

Review **Cold Plasma-Assisted Extraction of Phytochemicals: A Review**

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Abstract: In recent years, there has been growing interest in bioactive plant compounds for their beneficial effects on health and for their potential in reducing the risk of developing certain diseases such as cancer, cardiovascular diseases, and neurodegenerative disorders. The extraction techniques conventionally used to obtain these phytocompounds, however, due to the use of toxic solvents and high temperatures, tend to be supplanted by innovative and unconventional techniques, in line with the demand for environmental and economic sustainability of new chemical processes. Among nonthermal technologies, cold plasma (CP), which has been successfully used for some years in the food industry as a treatment to improve food shelf life, seems to be one of the most promising solutions in green extraction processes. CP is characterized by its low environmental impact, low cost, and better extraction yield of phytochemicals, saving time, energy, and solvents compared with other classical extraction processes. In light of these considerations, this review aims to provide an overview of the potential and critical issues related to the use of CP in the extraction of phytochemicals, particularly polyphenols and essential oils. To review the current knowledge status and future insights of CP in this sector, a bibliometric study, providing quantitative information on the research activity based on the available published scientific literature, was carried out by the VOSviewer software (v. 1.6.18). Scientometric analysis has seen an increase in scientific studies over the past two years, underlining the growing interest of the scientific community in this natural substance extraction technique. The literature studies analyzed have shown that, in general, the use of CP was able to increase the yield of essential oil and polyphenols. Furthermore, the composition of the phytoextract obtained with CP would appear to be influenced by process parameters such as intensity (power and voltage), treatment time, and the working gas used. In general, the studies analyzed showed that the best yields in terms of total polyphenols and the antioxidant and antimicrobial properties of the phytoextracts were obtained using mild process conditions and nitrogen as the working gas. The use of CP as a non-conventional extraction technique is very recent, and further studies are needed to better understand the optimal process conditions to be adopted, and above all, in-depth studies are needed to better understand the mechanisms of plasma–plant matrix interaction to verify the possibility of any side reactions that could generate, in a highly oxidative environment, potentially hazardous substances, which would limit the exploitation of this technique at the industrial level.

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1. Introduction

In recent years, growing consumer demand for natural-based products in the personal care and well-being sectors has sparked renewed interest in medicinal and aromatic plants as sources of bioactive and functional components, especially in the wake of the recent pandemic events. Globally, the phytochemicals market is experiencing a significant positive trend lately, with a Compound Annual Growth Rate (CARG) of about 8.5%, corresponding to a trade value of about USD 7 million in 2023 [\[1\]](#page-28-0). Phytochemicals (term generically referring to the secondary metabolites of plants such as polyphenols) are non-nutrient bioactive components of plant origin recognized to be of great interest for their beneficial effects on human health and beyond. They include a plethora of different chemical structures that can be grouped into polyphenols, terpenoids, alkaloids, glucosinolates, etc. [\[2\]](#page-28-1) (Table [1\)](#page-1-0).

Table 1. Main phytochemical classes and related biological properties (adapted from [\[3\]](#page-28-2)).

Yet, the choice of which extraction method needs to be applied to the plant matrix for their recovery is the first critical issue in maintaining their quality and biological potential unaltered. Extraction can be described as a critical transport phenomenon that transfers matrix components to the solvent $[4–7]$ $[4–7]$, which can be carefully controlled to preserve odor, flavor, and biologically active compounds $[8-11]$ $[8-11]$. Table [2](#page-2-0) shows an overview of the advantages and disadvantages of different approaches for the extraction of phytochemicals. The biological potential of natural extracts is influenced by both the technology used to obtain them and the quality of the plant matrix used [\[7,](#page-28-4)[10,](#page-28-7)[12](#page-28-8)[,13\]](#page-28-9). Conventional extraction strategies to isolate bioactive compounds from plants are based on the use of organic

solvents, most of which have a negative environmental impact [\[14–](#page-28-10)[18\]](#page-28-11). These extraction methods, including steam distillation, hydro distillation, and liquid–solvent extraction, require high temperatures, long processing times, and at times many extraction steps, limited extraction efficiency, safety issues, and environmental concerns regarding the use of toxic solvents [\[19–](#page-28-12)[24\]](#page-29-0). Recently, green, and non-thermal alternatives such as ultrasounds, pressurized liquids, supercritical fluid extraction, pulsed electric fields, and cold plasma (CP) have been introduced to partially overcome these problems [\[7](#page-28-4)[,25–](#page-29-1)[28\]](#page-29-2). These new methods can reduce energy consumption, process time, use of organic solvents, and loss of nutrient/nutraceutical compounds, enabling high-quality functional plant extracts [\[14](#page-28-10)[,29–](#page-29-3)[31\]](#page-29-4).

Table 2. Comparison of different extraction methods for biologically active compounds.

ex

Table 2. *Cont.*

• Accurate production of usable plasmas • Removal of food volatile compounds • Enzymes deactivate in solid and liquid

food processing industries

CP is an emerging and relatively unexplored non-thermal technology with promising applications in various areas of food processing [\[72](#page-30-9)[–74\]](#page-30-10). It is regarded as an innovative method, which uses highly reactive charged molecules and gaseous species to inactivate contaminating microorganisms in food. From a chemical-physical perspective, plasma represents the fourth state of matter [\[73,](#page-30-11)[75\]](#page-30-12). When the energy of molecules in a system increases, solids turn into liquids and liquids into gases. The intermolecular configuration is altered. A further increase in the energy of gases causes all interactions to vanish, releasing positive and negative ions and causing some molecules and atoms to ionize, giving rise to plasma, which can then be defined as an ionized gas [\[72–](#page-30-9)[74\]](#page-30-10). Currently, CP is used to improve the shelf life of food [\[75,](#page-30-12)[76\]](#page-30-13), reduce food contamination [\[77\]](#page-30-14), and improve the functional properties of proteins [\[78\]](#page-31-0). It is also used for structural modification [\[79\]](#page-31-1), enzyme inactivation [\[80\]](#page-31-2), removal of toxins [\[81\]](#page-31-3), change of food packaging characteristics [\[82\]](#page-31-4), wastewater treatment [\[8,](#page-28-5)[83,](#page-31-5)[84\]](#page-31-6), control of biofilm [\[85\]](#page-31-7), and food waste valorization [\[24,](#page-29-0)[86\]](#page-31-8) (Figure [1\)](#page-4-0). CP can also reduce the unpleasant effects of thermal treatments, such as discoloration and loss of nutrients [\[72,](#page-30-9)[87\]](#page-31-9).

In recent years, a growing number of scientific studies have highlighted the potential of using CP to increase the extraction yield of phytochemicals due to its non-thermal properties and energy efficiency [\[19,](#page-28-12)[21](#page-28-13)[,30,](#page-29-14)[77](#page-30-14)[,88](#page-31-10)[,89\]](#page-31-11), ensuring faster times, reducing the degradation of heat-sensitive substances, and achieving sophisticated extraction without impacting the environment. However, to date, most studies have focused on the microbial decontamination properties of CP, while there are few studies on the effect of CP treatment on the production of plant extracts and their phytochemical profile and relative biological potential. Thus, this work aims to review up-to-date information on the application of CP technology for the extraction of valuable plant compounds, considering its effect on (a) extraction yield, (b) total phenolic content and phytochemical profile, (c) antioxidant capacity, and (d) antimicrobial properties of the obtained extracts.

Figure 1. Applications of cold plasma in food-related fields. **Figure 1.** Applications of cold plasma in food-related fields.

In recent years, a growing number of scientific studies have highlighted the potential **2. Scientometric Analysis**

To obtain a general overview of international studies published up to the 24 April 2023 on the use of CP in the extraction of phytochemicals, the scientific literature was analyzed using the Scopus database (Elsevier) and the VOSviewer software (v. 1.6.18).

The Scopus search was performed (Search within Article title, Abstract, Keywords) by typing the keywords (Search documents): "cold plasma" AND "extraction". The resulting 124 papers were examined, selecting for analysis only those documents that had all keywords typed between the title, abstract, or author keywords. This sub-selection resulted in the collection of 77 documents, which constituted the bibliographic dataset, first analyzed using the Scopus Analyze search results function (Documents by year, Documents by type, Documents by subject area, and Documents by country or territory), from which all Scopus information data analyzed with the VOSviewer software were exported.

Figure S1a shows the trend in the number of articles by year of publication, highlighting that the application of CP as an extraction technique is a very recent research topic, developed mainly in the last 10 years.

Looking at the graph in detail, it can be seen that from 2012 to 2016, the trend of publications has been constant, even the number of articles has been very low. There has been a significant increase in publications since 2017, with the highest peak reached in 2022, with 23 total articles. This value is likely to be exceeded in the current year, which has produced 12 papers up to the time of the Scopus search (April 2023), indicating that the research field has attracted even more interest in the last three years. Figure S1b also points out that CP is an innovative, start-up research topic, with 68.8% of the papers consisting of articles, while only 22.1% and 6.5% of the papers consist of reviews (17 papers) or book chapters (5 papers). Furthermore, Figure S1c shows the highest interest of the scientific community in the subject area 'Agricultural and Biological Science' (31.5%—52 articles), with the highest number of studies concerning the use of CP for the extraction of bioactive compounds from various plant/agricultural matrices published in journals belonging to the subject area "Biochemistry, Genetics, and Molecular Biology" (7.9%—13 articles). The second and third areas of interest are represented by "Engineering" (12.7%—21 documents) and "Chemical Engineering" (10.9%—18 documents), underscoring the engineering effort

required to develop CP technology for chemical extraction. The bar graph shown in Figure S1d shows the number of papers for each country/territory and, interestingly, highlights that the use of CP as an extractive technique has so far attracted the interest of mainly three nations, all from Asia, with China being the top country in terms of related scientific production (21 papers), followed by Iran (13 papers) and India (12 papers).To explore the knowledge structure in this field of research, the extracted bibliographic dataset was analyzed with VOSviewer to calculate and display a map of the keyword co-occurrence network. From the dataset, the software extracted 974 total keywords, of which 39 exceeded the predefined minimum threshold of 5 occurrences. The keyword "article" was manually excluded from the final calculation to avoid any bias in the topology of the graph. All other software settings were left as default.

The Network Visualization in VOSviewer shows the keyword co-occurrence network map, which graphically represents the keywords extracted from the bibliographical dataset as points or nodes, and the co-occurrence of two keywords into the same publications as a line or link. The larger the area of a node, the higher the absolute number of occurrences of a keyword in the dataset. Nodes proximity is directly related to keywords co-occurrence: the closer two nodes are, the more the two keywords are related by similar research publications. VOSviewer identified 3 clusters (see Table S1 for cluster composition), each highlighted by a different color (red, green, light blue) in the Network Visualization map and by the same colors in the Density Visualization map (Cluster density; CS) (Figure [2a](#page-6-0),b).

The network map shows a general topology of highly interconnected nodes among the three clusters, with the absence of any overlap and the absence of a single central node, but with two nearby central nodes represented by the keywords 'cold plasma' (30 occurrences) and 'extraction' (27 occurrences), both in cluster 1 (CS_1). The small size of all nodes (low number of total occurrences of each keyword) and the low number of nodes that make up the graph, together with the absence of areas of overlap between clusters, confirm that the use of CP for chemical extraction is a very young field of research with strong growth potential for the future. CS_1 has "cold plasma" as the most recurrent keyword in the cluster (n. of occurrences 30), which is related to almost all nodes (keywords) in the entire graph. CS_1 is composed of 14 keywords extracted from publications on the characterization of the effects of CP technology on complex matrices, subsequently used for chemical extraction at the microscopical and chemical level: "extraction", "plasma application", "scanning electron microscopy", "gas chromatography", and "mass spectrometry". In CS_2, the 'electric fields' keyword is the most recurrent (9) out of a total of 13 keywords, all related to the testing of different extraction technologies coupled with CP for different applications: "pulsed electric field", "hydrostatic pressure", "ultrasound", "ultrasonics", "non-thermal processing", and "food handling". CS_3, in which the most recurrent keyword is 'antioxidants' with 15 total occurrences in the analyzed documents, is composed of 11 nodes, graphically representing research on the use of plasma gas for the chemical extraction of phenolic compounds and antioxidants from plant matrices: "plasma gas", "polyphenols", "phenolic compounds", "phenols", "plant extract", and "antioxidant activity".

The overlay view of the network map drawn by VOSviewer (Figure [3\)](#page-7-0) shows, for each keyword, the average year of publication (AYP), with a color scale ranging from dark blue, for older publications, to bright yellow, for more recent ones.

The analysis conducted suggests that the earliest applications of CP for the extraction of phytocompounds were in 'food processing' using 'ultrasound' and/or 'electric fields'. In contrast, the results highlight very recent applications studies on cold plasma for the 'extraction' of 'phenolic compounds' and 'anthocyanins' from complex matrices, analyzing the effects of this technology using 'scanning electron microscopy'.

VOSviewer

hydrostatic pressur cold plasma plasma gase plasma gas antioxidants **& VOSviewer**

(**a**)

(**b**)

Figure 2. (a) Network Visualization of the keywords co-occurrence network map. (b) Density visualization (Cluster density setting). Each density representing a cluster has a different color $r_{\rm sc}$ (CS₋₂): $r_{\rm acc}$ creen; CS₋₂: light blue). All the visualizations were generated by the VOSViewer (CS_1: red; CS_2: green; CS_3: light blue). All the visualizations were generated by the VOSViewer software from the search results in Scopus (performed on 24 April 2023) after the search: "cold plasma" AND "extraction" followed by the sub-selection of the dataset.

blue, for older publications, to bright yellow, for more recent ones.

VOSviewer

Figure 3. Overlay visualization of the keyword co-occurrence network map which highlights the **Figure 3.** Overlay visualization of the keyword co-occurrence network map which highlights the average year of publication of the keywords in the map (AYP). All the visualizations were generated by the VOSViewer software from the search results in Scopus (performed on 24 April 2023) after the by the VOSViewer software from the search results in Scopus (performed on 24 April 2023) after the search: "cold plasma" AND "extraction" followed by the sub-selection of the dataset. search: "cold plasma" AND "extraction" followed by the sub-selection of the dataset.

3. Principles, Types, and Sources of Cold Plasma Production

Plasma is considered the fourth state of the matter. It is a partially or fully ionized gas, macroscopically neutral, in which the species that can be identified are the molecules of the gas, fragments of the same (atoms, positive and negative ions, and radicals), and reaction products between all the species present [\[90\]](#page-31-12) (Figure [4\)](#page-8-0); these may be in different states of excitation depending on how the energy is distributed within the system.

