

Article

Using Zebrafish to Screen Developmental Toxicity of Per- and Polyfluoroalkyl Substances (PFAS)

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Abstract: Per- and polyfluoroalkyl substances (PFAS) are found in many consumer and industrial products. While some PFAS, notably perfluorooctanoic acid (PFOA) and perfluorooctanesulfonic acid (PFOS), are developmentally toxic in mammals, the vast majority of PFAS have not been evaluated for developmental toxicity potential. A concentration–response study of 182 unique PFAS chemicals using the zebrafish medium-throughput, developmental vertebrate toxicity assay was conducted to investigate chemical structural identifiers for toxicity. Embryos were exposed to each PFAS compound ($\leq 100 \mu\text{M}$) beginning on the day of fertilization. At 6 days post-fertilization (dpf), two independent observers graded developmental landmarks for each larva (e.g., mortality, hatching, swim bladder inflation, edema, abnormal spine/tail, or craniofacial structure). Thirty percent of the PFAS were developmentally toxic, but there was no enrichment of any OECD structural category. PFOS was developmentally toxic (benchmark concentration [BMC] = $7.48 \mu\text{M}$); however, other chemicals were more potent: perfluorooctanesulfonamide (PFOSA), N-methylperfluorooctane sulfonamide (N-MeFOSA), ((perfluorooctyl)ethyl)phosphonic acid, perfluoro-3,6,9-trioxatridecanoic acid, and perfluorohexane sulfonamide. The developmental toxicity profile for these more potent PFAS is largely unexplored in mammals and other species. Based on these zebrafish developmental toxicity results, additional screening may be warranted to understand the toxicity profile of these chemicals in other species.

Keywords: PFAS; perfluorooctanesulfonamide; larval zebrafish; in vivo; gross morphology; benchmark concentration (BMC); potency; high-throughput screening

1. Introduction

Per- and polyfluoroalkyl substances (PFAS) are a class of synthetic compounds used in everyday items, including cookware, textiles, food packaging, and electronics [1–3]. In the last decade, attention has been paid to PFAS due to their ubiquitous presence in environmental matrices, such as soil, water (ground and surface), as well as human blood (maternal and fetal cord), contributing to a better understanding and delineation of PFAS-related toxicity to human health and the environment [1,4–7]. The Organisation for Economic Co-operation and Development (OECD) has recently characterized PFAS as having at least one fully fluorinated methyl or methylene carbon without any H/Cl/Br/I atoms attached to it [8]. The fluorinated methyl or methylene bond imparts properties that render PFAS as generally possessing high stability, low reactivity, and varying levels of bioactivity [2]. Estimates of the number of PFAS in the environment vary depending on the PFAS definition applied. As per the Toxic Substances Control Act (TSCA) 8(a)(7) rule (EPA, 2024), there are approximately 13,000 PFAS (which can be represented by a discrete chemical structure). It is estimated that ~650 are included as part of the non-confidential TSCA inventory and are still actively being produced and used in commercial products, with an unknown number of degradation products and manufacturing byproducts [9].

While much is still unknown about the adverse effects of PFAS, a subset of legacy compounds, namely perfluorooctanesulfonic acid (PFOS), perfluorooctanoic acid (PFOA), and related perfluoroalkyl acids (PFAAs) [10,11], have been well-studied, leading to the publication of lifetime drinking water health advisories for selected PFAS [12]. Many of the health effects linked to PFAS exposure were also associated with some legacy compounds, including changes in immune and thyroid function, reproductive challenges, liver disease, and cancer [10]. This extensive range of effects could be due to PFAS exposures varying by structure, route, or duration. In addition, the bioaccumulation potential of these compounds could lead to differences in internal doses for both human and environmentally relevant species [7,10,13,14]. A recent systematic review of the PFAS literature focusing on mammalian toxicological and epidemiological studies revealed that one of the primary targets of PFAS toxicity was developmental processes, and that there is an overall poor understanding of developmental toxicity potential for the majority of PFAS included in that review [15]. Although these reviews and mammalian studies have advanced the understanding of PFAS-related human adverse effects, only a small proportion of PFAS have any *in vivo* toxicity data. Thus, there is a need for rapid, high-throughput, screening tools to prioritize this diverse set of chemicals, especially for developmental toxicity.

Zebrafish (*Danio rerio*), small freshwater fish native to Southeast Asia, have been utilized in many toxicological studies as a model for developmental toxicity assessments (e.g., [16–19]). Zebrafish share many developmental signaling pathways, organ systems, metabolism, and brain structure/functions with mammals [20–24], leading to the relatively high concordance of developmental toxicity outcomes with other vertebrates [16,19,25,26]. There has been an increase in zebrafish PFAS investigations for developmental toxicity, but most of the research has only focused on a few PFAS (selected examples, [27–31]). Recently, however, larger PFAS chemical screens have utilized early life-stage zebrafish. One study, using dechorionated embryos exposed for five days to 139 PFAS, showed developmental and neurodevelopmental toxicity linked to chemical volatility and structural features, suggesting that grouping these chemicals may aid in identifying toxicity [32]. Similarly, it has been reported [33] that, by varying exposure windows and duration (4–72 h), a selection of PFAS chemicals ($n = 38$) induced developmental toxicity, in particular hepatotoxicity, with lipid transport potentially playing a role in these observations. Another screening effort testing 74 PFAS in developing zebrafish [34] associated chemical structural features to bioconcentration factors and metabolic pathways in larvae exposed to either 0.5 or 5.0 μM of each PFAS. Despite using different experimental designs and endpoints, perfluorooctanesulfonamide (PFOSA) was among the most potent developmentally toxic PFAS across these studies [32–34].

In 2019, the U.S. Environmental Protection Agency (EPA) began a coordinated research effort to screen a large PFAS library using an array of different *in vitro* and *in vivo* high- and medium-throughput toxicity assays to inform chemical category and read-across approaches [35]. As part of that screening effort, an *in vivo*, developing zebrafish model was employed to assess the developmental toxicity of 182 unique PFAS from the EPA PFAS chemical library. Various landmarks of development in zebrafish larvae were assessed following PFAS developmental exposure, and the results were combined with previously published information on the chemical purity assessments of the stocks used to treat the zebrafish [36]. This type of quality control (QC) check at this stage of experimentation has not been included in all the PFAS screens mentioned above and, given that a significant number (55/182: 30%) of the chemicals failed this basic stock quality assessment, analytical chemical QC could be a significant confounder of data interpretation. The present results were also compared to previously published data on the developmental effects of PFAS in zebrafish embryos/larvae to gain a better understanding of the manner in which these chemicals affect zebrafish in various developmental assays.

2. Materials and Methods

2.1. Experimental Animals

All research and breeding procedures in this study were reviewed and approved by the Office of Research and Development's Health Institutional Animal Care and Use Committee (IACUC) at the U.S. EPA in Research Triangle Park, NC (Protocol #21-08-003; approved 8 August 2018, and Protocol #24-09-002; approved 2 September 2021). The animal facility is an internationally accredited Association for Assessment and Accreditation of Laboratory Animal Care (AAALAC) facility (Unit# 000509). The parental fish were wild-type adult zebrafish (*Danio rerio*) descended from an undefined outbred stock originally supplied by both Aquatic Research Organisms, Hampton, NH, and EkkWill Waterlife Resources, Ruskin, FL, USA. The adult zebrafish were maintained at a density of 7 fish/L in 3.5 L tanks and housed in recirculating zebrafish housing racks (Tecniplast USA, West Chester, PA, USA) with reverse osmosis-purified tap water (Durham, NC, USA), which was buffered with Instant Ocean Sea Salt (Spectrum Brands, Blacksburg, VA, USA) and sodium bicarbonate (Church & Dwight, Co., Ewing, NJ, USA). The water was maintained at 28 °C, pH 7.4, and conductivity (1000 µS/cm), with ammonia and nitrite (maintained at 0 ppm) and nitrate (allowed in insignificant amounts). The fish were fed twice a day with decapsulated artemia (E-Z Egg; Brine Shrimp Direct, Ogden, UT, USA) and Gemma Micro 300 formulated diet (Skretting, Westbrook, ME, USA). The housing rooms were illuminated according to a 14:10 h light:dark cycle (lights on at 07:00 h). For embryo production, groups of approximately 150 same-age mixed sex zebrafish (ages ranging from 3 to 15 months old) were moved into 16 L on rack recirculating spawning tanks (Z-Park tanks, Tecniplast USA, West Chester, PA, USA) about one week before embryos were needed. Then, on the afternoon before embryos were needed, mesh spawning platform inserts were added. Embryos were collected the following morning approximately 45 min after the room lights came on (07:45 h) and were maintained at 28 °C for 1 to 2 h until washing. A diagram of the experimental methods is included in Figure 1.

2.2. Embryo Rearing

The embryos were placed in 600 mL beakers and kept at 28 °C until washing [37] two times with 0.06% bleach (*v/v*) in 10% Hanks' Balanced Salt Solution (13.7 mM NaCl, 0.54 mM KCl, 25 µM Na₂HPO₄, 44 µM KH₂PO₄, 130 µM CaCl₂, 100 µM MgSO₄, and 420 µM NaHCO₃ [hereinafter referred to as Hanks']) for 5 min each wash, then rinsed with Hanks' alone after each bleach wash. Immediately following washing, the embryos were examined, and healthy embryos were separated from dead or unfertilized eggs and moved into fresh Hanks'.

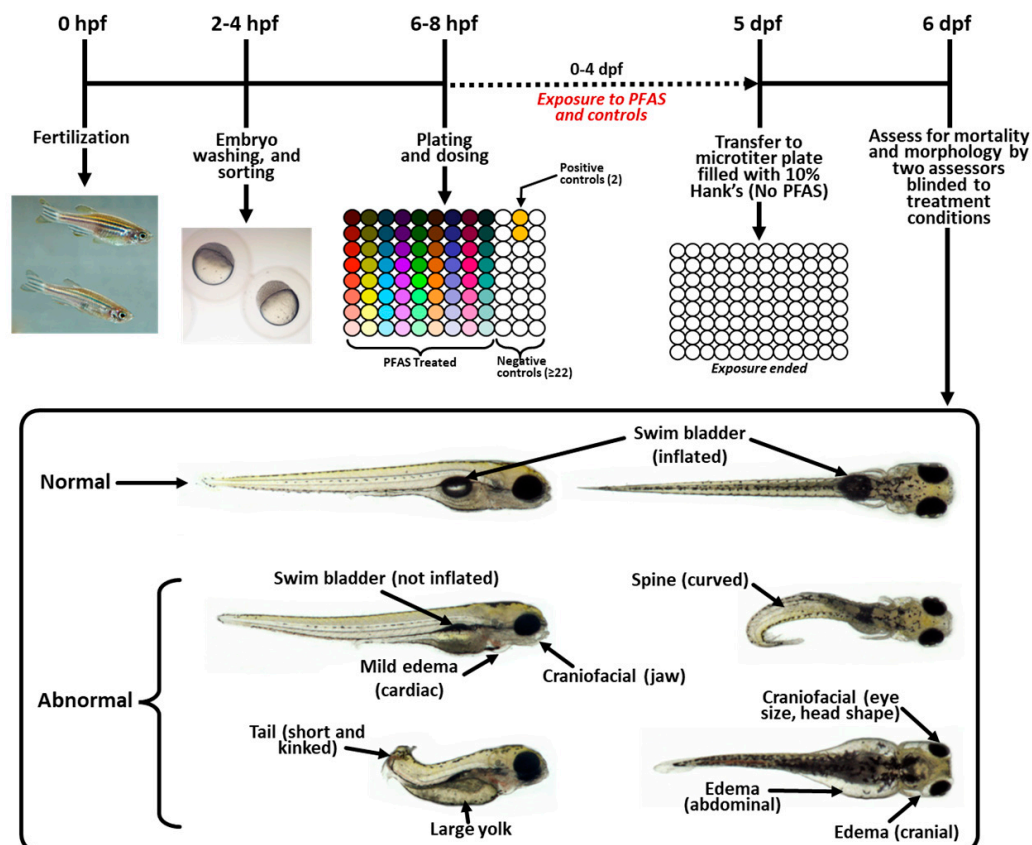


Figure 1. Experimental design: Experimental events from 0 h post-fertilization (hpf) through 6 days post-fertilization (dpf). Embryo washing, plating, and chemical exposure occurred on 0 dpf. Each column of colored circles on the 6–8 hpf plate represents a different chemical. Morphological assessments were conducted on 6 dpf by two experimenters blinded to treatment group information. Images show a normal embryo as well as several of the common developmental malformations identified in this analysis.

2.3. Chemical Exposure

All PFAS were received from Evotec Inc. (Branford, CT, USA) at concentrations ranging from 5 to 30 mM, solubilized in dimethyl sulfoxide (DMSO). All researchers were blinded to the chemical identity until after all data were collected and analyzed. Stock plates were first prepared with the highest concentration of each chemical, dependent on their level of initial solubilization provided by Evotec. The final concentration of the DMSO vehicle in every well was 0.4% (*v/v*). A single-concentration screening approach was implemented whereby the highest concentration of each chemical was tested to determine which PFAS were likely to be positive for developmental toxicity. A positive chemical was liberally defined, selecting those where 2/6 (33.3%) or more of the larvae per condition appeared to be affected either by death and/or malformations. An additional multiple-concentration screening of the positive chemicals ($n = 87$) was then conducted along with $\geq 15\%$ ($n = 13$; PFAS selected by a random number generator) of the chemicals that were negative in the single-concentration screen.

The multiple-concentration PFAS screening plates were made up with the highest concentration of each positive chemical as well as the chosen negatives and then were serially diluted with DMSO to produce an 8-point concentration–response curve (half-log dilution interval). Each experimental plate for both the single high-concentration test (described above) and the multiple concentration–response assessment contained ≥ 22 vehicle control wells (DMSO; $>99.9\%$ purity; Sigma-Aldrich (St. Louis, MO, USA); Chemical Abstract Services [CAS] number 67-68-5; 0.4% *v/v* final concentration) and

2 positive control wells (both containing chlorpyrifos [97% purity; Sigma-Aldrich]); CAS number 2921-88-2; 30 μM final concentration; extensive historical data show that this chlorpyrifos concentration will cause either death or severe malformations in zebrafish embryos by 6 dpf. A typical stock plate held nine PFAS as part of the 8-point concentration response curve and was used to dose six identical plates for each group of nine chemicals. Since the plates contained an arrangement with a single well for each concentration of each of the nine chemicals, six identical plates dosed from that same stock plate that produced a total $n = 6$ wells per chemical per dose. All final exposure concentrations for each chemical are listed in Supplemental Table S1, Col F of the second sheet.

Between 6 and 8 h post-fertilization (hpf), healthy embryos (with chorions) were transferred, one embryo per well, into 96-well (0.5 mL) microtiter plates (Cell Culture-Treated, Flat-Bottom Microplate 96 well [Corning™ Costar™, Kennebunk, ME, USA; Cat # 09-761-145]) filled with 200 μL of Hanks' solution. A random number generator was used to assign the order in which the rows of embryos were plated in the 96-well plates each week. After plating, the embryos were dosed with 0.8 μL of the chemical dosing solution, then immediately sealed with AlumaSeal II™ (Excel Scientific Inc., Victorville, CA, USA) to prevent the volatilization of the chemicals, and then placed in a leakproof secondary container. These containers were placed in an incubator maintained on a 14:10 h light:dark cycle at 26 °C for rearing. On 5 dpf, live larvae were gently moved out of the plate with test chemicals and into a new 96-well mesh plate (Millipore Corp., Bedford, MA, USA), just with the Hanks' solution. Once within the mesh well plate, the animals were rinsed with 400 μL of fresh Hanks' three times, and then the plates were covered with a non-adhesive material (Microseal® A, BioRad, Hercules, CA, USA), the plate lid was added, and then the plate sides were wrapped in Parafilm™ (PM992, Bermis Company, Neenah, WI, USA) and returned to the incubator. This rinse on 5 dpf was conducted to lessen the possible exposure of the human assessors to the chemicals during the detailed examination of each larva on 6 dpf.

