

Supplementary Material

Table S1. Concentrations and impact of AD inhibitors reported in the literature.

Inhibitor	Concentration (mg/L)	Substrate	Conditions	Performance reduction	Ref.
TAN	4000	Cattle manure	CSTR 55 °C	25% reduction in methane yield	[1]
TAN	5 and 7 g-N/L TAN	Cattle manure	UASB	25% reduction in methane yield	[2]
TAN	4 and 7.5 g-N/L TAN	Cattle manure	SMA test	50% reduced growth rate for methanogens	[2]
TAN	1500-3000	-	pH>7.4-7.6	Inhibitory	[3]
TAN	10000 (1200mg/L FAN)	Cattle manure	CSTR 55 °C	Inhibition of the methanogenesis	[4]
FAN	55 at 40 d 32 at 15 and 25 d	Acetate & Propionate	CSTR	Maximum tolerable concentration	[5]
FAN	61	Crab cooker wastewater	UASB 37 °C	40% inhibition of methanogenic activity	[6]
FAN	85	Crab cooker wastewater	ATA test 37 °C	50% inhibition	[6]
FAN	>250	Chicken manure	Batch test	Methane production ceased	[7]
NO ₃ ⁻	150 mg-N/L	Dextrin peptone	Batch test 35 °C	52% reduction in methane production rate	[8]
NO ₃ ⁻	300 mg-N/L	Dextrin peptone	CSTR 35 °C	90% reduction in methane production rate 95% reduction in methane production rate (after recover of reactor)	[8]
NO ₃ ⁻	156 mg-N/L	Acetate	Batch test	Methanogenesis inhibition	[9]
NO ₃ ⁻	10 mM	Soil	Soil test	Stopped methanogenesis, recovery 70% methane production	[10]
NO ₃ ⁻	5mM	H ₂ /CO ₂ and/or Acetate	<i>M. bakery</i> Batch test	65% reduction in methane production	[10]
NO ₃ ⁻	30 mM	H ₂ /CO ₂ and/or Acetate	<i>M. bryantii</i> Batch test	59% reduction in methane production	[10]
NO ₃ ⁻	8 COD/NO ₃ -N	Glucose	Batch test 35 °C	86% reduction in methane production	[11]
NO ₃ ⁻	8 COD/NO ₃ -N	Glucose	CSTR 35 °C	34.8% reduction of % of methane in biogas	[11]
NO ₃ ⁻	14.3 mM	Acetate or methanol	Batch test	46% inhibition on <i>M. mazy</i> by competition	[12]
NO ₂ ⁻	50 mg-N/L	Dextrin peptone	Batch test 35 °C	20% reduction in methane production rate	[12]
NO ₂ ⁻	315 mg-N/L	Glucose	UASB 30 °C	12% reduction	[13]
NO ₂ ⁻	50	Glucose	Batch test Non-acclimated dispersed	50% reduction SMA (Specific methanogenic activity)	[13]

NO ₂ ⁻	100	Glucose	Batch test Non-acclimated granular	50% reduction SMA	[13]
NO ₂ ⁻	100	Glucose	Batch test Acclimated dispersed	50% reduction SMA	[13]
NO ₂ ⁻	0.18mM 0.32mM NO _x	Acetate or methanol	Batch test	96-97% inhibition <i>M. mazey</i> 95% inhibition <i>M. mazey</i>	[12]
NO ₂ ⁻	50uM		<i>M. bakery</i> Batch test	50% reduction in methane production	[10]
NO ₂ ⁻	1mM	H ₂ /CO ₂ and/or Acetate	<i>M. bryantii</i> Batch test	50% reduction in methane production	[10]
CN ⁻	4.5, possible recovery 50, no recovery	Fibber chemical plant wastewater	Batch test	50% IC (toxic to the sludge's methane-producing activity)	[14]
CN ⁻	5 mg/L	Cassava derived starch and VFA	UASB 33 °C	70% reduction in methane production	[15]
CN ⁻	125 mg/L	Cassava derived starch and VFA	UASB 33 °C	96% methane reduction with acclimation	[10]
SO ₄ ²⁻	150	Glucose for semi-batch, VFAs for SMA	Granular sludge, Semi-batch, SMA to the outlet	65% SMA decrease using butyrate at 59 d, recovering at 114 d to 8% SMA decrease	[16]
SO ₄ ²⁻	Change of COD/SO ₄ from 20 to 0.5 (6000 mg/L)	Synthetic organic chemical industry wastewater	UASB 35 °C, Batch tests to the outlet	27% reduction in methane production rate	[17]
SO ₄ ²⁻	2.2 kg SO ₄ /m ³ /d	Mussels process wastewater	Anaerobic filter 37 °C	15% and 2% of Propionate and acetate was converted by sulphate reducing microorganism instead of the methanogens	[18]
SO ₄ ²⁻	1500 (6.7 COD/SO ₄ ratio)	Sewage sludge	CSTR 35 °C	Decrease of 33% of methane production	[19]
Cd	170 Pulse feed:680	Sludge	Fixed film bed reactor Semi-continuous 22 °C	55% biogas reduction Pulse feed: 84.8% decrease	[20]
Cd	15-23 11-34 (with Nickel)	Acetate solution	Pure culture 60 °C	50% inhibition of thermophilic methanogens	[21]
Cd	3300	Sucrose	Batch Hydrogen generation Granular sludge 26 °C	50% reduction of hydrogen potential	[22]
Cd	36	Glucose	Batch 5 d Granular sludge 35 °C	50% reduction of methane production	[23]
Fe	200	Sludge	Batch 55 °C	50% biogas production using Fe NO ₃	[24]
Fe (III)	1 mM	Acetate	Batch, Pure culture, <i>M. hungatei</i> 30 °C	Almost completely inhibited methane production	[25]