Any gas at a temperature above 0° K contains a certain concentration of charged species (electrons and ions), but it is only considered a plasma if the concentration of charged species is such as to affect its motion. Any form of energy, including UV and gamma radiations, electrical energy, and electromagnetic radiation, has the potential to generate plasma. Plasma can be classified based on temperature (hot plasma or cold plasma), pressure (low, atmospheric, or high pressure), working gas (air, oxygen, argon, or helium), and mode of production. Non-thermal atmospheric pressure plasma (non-equilibrium plasma) is commonly described as CP, in which the temperature of the ions and neutral atoms is significantly lower than that of the electrons. Thus, it is characterized by a nonuniform distribution of energy among the constituent particles. Since the ions and neutral atoms remain relatively cold, CP can be successfully used with heat-sensitive compounds.

CP emits light with wavelengths in both the visible and ultraviolet spectral regions. In addition to the emission of UV radiation (wavelength range: 100–380 nm), an important property of low-temperature plasma is the presence of high-energy, highly reactive electrons, which cause several chemical and physical processes such as oxidation, the excitation of atoms and molecules, the production of free radicals, and UV photons that can decompose covalent bonds and produce many chemical reactions, such as surface etching (creation of pores/tissue damage), depolymerization (formation of new compounds through the breakdown of cell wall polysaccharides), and cross-linking (cleavage of C-OH polymeric chains for adding new C-O-C bond by eliminating water) [\[72,](#page-30-9)[87\]](#page-31-9). In addition, UV

photons can increase the activity of specific enzymes such as phenylalanine ammonia-lyase, thereby increasing the amount of total phenols extracted from the plant matrix [\[91\]](#page-31-13). Plasma can be generated artificially by supplying a gas with sufficiently high energy employing lasers, shock waves, electric arcs, and electric and magnetic fields, i.e., by applying energy to a gas in such a way as to reorganize the electronic structure of species (atoms, molecules) and produce excited species and ions. One of the most common ways to artificially create and maintain plasma is by using an electric discharge in a gas. Other ways include the use of jet plasma (a jet plasma system is connected to a power source, which induces ionization in the surrounding gas) and microwave plasma (plasma torches with two dielectric tubes that let the gas pass through). In the case of CP, so-called non-thermal discharges are used [\[30,](#page-29-14)[72,](#page-30-9)[92\]](#page-31-14). The two main types of non-thermal discharge at atmospheric pressure are corona discharge and dielectric barrier discharge (DBD) [\[93\]](#page-31-15). DBD is the commonly used **3. Principles, Types, and Sources of Cold Plasma Production**

system for the extraction of bioactive compounds from plant matrices $[20,72,88,94]$ $[20,72,88,94]$ $[20,72,88,94]$ $[20,72,88,94]$. In a DBD system, an electrical discharge is generated by two electrodes that are separated by a dielectric layer (a material with high electrical resistance) (Figure 5). It operates at approximately atmospheric pressure (0.1–1 atm), at frequencies up to 104 Hz, and alternating voltages up to $100 \, \mathrm{kV}$ [\[72\]](#page-30-9).

Figure 4. Plasma reactive species. **Figure 4.** Plasma reactive species.

The most important parameters influencing the CP process are the plasma generation voltage or power (higher power increases the electron density, generating more reactive species capable of interacting with the matrix to be treated), the amount of material to be treated (efficiency decreases as the amount increases), the duration of exposure (time), the gas pressure (affects the rate of plasma volatilization), the gas flow rate (higher gas flow $\overline{}$ rate can improve treatment efficiency), and the type of feed gas used $[30,92]$ $[30,92]$. The degree to which reactive species are produced depends crucially on the gas used to generate the plasma (working gas). Plasma gases, such as oxygen, nitrogen, and dry air, are commonly plasma in which the ions of the ions o employed in the food processing industry [\[72,](#page-30-9)[87,](#page-31-9)[89\]](#page-31-11). However, gas mixtures such as $W(\lambda, \mu) \propto \lambda / 2$ He/N_2 , He/O_2 , N_2/N_2O , Ar/O_2 , etc., can also be used.

 $\frac{\text{ln}(1+\sqrt{2}, 1+\sqrt{2}, 1+\sqrt{2}, \text{ln}(1-\sqrt{2})}{\text{ln}(\sqrt{2})}$, the $\sqrt{2}$ and neutral atoms remain relatively cold, ϵ cold, ϵ can be successfully used with heat-sensitively used wit extracting phytochemicals is the frequency of plasma generation, due to the inverse relation-

ship between the frequency and the activity of certain crucial enzymes such as peroxidase and polyphenol oxidase [\[95\]](#page-31-17).

Figure 5. Dielectric barrier discharge system to generate cold plasma. **Figure 5.** Dielectric barrier discharge system to generate cold plasma.

The most important parameters influencing the CP process are the plasma generation **4. Mode of Action of Cold Plasma on the Plant Matrix and Its Critical Issues**

The extraction yield of plant components can be affected by several factors [\[30](#page-29-14)[,76](#page-30-13)[,92\]](#page-31-14), including the level of disruption of the cell wall, CP process parameters (type of extraction method), plant species, material surfaces, and composition of the bioactive compounds that occurred in the plant (Table [3\)](#page-10-0). It has been demonstrated that CP treatment can cause changes in the surface physical properties of the plant matrix, with the formation of cracks and depressions on its surface that allow better outward release of the compounds of interest, thus increasing the extraction yield. In addition, CP appears to be able to increase the hydrophilicity of the matrix surface through the degradation of the cuticular layer, the Hydrogrammary of the final startive through the digenometric of the calculation in year, thereby facilitating the diffusion of internal molecules to the solvent and consequently increasing the extractability of hydrophilic compounds, such as phenols. These mechanisms are attributable to the action of plasma-generated reactive species, mainly ROS and RNS, which can modify the surfaces of materials and change their functional groups, as stated previously.

Table 3. An overview of the effects of cold plasma on the extraction yield and quality of plant extracts.

* HVACP: high voltage atmospheric cold plasma; TPC: total phenolic content; PC: phenolic compounds; DBD: dielectric barrier discharge; GDP: glow discharge plasma; EGCG: epigallocatechin gallate; LPCP: low-pressure cold plasma; NA: not available.

Figure [6](#page-15-0) shows the possible mechanism of action of CP responsible for the release of bioactive substances from plant matrices. bioactive substances from plant matrices.

Figure 6. Possible release mechanisms of phytochemicals through the interaction between cold **Figure 6.** Possible release mechanisms of phytochemicals through the interaction between cold plasma and plant matrix.

This mechanism is supported by several findings. Rashid et al. (2020) observed that This mechanism is supported by several findings. Rashid et al. (2020) observed that the extraction yield of galactomannan from fenugreek increased before (crushed dry the extraction yield of galactomannan from fenugreek increased before (crushed dry seeds) and after (crushed seeds soaked in extraction solutions) treatment with high voltage cold age cold atmospheric plasma (HVACP) by 67% and 122%, respectively, in a statistically atmospheric plasma (HVACP) by 67% and 122%, respectively, in a statistically significant manner. They concluded that the increase in dry extract yield could be due to HVACPinduced changes in the surface morphology of fenugreek seeds. Exposure to plasma active species resulted in specific fragments and cracks in the epidermal structure of the seeds \overrightarrow{f} (Figure [7b](#page-16-0),d), which were able to retain and absorb the extractive solution more effectively than the control, which showed a smooth, intact surface on the microscopic investigation (Figure [7a](#page-16-0),c) [\[112\]](#page-32-20).

Figure 7. Scanning electron microscope images of (a) untreated dry fenugreek seeds, (b) HVACPtreated dry fenugreek seeds, (**c**) untreated soaked fenugreek seeds, and (**d**) HVACP-treated soaked fenugreek seeds. Adapted with permission from Ref. [\[112\]](#page-32-20), 2020, Elsevier.

Ebadi et al. (2019) showed that the yield of lemon verbena leaf essential oil was im-Ebadi et al. (2019) showed that the yield of lemon verbena leaf essential oil was im p_{total} of p_{CUT} showed black plasma (LPCP) treatment p_{total} . This result was first proved by the low-pressure cold plasma (LPCP) treatment [\[118\]](#page-32-21). This result was confirmed
here the non-produced above the constitution of the low-plate of 2014) found that plasma treatment by other researchers' observations. Kodama et al. (2014) found that plasma treatment that the extraction efficiency of essential oil from fennel seeds (*Foeniculum vulgare* Mill.) and mint leaves (*Mentha spicata* L.) can be influenced by DBD CP treatment followed by hand mint leaves (*Mentha spicata* L.) can be influenced by DBD CP treatment followed by and nunt reaves (*Wentha spidua E.*) can be inhactived by *BBB* Cr treatment followed by hydrodistillation [\[115\]](#page-32-23). In particular, they observed a higher extraction yield with a 15 kV my discussimation [110]. In particular, they observed a higher extraction yield with a 15 KV plasma treatment applied for 5 min (1.83 (% v/w)) than with a 23 kV treatment applied for 17 min (1.81 (% v/w)). According to Figure [8,](#page-17-0) CP disrupted the structural integrity for 17 min (1.81 (% v/w)). According to Figure 8, CP disrupted the structural integrity of the seed coat, disrupting the surface oil glands and creating permeability during dis-
of the seed coat, disrupting the surface oil glands and creating permeability during disof the seed estay, distributed in a higher oil extraction yield. In contrast, intense treatments, tillation, which resulted in a higher oil extraction yield. In contrast, intense treatments, exposure, and leading the definition of the contraction plasma in contract, and leading to a definition of due
due to prolonged plasma exposure, adversely affected the glands, producing essential oil and the process practical expression, and the same authors, [\[113\]](#page-32-24), in a previous paper, vapors and leading to a decrease in yield. The same authors, [113], in a previous paper, CP treatment was able to increase the oil yield extracted from *Camelina sativa* seeds by 31.5%, showed that the application of CP treatment was able to increase the oil yield extracted correlation to the results obtained to the results of the more intense and higher treatment and processing $\cos \theta$ is 21.5% correlation the more intense and processing $\cos \theta$ from *Camelina sativa* seeds by 31.5%, correlating the results obtained to the more intense and increased the essential oil yield of lemon peel [\[119\]](#page-32-22), while Rezaei et al. (2021) reported higher treatment and processing time used (18 kV and 16 min), which could further damage the cell wall [\[113](#page-32-24)[,123\]](#page-32-25). Similarly, Afshar et al. (2022) indicated that the extraction of sunflower and sesame oil seeds by non-thermal plasma in oxygen and nitrogen atmospheres increased the extraction yield through wall degradation of the seed cell [114,124]. Also, Pragna et al. (2019) and Kodama and Sekiguchi (2015) observed a similar effect of DBD CP-assisted hydrodistillation on the enhancement of lemon peel extraction yield [\[116,](#page-32-28)[119\]](#page-32-22).

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Magnification of 200× Magnification of 800×

Figure 8. Scanning electron microscope images of (a,b) Control (spearmint leaf), (c,d) CP-treated leaf (15 kV, 5 min), (e,f) CP-treated leaf treated leaf (23 kV, 17 min) at different magnifications. Adapted with permission from Ref. $[115]$, 2021, Elsevier.

In addition, according to Fernandes and Rodrigues (2021), CP can affect the final polyphenol content and profile of the plant extract, either positively or negatively, through other different mechanisms in addition to those described so far: (i) depolymerization of tannins, and (ii) oxidation of phenolic structures [\[90\]](#page-31-12). Lastly, it has been shown that CP treatment can in some cases produce a stress-induced accumulation of phenolic antioxidants, thereby increasing the extraction yield by acting on the regulation of gene expression and the activity of crucial enzymes of the phenylpropanoid pathway [\[90\]](#page-31-12). Conversely, one of the main critical issues related to the mechanisms of interaction between plasma and plant matrix is the possible generation, within the extract, of reactive oxygen and nitrogen species that are harmful to human health, such as hydrogen peroxide, which is capable of exerting antimicrobial and cytotoxic activity [\[125\]](#page-32-29), or degradation compounds resulting from interactions between the reactive species and plant matrix nutrients (i.e., proteins, lipids, and carbohydrates) [\[126\]](#page-32-30) such as malondialdehyde or 4-hydroxynonenal from unsaturated fatty acids [\[127\]](#page-32-31).

An important prerequisite for the applicability of a plant extract, whether in the food, herbal, or cosmetic industries, is safety. In the case of plasma, in particular, the use of which generates harmful reactive species, knowledge of the potential threats from its use is even more important.

In this regard, Wende et al. (2018) pointed out, in their review of potential risks from the use of plasma in clinical applications, that the plasma sources considered were safe biologically, chemically, and physically [\[128\]](#page-33-0).

However, if we go outside the clinical field, to date there is very little research on the safety of plasma-treated foods for human or animal consumption, while, as far as we know, there are even fewer studies in the literature on the toxic effects of phytoextracts obtained by CP.

Mehta et al. (2022) reported no toxicity towards HepG2 cells when using plasma polyphenol extracts from rice and corn at concentrations under 250 µg/mL [\[129\]](#page-33-1).

Conversely, Heslin et al. (2020) reported contrasting effects using lettuce broth treated with plasma for different times in in vitro and in vivo experiments on *Galleria mellonella* larvae. The authors observed low cytotoxic effects in vitro and acute toxicity of the broth treated for 5 min in vivo, emphasizing the need to evaluate plasma-treated products both in isolation and in the context of a biological system and highlighting the need for detailed exposure studies and standardized evaluation procedures [\[130\]](#page-33-2). Los et al. (2020) reported that the biochemistry and cytotoxic potential of food models changed more when the plasma-treated sample was in liquid (aqueous wheat extract) rather than solid (whole wheat kernels) form and that the reduction in cell growth also depended on the parameters of the plasma treatment. As in other studies, the authors emphasized increased cytotoxicity when samples were exposed for long periods to plasma treatment (up to 20 min) and when cell cultures were treated with high concentrations of the test solutions (up to 10%) [\[131\]](#page-33-3). These results are very interesting because they highlight the possibility of low risk in the use of plasma as an extractive technique for the preparation of phytochemical extracts from a solid plant matrix. However, the lack of in-depth studies on the variation in the chemical and biochemical composition of the final extracts obtained with this technology does not allow the technique to be defined as safe for the preparation of phytochemical extracts to date, but opens up a very exciting field of research for scholars in the field.

5. Influence of Cold Plasma on the Phytochemical Profile and Biological Activity of Plant Extracts

5.1. Phytochemical Profile

From a chemical point of view, the term 'phenolic' or 'polyphenol' can be defined as a substance that possesses one or more aromatic ring and various functional derivatives such as esters, methyl ethers, glycosides, etc. [\[132\]](#page-33-4) (Figure [9\)](#page-19-0). In addition, most phenolic compounds possess two or more hydroxyl groups, which contribute to their potential hydrogen binding with other compounds [\[132\]](#page-33-4).