2.4. Larval Assessments

On 6 dpf between 8 and 10 AM, two individual assessors, blinded to the chemical treatments, independently examined each larva for mortality, hatching status, and malformations using an Olympus SZH10 stereo microscope. Mortality was defined as a lack of heartbeat or presence of coagulation. Malformations were defined as uninflated swim bladder, craniofacial defects, edema, spinal defects, decreased pigmentation, abnormal position in water column, tail defects, or blood pooling (some examples can be viewed in Figure 1; data are in Supplemental Table S1). If more than 15% of the negative control larvae were abnormal (i.e., dead, not hatched, or malformed) or if the positive control larvae were not at least 50% abnormal, that plate was not used for any analysis. Over the course of the experiment, there were no plates that needed to be removed from analysis based on the criteria above. Additionally, the overall rate of normal animals in the controls for the entire study was 97% (Supplemental Table S1). After all plates were assessed, the larvae were anesthetized using cold shock, and then euthanized with a cold 20% bleach solution.

2.5. Concentration–Response Modeling

The raw data from the larval assessments consisted of counts of larvae observed at each of the twelve endpoints: living (dead or alive), hatching (hatched or unhatched), swim bladder (non-inflation), craniofacial defects (dysmorphology of the head or eyes), edema, spinal defects (curved spine), pigmentation, position (in water column, either persistent lying on one side or upside down), tail defects (e.g., kinks), or blood pooling (Supplemental Table S1). Each assessor also assigned every larva a general ranking for the overall condition: normal (no defects present), abnormal (defects present), or severely abnormal (life-threatening defects present). Observations were recorded for each chemical and concentration. If any of these defects were present in a larva, the additional endpoint “any” was set to 1.

Concentration–response data were processed using the ToxCast Data Analysis Pipeline (tcpl) R-package (tcpl v.3.2.0; <https://cran.r-project.org/web/packages/tcpl/index.html>; accessed on 4 March 2024 [38,39]). These endpoints are anticipated to be released in Fall 2024 with ToxCast’s InvitroDB v4.2 at <https://doi.org/10.23645/epacomptox.6062623> (accessed on 8 April 2024). For the endpoint–chemical per concentration index, counts were aggregated to a percentage, called endpoint scores, with dead larvae excluded. For example, if there were 5 (out of 6) living larvae and 2 had edema at a tested concentration, the edema score would equal $2/5 \times 100 = 40\%$. No additional normalization was performed, and outliers were not excluded. For each endpoint–chemical pair, the concentration–response series was fit to 5 bounded models (constant, hill, gain–loss, exponential 4–5), with the winning model selected by the lowest Akaike Information Criteria (AIC) score, a statistical calculation that compares model quality. Unbounded models available in tcpl were excluded given the dichotomous nature of observations. To estimate activity, a cutoff threshold was set at 16% and a continuous hit call (hitc) value was determined as the product of the following components: (1) at least one median response greater than the assay cutoff threshold, (2) the maximal efficacy in the fitted response is larger than the assay cutoff, and (3) the AIC score of the winning model is less than the constant model [40]. Classification criteria for continuous hit calls were set in line with other in vitro screening efforts as: hitc = 0 as negative, $0 < \text{hitc} < 0.9$ as equivocal, and $\text{hitc} \geq 0.9$ as positive [41]. In addition to the estimated activity concentrations inducing a specified level of responses (e.g., 10%, 50%, etc.), a benchmark concentration (BMC) was also derived in tcpl using a specified benchmark response level (BMR) of 1.349 times the standard deviation of the baseline (10%) [42]. Given the lack of baseline variability in the dichotomous observations, the baseline median absolute deviation was set as 5%.

3. Results

The raw data collected for each larva at 6 days post-fertilization (dpf) from each assessor are shown in Supplemental Table S1. These data then underwent concentration–response modeling (described in detail in Section 2) for the twelve developmental endpoints, and benchmark concentrations (BMCs) were calculated for each positive endpoint/chemical. All the BMC graphs for the positive endpoints for each chemical are presented in Supplemental Figure S1. The calculations and results for the BMC determinations are presented in Supplemental Table S2. Figure 2 shows an example of the best fit BMC dose–response modeling results with a description of the information provided on each graph. In the example, the chemical is PFOSA, and the endpoint is swim bladder non-inflation (“score.swim_bladder”). The black circles represent the fraction of larvae that were positive for that morphological endpoint at the given concentration in micromolar (noted on the *x*-axis; “1e+00” $\equiv 1 \times 10^0 \equiv 1 \mu\text{M}$). The black Xs are the fraction of animals that were alive and morphologically normal at that concentration; in this example, 6/6 animals at the four highest concentrations (3, 10, 30, and 100 μM) were dead at 6 dpf. The black curve is the best fit model to the concentration–response data, and the “mthd” listed at the top is the name of best fit model function (all possible functions are shown in Figure 2 of Ref. [38]). The gray band surrounding the *x*-axis indicates the background noise cutoff level.

A total of 185 PFAS chemicals (182 unique) were assessed for developmental toxicity in zebrafish embryos/larvae, which included measurements of malformation or mortality. Of those 185 chemicals, 56 (30%) caused developmental toxicity. Three PFAS were tested twice and were highly reproducible: 1,6-diiodoperfluorohexane tested positive both times, with potencies within an order of magnitude; and pentafluoropropanoic anhydride and 1H,1H-perfluorooctylamine, both of which tested negative both times. These duplicate chemicals are highlighted in yellow in Supplemental Table S2. Therefore, ignoring the duplicates, 55/182 (30%) PFAS caused developmental toxicity.

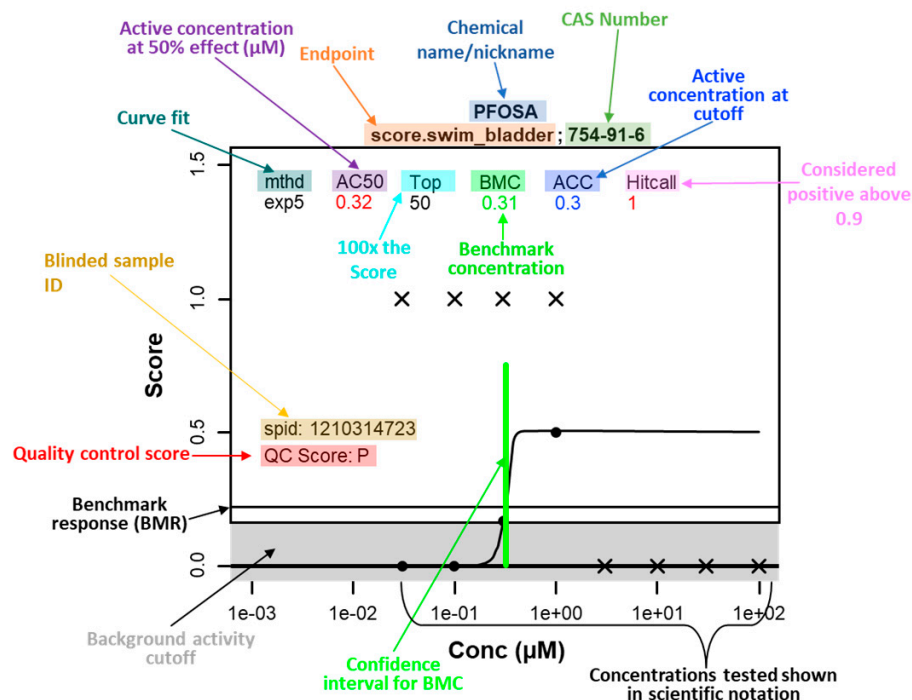


Figure 2. BMC determination example: Concentration–response data for a given chemical and endpoint are presented in this example; the chemical name is noted in the first line of the title, and the endpoint and CAS numbers are noted in the second line of the title. In the example, the chemical is PFOSA, and the endpoint is swim bladder non-inflation (“score.swim_bladder”). Score refers to percentage of animals affected divided by 100. The black circles represent the fraction of larvae that were positive for that morphological endpoint at the given concentration in micromolar (noted on the x -axis; “1e+00” $\equiv 1 \times 10^0 \equiv 1 \mu\text{M}$). The black Xs are the fraction of larvae that were alive and morphologically normal at that concentration; in this example, all 6/6 animals at the four highest concentrations (3, 10, 30, and 100 μM) were dead at 6 dpf. The black curve is the best fit model to the concentration–response data, and the “mthd” listed at the top is the name of best fit model function (all possible functions are shown in Figure 2 of Ref. [38]). The gray band near the bottom of the graph (0–0.16) indicates the background noise cutoff level. The horizontal line above the cutoff is the benchmark response (BMR) and is equal to $1.349 \times$ cutoff of 0.16 [42]. For an active response (i.e., hitcall), the point at which the BMR intersects the model curve is the benchmark concentration (BMC, μM), indicated by the vertical green line, with the width representing the 95% confidence interval. In the lower left corner of each graph, the sample ID (spid) and the analytical quality control (QC) score for that sample (P = pass or F = fail: [36]) are shown. For more information on curve fitting visit <https://clowder.edap-cluster.com/files/659c5239e4b063812d5d00cc?dataset=64b81ac2e4b08a6b5a3c2528&space=647f710ee4b08a6b394e426b> (accessed on 6 May 2024).

One strength of the present study is that the same stock chemicals tested in the zebrafish were also assessed for chemical presence and purity [36]. Chemicals that passed quality control (QC) were the ones where the compound was present by molecular match; fragmentation patterns putatively confirmed structure; peak area percentage was $>85\%$ of the total response; and/or instrument attenuation was less than the threshold based on signal-to-noise ratio or general peak height. A chemical passed the quality control check if it was detected with analytical instrumentation according to the above criteria and failed if either it was not detected or if degradation had occurred. Note that concentration was not determined for most of the chemical stocks, primarily because standards were not available. In the few situations where a standard was available and the concentration was estimated (Table 5 of Ref. [36]), many of the chemicals were only 60% to 90% of their target concentration.

If the quality control data for the chemical stocks were combined with the zebrafish developmental data (Table 1 or Figure 3), 127 (70%) of the 182 unique chemicals passed the quality control. Considering only the 55 chemicals that affected zebrafish development, 49 of those chemicals passed the quality control. Interestingly, six of the chemicals that affected zebrafish development failed the quality control. This suggests that the toxicity may have been caused by an unknown substance or a mixture of the parent PFAS and its degradation products. Table 1 indicates the chemicals that failed the quality control labeled with an “F” and the ones that passed labeled with a “P”. In Figure 3, the chemicals that failed the chemical quality control are designated by one of three symbols, “⊗” or “x” or “Δ”; “⊗” shows the chemicals that failed the QC and did not result in developmental toxicity. “x” represents the chemicals that failed the quality control and were also likely to be volatile (vapor pressure exceeded 100 Tor [100 mm Hg]). The high volatility indicates that the chemical was more likely to be lost during the solubilization and/or testing process. The six chemicals that failed the quality control but were positive for eliciting developmental toxicity are designated with an open triangle (Δ) in Figure 3. These six chemicals were removed from further consideration in subsequent analyses. The chemicals that did not cause developmental toxicity are noted in Table 1 or Supplemental Table S3 as having a BMC of “1000”; chemicals positive for developmental toxicity have a numerical BMC listed with values equal to or less than 100 μM, the highest concentration tested.

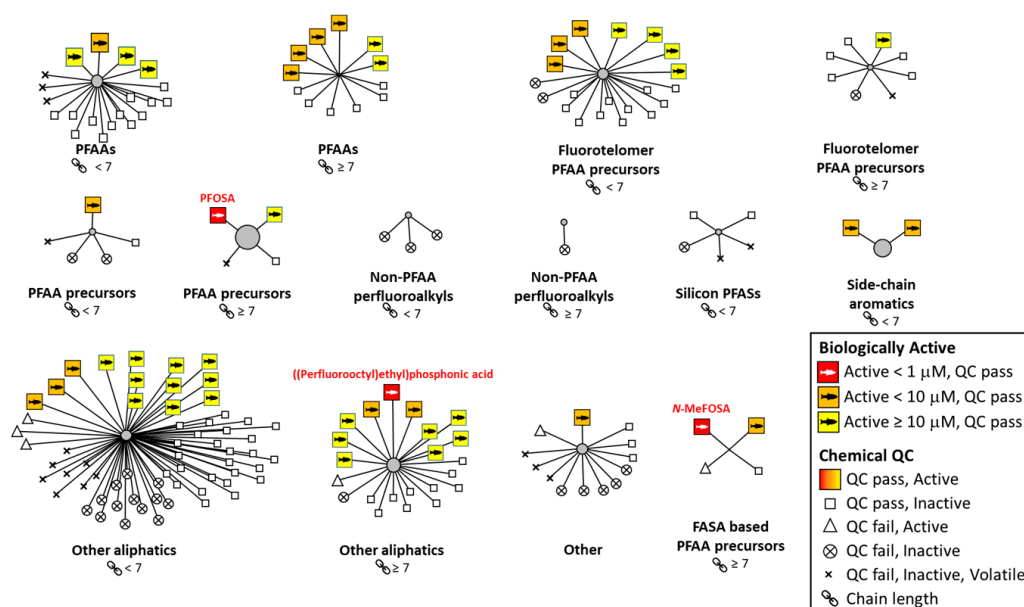


Figure 3. Visualization of toxicity, QC verdict, and structural class: This diagram summarizes and compares the biological activity and chemical quality control (QC) status as a function of OECD PFAS structural category [43]. Each group is one chemical category separated by chain length groups of those less than 7 (<7) or those greater than or equal to 7 (≥7). The lengths of the lines from the center of each radial plot were arbitrarily selected for visual spacing purposes and confer no additional information. Each point is one chemical representing the 182 unique ones tested. A large square is assigned to a chemical that was active in at least one endpoint, and its color indicates the potency of the most active endpoint. Each of the three most potent chemicals is indicated by a red chemical name above its respective red square. Small squares indicate an inactive chemical that passed the QC. Chemicals that failed the QC are broken down into three groups based on volatility and activity. PFAA = perfluoroalkyl acid, PFAS = perfluoroalkyl substance, FASA = fluoroalkyl sulfonamide.