		Carbon dioxide and hydrogen			
Fe (III)	5-10 mM	Acetate Carbon dioxide and hydrogen	Batch, Pure culture, <i>M. barkeri</i> 30 °C	(H ₂ /CO ₂) significantly decreased methane production (acetate) methane production was also significantly reduced but less	[25]
Zn	1500	Sucrose	Batch, Hydrogen generation, Granular sludge 26 °C	50% reduction of hydrogen potential	[22]
Zn	4.5	Sucrose	Batch 72 h, Hydrogen generation 35 °C	50% reduction of hydrogen production	[20]
Zn	7.5	Glucose	Batch 5 d, Granular sludge 35 °C	50% reduction in methane production	[23]
Ni	1300	Sucrose	Batch, Hydrogen generation, Granular sludge 26 °C	50% reduction of hydrogen potential	[22]
Ni	50	Ethanol and acetate	EGSB	18% inhibition methanogen	[26]
Ni	14.5-130	Acetate solution	Pure culture 60 °C	50% inhibition of thermophilic methanogens	[21]
Ni	35	Glucose	Batch 5 d, Granular sludge 35 °C	50% reduction of methane production	[23]
Cr	3000	Sucrose	Batch, Hydrogen generation, Granular sludge 26 °C	50% reduction of hydrogen potential	[22]
Cr	60	Sucrose	Batch 72 h Hydrogen generation 35 °C	50% reduction of hydrogen production	[27]
Cr	50	Ethanol and acetate	EGSB	11% inhibition methanogens	[26]
Cr (III)	775 Pulse feed: 3100	Sludge	Fixed film bed reactor Semi-continuous 22 °C	52% biogas reduction Pulse feed: 83% decrease	[28]
Cr	27	Glucose	Batch 5 d, Granular sludge 35 °C	50% reduction in methane production	[23]
Al	1000	Glucose for semi-batch, VFAs for SMA	Granular sludge Semi-batch SMA to the outlet	72% SMA decrease using butyrate at 59 d, recovering at 114 d to 21% SMA decrease and 36% SMA when 150 mg/L of sulphate was added	[16]
Al	2500	Synthetic waste	UASB semi-continuous	Digester failure (85% decrease of biogas volume approximately)	[29]

Cu	6.5	Sucrose	Batch 72 h, Hydrogen generation 35 °C	50% reduction of hydrogen production	[27]
Cu	30	Sucrose	Batch, Hydrogen generation, Granular sludge 26 °C	50% reduction of hydrogen potential	[22]
Cu	2.4-14, 2.2-28 (with Ni)	Acetate solution	Pure culture 60 °C	50% inhibition of thermophilic methanogens	[21]
Cu	50	Ethanol and acetate	EGSB	21% methanogen inhibition with the presence of accumulated Cr 69% methanogen inhibition	[26]
Pb	>5000	Sucrose	Batch, Hydrogen generation, Granular sludge 26 °C	50% reduction of hydrogen potential	[22]
As III	15.5 uM 27.1 uM 4.4 uM	Acetate Hydrogen Lactate	Batch test 1 d, Granular sludge 30 °C	50% inhibition of methanogens	[30]
As III	23.5	Acetate	Batch test for 1 week Granular sludge 30 °C	36.9% inhibition of methanogens	[30]
As III	100uM	Acetate	Batch test 1 d, Flocculent sludge 30 °C	50% inhibition of methanogens	[30]
As V	>500 uM	Acetate or Hydrogen or Lactate	Batch test 1 d, Granular and flocculent sludge 30 °C	Non-inhibition	[30]
As V	500 uM	Acetate	Batch test for 4 d, flocculent sludge 30 °C	Complete inhibition	[30]
SeO ₃	10 ⁻³ M 10 ⁻² M	H ₂ /CO ₂ Acetate	Granular sludge Semi-batch	>96% inhibition	[31]
SeO ₄	10 ⁻³ M	Acetate H ₂ /CO ₂	Granular sludge Semi-batch	>96% inhibition	[31]
Hg	125 Pulse feed: 500	Sludge	Fixed film bed reactor Semi-continuous 22 °C	60% biogas reduction Pulse feed: 92% decrease after 2 days	[20]
F ⁻	325	Glucose	Batch tests, Granular fermenters 30 °C	50% reduction of control activity	[32]
F ⁻	36.5	Propionate	Batch test, acetogenic bacteria 30 °C	50% reduction of control activity	[32]
	25.5	Butyrate			
F ⁻	34.5	Acetate	Batch test, Flocculent sludge 30 °C	50% reduction of methanogenesis	[32]
	82	Hydrogen			
F ⁻	18.1	Acetate	Batch test, Flocculent sludge 55 °C	50% reduction of methanogenesis	[32]
	433	Hydrogen			
F ⁻	160	Acetate	Batch test, Granular 30 °C	50% reduction of methanogenesis	[32]