Figure 9. Chemical structures of the most representative polyphenol compounds. **Figure 9.** Chemical structures of the most representative polyphenol compounds.

Phenolic compounds are positively correlated with the nutraceutical and sensory Phenolic compounds are positively correlated with the nutraceutical and sensory quality of fresh and processed plant f[oods](#page-33-5) [133]. They possess a broad spectrum of biological properties such as antioxidant, antiproliferative, and anti-inflammator[y](#page-1-0) (Table 1), making them excellent health-promoting compounds for the food i[ndu](#page-33-6)[stry \[](#page-33-7)134–136].

Several studies have shown how CP treatment can affect the extraction of these pounds, by increasing the total polyphenol content (TPC) in the obtained plant extracts. compounds, by increasing the total polyphenol content (TPC) in the obtained plant extracts. Ahmadian et al. (2023) demonstrated that ultrasounds coupled with cold atmospheric Ahmadian et al. (2023) demonstrated that ultrasounds coupled with cold atmospheric plasma (CAP), as a pre-treatment, improved the extraction of phenolic components from plasma (CAP), as a pre-treatment, improved the extraction of phenolic components from *Hyssopus officinalis* L. (Hyssop) by about 22% compared with ultrasound alone [96]. Phe-*Hyssopus officinalis* L. (Hyssop) by about 22% compared with ultrasound alone [\[96\]](#page-31-27). Phenolic components increased in connection with cell membrane destruction by active chemical species, UV photons, and charged particles generated from CP, simplifying the extraction of phenolic compounds from the plant matrix. Also, CP supplied enough energy to break phenolic covalent bonds within polysaccharides (plant cell walls). Moreover, nitrogen plasma-pretreated samples exhibited a higher TPC than the other samples (air plasma and conventional solvent extraction). This was probably due to the breakdown of the aromatic rings of phenolic compounds by molecular ozone, which caused their destruction. However,

a longer pre-treatment time with CP can create more reactive components, providing sufficient energy to separate and release the bound phenols [\[88,](#page-31-10)[94,](#page-31-16)[96,](#page-31-27)[99\]](#page-31-28). Although CPgenerated ozone can affect the properties of plant extracts, it is very unstable to remain in the final product [\[137–](#page-33-8)[139\]](#page-33-9). Bao et al. (2020) tested a high-voltage atmospheric cold plasma (HVACP) pretreatment using different working gases (air, Ar, He, and N_2) to evaluate the extraction of phenolics from tomato pomace. According to the main results, samples treated with He and N_2 plasma showed significantly higher extraction yields than untreated samples. In contrast, argon and air plasma treatments showed no significant differences compared with the control. Eight phenolic compounds (such as phenolic acids and flavonoids) were also identified and confirmed by liquid chromatography massspectroscopy (LC-MS). CP treatments showed no effect on the concentrations of trans-ferulic acid, gallic acid, rutin, and isoquercetin, while concentrations of caffeic acid, chlorogenic acid, quercetin, and naringenin increased depending on the working gas. Additionally, the extraction rate of flavonoids was more significant than that of phenolic acids, since the release of bound flavonoids required less energy [\[88\]](#page-31-10).

In a similar study, Bao et al. (2020) investigated the effect of HVACP on the extraction of phenolics and anthocyanins from grape pomace. Treatment with CP (5 and 15 min) significantly increased the extraction of phenols, while the sample treated for 10 min showed no significant differences compared with the control. Similarly to the phenol content, the anthocyanin content increased after treatment with CP (processing time: 5 and 15 min) compared with the untreated sample; however, the authors noted no significant difference from the control for samples treated for 10 min with CP. In the analyzed samples, anthocyanins, quercetin, and phenolic acids (such as protocatechuic and gallic acids) were identified by ultra-performance liquid chromatography (UPLC). About these compounds specifically, after treatment with CP for 5, 10, and 15 min, the results showed higher quercetin concentrations than in the control, while CP treatments lasting 5 and 15 min were able to produce increased anthocyanidin concentrations in the respective extracts. The authors also reported that, under the experimental conditions tested, flavonoids were more extractable than phenolic acids [\[94\]](#page-31-16). Umair et al. (2022) observed an increase in the TPC of carrot juice due to the combined effect of HVACP and ultra-high hydrostatic pressure (UHHP). According to the authors, the increase in TPC by plasma-induced active species was related to the disruption of cell membrane bonds and the biosynthesis of phenolic compounds through the metabolism of phenylpropanoid enzymes [\[111\]](#page-32-32). An investigation by Kashfi et al. (2020) indicated that treatment of peppermint with low-pressure cold plasma (LPCP) (20 and 50 W, for 20 min) increased the TPC in the extract compared with the control sample [\[109\]](#page-32-33).

Keshavarzi et al. (2020) evaluated the efficacy of DBD cold plasma variables (gas, time, and power) on the TPC extraction rate from green tea leaves. They reported that the TPC of the samples decreased and increased after air and nitrogen DBD plasma treatment, respectively. In the first case, oxygen radicals formed during treatment with air CP led to the degradation of phenolic compounds. In contrast, in nitrogen CP treatment, ion bombardment led to erosion of the superficial epidermal layer of the leaves, facilitating solvent penetration and the extraction of more phenolic compounds. The study also showed that the interaction between time and power required an increase in both parameters to achieve the best TPC. Finally, the analysis of the individual phenolic compounds under optimal nitrogen CP conditions (15 W and 15 min) showed an increase in the concentration of gallic acid and catechin, with a slight decrease in epicatechin and epigallocatechin gallate (EGCG) [\[21\]](#page-28-13).

Pogorzelska-Nowicka et al. (2021) evaluated the effect of CP pretreatment on the TPC of 12 types of grounded and water-suspended herbs. This approach has not been found in previous studies. In 11 extracts from 12 herbs, they observed a significant increase in total phenols (approximately 10%) after the CP treatment (8 min, 20 kHz). This was probably due to the destruction of the cell membrane by reactive plasma species. In addition, the hydration of ground herbs can facilitate the extraction process because of the equal penetration of radicals in the whole sample surface. They also stated that the breakdown of larger polyphenols into minor compounds can increase the TPC of the extract. Anthocyanin and flavonoid content also increased but only in four herb extracts. However, in 8 out of the 12 samples, no significant difference was observed in individual phenolic compounds [\[19\]](#page-28-12). Rodríguez et al. (2017) reported that nitrogen-CP treatment significantly increased the TPC of cashew apple juice. However, in the case of treatment performed for 5, 10, and 15 min, no significant differences between the samples were observed [\[97\]](#page-31-29). Lee et al. (2023) studied the effect of surface dielectric barrier discharge (SDBD) on oat (*Avena sativa* L.) sprout extracts [\[117\]](#page-32-34). In this study, oat seeds, after hydration for 12 h, were exposed to plasma for 6 min per day for 3 days after sowing, with no significant effect on sprout growth, but with a significant increase in the content of bioactive metabolites. They stated that the phenolic content of oat sprouts increased through the stimulation of the antioxidant system, the release of phenolic compounds, and their decomposition into minor compounds, similar to the study conducted by Kim et al. (2014) [\[108\]](#page-32-35). According to the results reported by the authors, the single plasma treatment (1 day) produced a significant increase in the content of free amino acids (39.4%), $γ$ -aminobutyric acid (53%), and avenacoside B (23%) compared with the control, while the increase in the number of CP treatments (3-days) increased the content of hexacosanol, the most plentiful polycosanol found in oat sprouts [\[117\]](#page-32-34). Herceg et al. (2016) demonstrated that processing time and gas flow rate were able to improve the TPC of pomegranate juice [\[98\]](#page-31-30).

Several researchers have also reported that CP treatment/pretreatment may reduce TPC. Faria et al. (2020) investigated the effect of glow discharge plasma (GDP) pretreatment on the ultrasound-assisted extraction of total phenolic compounds from sea asparagus. CP pretreatment for 60 min resulted in a significant decrease (83%) in TPC [\[20\]](#page-28-14). Afshar et al. (2021) investigated the effect of oxygen and nitrogen CP treatment on the physicochemical properties of oil extracted from sunflower and sesame seeds. They stated that the TPC in both extracted oils decreased significantly as exposure time increased, especially when an oxygen atmosphere was applied. Indeed, ROS in an oxygen-CP atmosphere can interact with phenolic compounds, leading to structural degradation that results in a decrease in total phenolic compounds [\[114\]](#page-32-26). Hemmati et al. (2021) reported that the total phenolic content of green tea powder decreased with the increase in exposure voltage (20, 22, and 25 kV) and time (2, 4, 6, and 8 min). Similarly, the TPC of apple juice was also reduced with increasing cold plasma treatment time [\[99](#page-31-28)[,140\]](#page-33-10).

Zielinska et al. (2022) investigated the modification of okra pod cell wall polysaccharides and phytochemicals using cold plasma. CP treatment (5, 15, and 30 s) slightly reduced the TPC of okra pods by 5, 13, and 20%, respectively. The results indicated that treatment with CP destroyed the basic structures of phenolic compounds and that oxidation of these compounds, such as phenolic acids and their derivatives, reduced TPC [\[100,](#page-31-31)[106\]](#page-32-36).

Almeida et al. (2015) reported the effect of CAP and ozone on prebiotic orange juice. Orange juice can be affected by direct and indirect DBD CP treatment exposure. They observed that the TPC decreased with 70 kV plasma treatment applied for 15–60 s [\[101\]](#page-31-32). Liu et al. (2021) and Leite et al. (2021) found a similar effect of DBD CP on the reduction in TPC in kiwi turbid juice and cashew apple juice extraction [\[102](#page-31-33)[,103\]](#page-32-37). In other studies, Abedelmaksoud et al. (2022) stated that the TPC of mango pulp increased with the application of dielectric barrier discharge plasma (DBDP) for up to 6 min and then decreased at 8 and 10 min. According to the authors, the increase in phenolic compounds was caused by plasma polymerization and phenylalanine ammonia-lyase activity, which caused the decomposition of cell wall polysaccharides and the release of conjugated phenolic compounds [\[104\]](#page-32-38). Furthermore, Yodpitak et al. (2019) reported that the phenolic content of DBD-treated brown rice (PGBR) reached its maximum value and increased during germination within 0.5 days, whereas in the control sample (GBR), germination peaked after 1.5 days and then decreased rapidly [\[105,](#page-32-39)[107,](#page-32-40)[122\]](#page-32-41). Noteworthily, Seelarat et al. (2023) reported that the TPC of white *Cordyceps militaris* blended by *Cordyceps militaris* through cold plasma jet (CPJ), increased as time did too, from 30 s to 90 s, and decreased when the

treatment time reached 120 s [\[110\]](#page-32-42). These studies have shown that the quantity and quality of the phytochemical profiles in the extracts are mainly related to the interaction of CP with the treated plant matrix.

Among the operating parameters of the CP process, the processing time can play a crucial role in the reaction between plasma reactive species and phenolics, e.g., by increasing the oxidation of flavanols [\[90\]](#page-31-12). The impact of processing time also depends very much on the chemical structure of the individual bioactive compound being considered. For example, in the case of anthocyanins, short CP treatment facilitates their extraction by disrupting the cell vacuoles where these substances are enclosed. On the other hand, longer treatment times (>20 min) can lead to a loss of anthocyanins due to photodegradation by UV photons generated in the plasma, which transform these compounds into chalcones and thus into benzaldehyde and benzoic acid [\[141\]](#page-33-11).

These reactions are also affected by the type of plasma, and more operating variables, such as the power or voltage applied, the type of feed gas, the target phenolic compound, and the food matrix. Generally, DBD CP using high voltages (>60 kV) decreases the phenolic content of plant extracts due to oxidation side reactions because of the generation of a high concentration of ROS and ozone in the plasma. However, when DBD CP is used at high voltages but low frequencies (50 Hz), an increase in the hydroxybenzoic acid content can be observed. It is important to note that plasma-side reactions are not just limited to oxidation ones; they also include hydrogenation, dehydrogenation, hydrolysis, and dehydration reactions, which may boost or reduce the phenolic content of the plant extract [\[98\]](#page-31-30). For a more in-depth look at the effect of CP on the chemical structure of individual phenols, we recommend reading the review by Kumar et al. (2023) [\[30\]](#page-29-14). Considering these considerations, it seems clear that after treatment with CP, the phytochemical profile of a plant extract may differ due to the different degrees of structural decomposition of the cells or different side reactions that may occur during the extraction process. Further studies are therefore needed to determine the optimal CP processing conditions to maximize the phenolic compound yield of the extract and optimize its phytochemical profile. Table [3](#page-10-0) summarizes the changes in phytochemicals induced by CP application.

5.2. Antioxidant Capacity

Antioxidants are key substances that can control free radicals, bind oxygen, and block oxidation, maintaining the nutritional and nutraceutical value of food [\[142\]](#page-33-12). Among the bioactive substances present in food, polyphenols are considered to contribute most to the antioxidant potential of foods and extracts of plant origin. Any chemical changes in the profile of these compounds, such as oxidative degradation and the cleavage of double bonds by reactive oxygen species generated through CP treatment, such as OH[.], O³, and *O*₂, may increase or decrease the antioxidant capacity of the extract [\[77](#page-30-14)[,92](#page-31-14)[,143\]](#page-33-13).

Several studies have therefore analyzed the effect of cold plasma on the antioxidant capacity of extracts and essential oils (EO) [\[144\]](#page-33-14). For example, Bao et al. (2020) reported an improvement in the antioxidant potential of the phenolic extract from tomato pomace when samples were subjected to HVACP pretreatment produced with different gases. Samples treated with nitrogen CP showed the highest improvement in antioxidant capacity (30%) compared with samples treated with argon, helium, and air plasma. They stated that this observed general improvement was probably related to the higher levels of more potent phenolic antioxidants and hydrophilic phenols present in the final extract due to the increase in surface hydrophilicity following the treatment [\[88\]](#page-31-10). Similarly, Poomanee et al. (2021) indicated that Luem Pua (LP) pre-treated with CP showed the highest increase in antioxidant capacity compared with the other tested species and untreated LP. The IC_{50} value recorded for the pretreated LP sample was 0.080 ± 0.002 mg/mL, while the value for the untreated LP was 0.187 ± 0.025 mg/mL. According to the authors, this improvement may result from the higher concentration of cyanidin-3-O-glucoside (CNG) and phenolic compounds in the pretreated LP extract, as no other new compounds were observed in the HPLC analysis to which this improvement could be attributed [\[122\]](#page-32-41).