Table 1. Chemical list and properties: Each of the unique 182 chemicals tested are listed one per row and include (from left to right) chemical name; an abbreviation or synonym, if appropriate; structural category; CAS number; DTXSID (which links directly to the CompTox dashboard; <https://comptox.epa.gov/dashboard/>; accessed on 8 February 2024); molecular weight; formula; chain length; quality control data (QC) verdict [36]; vapor pressure (mm Hg) OPERA model values from <https://comptox.epa.gov/dashboard/> (accessed on 16 March 2024); and benchmark concentration (BMC) in micromolar (μM). The table is sorted by BMC from the lowest value to the highest.

| Chemical Name | Abbreviation/ Synonym | Structural Category | CAS Number | DTXSID | Molecular Weight | Formula | Chain Length | QC | Vapor Pressure (mm Hg) | BMC (μM) |
|--|--------------------------|-----------------------------------|---------------|--------------------------------|---------------------|--------------|-----------------|----|------------------------------|--------------------------|
| Perfluorooctanesulfonamide | PFOSA | PFAA precursors gte7 | 754-91-6 | DTXSID3038939 | 499.14 | C8H2F17NO2S | 8 | P | 2.45×10^{-1} | 0.26 |
| <i>N</i> -Methylperfluorooctanesulfonamide | N-MeFOSA | FASA based PFAA precursors gte7 | 31506-32-8 | DTXSID1067629 | 513.17 | C9H4F17NO2S | 8 | P | 1.20×10^{-4} | 0.44 |
| ((Perfluorooctyl)ethyl)phosphonic acid | | Other aliphatics gte7 | 80220-63-9 | DTXSID30627108 | 528.10 | C10H6F17O3P | 8 | P | 1.27×10^{-4} | 0.58 |
| Perfluoro-3,6,9-trioxatridecanoic acid | | PFAAs lt7 | 330562-41-9 | DTXSID50375114 | 562.08 | C10HF19O5 | 4 | P | 6.61×10^{-5} | 1.30 |
| Perfluorohexanesulfonamide | FHxSA | PFAA precursors lt7 | 41997-13-1 | DTXSID50469320 | 399.13 | C6H2F13NO2S | 6 | P | 2.49×10^{-4} | 1.45 |
| 1H,1H,6H,6H-Perfluorohexane-1,6-diol diacrylate | | Other aliphatics lt7 | 2264-01-9 | DTXSID80379721 | 370.20 | C12H10F8O4 | 4 | P | 5.06×10^{-3} | 1.52 |
| 1,6-Diiodoperfluorohexane | | Other aliphatics lt7 | 375-80-4 | DTXSID90190949 | 553.86 | C6F12I2 | 6 | P | 2.91×10^{-1} | 1.85 |
| Perfluoropinacol | | other | 918-21-8 | DTXSID60238701 | 334.06 | C6H2F12O2 | 1 | P | 1.41×10^0 | 1.94 |
| <i>N</i> -Ethylperfluorooctane sulfonamide | NEtFOSA | FASA based PFAA precursors gte7 | 4151-50-2 | DTXSID1032646 | 527.20 | C10H6F17NO2S | 8 | P | 5.02×10^{-6} | 2.37 |
| Perfluoroundecanoic acid | PFUnDA | PFAAs gte7 | 2058-94-8 | DTXSID8047553 | 564.09 | C11HF21O2 | 10 | P | 6.48×10^{-4} | 2.74 |
| Potassium perfluorooctanesulfonate | PFOS-K | PFAAs gte7 | 2795-39-3 | DTXSID8037706 | 538.22 | C8F17KO3S | 8 | P | 2.48×10^{-6} | 2.77 |
| (Perfluorobutyryl)-2-thenoilmethane | | Side-chain aromatics lt7 | 559-94-4 | DTXSID7060332 | 322.20 | C10H5F7O2S | 3 | P | 4.23×10^{-2} | 3.11 |
| 2-(Perfluorohexyl)ethanol | 6:2 FTOH | Fluorotelomer PFAA precursors lt7 | 647-42-7 | DTXSID5044572 | 364.11 | C8H5F13O | 6 | P | 6.01×10^{-1} | 3.37 |
| 1H,1H,10H,10H-Perfluorodecane-1,10-diol | | Other aliphatics gte7 | 754-96-1 | DTXSID50369896 | 462.13 | C10H6F16O2 | 8 | P | 3.14×10^{-2} | 3.47 |
| 1-(Perfluorofluorooctyl)propane-2,3-diol | | Other aliphatics gte7 | 94159-84-9 | DTXSID80881157 | 494.15 | C11H7F17O2 | 8 | F | 1.27×10^{-2} | 4.42 |
| 9-Chloro-perfluorononanoic acid | Cl-PFNA | Other aliphatics gte7 | 865-79-2 | DTXSID30382104 | 480.53 | C9HClF16O2 | 8 | P | 1.17×10^{-2} | 6.38 |
| <i>N</i> -Methyl- <i>N</i> -(2-hydroxyethyl)perfluorooctanesulfonamide | N-MeFOSE | FASA based PFAA precursors gte7 | 24448-09-7 | DTXSID7027831 | 557.22 | C11H8F17NO3S | 8 | F | 1.01×10^{-5} | 7.06 |
| Perfluorooctanesulfonic acid | PFOS | PFAAs gte7 | 1763-23-1 | DTXSID3031864 | 500.13 | C8HF17O3S | 8 | P | 2.48×10^{-6} | 7.48 |

Table 1. Cont.

| Chemical Name | Abbreviation/ Synonym | Structural Category | CAS Number | DTXSID | Molecular Weight | Formula | Chain Length | QC | Vapor Pressure (mm Hg) | BMC (μM) |
|--|--------------------------|-----------------------------------|---------------|--------------------------------|---------------------|-----------------|-----------------|----|------------------------------|--------------------------|
| 1H,1H,5H,5H-Perfluoro-1,5-pentanediol diacrylate | | Other aliphatics lt7 | 678-95-5 | DTXSID5060986 | 320.19 | C11H10F6O4 | 3 | P | 1.67×10^{-2} | 7.62 |
| 2,2,3,3-Tetrafluoropropyl acrylate | | Other aliphatics lt7 | 7383-71-3 | DTXSID10224331 | 186.11 | C6H6F4O2 | 2 | F | 3.51×10^0 | 7.81 |
| 3-(Perfluorohexyl)propanoic acid | 6:3 FTCA | Fluorotelomer PFAA precursors lt7 | 27854-30-4 | DTXSID70379917 | 392.12 | C9H5F13O2 | 6 | P | 1.06×10^{-1} | 8.20 |
| 2H,2H,3H,3H-Perfluorooctanoic acid | 5:3 PFOA | Fluorotelomer PFAA precursors lt7 | 914637-49-3 | DTXSID20874028 | 342.11 | C8H5F11O2 | 5 | P | 4.39×10^{-1} | 8.77 |
| 1H,1H,7H-Perfluoroheptyl 4-methylbenzenesulfonate | | Side-chain aromatics lt7 | 424-16-8 | DTXSID30340244 | 486.27 | C14H10F12O3S | 6 | P | 1.95×10^{-4} | 8.97 |
| 3H-Perfluoro-2,2,4,4-tetrahydroxypentane | | other | 77953-71-0 | DTXSID70379295 | 262.08 | C5H5F7O4 | 1 | F | 3.89×10^{-8} | 9.29 |
| Perfluorodecanoic acid | PFDA | PFAAs gte7 | 335-76-2 | DTXSID3031860 | 514.09 | C10HF19O2 | 8 | P | 1.46×10^{-3} | 9.34 |
| 1H,1H,8H,8H-Perfluorooctane-1,8-diol | | Other aliphatics lt7 | 90177-96-1 | DTXSID30396867 | 362.12 | C8H6F12O2 | 6 | P | 1.18×10^{-1} | 10.37 |
| Perfluoro-1-octanesulfonyl chloride | | Other aliphatics gte7 | 423-60-9 | DTXSID90315130 | 518.57 | C8ClF17O2S | 8 | P | 1.75×10^1 | 10.42 |
| ((2,2,3,3-Tetrafluoropropoxy)methyl)oxirane | | Other aliphatics lt7 | 19932-26-4 | DTXSID70880230 | 188.12 | C6H8F4O2 | 2 | P | 3.13×10^0 | 10.44 |
| 1-Iodopentadecafluoroheptane | | PFAA precursors gte7 | 335-58-0 | DTXSID5059828 | 495.96 | C7F15I | 7 | P | 1.04×10^2 | 10.89 |
| 11-H-Perfluoroundecanoic acid | H-PFUnA | Other aliphatics gte7 | 1765-48-6 | DTXSID5061954 | 546.10 | C11H2F20O2 | 10 | P | 1.37×10^{-3} | 11.17 |
| Perfluorononanoyl chloride | | Other aliphatics gte7 | 52447-23-1 | DTXSID00379925 | 482.52 | C9ClF17O | 8 | P | 2.33×10^2 | 13.10 |
| Perfluoro-1,4-diiodobutane | | Other aliphatics lt7 | 375-50-8 | DTXSID30190948 | 453.84 | C4F8I2 | 4 | P | 3.32×10^1 | 14.80 |
| Perfluoroheptanesulfonic acid | PFHpS | PFAAs gte7 | 375-92-8 | DTXSID8059920 | 450.12 | C7HF15O3S | 7 | P | 3.33×10^{-7} | 15.55 |
| Perfluorooctanamide | | Other aliphatics gte7 | 307-31-3 | DTXSID70381151 | 412.10 | C8H3F15N2 | 7 | P | 3.12×10^{-1} | 19.45 |
| N-[(Perfluorooctylsulfonamido)propyl]-N,N,N-trimethylammonium iodide | | Other aliphatics gte7 | 1652-63-7 | DTXSID8051419 | 726.23 | C14H16F17IN2O2S | 8 | P | 1.17×10^{-4} | 19.75 |
| 2-(Perfluorobutyl)ethyl acrylate | | Fluorotelomer PFAA precursors lt7 | 52591-27-2 | DTXSID1068772 | 318.14 | C9H7F9O2 | 4 | P | 9.39×10^{-1} | 20.74 |
| 1H,1H-Perfluoro-3,6,9-trioxadecan-1-ol | | Other aliphatics lt7 | 147492-57-7 | DTXSID40380797 | 398.08 | C7H3F13O4 | 2 | F | 1.64×10^{-2} | 21.25 |
| 2-(Perfluorohexyl)ethylphosphonic acid | | Other aliphatics lt7 | 252237-40-4 | DTXSID20179883 | 428.09 | C8H6F13O3P | 6 | P | 3.40×10^{-6} | 21.90 |
| (Heptafluorobutanoyl)pivaloylmethane | | Other aliphatics lt7 | 17587-22-3 | DTXSID3066215 | 296.19 | C10H11F7O2 | 3 | P | 1.01×10^{-1} | 22.97 |
| Perfluorononanoic acid | PFNA | PFAAs gte7 | 375-95-1 | DTXSID8031863 | 464.08 | C9HF17O2 | 8 | P | 8.44×10^{-3} | 25.45 |

Table 1. Cont.

| Chemical Name | Abbreviation/ Synonym | Structural Category | CAS Number | DTXSID | Molecular Weight | Formula | Chain Length | QC | Vapor Pressure (mm Hg) | BMC (μM) |
|---|--------------------------|---------------------------------------|---------------|--------------------------------|---------------------|------------|-----------------|----|------------------------------|--------------------------|
| 4:4 Fluorotelomer alcohol | 4:4 FTOH | Other aliphatics lt7 | 3792-02-7 | DTXSID60377821 | 292.15 | C8H9F9O | 4 | P | 6.28×10^0 | 25.53 |
| 1H,1H-Perfluorononylamine | | Other aliphatics gte7 | 355-47-5 | DTXSID50379930 | 449.11 | C9H4F17N | 8 | P | 9.67×10^{-2} | 27.23 |
| Perfluoro(2-(2-propoxypropoxy)-1H,1H-propan-1-ol) | | Other aliphatics lt7 | 14548-74-4 | DTXSID80371164 | 482.09 | C9H3F17O3 | 3 | F | 1.43×10^{-3} | 27.37 |
| 1H,1H,5H-Perfluoropentyl methacrylate | | Other aliphatics lt7 | 355-93-1 | DTXSID90880131 | 300.15 | C9H8F8O2 | 4 | P | 4.39×10^{-1} | 27.89 |
| 1-Iodo-1H,1H,2H,2H-perfluoroheptane | | Fluorotelomer PFAA precursors lt7 | 1682-31-1 | DTXSID9061881 | 424.00 | C7H4F11I | 5 | P | 8.54×10^{-1} | 29.34 |
| 2,2,2-Trifluoroethyl perfluorobutanesulfonate | | Other aliphatics lt7 | 79963-95-4 | DTXSID60380390 | 382.12 | C6H2F12O3S | 4 | P | 3.01×10^1 | 29.36 |
| Perfluorohexanesulfonic acid | PFHxS | PFAAs lt7 | 355-46-4 | DTXSID7040150 | 400.11 | C6HF13O3S | 6 | P | 8.19×10^{-9} | 30.15 |
| 1H,1H,11H,11H-Perfluorotetraethylene glycol | | Other aliphatics lt7 | 330562-44-2 | DTXSID00380798 | 410.11 | C8H6F12O5 | 2 | P | 7.85×10^{-6} | 31.27 |
| Potassium perfluorohexanesulfonate | PFHS-K | PFAAs lt7 | 3871-99-6 | DTXSID3037709 | 438.20 | C6F13KO3S | 6 | P | 8.19×10^{-9} | 31.30 |
| 7:3 Fluorotelomer alcohol | 7:3 FTOH | Other aliphatics gte7 | 25600-66-2 | DTXSID50382621 | 428.14 | C10H7F15O | 7 | P | 3.00×10^{-1} | 38.32 |
| 6:1 Fluorotelomer alcohol | 6:1 FTOH | Fluorotelomer PFAA precursors lt7 | 375-82-6 | DTXSID00190950 | 350.08 | C7H3F13O | 6 | P | 5.05×10^{-1} | 73.35 |
| Perfluoro-3,6-dioxadecanoic acid | | PFAAs lt7 | 137780-69-9 | DTXSID50381073 | 446.07 | C8HF15O4 | 4 | P | 3.42×10^{-3} | 75.28 |
| 1H,1H-Perfluoroheptylamine | | Other aliphatics lt7 | 423-49-4 | DTXSID10379835 | 349.10 | C7H4F13N | 6 | P | 5.01×10^{-1} | 79.17 |
| 3:3 Fluorotelomer carboxylic acid | 3:3 FTCA | Fluorotelomer PFAA precursors lt7 | 356-02-5 | DTXSID00379268 | 242.09 | C6H5F7O2 | 3 | P | 5.19×10^{-1} | 87.79 |
| 2-(Perfluorooctyl)ethanol | 8:2 FTOH | Fluorotelomer PFAA precursors gte7 | 678-39-7 | DTXSID7029904 | 464.12 | C10H5F17O | 8 | P | 2.07×10^{-1} | 89.03 |
| (Heptafluoropropyl)trimethylsilane | | Silicon PFASs lt7 | 3834-42-2 | DTXSID70400078 | 242.21 | C6H9F7Si | 3 | F | 1.28×10^2 | 1000 |
| (Perfluoro-5-methylhexyl)ethyl 2-methylprop-2-enoate | | Fluorotelomer PFAA precursors lt7 | 50836-66-3 | DTXSID60379901 | 482.19 | C13H9F15O2 | 4 | P | 1.13×10^{-2} | 1000 |
| (Perfluorobutyl)ethene | | Fluorotelomer PFAA precursors lt7 | 19430-93-4 | DTXSID6047575 | 246.08 | C6H3F9 | 4 | F | 2.14×10^3 | 1000 |
| (Perfluoroheptyl)methyl methacrylate | | Other aliphatics gte7 | 3934-23-4 | DTXSID5063235 | 468.16 | C12H7F15O2 | 7 | P | 4.03×10^0 | 1000 |

