

Article

Seroprevalence and Risk Factors Associated with Canine Leishmaniasis in Egypt

Abdelfattah Selim ^{1,*}, Salma Shoulah ¹, Abdelhamed Abdelhady ², Abdulaziz Alouffi ^{3,4}, Yasser Alraey ⁵ and Waleed S. Al-Salem ^{6,7}

- ¹ Department of Animal Medicine (Infectious Diseases), College of Veterinary Medicine, Benha University, Toukh 13736, Egypt; salma.soulah@fvmtm.bu.edu.eg
- ² Department of Parasitology and Animal Diseases, National Research Center, Giza 8655, Egypt; afanrc@yahoo.com
- ³ King Abdulaziz City for Science and Technology, Riyadh 12354, Saudi Arabia; asn1950r@gmail.com
- ⁴ Chair Vaccines Research of Infectious Diseases, King Saud University, Riyadh 11495, Saudi Arabia
- ⁵ Department of Clinical Laboratory Sciences, College of Applied Medical Sciences, King Khalid University, Abha 62529, Saudi Arabia; yahamd@kku.edu.sa
- ⁶ Department of Parasitology, Liverpool School of Tropical Medicine, Liverpool L3 5QA, UK; Waleed-alsalem@hotmail.com
- ⁷ Minister of Environment, Water and Agriculture, Riyadh 11195, Saudi Arabia
- * Correspondence: Abdelfattah.selim@fvmtm.bu.edu.eg

Abstract: Background: Canine leishmaniasis (CanL) is caused by *Leishmania infantum* (*L. infantum*) that is transmitted by sand fly vectors with dogs acting as the main reservoir. Methods: The present study aimed to determine the seroprevalence of CanL in dogs from Egypt and assessed the associated risk factors. The study was conducted from 2019 to 2020 in five governorates situated in Northern Egypt. Serum samples from 450 asymptomatic dogs were serologically examined by use of enzyme-linked immunosorbent assay (ELISA). Results: Overall, the seroprevalence rate of CanL was 21.3% and the highest rates were observed in Cairo and Giza governorates. The univariable analysis revealed that the seropositivity of CanL was strongly related to the dogs' ages, length of hair, absence of veterinary care or application of insecticides, and the type of floor of their shelters. The risk factors that were found to be associated with CanL in exposed dogs were: age group 2–4 years old (OR = 12, 95% CI: 1.6–92.3); short hair (OR = 2.07, 95% CI: 1.2–3.6); absence of veterinary care (OR = 2.7, 95% CI: 1.3–5.8); no application of insecticides (OR = 3.09, 95% CI: 1.5–6.5) and their residence in a shelter with an earthen floor (OR = 1.42, 95% CI: 0.7–2.9). Conclusions: Based on the present results, CanL is present in Egyptian dogs and this increases the possibility of transmission by sand fly to humans with whom they have contact. Consequently, an efficient monitoring programme and effective control measures are important to reduce the risk of infection.

Keywords: *L. infantum*; dogs; ELISA; Egypt



Citation: Selim, A.; Shoulah, S.; Abdelhady, A.; Alouffi, A.; Alraey, Y.; Al-Salem, W.S. Seroprevalence and Risk Factors Associated with Canine Leishmaniasis in Egypt. *Vet. Sci.* **2021**, *8*, 236. <https://doi.org/10.3390/vetsci8100236>

Academic Editor: Stefania Hanau

Received: 21 September 2021

Accepted: 13 October 2021

Published: 15 October 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Canine leishmaniasis (CanL) is a neglected zoonotic disease that is caused by *Leishmania* spp. [1]. Leishmaniasis is a vector-borne disease that is caused by protozoan flagellates. It affects several animal species including humans and is transmitted via bites of the infected female *Phlebotomus* sand fly [2,3].

The disease is endemic in tropical and temperate regions in 98 countries, and visceral leishmaniasis (VL) is the most fatal and common form in the Mediterranean region [4,5]. Zoonotic leishmaniasis caused by *Leishmania infantum* (*L. infantum*) is endemic in most of the Middle East, North Africa and the Mediterranean, including Egypt [6].

Domestic dogs are the main reservoir for leishmaniasis and can become infected at any age [7]. The clinical features of CanL vary from asymptomatic through self-limiting to severe viscerocutaneous infection [8,9]. However, asymptomatic dogs can develop clinical

signs during their lives, while *L. infantum* can spread unnoticed within the dog population and remain as a source of infection for sand flies, which transmit the parasite to other hosts [10]. Hence, early diagnosis of these asymptomatic carriers is critical for disease control in both endemic and non-endemic countries [11].

Leishmaniasis was reported in Egypt over 4000 years ago, in ancient Egyptian mummies. In past decades, *L. major* has been detected in dogs from Egypt [12] and antibodies against *Leishmania* spp. were detected in dogs from Egypt by Morsy, et al. [13]. In addition, there have been several reports from the mid to late 20th century of sporadic cases of cutaneous leishmaniasis (CL) and VL among Egyptian people, particularly those in the Suez Canal, Sinai and Agamy regions in Alexandria [4,14–18]. Recently, the presence of *L. tropica* in humans and *L. infantum* in dogs has been proven [1].