Cl ⁻	>5500	Tannery wastewater	Contact filter reactor	Decreasing trend	[33]
Phenol	300	Phenol, polyphenols, acetate and propionate	ATA and BMP 37 °C	Reduction of expected ratio, 2.03, to 1.62	[28]
Phenol	250	Phenol	2 stage AD 35 °C	62% biogas production reduction	[34]

Continue stir reactor: CSTR; Up-flow anaerobic sludge blanket: UASB; Specific methanogenic activity: SMA; Expanded granular sludge bed: EGSB

Table S2. Statistics of the biogas and methane yield using ANOVA compared with the control test (if $F > F_{crit}$ they are different).

<i>CuSO₄</i>	<i>mg Cu/L</i>	20	32	73	123	167
Biogas	F	0.77	0.01	0.26	0.18	0.15
Biogas	F _{crit}	7.71	7.71	7.71	7.71	7.71
Methane	F	0.00	0.61	0.03	0.19	1.64
Methane	F _{crit}	7.71	7.71	7.71	7.71	7.71
<i>Al₂(SO₄)₃</i>	<i>mg Al/L</i>	85	210	771	1308	1534
Biogas	F	0.14	0.00	24.59	39.19	309.83
Biogas	F _{crit}	7.71	7.71	7.71	7.71	7.71
Methane	F	0.13	10.82	29.84	155.46	1728.87
Methane	F _{crit}	7.71	7.71	7.71	7.71	7.71
<i>NaCl</i>	<i>mg Cl/L</i>	1066	1941	2821	3666	4535
Biogas	F	22.36	71.65	13.08	5.07	19.78
Biogas	F _{crit}	7.71	7.71	7.71	7.71	7.71
Methane	F	4.29	2.99	1.67	5.07	20.12
Methane	F _{crit}	7.71	7.71	7.71	7.71	7.71
<i>NaSO₄</i>	<i>mg SO₄/L</i>	162	308	600	1032	1466
Biogas	F	5.18	59.24	66.89	247.08	47.18
Biogas	F _{crit}	7.71	7.71	7.71	7.71	7.71
Methane	F	65.10	115.08	193.43	715.63	204.86
Methane	F _{crit}	7.71	7.71	7.71	7.71	7.71
<i>CuCl₂</i>	<i>mg Cu/L</i>	22	91	198	501	804
Biogas	F	802.48	823.00	567.04	292.74	2496.73
Biogas	F _{crit}	7.71	7.71	7.71	7.71	7.71
Methane	F	446.98	1105.54	239.50	241.92	4729.63
Methane	F _{crit}	7.71	7.71	7.71	7.71	7.71
<i>AlCl₃</i>	<i>mg Al/L</i>	82	203	760	1312	1531
Biogas	F	5.69	19.16	284.00	2483.70	3709.46
Biogas	F _{crit}	7.71	7.71	7.71	7.71	7.71
Methane	F	8.59	11.85	549.89	4231.14	4233.74
Methane	F _{crit}	7.71	7.71	7.71	7.71	7.71
<i>ZnCl₂</i>	<i>mg Zn/L</i>	10	61	87	592	1307
Biogas	F	0.22	0.23	6.07	1348.11	2973.37
Biogas	F _{crit}	7.71	7.71	7.71	7.71	7.71
Methane	F	0.00	4.87	3.76	1421.18	2923.37
Methane	F _{crit}	7.71	7.71	7.71	7.71	7.71
<i>NH₄SO₄</i>	<i>mg NH₄/L</i>	1460	1602	1764	2201	3405
Biogas	F	3.61	0.03	10.75	47.23	28.78
Biogas	F _{crit}	7.71	7.71	7.71	7.71	7.71
Methane	F	36.82	4.68	49.21	19.93	18.83
Methane	F _{crit}	7.71	7.71	7.71	7.71	7.71
<i>NaNO₃</i>	<i>mg NO₃/L</i>	92	174	254	332	427
Biogas	F	100.75	11.85	176.30	43.46	1.36
Biogas	F _{crit}	7.71	7.71	7.71	7.71	7.71
Methane	F	684.23	80.07	270.19	653.99	65.37
Methane	F _{crit}	7.71	7.71	7.71	7.71	7.71
<i>HgSO₄</i>	<i>mg Hg/L</i>	25	20	920	239	495
Biogas	F	1.63	7.65	15.86	110.65	8.19
Biogas	F _{crit}	7.71	7.71	7.71	7.71	7.71
Methane	F	13.91	42.45	1.96	11.47	23.17
Methane	F _{crit}	7.71	7.71	7.71	7.71	7.71
<i>NH₄Cl</i>	<i>mg NH₄/L</i>	976	1958	2946	3923	5162
Biogas	F	0.09	0.16	0.87	5.62	28.21
Biogas	F _{crit}	7.71	7.71	7.71	7.71	7.71
Methane	F	0.57	0.00	0.06	1.73	23.79
Methane	F _{crit}	7.71	7.71	7.71	7.71	7.71
<i>NaHASO₄</i>	<i>mg As/L</i>	0.1	0.4	2	4	17

