

Figure S1. Resistance rates of ten antibiotics against 37 *tet(X)*-positive *Empedobacter* sp. strains. The resistance breakpoints of different antibiotics are also present. TGC, tigecycline; TC, tetracycline; CS, colistin; CIP, ciprofloxacin; MEM, meropenem; FFC, florfenicol; CTX, cefotaxime; GEN, gentamicin; AMK, amikacin; SXT, trimethoprim/sulfamethoxazole.

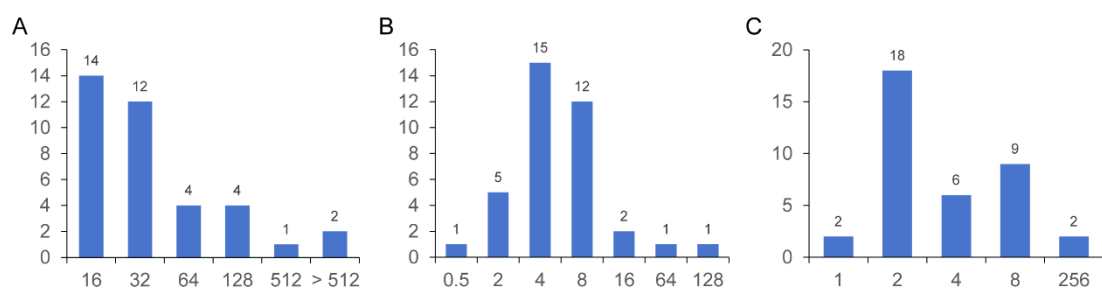


Figure S2. MICs of tylosin (A), tilmicosin (B), and tildipirosin (C) against 37 *tet(X)*-positive *Empedobacter* sp. strains. Bacterial numbers per dilution (mg/L) are present.