Serological examination is a useful tool to detect specific antibodies and to determine the spread of the disease, because a large proportion of dogs are asymptomatic [19]. Enzyme-linked immunosorbent assay (ELISA) and indirect fluorescent antibody tests (IFAT) are the most widely used serological methods [20]. ELISA shows potential as a sensitive tool for mass screening in epidemiological studies and is suited to field conditions [21,22].

In Egypt, the prevalence of *L. infantum* in dogs is uncertain, especially with changes of climatic conditions, dog populations and distributions of sand flies, all of which affect the epidemiology of the disease.

To fill this knowledge gap, the present study aimed to determine the seroprevalence of CanL and the risk factors that were associated with infection among dogs in some governorates in Northern Egypt.

2. Materials and Methods

2.1. Ethics Statement

All procedures involving the handling and collection of samples from dogs used in this study were approved by the ethical committee for Animal Experiment of Benha University and informed consent was obtained from owners. All methods regarding animals and human participant in the study were performed in accordance with the relevant guidelines and regulations and were approved by ethical committee of faculty of veterinary medicine, Benha University.

2.2. Study Area

The study was conducted during the period from June 2019 to May 2020 in five governorates located in Northern Egypt. These governorates were Cairo (30.0444° N, 31.2357° E), Giza (30.0131° N, 31.2089° E), Qalyubia (30.3292° N, 31.2168° E), Kafr ElSheikh (31.1107° N, 30.9388° E) and Gharbia (30.8754° N, 31.0335° E), Figure 1. The climate of the selected areas is Mediterranean with dry, hot summers and wet winters.

2.3. Sampling and Data Collection

The required samples size was determined using Danial's formula as follows:

$$n = \frac{z^2 p(1 - P)}{d^2} \quad (1)$$

where n is number of appropriate sample, z is level of confidence (z value is 1.96 if 95% confidence level is conventional), P is prevalence level which was 10% based on previously study of [12] and e is precision whereas this study's precision (e) was 5% based on [23]. According to dog population in each examined area and based on Danial's formula, the estimated sample size from each area was 95, 115, 75, 80 and 85 from Cairo, Giza, Qalyubia, Kafr ElSheikh and Gharbia governorates.