Biogas	F	0.43	5.15	0.09	0.77	0.39	
Biogas	Fcrit	7.71	7.71	7.71	7.71	7.71	
Methane	F	0.34	1.15	0.04	5.17	0.59	
Methane	Fcrit	7.71	7.71	7.71	7.71	7.71	
<i>Al+OH/pH</i>		<i>pH 5</i>	<i>pH 4.5</i>	<i>pH 6.55</i>	<i>Al 608 mg/L</i>	<i>Al 262 mg/L</i>	<i>Al 464 mg/L</i>
Biogas	F	37.14	54.77	2.26	13.43	8.29	155.18
Biogas	Fcrit	18.51	18.51	18.51	18.51	18.51	18.51
Methane	F	600.01	1761.45	0.97	17.10	18.25	104.83
Methane	Fcrit	18.51	18.51	18.51	18.51	18.51	18.51

Table S3. Results from first-order and Gompertz model for the biogas formation kinetics on the BMP experiments (B=biogas; Conc= concentration on the reactor; C=control).

Salt	Ion	Conc (mg/L)	1st order				Gompertz model				
			k_h (d ⁻¹)	P_{max} (mL B/g VS)	RMSE _n	r^2	R_m (mL B/g VS/d)	λ (d)	P_{max} (mL B/g VS)	RMSE _n	r^2
(NH ₄) ₂ SO ₄	NH ₄	635 (C)	0.46	598	0.15	0.91	111	0.0	611	0.11	0.98
(NH ₄) ₂ SO ₄	NH ₄	1460	0.59	553	0.11	0.928	132	0.0	566	0.12	0.992
(NH ₄) ₂ SO ₄	NH ₄	1602	0.62	604	0.10	0.939	150	0.0	617	0.13	0.983
(NH ₄) ₂ SO ₄	NH ₄	1764	0.58	565	0.11	0.932	132	0.0	578	0.12	0.990
(NH ₄) ₂ SO ₄	NH ₄	2201	0.43	566	0.12	0.897	113	0.0	556	0.03	0.998
(NH ₄) ₂ SO ₄	NH ₄	3405	0.32	567	0.17	0.824	96	0.1	536	0.02	0.996
NH ₄ Cl	NH ₄	586 (C)	0.34	635	0.24	0.793	98	0.0	570	0.10	0.962
NH ₄ Cl	NH ₄	976	0.36	598	0.21	0.835	90	0.0	535	0.13	0.931
NH ₄ Cl	NH ₄	1958	0.19	642	0.30	0.841	57	0.0	556	0.06	0.989
NH ₄ Cl	NH ₄	2946	0.03	889	0.50	0.672	32	2.4	543	0.16	0.977
NH ₄ Cl	NH ₄	3923	0.02	889	0.47	0.682	22	3.3	505	0.17	0.993
NH ₄ Cl	NH ₄	5162	0.00	79	0.92	0.654	24	9.2	426	0.65	0.991
ZnSO ₄	Zn	0.26(C)	0.01	2202	0.55	0.278	4	4.5	261	0.17	0.935
ZnSO ₄	Zn	2	0.04	1662	0.41	0.767	69	1.4	830	0.07	0.992
ZnSO ₄	Zn	7	0.03	1988	0.45	0.753	80	1.4	821	0.11	0.983
ZnSO ₄	Zn	112	0.00	30016	0.53	0.310	28	2.0	655	0.04	0.998
ZnSO ₄	Zn	746	0.00	20620	0.63	0.308	12	7.9	522	0.07	0.994
ZnSO ₄	Zn	760	0.00	1725	0.75	0.530	18	21.4	477	0.03	0.998
ZnSO ₄	Zn	1676	-	-	-	-	-	-	-	-	-
ZnCl ₂	Zn	3 (C)	0.46	598	0.15	0.911	112	0.0	611	0.11	0.984
ZnCl ₂	Zn	10	0.49	615	0.15	0.909	117	0.0	638	0.12	0.992
ZnCl ₂	Zn	61	0.41	628	0.14	0.910	111	0.0	627	0.07	0.990
ZnCl ₂	Zn	87	0.35	659	0.16	0.895	102	0.0	640	0.05	0.984
ZnCl ₂	Zn	592	-	-	-	-	-	-	-	-	-
ZnCl ₂	Zn	1307	-	-	-	-	-	-	-	-	-
CuSO ₄	Cu	7 (C)	0.22	621	0.14	0.801	86	0.0	546	0.04	0.977
CuSO ₄	Cu	20	0.27	655	0.10	0.878	117	0.0	566	0.06	0.955
CuSO ₄	Cu	32	0.29	611	0.10	0.871	119	0.0	538	0.04	0.969