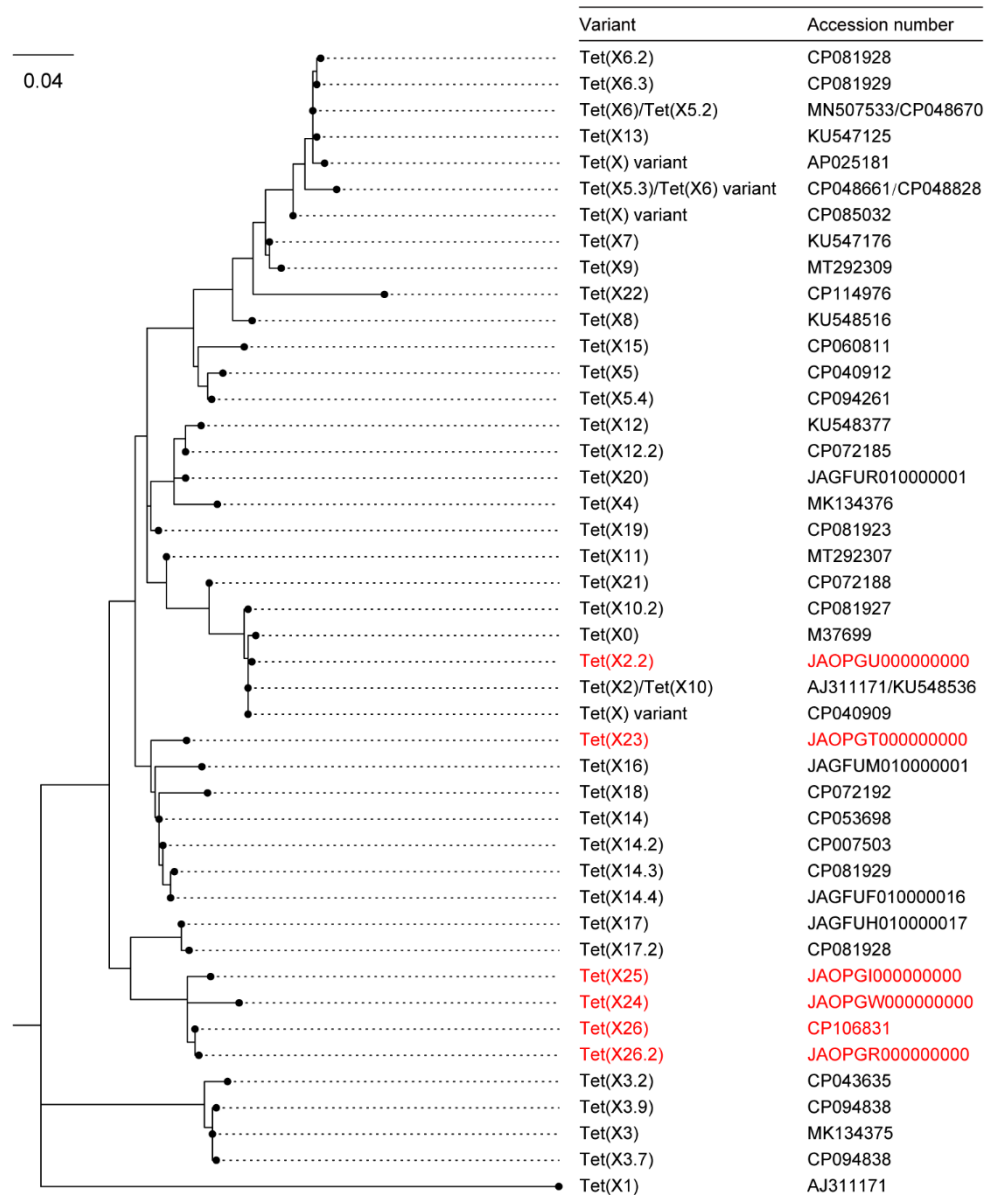


Figure S3. Maximum-likelihood phylogenetic tree of the Tet(X) variants. GenBank accession numbers are provided, of which the novel Tet(X) variants ($n = 6$) are marked in red. The proteins encoded by previously reported *tet(X3.3)* (CP094246), *tet(X3.4)* (CP094246), *tet(X3.5)* (CP094246), *tet(X3.6)* (CP094246), and *tet(X3.8)* (CP094838) are frameshifted, and therefore aren't involved in the phylogenetic analysis. Bar, 0.04 amino acid substitutions per site.