Table 1. Cont.

| Chemical Name | Abbreviation/ Synonym | Structural Category | CAS Number | DTXSID | Molecular Weight | Formula | Chain Length | QC | Vapor Pressure (mm Hg) | BMC (μM) |
|--|--------------------------|---------------------------------------|---------------|--------------------------------|---------------------|------------|-----------------|----|------------------------------|--------------------------|
| (Perfluoropropyl)methyl methacrylate | | Other aliphatics lt7 | 13695-31-3 | DTXSID3065586 | 268.13 | C8H7F7O2 | 3 | P | 1.19×10^0 | 1000 |
| 1-(Perfluorohexyl)octane | | Fluorotelomer PFAA precursors lt7 | 133331-77-8 | DTXSID20440585 | 432.27 | C14H17F13 | 6 | P | 2.67×10^{-3} | 1000 |
| 1,1,1,3,3-Pentafluorobutane | | Other | 406-58-6 | DTXSID5073901 | 148.08 | C4H5F5 | 1 | F | 2.81×10^3 | 1000 |
| 1,1,1,5,5,5-Hexafluoro-2,4-pentanedione | | Other | 1522-22-1 | DTXSID4061753 | 208.06 | C5H2F6O2 | - | - | 2.81×10^2 | 1000 |
| 1,6-Dibromododecafluorohexane | | Other aliphatics lt7 | 918-22-9 | DTXSID20335129 | 459.86 | C6Br2F12 | 6 | P | 8.38×10^1 | 1000 |
| 11:1 Fluorotelomer alcohol | 11:1 FTOH | Fluorotelomer PFAA precursors gte7 | 423-65-4 | DTXSID80375107 | 600.12 | C12H3F23O | 11 | P | 4.16×10^{-3} | 1000 |
| 1-Bromopentadecafluoroheptane | | Other aliphatics gte7 | 375-88-2 | DTXSID9059919 | 448.96 | C7BrF15 | 7 | F | 2.93×10^2 | 1000 |
| 1H,1H,2H,2H-Perfluorohexyl iodide | | Fluorotelomer PFAA precursors lt7 | 2043-55-2 | DTXSID1047578 | 373.99 | C6H4F9I | 4 | P | 5.38×10^0 | 1000 |
| 1H,1H,2H-Perfluoro-1-decene | | Fluorotelomer PFAA precursors gte7 | 21652-58-4 | DTXSID7074616 | 446.11 | C10H3F17 | 8 | F | 4.07×10^2 | 1000 |
| 1H,1H,5H-Perfluoropentanol | | Other aliphatics lt7 | 355-80-6 | DTXSID0059879 | 232.07 | C5H4F8O | 4 | P | 2.85×10^1 | 1000 |
| 1H,1H,7H-Dodecafluoro-1-heptanol | | Other aliphatics lt7 | 335-99-9 | DTXSID9059832 | 332.09 | C7H4F12O | 6 | P | 6.18×10^{-1} | 1000 |
| 1H,1H,8H,8H-Perfluoro-3,6-dioxaoctane-1,8-diol | | Other aliphatics lt7 | 129301-42-4 | DTXSID70381090 | 294.10 | C6H6F8O4 | 2 | P | 1.72×10^{-4} | 1000 |
| 1H,1H,9H-Perfluorononyl acrylate | | Other aliphatics gte7 | 4180-26-1 | DTXSID00194615 | 486.15 | C12H6F16O2 | 8 | P | 7.06×10^{-1} | 1000 |
| 1H,1H-Heptafluorobutyl epoxide | | Other aliphatics lt7 | 1765-92-0 | DTXSID10379254 | 226.09 | C6H5F7O | 3 | P | 1.58×10^2 | 1000 |
| 1H,1H-Perfluorooctyl acrylate | | Other aliphatics gte7 | 307-98-2 | DTXSID5059799 | 454.14 | C11H5F15O2 | 7 | P | 6.31×10^0 | 1000 |
| 1H,1H-Perfluorooctylamine | | Other aliphatics gte7 | 307-29-9 | DTXSID50184723 | 399.10 | C8H4F15N | 7 | P | 1.06×10^{-1} | 1000 |
| 1H,1H-Perfluoropentylamine | | Other aliphatics lt7 | 355-27-1 | DTXSID60377826 | 249.08 | C5H4F9N | 4 | F | 4.22×10^1 | 1000 |
| 1H,2H-Hexafluorocyclopentene | | Other aliphatics lt7 | 1005-73-8 | DTXSID10461880 | 176.06 | C5H2F6 | 3 | F | 5.15×10^2 | 1000 |
| 1H-Perfluoro-1,1-propanediol | | Other aliphatics lt7 | 422-63-9 | DTXSID9059969 | 166.05 | C3H3F5O2 | 2 | F | 1.41×10^{-1} | 1000 |
| 1-Iodo-1H,1H,2H,2H-perfluorononane | | Fluorotelomer PFAA precursors gte7 | 2043-52-9 | DTXSID90880156 | 524.01 | C9H4F15I | 7 | P | 1.14×10^{-1} | 1000 |
| 1-Pentafluoroethylethanol | | Other aliphatics lt7 | 374-40-3 | DTXSID70880134 | 164.08 | C4H5F5O | 2 | F | 3.95×10^1 | 1000 |

Table 1. Cont.

| Chemical Name | Abbreviation/ Synonym | Structural Category | CAS Number | DTXSID | Molecular Weight | Formula | Chain Length | QC | Vapor Pressure (mm Hg) | BMC (μ M) |
|--|--------------------------|---------------------------------------|---------------|--------------------------------|---------------------|------------|-----------------|----|------------------------------|-------------------|
| 1-Propenylperfluoropropane | | Fluorotelomer PFAA precursors lt7 | 355-95-3 | DTXSID70379270 | 210.10 | C6H5F7 | 3 | F | 2.45×10^3 | 1000 |
| 2-(Perfluorohexyl)ethanethiol | 6:2 FtSH | Other aliphatics lt7 | 34451-26-8 | DTXSID20379947 | 380.17 | C8H5F13S | 6 | P | 2.01×10^0 | 1000 |
| 2-(Perfluorohexyl)ethyl methacrylate | 6:2 FTMAc | Fluorotelomer PFAA precursors lt7 | 2144-53-8 | DTXSID3047558 | 432.18 | C12H9F13O2 | 6 | P | 4.32×10^{-2} | 1000 |
| 2-(Perfluorooctyl)ethanethiol | | Other aliphatics gte7 | 34143-74-3 | DTXSID20337446 | 480.18 | C10H5F17S | 8 | P | 4.90×10^{-2} | 1000 |
| 2-(Perfluorooctyl)ethyl acrylate | 8:2 FTAc | Fluorotelomer PFAA precursors gte7 | 27905-45-9 | DTXSID5067348 | 518.17 | C13H7F17O2 | 8 | P | 1.53×10^{-1} | 1000 |
| 2-(Perfluorooctyl)ethyl methacrylate | 8:2 FTMAc | Fluorotelomer PFAA precursors gte7 | 1996-88-9 | DTXSID8062101 | 532.20 | C14H9F17O2 | 8 | P | 2.20×10^{-2} | 1000 |
| 2-(Trifluoromethoxy)ethyl trifluoromethanesulfonate | | other | 329710-76-1 | DTXSID00442840 | 262.12 | C4H4F6O4S | 1 | F | 4.18×10^{-1} | 1000 |
| 2,2-Difluoroethyl triflate | | other | 74427-22-8 | DTXSID30378880 | 214.11 | C3H3F5O3S | 1 | F | 1.94×10^1 | 1000 |
| 2-Amino-2H-perfluoropropane | | other | 1619-92-7 | DTXSID70481246 | 167.05 | C3H3F6N | 1 | F | 1.36×10^2 | 1000 |
| 2-Aminohexafluoropropan-2-ol | | other | 31253-34-6 | DTXSID80382093 | 183.05 | C3H3F6NO | 1 | F | 2.45×10^{-1} | 1000 |
| 2H-Perfluoroisopropyl 2-fluoroacrylate | | other | 74359-06-1 | DTXSID30622698 | 240.08 | C6H3F7O2 | 1 | F | 2.88×10^2 | 1000 |
| 3-(Perfluoro-2-butyl)propane-1,2-diol | | Other aliphatics lt7 | 125070-38-4 | DTXSID10382147 | 294.12 | C7H7F9O2 | 4 | F | 4.22×10^{-2} | 1000 |
| 3-(Perfluoro-3-methylbutyl)-1,2- propenoxide | | Other aliphatics lt7 | 54009-81-3 | DTXSID00379884 | 326.11 | C8H5F11O | 2 | P | 4.98×10^1 | 1000 |
| 3-(Perfluoroheptyl)propanoic acid | 7:3 FTCA | Fluorotelomer PFAA precursors gte7 | 812-70-4 | DTXSID90382620 | 442.12 | C10H5F15O2 | 7 | F | 5.47×10^{-3} | 1000 |
| 3-(Perfluorohexyl)-1,2-epoxypropane | | Other aliphatics lt7 | 38565-52-5 | DTXSID30880413 | 376.12 | C9H5F13O | 6 | F | 2.95×10^1 | 1000 |
| 3-(Perfluoroisopropyl)-2-propenoic acid | | Fluorotelomer PFAA precursors lt7 | 243139-64-2 | DTXSID40380257 | 240.08 | C6H3F7O2 | 1 | P | 2.68×10^0 | 1000 |
| 3-(Perfluorooctyl)propanol | 8:3 FTOH | Other aliphatics gte7 | 1651-41-8 | DTXSID10379991 | 478.15 | C11H7F17O | 8 | P | 7.39×10^{-2} | 1000 |
| 3-(Perfluoropropyl)propanol | | Other aliphatics lt7 | 679-02-7 | DTXSID60379269 | 228.11 | C6H7F7O | 3 | F | 1.70×10^1 | 1000 |
| 3,3-Bis(trifluoromethyl)-2-propenoic acid | | other | 1763-28-6 | DTXSID30170109 | 208.06 | C5H2F6O2 | 1 | P | 2.55×10^0 | 1000 |

Table 1. Cont.

| Chemical Name | Abbreviation/ Synonym | Structural Category | CAS Number | DTXSID | Molecular Weight | Formula | Chain Length | QC | Vapor Pressure (mm Hg) | BMC (μ M) |
|---|--------------------------|---------------------------------------|---------------|--------------------------------|---------------------|---------------|-----------------|----|------------------------------|-------------------|
| 3:1 Fluorotelomer alcohol | | Fluorotelomer PFAA precursors lt7 | 375-01-9 | DTXSID4059914 | 200.06 | C4H3F7O | 3 | P | 3.32×10^1 | 1000 |
| 3H,3H-Perfluoro-2,4-hexanedione | | Other aliphatics lt7 | 20825-07-4 | DTXSID90174941 | 258.07 | C6H2F8O2 | 2 | F | 1.25×10^3 | 1000 |
| 3-Methoxyperfluoro(2-methylpentane) | | Other aliphatics lt7 | 132182-92-4 | DTXSID20881338 | 350.08 | C7H3F13O | 2 | F | 3.56×10^2 | 1000 |
| 4,4-bis(Trifluoromethyl)-4-fluoropropanoic acid | | Fluorotelomer PFAA precursors lt7 | 243139-62-0 | DTXSID80380256 | 242.09 | C6H5F7O2 | 1 | P | 1.74×10^0 | 1000 |
| 4:2 Fluorotelomer alcohol | 4:2 FTOH | Fluorotelomer PFAA precursors lt7 | 2043-47-2 | DTXSID1062122 | 264.09 | C6H5F9O | 4 | P | 3.62×10^0 | 1000 |
| 4:2 Fluorotelomer sulfonic acid | 4:2 FTSA | Fluorotelomer PFAA precursors lt7 | 757124-72-4 | DTXSID30891564 | 328.15 | C6H5F9O3S | 4 | P | 1.32×10^{-6} | 1000 |
| 4H-Perfluorobutanoic acid | | Other aliphatics lt7 | 679-12-9 | DTXSID50892417 | 196.05 | C4H2F6O2 | 3 | P | 1.40×10^0 | 1000 |
| 5H-Perfluoropentanal | | Other aliphatics lt7 | 2648-47-7 | DTXSID20337466 | 230.06 | C5H2F8O | 4 | F | 1.04×10^3 | 1000 |
| 6:2 Fluorotelomer phosphate monoester | 6:2 monoPAP | Fluorotelomer PFAA precursors lt7 | 57678-01-0 | DTXSID90558000 | 444.09 | C8H6F13O4P | 6 | P | 1.32×10^{-6} | 1000 |
| 6:2 Fluorotelomer sulfonic acid | 6:2 FTSA | Fluorotelomer PFAA precursors lt7 | 27619-97-2 | DTXSID6067331 | 428.16 | C8H5F13O3S | 6 | P | 8.24×10^{-7} | 1000 |
| 6H-Perfluorohex-1-ene | | Other aliphatics lt7 | 1767-94-8 | DTXSID10379850 | 282.06 | C6HF11 | 4 | F | 3.57×10^2 | 1000 |
| 8:2 Fluorotelomer sulfonic acid | 8:2 FTS | Fluorotelomer PFAA precursors gte7 | 39108-34-4 | DTXSID00192353 | 528.18 | C10H5F17O3S | 8 | P | 1.00×10^{-5} | 1000 |
| 8H-Perfluorooctanoic acid | H-PFOA | Other aliphatics gte7 | 13973-14-3 | DTXSID70565479 | 396.08 | C8H2F14O2 | 7 | P | 5.35×10^{-2} | 1000 |
| 9H-Perfluorononanoic acid | H-PFNA | Other aliphatics gte7 | 76-21-1 | DTXSID50226894 | 446.09 | C9H2F16O2 | 8 | P | 2.62×10^{-2} | 1000 |
| Allyl perfluoroisopropyl ether | | Other aliphatics lt7 | 15242-17-8 | DTXSID10370988 | 226.09 | C6H5F7O | 1 | F | 2.73×10^2 | 1000 |
| Ammonium perfluorooctanoate | PFOAA | PFAAs gte7 | 3825-26-1 | DTXSID8037708 | 431.10 | C8H4F15NO2 | 7 | P | 1.11×10^{-1} | 1000 |
| Bis(1H,1H-perfluoropropyl)amine | | Other aliphatics lt7 | 883498-76-8 | DTXSID50381992 | 281.10 | C6H5F10N | 2 | P | 1.42×10^0 | 1000 |
| Dichloromethyl((perfluorohexyl)ethyl)silane | | Silicon PFASs lt7 | 73609-36-6 | DTXSID00223797 | 461.12 | C9H7Cl2F13Si | 6 | F | 1.13×10^{-1} | 1000 |
| Dimethoxymethyl((perfluorohexyl)ethyl)silan | | Silicon PFASs lt7 | 85857-17-6 | DTXSID40235137 | 452.29 | C11H13F13O2Si | 6 | P | 8.37×10^{-2} | 1000 |
| Ethyl pentafluoropropionyl acetate | | Other aliphatics lt7 | 663-35-4 | DTXSID20880144 | 234.12 | C7H7F5O3 | 2 | P | 1.08×10^0 | 1000 |
| Ethyl perfluorobutyl ether | | Other aliphatics lt7 | 163702-05-4 | DTXSID0073118 | 264.09 | C6H5F9O | 4 | F | 1.89×10^2 | 1000 |