CuSO ₄	Cu	73	0.26	555	0.07	0.921	112	0.0	556	0.06	0.958
CuSO ₄	Cu	123	0.23	624	0.13	0.853	98	0.0	524	0.06	0.957
CuSO ₄	Cu	167	0.22	639	0.15	0.825	94	0.0	535	0.04	0.982
CuCl ₂	Cu	0.1 (C)	0.41	890	0.12	0.890	131	0.0	908	0.05	0.994
CuCl ₂	Cu	22	0.37	351	0.12	0.897	51	0.0	345	0.02	0.996
CuCl ₂	Cu	91	0.14	481	0.24	0.662	45	0.8	373	0.02	0.996
CuCl ₂	Cu	198	0.00	5679	0.65	0.230	23	2.9	256	0.11	0.955
CuCl ₂	Cu	501	-	-	-	-	-	-	-	-	-
CuCl ₂	Cu	804	-	-	-	-	-	-	-	-	-
Al ₂ (SO ₄) ₃	Al	4 (C)	0.22	621	0.14	0.801	86	0.0	546	0.04	0.977
Al ₂ (SO ₄) ₃	Al	85	0.27	611	0.10	0.884	106	0.0	544	0.05	0.955
Al ₂ (SO ₄) ₃	Al	210	0.24	614	0.11	0.884	94	0.0	532	0.07	0.929
Al ₂ (SO ₄) ₃	Al	771	0.10	572	0.28	0.668	60	0.7	395	0.04	0.989
Al ₂ (SO ₄) ₃	Al	1309	4.00	21	0.09	0.000	15	50.0	19	0.82	0.001
Al ₂ (SO ₄) ₃	Al	1534	-	-	-	-	-	-	-	-	-
AlCl ₃	Al	1 (C)	0.41	890	0.12	0.89	131	0.0	908	0.05	0.99
AlCl ₃	Al	82	0.49	941	0.08	0.954	148	0.0	974	0.44	0.996
AlCl ₃	Al	204	0.43	961	0.09	0.950	153	0.0	1008	0.07	0.996
AlCl ₃	Al	760	-	-	-	-	-	-	-	-	-
AlCl ₃	Al	1312	-	-	-	-	-	-	-	-	-
AlCl ₃	Al	1531	-	-	-	-	-	-	-	-	-
Al ₂ (SO ₄) ₃ +OH	Al	0 (C)	1.00	546	0.46	0.014	154	0.2	524	0.03	0.957
Al ₂ (SO ₄) ₃ +OH	Al	309	28.00	130	0.76	0.011	118	0.8	436	0.02	0.987
Al ₂ (SO ₄) ₃ +OH	Al	511	0.03	2602	0.77	0.025	95	0.9	398	0.02	0.997
Al ₂ (SO ₄) ₃ +OH	Al	607	23.21	41	0.89	0.012	80	1.2	373	0.02	0.994
HCl	pH	7.3 (C)	1.00	546	0.46	0.014	154	0.2	524	0.03	0.957
HCl	pH	6.6	1.00	536	0.45	0.019	131	0.1	516	0.03	0.974
HCl	pH	5.2	0.02	144	0.88	0.200	48	15.7	111	0.63	0.957
HCl	pH	4.5	0.03	31	0.92	0.135	25	25.1	118	0.34	0.992
NaCl	Na	3360 (C)	0.21	636	0.10	0.881	103	0.0	560	0.04	0.982
NaCl	Na	3915	0.26	700	0.09	0.895	134	0.0	643	0.04	0.978
NaCl	Na	4483	0.23	704	0.06	0.939	119	0.0	574	0.04	0.974
NaCl	Na	5054	0.22	705	0.08	0.921	112	0.0	579	0.04	0.977

NaCl	Na	5601	0.19	626	0.09	0.911	88	0.0	494	0.05	0.971
NaCl	Na	6165	0.09	913	0.18	0.774	56	0.0	512	0.02	0.995
Na ₂ SO ₄	Na	3360 (C)	0.21	636	0.10	0.881	103	0.0	560	0.04	0.982
Na ₂ SO ₄	Na	3430	0.25	637	0.05	0.952	115	0.0	538	0.05	0.958
Na ₂ SO ₄	Na	3499	0.27	595	0.05	0.962	116	0.0	512	0.05	0.951
Na ₂ SO ₄	Na	3639	0.27	575	0.05	0.956	114	0.0	499	0.05	0.959
Na ₂ SO ₄	Na	3846	0.31	523	0.03	0.978	118	0.0	466	0.05	0.947
Na ₂ SO ₄	Na	4054	0.27	593	0.05	0.954	115	0.0	511	0.05	0.952
NaNO ₃	Na	1370 (C)	0.19	784	0.41	0.775	64	0.0	748	0.04	0.995
NaNO ₃	Na	1403	0.05	1073	0.39	0.722	32	1.6	694	0.09	0.997
NaNO ₃	Na	1433	0.00	12884	0.47	0.658	23	3.3	657	0.10	0.988
NaNO ₃	Na	1463	0.00	9507	0.39	0.677	25	4.0	652	0.15	0.996
NaNO ₃	Na	1491	0.02	1689	0.39	0.704	26	2.9	713	0.13	0.993
NaNO ₃	Na	1527	0.01	4685	0.42	0.702	30	3.0	780	0.10	0.997
HgSO ₄	Hg	0.03 (C)	0.19	784	0.41	0.775	64	0.0	748	0.04	0.995
HgSO ₄	Hg	25	0.19	826	0.29	0.796	75	0.2	760	0.03	0.994
HgSO ₄	Hg	20	0.32	855	0.35	0.761	107	0.0	840	0.07	0.987
HgSO ₄	Hg	920	0.36	824	0.34	0.750	113	0.0	824	0.10	0.982
HgSO ₄	Hg	239	0.23	889	0.38	0.781	90	0.2	820	0.03	0.992
HgSO ₄	Hg	495	0.12	948	0.49	0.706	82	0.8	800	0.08	0.991
Na ₂ HAsO ₄	As	0.03 (C)	0.34	635	0.24	0.793	98	0.0	570	0.10	0.962
Na ₂ HAsO ₄	As	0.1	0.32	612	0.25	0.787	87	0.0	554	0.10	0.975
Na ₂ HAsO ₄	As	0.4	0.23	562	0.24	0.869	53	0.0	507	0.12	0.984
Na ₂ HAsO ₄	As	2	0.24	594	0.22	0.901	56	0.0	511	0.16	0.920
Na ₂ HAsO ₄	As	4	0.15	679	0.26	0.924	41	0.0	552	0.14	0.932
Na ₂ HAsO ₄	As	17	0.03	883	0.48	0.761	25	2.2	536	0.18	0.982