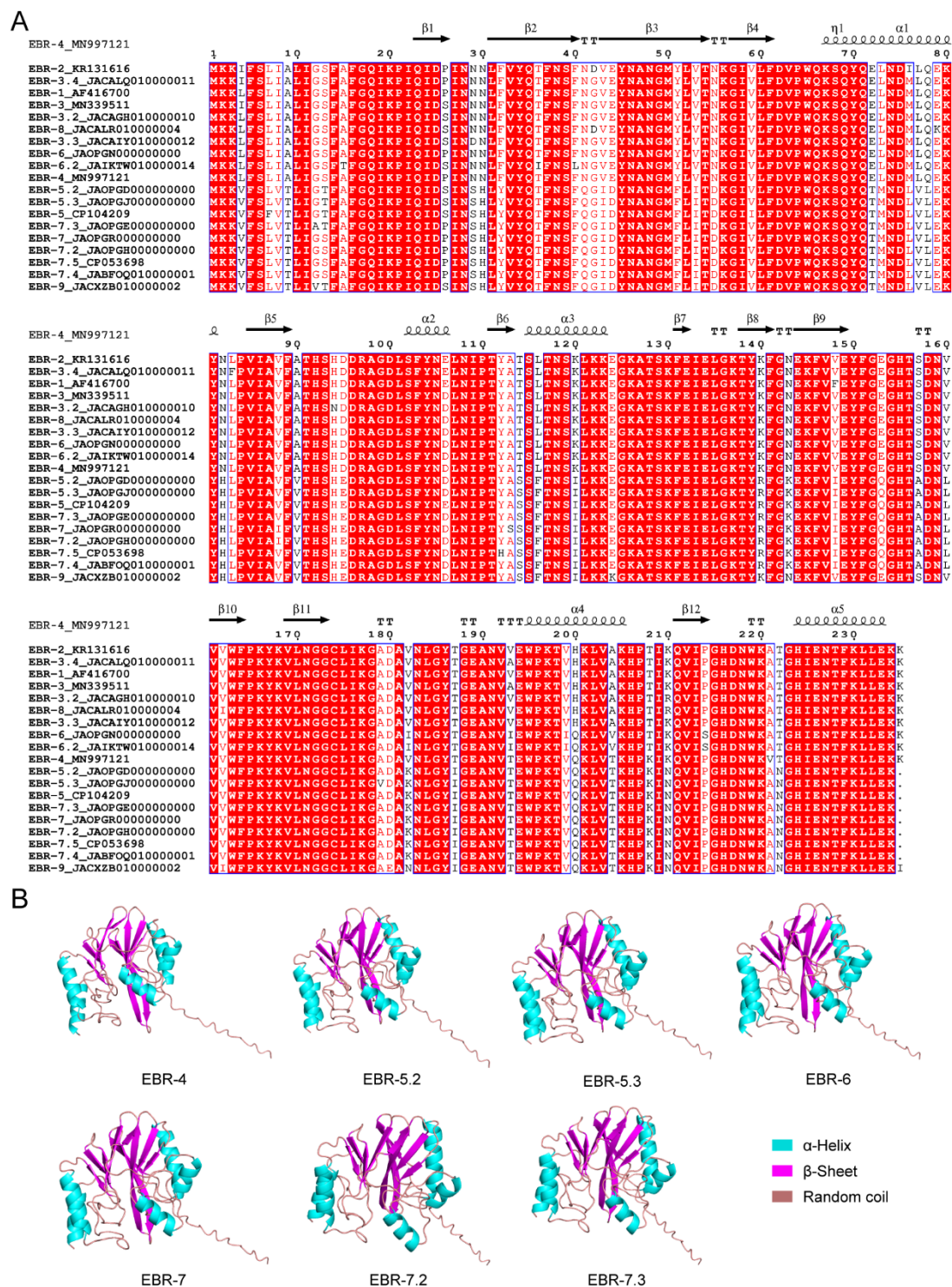


Figure S4. Structural characteristics of the EBR proteins. (A) Result of multiple sequence alignment by ESPrpt version 3.0. GenBank accession numbers of the EBR proteins are provided. (B) Prediction of three-dimensional structures by AlphaFold 3. The newly identified EBR variants are colored by their secondary structures and EBR-4 (MN997121) is used as the reference protein.