Table 1. Cont.

| Chemical Name | Abbreviation/ Synonym | Structural Category | CAS Number | DTXSID | Molecular Weight | Formula | Chain Length | QC | Vapor Pressure (mm Hg) | BMC (μ M) |
|---|--------------------------|---------------------------------|---------------|--------------------------------|---------------------|---------------|-----------------|----|------------------------------|-------------------|
| Flurothyl | | other | 333-36-8 | DTXSID5046516 | 182.07 | C4H4F6O | 1 | F | 1.57×10^2 | 1000 |
| Heptafluorobutyl iodide | | Other aliphatics lt7 | 374-98-1 | DTXSID4059912 | 309.95 | C4H2F7I | 3 | P | 1.07×10^2 | 1000 |
| Heptafluorobutyramide | | Other aliphatics lt7 | 662-50-0 | DTXSID2060965 | 213.06 | C4H2F7NO | 3 | P | 2.15×10^{-1} | 1000 |
| Hexafluoroamylene glycol | | Other aliphatics lt7 | 376-90-9 | DTXSID3059927 | 212.09 | C5H6F6O2 | 3 | P | 1.25×10^{-1} | 1000 |
| Hexafluoroglutaryl chloride | | Other aliphatics lt7 | 678-77-3 | DTXSID0060985 | 276.94 | C5Cl2F6O2 | 3 | P | 3.05×10^2 | 1000 |
| Methyl 3H-perfluoroisopropyl ether | | Other aliphatics lt7 | 568550-25-4 | DTXSID70537191 | 182.07 | C4H4F6O | 1 | F | 4.08×10^2 | 1000 |
| Methyl perfluoro(3-(1-ethenyloxypropan-2-yloxy)propanoate) | EVE | Other aliphatics lt7 | 63863-43-4 | DTXSID8044969 | 422.10 | C9H3F13O4 | 2 | P | 1.12×10^{-1} | 1000 |
| Methyl perfluorohexanoate | | Other aliphatics lt7 | 424-18-0 | DTXSID20335700 | 328.08 | C7H3F11O2 | 5 | P | 6.20×10^1 | 1000 |
| <i>N-Ethyl-N</i> -(2-hydroxyethyl)perfluorooctane sulfonamide | N-EtFOSE | FASA based PFAA precursors gte7 | 1691-99-2 | DTXSID6027426 | 571.25 | C12H10F17NO3S | 8 | P | 8.78×10^{-4} | 1000 |
| <i>N-Methyl-N-trimethylsilylheptafluorobutyramide</i> | | Silicon PFAAs lt7 | 53296-64-3 | DTXSID40379666 | 299.26 | C8H12F7NOSi | 3 | P | 3.76×10^{-1} | 1000 |
| Nonafluoropentanamide | | Other aliphatics lt7 | 13485-61-5 | DTXSID60400587 | 263.06 | C5H2F9NO | 4 | P | 4.96×10^0 | 1000 |
| Octafluoroadipamide | | Other aliphatics lt7 | 355-66-8 | DTXSID80310730 | 288.10 | C6H4F8N2O2 | 4 | P | 5.99×10^{-8} | 1000 |
| Pentadecafluorooctanoyl chloride | | Other aliphatics gte7 | 335-64-8 | DTXSID40187142 | 432.51 | C8ClF15O | 7 | P | 1.00×10^2 | 1000 |
| Pentafluoropropanoic anhydride | | Other aliphatics lt7 | 356-42-3 | DTXSID70870515 | 310.05 | C6F10O3 | 2 | F | 1.24×10^2 | 1000 |
| Pentafluoropropionamide | | Other aliphatics lt7 | 354-76-7 | DTXSID0059871 | 163.05 | C3H2F5NO | 2 | P | 3.89×10^{-1} | 1000 |
| Perfluamine | FTPA | Other aliphatics lt7 | 338-83-0 | DTXSID9059834 | 521.07 | C9F21N | 3 | F | 1.94×10^2 | 1000 |
| Perfluoro-(2,5,8-trimethyl-3,6,9-trioxadodecanoic) acid | | PFAAs lt7 | 65294-16-8 | DTXSID70276659 | 662.10 | C12HF23O5 | 3 | F | 7.55×10^{-5} | 1000 |
| Perfluoro(4-methoxybutanoic acid) | PFMOBA | PFAAs lt7 | 863090-89-5 | DTXSID60500450 | 280.05 | C5HF9O3 | 3 | P | 1.75×10^0 | 1000 |
| Perfluoro(<i>N-methylmorpholine</i>) | | Other aliphatics lt7 | 382-28-5 | DTXSID7059933 | 299.04 | C5F11NO | 2 | F | 7.27×10^1 | 1000 |
| Perfluoro-1,3-dimethylcyclohexane | | Non-PFAA perfluoroalkyls lt7 | 335-27-3 | DTXSID0036926 | 400.06 | C8F16 | 3 | F | 3.60×10^2 | 1000 |
| Perfluoro-1-iodohexane | | PFAA precursors lt7 | 355-43-1 | DTXSID7047566 | 445.95 | C6F13I | 6 | P | 2.88×10^2 | 1000 |
| Perfluoro-2,5-dimethyl-3,6-dioxanonanoic acid | | PFAAs lt7 | 13252-14-7 | DTXSID00892442 | 496.08 | C9HF17O4 | 3 | F | 1.38×10^{-3} | 1000 |

Table 1. Cont.

| Chemical Name | Abbreviation/ Synonym | Structural Category | CAS Number | DTXSID | Molecular Weight | Formula | Chain Length | QC | Vapor Pressure (mm Hg) | BMC (μ M) |
|--|--------------------------|----------------------------------|---------------|--------------------------------|---------------------|-----------|-----------------|----|------------------------------|-------------------|
| Perfluoro-2-ethoxyethanesulfonic acid | PES | PFAAs lt7 | 113507-82-7 | DTXSID50379814 | 316.09 | C4HF9O4S | 2 | P | 1.25×10^{-6} | 1000 |
| Perfluoro-2-methyl-3-oxahexanoic acid | GenX | PFAAs lt7 | 13252-13-6 | DTXSID70880215 | 330.05 | C6HF11O3 | 3 | F | 2.41×10^{-1} | 1000 |
| Perfluoro-3-(1H-perfluoroethoxy)propane | Freon E1 | Other aliphatics lt7 | 3330-15-2 | DTXSID8052017 | 286.04 | C5HF11O | 3 | F | 3.61×10^2 | 1000 |
| Perfluoro-3,6,9-trioxadecanoic acid | | PFAAs lt7 | 151772-59-7 | DTXSID80380837 | 412.06 | C7HF13O5 | 2 | P | 3.20×10^{-4} | 1000 |
| Perfluoro-3,6-dioxaheptanoic acid | PFECA B | PFAAs lt7 | 151772-58-6 | DTXSID30382063 | 296.05 | C5HF9O4 | 2 | P | 6.90×10^{-4} | 1000 |
| Perfluoro-3,6-dioxaoctane-1,8-dioic acid | | Other aliphatics lt7 | 55621-21-1 | DTXSID20375106 | 322.06 | C6H2F8O6 | 2 | P | 6.78×10^{-5} | 1000 |
| Perfluoro-3-methoxypropanoic acid | PFMOPrA | PFAAs lt7 | 377-73-1 | DTXSID70191136 | 230.04 | C4HF7O3 | 2 | P | 6.93×10^{-2} | 1000 |
| Perfluoro-4-isopropoxybutanoic acid | PFECA G | PFAAs lt7 | 801212-59-9 | DTXSID60663110 | 380.06 | C7HF13O3 | 3 | P | 4.35×10^{-2} | 1000 |
| Perfluorobutanedioic acid | | Other aliphatics lt7 | 377-38-8 | DTXSID8059928 | 190.05 | C4H2F4O4 | 2 | P | 9.24×10^{-5} | 1000 |
| Perfluorobutanesulfonic acid | PFBS | PFAAs lt7 | 375-73-5 | DTXSID5030030 | 300.09 | C4HF9O3S | 4 | P | 1.14×10^{-8} | 1000 |
| Perfluorobutanesulfonyl fluoride | PFBS-F | PFAA precursors lt7 | 375-72-4 | DTXSID20861913 | 302.09 | C4F10O2S | 4 | F | 3.93×10^3 | 1000 |
| Perfluorobutanoic acid | PFBA | PFAAs lt7 | 375-22-4 | DTXSID4059916 | 214.04 | C4HF7O2 | 3 | P | 3.36×10^1 | 1000 |
| Perfluorobutyraldehyde | | PFAA precursors lt7 | 375-02-0 | DTXSID10190946 | 198.04 | C4HF7O | 3 | F | 2.61×10^3 | 1000 |
| Perfluorocyclohexanecarbonyl fluoride | | PFAA precursors lt7 | 6588-63-2 | DTXSID80379781 | 328.06 | C7F12O | 5 | F | 6.10×10^1 | 1000 |
| Perfluoroglutaryl difluoride | | Other aliphatics lt7 | 678-78-4 | DTXSID50218052 | 244.04 | C5F8O2 | 3 | F | 8.64×10^3 | 1000 |
| Perfluoroheptanoic acid | PFHpA | PFAAs lt7 | 375-85-9 | DTXSID1037303 | 364.06 | C7HF13O2 | 6 | P | 6.68×10^{-2} | 1000 |
| Perfluoroheptanoyl chloride | | Other aliphatics lt7 | 52447-22-0 | DTXSID80382154 | 382.51 | C7ClF13O | 6 | P | 6.71×10^1 | 1000 |
| Perfluorohexanedioic acid | | Other aliphatics lt7 | 336-08-3 | DTXSID4059833 | 290.07 | C6H2F8O4 | 4 | P | 7.28×10^{-5} | 1000 |
| Perfluorohexanoic acid | PFHxA | PFAAs lt7 | 307-24-4 | DTXSID3031862 | 314.05 | C6HF11O2 | 5 | P | 9.03×10^{-1} | 1000 |
| Perfluoromethylcyclopentane | PMCP | Non-PFAA perfluoroalkyls lt7 | 1805-22-7 | DTXSID7061982 | 300.05 | C6F12 | 4 | F | 2.58×10^2 | 1000 |
| Perfluorooct-1-ene | | Non-PFAA perfluoroalkyls lt7 | 559-14-8 | DTXSID40204489 | 400.06 | C8F16 | 6 | F | 2.98×10^2 | 1000 |
| Perfluorooctanamide | | Other aliphatics gte7 | 423-54-1 | DTXSID60195123 | 413.09 | C8H2F15NO | 7 | P | 1.14×10^{-1} | 1000 |
| Perfluorooctane | | Non-PFAA perfluoroalkyls gte7 | 307-34-6 | DTXSID0059794 | 438.06 | C8F18 | 8 | F | 9.34×10^3 | 1000 |
| Perfluorooctanesulfonyl fluoride | PFOS-F | PFAA precursors gte7 | 307-35-7 | DTXSID5027140 | 502.12 | C8F18O2S | 8 | F | 2.53×10^1 | 1000 |

Table 1. Cont.

| Chemical Name | Abbreviation/ Synonym | Structural Category | CAS Number | DTXSID | Molecular Weight | Formula | Chain Length | QC | Vapor Pressure (mm Hg) | BMC (μ M) |
|--|--------------------------|----------------------|---------------|--------------------------------|---------------------|---------------|-----------------|----|------------------------------|-------------------|
| Perfluorooctanoic acid | PFOA | PFAAs gte7 | 335-67-1 | DTXSID8031865 | 414.07 | C8HF15O2 | 7 | P | 1.11×10^{-1} | 1000 |
| Perfluorooctanoyl fluoride | PFOA-F | PFAA precursors gte7 | 335-66-0 | DTXSID0059829 | 416.06 | C8F16O | 7 | P | 3.26×10^2 | 1000 |
| Perfluoropentanamide | | Other aliphatics lt7 | 355-81-7 | DTXSID70366226 | 245.07 | C5H3F8NO | 4 | P | 5.22×10^{-2} | 1000 |
| Perfluoropentanedioic acid | | Other aliphatics lt7 | 376-73-8 | DTXSID8059926 | 240.06 | C5H2F6O4 | 3 | P | 8.32×10^{-5} | 1000 |
| Perfluoropentanoic acid | PFPeA | PFAAs lt7 | 2706-90-3 | DTXSID6062599 | 264.05 | C5HF9O2 | 4 | P | 6.62×10^0 | 1000 |
| Perfluoropropanoic acid | PPF | PFAAs lt7 | 422-64-0 | DTXSID8059970 | 164.03 | C3HF5O2 | 2 | P | 1.03×10^1 | 1000 |
| Perfluoropropyl trifluorovinyl ether | PPVE | Other aliphatics lt7 | 1623-05-8 | DTXSID0061826 | 266.04 | C5F10O | 3 | F | 2.13×10^2 | 1000 |
| Perfluorosuccinic anhydride | | Other aliphatics lt7 | 699-30-9 | DTXSID6061022 | 172.04 | C4F4O3 | 1 | F | 9.71×10^1 | 1000 |
| Perfluorotetradecanoic acid | PFTeDA | PFAAs gte7 | 376-06-7 | DTXSID3059921 | 714.12 | C14HF27O2 | 13 | P | 1.02×10^{-3} | 1000 |
| Perfluorotridecanoic acid | PFTriDA | PFAAs gte7 | 72629-94-8 | DTXSID90868151 | 664.11 | C13HF25O2 | 12 | P | 6.60×10^{-4} | 1000 |
| Potassium perfluorobutanesulfonate | KPFBS | PFAAs lt7 | 29420-49-3 | DTXSID3037707 | 338.18 | C4F9KO3S | 4 | P | 1.14×10^{-8} | 1000 |
| Potassium perfluorooctanoate | PFOA-K | PFAAs gte7 | 2395-00-8 | DTXSID00880026 | 452.16 | C8F15KO2 | 7 | P | 1.11×10^{-1} | 1000 |
| Sevoflurane | | other | 28523-86-6 | DTXSID8046614 | 200.06 | C4H3F7O | 1 | F | 1.94×10^2 | 1000 |
| Sodium perfluorooctanoate | PFOA-Na | PFAAs gte7 | 335-95-5 | DTXSID40880025 | 436.05 | C8F15NaO2 | 7 | P | 1.11×10^{-1} | 1000 |
| Triethoxy((perfluorohexyl)ethyl)silane | | Silicon PFASs lt7 | 51851-37-7 | DTXSID1074915 | 510.37 | C14H19F13O3Si | 6 | F | 1.23×10^{-1} | 1000 |
| tris(Trifluoroethoxy)methane | | other | 58244-27-2 | DTXSID30395037 | 310.12 | C7H7F9O3 | 1 | P | 4.55×10^{-1} | 1000 |

Figure 3 visualizes the toxicity of the 182 unique chemicals across OECD structural categories [43] to assess whether there are PFAS categories that were more or less likely to be developmentally toxic. The lowest BMC was used in this figure for each chemical; the specific endpoint that was used is noted in Supplemental Table S2 (Column K). The large colored squares with the cartoon of a fish inside represent the PFAS that produced developmental toxicity; the color relates to the potency (i.e., BMC; yellow is the least potent and red is the most potent). Almost any class with three or more chemicals that passed the chemical quality control contained at least one positive chemical; so, it is difficult to declare whether a particular chemical class is more or less likely to contain developmentally toxic chemicals. The three most potent chemicals were PFOSA (BMC = 0.26 μ M), N-MeFOSA (BMC = 0.44 μ M), and ((Perfluorooctyl)ethyl)phosphonic acid (BMC = 0.58 μ M), all of which had BMCs of less than 1 μ M and represent three different structural classifications: PFAA precursors, other aliphatics, and FASA-based PFAA precursors. There were 18 PFAS with BMCs less than 10 μ M representing nine different classes, and 28 PFAS with a BMC between 10 and 100 μ M representing seven different classes. The highest chemical concentration tested was 100 μ M; therefore, all BMC results for positive chemicals fall below that threshold (Table 1 and Figure 3). Interestingly, the sulfonamide structure seemed especially toxic to the developing zebrafish as five out of the six sulfonamide containing PFAS chemicals that also passed the QC produced developmental toxicity (Table 1). All five of these sulfonamides in the zebrafish were markedly toxic having a BMC below 20 μ M, whereas only one did not and was considered a negative. That negative sulfonamide (CAS number 1691-99-2; N-ethyl-N-(2-hydroxyethyl) perfluorooctanesulfonamide) has the same basic structure as the most potent sulfonamide (PFOSA; CAS number 754-91-6), except with more extensive functional groups attached to the sulfonamide moiety. Perhaps these extra ethyl and hydroxyethyl functional groups block the toxic action of the sulfonamide group in some manner or impair its uptake into the larva. The apparent toxicity of the sulfonamide structure was reinforced by a ToxPrint chemotype enrichment analysis [35,44,45], showing that three structures were significantly associated with developmental toxicity in the zebrafish embryos/larvae: sulfonamide, sulfonyl, or an 8-carbon chain length (the results are in column L of Supplemental Table S2). The sulfonamide structure is a subset of the sulfonyl structure, i.e., a chemical cannot be classified as a sulfonamide without also being classified as a sulfonyl, but the opposite is not necessarily true. The sulfonamide association does not account for all the sulfonyl-associated toxicity as there were sulfonyl containing PFAS that were not sulfonamides but were also active.