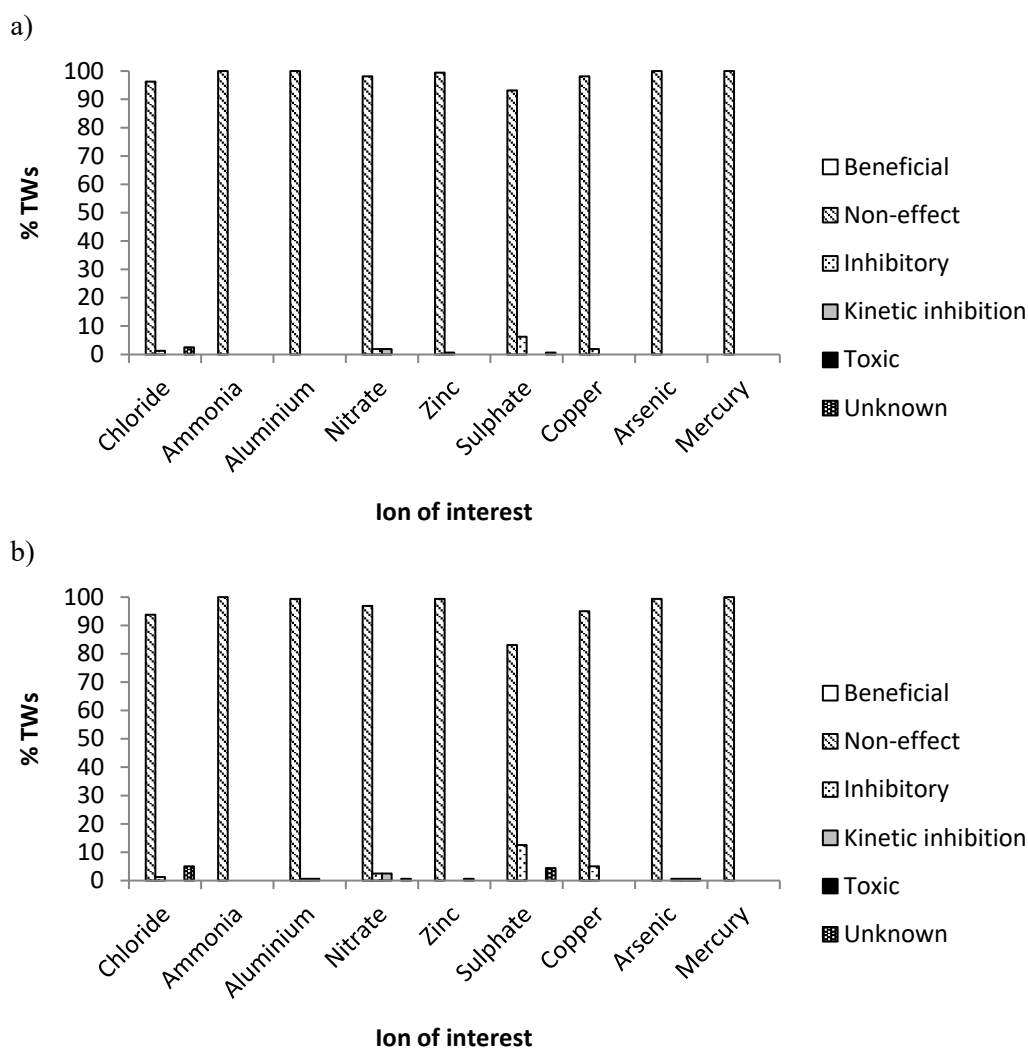


Figure S1. Percentage of TWs within the 160 studied that will incur in beneficial, non-effect, yield inhibition, kinetic inhibition, toxic or unknown impact if dosed to sewage sludge anaerobic digesters as 10% of the feedstock (a) or 30% of the feedstock (%) (b).