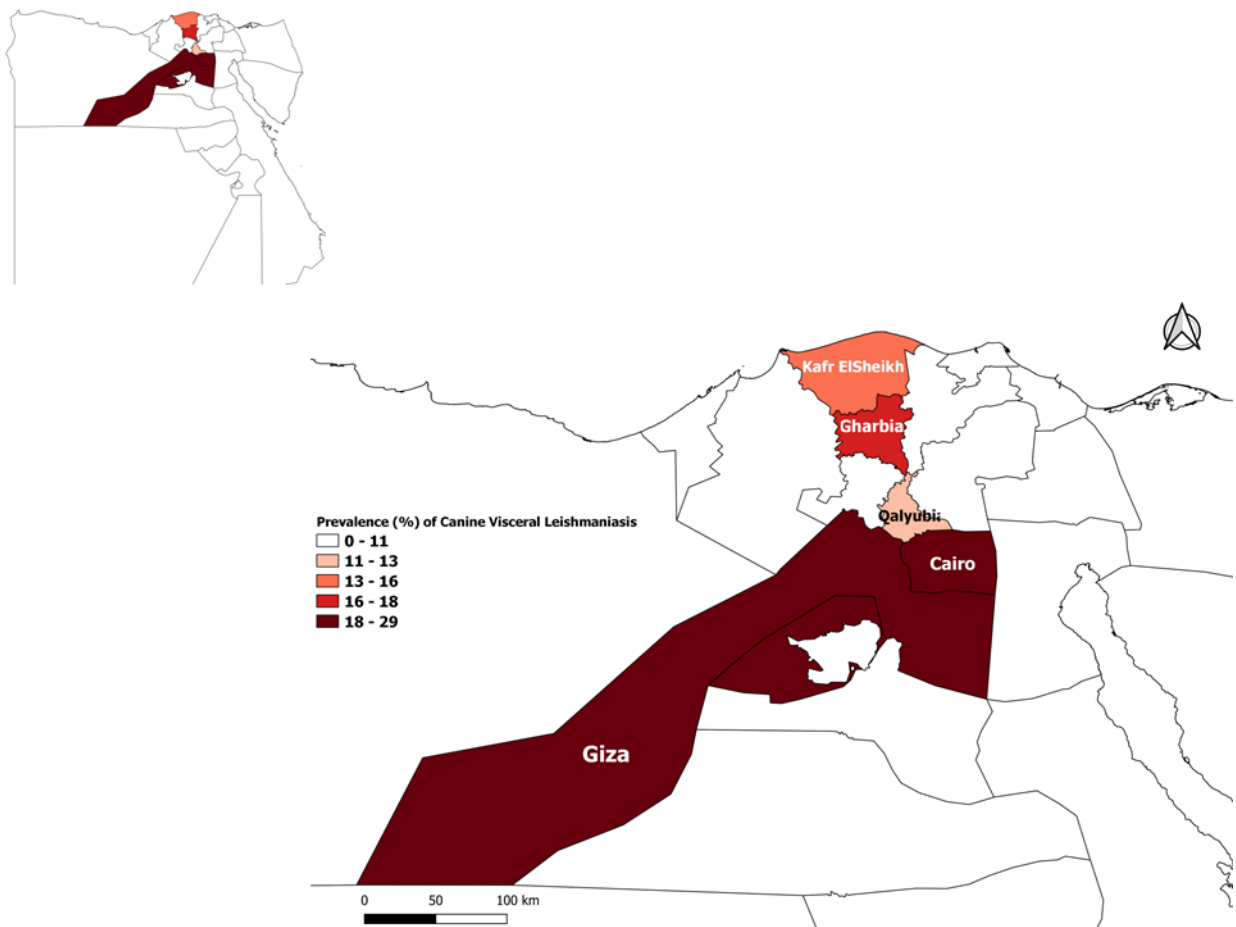


Figure 1. Map of Egypt showing governorates that were considered in the study.

During June 2019 to May 2020, a total of 450 blood samples (2 mL) were collected from the saphenous or cephalic veins of dogs that had been admitted to veterinary clinics distributed across the five governorates. Serum samples were separated by centrifugation at $3500 \times g$ for 10 min and preserved at $-20\text{ }^{\circ}\text{C}$ for serological examination. In addition, the data for each dog were gathered when the samples were collected. These data included location of the dog, its age, sex and hair length. Additionally, owners were questioned regarding the veterinary care they supplied to the dog, whether any insecticides had been applied to the animals against sand flies and the type of floor on which they slept.

2.4. Statistical Analysis

The study data were analysed through use of the statistical package for the social sciences (SPSS) software (ver. 24.0, IBM, Endicott, NY, USA). A chi-square test was applied to compare seropositivity to each variable for *L. infantum* and the results were considered significant if p was ≤ 0.05 . Univariate logistic regression was performed to determine any association between the seropositivity of exposed dogs for *L. infantum* and variables of location (five governorates), age (6–12 months, 1–2 years, 2–4 years, 4–6 years or >6 years old), sex (male or female), hair length (short or long), floor of shelter (paved or soil), level of veterinary care and whether or not insecticide had been applied. The variables with $p \leq 0.2$ were included in a multivariable logistic model to determine risk factors, odds ratios (ORs) and confidence intervals (CIs) for each significant variable. The Hosmer–Lemeshow goodness-of-fit test was used to determine the fit of the multivariable logistic regression model.

3. Results

A total of 450 blood samples were collected from asymptomatic dogs that were distributed across five governorates of Northern Egypt. In this study, antibodies against *L. infantum* were detected in 96 (21.3%) of 450 examined dogs. In general, there were significant differences ($p \leq 0.05$) between the different localities that were visited in the study. The seroprevalence rate ranged between 17.6% and 28.4%. The governorates of Cairo (28.4%) and Giza (21.7%) showed the highest rates of infection, Table 1.

Table 1. Seroprevalence of canine leishmaniasis in relation to different variables.

Factor	No of Examined Dogs	No of Positive	No of Negative	%	95% CI	Statistics
Location						
Cairo	95	27	68	28.4	20.33–38.19	$\chi^2 = 4.179$ df = 4 $p = 0.382$
Giza	115	33	82	21.7	15.2–30.1	
Qalyubia	75	9	66	18.7	11.5–28.9	
Kafr ElSheikh	80	12	68	18.8	11.7–28.7	
Gharbia	85	15	70	17.6	11–27.1	
Age						
6–12 months	30	1	29	3.3	0.2–19.1	$\chi^2 = 11.483$ df = 4 $p = 0.02^*$
1–2 years	65	9	56	13.8	6.0–25.2	
2–4 years	230	60	170	26.1	20.6–32.3	
4–6 years	95	21	74	22.1	14.5–32.0	
>6 years	30	5	25	16.7	6.3–35.4	
Sex						
Male	280	67	213	23.9	19.1–29.5	$\chi^2 = 2.975$ df = 1 $p = 0.08$
Female	170	29	141	17.1	11.9–23.7	
Breed						
German Shepherd	170	31	139	18.2	13.2–24.7	$\chi^2 = 2.891$ df = 2 $p = 0.236$
Rott Weiler	100	27	73	27	19.3–36.4	
Mongrel	180	38	142	21.1	15.7–27.6	
Hair length						
Long	150	21	129	14	9.1–20.8	$\chi^2 = 7.210$ df = 1 $p = 0.007^*$
Short	300	75	225	25	20.3–30.4	
Veterinary care						
Yes	260	32	228	12.3	8.7–17.1	$\chi^2 = 29.891$ df = 1 $p = 0.0001^*$
No	190	64	126	33.7	27.0–40.9	
Application of insecticides						
Yes	270	32	238	11.9	8.4–16.5	$\chi^2 = 36.158$ df = 1 $p = 0.0001^*$
No	180	64	116	35.6	28.7–43.1	
Floor of shelter						
Paved	310	45	265	14.5	10.8–19.1	$\chi^2 = 27.594$ df = 1 $p = 0.0001^*$
Soil	140	51	89	36.4	28.5–45.0	

* The result is significant at $p < 0.05$.