A commonly used general method to assess the influence of structure on the toxicity of PFAS chemicals is to compare the chain length of the PFAS versus its likelihood to cause toxicity. For the PFAS that passed the QC, a longer chain length was associated with a higher likelihood of developmental toxicity: if the chain length of the toxic PFAS is compared to the chain length of the non-toxic PFAS, the average chain length of the toxic PFAS was significantly longer than the chain length of the non-toxic PFAS ($p = 0.035$) (Figure 4A). There was, however, no correlation between the chain length and the increased potency of the chemical ($r^2 = 0.24$; a p -value for slope = 0.28; $n = 49$) (Figure 4B). When the degree of potency (BMC) is plotted against the chain length for each chemical that produced developmental toxicity, no relationship was found (Figure 4B); therefore, the potency of the chemical does not tend to change as the chain length increases.

The evidence of the lack of a significant relationship between OECD structural groups and the toxicity profile is also supported by the heat maps depicted in Figure 5. In this case, the potency of each positive endpoint for each chemical that passed the QC is indicated with the color representing the degree of potency. This was performed to discern any patterns of specific endpoints that might be associated with the different PFAS structural classifications. The left panel (A) shows the independent hierarchical clustering of the chemicals by the pattern of endpoint affected to evaluate if any of the chemical categories (listed by color in the lower right legend) are enriched in certain endpoints or endpoint patterns. As can be seen by the extremely varied representation of the PFAS category

colors in the far-left column, there does not appear to be any pattern of effects related to a particular PFAS structural category. This is further explained by the results shown in the righthand panel (Figure 5B) showing the pattern of effect by PFAS category, again visualizing that any one category (B, left column) does not present with a signature profile of endpoints.

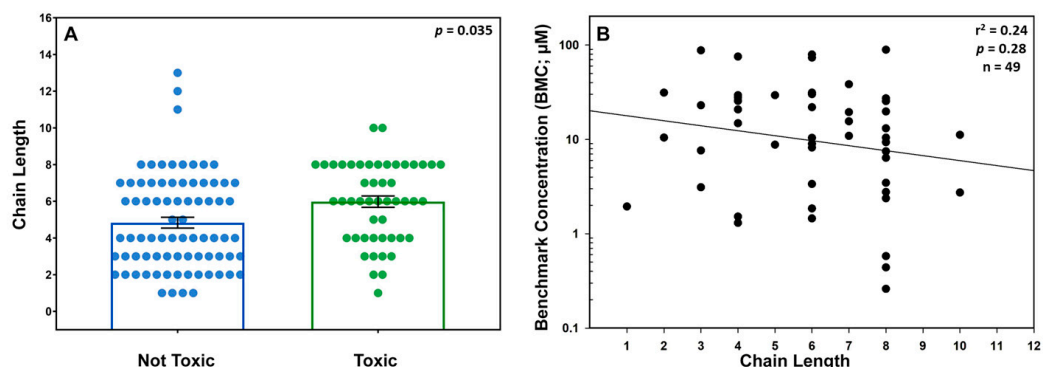


Figure 4. Comparison of toxicity and chain length: (A) Bar graph showing the relationship between the chain length and developmental toxicity of the chemical. Each dot represents one unique chemical. Only chemicals that passed the QC are included ($n = 49$ green dots for toxic [has a BMC] and $n = 78$ blue dots for non-toxic [no concentration relation or BMC] chemicals). A Mann–Whitney U non-parametric test showed that there was a significant difference ($p = 0.0035$) in average chain length between the two groups. (B) Linear regression showing that an increase in chain length for the positive chemicals was not associated with increased toxicity. $r^2 = 0.24$; a p -value for slope = 0.28; $n = 49$.

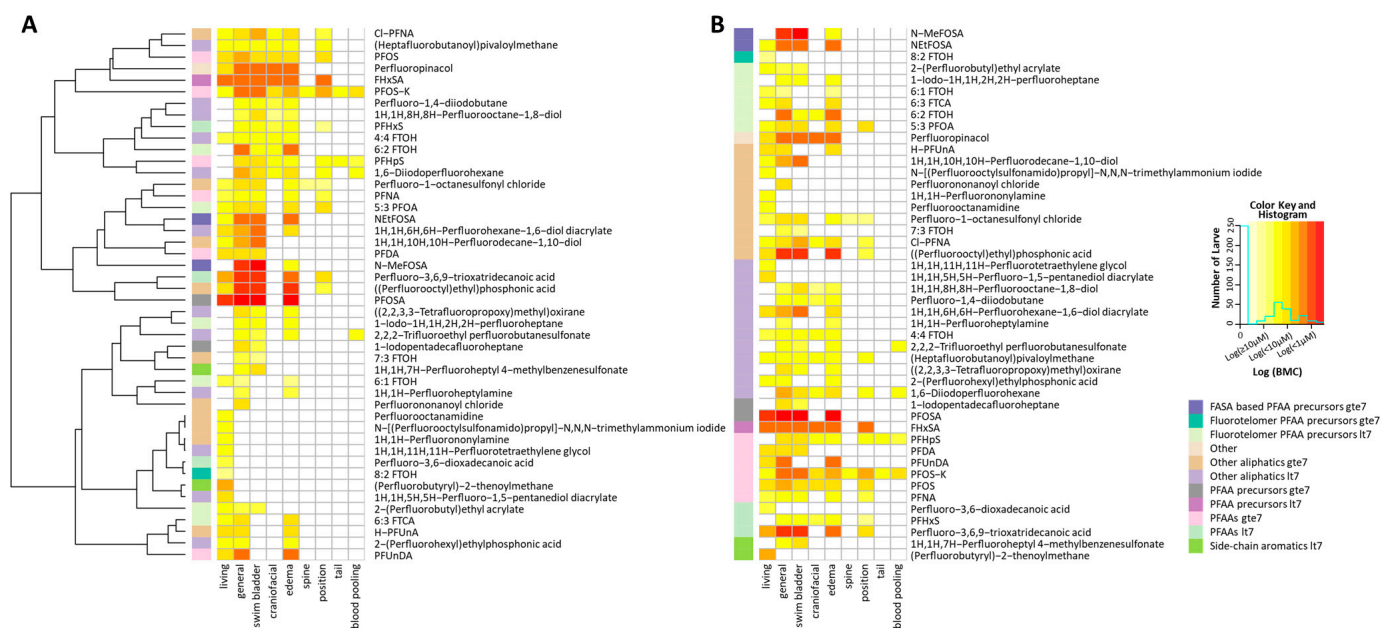


Figure 5. Heat maps showing the relationships among specific endpoints and structural class: Colors (upper right legend) indicate (BMC; μM), with deeper red colors indicating increased potency. Only chemicals that passed the analytical quality control (QC) and were active in at least one endpoint are shown. Left panel (A): Chemicals are sorted by unsupervised hierarchical clustering to discern if any of the chemical categories (listed by color in the lower right legend) are enriched in certain endpoints. Right panel (B): Endpoint enrichment is also explored with data sorted by chemical category.

4. Discussion

Although there did not appear to be any OECD structural groups that were more or less likely to contain chemicals positive for developmental toxicity, another type of analysis revealed that the sulfonamide containing PFAS were especially likely to be developmentally toxic to zebrafish and possibly other vertebrates. Like other reports [46–48], there was a general tendency for the longer chain compounds to be toxic, but that did not extend to a direct relationship between potency and chain length.

The QC testing of the chemical stocks was a definite advantage in this study as substances could volatilize during chemical handling, such as those with higher vapor pressure, which could be interpreted as false negatives for developmental toxicity. Given that a large percentage of the PFAS chemicals failed the QC, it seems irresponsible for future PFAS screens to proceed without analytical QC. Despite this library being selected for its structural diversity [35], in order to identify chemical grouping(s) with a higher rate of positives, there is still additional testing of the PFAS chemicals that will be needed to inform health and environmental hazard and enable accurate read-across comparisons for this class of compounds.

This study evaluated a large group of PFAS to determine developmental toxicity during early-life exposure. As part of the analysis, the results were compared to those from other PFAS library screening studies (Table 2). Notably, three screens using a zebrafish developmental model with similar assessment methods were reviewed [32–34], with results compared in Table 2. Differences in experimental design choices among laboratories, including rearing temperature, chorion status, dosing range, windows of exposure, and statistical methods for potency determination, made direct comparisons difficult among the studies. Even with those caveats, some common trends can be discerned in the results. There were fifteen chemicals that were tested in all four laboratories. Of those fifteen, fourteen passed the QC based on the testing associated with the present study. Of all chemicals, PFOSA was the most potent in every laboratory. PFOS (CAS number 754-91-6) was also tested in all four laboratories, but only tested positive for developmental toxicity in two out of the four laboratories, though this comparison is confounded by dosing range choices in each laboratory. In all four laboratories, PFOS was less potent than PFOSA. Although PFOSA is metabolized to PFOS in zebrafish [34] and other species [49], PFOSA has a markedly higher bioaccumulation factor than PFOS [34], which could increase the time \times tissue concentration of PFOSA and may help to explain why the parent chemical is more toxic than the metabolite. Even though PFOSA was so potent across multiple laboratory settings in larval zebrafish, no PFOSA developmental toxicity studies in mammals were identified. Using the Expanded PFAS Evidence Map Dashboard [50], only two studies of PFOSA in mammals were identified: both involved acute dosing and resulted in evidence of systemic effects in rats, but neither contained any assessment of developmental toxicity. Therefore, it may be important going forward to focus on more developmental toxicity testing among chemicals such as PFOSA. It should be noted that, in the present study as well as in the other studies listed in Table 2, the endpoints employed could best be classified as gross morphological changes. Normal-appearing animals do not necessarily signal the lack of developmental toxicity for the test chemical. Internal anatomy, physiology, and function may have been affected by the PFAS exposure without affecting gross morphology.

It has been widely reported that PFAS exposure causes changes in the thyroid axis in adults and developing animals. In a recently published meta-analysis of 13 human studies, a correlation between PFAS exposure and changes in maternal or newborn thyroid function was noted [51], while in an analysis of National Health and Nutrition Examination Survey (NHANES) data, an association between PFAS exposure and thyroid disorders was also revealed [52]. In a separate study of a smaller population of mothers and newborns, it was noted that the newer “replacement” PFAS disrupt newborn thyroid status as much as or more than the legacy PFAS, like PFOS [53]. Zebrafish possess a thyroid axis very similar to the mammalian thyroid axis [23,54–58] and are sensitive to the developmental effects of many confirmed thyroid-disrupting chemicals (reviewed in [24]). Likewise, as a

major regulatory organ, the disruption of thyroid axis development in zebrafish may cause malformations or alter physiology and behavior during development. Mirroring results in mammals, zebrafish exposed to PFAS show disrupted thyroid development and function: changes in thyroid hormone levels and/or of thyroid-related genes have been noted in zebrafish after exposure to PFAS [59–63]. Given this previous literature, the present study's morphological analysis concentrated on the known effects of thyroid hormone disruption, like decreased pigmentation and disrupted swim bladder inflation (reviewed in [64]). The results do not reveal any significant changes in pigmentation (Supplemental Table S1), but many of the chemicals (31 out of the 49; 63%) that were positive for developmental toxicity resulted in an uninflated swim bladder (Figure 5), with that effect being one of the most sensitive endpoints. The effects of PFAS chemicals on zebrafish swim bladder non-inflation have been reported by others [28,65]. As noted above, this effect of PFAS on swim bladder inflation could be due to an indirect effect through the perturbation of the thyroid axis, or it could possibly be due to the direct action of the PFAS on the surfactant lipid profile of the swim bladder, which is necessary for normal structure and function [66]. Failure to inflate could be related to problems with surfactant lipid function as it has been noted in a study of the effects of PFAS in human bronchial epithelial cells [67] or general membrane disruption [68]. While the present study did not assess the mechanistic underpinnings of the swim bladder non-inflation or possible thyroid disrupting effects, this could be a promising area for future work. To that end, the comparison of the present results on zebrafish development with a recently published *in vitro* thyroid screen [69] using many of the same chemicals tested in this paper revealed some possible insights into mechanism. One hundred percent of the PFAS that affected human or *Xenopus* iodotyrosine deiodinase (hIYD or xIYD) also affected development in the zebrafish embryos. Although zebrafish do possess IYD [70], the authors [69] of the *in vitro* screening study felt that the IYD enzymes were not a “sensitive” target of the tested PFAS. Perhaps zebrafish deiodinases are more sensitive to PFAS inhibition than the human or *Xenopus* counterparts, or the consequences of enzyme inhibition during development are more significant than anticipated.

In general, longer chain PFAS are more lipophilic, less water soluble, and tend to have longer half-lives, which is thought to contribute to their increased toxicity [46–48]. It was observed that whether a PFAS was developmentally toxic or not was related to chain length, but these data were quite variable. For the 49 chemicals assessed as positive for developmental toxicity, there was no direct relationship between potency and chain length, i.e., that the increase in chain length was not associated with an increase in toxicity. This lack of a clear trend between chain length and potency mirrors results from other PFAS zebrafish developmental toxicity studies [28] as well as other developmental *in vitro* studies [44]. It, however, contrasts with results of clinical chemistry endpoints in rodent subchronic oral toxicity studies using PFAS of various chain lengths [71], where the chain length was shown to be associated with the health outcomes for PFAS. At least in developing vertebrates, there may be more structural variables besides chain length contributing to the toxicity profile. We also found a lack of association between OECD structural categories and developmental toxicity potential, which was also observed in a study of developmental neurotoxicity *in vitro* assays [44]. The absence of structure-based bioactivity relationships may be explained by limitations in biologically informative groupings of PFAS structures and ongoing work towards reproducible PFAS structure-based chemical categories, which may further inform toxicity [35]. An alternate chemotyping analysis [35,44,45] showed that three substructures were associated with zebrafish developmental toxicity: sulfonyl, sulfonamide, and an 8-carbon chain length; the sulfonamide structure has also been highlighted by others exploring PFAS developmental toxicity [29]. In addition, there was a relationship between developmental toxicity in zebrafish and certain PFAS as has been noted previously in other zebrafish *in vivo* [32–34] and *in vitro* [44,72] PFAS screening studies.