However, Bao et al. (2020) believed that another reason may be responsible for the improved antioxidant potential in plasma-treated samples. Indeed, they showed that when exposing the grape pomace to HVACP pretreatment, the phenolic extract obtained had a higher antioxidant potential regardless of the treatment duration (5, 10, and 15 min) than the untreated sample, due to the release of the intercellular compounds caused by the strong breakdown of the plant tissue following the experimental treatment. However, regardless of the overall increase in antioxidant capacity, it was observed a slight, though not significant, reduction in it from 5 to 10 min [\[94\]](#page-31-16).

Interestingly, some researchers observed that although CP treatment reduces TPC, total flavan content (TFC), or vitamin C, the antioxidant potential remains unaffected. The possible reason for this stability is that the unbounded antioxidant compounds, which are released from carbohydrate and protein complexes during CP treatment, can be replaced by part of the antioxidant compounds that are decomposed by the plasma reactive species generated during CP treatment [\[96,](#page-31-27)[100\]](#page-31-31).

For instance, Faria et al. (2020) reported that despite the drastic loss of phenolic content in sea asparagus (*Salicornia neei*) extract, when the pre-treatment time with CP was extended to 60 min, the antioxidant capacity did not change significantly. One possible reason for this stability, according to the authors, could be the presence of more active antioxidants formed by highly reactive species by the CP pretreatment employed before ultrasound-assisted extraction (UAE). Notably, with a shorter CP exposure time (5 min), an improvement in the antioxidant capacity of the extract was observed, probably due to a more severe cell wall cleavage that led to greater solvent penetration [\[20\]](#page-28-14).

Hou et al. (2019) also mentioned the importance of CP treatment exposure time since the antioxidant potential of blueberry juice (assessed by FRAP, ABTS, and DPPH techniques) declined when the operation time was extended from 2 to 6 min [\[145\]](#page-33-15). Similar results were reported by Abedelmaksoud et al. (2022) for fresh mango pulp treated with DBDP for four minutes [\[104\]](#page-32-38).

It is worth noting that CP treatment does not always positively affect antioxidant capacity. Keshavarzi et al. (2020) indicated that air-working gas in CP pretreatment before the extraction of phenols from green tea leaves led to a significant reduction in the antioxidant capacity in all treatments evaluated [\[21\]](#page-28-13). According to the authors, ROS formed through air-CP might be responsible for the cleavage of phenolic compounds, thus reducing the antioxidant potential of the extracts. In contrast, cold nitrogen plasma increased the antioxidant potential. This increase can be due to the presence of specific phenolic compounds, usually restricted to the membrane and cell wall, which are now rapidly released as sufficient energy is supplied under CP conditions. Another potential reason can be the accelerated solvent permeability due to the severe tissue breakdown caused by ion bombardment during CP pretreatment. This investigation continued by optimizing nitrogen CP pretreatment conditions (exposure time of 15 min and generation power of 15 W) to reach the highest antioxidant capacity of the final extract (152.114 μ M Trolox/g) by central composite design $[21]$.

Kashfi et al. (2020) reported that plasma power values (20, 50, and 60 W for 20 min) during radiofrequency LPCP treatment affected the antioxidant potential of plant extracts. The authors showed that when peppermint (*Mentha piperita* L.) was subjected to LPCP immediately before the drying process, an increase in the antioxidant capacity of the extract was observed at a plasma power value of 50 W (IC_{50} : 0.240 mg/mL), probably resulting from an increase in the concentration of bioactive compounds in the peppermint extract pretreated with plasma compared with the control sample. However, by increasing the power to 60 W, a significant decrease in the antioxidant capacity was observed. Furthermore, all the LPCP-treated samples demonstrated higher ferric-reducing ability compared with the untreated control sample [\[109\]](#page-32-33).

In agreement with this study, Kim et al. (2019) pointed out that the intensity of active species generated during CP treatment was highly dependent on different CP parameters such as exposure time, generating power, plasma-working gas, and plasma source [\[120\]](#page-32-43). Pogorzelska-Nowicka et al. (2021) reported that, when subjecting various herbs (12 species) to cold plasma pre-treatment, the extract obtained from nine species showed a significant increase in their antioxidant potential, while *Sanguisorba officinalis* showed a slight reduction, and the extracts of *Andrographis paniculata* and *Polygonum aviculare* showed no significant changes following CP treatment [\[19\]](#page-28-12). As far as the effect of CP on the biological properties of essential oils (EOs) is concerned, Buonopane et al. (2019) reported that the EOs from CP-treated sweet basil plants showed a higher antioxidant capacity than untreated samples due to the higher eugenol extraction (48–94.82%) [\[121\]](#page-32-44). In contrast, Afshar et al. (2022) reported that for sunflower and sesame seeds pre-treated with CP, there was a significant decrease in the antioxidant capacity of the extracted oils, attributable to the degradation of phytochemicals during treatment, especially in cold air plasma, caused by the presence of ROS in the plasma. Furthermore, sunflower oil samples showed a higher scavenging capacity than sesame oil samples in the DPPH assay [\[114\]](#page-32-26). In this regard, it is worth noting that employing different methods provides a more accurate view of the potential antioxidant alteration in food products [\[97,](#page-31-29)[146\]](#page-33-16). Several studies have shown different trends in antioxidant potential values about the type of in vitro test used to measure it, emphasizing the need to use more than one analytical test to more reliably determine the antioxidant potential of an extract or food, also depending on the fact that the antioxidant compounds most reactive to a specific type of method may be affected differently during plasma processing [\[96](#page-31-27)[,145\]](#page-33-15).

In agreement with the above statements, Ahmadian et al. (2023) reported inconsistent results when examining the antioxidant potential of extracts from hyssop (*Hyssopus officinalis* L.) pretreated with CAP using FRAP and DPPH techniques, respectively. According to the FRAP assay, pretreated samples with N_2 and air plasma showed an increasing trend in antioxidant properties compared with control samples. In contrast, the DPPH assay showed that CAP treatment led to lower antioxidant capacity. However, N_2 plasma-treated samples exhibited a higher radical scavenging rate than the untreated ones [\[96\]](#page-31-27).

It is worth emphasizing that the different process parameters of the CP treatment, influencing the phytochemical profile of the extract, can similarly modulate its antioxidant potential. Rodríguez et al. (2017) reported that among different process parameters during the CP treatment of cashew apple juice, N_2 plasma flow rate and exposure time played a determinant role in the FRAP assay. The most significant changes were observed at the 15th min of treatment and flow rates of 30 mL/min (164% relative antioxidant activity) and 50 mL/min (163% relative antioxidant activity). In addition, the results showed that the DPPH and ABTS assays were significantly dependent on the period of plasma treatment of the sample, where a longer exposure time, regardless of the rate of N_2 flow, resulted in a continuous decrease in the antioxidant capacity (the highest antioxidant potential was recorded at the 5th min of CP treatment) [\[97\]](#page-31-29).

Almeida et al. (2015) considered the ABTS assay a more accurate method in comparison with the DPPH one to reflect the antioxidant capacity alteration, since the CAP-treated prebiotic orange juice showed lower antioxidant activity in ABTS technique, while no significant difference was observed among treated and untreated samples through DPPH assay [\[101\]](#page-31-32). Ramazzina et al. (2016) evaluated some functional properties of minimally processed Pink Lady apples treated with DBD CP in comparison with untreated samples, through different in vitro and ex vivo tests, pointing out that plasma treatment caused only a slight reduction in antioxidant content and antioxidant capacity (up to 10%), mainly limited to the amphiphilic fraction [\[147\]](#page-33-17).

It is worth noting that CP treatment may reduce antioxidant properties or even have no significant effect, due to the oxygen in the air plasma, which plays a role in suppressing antioxidant capacity by creating highly destructive ROS. Regardless of different types of working gases, the CP process parameters such as exposure time, power, and storage period have an important role in the antioxidant capacity changes. However, further investigation is recommended for a more accurate determination of different action mechanisms of the CP treatment on the antioxidant, through simultaneous assays such as ABTS, FRAP, and ORAC, to gain more reliable information. An overview of the effects of CP on antioxidant capacity is shown in Table [3.](#page-10-0)

5.3. Antimicrobial Properties

The use of plant extracts for the control of microbial infections and food spoilage is becoming increasingly popular in recent years. The ability to eliminate or control an exhaustive range of harmful organisms by complex plant extracts or essential oils, rich in polyphenols, terpenoids, and alkaloids, has led to the use of these bioactive compounds as food preservatives [\[148](#page-33-18)[,149\]](#page-33-19). In this regard, the quality of the extraction process is one of the essential factors affecting the antimicrobial properties of plant extracts. In some studies, CP has been introduced as a pioneering technique for obtaining a high-quality extraction process with minimal loss of valuable compounds, thus preserving their antimicrobial properties. In one of these studies by Pogorzelska-Nowicka and coworkers (2021), the antibacterial potential of various dried herb extracts obtained by pretreatment with CP was evaluated against aerobic bacteria. Overall, the CP samples reduced total aerobic bacteria and most of them showed germicidal properties. Depending on herb species, the number of aerobic bacteria decreased by 1.27 up to 3.48 logCFU/g [\[19\]](#page-28-12).

Kashfi et al. (2020) evaluated in vivo the efficacy of dried peppermint (*Mentha piperita* L.) extracts obtained by LPCP at three different powers (20, 50, and 60 W) for 20 min against the pathogen *E. coli* [\[109\]](#page-32-33). According to experimental observations, the bacterial growth was completely controlled with peppermint extract using high CP powers (50 and 60 W), whereas this process was unable to inhibit *E. coli* at 20 W. Faria et al. (2020) investigated the effect of ultrasound-assisted CP pretreatment on in vitro antimicrobial activity of the *S. neei* extract against *S. aureus* and *E. coli* by the disc diffusion method [\[20\]](#page-28-14). The *S. neei* extract (0.1 g/mL) showed an inhibition halo (4 mm) against *E. coli* and no inhibition halo against *S. aureus*. In general, the decomposition of the extract caused by CP treatment produces bioactive compounds such as ferulic, caffeic, and *p*-coumaric acids, which have shown synergistic antibacterial properties. Despite the few studies on the indirect antimicrobial effects of CP through the use of the plant extract obtained by this technique, many results confirm the direct application of CP in food disinfection.

Most studies stated that the microbial count decreased while increasing plasma treatment time and voltage [\[99](#page-31-28)[,108\]](#page-32-35). In general, it has been found that Gram-negative bacteria are more sensitive to plasma treatment than Gram-positive ones. CP efficacy seems to be directly correlated to bacterial cell wall thickness. Gram-negative bacteria are characterized by a thin (<10 nm) inner cell wall layer and an additional outer membrane, consisting of phospholipids and lipopolysaccharides that are highly sensitive to plasma-generated ROS. Moreover, due to the presence of porins on the outer membrane of these bacteria, plasma penetration is certainly more effective than in the case of Gram-positive bacteria, which are characterized by a thicker peptidoglycan-based cell wall (20–80 nm) that cannot easily break down by reactive plasma species [\[150](#page-33-20)[–152\]](#page-33-21). However, Hemmati et al. (2021) noticed that CAP was more effective in inhibiting Gram-positive bacteria (*E. faecalis*) than Gram-negative ones (*E. coli*) [\[153\]](#page-33-22).

A summary of the effects of CP on the antimicrobial properties of the plant extracts obtained is given in Table [3.](#page-10-0) To date, studies on the antibacterial properties of these extracts are lacking, making it difficult to assess their efficacy. Therefore, it is necessary to extend research studies in this area with the help of statistical strategies to model and optimize the extraction conditions.

5.4. Other Relevant Biological Properties

The use of cold plasma to produce phytoextracts is a very recent technique, and most studies have focused on the effect of the extraction technique on the antioxidant and antimicrobial activity of the extracts obtained. However, some studies have evaluated other biological properties of phytoextracts obtained using plasma. Poomanee et al. (2021) optimized the CP extraction of some purple rice varieties, obtaining up to a five-fold higher

amount of Cyanidin-3-O-glucoside in plasma-treated samples compared with the untreated one, with a significantly more substantial anti-ageing potential [\[122\]](#page-32-41).

Mehta et al. (2022) optimized the extraction process of polyphenols by atmospheric and vacuum CP from rice and maize bran. The resulting phytoextracts showed not only higher antioxidant activity, but also better in vitro digestion, higher anti-inflammatory responses, and better overall quality than a conventional extraction [\[129\]](#page-33-1). In another study, they extracted xylooligosaccharides (XOS) from rice and corn bran dietary fibers by CP. Similar to polyphenols in their previous study, the extracted XOS showed better gastric digestion and anti-inflammatory responses with no cytotoxicity against RAW 264.7 and HepG2 cell lines [\[154\]](#page-34-0).

Jeong et al. (2019) found that atmospheric DBD cold plasma treatment resulted in the dimerization of *trans*-resveratrol in a methanol solution. The generated *trans*-resveratrol dimers showed higher inhibitory activities against α -glucosidase and α -amylase than parent *trans*-resveratrol. Inhibiting the activity of these enzymes can contribute to the improvement of anti-diabetic properties through controlling postprandial hyperglycemia, and reducing the risk of developing diabetes [\[155\]](#page-34-1). The lack of studies on the biological properties, such as anticarcinogenic or anti-mutagenic, of plasma-derived phytoextracts, opens exciting new research prospects for scholars in the field to implement this technique commercially.

6. Conclusions and Future Perspective

Cold plasma is a new non-thermal technology that has shown considerable potential for food disinfection at low temperatures, with minimal energy consumption and cost. It has been introduced in recent years also as an advanced technique to produce high-quality phytoextracts with minimal side effects. The preservation of the main plant components and low impact on the internal matrix of the product is an essential advantage of CP compared with other traditional extraction methods. In the extraction industry, it represents an environmentally friendly process, with no production of toxic and hazardous waste compounds and no need to use water or solvents, which is a high added value in the preparation of high-quality plant extracts. However, like all nascent technologies, for its full exploitation at the industrial level, many studies are needed to design the system, its scalability, etc.

CP leads to an increase in extraction efficiency and, in general, improves the antioxidant properties of the extracts, which mainly depend on the phenolic compounds present in them. However, the complete control of CP treatment to achieve optimal extraction conditions is rather difficult due to the involvement of several process parameters that may enhance or suppress the antioxidant potential of the extracts. For the technique to take off, especially on a commercial scale, further investigations are therefore needed to in particular the mechanisms of plant cell wall disruption caused by CP and the interactions between phenolic compounds and plasma reactive species on a molecular or atomic level.

More studies are also needed in the food sector, where researchers have recently shared a new perspective on the ability of CP to reduce allergens in plant-derived foods. However, the mechanisms involved in this process are not entirely clear and require further investigation.