According to univariate analysis, the seroprevalence rate increased significantly with exposed dogs in the >2–4-year age group that were most likely to be infected. However, no association ($p > 0.05$) was found between sex and CanL infection.

A further analysis revealed that the seropositive rate was higher among German Shepherd (18.2%, 95% CI: 13.2–24.7) and rottweiler (27%, 95% CI: 19.3–36.4) vs another breed and in short-haired dogs (25%, 95% CI: 20.3–30.4) was higher than that in long-haired dogs, particularly among those raised in earthen-floor shelters (36.4%, 95% CI: 28.5–45). Likewise, the seroprevalence of *L. infantum* was strongly associated with lack of veterinary care (33.7%, 95% CI: 27–40.9) and with no insecticide having been applied (35.6%, 95% CI: 28.7–43.1), Table 1.

Application of a multivariate logistical regression model identified age, hair length, veterinary care, application of insecticides and life in a shelter with earthen floor as definitive predictors of *L. infantum* infection in exposed dogs. According to the multivariate model, the risk of infection increased in middle age, peaking in dogs aged 2–4 years old (OR = 12, 95% CI: 1.6–92.3), male dogs (OR = 1.93, 95% CI: 1.1–3.3) German Shepherd breed (OR = 1.59, 95% CI: 0.9–2.9) and in short-haired dogs (OR = 2.07, 95% CI: 1.2–3.6). Moreover, an absence of veterinary care (OR = 2.7, 95% CI: 1.3–5.8) or no application of insecticides (OR = 3.09, 95% CI: 1.5–6.5) were associated with higher risk of seropositivity. Regarding the floor of the shelter, dogs that lived in earthen-floored shelters seemed to be at higher risk of infection (OR = 1.42, 95% CI: 0.7–2.9), Table 2.

Table 2. Multivariable logistic analysis for risk factors of *Leishmania infantum* infection in dogs.

Variable	B ^a	SE ^b	OR ^c	95% CI ^d	p Value
Age					
1–2 years	1.602	1.094	4.96	0.6–42.3	0.143
2–4 years	2.485	1.041	12.00	1.6–92.3	0.017 *
4–6 years	2.249	1.060	9.48	1.2–75.8	0.034 *
>6 years	1.678	1.146	5.35	0.6–50.6	0.143
Sex					
Male	0.655	0.269	1.93	1.1–3.3	0.015 *
Breed					
German Shepherd	0.462	0.320	1.59	0.9–2.9	0.15
Hair length					
Short	0.729	0.285	2.07	1.2–3.6	0.011 *
Veterinary Care					
No	0.994	0.387	2.7	1.3–5.8	0.010 *
Application of Insecticides					
No	1.127	0.378	3.09	1.5–6.5	0.003 *
Floor of shelter					
Soil	0.353	0.373	1.42	0.7–2.9	0.343

^a Logistic regression coefficient; ^b Standard error; ^c Odds ratio; ^d Confidence interval; * The result is significant at $p < 0.05$.

4. Discussion

CanL caused by *L. infantum* is an emerging parasitic disease in the Mediterranean region, including Egypt. Dogs form the main reservoir and remain asymptomatic for long periods of their lives, so are considered to be a source of infection for other hosts. Until now, data on CanL in Egypt are fragmented and scarce. This first survey presents a picture of the current epidemiology of CanL in Egypt, including the associated risk factors.

In order to facilitate and improve disease control, sensitive diagnostic tests that may be used in the field are becoming increasingly important. IFATs, ELISAs and direct agglutination tests (DATs) are the most widely used serological tests in CanL screening [14,20]. ELISA is a highly applicable technique that is characterised by high sensitivity, but its specificity depends on the antigen. The specificity and sensitivity of the ELISA test that was employed to detect CanL in this investigation were 99.1% and 98.5%, respectively [21].

Overall, the seroprevalence rate of CanL in exposed dogs was 21.3%, which was in accordance with a previous rate (19.5%) that was reported in the North-Eastern and Pyrenean areas of Spain [19]. In the present study, the seroprevalence rate showed non-significant disparity ($p = 0.382$) between governorates that were visited for the study. The highest prevalence rates among the governorates were observed in Cairo and Giza. In previous studies performed in Egypt, the prevalence rate of CanL was estimated to be 66.6% by polymerase chain reaction [1] and 10% according to immunochromatography [12].

In other studies, the seroprevalence rate of CanL in exposed dogs as measured by ELISA was estimated to be 15.4% in Iran [21], 10.5% in the west of Iran [24], 5.5% in Palestine [25] and 26.6% in Pakistan [26]. Through use of IFAT, the seroprevalence rate

was estimated to be 15.4% in Sardinia, Italy [5] and ranged between 42.9% and 74.3% in Sudan [27].

These disparities could be explained by differences in sampling techniques, serological tests, ecological factors and encroachment on urban areas [2,6,12,28–30].

In the present study, dogs older than two years were more likely to become infected. This finding was in accordance with those of previous studies, which confirmed that the risk of infection increased with the age of the dogs [20,31,32]. A popular explanation is that adult dogs remain outside for long periods and that increases their chance of contact with vectors [33–36], and that the immune response against latent infection in resistant dogs may develop in older dogs [37].