Table 2. Comparison with other published PFAS zebrafish screens: Developmental toxicity endpoint values (i.e., mortality and morphology) were compared among screening studies, and the results are presented. Provided is the chemical name and a CAS number used commonly across all four laboratories. For “Britton et al. BMC”, a benchmark concentration (BMC) as defined by benchmark response (BMR) is reported herein. A lower BMC indicates higher potency. For “[32], Morph.BMD10”, a benchmark concentration (BMD10) defined as a 10% change relative to the background response is reported (data from Ref. [32] Supplemental Table S3, Column F, Morph.BMD10). For “[34], Percent Mortality”, a percentage of mortality calculated for chemicals tested only at the nominal concentration of 5 μ M is reported. (Data from Ref. [34]; Supplemental Table S4 Mortality (%).) For “[33], survival or nominal % 6–72 hpf”, a percentage of survival or percent normal at 6 dpf is presented; for simplicity, columns for percent survival and percent normal were combined, and if the values varied, the lowest was selected for use (Data from Ref. [33] Figure 1). Colors represent potency with red and bolded text being compounds with a BMC less than 1 μ M for Britton and Truong, and active compounds for Han and Dasgupta. Yellow represents those with a BMC greater than 1 and less than 10 μ M for Britton and Truong. Green represents compounds with a BMC greater than 10 μ M for Britton and Truong. Dark gray indicates inactive compounds. Light gray indicates compounds not tested in other screens.

| Chemical Name | CAS Number | Britton et al., 2024—Chemical QC | Britton et al., 2024—BMC (μ M) | [32]—Morph.BMD10 (μ M) | [34]—Percent Mortality | [33]—Percent Survival or Normal at 6–72 hpf |
|---|-------------|----------------------------------|-------------------------------------|-----------------------------|------------------------|---|
| Perfluorooctanesulfonamide | 754-91-6 | P | 0.26 | 2.88 | 100 | 0 |
| N-Methylperfluorooctanesulfonamide | 31506-32-8 | P | 0.44 | 28.98 | 34.4 | Not Tested |
| ((Perfluorooctyl)ethyl)phosphonic acid | 80220-63-9 | P | 0.58 | Inactive | Not Tested | Not Tested |
| Perfluoro-3,6,9-trioxatridecanoic acid | 330562-41-9 | P | 1.30 | 9.78 | Inactive | Inactive |
| Perfluorohexanesulfonamide | 41997-13-1 | P | 1.45 | 9.76 | Not Tested | Not Tested |
| 1H,1H,6H,6H-Perfluorohexane-1,6-diol diacrylate | 2264-01-9 | P | 1.52 | 12.07 | Not Tested | Not Tested |
| 1,6-Diiodoperfluorohexane | 375-80-4 | P | 1.85 | Not Tested | Not Tested | Not Tested |
| Perfluoropinacol | 918-21-8 | P | 1.94 | Not Tested | Not Tested | Not Tested |
| N-Ethylperfluorooctanesulfonamide | 4151-50-2 | P | 2.37 | 43.70 | Inactive | Not Tested |
| Perfluoroundecanoic acid | 2058-94-8 | P | 2.74 | 55.82 | Not Tested | Not Tested |
| Potassium perfluorooctanesulfonate | 2795-39-3 | P | 2.77 | 11.02 | Inactive | Not Tested |
| (Perfluorobutyl)-2-thenoylmethane | 559-94-4 | P | 3.11 | 54.32 | Not Tested | Not Tested |
| 6:2 Fluorotelomer alcohol | 647-42-7 | P | 3.37 | Inactive | Inactive | Not Tested |
| 1H,1H,10H,10H-Perfluorodecane-1,10-diol | 754-96-1 | P | 3.47 | 11.16 | Not Tested | Not Tested |
| 1-(Perfluorooctyl)propane-2,3-diol | 94159-84-9 | F | 4.42 | 18.86 | Not Tested | Not Tested |
| 9-Chloro-perfluorononanoic acid | 865-79-2 | P | 6.38 | 20.97 | Not Tested | Not Tested |
| N-Methyl-N-(2-hydroxyethyl)perfluorooctanesulfonamide | 24448-09-7 | F | 7.06 | 69.09 | Not Tested | Not Tested |
| Perfluorooctanesulfonic acid | 1763-23-1 | P | 7.48 | 15.50 | Inactive | Inactive |
| 1H,1H,5H,5H-Perfluoro-1,5-pentenediol diacrylate | 678-95-5 | P | 7.62 | 2.47 | Not Tested | Not Tested |
| 2,2,3,3-Tetrafluoropropyl acrylate | 7383-71-3 | F | 7.81 | 31.89 | Not Tested | Not Tested |
| 8:2 Fluorotelomer acrylate | 27854-30-4 | P | 8.20 | Not Tested | Not Tested | Inactive |
| 2H,2H,3H,3H-Perfluorooctanoic acid | 914637-49-3 | P | 8.77 | Inactive | Inactive | Inactive |

Table 2. Cont.

| Chemical Name | CAS Number | Britton et al., 2024—Chemical QC | Britton et al., 2024—BMC (μM) | [32]—Morph.BMD10 (μM) | [34]—Percent Mortality | [33]—Percent Survival or Normal at 6–72 hpf |
|---|-------------|----------------------------------|-------------------------------|-----------------------|------------------------|---|
| 1H,1H,7H-Perfluoroheptyl 4-methylbenzenesulfonate | 424-16-8 | P | 8.97 | Inactive | Not Tested | Not Tested |
| 3H-Perfluoro-2,2,4,4-tetrahydroxypentane | 77953-71-0 | F | 9.29 | 30.23 | Inactive | Not Tested |
| Perfluorodecanoic acid | 335-76-2 | P | 9.34 | 0.22 | Not Tested | Not Tested |
| 1H,1H,8H,8H-Perfluorooctane-1,8-diol | 90177-96-1 | P | 10.37 | Inactive | Not Tested | Not Tested |
| Perfluoro-1-octanesulfonyl chloride | 423-60-9 | P | 10.42 | Inactive | Not Tested | Not Tested |
| ((2,2,3,3-Tetrafluoropropoxy)methyl)oxirane | 19932-26-4 | P | 10.44 | Inactive | Not Tested | Not Tested |
| 1-Iodopentadecafluoroheptane | 335-58-0 | P | 10.89 | Inactive | Not Tested | Not Tested |
| 11-H-Perfluoroundecanoic acid | 1765-48-6 | P | 11.17 | 22.11 | Not Tested | Not Tested |
| 8:2 Fluorotelomer methacrylate | 52447-23-1 | P | 13.10 | Not Tested | Not Tested | Not Tested |
| 3-(Perfluorooctyl)propanol | 375-50-8 | P | 14.80 | Not Tested | Not Tested | Not Tested |
| Perfluoroheptanesulfonic acid | 375-92-8 | P | 15.55 | 35.16 | Not Tested | Not Tested |
| Perfluorooctanamide | 307-31-3 | P | 19.45 | Inactive | Not Tested | Not Tested |
| Perfluorooctanesulfonamido ammonium iodide | 1652-63-7 | P | 19.75 | 74.00 | Inactive | Not Tested |
| 2-(Perfluorobutyl)ethyl acrylate | 52591-27-2 | P | 20.74 | Inactive | Not Tested | Not Tested |
| 1H,1H-Perfluoro-3,6,9-trioxadecan-1-ol | 147492-57-7 | F | 21.25 | Inactive | Not Tested | Not Tested |
| 2-(Perfluorohexyl)ethylphosphonic acid | 252237-40-4 | P | 21.90 | Inactive | Not Tested | Not Tested |
| (Heptafluorobutanoyl)pivaloylmethane | 17587-22-3 | P | 22.97 | Inactive | Not Tested | Not Tested |
| Perfluorononanoic acid | 375-95-1 | P | 25.45 | Inactive | Inactive | Inactive |
| 4:4 Fluorotelomer alcohol | 3792-02-7 | P | 25.53 | Inactive | Inactive | Not Tested |
| Dimethoxymethyl((perfluorohexyl)ethyl)silane | 355-47-5 | P | 27.23 | Not Tested | Not Tested | Not Tested |
| Perfluoro(2-(2-propoxypropoxy)-1H,1H-propan-1-ol) | 14548-74-4 | F | 27.37 | Not Tested | Not Tested | Not Tested |
| 1-Iodo-1H,1H,2H,2H-perfluorononane | 355-93-1 | P | 27.89 | Not Tested | Not Tested | Not Tested |
| 1-Iodo-1H,1H,2H,2H-perfluoroheptane | 1682-31-1 | P | 29.34 | Inactive | Not Tested | Not Tested |
| 2,2,2-Trifluoroethyl perfluorobutanesulfonate | 79963-95-4 | P | 29.36 | Inactive | Not Tested | Not Tested |
| Perfluorohexanesulfonic acid | 355-46-4 | P | 30.15 | 65.28 | Not Tested | Not Tested |
| 5H-Perfluoropentanal | 330562-44-2 | P | 31.27 | 10.53 | Not Tested | Not Tested |
| Potassium perfluorohexanesulfonate | 3871-99-6 | P | 31.30 | 76.85 | Inactive | Inactive |
| 7:3 Fluorotelomer alcohol | 25600-66-2 | P | 38.32 | Inactive | Not Tested | Not Tested |
| 6:1 Fluorotelomer alcohol | 375-82-6 | P | 73.35 | Inactive | Not Tested | Not Tested |
| Perfluamine | 137780-69-9 | P | 75.28 | Not Tested | Not Tested | Inactive |
| 1H,1H-Heptafluorobutyl epoxide | 423-49-4 | P | 79.17 | Not Tested | Not Tested | Not Tested |
| 3:3 Fluorotelomer carboxylic acid | 356-02-5 | P | 87.79 | Inactive | Not Tested | Inactive |
| 1H,1H-Heptafluorobutanol | 678-39-7 | P | 89.03 | Inactive | Inactive | Not Tested |

Table 2. Cont.

| Chemical Name | CAS Number | Britton et al., 2024—Chemical QC | Britton et al., 2024—BMC (μM) | [32]—Morph.BMD10 (μM) | [34]—Percent Mortality | [33]—Percent Survival or Normal at 6–72 hpf |
|--|-------------|----------------------------------|-------------------------------|-----------------------|------------------------|---|
| Perfluorobutyraldehyde | 423-54-1 | P | Inactive | 41.48 | Not Tested | Not Tested |
| (Perfluoro-5-methylhexyl)ethyl 2-methylprop-2-enoate | 72629-94-8 | P | Inactive | 41.56 | Not Tested | Not Tested |
| Perfluoro-1,3-dimethylcyclohexane | 678-77-3 | P | Inactive | 50.87 | Not Tested | Not Tested |
| 3-(Perfluoroisopropyl)-2-propenoic acid | 39108-34-4 | P | Inactive | 55.68 | Not Tested | Inactive |
| Perfluoro-4-isopropoxybutanoic acid | 63863-43-4 | P | Inactive | 58.77 | Not Tested | Not Tested |
| 1-Propenylperfluoropropane | 57678-01-0 | P | Inactive | 72.20 | Not Tested | Not Tested |
| (Heptafluoropropyl)trimethylsilane | 307-35-7 | F | Inactive | 86.05 | Not Tested | Not Tested |
| Perfluorotridecanoic acid | 151772-58-6 | P | Inactive | Inactive | Inactive | Inactive |
| Perfluorobutanoic acid | 863090-89-5 | P | Inactive | Inactive | Inactive | Inactive |
| Octafluoroadipamide | 335-67-1 | P | Inactive | Inactive | Inactive | Inactive |
| Perfluorooctanoic acid | 55621-21-1 | P | Inactive | Inactive | Inactive | Inactive |
| Potassium perfluorobutanesulfonate | 13252-13-6 | F | Inactive | Inactive | Inactive | Inactive |
| Perfluoro-3,6-dioxaoctane-1,8-dioic acid | 307-24-4 | P | Inactive | Inactive | Inactive | Inactive |
| 1H,1H,2H,2H-Perfluorohexyl iodide | 375-22-4 | P | Inactive | Inactive | Inactive | Inactive |
| 8:2 Fluorotelomer alcohol | 29420-49-3 | P | Inactive | Inactive | Inactive | Inactive |
| Perfluoro(4-methoxybutanoic) acid | 757124-72-4 | P | Inactive | Inactive | Inactive | Inactive |
| Perfluorooct-1-ene | 375-85-9 | P | Inactive | Inactive | Not Tested | Inactive |
| 3,3-Bis(trifluoromethyl)-2-propenoic acid | 377-73-1 | P | Inactive | Inactive | Not Tested | Inactive |
| Hexafluoroglutaryl chloride | 13252-14-7 | F | Inactive | Inactive | Not Tested | Inactive |
| tris(Trifluoroethoxy)methane | 2706-90-3 | P | Inactive | Inactive | Not Tested | Inactive |
| Pentafluoropropanoic anhydride | 65294-16-8 | F | Inactive | Inactive | Not Tested | Inactive |
| Sevoflurane | 336-08-3 | P | Inactive | Inactive | Not Tested | Inactive |
| Perfluorocyclohexanecarbonyl fluoride | 422-64-0 | P | Inactive | Inactive | Not Tested | Inactive |
| Perfluorooctane | 377-38-8 | P | Inactive | Inactive | Not Tested | Inactive |
| Perfluoro-1-iodohexane | 355-81-7 | P | Inactive | Inactive | 17.8 | Not Tested |
| Perfluoro(N-methylmorpholine) | 31253-34-6 | F | Inactive | Inactive | Inactive | Not Tested |
| 3H,3H-Perfluoro-2,4-hexanedione | 355-66-8 | P | Inactive | Inactive | Inactive | Not Tested |
| Perfluorooctanamide | 374-40-3 | F | Inactive | Inactive | Inactive | Not Tested |
| Perfluoroglutaryl difluoride | 375-73-5 | P | Inactive | Inactive | Inactive | Not Tested |
| 1H,1H,5H-Perfluoropentanol | 243139-64-2 | P | Inactive | Inactive | Inactive | Not Tested |
| 3-(Perfluorohexyl)-1,2-epoxypropane | 129301-42-4 | P | Inactive | Inactive | Inactive | Not Tested |
| 2,2,3,3,4,4,4-Heptafluorobutyl methacrylate | 662-50-0 | P | Inactive | Inactive | Inactive | Not Tested |
| 3-(Perfluoro-2-butyl)propane-1,2-diol | 1763-28-6 | P | Inactive | Inactive | Inactive | Not Tested |

Table 2. Cont.