Studies in the literature have also highlighted the possible cytotoxic activity of plasmatreated solutions, which can be successfully used in the treatment of cancer cells, but parallel data on the effect these same solutions would have on healthy cell lines are lacking. In contrast, the results of studies on the mutagenic activity of plasma-treated protein solutions are mixed. Currently, the technique appears very promising for the preparation of phytoextracts, but for it to find industrial application, further studies are needed on the wholesomeness of the products obtained by this technique and thus on the interactions between the reactive species generated during plasma treatment and the primary and secondary metabolites present in the plant matrix.

Supplementary Materials: The following supporting information can be downloaded at: [https:](https://www.mdpi.com/article/10.3390/foods12173181/s1) [//www.mdpi.com/article/10.3390/foods12173181/s1,](https://www.mdpi.com/article/10.3390/foods12173181/s1) Figure S1a: Documents by year; Figure S1b: Documents by type; Figure S1c: Documents by subject area; Figure S1d: Documents by country or territory; Reprinted with permission from Scopus®. 2023, Elsevier. Table S1. Cluster composition of keyword co-occurrence network analysis from the cleaned dataset after SCOPUS search: "*cold plasma*" AND "*extraction*". Descending order of total number of occurrences for each keyword in the cluster.

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Conflicts of Interest: The authors declare no conflict of interest.