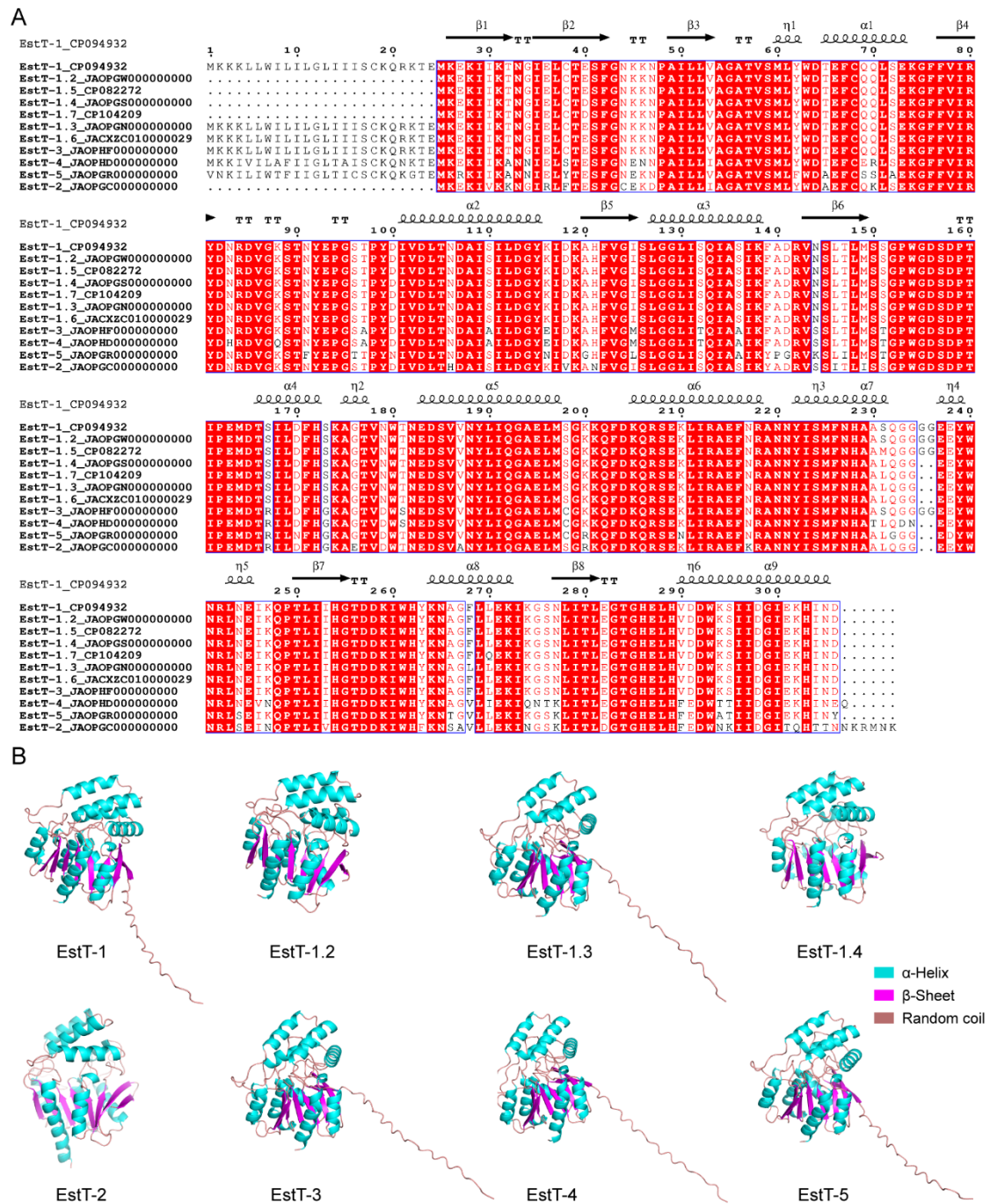


Figure S5. Structural characteristics of the EstT proteins. (A) Multiple sequence alignment of the EstT proteins by ESPrpt version 3.0. GenBank accession numbers of them are provided. (B) Prediction of three-dimensional structures by AlphaFold 3. The newly identified EstT variants are colored by their secondary structures and EstT-1 (CP094932) is used as the reference protein.