Description of Figure S1:

A more detailed investigation was conducted for scenarios with TW as 10% and 30% of the feedstock (10/90 and 30/70), for which potential impact as beneficial, non-effect, yield inhibition for methane yields, kinetic inhibition and toxic were investigated for the individual constituents present in the TWs. Another division called unknown was added to refer to conditions above the maximum concentration studied where no toxicity was observed. The 30/70 scenario was selected because it is reported to be the average spare capacity in municipal wastewater treatment plants digesters in Europe [35]. The 10/90 scenario was selected as 10% is commonly accepted by wastewater utilities to be the maximum spare capacity that could be allocated to TWs digestion without hindering sewage sludge treatment.

References of Supplementary Materials

1. Angelidaki I, Ahring BK. Thermophilic anaerobic digestion of livestock waste: the effect of ammonia. *Appl Microbiol Biotechnol* 1993;38:560–4. <https://doi.org/10.1007/BF00242955>.
2. Borja R, Sánchez E, Weiland P. Influence of ammonia concentration on thermophilic anaerobic digestion of cattle manure in upflow anaerobic sludge blanket (UASB) reactors. *Process Biochem* 1996;31:477–83. [https://doi.org/http://dx.doi.org/10.1016/0032-9592\(95\)00099-2](https://doi.org/http://dx.doi.org/10.1016/0032-9592(95)00099-2).
3. McCarty PL. Anaerobic waste treatment fundamentals. *Public Work* 1964;95:107–12.
4. Nielsen HB, Angelidaki I. Strategies for optimizing recovery of the biogas process following ammonia inhibition. *Bioresour Technol* 2008;99:7995–8001. <https://doi.org/https://doi.org/10.1016/j.biortech.2008.03.049>.
5. Bhattacharya SK, Parkin GF. The Effect of Ammonia on Methane Fermentation Processes. *J (Water Pollut Control Fed)* 1989;61:55–9.
6. Boardman GD, McVeigh PJ. Use of UASB technology to treat crab processing wastewaters. *J Environ Eng* 1997;123:776.
7. Bujoczek G, Oleszkiewicz J, Sparling R, Cenkowski S. High Solid Anaerobic Digestion of Chicken Manure. *J Agric Eng Res* 2000;76:51–60. <https://doi.org/http://dx.doi.org/10.1006/jaer.2000.0529>.
8. Tugtas AE, Pavlostathis SG. Inhibitory effects of nitrogen oxides on a mixed methanogenic culture. *Biotechnol Bioeng* 2007;96:444–55. <https://doi.org/10.1002/bit.21105>.
9. Akunna JC, Bizeau C, Moletta R. Nitrate reduction by anaerobic sludge using glucose at various nitrate concentrations: Ammonification, denitrification and methanogenic activities. *Environ Technol* 1994;15:41–9. <https://doi.org/10.1080/09593339409385402>.
10. Klüber HD, Conrad R. Inhibitory effects of nitrate, nitrite, NO and N₂O on methanogenesis by *Methanosarcina barkeri* and *Methanobacterium bryantii*. *FEMS Microbiol Ecol* 1998;25:331–9. <https://doi.org/10.1111/j.1574-6941.1998.tb00484.x>.
11. Yi X-H, Wan J, Ma Y, Wang Y, Guan Z, Jing D-D. Structure and Succession of Bacterial Communities of the Granular Sludge during the Initial Stage of the Simultaneous Denitrification and Methanogenesis Process. *Water, Air, Soil Pollut* 2017;228:121. <https://doi.org/10.1007/s11270-016-3168-5>.
12. Clarens M, Bernet N, Delgenès J-P, Moletta R. Effects of nitrogen oxides and denitrification by *Pseudomonas stutzeri* on acetotrophic methanogenesis by *Methanosarcina mazei*. *FEMS Microbiol Ecol* 1998;25:271–6. <https://doi.org/10.1111/j.1574-6941.1998.tb00479.x>.
13. Borges LI, López-Vazquez CM, García H, van Lier JB. Nitrite reduction and methanogenesis in a single-stage UASB reactor. *Water Sci Technol* 2015;72:2236 LP – 2242.
14. Yan G, Wang J, Guo S. Anaerobic Biochemical Treatment of Wastewater Containing Highly Concentrated Organic Cyanogen. *Energy Sources, Part A Recover Util Environ Eff* 2007;29:529–35. <https://doi.org/10.1080/009083190966342>.
15. Gijzen HJ, Bernal E, Ferrer H. Cyanide toxicity and cyanide degradation in anaerobic wastewater treatment. *Water Res* 2000;34:2447–54. [https://doi.org/10.1016/S0043-1354\(99\)00418-2](https://doi.org/10.1016/S0043-1354(99)00418-2).
16. Cabirol N, Barragán EJ, Durán A, Noyola A. Effect of aluminium and sulphate on anaerobic digestion of sludge from wastewater enhanced primary treatment. *Water Sci Technol* 2003;48:235 LP – 240.
17. Hu Y, Jing Z, Sudo Y, Niu Q, Du J, Wu J, et al. Effect of influent COD/SO₄²⁻ ratios on UASB treatment of a synthetic sulfate-containing wastewater. *Chemosphere* 2015;130:24–33. <https://doi.org/https://doi.org/10.1016/j.chemosphere.2015.02.019>.
18. Soto M, Méndez R, Lema JM. Sodium inhibition and sulphate reduction in the anaerobic treatment of mussel processing wastewaters. *J Chem Technol Biotechnol* 1993;58:1–7. <https://doi.org/10.1002/jctb.280580102>.
19. Jeong T-Y, Chung H-K, Yeom SH, Choi SS. Analysis of methane production inhibition for treatment of sewage sludge containing sulfate using an anaerobic continuous degradation process. *Korean J Chem Eng* 2009;26:1319–22. <https://doi.org/10.1007/s11814-009-0229-0>.
20. Abdel-Shafy HI, Mansour MSM. Biogas production as affected by heavy metals in the