Concerning the sex of the dogs, the males showed higher prevalence rates but without significant differences. This finding is consistent with those of [5,37]. This result may be attributable to the roaming behaviour of males [20] or a host immune response that results from the properties of the testosterone hormone in males [38–41].

CanL susceptibility is known to vary depending on a variety of host-related factors, such as dog breed. German Shepherd was more likely to get an infection than other examined breeds. This result ties well with previous studies [42] wherein certain breeds such as German Shepherd and Rottweiler are more susceptible to being infected. It has been suggested that the breed's relative immunocompetence derived from a cellular, parasite-specific immune response, which is linked to clinical wellness [43].

Interestingly, the risk of being seropositive to *L. infantum* increased significantly among short-haired dogs, which was also in agreement with other previous studies [44,45]. Indeed, phlebotomine sand flies are known to feed on hairless areas such as the border of the canine muzzle, which is always exposed [44]. Moreover, long hair decreases emissions of CO₂ and heat radiation from the host's body, making it less appealing to vectors [46].

Furthermore, the strong association that was found between the seropositivity of the exposed dogs and the absence of veterinary care or application of insecticides was also as previously reported [5]. This result could be due to the ineffectiveness of control measures such as use of repellent collars or insecticides and the absence of a vaccination protocol against CanL, which is related to a high risk of infection [47].

Another important risk factor that was identified in this study was the material used for the floor of the shelter. The risk of infection with *L. infantum* increased significantly among dogs that were raised in shelters made of earthen floors in comparison with those that were kept in shelters with paved floors. Since paved floors are easy to clean and covered with less organic matter than soil floors, they offer unfavourable conditions for the spreading of the vector larvae [48,49].

5. Conclusions

The results that were obtained in this study confirm the presence of antibodies against CanL in exposed dogs in some governorates of Northern Egypt and identify the risk factors that are associated with the infection. A high *L. infantum* seroprevalence rate was observed in older, male, short-haired dogs that were kept in earthen-floor shelters, particularly in the absence of veterinary care or any application of insecticides. Thus, the results show an increased number of asymptomatic dogs that acted as reservoirs for the disease and should be considered a great risk to public health. Consequently, additional information about risk factors and application of efficient control measures is a potential tool to reduce the zoonotic hazard.

Author Contributions: Conceptualization, A.S., A.A. (Abdulaziz Alouffi), Y.A., S.S. and A.A. (Abdelhamed Abdelhady); methodology, A.S.; formal analysis, A.S.; investigation, A.S.; resources, A.A. (Abdelhamed Abdelhady) and Y.A.; data curation, A.A. (Abdelhamed Abdelhady); writing—original draft preparation, A.S.; writing—review and editing, A.S., A.A. (Abdelhamed Abdelhady), Y.A., S.S. and W.S.A.-S.; project administration, A.A. (Abdulaziz Alouffi); funding acquisition, A.A. (Abdelhamed Abdelhady). All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: The study was conducted according to the guidelines of the Declaration of Benha University, and approved by Ethics Committee of Faculty of Veterinary Medicine, Benha Univeristy (BUFVTM).

Informed Consent Statement: Informed consent was obtained from all owner involved in the study.

Data Availability Statement: All data analyzed during this study are included in this published article.