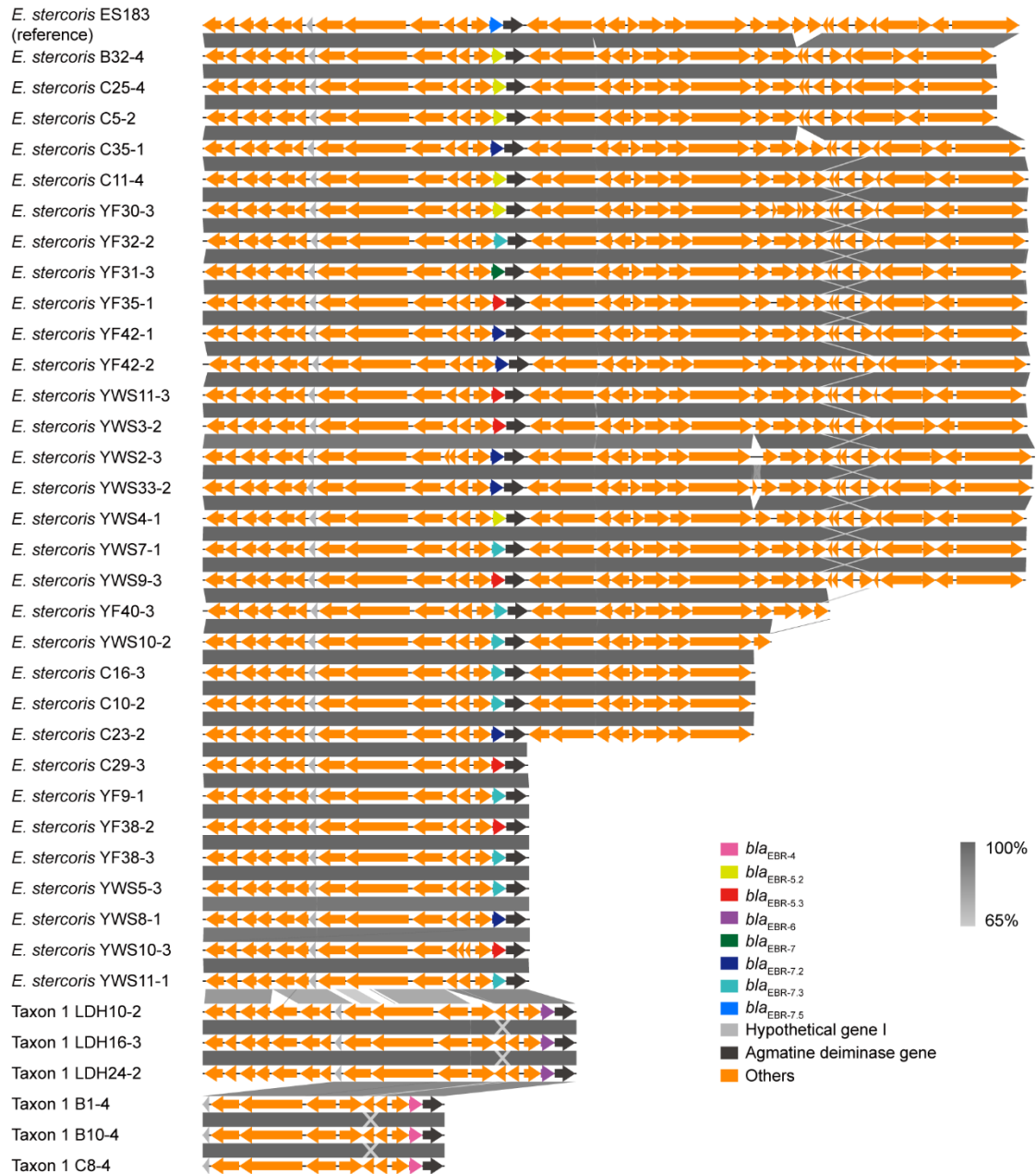


Figure S6. Conserved genetic backgrounds of the bla_{EBR} genes. The $bla_{EBR-7.5}$ -positive *E. stercoris* ES183 (GCA_014621655.1) is used as the reference strain. Regions of > 65% nucleotide identity are marked by shading.