- anaerobic digestion of sludge. Egypt J Pet 2014;23:409–17. <https://doi.org/https://doi.org/10.1016/j.ejpe.2014.09.009>.
21. Ahring BK, Westermann P. Sensitivity of thermophilic methanogenic bacteria to heavy metals. Curr Microbiol 1985;12:273–6. <https://doi.org/10.1007/BF01567977>.
 22. Li C, Fang HHP. Inhibition of heavy metals on fermentative hydrogen production by granular sludge. Chemosphere 2007;67:668–73. <https://doi.org/https://doi.org/10.1016/j.chemosphere.2006.11.005>.
 23. Altaş L. Inhibitory effect of heavy metals on methane-producing anaerobic granular sludge. J Hazard Mater 2009;162:1551–6. <https://doi.org/https://doi.org/10.1016/j.jhazmat.2008.06.048>.
 24. Yu B, Lou Z, Zhang D, Shan A, Yuan H, Zhu N, et al. Variations of organic matters and microbial community in thermophilic anaerobic digestion of waste activated sludge with the addition of ferric salts. Bioresour Technol 2015;179:291–8. <https://doi.org/https://doi.org/10.1016/j.biortech.2014.12.011>.
 25. Van Bodegom PM, Scholten JCM, Stams AJM. Direct inhibition of methanogenesis by ferric iron. FEMS Microbiol Ecol 2004;49:261–8. <https://doi.org/10.1016/j.femsec.2004.03.017>.
 26. Colussi I, Cortesi A, Vedova L Della, Gallo V, Robles FKC. Start-up procedures and analysis of heavy metals inhibition on methanogenic activity in EGSB reactor. Bioresour Technol 2009;100:6290–4. <https://doi.org/https://doi.org/10.1016/j.biortech.2009.07.041>.
 27. Lin C-Y, Shei S-H. Heavy metal effects on fermentative hydrogen production using natural mixed microflora. Int J Hydrogen Energy 2008;33:587–93. <https://doi.org/https://doi.org/10.1016/j.ijhydene.2007.09.030>.
 28. O'Connor OA, Young LY. Toxicity and anaerobic biodegradability of substituted phenols under methanogenic conditions. Environ Toxicol Chem 1989;8:853–62. <https://doi.org/10.1002/etc.5620081003>.
 29. Hansen KH, Angelidaki I, Ahring BK. Anaerobic digestion of swine manure: Inhibition by ammonia. Water Res 1998;32:5–12. [https://doi.org/http://dx.doi.org/10.1016/S0043-1354\(97\)00201-7](https://doi.org/http://dx.doi.org/10.1016/S0043-1354(97)00201-7).
 30. Sierra-Alvarez R, Cortinas I, Yenal U, Field JA. Methanogenic Inhibition by Arsenic Compounds. Appl Environ Microbiol 2004;70:5688–91. <https://doi.org/10.1128/AEM.70.9.5688-5691.2004>.
 31. Lenz M, Janzen N, Lens PNL. Selenium oxyanion inhibition of hydrogenotrophic and acetoclastic methanogenesis. Chemosphere 2008;73:383–8. <https://doi.org/10.1016/J.CHEMOSPHERE.2008.05.059>.
 32. Ochoa-Herrera V, Banihani Q, León G, Khatri C, Field JA, Sierra-Alvarez R. Toxicity of fluoride to microorganisms in biological wastewater treatment systems. Water Res 2009;43:3177–86. <https://doi.org/https://doi.org/10.1016/j.watres.2009.04.032>.
 33. Vijayaraghavan K, Ramanujam TK. Effect of chloride and condensable tannin in anaerobic degradation of tannery wastewaters. Bioprocess Eng 1999;20:499. <https://doi.org/10.1007/s004490050621>.
 34. Hernandez JE, Edyvean RGJ. Comparison between a two-stage and single-stage digesters when treating a synthetic wastewater contaminated with phenol. Water SA 2011. <https://doi.org/10.4314/wsa.v37i1.64103>.
 35. Maragkaki AE, Fountoulakis M, Kyriakou A, Lasaridi K, Manios T. Boosting biogas production from sewage sludge by adding small amount of agro-industrial by-products and food waste residues. Waste Manag 2018;71:605–11. <https://doi.org/10.1016/j.wasman.2017.04.024>.