List of Abbreviations

References

- 1. Phytochemicals Market. Available online: [https://www.persistencemarketresearch.com/market-research/phytochemicals](https://www.persistencemarketresearch.com/market-research/phytochemicals-market.asp)[market.asp](https://www.persistencemarketresearch.com/market-research/phytochemicals-market.asp) (accessed on 13 February 2023).
- 2. Thakur, M.; Singh, K.; Khedkar, R. Phytochemicals: Extraction process, safety assessment, toxicological evaluations, and regulatory issues. In *Functional and Preservative Properties of Phytochemicals*; Elsevier: Amsterdam, The Netherlands, 2020; pp. 341–361.
- 3. Saxena, M.; Saxena, J.; Nema, R.; Singh, D.; Gupta, A. Phytochemistry of medicinal plants. *J. Pharmacogn. Phytochem.* **2013**, *1*, 168–182.
- 4. Shirani, M.; Parandi, E.; Nodeh, H.R.; Akbari-Adergani, B.; Shahdadi, F. Development of a rapid efficient solid-phase microextraction: An overhead rotating flat surface sorbent based 3-D graphene oxide/lanthanum nanoparticles@ Ni foam for separation and determination of sulfonamides in animal-based food products. *Food Chem.* **2022**, *373*, 131421. [\[CrossRef\]](https://doi.org/10.1016/j.foodchem.2021.131421)
- 5. Shirani, M.; Aslani, A.; Sepahi, S.; Parandi, E.; Motamedi, A.; Jahanmard, E.; Nodeh, H.R.; Akbari-Adergani, B. An efficient 3D adsorbent foam based on graphene oxide/AgO nanoparticles for rapid vortex-assisted floating solid phase extraction of bisphenol A in canned food products. *Anal. Methods* **2022**, *14*, 2623–2630. [\[CrossRef\]](https://doi.org/10.1039/D2AY00426G) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35735028)
- 6. Bidhendi, M.E.; Parandi, E.; Meymand, M.M.; Sereshti, H.; Nodeh, H.R.; Joo, S.-W.; Vasseghian, Y.; Khatir, N.M.; Rezania, S. Removal of lead ions from wastewater using magnesium sulfide nanoparticles caged alginate microbeads. *Environ. Res.* **2023**, *216*, 114416. [\[CrossRef\]](https://doi.org/10.1016/j.envres.2022.114416)
- 7. Lourenço, S.C.; Moldão-Martins, M.; Alves, V.D. Antioxidants of natural plant origins: From sources to food industry applications. *Molecules* **2019**, *24*, 4132. [\[CrossRef\]](https://doi.org/10.3390/molecules24224132) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31731614)
- 8. Mosleh, N.; Ahranjani, P.J.; Parandi, E.; Nodeh, H.R.; Nawrot, N.; Rezania, S.; Sathishkumar, P. Titanium lanthanum three oxides decorated magnetic graphene oxide for adsorption of lead ions from aqueous media. *Environ. Res.* **2022**, *214*, 113831. [\[CrossRef\]](https://doi.org/10.1016/j.envres.2022.113831) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35841973)
- 9. Shirani, M.; Aslani, A.; Ansari, F.; Parandi, E.; Nodeh, H.R.; Jahanmard, E. Zirconium oxide/titanium oxide nanorod decorated nickel foam as an efficient sorbent in syringe filter based solid-phase extraction of pesticides in some vegetables. *Microchem. J.* **2023**, *189*, 108507. [\[CrossRef\]](https://doi.org/10.1016/j.microc.2023.108507)
- 10. De Santis, D.; Carbone, K.; Garzoli, S.; Laghezza Masci, V.; Turchetti, G. Bioactivity and chemical profile of *Rubus idaeus* L. leaves steam-distillation extract. *Foods* **2022**, *11*, 1455. [\[CrossRef\]](https://doi.org/10.3390/foods11101455)
- 11. Santarelli, V.; Neri, L.; Carbone, K.; Macchioni, V.; Pittia, P. Use of Conventional and Innovative Technologies for the Production of Food Grade Hop Extracts: Focus on Bioactive Compounds and Antioxidant Activity. *Plants* **2022**, *11*, 41. [\[CrossRef\]](https://doi.org/10.3390/plants11010041)
- 12. Abedi-Firoozjah, R.; Yousefi, S.; Heydari, M.; Seyedfatehi, F.; Jafarzadeh, S.; Mohammadi, R.; Rouhi, M.; Garavand, F. Application of red cabbage anthocyanins as pH-sensitive pigments in smart food packaging and sensors. *Polymers* **2022**, *14*, 1629. [\[CrossRef\]](https://doi.org/10.3390/polym14081629)
- 13. Macchioni, V.; Carbone, K.; Cataldo, A.; Fraschini, R.; Bellucci, S. Lactic acid-based deep natural eutectic solvents for the extraction of bioactive metabolites of *Humulus lupulus* L.: Supramolecular organization, phytochemical profiling and biological activity. *Sep. Purif. Technol.* **2021**, *264*, 118039. [\[CrossRef\]](https://doi.org/10.1016/j.seppur.2020.118039)
- 14. Chemat, F.; Vian, M.A.; Fabiano-Tixier, A.-S.; Nutrizio, M.; Jambrak, A.R.; Munekata, P.E.; Lorenzo, J.M.; Barba, F.J.; Binello, A.; Cravotto, G. A review of sustainable and intensified techniques for extraction of food and natural products. *Green Chem.* **2020**, *22*, 2325–2353. [\[CrossRef\]](https://doi.org/10.1039/C9GC03878G)
- 15. Sarlak, Z.; Rouhi, M.; Mirza Alizadeh, A.; Sadeghi, E.; Hosseini, H.; Mousavi Khaneghah, A. Pb exposure from plant foods in Iran: A review. *Int. J. Environ. Anal. Chem.* **2021**, 1–22. [\[CrossRef\]](https://doi.org/10.1080/03067319.2021.1970149)
- 16. Raesi, S.; Mohammadi, R.; Khammar, Z.; Paimard, G.; Abdalbeygi, S.; Sarlak, Z.; Rouhi, M. Photocatalytic detoxification of aflatoxin B1 in an aqueous solution and soymilk using nano metal oxides under UV light: Kinetic and isotherm models. *LWT* **2022**, *154*, 112638. [\[CrossRef\]](https://doi.org/10.1016/j.lwt.2021.112638)
- 17. Paimard, G.; Mohammadi, R.; Bahrami, R.; Khosravi-Darani, K.; Sarlak, Z.; Rouhi, M. Detoxification of patulin from juice simulator and apple juice via cross-linked Se-chitosan/L-cysteine nanoparticles. *LWT* **2021**, *143*, 111146. [\[CrossRef\]](https://doi.org/10.1016/j.lwt.2021.111146)
- 18. Sarlak, Z.; Khosravi-Darani, K.; Rouhi, M.; Garavand, F.; Mohammadi, R.; Sobhiyeh, M.R. Bioremediation of organophosphorus pesticides in contaminated foodstuffs using probiotics. *Food Control* **2021**, *126*, 108006. [\[CrossRef\]](https://doi.org/10.1016/j.foodcont.2021.108006)
- 19. Pogorzelska-Nowicka, E.; Hanula, M.M.; Brodowska-Trębacz, M.; Górska-Horczyczak, E.; Jankiewicz, U.; Mazur, T.; Marcinkowska-Lesiak, M.; Półtorak, A.; Wierzbicka, A. The Effect of Cold Plasma Pretreatment on Water-Suspended Herbs Measured in the Content of Bioactive Compounds, Antioxidant Activity, Volatile Compounds and Microbial Count of Final Extracts. *Antioxidants* **2021**, *10*, 1740. [\[CrossRef\]](https://doi.org/10.3390/antiox10111740) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34829611)
- 20. Faria, G.; Souza, M.; Oliveira, J.; Costa, C.; Collares, M.; Prentice, C. Effect of ultrasound-assisted cold plasma pretreatment to obtain sea asparagus extract and its application in Italian salami. *Food Res. Int.* **2020**, *137*, 109435. [\[CrossRef\]](https://doi.org/10.1016/j.foodres.2020.109435)
- 21. Keshavarzi, M.; Najafi, G.; Ahmadi Gavlighi, H.; Seyfi, P.; Ghomi, H. Enhancement of polyphenolic content extraction rate with maximal antioxidant activity from green tea leaves by cold plasma. *J. Food Sci.* **2020**, *85*, 3415–3422. [\[CrossRef\]](https://doi.org/10.1111/1750-3841.15448)
- 22. Moreira, S.A.; Alexandre, E.M.; Pintado, M.; Saraiva, J.A. Effect of emergent non-thermal extraction technologies on bioactive individual compounds profile from different plant materials. *Food Res. Int.* **2019**, *115*, 177–190. [\[CrossRef\]](https://doi.org/10.1016/j.foodres.2018.08.046) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30599930)
- 23. Ranjha, M.M.A.; Kanwal, R.; Shafique, B.; Arshad, R.N.; Irfan, S.; Kieliszek, M.; Kowalczewski, P.Ł.; Irfan, M.; Khalid, M.Z.; Roobab, U. A critical review on pulsed electric field: A novel technology for the extraction of phytoconstituents. *Molecules* **2021**, *26*, 4893. [\[CrossRef\]](https://doi.org/10.3390/molecules26164893) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34443475)
- 24. Aghel, B.; Gouran, A.; Parandi, E.; Jumeh, B.H.; Nodeh, H.R. Production of biodiesel from high acidity waste cooking oil using nano GO@ MgO catalyst in a microreactor. *Renew. Energy* **2022**, *200*, 294–302. [\[CrossRef\]](https://doi.org/10.1016/j.renene.2022.09.045)
- 25. Fierascu, R.C.; Fierascu, I.; Avramescu, S.M.; Sieniawska, E. Recovery of natural antioxidants from agro-industrial side streams through advanced extraction techniques. *Molecules* **2019**, *24*, 4212. [\[CrossRef\]](https://doi.org/10.3390/molecules24234212) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31757027)
- 26. Heydari, M.; Rostami, O.; Mohammadi, R.; Banavi, P.; Farhoodi, M.; Sarlak, Z.; Rouhi, M. Hydrodistillation ultrasound-assisted green extraction of essential oil from bitter orange peel wastes: Optimization for quantitative, phenolic, and antioxidant properties. *J. Food Process. Preserv.* **2021**, *45*, e15585. [\[CrossRef\]](https://doi.org/10.1111/jfpp.15585)
- 27. Carbone, K.; Macchioni, V.; Petrella, G.; Cicero, D.O. Exploring the potential of microwaves and ultrasounds in the green extraction of bioactive compounds from *Humulus lupulus* for the food and pharmaceutical industry. *Ind. Crops Prod.* **2020**, *156*, 112888. [\[CrossRef\]](https://doi.org/10.1016/j.indcrop.2020.112888)
- 28. Ramazzina, I.; Macchioni, V.; Carbone, K. Antioxidant and pro-oxidant phytochemicals in ultrasound and microwave assisted extracts from hop cones: A statistical modelling approach. *Food Funct.* **2022**, *13*, 9589–9601. [\[CrossRef\]](https://doi.org/10.1039/D2FO02020C)
- 29. Hanula, M.; Wyrwisz, J.; Moczkowska, M.; Horbańczuk, O.K.; Pogorzelska-Nowicka, E.; Wierzbicka, A. Optimization of microwave and ultrasound extraction methods of açai berries in terms of highest content of phenolic compounds and antioxidant activity. *Appl. Sci.* **2020**, *10*, 8325. [\[CrossRef\]](https://doi.org/10.3390/app10238325)
- 30. Kumar, S.; Pipliya, S.; Srivastav, P.P. Effect of cold plasma on different polyphenol compounds: A review. *J. Food Process Eng.* **2023**, *46*, e14203. [\[CrossRef\]](https://doi.org/10.1111/jfpe.14203)
- 31. Carbone, K.; Amoriello, T.; Iadecola, R. Exploitation of kiwi juice pomace for the recovery of natural antioxidants through microwave-assisted extraction. *Agriculture* **2020**, *10*, 435. [\[CrossRef\]](https://doi.org/10.3390/agriculture10100435)
- 32. Patil, P.S.; Shettigar, R. An advancement of analytical techniques in herbal research. *J. Adv. Sci. Res.* **2010**, *1*, 8–14.
- 33. Naude, Y.; De Beer, W.; Jooste, S.; Van Der Merwe, L.; Van Rensburg, S. Comparison of supercritical fluid extraction and Soxhlet extraction for the determination of DDT, DDD and DDE in sediment. *Water* **1998**, *24*, 205–214.
- 34. Handa, S. An overview of extraction techniques for medicinal and aromatic plants. *Extr. Technol. Med. Aromat. Plants* **2008**, *1*, 21–40.
- 35. Gupta, A.; Naraniwal, M.; Kothari, V. Modern extraction methods for preparation of bioactive plant extracts. *Int. J. Appl. Nat. Sci.* **2012**, *1*, 8–26.
- 36. Mohammad Azmin, S.N.H.; Abdul Manan, Z.; Wan Alwi, S.R.; Chua, L.S.; Mustaffa, A.A.; Yunus, N.A. Herbal processing and extraction technologies. *Sep. Purif. Rev.* **2016**, *45*, 305–320. [\[CrossRef\]](https://doi.org/10.1080/15422119.2016.1145395)
- 37. Safaripour, M.; Parandi, E.; Aghel, B.; Gouran, A.; Saidi, M.; Nodeh, H.R. Optimization of the microreactor-intensified transesterification process using silver titanium oxide nanoparticles decorated magnetic graphene oxide nanocatalyst. *Process Saf. Environ. Prot.* **2023**, *173*, 495–506. [\[CrossRef\]](https://doi.org/10.1016/j.psep.2023.03.039)
- 38. Trusheva, B.; Trunkova, D.; Bankova, V. Different extraction methods of biologically active components from propolis: A preliminary study. *Chem. Cent. J.* **2007**, *1*, 13. [\[CrossRef\]](https://doi.org/10.1186/1752-153X-1-13)
- 39. Laghari, A.Q.; Memon, S.; Nelofar, A.; Laghari, A.H. Extraction, identification and antioxidative properties of the flavonoid-rich fractions from leaves and flowers of Cassia angustifolia. *Am. J. Anal. Chem.* **2011**, *2*, 871. [\[CrossRef\]](https://doi.org/10.4236/ajac.2011.28100)
- 40. Zhang, H.-F.; Yang, X.-H.; Zhao, L.-D.; Wang, Y. Ultrasonic-assisted extraction of epimedin C from fresh leaves of Epimedium and extraction mechanism. *Innov. Food Sci. Emerg. Technol.* **2009**, *10*, 54–60. [\[CrossRef\]](https://doi.org/10.1016/j.ifset.2008.09.007)
- 41. Zhang, H.-F.; Yang, X.-H.; Wang, Y. Microwave assisted extraction of secondary metabolites from plants: Current status and future directions. *Trends Food Sci. Technol.* **2011**, *22*, 672–688. [\[CrossRef\]](https://doi.org/10.1016/j.tifs.2011.07.003)
- 42. Kaufmann, B.; Christen, P. Recent extraction techniques for natural products: Microwave-assisted extraction and pressurised solvent extraction. *Phytochem. Anal. Int. J. Plant Chem. Biochem. Tech.* **2002**, *13*, 105–113. [\[CrossRef\]](https://doi.org/10.1002/pca.631)
- 43. Parandi, E.; Safaripour, M.; Abdellattif, M.H.; Saidi, M.; Bozorgian, A.; Nodeh, H.R.; Rezania, S. Biodiesel production from waste cooking oil using a novel biocatalyst of lipase enzyme immobilized magnetic nanocomposite. *Fuel* **2022**, *313*, 123057. [\[CrossRef\]](https://doi.org/10.1016/j.fuel.2021.123057)
- 44. Chemat, F.; Rombaut, N.; Sicaire, A.-G.; Meullemiestre, A.; Fabiano-Tixier, A.-S.; Abert-Vian, M. Ultrasound assisted extraction of food and natural products. Mechanisms, techniques, combinations, protocols and applications. A review. *Ultrason. Sonochem.* **2017**, *34*, 540–560. [\[CrossRef\]](https://doi.org/10.1016/j.ultsonch.2016.06.035) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27773280)
- 45. Chandrapala, J.; Oliver, C.M.; Kentish, S.; Ashokkumar, M. Use of power ultrasound to improve extraction and modify phase transitions in food processing. *Food Rev. Int.* **2013**, *29*, 67–91. [\[CrossRef\]](https://doi.org/10.1080/87559129.2012.692140)
- 46. Wu, J.; Lin, L.; Chau, F.-t. Ultrasound-assisted extraction of ginseng saponins from ginseng roots and cultured ginseng cells. *Ultrason. Sonochem.* **2001**, *8*, 347–352. [\[CrossRef\]](https://doi.org/10.1016/S1350-4177(01)00066-9)
- 47. De Castro, M.L.; Priego-Capote, F. Soxhlet extraction: Past and present panacea. *J. Chromatogr. A* **2010**, *1217*, 2383–2389. [\[CrossRef\]](https://doi.org/10.1016/j.chroma.2009.11.027) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/19945707)
- 48. Parandi, E.; Pero, M.; Kiani, H. Phase change and crystallization behavior of water in biological systems and innovative freezing processes and methods for evaluating crystallization. *Discov. Food* **2022**, *2*, 6. [\[CrossRef\]](https://doi.org/10.1007/s44187-021-00004-2)
- 49. Abedi-Firoozjah, R.; Parandi, E.; Heydari, M.; Kolahdouz-Nasiri, A.; Bahraminejad, M.; Mohammadi, R.; Rouhi, M.; Garavand, F. Betalains as promising natural colorants in smart/active food packaging. *Food Chem.* **2023**, *424*, 136408. [\[CrossRef\]](https://doi.org/10.1016/j.foodchem.2023.136408) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37245469)
- 50. De Castro, M.L.; Garcıa-Ayuso, L. Soxhlet extraction of solid materials: An outdated technique with a promising innovative future. *Anal. Chim. Acta* **1998**, *369*, 1–10. [\[CrossRef\]](https://doi.org/10.1016/S0003-2670(98)00233-5)
- 51. Gao, X.; Yani, S.; Wu, H. Pyrolysis of spent biomass from mallee leaf steam distillation: Biochar properties and recycling of inherent inorganic nutrients. *Energy Fuels* **2014**, *28*, 4642–4649. [\[CrossRef\]](https://doi.org/10.1021/ef501114v)