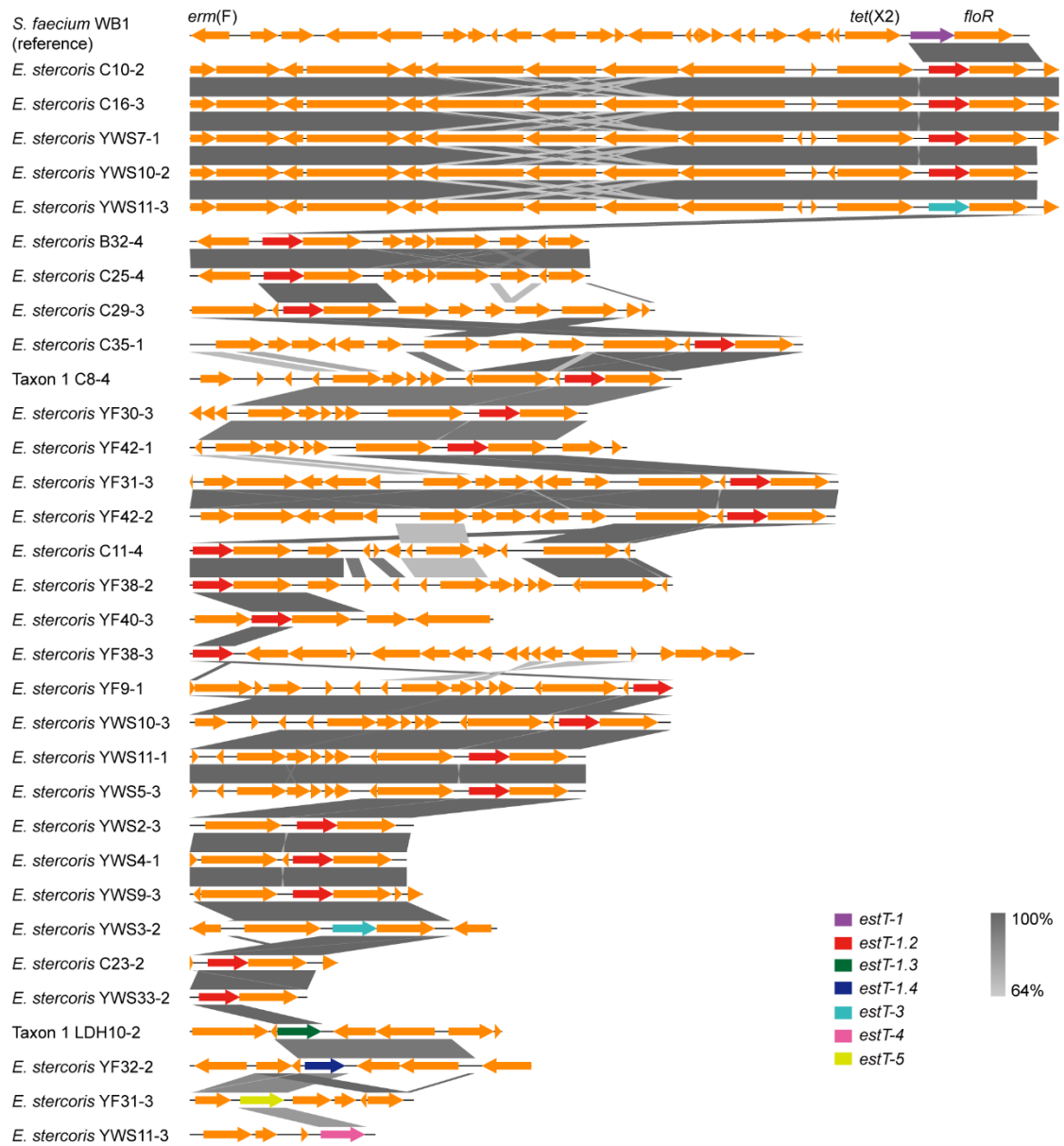


Figure S7. Diverse genetic backgrounds of *estT* genes. The *estT*-1-positive *S. faecium* WB1 (GCA_025340125.1) is used as the reference strain. Regions of > 64% nucleotide identity are marked by shading.

Table S1. Distribution of *tet(X)*- positive *Empedobacter* sp. isolates of different sources.

Province	Detection rate (percentage of positive strain/sample)				
	Chicken feces	Chicken carcass	Sheep feces	Soil	Sewage
Shandong	11.3% (22/195)	0.0% (0/60)	0.0% (0/0)	5.6% (1/18)	61.1% (11/18)
Jiangsu	2.5% (3/122)	0.0% (0/0)	0.0% (0/0)	0.0% (0/30)	0.0% (0/18)
Ningxia	0.0% (0/0)	0.0% (0/0)	0.0% (0/95)	0.0% (0/0)	0.0% (0/0)
In total	7.9% (25/317)	0.0% (0/60)	0.0% (0/95)	2.1% (1/48)	30.6% (11/36)

Table S2. Primers used in this study.

Primer	Nucleotide sequence (5' to 3') [†]	Size (bp)
<i>tet</i> (X)-uniF	TTCAGGTCAGGAAGCAATGAAAA	780 [1]
<i>tet</i> (X)-uniR	TTTACGCCTTGTCTGCAAAAGG	
16S rDNA-F	AGAGTTTGATCCTGGCTCAG	1450 [2]
16S rDNA-R	GGTTACCTTGTTACGACTT	
<i>tet</i> (X2.2)-F	<u>ttgggctagcaggaggaattc</u> ATGACAATGCGAATAGATACAGACAA	1167 ^{††}
<i>tet</i> (X2.2)-R	<u>cttgcctgcctgcagggtcgac</u> TTATACATTTAACAATTGCTGAAACGTAA	
<i>tet</i> (X23)-F	<u>ttgggctagcaggaggaattc</u> ATGAATTTACTTAGTGATAAGAACGTTGC	1137
<i>tet</i> (X23)-R	<u>cttgcctgcctgcagggtcgac</u> TTATACATTTAACAATTGCTGAAACGTAA	
<i>tet</i> (X24)-F	<u>ttgggctagcaggaggaattc</u> ATGACAATGCGAATAGATACAGACAA	1167