Acknowledgments: The authors also thank the veterinarians for their support and help in providing data and samples collection throughout the study.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Abuowarda, M.; AbuBakr, H.O.; Ismael, E.; Shaalan, M.; Mohamed, M.A.; Aljuaydi, S.H. Epidemiological and genetic characteristics of asymptomatic canine leishmaniasis and implications for human Leishmania infections in Egypt. *Zoonoses Public Health* **2021**, *68*, 413–430. [[CrossRef](#)]
2. Malmasi, A.; Janitabar, S.; Mohebbali, M.; Akhoundi, B.; Maazi, N.; Aramoon, M.; Khorrami, N.; Seifi, H.A. Seroepidemiologic survey of canine visceral leishmaniasis in Tehran and Alborz Provinces of Iran. *J. Arthropod Borne Dis.* **2014**, *8*, 132. [[PubMed](#)]
3. Cardoso, L.; Rodrigues, M.; Santos, H.; Schoone, G.J.; Carreta, P.; Varejão, E.; van Benthem, B.; Afonso, M.O.; Alves-Pires, C.; Semião-Santos, S.J. Sero-epidemiological study of canine Leishmania spp. infection in the municipality of Alijó (Alto Douro, Portugal). *Vet. Parasitol.* **2004**, *121*, 21–32. [[CrossRef](#)] [[PubMed](#)]
4. Alvar, J.; Vélez, I.D.; Bern, C.; Herrero, M.; Desjeux, P.; Cano, J.; Jannin, J.; Boer, M.D.; Team, W.L.C. Leishmaniasis worldwide and global estimates of its incidence. *PLoS ONE* **2012**, *7*, e35671. [[CrossRef](#)] [[PubMed](#)]
5. Tamponi, C.; Scarpa, F.; Carta, S.; Knoll, S.; Sanna, D.; Gai, C.; Pipia, A.P.; Dessì, G.; Casu, M.; Varcasia, A. Seroprevalence and risk factors associated with Leishmania infantum in dogs in Sardinia (Italy), an endemic island for leishmaniasis. *Parasitol. Res.* **2020**, *120*, 289–300. [[CrossRef](#)]
6. Tabbabi, A. Review of leishmaniasis in the Middle East and North Africa. *Afr. Health Sci.* **2019**, *19*, 1329–1337. [[CrossRef](#)]
7. Medkour, H.; Davoust, B.; Dulieu, F.; Maurizi, L.; Lamour, T.; Marié, J.-L.; Mediannikov, O. Potential animal reservoirs (dogs and bats) of human visceral leishmaniasis due to Leishmania infantum in French Guiana. *PLoS Negl. Trop. Dis.* **2019**, *13*, e0007456. [[CrossRef](#)]
8. Dantas-Torres, F.; de Brito, M.E.F.; Brandão-Filho, S.P. Seroepidemiological survey on canine leishmaniasis among dogs from an urban area of Brazil. *Vet. Parasitol.* **2006**, *140*, 54–60. [[CrossRef](#)]
9. Najafi, L.; Omidian, M.; Rezaei, Z.; Shahabi, S.; Ghorbani, F.; Arefkhan, N.; Mohebbali, M.; Zariaei, Z.; Sarkari, B. Molecular and serological evaluation of zoonotic visceral leishmaniasis in dogs in a rural area of Fars province, southern Iran, as a source of Leishmania infantum infection. *Vet. Med. Sci.* **2021**, *7*, 1082–1089. [[CrossRef](#)]
10. Solano-Gallego, L.; Miró, G.; Koutinas, A.; Cardoso, L.; Pennisi, M.G.; Ferrer, L.; Bourdeau, P.; Oliva, G.; Baneth, G. LeishVet guidelines for the practical management of canine leishmaniosis. *Parasites Vectors* **2011**, *4*, 1–16. [[CrossRef](#)]
11. Esteve, L.; Vargas, C.; de León, C.V. The role of asymptomatics and dogs on leishmaniasis propagation. *Math. Biosci.* **2017**, *293*, 46–55. [[CrossRef](#)]
12. Rosypal, A.C.; Bowman, S.S.; Epps, S.A.; El Behairy, A.; Hilali, M.; Dubey, J. Serological survey of dogs from Egypt for antibodies to Leishmania species. *J. Parasitol.* **2013**, *99*, 170–171. [[CrossRef](#)]
13. Morsy, T.A.; Schnur, L.F.; Feinsod, F.M.; Salem, A.M.; Wahba, M.M.; El Said, S.M. Natural infections of Leishmania major in domestic dogs from Alexandria, Egypt. *Am. J. Trop. Med. Hyg.* **1987**, *37*, 49–52. [[CrossRef](#)]
14. Bessat, M.; Okpanma, A.; Shanat, E. Leishmaniasis: Epidemiology, control and future perspectives with special emphasis on Egypt. *J. Trop. Dis.* **2015**, *2*, 1–10.
15. Awadalla, H.; Mansour, N.; Mohareb, E. Further characterization of Leishmania isolates from children with visceral infection in Alexandria area, Egypt. *Trans. R. Soc. Trop. Med. Hyg.* **1987**, *81*, 915–917. [[CrossRef](#)]
16. Faris, R.; Massoud, A.; El Said, S.; Gadallah, M.; Feinsod, F.; Saar, A.; Londner, M.; Rosen, G. The epidemiology of human visceral leishmaniasis in El Agamy (Alexandria Governorate), Egypt: Serosurvey and case/control study. *Ann. Trop. Med. Parasitol.* **1988**, *82*, 445–452. [[CrossRef](#)] [[PubMed](#)]
17. Fryauff, D.J.; Modi, G.B.; Mansour, N.S.; Kreutzer, R.D.; Soliman, S. Epidemiology of Cutaneous Leishmaniasis at a Focus Monitored by the Multinational Force and Observers in the Northeastern Sinai Desert of Egypt. *Am. J. Trop. Med. Hyg.* **1993**, *49*, 598–607. [[CrossRef](#)] [[PubMed](#)]
18. Hamadto, H.A.; El Fkahan, A.F.; Morsy, T.A.; Farrag, A.; MK, A.M. Re-evaluation of zoonotic cutaneous leishmaniasis status in North Sinai Governorate, Egypt. *J. Egypt. Soc. Parasitol.* **2003**, *33*, 687–694.
19. Vélez, R.; Ballart, C.; Domenech, E.; Abras, A.; Fernández-Arévalo, A.; Gómez, S.A.; Tebar, S.; Muñoz, C.; Cairó, J.; Gállego, M. Seroprevalence of canine Leishmania infantum infection in the Mediterranean region and identification of risk factors: The example of North-Eastern and Pyrenean areas of Spain. *Prev. Vet. Med.* **2019**, *162*, 67–75. [[CrossRef](#)]