- 52. Alupului, A.; Calinescu, I.; Lavric, V. Microwave extraction of active principles from medicinal plants. *UPB Sci. Bull. Ser. B* **2012**, *74*, 129–142.
- 53. Ridgway, K.; Lalljie, S.P.; Smith, R.M. Sample preparation techniques for the determination of trace residues and contaminants in foods. *J. Chromatogr. A* **2007**, *1153*, 36–53. [\[CrossRef\]](https://doi.org/10.1016/j.chroma.2007.01.134)
- 54. Zhang, Q.-W.; Lin, L.-G.; Ye, W.-C. Techniques for extraction and isolation of natural products: A comprehensive review. *Chin. Med.* **2018**, *13*, 20. [\[CrossRef\]](https://doi.org/10.1186/s13020-018-0177-x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29692864)
- 55. Płotka-Wasylka, J.; Szczepańska, N.; de La Guardia, M.; Namieśnik, J. Miniaturized solid-phase extraction techniques. *TrAC Trends Anal. Chem.* **2015**, *73*, 19–38. [\[CrossRef\]](https://doi.org/10.1016/j.trac.2015.04.026)
- 56. Andrade-Eiroa, A.; Canle, M.; Leroy-Cancellieri, V.; Cerdà, V. Solid-phase extraction of organic compounds: A critical review. part ii. *TrAC Trends Anal. Chem.* **2016**, *80*, 655–667. [\[CrossRef\]](https://doi.org/10.1016/j.trac.2015.08.014)
- 57. Jume, B.H.; Valizadeh Dana, N.; Rastin, M.; Parandi, E.; Darajeh, N.; Rezania, S. Sulfur-Doped Binary Layered Metal Oxides Incorporated on Pomegranate Peel-Derived Activated Carbon for Removal of Heavy Metal Ions. *Molecules* **2022**, *27*, 8841. [\[CrossRef\]](https://doi.org/10.3390/molecules27248841)
- 58. Sereshti, H.; Soltani, S.; Sridewi, N.; Salehi, E.; Parandi, E.; Rashid Nodeh, H.; Shahabuddin, S. Solid Phase Extraction Penicillin and Tetracycline in Human Serum Using Magnetic Graphene Oxide-Based Sulfide Nanocomposite. *Magnetochemistry* **2023**, *9*, 132. [\[CrossRef\]](https://doi.org/10.3390/magnetochemistry9050132)
- 59. Hadi Jume, B.; Parandi, E.; Nouri, M.; Aghel, B.; Gouran, A.; Rashidi Nodeh, H.; Kamyab, H.; Cho, J.; Rezania, S. Optimization of microreactor-assisted transesterification for biodiesel production using bimetal zirconium-titanium oxide doped magnetic graphene oxide heterogeneous nanocatalyst. *Chem. Eng. Process. Process Intensif.* **2023**, *191*, 109479. [\[CrossRef\]](https://doi.org/10.1016/j.cep.2023.109479)
- 60. Parandi, E.; Safaripour, M.; Mosleh, N.; Saidi, M.; Rashidi Nodeh, H.; Oryani, B.; Rezania, S. Lipase enzyme immobilized over magnetic titanium graphene oxide as catalyst for biodiesel synthesis from waste cooking oil. *Biomass Bioenergy* **2023**, *173*, 106794. [\[CrossRef\]](https://doi.org/10.1016/j.biombioe.2023.106794)
- 61. Hidayah, N.N.; Abidin, S.Z. The evolution of mineral processing in extraction of rare earth elements using liquid-liquid extraction: A review. *Miner. Eng.* **2018**, *121*, 146–157. [\[CrossRef\]](https://doi.org/10.1016/j.mineng.2018.03.018)
- 62. Silvestre, C.I.; Santos, J.L.; Lima, J.L.; Zagatto, E.A. Liquid–liquid extraction in flow analysis: A critical review. *Anal. Chim. Acta* **2009**, *652*, 54–65. [\[CrossRef\]](https://doi.org/10.1016/j.aca.2009.05.042)
- 63. Kruk, Z.A.; Yun, H.; Rutley, D.L.; Lee, E.J.; Kim, Y.J.; Jo, C. The effect of high pressure on microbial population, meat quality and sensory characteristics of chicken breast fillet. *Food Control* **2011**, *22*, 6–12. [\[CrossRef\]](https://doi.org/10.1016/j.foodcont.2010.06.003)
- 64. Kulawik, P.; Kumar Tiwari, B. Recent advancements in the application of non-thermal plasma technology for the seafood industry. *Crit. Rev. Food Sci. Nutr.* **2019**, *59*, 3199–3210. [\[CrossRef\]](https://doi.org/10.1080/10408398.2018.1510827) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30277810)
- 65. Preis, S.; Klauson, D.; Gregor, A. Potential of electric discharge plasma methods in abatement of volatile organic compounds originating from the food industry. *J. Environ. Manag.* **2013**, *114*, 125–138. [\[CrossRef\]](https://doi.org/10.1016/j.jenvman.2012.10.042) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23238056)
- 66. Niemira, B.A.; Sites, J. Cold plasma inactivates Salmonella Stanley and Escherichia coli O157: H7 inoculated on golden delicious apples. *J. Food Prot.* **2008**, *71*, 1357–1365. [\[CrossRef\]](https://doi.org/10.4315/0362-028X-71.7.1357)
- 67. Yang, L.; Chen, J.; Gao, J. Low temperature argon plasma sterilization effect on Pseudomonas aeruginosa and its mechanisms. *J. Electrost.* **2009**, *67*, 646–651. [\[CrossRef\]](https://doi.org/10.1016/j.elstat.2009.01.060)
- 68. Thirumdas, R.; Sarangapani, C.; Annapure, U.S. Cold plasma: A novel non-thermal technology for food processing. *Food Biophys.* **2015**, *10*, 1–11. [\[CrossRef\]](https://doi.org/10.1007/s11483-014-9382-z)
- 69. Pankaj, S.K.; Bueno-Ferrer, C.; Misra, N.; Milosavljević, V.; O'donnell, C.; Bourke, P.; Keener, K.; Cullen, P. Applications of cold plasma technology in food packaging. *Trends Food Sci. Technol.* **2014**, *35*, 5–17. [\[CrossRef\]](https://doi.org/10.1016/j.tifs.2013.10.009)
- 70. Misra, N.; Patil, S.; Moiseev, T.; Bourke, P.; Mosnier, J.; Keener, K.; Cullen, P. In-package atmospheric pressure cold plasma treatment of strawberries. *J. Food Eng.* **2014**, *125*, 131–138. [\[CrossRef\]](https://doi.org/10.1016/j.jfoodeng.2013.10.023)
- 71. Yun, H.; Kim, B.; Jung, S.; Kruk, Z.A.; Kim, D.B.; Choe, W.; Jo, C. Inactivation of Listeria monocytogenes inoculated on disposable plastic tray, aluminum foil, and paper cup by atmospheric pressure plasma. *Food Control* **2010**, *21*, 1182–1186. [\[CrossRef\]](https://doi.org/10.1016/j.foodcont.2010.02.002)
- 72. Jadhav, H.B.; Annapure, U. Consequences of non-thermal cold plasma treatment on meat and dairy lipids—A review. *Future Foods* **2021**, *4*, 100095. [\[CrossRef\]](https://doi.org/10.1016/j.fufo.2021.100095)
- 73. Paixão, L.; Fonteles, T.V.; Oliveira, V.S.; Fernandes, F.A.; Rodrigues, S. Cold plasma effects on functional compounds of siriguela juice. *Food Bioprocess Technol.* **2019**, *12*, 110–121. [\[CrossRef\]](https://doi.org/10.1007/s11947-018-2197-z)
- 74. Pankaj, S.K.; Wan, Z.; Keener, K.M. Effects of cold plasma on food quality: A review. *Foods* **2018**, *7*, 4. [\[CrossRef\]](https://doi.org/10.3390/foods7010004) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29301243)
- 75. Subrahmanyam, K.; Gul, K.; Sehrawat, R.; Allai, F.M. Impact of in-package cold plasma treatment on the physicochemical properties and shelf life of button mushrooms (*Agaricus bisporus*). *Food Biosci.* **2023**, *52*, 102425. [\[CrossRef\]](https://doi.org/10.1016/j.fbio.2023.102425)
- 76. Abouelenein, D.; Mustafa, A.M.; Nzekoue, F.K.; Caprioli, G.; Angeloni, S.; Tappi, S.; Castagnini, J.M.; Dalla Rosa, M.; Vittori, S. The Impact of Plasma Activated Water Treatment on the Phenolic Profile, Vitamins Content, Antioxidant and Enzymatic Activities of Rocket-Salad Leaves. *Antioxidants* **2023**, *12*, 28. [\[CrossRef\]](https://doi.org/10.3390/antiox12010028) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36670890)
- 77. Rao, W.; Li, Y.; Dhaliwal, H.; Feng, M.; Xiang, Q.; Roopesh, M.; Pan, D.; Du, L. The Application of Cold Plasma Technology in Low-Moisture Foods. *Food Eng. Rev.* **2023**, *15*, 86–112. [\[CrossRef\]](https://doi.org/10.1007/s12393-022-09329-9)
- 78. Qu, Z.; Chen, G.; Wang, J.; Xie, X.; Chen, Y. Preparation, structure evaluation, and improvement in foaming characteristics of fibrotic pea protein isolate by cold plasma synergistic organic acid treatment. *Food Hydrocoll.* **2023**, *134*, 108057. [\[CrossRef\]](https://doi.org/10.1016/j.foodhyd.2022.108057)
- 79. Zhou, J.; Qaing, S.; Yang, B.; Wang, Y.; Wang, J.; Yang, T.; Zhang, Y.; Chen, Y.; Li, S. Cold plasma treatment with alginate oligosaccharide improves the digestive stability and bioavailability of nutrient-delivered particles: An in vitro INFOGEST gastrointestinal study. *Int. J. Biol. Macromol.* **2023**, *232*, 123309. [\[CrossRef\]](https://doi.org/10.1016/j.ijbiomac.2023.123309)
- 80. Zeng, Z.; Wang, Y.; Xu, G.; Zhou, L.; Liu, C.; Luo, S. Peroxidase inactivation by cold plasma and its effects on the storage, physicochemical and bioactive properties of brown rice. *Food Biosci.* **2023**, *52*, 102383. [\[CrossRef\]](https://doi.org/10.1016/j.fbio.2023.102383)
- 81. Guo, J.; He, Z.; Ma, C.; Li, W.; Wang, J.; Lin, F.; Liu, X.; Li, L. Evaluation of cold plasma for decontamination of molds and mycotoxins in rice grain. *Food Chem.* **2023**, *402*, 134159. [\[CrossRef\]](https://doi.org/10.1016/j.foodchem.2022.134159)
- 82. Wang, S.; Liu, Z.; Zhao, M.; Gao, C.; Wang, J.; Li, C.; Dong, X.; Liu, Z.; Zhou, D. Chitosan-wampee seed essential oil composite film combined with cold plasma for refrigerated storage with modified atmosphere packaging: A promising technology for quality preservation of golden pompano fillets. *Int. J. Biol. Macromol.* **2023**, *224*, 1266–1275. [\[CrossRef\]](https://doi.org/10.1016/j.ijbiomac.2022.10.212)
- 83. Giannoulia, S.; Triantaphyllidou, I.-E.; Tekerlekopoulou, A.G.; Aggelopoulos, C.A. Mechanisms of Individual and Simultaneous Adsorption of Antibiotics and Dyes onto Halloysite Nanoclay and Regeneration of Saturated Adsorbent via Cold Plasma Bubbling. *Nanomaterials* **2023**, *13*, 341. [\[CrossRef\]](https://doi.org/10.3390/nano13020341) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36678094)
- 84. Mosleh, N.; Najmi, M.; Parandi, E.; Nodeh, H.R.; Vasseghian, Y.; Rezania, S. Magnetic sporopollenin supported polyaniline developed for removal of lead ions from wastewater: Kinetic, isotherm and thermodynamic studies. *Chemosphere* **2022**, *300*, 134461. [\[CrossRef\]](https://doi.org/10.1016/j.chemosphere.2022.134461) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35395264)
- 85. Wang, Q.; Lavoine, N.; Salvi, D. Cold atmospheric pressure plasma for the sanitation of conveyor belt materials: Decontamination efficacy against adherent bacteria and biofilms of Escherichia coli and effect on surface properties. *Innov. Food Sci. Emerg. Technol.* **2023**, *84*, 103260. [\[CrossRef\]](https://doi.org/10.1016/j.ifset.2022.103260)
- 86. Asghari, M.; Hosseinzadeh Samani, B.; Ebrahimi, R. Review on non-thermal plasma technology for biodiesel production: Mechanisms, reactors configuration, hybrid reactors. *Energy Convers. Manag.* **2022**, *258*, 115514. [\[CrossRef\]](https://doi.org/10.1016/j.enconman.2022.115514)
- 87. Ebrahimi, P.; Lante, A. Environmentally friendly techniques for the recovery of polyphenols from food by-products and their impact on polyphenol oxidase: A critical review. *Appl. Sci.* **2022**, *12*, 1923. [\[CrossRef\]](https://doi.org/10.3390/app12041923)
- 88. Bao, Y.; Reddivari, L.; Huang, J.-Y. Development of cold plasma pretreatment for improving phenolics extractability from tomato pomace. *Innov. Food Sci. Emerg. Technol.* **2020**, *65*, 102445. [\[CrossRef\]](https://doi.org/10.1016/j.ifset.2020.102445)
- 89. Yepez, X.; Illera, A.E.; Baykara, H.; Keener, K. Recent advances and potential applications of atmospheric pressure cold plasma technology for sustainable food processing. *Foods* **2022**, *11*, 1833. [\[CrossRef\]](https://doi.org/10.3390/foods11131833)
- 90. Fernandes, F.A.; Rodrigues, S. Cold plasma processing on fruits and fruit juices: A review on the effects of plasma on nutritional quality. *Processes* **2021**, *9*, 2098. [\[CrossRef\]](https://doi.org/10.3390/pr9122098)
- 91. Stevens, C.; Wilson, C.; Lu, J.; Khan, V.; Chalutz, E.; Droby, S.; Kabwe, M.; Haung, Z.; Adeyeye, O.; Pusey, L. Plant hormesis induced by ultraviolet light-C for controlling postharvest diseases of tree fruits. *Crop Prot.* **1996**, *15*, 129–134. [\[CrossRef\]](https://doi.org/10.1016/0261-2194(95)00082-8)
- 92. Cavalcanti, R.N.; Pimentel, T.C.; Esmerino, E.A.; de Freitas, M.Q.; Verruck, S.; Silva, M.C.; da Cruz, A.G. Cold Plasma. *Nov. Technol. Food Sci.* **2023**, 109–169. [\[CrossRef\]](https://doi.org/10.1002/9781119776376.ch4)
- 93. Surowsky, B.; Schlüter, O.; Knorr, D. Interactions of non-thermal atmospheric pressure plasma with solid and liquid food systems: A review. *Food Eng. Rev.* **2015**, *7*, 82–108. [\[CrossRef\]](https://doi.org/10.1007/s12393-014-9088-5)
- 94. Bao, Y.; Reddivari, L.; Huang, J.-Y. Enhancement of phenolic compounds extraction from grape pomace by high voltage atmospheric cold plasma. *LWT* **2020**, *133*, 109970. [\[CrossRef\]](https://doi.org/10.1016/j.lwt.2020.109970)
- 95. Farias, T.R.; Rodrigues, S.; Fernandes, F.A. Effect of dielectric barrier discharge plasma excitation frequency on the enzymatic activity, antioxidant capacity and phenolic content of apple cubes and apple juice. *Food Res. Int.* **2020**, *136*, 109617. [\[CrossRef\]](https://doi.org/10.1016/j.foodres.2020.109617)
- 96. Ahmadian, S.; Kenari, R.E.; Amiri, Z.R.; Sohbatzadeh, F.; Khodaparast, M.H.H. Effect of ultrasound-assisted cold plasma pretreatment on cell wall polysaccharides distribution and extraction of phenolic compounds from hyssop (*Hyssopus officinalis* L.). *Int. J. Biol. Macromol.* **2023**, *233*, 123557. [\[CrossRef\]](https://doi.org/10.1016/j.ijbiomac.2023.123557) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36740126)
- 97. Rodríguez, Ó.; Gomes, W.F.; Rodrigues, S.; Fernandes, F.A. Effect of indirect cold plasma treatment on cashew apple juice (*Anacardium occidentale* L.). *LWT* **2017**, *84*, 457–463. [\[CrossRef\]](https://doi.org/10.1016/j.lwt.2017.06.010)
- 98. Herceg, Z.; Kovačević, D.B.; Kljusurić, J.G.; Jambrak, A.R.; Zorić, Z.; Dragović-Uzelac, V. Gas phase plasma impact on phenolic compounds in pomegranate juice. *Food Chem.* **2016**, *190*, 665–672. [\[CrossRef\]](https://doi.org/10.1016/j.foodchem.2015.05.135)
- 99. Hemmati, V.; Garavand, F.; Khorshidian, N.; Cacciotti, I.; Goudarzi, M.; Chaichi, M.; Tiwari, B.K. Impact of cold atmospheric plasma on microbial safety, total phenolic and flavonoid contents, antioxidant activity, volatile compounds, surface morphology, and sensory quality of green tea powder. *Food Biosci.* **2021**, *44*, 101348. [\[CrossRef\]](https://doi.org/10.1016/j.fbio.2021.101348)
- 100. Zielinska, S.; Staniszewska, I.; Cybulska, J.; Zdunek, A.; Szymanska-Chargot, M.; Zielinska, D.; Liu, Z.-L.; Pan, Z.; Xiao, H.-W.; Zielinska, M. Modification of the cell wall polysaccharides and phytochemicals of okra pods by cold plasma treatment. *Food Hydrocoll.* **2022**, *131*, 107763. [\[CrossRef\]](https://doi.org/10.1016/j.foodhyd.2022.107763)
- 101. Almeida, F.D.L.; Cavalcante, R.S.; Cullen, P.J.; Frias, J.M.; Bourke, P.; Fernandes, F.A.; Rodrigues, S. Effects of atmospheric cold plasma and ozone on prebiotic orange juice. *Innov. Food Sci. Emerg. Technol.* **2015**, *32*, 127–135. [\[CrossRef\]](https://doi.org/10.1016/j.ifset.2015.09.001)
- 102. Liu, Z.; Zhao, W.; Zhang, Q.; Gao, G.; Meng, Y. Effect of cold plasma treatment on sterilizing rate and quality of kiwi turbid juice. *J. Food Process Eng.* **2021**, *44*, e13711. [\[CrossRef\]](https://doi.org/10.1111/jfpe.13711)
- 103. Leite, A.K.; Fonteles, T.V.; Miguel, T.B.; da Silva, G.S.; de Brito, E.S.; Alves Filho, E.G.; Fernandes, F.A.; Rodrigues, S. Atmospheric cold plasma frequency imparts changes on cashew apple juice composition and improves vitamin C bioaccessibility. *Food Res. Int.* **2021**, *147*, 110479. [\[CrossRef\]](https://doi.org/10.1016/j.foodres.2021.110479) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34399475)