<i>tet</i> (X24)-R	<u>cttgcctgcctgcagggtcgac</u> TTATAGATTTCATTAGTTTTTGAACGAA	
<i>bla</i> _{EBR-4} -F	ttgggctagcaggaggaattcATGAAGAAATTATTTTCACTTATAGCATTG	708 ^{†††}
<i>bla</i> _{EBR-4} -R	<u>cttgcctgcctgcagggtcgac</u> TTATTTCTTTTCTAAAAGTTTAAACGTATTTT	
<i>bla</i> _{EBR-5.2} -F	<u>ttgggctagcaggaggaattc</u> ATGAAAAAAGTATTTTCTCTGTGCACATT	705 ^{††††}
<i>bla</i> _{EBR-5.2} -R	<u>cttgcctgcctgcagggtcgac</u> TTATTTTCTAAAAGTTTAAACGTATTTTCTAT	
<i>estT</i> -1.2-F	<u>ttgggctagcaggaggaattc</u> ATGAAAGAGAAAATAATTAACAAACGG	849
<i>estT</i> -1.2-R	<u>cttgcctgcctgcagggtcgac</u> TCAGTCATTTATGTGTTTTTCTATTCCA	
<i>estT</i> -1.3-F	<u>taccggttttttgggctagc</u> ATGAAAAAATACTTTTGATATTAATTT	915 ^{†††††}
<i>estT</i> -1.3-R	<u>cttgcctgcctgcagggtcgac</u> TCAGTCATTTATGTGTTTTTCTATTCCA	
<i>estT</i> -1.4-F	<u>taccggttttttgggctagc</u> ATGAAAGAGAAAATAATTAACAAACGG	843
<i>estT</i> -1.4-R	<u>cttgcctgcctgcagggtcgac</u> TCAGTCATTTATGTGTTTTTCTATTCCA	
<i>estT</i> -2-F	<u>taccggttttttgggctagc</u> ATGAAAGAGAAAATAGTTAAAAAAAACGG	861
<i>estT</i> -2-R	<u>cttgcctgcctgcagggtcgac</u> CTATTTGTTTCATTCTTTTGTTATTTGTTG	
<i>estT</i> -4-F	<u>taccggttttttgggctagc</u> ATGAAGAAAATAGTAATTTTAGCATTTATAATT	918
<i>estT</i> -4-R	<u>cttgcctgcctgcagggtcgac</u> TTATTGCTCATTATATGTTTTTCAATTC	
<i>estT</i> -5-F	<u>taccggttttttgggctagc</u> GTGAATAAAATACTAATTTGGACTTTTATAATT	915
<i>estT</i> -5-R	<u>cttgcctgcctgcagggtcgac</u> TTAGTAATTTATGTGCTTTTCAATTCCTT	
pBAD24-F	CGGCGTCACACTTTGCTATG	Cloning confirmation [3]
pBAD24-R	ACGGCGTTTCACTTCTGAGT	