20. Rombolà, P.; Barlozzari, G.; Carvelli, A.; Scarpulla, M.; Iacoponi, F.; Macrì, G. Seroprevalence and risk factors associated with exposure to *Leishmania infantum* in dogs, in an endemic Mediterranean region. *PLoS ONE* **2021**, *16*, e0244923. [[CrossRef](#)]
21. Mahshid, M.; Baharak, A.; Iraj, S.; Sina, K.; Javad, K.; Mehdi, B. Seroprevalence of canine visceral leishmaniasis in southeast of Iran. *J. Parasit. Dis.* **2014**, *38*, 218–222. [[CrossRef](#)]
22. Costa, M.M.; Penido, M.; Dos Santos, M.S.; Doro, D.; de Freitas, E.; Michalick, M.S.M.; Grimaldi, G.; Gazzinelli, R.T.; Fernandes, A.P. Improved canine and human visceral leishmaniasis immunodiagnosis using combinations of synthetic peptides in enzyme-linked immunosorbent assay. *PLoS Negl. Trop. Dis.* **2012**, *6*, e1622. [[CrossRef](#)] [[PubMed](#)]
23. Pourhoseingholi, M.A.; Vahedi, M.; Rahimzadeh, M. Sample size calculation in medical studies. *Gastroenterol. Hepatol. Bed Bench* **2013**, *6*, 14. [[PubMed](#)]
24. Gharekhani, J.; Pourmahdi Borujeni, M.; Sazmand, A. Seroprevalence of Visceral Leishmaniasis in Stray Dogs of Hamedan, West of Iran in 2018. *J. Med Microbiol. Infect. Dis.* **2020**, *8*, 71–75. [[CrossRef](#)]
25. Abdeen, Z.A.; Sawalha, S.S.; Eisenberger, C.L.; Khanfar, H.M.; Greenblatt, C.L.; Yousef, O.; Schnur, L.F.; Azmi, K.; Warburg, A.; Bader, K.A. Epidemiology of visceral leishmaniasis in the Jenin District, West Bank: 1989–1998. *Am. J. Trop. Med. Hyg.* **2002**, *66*, 329–333. [[CrossRef](#)] [[PubMed](#)]
26. Rab, M.; Frame, I.; Evans, D. The role of dogs in the epidemiology of human visceral leishmaniasis in northern Pakistan. *Trans. R. Soc. Trop. Med. Hyg.* **1995**, *89*, 612–615. [[CrossRef](#)]
27. Dereure, J.; El-Safi, S.H.; Bucheton, B.; Boni, M.; Kheir, M.M.; Davoust, B.; Pralong, F.; Feugier, E.; Lambert, M.; Dessein, A. Visceral leishmaniasis in eastern Sudan: Parasite identification in humans and dogs; host-parasite relationships. *Microbes Infect.* **2003**, *5*, 1103–1108. [[CrossRef](#)]
28. Selim, A.; Almohammed, H.; Abdelhady, A.; Alouffi, A.; Alshammari, F.A. Molecular detection and risk factors for *Anaplasma platys* infection in dogs from Egypt. *Parasites Vectors* **2021**, *14*, 1–6. [[CrossRef](#)]
29. Selim, A.; Manaa, E.; Khater, H. Seroprevalence and risk factors for lumpy skin disease in cattle in Northern Egypt. *Trop. Anim. Health Prod.* **2021**, *53*, 1–8. [[CrossRef](#)]
30. Selim, A.; Manaa, E.A.; Waheed, R.M.; Alanazi, A.D. Seroprevalence, associated risk factors analysis and first molecular characterization of *Chlamydia abortus* among Egyptian sheep. *Comp. Immunol. Microbiol. Infect. Dis.* **2021**, *74*, 101600. [[CrossRef](#)]
31. Martín-Sánchez, J.; Morales-Yuste, M.; Acedo-Sánchez, C.; Barón, S.; Díaz, V.; Morillas-Márquez, F. Canine leishmaniasis in southeastern Spain. *Emerg. Infect. Dis.* **2009**, *15*, 795. [[CrossRef](#)]
32. Sauda, F.; Malandrucchio, L.; Macrì, G.; Scarpulla, M.; De Liberato, C.; Terracciano, G.; Fichi, G.; Berrilli, F.; Perrucci, S. *Leishmania infantum*, *Dirofilaria* spp. and other endoparasite infections in kennel dogs in central Italy. *Parasite* **2018**, *25*, 2–10. [[CrossRef](#)]
33. Matos, M.M.; Figueira, K.D.; Amora, S.; Suassuna, A.; Ahid, S.M.M.; Alves, N. Ocorrência da leishmaniose visceral em cães em Mossoró, Rio Grande do Norte. *Ciênc. Anim.* **2006**, *16*, 51–54.
34. Selim, A.; Megahed, A.A.; Kandeel, S.; Abdelhady, A. Risk factor analysis of bovine leukemia virus infection in dairy cattle in Egypt. *Comp. Immunol. Microbiol. Infect. Dis.* **2020**, *72*, 101517. [[CrossRef](#)]
35. Selim, A.; Radwan, A. Seroprevalence and molecular characterization of West Nile Virus in Egypt. *Comp. Immunol. Microbiol. Infect. Dis.* **2020**, *71*, 101473. [[CrossRef](#)] [[PubMed](#)]
36. Selim, A.; Radwan, A.; Arnaout, F.; Khater, H. The Recent Update of the Situation of West Nile Fever among Equids in Egypt after Three Decades of Missing Information. *Pak. Vet. J.* **2020**, *40*, 390–393.
37. Gálvez, R.; Miró, G.; Descalzo, M.; Nieto, J.; Dado, D.; Martín, O.; Cubero, E.; Molina, R. Emerging trends in the seroprevalence of canine leishmaniasis in the Madrid region (central Spain). *Vet. Parasitol.* **2010**, *169*, 327–334. [[CrossRef](#)] [[PubMed](#)]
38. Živičnjak, T.; Martinković, F.; Marinculić, A.; Mrljak, V.; Kučer, N.; Matijatko, V.; Mihajević, Ž.; Barić-Rafaj, R. A seroepidemiologic survey of canine visceral leishmaniasis among apparently healthy dogs in Croatia. *Vet. Parasitol.* **2005**, *131*, 35–43. [[CrossRef](#)]
39. Selim, A.; Abdelhady, A. The first detection of anti-West Nile virus antibody in domestic ruminants in Egypt. *Trop. Anim. Health Prod.* **2020**, *52*, 3147–3151. [[CrossRef](#)]
40. Selim, A.; Ali, A.-F. Seroprevalence and risk factors for *C. burnetii* infection in camels in Egypt. *Comp. Immunol. Microbiol. Infect. Dis.* **2020**, *68*, 101402. [[CrossRef](#)]
41. Selim, A.; Marawan, M.A.; Ali, A.-F.; Manaa, E.; AbouelGhaut, H.A. Seroprevalence of bovine leukemia virus in cattle, buffalo, and camel in Egypt. *Trop. Anim. Health Prod.* **2020**, *52*, 1207–1210. [[CrossRef](#)]
42. Burnham, A.C.; Ordeix, L.; Alcover, M.M.; Martínez-Orellana, P.; Montserrat-Sangrà, S.; Willen, L.; Spitzova, T.; Volf, P.; Solano-Gallego, L. Exploring the relationship between susceptibility to canine leishmaniasis and anti-*Phlebotomus perniciosus* saliva antibodies in Ibizan hounds and dogs of other breeds in Mallorca, Spain. *Parasites Vectors* **2020**, *13*, 1–15. [[CrossRef](#)] [[PubMed](#)]
43. Solano-Gallego, L.; Llull, J.; Ramis, A.; Fernández-Bellón, H.; Rodríguez, A.; Ferrer, L.; Alberola, J. Longitudinal study of dogs living in an area of Spain highly endemic for leishmaniasis by serologic analysis and the leishmanin skin test. *Am. J. Trop. Med. Hyg.* **2005**, *72*, 815–818. [[CrossRef](#)] [[PubMed](#)]
44. Cortes, S.; Vaz, Y.; Neves, R.; Maia, C.; Cardoso, L.; Campino, L. Risk factors for canine leishmaniasis in an endemic Mediterranean region. *Vet. Parasitol.* **2012**, *189*, 189–196. [[CrossRef](#)] [[PubMed](#)]
45. Silva, J.C.F.D.; Costa, R.T.D.; Siqueira, A.M.; Coelho, G.L.L.M.; Costa, C.A.D.; Mayrink, W.; Vieira, E.P.; Silva, J.C.D. Epidemiology of canine visceral leishmaniasis in the endemic area of Montes Claros Municipality, Minas Gerais State, Brazil. *Vet. Parasitol.* **2003**, *111*, 161–173. [[CrossRef](#)]