| Chemical Name | CAS Number | Britton et al., 2024—Chemical QC | Britton et al., 2024—BMC (μM) | [32]—Morph.BMD10 (μM) | [34]—Percent Mortality | [33]—Percent Survival or Normal at 6–72 hpf |
|--|-------------|----------------------------------|--|------------------------------------|------------------------|---|
| Flurothyl | 883498-76-8 | P | Inactive | Inactive | Inactive | Not Tested |
| 1,1,1,3,3-Pentafluorobutane | 74427-22-8 | F | Inactive | Inactive | Inactive | Not Tested |
| 4:2 Fluorotelomer sulfonic acid | 355-27-1 | F | Inactive | Inactive | Inactive | Not Tested |
| Perfluoropropanoic acid | 375-01-9 | P | Inactive | Inactive | Inactive | Not Tested |
| 8:2 Fluorotelomer sulfonic acid | 355-80-6 | P | Inactive | Inactive | Inactive | Not Tested |
| Tetrafluorosuccinic acid | 1691-99-2 | P | Inactive | Inactive | Inactive | Not Tested |
| 3-Perfluoroheptylpropanoic acid | 679-02-7 | F | Inactive | Inactive | Inactive | Not Tested |
| 1H,2H-Hexafluorocyclopentene | 13485-61-5 | P | Inactive | Inactive | Inactive | Not Tested |
| Perfluorotetradecanoic acid | 423-65-4 | P | Inactive | Inactive | Inactive | Not Tested |
| Perfluoro-3,6-dioxaheptanoic acid | 125070-38-4 | F | Inactive | Inactive | Inactive | Not Tested |
| Perfluoroheptanoic acid | 58244-27-2 | P | Inactive | Inactive | Inactive | Not Tested |
| Perfluoro-3-methoxypropanoic acid | 329710-76-1 | F | Inactive | Inactive | Inactive | Not Tested |
| Perfluoro-2-methyl-3-oxahexanoic acid | 335-99-9 | P | Inactive | Inactive | Inactive | Not Tested |
| Perfluoropentanoic acid | 2144-53-8 | P | Inactive | Inactive | Inactive | Not Tested |
| Octafluoroadipic acid | 2043-47-2 | P | Inactive | Inactive | Inactive | Not Tested |
| Perfluorohexanoic acid | 376-90-9 | P | Inactive | Inactive | Inactive | Not Tested |
| Perfluoro-(2,5,8-trimethyl-3,6,9-trioxadodecanoic)acid | 679-12-9 | P | Inactive | Inactive | Inactive | Not Tested |
| Perfluoro-2,5-dimethyl-3,6-dioxanonanoic acid | 3825-26-1 | P | Inactive | Inactive | Inactive | Not Tested |
| Allyl perfluoroisopropyl ether | 812-70-4 | F | Inactive | Inactive | Not Tested | Not Tested |
| N-Ethyl-N-(2-hydroxyethyl)perfluorooctanesulfonamide | 376-06-7 | P | Inactive | Inactive | Not Tested | Not Tested |
| 3-(Perfluoropropyl)propanol | 13695-31-3 | P | Inactive | Inactive | Not Tested | Not Tested |
| Bis(1H,1H-perfluoropropyl)amine | 21652-58-4 | F | Inactive | Inactive | Not Tested | Not Tested |
| Perfluoropentanamide | 38565-52-5 | F | Inactive | Inactive | Not Tested | Not Tested |
| 2-(Trifluoromethoxy)ethyl trifluoromethanesulfonate | 374-98-1 | P | Inactive | Inactive | Not Tested | Not Tested |
| 1H,1H,7H-Dodecafluoro-1-heptanol [Dodecafluoroheptanol] | 2043-55-2 | P | Inactive | Inactive | Not Tested | Not Tested |
| 6:2 Fluorotelomer methacrylate | 4180-26-1 | P | Inactive | Inactive | Not Tested | Not Tested |
| Perfluorooctanesulfonyl fluoride | 20825-07-4 | F | Inactive | Inactive | Not Tested | Not Tested |
| 1H,1H-Perfluorooctyl acrylate | 355-43-1 | P | Inactive | Inactive | Not Tested | Not Tested |
| 4:2 Fluorotelomer alcohol | 559-14-8 | F | Inactive | Inactive | Not Tested | Not Tested |
| 2-(Perfluorooctyl)ethanthiol | 27905-45-9 | P | Inactive | Inactive | Not Tested | Not Tested |
| Hexafluoroamylene glycol | 338-83-0 | F | Inactive | Inactive | Not Tested | Not Tested |
| 2-Amino-2H-perfluoropropane | 6588-63-2 | F | Inactive | Inactive | Not Tested | Not Tested |

Table 2. Cont.

| Chemical Name | CAS Number | Britton et al., 2024—Chemical QC | Britton et al., 2024—BMC (μM) | [32]—Morph.BMD10 (μM) | [34]—Percent Mortality | [33]—Percent Survival or Normal at 6–72 hpf |
|--|-------------|----------------------------------|-------------------------------|-----------------------|------------------------|---|
| Nonafluoropentanamide | 307-34-6 | F | Inactive | Inactive | Not Tested | Not Tested |
| Triethoxy((perfluorohexyl)ethyl)silane | 50836-66-3 | P | Inactive | Inactive | Not Tested | Not Tested |
| 1-Bromopentadecafluoroheptane | 801212-59-9 | P | Inactive | Inactive | Not Tested | Not Tested |
| 3-Methoxyperfluoro(2-methylpentane) | 3834-42-2 | F | Inactive | Inactive | Not Tested | Not Tested |
| 4H-Perfluorobutanoic acid | 307-98-2 | P | Inactive | Inactive | Not Tested | Not Tested |
| Ammonium perfluorooctanoate | 34143-74-3 | P | Inactive | Inactive | Not Tested | Not Tested |
| Perfluoro(propyl vinyl ether) [Heptafluoropropyltrifluorovinyl ether] | 51851-37-7 | F | Inactive | Inactive | Not Tested | Not Tested |
| 11:1 Fluorotelomer alcohol | 375-88-2 | F | Inactive | Inactive | Not Tested | Not Tested |
| 1H,1H,8H,8H-Perfluoro-3,6-dioxaoctane-1,8-diol | 27619-97-2 | P | Inactive | Inactive | Not Tested | Not Tested |
| Perfluorobutanesulfonic acid | 422-63-9 | F | Inactive | Inactive | Not Tested | Not Tested |
| 1-Pentafluoroethylethanol | 663-35-4 | P | Inactive | Inactive | Not Tested | Not Tested |
| Perfluoromethylcyclopentane | 3934-23-4 | P | Inactive | Inactive | Not Tested | Not Tested |
| 2,2-Difluoroethyl triflate | 335-95-5 | P | Inactive | Inactive | Not Tested | Not Tested |
| 6:2 Fluorotelomer sulfonic acid | 73609-36-6 | F | Inactive | Inactive | Not Tested | Not Tested |
| Methyl perfluoro(3-(1-ethenyloxypropan-2- yloxy)propanoate) | 1651-41-8 | P | Inactive | Inactive | Not Tested | Not Tested |
| Perfluorononanoyl chloride | 1765-92-0 | P | Inactive | Inactive | Not Tested | Not Tested |
| Perfluoro-1,4-diiodobutane | 354-76-7 | P | Inactive | Inactive | Not Tested | Not Tested |
| Perfluoroheptanoyl chloride | 133331-77-8 | P | Inactive | Inactive | Not Tested | Not Tested |
| 1,6-Dibromododecafluorohexane | 2395-00-8 | P | Inactive | Inactive | Not Tested | Not Tested |
| 1H,1H-Perfluoroheptylamine | 2043-52-9 | P | Inactive | Inactive | Not Tested | Not Tested |
| Perfluorooctanoyl fluoride | 85857-17-6 | P | Inactive | Inactive | Not Tested | Not Tested |
| Pentadecafluorooctanoyl chloride | 54009-81-3 | P | Inactive | Inactive | Not Tested | Not Tested |
| 8H-Perfluorooctanoic acid | 376-73-8 | P | Inactive | Inactive | Not Tested | Not Tested |
| 1H,1H,5H-Perfluoropentyl methacrylate | 568550-25-4 | F | Inactive | Inactive | Not Tested | Not Tested |
| 1H,1H-Perfluorononylamine | 53296-64-3 | P | Inactive | Inactive | Not Tested | Not Tested |
| Ethyl perfluorobutyl ether | 151772-59-7 | P | Inactive | Not Tested | Not Tested | Inactive |
| Perfluoro-3-(1H-perfluoroethoxy)propane | 678-78-4 | F | Inactive | Not Tested | Inactive | Not Tested |
| 1H,1H,9H-Perfluorononyl acrylate | 356-42-3 | F | Inactive | Not Tested | Inactive | Not Tested |
| Perfluorosuccinic anhydride | 19430-93-4 | F | Inactive | Not Tested | Inactive | Not Tested |
| Perfluorobutanesulfonyl fluoride | 424-18-0 | P | Inactive | Not Tested | Inactive | Not Tested |
| 3,3,4,4,5,5,6,6,6-Nonafluorohexene [1H,1H,2H-Perfluoro1-hexene] | 406-58-6 | F | Inactive | Not Tested | Inactive | Not Tested |

Table 2. Cont.

| Chemical Name | CAS Number | Britton et al., 2024—Chemical QC | Britton et al., 2024—BMC (μM) | [32]—Morph.BMD10 (μM) | [34]—Percent Mortality | [33]—Percent Survival or Normal at 6–72 hpf |
|---|-------------|----------------------------------|-------------------------------|-----------------------|------------------------|---|
| Heptafluorobutyl iodide | 2648-47-7 | F | Inactive | Not Tested | Inactive | Not Tested |
| 2-Aminohexafluoropropan-2-ol | 355-95-3 | F | Inactive | Not Tested | Inactive | Not Tested |
| Methyl perfluorohexanoate | 15242-17-8 | F | Inactive | Not Tested | Inactive | Not Tested |
| Heptafluorobutyramide | 1619-92-7 | F | Inactive | Not Tested | Inactive | Not Tested |
| 1,1,1,5,5,5-Hexafluoroacetylacetone | 375-72-4 | F | Inactive | Not Tested | Inactive | Not Tested |
| 1H,1H,2H-Perfluoro-1-decene | 1623-05-8 | F | Inactive | Not Tested | Inactive | Not Tested |
| 1H,1H-Perfluoropentylamine | 333-36-8 | F | Inactive | Not Tested | Inactive | Not Tested |
| 6H-Perfluorohex-1-ene | 28523-86-6 | F | Inactive | Not Tested | Inactive | Not Tested |
| Perfluoro-3,6,9-trioxadecanoic acid | 1767-94-8 | F | Inactive | Not Tested | Inactive | Not Tested |
| 4,4,5,5,6,6,7,7,8,8,9,9,9-Tridecafluorononanoic acid | 163702-05-4 | F | Inactive | Not Tested | Inactive | Not Tested |
| Perfluoro-3,6-dioxadecanoic acid | 375-02-0 | F | Inactive | Not Tested | Inactive | Not Tested |
| 1H,1H-Perfluorooctylamine | 307-29-9 | P | Inactive | Not Tested | Not Tested | Not Tested |
| 9-H-Perfluorononanoic acid | 76-21-1 | P | Inactive | Not Tested | Not Tested | Not Tested |
| 2-(Perfluorohexyl)ethanethiol | 1005-73-8 | F | Inactive | Not Tested | Not Tested | Not Tested |
| 4,4-bis(Trifluoromethyl)-4-fluoropropanoic acid | 1522-22-1 | - | Inactive | Not Tested | Not Tested | Not Tested |
| 1H-Perfluoro-1,1-propanediol | 699-30-9 | F | Inactive | Not Tested | Not Tested | Not Tested |
| Perfluoro(2-ethoxyethane)sulfonic acid | 3330-15-2 | F | Inactive | Not Tested | Not Tested | Not Tested |
| Ethyl pentafluoropropionyl acetate | 382-28-5 | F | Inactive | Not Tested | Not Tested | Not Tested |
| 2,2,3,3,4,4,5,5,6,6,7,7,8,8,8-Pentadecafluorooctyl methacrylate | 335-27-3 | F | Inactive | Not Tested | Not Tested | Not Tested |
| Sodium perfluorooctanoate | 132182-92-4 | F | Inactive | Not Tested | Not Tested | Not Tested |
| Dichloromethyl((perfluorohexyl)ethyl)silane | 1805-22-7 | F | Inactive | Not Tested | Not Tested | Not Tested |
| 6:2 Fluorotelomer phosphate monoester | 52447-22-0 | P | Inactive | Not Tested | Not Tested | Not Tested |
| 1H,1H,11H,11H-Perfluorotetraethylene glycol | 918-22-9 | P | Inactive | Not Tested | Not Tested | Not Tested |
| Pentafluoropropionamide | 335-66-0 | P | Inactive | Not Tested | Not Tested | Not Tested |
| 1-(Perfluorohexyl)octane | 335-64-8 | P | Inactive | Not Tested | Not Tested | Not Tested |
| Potassium perfluorooctanoate | 13973-14-3 | P | Inactive | Not Tested | Not Tested | Not Tested |
| 3-(Perfluoro-3-methylbutyl)-1,2-propenoxide | 34451-26-8 | P | Inactive | Not Tested | Not Tested | Not Tested |
| Hexafluoroglutaric acid | 243139-62-0 | P | Inactive | Not Tested | Not Tested | Not Tested |
| Methyl 3H-perfluoroisopropyl ether | 113507-82-7 | P | Inactive | Not Tested | Not Tested | Not Tested |
| N-Methyl-N-trimethylsilylheptafluorobutyramide | 1996-88-9 | P | Inactive | Not Tested | Not Tested | Not Tested |
| 2H-Perfluoroisopropyl 2-fluoroacrylate | 74359-06-1 | F | Inactive | Not Tested | Not Tested | Not Tested |

This study of developmental toxicity in larval zebrafish using a diverse structural library of PFAS has illustrated the usefulness of this type of medium-throughput screening for potency and possible effects related to chemical structure. Despite no clear pattern regarding structural features related to toxic or non-toxic PFAS using the OECD structural groupings of the 182 chemicals, other structural identifiers were noted. Finally, leveraging the analytical QC results for the PFAS chemicals proved useful and should be encouraged in future studies, not only to eliminate the possibility for false negatives or positives, but also to promote the standardization of methodologies across laboratories.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/toxics12070501/s1>. Figure S1: BMC curves. Table S1: Fish assessments. Table S2: Data to make visualization of toxicity (Figure 3). Table S3: Data to make BMC curves.

Author Contributions: Conceptualization, K.N.B., S.P. and K.A.J.; methodology, K.N.B., R.S.J., K.A.J. and S.P.; software, R.S.J., M.F. and J.B.; validation, K.N.B. and J.K.O.; formal analysis, K.N.B. and R.S.J.; investigation, K.N.B., R.S.J., B.N.H., K.A.J., B.R.K. and S.P.; resources, J.K.O. and S.P.; data curation, K.N.B., R.S.J., K.A.J., M.L. and M.F.; writing—original draft preparation, K.N.B., B.N.H. and S.P.; writing—review and editing, K.N.B., R.S.J., B.N.H., K.A.J., J.K.O., B.R.K., M.F. and S.P.; visualization, K.N.B., R.S.J., K.A.J. and J.K.O.; supervision, K.N.B. and S.P.; project administration, K.N.B., K.A.J. and S.P. All authors have read and agreed to the published version of the manuscript.

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