[†] The homologous sequences are underlined.

^{††} The primers are used for cloning of *tet*(X2.2), *tet*(X25), *tet*(X26), and *tet*(X26.2) genes.

^{†††} The primers are used for cloning of *bla*_{EBR-4} and *bla*_{EBR-6} genes.

^{††††} The primers are used for cloning of *bla*_{EBR-5.2}, *bla*_{EBR-5.3}, *bla*_{EBR-7}, *bla*_{EBR-7.2}, and *bla*_{EBR-7.3} genes.

^{†††††} The primers are used for cloning of *estT*-1.3 and *estT*-3 genes.

Table S3. Summary of the *tet*(X) variants (n = 52, accessed on 16 Apr 2024).

Subtype	Accession number	Bacterial species	Size (bp)	Reference
<i>tet</i> (X0)	M37699	<i>Bacteroides fragilis</i>	1167	[4]
<i>tet</i> (X1)	AJ311171	<i>Bacteroides thetaiotaomicron</i>	1080	[5]
<i>tet</i> (X2) ⁺	AJ311171	<i>Bacteroides thetaiotaomicron</i>	1167	[5]
<i>tet</i> (X2.2)	JAOPGU000000000	<i>Empedobacter falsenii</i>	1167	This study
<i>tet</i> (X) variant	CP040909	<i>Empedobacter falsenii</i>	1137	[6]
<i>tet</i> (X3)	MK134375	<i>Acinetobacter baumannii</i>	1167	[7]
<i>tet</i> (X3.2)	CP043635	<i>Empedobacter brevis</i>	1167	[8]
<i>tet</i> (X3.3) ⁺⁺	CP094246	<i>Acinetobacter variabilis</i>	1166	[9]
<i>tet</i> (X3.4) ⁺⁺	CP094246	<i>Acinetobacter variabilis</i>	1164	[9]
<i>tet</i> (X3.5) ⁺⁺	CP094246	<i>Acinetobacter variabilis</i>	1165	[9]
<i>tet</i> (X3.6) ⁺⁺	CP094246	<i>Acinetobacter variabilis</i>	1165	[9]
<i>tet</i> (X3.7)	CP094838	<i>Acinetobacter schindleri</i>	1167	[9]
<i>tet</i> (X3.8) ⁺⁺	CP094838	<i>Acinetobacter schindleri</i>	1165	[9]
<i>tet</i> (X3.9)	CP094838	<i>Acinetobacter schindleri</i>	1167	[9]
<i>tet</i> (X4)	MK134376	<i>Escherichia coli</i>	1158	[7]
<i>tet</i> (X5)	CP040912	<i>Acinetobacter baumannii</i>	1167	[10]
<i>tet</i> (X5.2) ⁺⁺⁺	CP048670	<i>Acinetobacter</i> sp.	1137	[3]
<i>tet</i> (X5.3) ⁺⁺⁺⁺	CP048661	<i>Acinetobacter</i> sp.	1137	[3]
<i>tet</i> (X5.4)	CP094261	<i>Acinetobacter indicus</i>	1167	[11]
<i>tet</i> (X6) ⁺⁺⁺	MN507533	<i>Proteus genomospecies 6</i>	1137	[12]
<i>tet</i> (X6.2)	CP081928	<i>Riemerella anatipestifer</i>	1137	[13]
<i>tet</i> (X6.3)	CP081929	<i>Riemerella anatipestifer</i>	1137	[13]
<i>tet</i> (X6) variant ⁺⁺⁺⁺	CP048828	<i>Acinetobacter baumannii</i>	1137	[14]
<i>tet</i> (X) variant	AP025181	<i>Escherichia coli</i>	1137	[15]
<i>tet</i> (X) variant	CP085032	<i>Proteus terrae</i> subsp. <i>cibarius</i>	1137	[16]
<i>tet</i> (X7)	KU547176	Human enteric microorganism	1137	[17]
<i>tet</i> (X8)	KU548516	Human enteric microorganism	1137	[17]
<i>tet</i> (X9)	MT292309	Human enteric microorganism	1137	[17]
<i>tet</i> (X10) ⁺	KU548536	Human enteric microorganism	1167	[17]
<i>tet</i> (X10.2)	CP081927	<i>Riemerella anatipestifer</i>	1167	[13]
<i>tet</i> (X11)	MT292307	Human enteric microorganism	1167	[17]
<i>tet</i> (X12)	KU548377	Human enteric microorganism	1167	[17]
<i>tet</i> (X12.2)	CP072185	<i>Riemerella anatipestifer</i>	1167	[13]
<i>tet</i> (X13)	KU547125	Human enteric microorganism	1137	[17]
<i>tet</i> (X14)	CP053698	<i>Empedobacter stercoris</i>	1167	[18]
<i>tet</i> (X14.2)	CP007503	<i>Riemerella anatipestifer</i>	1167	[18]
<i>tet</i> (X14.3)	CP081929	<i>Riemerella anatipestifer</i>	1167	[13]
<i>tet</i> (X14.4)	JAGFUF010000016	<i>Riemerella anatipestifer</i>	1167	[13]
<i>tet</i> (X15)	CP060811	<i>Acinetobacter variabilis</i>	1158	[19]
<i>tet</i> (X16)	JAGFUM010000001	<i>Riemerella anatipestifer</i>	1167	[13]
<i>tet</i> (X17)	JAGFUH010000017	<i>Riemerella anatipestifer</i>	1167	[13]
<i>tet</i> (X17.2)	CP081928	<i>Riemerella anatipestifer</i>	1167	[13]
<i>tet</i> (X18)	CP072192	<i>Riemerella anatipestifer</i>	1137	[13]
<i>tet</i> (X19)	CP081923	<i>Riemerella anatipestifer</i>	1167	[13]
<i>tet</i> (X20)	JAGFUR010000001	<i>Riemerella anatipestifer</i>	1167	[13]
<i>tet</i> (X21)	CP072188	<i>Riemerella anatipestifer</i>	1167	[13]
<i>tet</i> (X22)	CP114976	<i>Pseudomonas caeni</i>	1137	[20]
<i>tet</i> (X23)	JAOPGT000000000	<i>Empedobacter falsenii</i>	1137	This study
<i>tet</i> (X24)	JAOPGW000000000	<i>Empedobacter falsenii</i>	1167	This study

<i>tet</i> (X25)	JAOPGI000000000	<i>Empedobacter falsenii</i>	1167	This study
<i>tet</i> (X26)	CP106831	<i>Empedobacter falsenii</i>	1167	This study
<i>tet</i> (X26.2)	JAOPGR000000000	<i>Empedobacter falsenii</i>	1167	This study

[†] *tet*(X2) (AJ311171) and *tet*(X10) (KU548536) share an 100% nucleotide identity.

^{††} The variants are frameshifted.

^{††} *tet*(X5.2) (CP048670) and *tet*(X6) (MN507533) share an 100% nucleotide identity.

^{†††} *tet*(X5.3) (CP048661) and the *tet*(X6) variant (CP048828) share an 100% nucleotide identity.

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