46. Belo, V.S.; Struchiner, C.J.; Werneck, G.L.; Barbosa, D.S.; de Oliveira, R.B.; Neto, R.G.T.; da Silva, E.S. A systematic review and meta-analysis of the factors associated with *Leishmania infantum* infection in dogs in Brazil. *Vet. Parasitol.* **2013**, *195*, 1–13. [[CrossRef](#)] [[PubMed](#)]
47. Lopes, P.M.; Sorte, E.D.C.B.; Gasparetto, N.D.; Oliveira, C.M.; Almeida, A.D.B.P.F.D.; Sousa, V.R.F. Seroprevalence and risk factors associated with visceral leishmaniasis in dogs in Jaciara, State of Mato Grosso. *Rev. Soc. Bras. Med. Trop.* **2014**, *47*, 791–795. [[CrossRef](#)] [[PubMed](#)]
48. Coura-Vital, W.; Marques, M.J.; Veloso, V.M.; Roatt, B.M.; Aguiar-Soares, R.D.D.O.; Reis, L.E.S.; Braga, S.L.; Morais, M.H.F.; Reis, A.B.; Carneiro, M. Prevalence and factors associated with *Leishmania infantum* infection of dogs from an urban area of Brazil as identified by molecular methods. *PLoS Negl. Trop. Dis.* **2011**, *5*, e1291. [[CrossRef](#)]
49. De Almeida Leal, G.G.; Carneiro, M.; da Costa Pinheiro, A.; Marques, L.A.; Ker, H.G.; Reis, A.B.; Coura-Vital, W. Risk profile for *Leishmania* infection in dogs coming from an area of visceral leishmaniasis reemergence. *Prev. Vet. Med.* **2018**, *150*, 1–7. [[CrossRef](#)]