- 104. Abedelmaksoud, T.G.; Hesarinejad, M.A.; Shokrollahi Yancheshmeh, B. The effect of cold plasma on the enzymatic activity and quality characteristics of mango pulp. *Res. Innov. Food Sci. Technol.* **2022**, *10*, 341–350.
- 105. Yodpitak, S.; Mahatheeranont, S.; Boonyawan, D.; Sookwong, P.; Roytrakul, S.; Norkaew, O. Cold plasma treatment to improve germination and enhance the bioactive phytochemical content of germinated brown rice. *Food Chem.* **2019**, *289*, 328–339. [\[CrossRef\]](https://doi.org/10.1016/j.foodchem.2019.03.061) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30955620)
- 106. Zhou, Y.-H.; Vidyarthi, S.K.; Zhong, C.-S.; Zheng, Z.-A.; An, Y.; Wang, J.; Wei, Q.; Xiao, H.-W. Cold plasma enhances drying and color, rehydration ratio and polyphenols of wolfberry via microstructure and ultrastructure alteration. *LWT* **2020**, *134*, 110173. [\[CrossRef\]](https://doi.org/10.1016/j.lwt.2020.110173)
- 107. Dakshayani, R.; Paul, A.; Mahendran, R. Cold Plasma-Induced Effects on Bioactive Constituents and Antioxidant Potential of Lotus Petal Powder. *IEEE Trans. Plasma Sci.* **2020**, *49*, 507–512. [\[CrossRef\]](https://doi.org/10.1109/TPS.2020.2995918)
- 108. Kim, H.-J.; Yong, H.I.; Park, S.; Kim, K.; Kim, T.H.; Choe, W.; Jo, C. Effect of atmospheric pressure dielectric barrier discharge plasma on the biological activity of naringin. *Food Chem.* **2014**, *160*, 241–245. [\[CrossRef\]](https://doi.org/10.1016/j.foodchem.2014.03.101) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24799234)
- 109. Kashfi, A.S.; Ramezan, Y.; Khani, M.R. Simultaneous study of the antioxidant activity, microbial decontamination and color of dried peppermint (*Mentha piperita* L.) using low pressure cold plasma. *LWT* **2020**, *123*, 109121. [\[CrossRef\]](https://doi.org/10.1016/j.lwt.2020.109121)
- 110. Seelarat, W.; Sangwanna, S.; Panklai, T.; Chaosuan, N.; Bootchanont, A.; Wattanawikkam, C.; Subcharoen, A.; Subcharoen, N.; Chanchula, N.; Boonyawan, D. Enhanced fruiting body production and bioactive phytochemicals from white Cordyceps militaris by blending Cordyceps militaris and using cold plasma jet. *Plasma Chem. Plasma Process.* **2023**, *43*, 139–162. [\[CrossRef\]](https://doi.org/10.1007/s11090-022-10292-w)
- 111. Umair, M.; Jabbar, S.; Lin, Y.; Nasiru, M.M.; Zhang, J.; Abid, M.; Murtaza, M.A.; Zhao, L. Comparative study: Thermal and non-thermal treatment on enzyme deactivation and selected quality attributes of fresh carrot juice. *Int. J. Food Sci. Technol.* **2022**, *57*, 827–841. [\[CrossRef\]](https://doi.org/10.1111/ijfs.15535)
- 112. Rashid, F.; Bao, Y.; Ahmed, Z.; Huang, J.-Y. Effect of high voltage atmospheric cold plasma on extraction of fenugreek galactomannan and its physicochemical properties. *Food Res. Int.* **2020**, *138*, 109776. [\[CrossRef\]](https://doi.org/10.1016/j.foodres.2020.109776)
- 113. Rezaei, S.; Ghobadian, B.; Ebadi, M.T.; Ghomi, H. Qualitative and quantitative assessment of extracted oil from Camelina sativa seed treated by dielectric-barrier discharge cold plasma. *Contrib. Plasma Phys.* **2020**, *60*, e202000032. [\[CrossRef\]](https://doi.org/10.1002/ctpp.202000032)
- 114. Afshar, S.; Ramezan, Y.; Hosseini, S. Physical and chemical properties of oil extracted from sesame (*Sesamum indicum* L.) and sunflower (*Helianthus annuus* L.) seeds treated with cold plasma. *J. Food Meas. Charact.* **2022**, *16*, 740–752. [\[CrossRef\]](https://doi.org/10.1007/s11694-021-01205-0)
- 115. Rezaei, S.; Ebadi, M.-T.; Ghobadian, B.; Ghomi, H. Optimization of DBD-Plasma assisted hydro-distillation for essential oil extraction of fennel (*Foeniculum vulgare* Mill.) seed and spearmint (*Mentha spicata* L.) leaf. *J. Appl. Res. Med. Aromat. Plants* **2021**, *24*, 100300. [\[CrossRef\]](https://doi.org/10.1016/j.jarmap.2021.100300)
- 116. Pragna, C.; Gracy, T.R.; Mahendran, R.; Anandharamakrishnan, C. Effects of microwave and cold plasma assisted hydrodistillation on lemon peel oil extraction. *Int. J. Food Eng.* **2019**, *15*, 20190093. [\[CrossRef\]](https://doi.org/10.1515/ijfe-2019-0093)
- 117. Lee, M.J.; Lee, H.-J.; Lee, Y.; Yang, J.Y.; Song, J.S.; Woo, S.Y.; Kim, H.Y.; Song, S.-Y.; Seo, W.D.; Son, Y.-J. Cold Plasma Treatment Increases Bioactive Metabolites in Oat (*Avena sativa* L.) Sprouts and Enhances In Vitro Osteogenic Activity of their Extracts. *Plant Foods Hum. Nutr.* **2023**, *78*, 146–153. [\[CrossRef\]](https://doi.org/10.1007/s11130-022-01029-3)
- 118. Ebadi, M.T.; Abbasi, S.; Harouni, A.; Sefidkon, F. Effect of cold plasma on essential oil content and composition of lemon verbena. *Food Sci. Nutr.* **2019**, *7*, 1166–1171. [\[CrossRef\]](https://doi.org/10.1002/fsn3.876)
- 119. Kodama, S.; Thawatchaipracha, B.; Sekiguchi, H. Enhancement of essential oil extraction for steam distillation by DBD surface treatment. *Plasma Process. Polym.* **2014**, *11*, 126–132. [\[CrossRef\]](https://doi.org/10.1002/ppap.201300047)
- 120. Kim, S.Y.; Lee, S.Y.; Min, S.C. Improvement of the antioxidant activity, water solubility, and dispersion stability of prickly pear cactus fruit extracts using argon cold plasma treatment. *J. Food Sci.* **2019**, *84*, 2876–2882. [\[CrossRef\]](https://doi.org/10.1111/1750-3841.14791)
- 121. Buonopane, G.J.; Antonacci, C.; Lopez, J.L. Effect of cold plasma processing on botanicals and their essential oils. *Plasma Med.* **2016**, *6*, 315–324. [\[CrossRef\]](https://doi.org/10.1615/PlasmaMed.2017019125)
- 122. Poomanee, W.; Wattananapakasem, I.; Panjan, W.; Kiattisin, K. Optimizing anthocyanins extraction and the effect of cold plasma treatment on the anti-aging potential of purple glutinous rice (*Oryza sativa* L.) extract. *Cereal Chem.* **2021**, *98*, 571–582. [\[CrossRef\]](https://doi.org/10.1002/cche.10399)
- 123. Patist, A.; Bates, D. Ultrasonic innovations in the food industry: From the laboratory to commercial production. *Innov. Food Sci. Emerg. Technol.* **2008**, *9*, 147–154. [\[CrossRef\]](https://doi.org/10.1016/j.ifset.2007.07.004)
- 124. Grzegorzewski, F.; Ehlbeck, J.; Schlüter, O.; Kroh, L.W.; Rohn, S. Treating lamb's lettuce with a cold plasma–Influence of atmospheric pressure Ar plasma immanent species on the phenolic profile of Valerianella locusta. *LWT—Food Sci. Technol.* **2011**, *44*, 2285–2289. [\[CrossRef\]](https://doi.org/10.1016/j.lwt.2011.05.004)
- 125. Boehm, D.; Heslin, C.; Cullen, P.J.; Bourke, P. Cytotoxic and mutagenic potential of solutions exposed to cold atmospheric plasma. *Sci. Rep.* **2016**, *6*, 21464. [\[CrossRef\]](https://doi.org/10.1038/srep21464)
- 126. Park, J.H.; Kumar, N.; Uhm, H.S.; Lee, W.; Choi, E.H.; Attri, P. Effect of nanosecond-pulsed plasma on the structural modification of biomolecules. *RSC Adv.* **2015**, *5*, 47300–47308. [\[CrossRef\]](https://doi.org/10.1039/C5RA04993H)
- 127. Ayala, A.; Muñoz, M.F.; Argüelles, S. Lipid peroxidation: Production, metabolism, and signaling mechanisms of malondialdehyde and 4-hydroxy-2-nonenal. *Oxidative Med. Cell. Longev.* **2014**, *2014*, 360438. [\[CrossRef\]](https://doi.org/10.1155/2014/360438)
- 128. Wende, K.; Schmidt, A.; Bekeschus, S. Safety aspects of non-thermal plasmas. In *Comprehensive Clinical Plasma Medicine: Cold Physical Plasma for Medical Application*; Springer: Berlin/Heidelberg, Germany, 2018; pp. 83–109.
- 129. Mehta, D.; Yadav, K.; Chaturvedi, K.; Shivhare, U.; Yadav, S.K. Impact of cold plasma on extraction of polyphenol from de-oiled rice and corn bran: Improvement in extraction efficiency, in vitro digestibility, antioxidant activity, cytotoxicity and anti-inflammatory responses. *Food Bioprocess Technol.* **2022**, *15*, 1142–1156. [\[CrossRef\]](https://doi.org/10.1007/s11947-022-02801-8)
- 130. Heslin, C.; Boehm, D.; Gilmore, B.F.; Megaw, J.; Bourke, P. Safety evaluation of plasma-treated lettuce broth using in vitro and in vivo toxicity models. *J. Phys. D Appl. Phys.* **2020**, *53*, 274003. [\[CrossRef\]](https://doi.org/10.1088/1361-6463/ab7ac8)
- 131. Los, A.; Ziuzina, D.; Van Cleynenbreugel, R.; Boehm, D.; Bourke, P. Assessing the biological safety of atmospheric cold plasma treated wheat using cell and insect models. *Foods* **2020**, *9*, 898. [\[CrossRef\]](https://doi.org/10.3390/foods9070898)
- 132. Carbone, K.; Garrigos, M.; Jimenez, A. Polyphenols: From wastes to high added value bio-products. In *Frontiers in Natural Product Chemistry*; Atta-ur-Rhaman, Ed.; Publishing Bentham Science: Cambridge, UK, 2016; Volume 2, pp. 115–178.
- 133. Khezerlou, A.; Jafari, S.M. Nanoencapsulated bioactive components for active food packaging. In *Handbook of Food Nanotechnology*; Elsevier: Amsterdam, The Netherlands, 2020; pp. 493–532.
- 134. Tian, W.; Chen, G.; Zhang, G.; Wang, D.; Tilley, M.; Li, Y. Rapid determination of total phenolic content of whole wheat flour using near-infrared spectroscopy and chemometrics. *Food Chem.* **2021**, *344*, 128633. [\[CrossRef\]](https://doi.org/10.1016/j.foodchem.2020.128633)
- 135. Del Mondo, A.; Smerilli, A.; Ambrosino, L.; Albini, A.; Noonan, D.M.; Sansone, C.; Brunet, C. Insights into phenolic compounds from microalgae: Structural variety and complex beneficial activities from health to nutraceutics. *Crit. Rev. Biotechnol.* **2021**, *41*, 155–171. [\[CrossRef\]](https://doi.org/10.1080/07388551.2021.1874284)
- 136. Albuquerque, B.R.; Heleno, S.A.; Oliveira, M.B.P.; Barros, L.; Ferreira, I.C. Phenolic compounds: Current industrial applications, limitations and future challenges. *Food Funct.* **2021**, *12*, 14–29. [\[CrossRef\]](https://doi.org/10.1039/D0FO02324H)
- 137. Gao, Y.; Yeh, H.-Y.; Bowker, B.; Zhuang, H. Effects of different antioxidants on quality of meat patties treated with in-package cold plasma. *Innov. Food Sci. Emerg. Technol.* **2021**, *70*, 102690. [\[CrossRef\]](https://doi.org/10.1016/j.ifset.2021.102690)
- 138. Gao, Y.; Zhuang, H.; Yeh, H.-Y.; Bowker, B.; Zhang, J. Effect of rosemary extract on microbial growth, pH, color, and lipid oxidation in cold plasma-processed ground chicken patties. *Innov. Food Sci. Emerg. Technol.* **2019**, *57*, 102168. [\[CrossRef\]](https://doi.org/10.1016/j.ifset.2019.05.007)
- 139. Yeh, H.-Y.; Line, J.E.; Hinton, A.; Gao, Y.; Zhuang, H. The effect of rosemary Extract and cold plasma treatments on bacterial community diversity in poultry ground meats. *Heliyon* **2019**, *5*, e02719. [\[CrossRef\]](https://doi.org/10.1016/j.heliyon.2019.e02719) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31687526)
- 140. Liao, X.; Li, J.; Muhammad, A.I.; Suo, Y.; Chen, S.; Ye, X.; Liu, D.; Ding, T. Application of a dielectric barrier discharge atmospheric cold plasma (Dbd-Acp) for Eshcerichia coli inactivation in apple juice. *J. Food Sci.* **2018**, *83*, 401–408. [\[CrossRef\]](https://doi.org/10.1111/1750-3841.14045)
- 141. Malien-Aubert, C.; Dangles, O.; Amiot, M.J. Color stability of commercial anthocyanin-based extracts in relation to the phenolic composition. Protective effects by intra-and intermolecular copigmentation. *J. Agric. Food Chem.* **2001**, *49*, 170–176. [\[CrossRef\]](https://doi.org/10.1021/jf000791o) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11170573)
- 142. Sruthi, N.; Josna, K.; Pandiselvam, R.; Kothakota, A.; Gavahian, M.; Khaneghah, A.M. Impacts of cold plasma treatment on physicochemical, functional, bioactive, textural, and sensory attributes of food: A comprehensive review. *Food Chem.* **2022**, *368*, 130809. [\[CrossRef\]](https://doi.org/10.1016/j.foodchem.2021.130809)
- 143. Farias, T.R.; Rodrigues, S.; Fernandes, F.A. Comparative study of two cold plasma technologies on apple juice antioxidant capacity, phenolic contents, and enzymatic activity. *J. Food Process. Preserv.* **2022**, *46*, e16871. [\[CrossRef\]](https://doi.org/10.1111/jfpp.16871)
- 144. Li, J.; Li, Z.; Ma, Q.; Zhou, Y. Enhancement of anthocyanins extraction from haskap by cold plasma pretreatment. *Innov. Food Sci. Emerg. Technol.* **2023**, *84*, 103294. [\[CrossRef\]](https://doi.org/10.1016/j.ifset.2023.103294)
- 145. Hou, Y.; Wang, R.; Gan, Z.; Shao, T.; Zhang, X.; He, M.; Sun, A. Effect of cold plasma on blueberry juice quality. *Food Chem.* **2019**, *290*, 79–86. [\[CrossRef\]](https://doi.org/10.1016/j.foodchem.2019.03.123)
- 146. Maria do Socorro, M.R.; Alves, R.E.; de Brito, E.S.; Pérez-Jiménez, J.; Saura-Calixto, F.; Mancini-Filho, J. Bioactive compounds and antioxidant capacities of 18 non-traditional tropical fruits from Brazil. *Food Chem.* **2010**, *121*, 996–1002.
- 147. Ramazzina, I.; Tappi, S.; Rocculi, P.; Sacchetti, G.; Berardinelli, A.; Marseglia, A.; Rizzi, F. Effect of cold plasma treatment on the functional properties of fresh-cut apples. *J. Agric. Food Chem.* **2016**, *64*, 8010–8018. [\[CrossRef\]](https://doi.org/10.1021/acs.jafc.6b02730) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27709918)
- 148. Yousefi, M.; Rahimi-Nasrabadi, M.; Pourmortazavi, S.M.; Wysokowski, M.; Jesionowski, T.; Ehrlich, H.; Mirsadeghi, S. Supercritical fluid extraction of essential oils. *TrAC Trends Anal. Chem.* **2019**, *118*, 182–193. [\[CrossRef\]](https://doi.org/10.1016/j.trac.2019.05.038)
- 149. Álvarez-Martínez, F.; Barrajón-Catalán, E.; Herranz-López, M.; Micol, V. Antibacterial plant compounds, extracts and essential oils: An updated review on their effects and putative mechanisms of action. *Phytomedicine* **2021**, *90*, 153626. [\[CrossRef\]](https://doi.org/10.1016/j.phymed.2021.153626)
- 150. Han, L.; Boehm, D.; Amias, E.; Milosavljević, V.; Cullen, P.; Bourke, P. Atmospheric cold plasma interactions with modified atmosphere packaging inducer gases for safe food preservation. *Innov. Food Sci. Emerg. Technol.* **2016**, *38*, 384–392. [\[CrossRef\]](https://doi.org/10.1016/j.ifset.2016.09.026)
- 151. Smet, C.; Baka, M.; Dickenson, A.; Walsh, J.L.; Valdramidis, V.P.; Van Impe, J.F. Antimicrobial efficacy of cold atmospheric plasma for different intrinsic and extrinsic parameters. *Plasma Process. Polym.* **2018**, *15*, 1700048. [\[CrossRef\]](https://doi.org/10.1002/ppap.201700048)
- 152. Wiegand, C.; Beier, O.; Horn, K.; Pfuch, A.; Tölke, T.; Hipler, U.-C.; Schimanski, A. Antimicrobial impact of cold atmospheric pressure plasma on medical critical yeasts and bacteria cultures. *Ski. Pharmacol. Physiol.* **2014**, *27*, 25–35. [\[CrossRef\]](https://doi.org/10.1159/000351353)
- 153. Hemmati, V.; Garavand, F.; Goudarzi, M.; Sarlak, Z.; Cacciotti, I.; Tiwari, B.K. Cold atmospheric-pressure plasma treatment of turmeric powder: Microbial load, essential oil profile, bioactivity and microstructure analyses. *Int. J. Food Sci. Technol.* **2021**, *56*, 2224–2232. [\[CrossRef\]](https://doi.org/10.1111/ijfs.14838)
- 154. Mehta, D.; Purohit, A.; Bajarh, P.; Yadav, K.; Shivhare, U.; Yadav, S.K. Cold plasma processing improved the extraction of xylooligosaccharides from dietary fibers of rice and corn bran with enhanced in-vitro digestibility and anti-inflammatory responses. *Innov. Food Sci. Emerg. Technol.* **2022**, *78*, 103027. [\[CrossRef\]](https://doi.org/10.1016/j.ifset.2022.103027)
- 155. Jeong, G.H.; Park, E.K.; Kim, T.H. Anti-diabetic effects of trans-resveratrol byproducts induced by plasma treatment. *Food Res. Int.* **2019**, *119*, 119–125. [\[CrossRef\]](https://doi.org/10.1016/j.foodres.2019.01.035)

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