with hypertension, dyslipidemia and prediabetes in the PREDAPS Study. *Rev. Esp. Cardiol.* **2018**, *71*, 170–177. [[CrossRef](#)]
163. Hirano, T. Pathophysiology of diabetic dyslipidemia. *J. Atheroscler. Thromb.* **2018**, *25*, 771–782. [[CrossRef](#)]
164. Chi, J.H.; Lee, B.J. Risk factors for hypertension and diabetes comorbidity in a Korean population: A cross-sectional study. *PLoS ONE* **2022**, *17*, e0262757. [[CrossRef](#)]
165. Lee, S.-H.; Park, S.-Y.; Choi, C.S. Insulin resistance: From mechanisms to therapeutic strategies. *Diabetes Metab. J.* **2022**, *46*, 15. [[CrossRef](#)] [[PubMed](#)]
166. Cignarelli, A.; Genchi, V.A.; Perrini, S.; Natalicchio, A.; Laviola, L.; Giorgino, F. Insulin and insulin receptors in adipose tissue development. *Int. J. Mol. Sci.* **2019**, *20*, 759. [[CrossRef](#)] [[PubMed](#)]
167. Losada-Barragán, M. Physiological effects of nutrients on insulin release by pancreatic beta cells. *Mol. Cell. Biochem.* **2021**, *476*, 3127–3139. [[CrossRef](#)]
168. Gil-Rivera, M.; Medina-Gali, R.M.; Martínez-Pinna, J.; Soriano, S. Physiology of pancreatic β -cells: Ion channels and molecular mechanisms implicated in stimulus-secretion coupling. *Int. Rev. Cell Mol. Biol.* **2021**, *359*, 287–323. [[PubMed](#)]
169. Jing, J.; Li, J.; Yan, N.; Li, N.; Liu, X.; Li, X.; Zhang, J.; Wang, Q.; Yang, C.; Qiu, J. Increased TG Levels and HOMA-IR Score Are Associated With a High Risk of Prediabetes: A Prospective Study. *Asia Pac. J. Public Health* **2023**, *35*, 413–419. [[CrossRef](#)] [[PubMed](#)]
170. Hostalek, U. Global epidemiology of prediabetes-present and future perspectives. *Clin. Diabetes Endocrinol.* **2019**, *5*, 5. [[CrossRef](#)]
171. Carmichael, J.; Fadavi, H.; Ishibashi, F.; Shore, A.C.; Tavakoli, M. Advances in screening, early diagnosis and accurate staging of diabetic neuropathy. *Front. Endocrinol.* **2021**, *12*, 671257. [[CrossRef](#)]
172. Hill, J.O.; Galloway, J.M.; Goley, A.; Marrero, D.G.; Minners, R.; Montgomery, B.; Peterson, G.E.; Ratner, R.E.; Sanchez, E.; Aroda, V.R. Scientific statement: Socioecological determinants of prediabetes and type 2 diabetes. *Diabetes Care* **2013**, *36*, 2430–2439. [[CrossRef](#)]
173. Mohan, V.; Sudha, V.; Shobana, S.; Gayathri, R.; Krishnaswamy, K. Are unhealthy diets contributing to the rapid rise of type 2 diabetes in India? *J. Nutr.* **2023**, *153*, 940–948. [[CrossRef](#)]
174. Sosibo, A.M.; Mzimela, N.C.; Ngubane, P.S.; Khathi, A. Prevalence of pre-diabetes in adults aged 25–45 years in a Durban-based clinical setting, South Africa: A retrospective study. *Prim. Care Diabetes* **2023**, *17*, 650–654. [[CrossRef](#)]
175. Al-Rifai, R.H.; Majeed, M.; Qambar, M.A.; Ibrahim, A.; AlYammahi, K.M.; Aziz, F. Type 2 diabetes and pre-diabetes mellitus: A systematic review and meta-analysis of prevalence studies in women of childbearing age in the Middle East and North Africa, 2000–2018. *Syst. Rev.* **2019**, *8*, 268. [[CrossRef](#)]
176. Andes, L.J.; Cheng, Y.J.; Rolka, D.B.; Gregg, E.W.; Imperatore, G. Prevalence of prediabetes among adolescents and young adults in the United States, 2005–2016. *JAMA Pediatr.* **2020**, *174*, e194498. [[CrossRef](#)]
177. Bansal, N. Prediabetes diagnosis and treatment: A review. *World J. Diabetes* **2015**, *6*, 296. [[CrossRef](#)] [[PubMed](#)]
178. Mzimela, N.C.; Ngubane, P.S.; Khathi, A. The changes in immune cell concentration during the progression of pre-diabetes to type 2 diabetes in a high-fat high-carbohydrate diet-induced pre-diabetic rat model. *Autoimmunity* **2019**, *52*, 27–36. [[CrossRef](#)] [[PubMed](#)]
179. Naidoo, K.; Ngubane, P.S.; Khathi, A. Investigating the Effects of Diet-Induced Pre-Diabetes on the Functioning of Calcium-Regulating Organs in Male Sprague Dawley Rats: Effects on Selected Markers. *Front. Endocrinol.* **2022**, *13*, 914189. [[CrossRef](#)]
180. Chakraborty, S.; Verma, A.; Garg, R.; Singh, J.; Verma, H. Cardiometabolic Risk Factors Associated with Type 2 Diabetes Mellitus: A Mechanistic Insight. *Clin. Med. Insights Endocrinol. Diabetes* **2023**, *16*, 11795514231220780. [[CrossRef](#)] [[PubMed](#)]
181. Ludidi, A.; Siboto, A.; Nkosi, A.; Xulu, N.D.; Khathi, A.; Sibiya, N.H.; Ngubane, P.S. High-fat, high-carbohydrate diet-induced prediabetes preconception in Sprague–Dawley rats as a risk factor for the development of preeclampsia: Assessing changes in placental metabolic insults. *Front. Nutr.* **2023**, *10*, 1241785. [[CrossRef](#)]
182. Alatrach, M.; Agyin, C.; Adams, J.; DeFronzo, R.A.; Abdul-Ghani, M.A. Decreased basal hepatic glucose uptake in impaired fasting glucose. *Diabetologia* **2017**, *60*, 1325–1332. [[CrossRef](#)] [[PubMed](#)]
183. Hudish, L.I.; Reusch, J.E.; Sussel, L. β Cell dysfunction during progression of metabolic syndrome to type 2 diabetes. *J. Clin. Investig.* **2019**, *129*, 4001–4008. [[CrossRef](#)] [[PubMed](#)]
184. Katugampola, H.; Gevers, E.F.; Dattani, M.T. Fetal Endocrinology. In *Brook's Clinical Pediatric Endocrinology*; John Wiley & Sons Ltd.: Hoboken, NJ, USA, 2019; pp. 47–104.
185. Hoffman, D.J.; Reynolds, R.M.; Hardy, D.B. Developmental origins of health and disease: Current knowledge and potential mechanisms. *Nutr. Rev.* **2017**, *75*, 951–970. [[CrossRef](#)] [[PubMed](#)]
186. Zygula, A.; Kosinski, P.; Wielgos, M. Saliva, hair, tears, and other biological materials obtained non-invasively for diagnosis in pregnancy: A literature review. *Ginekol. Pol.* **2019**, *90*, 475–481. [[CrossRef](#)] [[PubMed](#)]

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